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\*CORRESPONDENCE Yu-Tong He Meyutong@hebmu.edu.cn

RECEIVED 29 November 2024 ACCEPTED 11 March 2025 PUBLISHED 31 March 2025

#### CITATION

Liu Y-Y, Fu Y-F, Yang W-Y, Li Z, Lu Q, Su X, Shi J, Wu S-Q, Liang D and He Y-T (2025) DKK3 and SERPINB5 as novel serum biomarkers for gastric cancer: facilitating the development of risk prediction models for gastric cancer. *Front. Oncol.* 15:1536491. doi: 10.3389/fonc.2025.1536491

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## DKK3 and SERPINB5 as novel serum biomarkers for gastric cancer: facilitating the development of risk prediction models for gastric cancer

Yan-Yu Liu<sup>1,2</sup>, Yan-Fang Fu<sup>1,2</sup>, Wan-Yu Yang<sup>1,2</sup>, Zheng Li<sup>1,2</sup>, Qian Lu<sup>1,2</sup>, Xin Su<sup>1,2</sup>, Jin Shi<sup>2</sup>, Si-Qi Wu<sup>2</sup>, Di Liang<sup>2</sup> and Yu-Tong He<sup>1,2\*</sup>

<sup>1</sup>School of Public Health, Hebei Medical University, Shijiazhuang, China, <sup>2</sup>Cancer Institute, The Fourth Hospital of Hebei Medical University, Shijiazhuang, China

The existing gastric cancer (GC) risk prediction models based on biomarkers are limited. This study aims to identify new promising biomarkers for GC to develop a risk prediction model for effective assessment, screening, and early diagnosis. This study was conducted utilizing a large combined cohort for upper gastrointestinal cancer that was established in Hebei Province, China. General macro risk factors, Helicobacter pylori (H.pylori) infection status, and protein biomarkers were collected through questionnaire surveys and laboratory tests. Novel GC biomarkers were explored using data-independent acquisition (DIA) proteomics and enzyme-linked immunosorbent assay (ELISA). Multiple machine learning algorithms were used to identify key predictors for the GC risk prediction model, which was validated with an independent external cohort from multiple hospitals. A total of 530 participants aged 40 to 74 were analyzed, with 104 ultimately diagnosed with GC. Significant biomarkers in GC patients were identified by DIA combined ELISA, including elevated Keratin 7 (KRT7) and Mammary fibrostatin (SERPINB5) (P<0.001) and decreased Dickkopf-associated protein 3 (DKK3) (P<0.001). Factors such as sex, age, smoking status, alcohol consumption, family history of GC, H. pylori infection, DKK3 and SERPINB5 were used to create a multidimensional risk prediction model for GC. This model achieved an area under the curve (AUC) of 0.938 (95% confidence interval: 0.913-0.962). The risk prediction model developed in this study shows high accuracy and practical utility, serving as an effective preliminary screening tool for identifying high-risk individuals for GC.

#### KEYWORDS

gastric cancer, risk prediction, proteomics, biomarkers, screening

Gastric cancer (GC) is the fifth most common cancer worldwide, with about 968,350 new cases annually, representing 4.9% of all new cancer diagnoses. China accounts for 37.1% of global GC incidence and 39.4% of related mortality (1). Despite advancements in diagnosis and treatment, patient prognosis remains unsatisfactory, with survival rates varying globally from 5% to 69% (2). In China, the five-year survival rate for GC patients is only 35.1% (3). Screening has been proven to be an effective approach to improve survival rates, and since the 21st century, China has implemented numerous screening programs for GC (4). However, these programs lack effective verification and evaluation. Meanwhile, GC screening programs rely mainly on endoscopy, but concerns regarding its invasive nature, the requirement for skilled endoscopists and pathologists, and the high costs in developing countries have been problems that have beset the prevention of GC (5). Therefore, accurate risk prediction is of significance as screening resources can be allocated more efficiently.

While some GC risk prediction models have been developed to support risk stratification strategies, there is a lack of comparative evaluation and no uniform conclusion, due to variations in study design, statistical methods, and model performance (6). Moreover, most diagnostic models suffer from inadequate sample sizes and rely solely on univariate analysis to select candidate variables, which may lead to the omission of important predictors. Additionally, comprehensive model evaluation is often neglected, and the vast majority of predictive models lack either internal or external validation, limiting their clinical applicability. Furthermore, most existing models are based solely on macro-epidemiological factors and known molecular markers. Apart from Helicobacter pylori (H. pylori), molecular markers of GC are largely unidentified, and the etiology underlying GC remains to be fully elucidated. Taken together, the published risk prediction models constructed based on GC molecular markers are extremely limited, particularly those based on prospective studies, and there is a critical need for wellperforming biomarkers.

Proteomics has gradually been applied to the early diagnosis of GC due to its high serum proteomic content combined with systemic and local tumor features. The equipment for measuring serum protein is also very mature (7, 8). Exploring novel GC screening markers based on proteomics may improve efficiency in risk stratification and further optimize GC risk prediction models. Currently, some proteomic studies have been applied to the therapeutic targets and prognosis prediction of GC, but few have been applied to population risk stratification. Most of these studies lack control for possible confounding factors and multiple comparison correction or are limited by small sample size (9).

This study aims to identify promising new biomarkers for GC screening. And based on the new markers, we will develop and optimize a population-specific risk prediction model for GC in Hebei Province, and provide an effective assessment and screening method for the general population.

## Methods

### Study design

Two large screening cohorts for upper gastrointestinal cancer were established in Hebei Province, China. These two cohorts were combined to serve as the model derivation cohort for this study. The Cancer Hospital of the Chinese Academy of Medical Sciences initiated a project aimed at creating a large-scale population screening cohort for upper gastrointestinal cancer across six provinces: Henan, Hebei, Shandong, Jiangsu, Shanxi, and Hunan (10). This project successfully developed a multicenter screening cohort, database, and biobank encompassing 110,000 individuals. Participants eligible for this study were residents aged 40-74 years old, with no personal history of cancer and who had not undergone endoscopy in the preceding 3 years. As of 2019, the baseline survey was completed at all project sites within Hebei Province. Specifically, a total of 24,679 individuals participated in this study.

Additionally, another initiative has been conducting upper gastrointestinal cancer screenings as part of the Cancer Screening Program in Urban China (CanSPUC) in Hebei since 2012 (11). Individuals are screened using questionnaires and H. pylori tests, then were advised to undergo endoscopic examinations and provide five milliliters of venous blood for biomarker analysis. Participants were aged between 45 and 74 years and had no prior history of diagnosis or treatment for significant heart, brain, lung, or kidney dysfunctions, serious mental disorders, or cancer. We selected data from a total of 65,435 individuals within the CanSPUC cohort at baseline survey out of an initial pool of 210,381 individuals, which included participants from all eleven cities in Hebei Province.

For the purpose of independent verification, we utilized a retrospective cohort comprising clinical early-stage GC patients and healthy individuals from multiple hospitals in Hebei Province. This cohort included 81 gastric cancer patients and 210 healthy participants. The detailed study design and cohorts are depicted in Figure 1.

This study has been approved by the Ethics Committees of the Cancer Hospital, Chinese Academy of Medical Sciences and the Fourth Hospital of Hebei Medical University, in accordance with the principles of the Helsinki Declaration. All participants have signed informed consent forms.

### Definition of macro variables

The questionnaire primarily consisted of general demographic information such as age and sex, behavioral factors including smoking and drinking habits, frequency of consumption of a total of 10 diets such as fresh vegetables and fruits, history of digestive diseases, family history of tumors, and body mass index (BMI). All variables are included in the model as categorical variables. Age is divided into two groups based on whether they have reached the age of 60. BMI is divided into three groups based on the threshold values of 18.5 and 23.9 (kg/m<sup>2</sup>). Marital status was dichotomized into unmarried (separated, divorced, widowed, or never married)



and married at the time of interview. Educational level was categorized as primary and below, secondary school, and college and above. Occupational exposure was categorized as absent and present. All dietary habits are categorized as either seldom or frequently based on self-reported questionnaire results from participants. All diseases history and family history are classified as present or absent. Smoking status was classified as smoker, exsmoker, and non-smoker based on current smoking status and past smoking history. Drinking status was categorized as drinker and non-drinker based on the history of alcohol consumption.

# Preliminary experiment for differential protein screening

A preliminary case-control study was conducted to identify potential biomarkers by comparing serum protein profiles between early GC patients and healthy controls. Five cases of early clinical GC were collected, and healthy controls were matched 1:1 based on age and sex. The GC patients were recruited from the Fourth Hospital of Hebei Medical University, while the healthy controls were obtained from the hospital's health examination population. This pre-experiment aimed to identify differential proteins that could serve as candidate biomarkers for further validation in the main cohort study. The findings from this preliminary study will guide the subsequent validation within the main cohort. The selection criteria for the early GC patients in the preliminary study were as follows: (1) age between 40 and 74 years; (2) histopathologically confirmed gastric cancer and clinically diagnosed as stage I; (3) no acute infections, allergies, or autoimmune diseases within the past three months that could affect blood biomarker expression; (4) no blood transfusions within the past three months; (5) normal liver and kidney function, as well as routine blood tests; (6) no history of other malignancies; and (7) no prior treatment with surgery, chemotherapy, or radiotherapy. The inclusion criteria for the healthy controls were as follows: (1) age between 40 and 74 years; (2) no history of malignancies or other major diseases; (3) no acute infections, allergies, or autoimmune diseases within the past three months that could affect blood biomarker expression; and (4) normal liver and kidney function, as well as routine blood tests.

Blood samples from GC patients who did not undergo chemoradiation were collected on an empty stomach before surgery and anesthesia. The serum was separated by centrifugation at 2000 rpm for 10 minutes after the coagulating tube was placed for 30 minutes as pre-experimental samples.

# Data-independent acquisition proteomic analysis

Data-independent acquisition (DIA) proteomic analysis was conducted on serum samples from both cases and controls in order

to identify differential proteins. The detailed DIA process followed the procedures outlined below, including protein extraction, determination of protein concentration, protease hydrolysis, desalination, peptide classification, establishment of a DDA reference map, data collection using DIA methodology, highprecision mass spectrometry detection, and qualitative database search for bioinformatics analysis (12). The DIA analysis was performed using a Q Exactive HF-X Hybrid Quadrupole-Orbitrap Mass Spectrometer (Thermo Fisher Scientific) coupled with a highperformance liquid chromatography system (EASY nLC 1200, Thermo Fisher Scientific). Quality control measures were implemented for each experimental session. These measures included instrument calibration, the use of internal standards, data replication, and peptide identification validation against a decoy database to control the false discovery rate (FDR) below 1%. Identification of target proteins involved a combination of domestic and international research findings as well as enrichment analysis of GO (Gene Ontology) and KEGG (Kyoto Encyclopedia of Genes and Genomes) databases. Given the complexity of biological samples and the potential for non-normal distributions in protein expression levels, differential proteins were analyzed using a twosided Wilcoxon rank-sum test (p < 0.05 and fold change > 2 or <0.5). Significantly enriched GO functions and KEGG pathways were determined using Fisher's exact test followed by Benjamini-Hochberg correction with p < 0.05.

### Enzyme-linked immunosorbent assay

The diagnostic validity of the new biomarkers for GC was confirmed through enzyme-linked immunosorbent assay (ELISA) in the serum samples of derivation cohort. The levels of the new biomarkers in both case and control groups were determined using the corresponding proteins ELISA kit (Jianglai, Shanghai), following the manufacturer's instructions. Briefly, 100  $\mu$ l of serum diluent was added to a 96-well plate coated with human target protein. Then 100  $\mu$ l of HRP-conjugated mixed solution was added to each well, and the plate was incubated for 0.5 h at 37°C. After several washes, the color reaction was developed with the substrate solution and blocked with stop solution. The optical densities were measured at 450 nm.

### Definition of outcome

The study outcome was a confirmed diagnosis of GC, as identified by the International Statistical Classification of Disease-10 code C16. The identification was achieved through screening detection, active follow-up, and passive follow-up. Participants with positive screening results were subsequently followed up by phone or retrieval of medical records, and the entire cohort population was also passively matched using the population-based Hebei Cancer Registry and Cause of Death Database until June 30, 2024 to obtain final diagnoses and outcomes.

## Development of cancer risk prediction models

The predictive model was developed in the derivation cohort and then validated in the independent validation cohort. Using GC occurrence as the dependent variable and the selected variables from the derivation cohort as independent variables, the GC risk prediction models were developed using multiple logistic regression. All potential macro risk factors are initially included in the underlying models. Four machine learning (ML) algorithms - logistic regression (LR), random forest (RF), lasso regression and eXtreme gradient boosting (XGBoost) - were employed to screen the variables ultimately included in the model. LR utilized a stepwise regression method for variable screening. RF used the reduction of average accuracy rate as the evaluation index and ranked the importance of variables. Lasso regression employed minimum mean to filter variables, while XGBoost ranked variables by importance. Predictive variables meeting the criterion of p < 0.05 were selected for inclusion in the model. Variable selection was achieved using R packages "MASS", "randomForest", "lasso" and "XGBoost".

Based on proteomic analysis, we investigated and identified novel GC markers. Subsequently, we integrated these markers with all variables in the cohort to identify predictors for GC through machine learning techniques. Therefore, the GC risk prediction model constructed by the final model contains the following risk factors:

Logit  $P = \beta_0 + \beta_{1*}$  macro risk factors  $+ \beta_{2*} H$ . pylori +  $\beta_{3*}$  novel markers

Where *Logit P* is the expected the risk of incident GC;  $\beta_0$  is the intercept.  $\beta_1$ ,  $\beta_2$ , and  $\beta_3$  are the coefficients of each variable.

### Statistical analysis

The descriptive statistics of baseline characteristics are presented according to the risk of GC. Categorical variables are displayed as frequencies and percentages. Chi-square tests were used for comparing differences in categorical variables. All these available factors were comprehensively screened in our prediction model to enhance its accuracy and to identify potential unrecognized risk factors for GC. The overall performance of the models was assessed based on the area under curve (AUC), while their calibration was evaluated using Hosmer-Lemeshow chi-square test results (H-L  $\chi^2$ ). The prediction models were internally validated using bootstrapping (40 replications).

Two-sided tests were used in all studies, and p < 0.05 was considered to be statistically significant. Statistical analyses were performed using R (v.4.2.0) software.

## Results

## Characteristics of the study population

The combined cohort consists of 90,114 individuals. After excluding 24,955 participants with missing key variables or those who were lost to follow-up regarding outcomes, we conducted ELISA assays on preserved serum samples from 530 individuals of the remaining population. During the study period, a total of 530 eligible

TABLE 1 Baseline characteristics of the derivation population.

participants were enrolled in the derivation cohort, among whom 104 eventually developed GC. The characteristics of all participants are presented in Table 1. The majority of the GC participants were aged 60 and above, accounting for 67.3%. Additionally, males comprised 72.1% of the GC participant group, and 52.9% of the GC participants were overweight or obese. A total of 291 participants, comprising 81 patients with GC and 210 healthy individuals, were included for independent validation. The baseline information for the independent validation cohort can be found in Supplementary Table 1.

Characteristics	Levels	Non-gastric cancer (N=426)	Gastric cancer (N=104)	р
Sam (0/)	Male	204 (47.9)	75 (72.1)	< 0.001
Sex (%)	Female	222 (52.1)	29 (27.9)	
A (	<60	228 (53.5)	34 (32.7)	<0.001
Age (years) (%)	≥60	198 (46.5)	70 (67.3)	
	<18.5	2 (0.5)	3 (2.9)	0.066
BMI (%)	18.5-23.9	182 (42.7)	46 (44.2)	
	≥23.9	242 (56.8)	55 (52.9)	
	Unmarried	16 (3.8)	4 (3.8)	1.000
Marriage (%)	Married	410 (96.2)	100 (96.2)	
	Primary and below	177 (41.5)	66 (63.5)	< 0.001
Education (%)	Secondary school	145 (34.0)	38 (36.5)	
	College and above	104 (24.4)	0 (0.0)	
	Absent	334 (78.4)	93 (89.4)	0.016
Occupational exposure (%)	Present	92 (21.6)	11 (10.6)	
	Never smoker	288 (67.6)	57 (54.8)	0.002
Smoking status (%)	Smoker	99 (23.2)	28 (26.9)	
	Ex-smoker	39 (9.2)	19 (18.3)	
	Absent	289 (67.8)	53 (51.0)	0.001
Drinking status (%)	Present	137 (32.2)	51 (49.0)	
	Seldom	98 (23.0)	3 (2.9)	< 0.001
Vegetables (%)	Frequently	328 (77.0)	101 (97.1)	
	Seldom	169 (39.7)	17 (16.3)	< 0.001
Fruits (%)	Frequently	257 (60.3)	87 (83.7)	
	Seldom	202 (47.4)	19 (18.3)	< 0.001
Milk, meat and egg production (%)	Frequently	224 (52.6)	85 (81.7)	
	Seldom	252 (59.2)	92 (88.5)	< 0.001
Pickled food (%)	Frequently	174 (40.8)	12 (11.5)	
	Seldom	313 (73.5)	99 (95.2)	< 0.001
Fried food (%)	Frequently	113 (26.5)	5 (4.8)	
Hot diet (%)	Seldom	210 (49.3)	92 (88.5)	< 0.001

(Continued)

#### TABLE 1 Continued

Characteristics	Levels	Non-gastric cancer (N=426)	Gastric cancer (N=104)	р
	Frequently	216 (50.7)	12 (11.5)	
	Absent	219 (51.4)	87 (83.7)	< 0.001
History of UD1Ds (%)	Present	207 (48.6)	17 (16.3)	
	Absent	381 (89.4)	100 (96.2)	0.053
Diabetes (%)	Present	45 (10.6)	4 (3.8)	
	Absent	280 (65.7)	77 (74.0)	0.133
rypertension (%)	Present	146 (34.3)	27 (26.0)	
	Absent	292 (68.5)	100 (96.2)	<0.001
riypernpenna (%)	Present	134 (31.5)	4 (3.8)	
	Absent	193 (45.3)	78 (75.0)	< 0.001
Family instory of cancer (%)	Present	233 (54.7)	26 (25.0)	
Family history of gastric	Absent	315 (73.9)	67 (64.4)	0.617
cancer (%)	Present	111 (26.1)	37 (35.6)	
	Negative	297 (69.7)	54 (51.9)	0.001
п.Руюп (%)	Positive	129 (30.3)	50 (48.1)	
KRT7 (%)	Negative	252 (59.2)	41 (39.4)	< 0.001
	Positive	174 (40.8)	63 (60.6)	
	Negative	148 (34.7)	68 (65.4)	< 0.001
DARS (%)	Positive	278 (65.3)	36 (34.6)	
CEDDINDS (0/)	Negative	328 (77.0)	21 (20.2)	< 0.001
SERFINDS (%)	Positive	98 (23.0)	83 (79.8)	

BMI, body mass index; History of UDTSs, History of upper digestive tract diseases; H.Pylori, Helicobacter pylori; KET 7, Keratin 7; DKK 3, Dickkopf-associated protein 3; SERPINB5, Mammary fibrostatin.

# Selection of novel markers for GC based on proteomics

Partial least squares discriminant analysis (PLS-DA) score plots revealed a significant disparity in proteomic components between GC patients and non-GC individuals (Figure 2A). A total of 4837 proteins were identified using DIA. Among them, 60 proteins showed differential expression between the case and control groups, with 37 up-regulated and 23 down-regulated proteins. A volcano plot was created to visually display the differential protein expression patterns (Figure 2B).

In the GO enrichment analysis of case and control, differential proteins were significantly enriched in biological functions such as intermediate filament organization, nucleolus, and chromatin structural components (Figure 2C). While in the KEGG enrichment analysis, pathways related to transcriptional dysregulation in cancer were significantly enriched (Figure 2D). Based on previous research and taking into account the enrichment findings of GO and KEGG, we have identified 3 proteins from the pool of differential proteins for further ELISA analysis (13–15). These proteins are Keratin 7 (KRT 7), Dickkopf-associated protein 3 (DKK 3), and Mammary fibrostatin (Maspin / SERPINB5).

## Validation of new markers by ELISA

We further assessed the expression levels of DKK3, KRT7, and SERPINB5 in serum samples from both GC patients and non-GC individuals using ELISA. In comparison to the non-GC samples, we observed a three-fold increase in the SERPINB5 level in GC samples (mean: 8.56 ng/ml vs 2.06 ng/ml), a significant increase in the KRT7 level (mean: 275.88 ng/ml vs 217.14 ng/ml), and a decrease in the DKK3 level (mean: 11.00 ng/ml vs 15.47 ng/ml) (Figure 3). Based on the median concentrations of biomarkers measured by ELISA, the three biomarkers are categorized into negative and positive groups. The categorical variables of the biomarkers are incorporated into the prediction models for variable screening.



Proteomic findings. (A) Partial least square discriminant analysis (PLS-DA) score. (B) Volcano plot of differential proteins. The blue circles on the left indicate down-regulated proteins, while the red circles on the right represent up-regulated proteins. The x-axis represents fold change, and the y-axis represents P value. The two-sided Wilcoxon rank-sum test was employed to identify proteins with an expression fold change of > 2 (either upregulated or down-regulated) and a p-value < 0.05 as differentially expressed proteins. Significantly enriched GO terms (C) and KEGG terms (D). Dot plots and enrichment plots display biological processes along the vertical axis, with circle size indicating gene counts. The depth of colors represents the P value.

## Variable selection and performance of the model

The discrimination performance metrics of the prediction models in derivation and independent validation cohorts are presented in Table 2. In the derivation cohort, the stepwise selection by LR model attained the highest AUC of 0.938 (95% confidence interval (CI): 0.913-0.962). All the ML algorithms showed statistically significant results. In independent validation, the LR model achieved a high AUC of 0.948 (95% CI: 0.916-0.980). Its AUC was significantly higher than the RF model, but comparable to other ML models with AUC in the range of 0.937-0.939. ROC curves of all prediction models are shown in Figure 4.

Given that the LR model demonstrated superior performance compared to other ML models in both derivation

and independent validation population, the final variables included in the prediction model were selected using logistic stepwise regression. These variables encompass sex, age group, smoking status, drinking status, family history of GC, H.pylori, DKK3, and SERPINB5. The association of these predictors with GC are detailed in Table 3. The findings suggest that the positivity of SERPINB5 are independent risk factors for GC, whereas DKK3 positivity is an independent protective factor against GC.

The ROC and calibration curves for the final prediction model in both derivation and validation cohort are shown in Figure 5. The predictive model demonstrates excellent performance, with an AUC ranging from 0.938 to 0.948. Additionally, the sensitivity and specificity have achieved values of 0.843-0.929 and 0.904-0.877, respectively.



#### FIGURE 3

Levels of novel biomarkers in the serum of gastric cancer patients (GC) and non-gastric cancer individuals (non-GC). The expression level of KRT 7, DKK3, and SERPINB5 in serum samples from GC (n=104) and non-GC (n=426) individuals was measured with ELISA. When the P value is less than 0.001, three asterisks (\*\*\*) are used to indicate statistical significance. KRT 7, Keratin 7; DKK 3, Dickkopf-associated protein 3; SERPINB5, Mammary fibrostatin.

## Discussion

To the best of our knowledge, this study represents the first attempt to develop a multidimensional GC risk prediction model that includes macro GC factors, identified GC markers, and novel GC biomarkers simultaneously. The multidimensional model in our study demonstrated strong performance in both derivation and validation cohorts, demonstrating its ability to identify high-risk patients for GC. As a result, this risk prediction model can serve as an initial pre-screening tool for gastroscopy to identify individuals at elevated risk of GC in Hebei Province.

Currently, numerous GC risk prediction models have been developed (6). However, due to variations in study design, statistical methods, and model performance, there is no

TABLE 2 Discrimination performance of gastric cancer risk prediction models derived from four machine learning algorithms.

Algorithm used in model development	Logistic regression	Random forest	eXtreme gradient boosting	Lasso regression	
Factors	Sex, Age group, Smoke, Drink, FGH, H.Pylori, DKK3, SERPINB5	BMI, Education, Smoke, MME production, History of UDTSs, DKK3, SERPINB5	Sex, Age group, Smoke, BMI, Education, H.Pylori, SERPINB5	Sex, Age group, Smoke, Vegetables, Pickled food, H.Pylori, DKK3	
Derivation cohort	Derivation cohort				
AUC (95% CI)	0.938 (0.913-0.962)	0.889 (0.857-0.921)	0.904 (0.876-0.933)	0.898 (0.867-0.930)	
Sensitivity	0.843	0.847	0.801	0.805	
Specificity	0.904	0.798	0.865	0.846	
PPV	0.973	0.945	0.961	0.955	
NPV	0.584	0.561	0.514	0.515	
Kappa	0.619	0.557	0.529	0.534	
Accuracy	0.855	0.838	0.813	0.813	
Independent validation					
AUC (95% CI)	0.948 (0.916-0.980)	0.830 (0.776-0.883)	0.939 (0.904-0.974)	0.937 (0.903-0.971)	
Sensitivity	0.929	0.724	0.945	0.948	
Specificity	0.877	0.827	0.827	0.803	
PPV	0.951	0.916	0.934	0.936	
NPV	0.826	0.536	0.848	0.855	
Kappa	0.790	0.472	0.776	0.765	
Accuracy	0.914	0.753	0.911	0.907	

FGH, Family history of gastric cancer; BMI, body mass index; MME production, Milk, meat and egg production; History of UDTSs, History of upper digestive tract diseases; H.Pylori, Helicobacter pylori; DKK 3, Dickkopf-associated protein 3; SERPINB5, Mammary fibrostatin; AUC, area under the receiver operating characteristic curve; CI, confidence interval; NPV, negative predictive value; PPV, positive predictive value.



Receiver operating characteristic (ROC) curves of the models by four machine algorithms in derivation and validation cohort. (A) Logistic Regression, (B) Random Forest, (C) EXtreme Gradient Boost, (D) Lasso Regression. X-axis: False positive rate (1-specificity); Y-axis: True positive rate (sensitivity). AUC: Area under the curve, indicating model discrimination ability, with higher values suggesting better performance in distinguishing gastric cancer patients.

standardized prediction rule for GC. Additionally, most diagnostic models lack an adequate sample size, select candidate variables through univariate analysis, or fail to undergo a comprehensive model evaluation. Furthermore, the vast majority of predictive models lack internal or external validation (16, 17). Our model is constructed based on a large combined cohort of individuals, adhering to a temporal sequence of cause and effect, with thorough internal and external validation. Various ML algorithms were employed to sift through candidate predictors from all questionnaire data, and the final chosen predictor circumvented the information loss typically associated with the traditional singlefactor to multi-factor screening process.

In this study, the developed model by various ML algorithms demonstrated good performance metrics that included known GC key risk factors in the analysis. These results were both internally and externally validated, indicating that the prediction rule is robust and effective. The macro variables included in the model of this study are sex, age, smoking status, drinking status, and family history of GC. All of these factors have been confirmed to be closely related to the occurrence and development of GC (18). A positive family history of GC in first-degree relatives is a known risk factor for GC (19). Previous studies have also indicated that a patient with a positive family history is at increased risk for developing gastric prelesions (20). Similarly, smoking and drinking have been identified as risk factors for GC according to several meta-analyses (21, 22).

In most studies that only consider general demographic factors in building models, there are also some models that take into account laboratory measures, but most of them only focus on routine tests for H. pylori infection and pepsinogen (23, 24). Protein and gene tests are commonly utilized in basic research of GC, rather than in the development of risk prediction models. Currently, there is a scarcity of studies that have incorporated proteomics into GC risk prediction models. In traditional research,

Characteristics	Univariate		Multivariate	
	OR (95%CI)	P value	OR (95%CI)	P value
Sex		< 0.001		< 0.001
Male	1.00		1.00	
Female	0.36 (0.22-0.57)		0.28 (0.14-0.54)	
Age(years)		< 0.001		0.007
< 60	1.00		1.00	
≥ 60	2.37 (1.51-3.73)		4.08 (2.08-8.02)	
Smoking status		0.013		0.164
Never smoker	1.00		1.00	
Smoker	1.43 (0.86-2.37)		1.40 (0.78-2.52)	
Ex-smoker	2.46 (1.33-4.57)		2.10 (0.94-4.67)	
Drinking status		0.001		0.078
Never-drinker	1.00		1.00	
Drinker	2.03 (1.31-3.14)		1.62 (0.95-2.78)	
Family history of gastric cancer		0.053		0.049
Absent	1.00		1.00	
Present	1.57 (0.99-2.47)		1.23 (1.00-5.46)	
H. pylori		0.001		0.037
Negative	1.00		1.00	
Positive	2.13 (1.38-3.30)		2.04 (1.05-4.00)	
DKK 3		< 0.001		< 0.001
Negative	1.00		1.00	
Positive	0.28 (0.18-0.44)		0.24 (0.13-0.43)	
SERPINB5		< 0.001		< 0.001
Negative	1.00		1.00	
Positive	13.23(7.80-22.46)		14.57 (7.80-27.19)	

TABLE 3 Association between the selected predictors and the risk of gastric cancer.

BMI, body mass index; H.Pylori, Helicobacter pylori; DKK 3, Dickkopf-associated protein 3; SERPINB5, Mammary fibrostatin; OR, odds ratio; CI, confidence interval.

basic research and clinical application often operate separately, with biomarker discovery and model development as distinct stages (25, 26). Our approach differs by integrating biomarker discovery into the model construction process, allowing for concurrent validation of biomarker utility. On one hand, most studies only conduct prognostic analysis, making it challenging to predict disease risk due to difficulties in cohort construction and lack of a control group (27–29). On the other hand, marker discovery in some studies is tissue-based, posing challenges in sample acquisition and complex detection techniques which hinder their application to population screening and risk stratification (30).

In our study, we also took into consideration previously known H. pylori infections. It is widely recognized that H. pylori infection is the most significant risk factor for GC (31, 32). Since the discovery of H. pylori, numerous studies have established a link

between H. pylori and GC as well as its precursors (33–35). Our study found that Helicobacter pylori infection was independently associated with the development of GC.

At the same time, the risk prediction model involving proteomics primarily focuses on typical carcinoembryonic antigens (36). It is crucial to identify additional biomarkers in the development of GC. A study investigated the humoral response of nearly all H. pylori immune proteome (1,527 proteins) in 50 GC cases and 50 control patients, and subsequently developed a GC prediction model. The findings revealed that the model, which incorporates four antibody proteins, achieved an AUC of 0.73 in distinguishing GC from control (37). Another study conducted in South Korea investigated blood-derived protein biomarkers for various types of cancer. Prediction models for six different types of cancer were developed using a panel of 12 blood proteins, with



carcinoembryonic antigen being the main component. The AUC for the prediction model of GC was found to be 0.97 (38). However, it is important to note that this model only utilizes blood markers and does not take into account general demographic factors. While models that use only biomarkers may achieve greater accuracy in a controlled setting, integrated models that combine biomarkers with population variables, such as our study, can provide better generality for population-level implementation.

Our research has identified there novel biomarkers that exhibit significant differences between GC patients and non-GC individuals. However, following machine learning variable selection, only DKK3 and SERPINB5 were incorporated into the GC risk prediction model. This inclusion markedly enhanced the predictive performance of the model, achieving an AUC of 0.938. KRT 7, also known as cytokeratin-7 (CK-7), is the primary component of the intermediate filament cytoskeleton. Studies indicate that KRT7 is associated with cancer cell behaviors such as proliferation, migration, and invasion (39, 40). Particularly, KRT7 has also been confirmed to be significantly up-regulated in GC tissues and cell lines (13). Knockdown of KRT7 impairs GC cell proliferation and migration, and its activation in GC cells is driven by FOXA1 transcription, which enhances these processes (41, 42). However, after adjusting for other protein markers, H.pylori, and macro variables, KRT7 was not included in the final model. In the future, there may be more additional biomarkers to develop novel GC models, and KRT7 warrants further investigation.

Another two biomarkers are also supported by other foundational studies and have a certain biological rationale.

Mammary fibrostatin (Maspin), or SERPINB5, is a member of the serine protease inhibitor superfamily. It is involved in regulating protein disassembly and has been implicated in various cancers, including colorectal and gallbladder cancers (43, 44). SERPINB5 has been confirmed as a novel serum diagnostic biomarker for highgrade intraepithelial neoplasia in GC and is involved in macrophage phenotype regulation (14). DKK 3, encoded by the DKK3 gene, plays critical roles in development, stem cell differentiation, and tissue homeostasis, and has immunomodulatory functions (45). DKK3 is a potential tumor suppressor, with downregulation observed in cancers such as prostate and ovarian cancer (46, 47). Previous studies have shown that reducing DKK3 enhances the migration and invasion of GC cells, which are consistent with our own findings. DKK3 regulates multiple pathways to suppress GC occurrence and progression (15, 48, 49). These biomarkers provide promising approaches for improving the performance of GC risk prediction model and understanding disease mechanisms.

Recent studies have elucidated the potential mechanisms of various molecular markers, including m1A-modified genes, miRNA-CD molecule interactions, and peptides encoded by lncRNAs, in the development, progression, and metastasis of gastrointestinal cancers (50–53). These markers are closely associated with key signaling pathways and influence the invasiveness and metastatic potential of tumors by modulating immune responses and cellular metabolism, showing high potential for predicting the occurrence and progression of gastric cancer. Future research should further explore their specific roles in the tumor microenvironment and integrate clinical samples with multi-omics data to identify additional potential markers and therapeutic targets, thereby advancing the precision diagnosis and treatment of gastrointestinal cancers.

The prediction rule developed in the novel GC model has good discrimination with an AUC of 0.938 in the derivation cohort and high sensitivity (84.3%). However, there are several potential limitations in the present study. Firstly, the serum samples were collected prior to the onset of the disease, and some GC patients were concurrently experiencing other gastric conditions at that time. This may have influenced the levels of serum biomarkers, thereby impacting the applicability of the GC prediction model we developed. However, previous research addressing this issue has indicated that stomach cancer cases accompanied by other gastric disorders are more likely to adhere to predictive guidelines and undergo endoscopy compared to those with isolated GC (54). Additionally, our questionnaire relied on self-reported data, and variables such as dietary habits could not be quantified accurately, which may introduce certain biases into our findings. However, our data collection was conducted by trained investigators following a standardized protocol to minimize potential biases. And the reliability of our data was internally verified through bootstrapping and confirmed in an independent external validation cohort, demonstrating the stability of our model. Moreover, the questionnaires used in this study are wellestablished and have been validated in multiple prior studies (55). Finally, our model is limited by the number of blood specimens and only included participants from Hebei province. However, the model developed in this study remains applicable for predicting the risk of GC in China and other Asian countries to a certain extent. This applicability is supported by the fact that over 90% of China's population is Han ethnic, which shares similar dietary habits and lifestyles with residents from other Asian nations such as Japan, Korea, and Singapore. Additionally, it provides foundational data for future cross-regional comparative studies. Future research should focus on verification and implementation in larger and diverse populations.

In conclusion, the risk prediction model established and validated in this study has shown good identification effectiveness for the high-risk population of GC in Hebei Province. Therefore, it can serve as an accurate and cost-effective initial large-scale prescreening tool to improve the detection rate of GC, reduce unnecessary invasive screening and diagnosis, and thus enhance secondary prevention of GC. In the future, this screening strategy can be extended to validate and test its feasibility in a larger population nationwide.

## Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

## **Ethics statement**

The studies involving humans were approved by the Ethics Committees of the Cancer Hospital, Chinese Academy of Medical Sciences and the Fourth Hospital of Hebei Medical University. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

## Author contributions

YL: Conceptualization, Investigation, Methodology, Software, Writing - original draft, Writing - review & editing, Data curation, Formal analysis, Funding acquisition, Project administration, Resources, Supervision, Validation, Visualization. YF: Data curation, Formal analysis, Methodology, Project administration, Resources, Supervision, Validation, Writing - review & editing. WY: Data curation, Methodology, Supervision, Writing - review & editing, Investigation. ZL: Supervision, Writing - review & editing, Project administration, Software, Validation, Visualization. QL: Project administration, Supervision, Writing - review & editing, Investigation, Methodology, Resources. XS: Methodology, Resources, Supervision, Writing - review & editing, Data curation, Formal analysis, Software. JS: Data curation, Formal analysis, Methodology, Software, Writing - review & editing, Investigation. SW: Methodology, Writing - review & editing, Conceptualization, Project administration, Resources. DL: Methodology, Resources, Writing - review & editing, Formal

Analysis, Software, Supervision, Validation. YH: Formal analysis, Methodology, Resources, Software, Supervision, Validation, Writing – review & editing, Conceptualization, Data curation, Funding acquisition, Investigation, Project administration, Visualization, Writing – original draft.

## Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was supported by the Natural Science Foundation of Hebei Province (grant number H2020206644).

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## References

1. Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, et al. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *Ca-Cancer J Clin.* (2024) 74:229–63. doi: 10.3322/caac.21834

2. Allemani C, Matsuda T, Di Carlo V, Harewood R, Matz M, Nikšić M, et al. Global surveillance of trends in cancer survival 2000-14 (CONCORD-3): analysis of individual records for 37 513 025 patients diagnosed with one of 18 cancers from 322 population-based registries in 71 countries. *Lancet.* (2018) 391:1023–75. doi: 10.1016/S0140-6736(17)3326-3

3. Zeng H, Chen W, Zheng R, Zhang S, Ji JS, Zou X, et al. Changing cancer survival in China during 2003-15: a pooled analysis of 17 population-based cancer registries. *Lancet Glob Health.* (2018) 6:e555–67. doi: 10.1016/S2214-109X(18)30127-X

4. Zhang X, Li M, Chen S, Hu J, Guo Q, Liu R, et al. Endoscopic screening in Asian countries is associated with reduced gastric cancer mortality: A meta-analysis and systematic review. *Gastroenterology*. (2018) 155:347–54.e9. doi: 10.1053/j.gastro.2018.04.026

5. Sun YX, Tang T, Zou JY, Yue QQ, Hu LF, Peng T, et al. Interventions to improve endoscopic screening adherence of cancer in high-risk populations: A scoping review. *Patient Prefer Adher*. (2024) 18:709–20. doi: 10.2147/PPA.S443607

6. He S, Sun D, Li H, Cao M, Yu X, Lei L, et al. Real-world practice of gastric cancer prevention and screening calls for practical prediction models. *Clin Transl Gastroen*. (2023) 14:e00546. doi: 10.14309/ctg.00000000000546

7. Kang C, Lee Y, Lee JE. Recent advances in mass spectrometry-based proteomics of gastric cancer. *World J Gastroentero*. (2016) 22:8283–93. doi: 10.3748/wjg.v22.i37.8283

8. Mohri Y, Toiyama Y, Kusunoki M. Progress and prospects for the discovery of biomarkers for gastric cancer: a focus on proteomics. *Expert Rev Proteomic*. (2016) 13:1131-9. doi: 10.1080/14789450.2016.1249469

9. Li X, Zheng NR, Wang LH, Li ZW, Liu ZC, Fan H, et al. Proteomic profiling identifies signatures associated with progression of precancerous gastric lesions and risk of early gastric cancer. *Ebiomedicine*. (2021) 74:103714. doi: 10.1016/j.ebiom.2021.103714

10. Zeng H, Sun K, Cao M, Zheng R, Sun X, Liu S, et al. Initial results from a multicenter population-based cluster randomized trial of esophageal and gastric cancer screening in China. *BMC Gastroenterol.* (2020) 20:398. doi: 10.1186/s12876-020-01517-3

11. Chen WQ, Li N, Shi JF, Ren JS, Chen HD, Li J, et al. Progress of cancer screening program in urban China [J]. *China Cancer*. (2019) 28(1):23–5. doi: 10.11735/j.issn.1004-0242.2019.01.A003

12. Cao L, Zhou Y, Lin S, Yang C, Guan Z, Li X, et al. The trajectory of vesicular proteomic signatures from HBV-HCC by chitosan-magnetic bead-based separation and DIA-proteomic analysis. *J Extracell Vesicles*. (2024) 13:e12499. doi: 10.1002/jev2.12499

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## Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2025.1536491/ full#supplementary-material

13. Huang B, Song JH, Cheng Y, Abraham JM, Ibrahim S, Sun Z, et al. Long noncoding antisense RNA KRT7-AS is activated in gastric cancers and supports cancer cell progression by increasing KRT7 expression. *Oncogene*. (2016) 35:4927–36. doi: 10.1038/onc.2016.25

14. Huang X, Xie X, Kang N, Qi R, Zhou X, Wang Y, et al. SERPINB5 is a novel serum diagnostic biomarker for gastric high-grade intraepithelial neoplasia and plays a role in regulation of macrophage phenotypes. *Transl Oncol.* (2023) 37:101757. doi: 10.1016/j.tranon.2023.101757

15. Pei Y, Tang Z, Cai M, Yao Q, Xie B, Zhang X. The E2F3/miR-125a/DKK3 regulatory axis promotes the development and progression of gastric cancer. *Cancer Cell Int.* (2019) 19:212. doi: 10.1186/s12935-019-0930-y

16. In H, Langdon-Embry M, Gordon L, Schechter CB, Wylie-Rosett J, Castle PE, et al. Can a gastric cancer risk survey identify high-risk patients for endoscopic screening? *A pilot study J Surg Res.* (2018) 227:246–56. doi: 10.1016/j.jss.2018.02.053

17. In H, Solsky I, Castle PE, Schechter CB, Parides M, Friedmann P, et al. Utilizing cultural and ethnic variables in screening models to identify individuals at high risk for gastric cancer: A pilot study. *Cancer Prev Res.* (2020) 13:687–98. doi: 10.1158/1940-6207.CAPR-19-0490

18. Yaghoobi M, McNabb-Baltar J, Bijarchi R, Hunt RH. What is the quantitative risk of gastric cancer in the first-degree relatives of patients? A meta-analysis. *World J Gastroentero*. (2017) 23:2435–42. doi: 10.3748/wjg.v23.i13.2435

19. Bernini M, Barbi S, Roviello F, Scarpa A, Moore P, Pedrazzani C, et al. Family history of gastric cancer: a correlation between epidemiologic findings and clinical data. *Gastric Cancer*. (2006) 9:9–13. doi: 10.1007/s10120-005-0350-7

20. Song M, Camargo MC, Weinstein SJ, Best AF, Mannisto S, Albanes D, et al. Family history of cancer in first-degree relatives and risk of gastric cancer and its precursors in a Western population. *Gastric Cancer*. (2018) 21:729–37. doi: 10.1007/ s10120-018-0807-0

21. Ferro A, Morais S, Rota M, Pelucchi C, Bertuccio P, Bonzi R, et al. Tobacco smoking and gastric cancer: meta-analyses of published data versus pooled analyses of individual participant data (StoP Project). *Eur J Cancer Prev.* (2018) 27:197–204. doi: 10.1097/CEJ.00000000000401

22. Wang PL, Xiao FT, Gong BC, Liu FN. Alcohol drinking and gastric cancer risk: a meta-analysis of observational studies. *Oncotarget*. (2017) 8:99013–23. doi: 10.18632/ oncotarget.20918

23. Eom BW, Joo J, Kim S, Shin A, Yang HR, Park J, et al. Prediction model for gastric cancer incidence in Korean population. *PLoS One.* (2015) 10:e0132613. doi: 10.1371/journal.pone.0132613

24. Afrash MR, Shafiee M, Kazemi-Arpanahi H. Establishing machine learning models to predict the early risk of gastric cancer based on lifestyle factors. *BMC Gastroenterol.* (2023) 23:6. doi: 10.1186/s12876-022-02626-x

25. Huang Y, Shao Y, Yu X, Chen C, Guo J, Ye G. Global progress and future prospects of early gastric cancer screening. *J Cancer.* (2024) 15:3045–64. doi: 10.7150/ jca.95311

26. Khalili-Tanha G, Khalili-Tanha N, Rouzbahani AK, Mahdieh R, Jasemi K, Ghaderi R, et al. Diagnostic, prognostic, and predictive biomarkers in gastric cancer: from conventional to novel biomarkers. *Transl Res.* (2024) 274:35–48. doi: 10.1016/ j.trsl.2024.09.001

27. Zhao J, Liu Y, Cui Q, He R, Zhao JR, Lu L, et al. A prediction model for prognosis of gastric adenocarcinoma based on six metabolism-related genes. *Biochem Biophys Rep.* (2023) 34:101440. doi: 10.1016/j.bbrep.2023.101440

28. Zhang W, Zhou D, Song S, Hong X, Xu Y, Wu Y, et al. Prediction and verification of the prognostic biomarker SLC2A2 and its association with immune infiltration in gastric cancer. *Oncol Lett.* (2024) 27:70. doi: 10.3892/ol.2023.14203

29. Ahn HS, Sohn TS, Kim MJ, Cho BK, Kim SM, Kim ST, et al. SEPROGADIC - serum protein-based gastric cancer prediction model for prognosis and selection of proper adjuvant therapy. *Sci Rep-Uk*. (2018) 8:16892. doi: 10.1038/s41598-018-34858-x

30. Kan Y, Lu X, Feng L, Yang X, Ma H, Gong J, et al. RPP30 is a novel diagnostic and prognostic biomarker for gastric cancer. *Front Genet.* (2022) 13:888051. doi: 10.3389/fgene.2022.888051

31. Nie Y, Wu K, Yu J, Liang Q, Cai X, Shang Y, et al. A global burden of gastric cancer: the major impact of China. *Expert Rev Gastroent*. (2017) 11:651–61. doi: 10.1080/17474124.2017.1312342

32. Leung WK, Wu MS, Kakugawa Y, Kim JJ, Yeoh KG, Goh KL, et al. Screening for gastric cancer in Asia: current evidence and practice. *Lancet Oncol.* (2008) 9:279–87. doi: 10.1016/S1470-2045(08)70072-X

33. Gonzalez CA, Agudo A. Carcinogenesis, prevention and early detection of gastric cancer: where we are and where we should go. *Int J Cancer*. (2012) 130:745–53. doi: 10.1002/ijc.v130.4

34. Karimi P, Islami F, Anandasabapathy S, Freedman ND, Kamangar F. Gastric cancer: descriptive epidemiology, risk factors, screening, and prevention. *Cancer Epidem Biomar.* (2014) 23:700–13. doi: 10.1158/1055-9965.EPI-13-1057

35. Hu Y, Zhu Y, Lu NH. The management of Helicobacter pylori infection and prevention and control of gastric cancer in China. *Front Cell Infect Mi*. (2022) 12:1049279. doi: 10.3389/fcimb.2022.1049279

36. Xie Y, Zhi X, Su H, Wang K, Yan Z, He N, et al. A novel electrochemical microfluidic chip combined with multiple biomarkers for early diagnosis of gastric cancer. *Nanoscale Res Lett.* (2015) 10:477. doi: 10.1186/s11671-015-1153-3

37. Song L, Song M, Rabkin CS, Williams S, Chung Y, Van Duine J, et al. Helicobacter pylori immunoproteomic profiles in gastric cancer. *J Proteome Res.* (2021) 20:409–19. doi: 10.1021/acs.jproteome.0c00466

38. Kim YS, Kang KN, Shin YS, Lee JE, Jang JY, Kim CW. Diagnostic value of combining tumor and inflammatory biomarkers in detecting common cancers in Korea. *Clin Chim Acta*. (2021) 516:169–78. doi: 10.1016/j.cca.2021.02.002

39. Song J, Wu Y, Chen Z, Zhai D, Zhang C, Chen S. Clinical significance of KRT7 in bladder cancer prognosis. *Int J Biol Marker*. (2024) 39:158–67. doi: 10.1177/03936155231224798

40. Song J, Ruze R, Chen Y, Xu R, Yin X, Wang C, et al. Construction of a novel model based on cell-in-cell-related genes and validation of KRT7 as a biomarker for predicting survival and immune microenvironment in pancreatic cancer. *BMC Cancer*. (2022) 22:894. doi: 10.1186/s12885-022-09983-6

41. Liu BL, Qin JJ, Shen WQ, Liu C, Yang XY, Zhang XN, et al. FOXA1 promotes proliferation, migration and invasion by transcriptional activating KRT7 in human gastric cancer cells. *J Biol Reg Homeos Ag.* (2019) 33:1041–50.

42. Liu Z, Sun L, Peng X, Zhu J, Wu C, Zhu W, et al. PANoptosis subtypes predict prognosis and immune efficacy in gastric cancer. *Apoptosis*. (2024) 29:799–815. doi: 10.1007/s10495-023-01931-4

43. Sinha KK, Vinay J, Parida S, Singh SP, Dixit M. Association and functional significance of genetic variants present in regulatory elements of SERPINB5 gene in gallbladder cancer. *Gene.* (2022) 808:145989. doi: 10.1016/j.gene.2021.145989

44. Liu BX, Xie Y, Zhang J, Zeng S, Li J, Tao Q, et al. SERPINB5 promotes colorectal cancer invasion and migration by promoting EMT and angiogenesis via the TNF-alpha/NF-kappaB pathway. *Int Immunopharmacol.* (2024) 131:111759. doi: 10.1016/j.intimp.2024.111759

45. Mourtada J, Thibaudeau C, Wasylyk B, Jung AC. The multifaceted role of human dickkopf-3 (DKK-3) in development, immune modulation and cancer. *Cells.* (2023) 13(1):75. doi: 10.3390/cells13010075

46. Nguyen Q, Park HS, Lee TJ, Choi KM, Park JY, Kim D, et al. DKK3, Downregulated in Invasive Epithelial Ovarian Cancer, Is Associated with Chemoresistance and Enhanced Paclitaxel Susceptibility via Inhibition of the beta-Catenin-P-Glycoprotein Signaling Pathway. *Cancers*. (2022) 14(4):924. doi: 10.3390/cancers14040924

47. Shareef ZA, Hachim MY, Talaat IM, Bhamidimarri PM, Ershaid M, Ilce BY, et al. DKK3's protective role in prostate cancer is partly due to the modulation of immune-related pathways. *Front Immunol.* (2023) 14:978236. doi: 10.3389/fimmu.2023.978236

48. Xia P, Xu XY. DKK3 attenuates the cytotoxic effect of natural killer cells on CD133(+) gastric cancer cells. *Mol Carcinogen*. (2017) 56:1712–21. doi: 10.1002/mc.22628

49. Pei Y, Tang Z, Cai M, Yao Q, Xie B. MicroRNA miR-425 promotes tumor progression by inhibiting Dickkopf-related protein-3 in gastric cancer. *Bioengineered*. (2021) 12:2045–54. doi: 10.1080/21655979.2021.1930743

50. Chen Y, Long W, Yang L, Zhao Y, Wu X, Li M, et al. Functional peptides encoded by long non-coding RNAs in gastrointestinal cancer. *Front Oncol.* (2021) 11:777374. doi: 10.3389/fonc.2021.777374

51. Zhang H, Li M, Kaboli PJ, Ji H, Du F, Wu X, et al. Identification of cluster of differentiation molecule-associated microRNAs as potential therapeutic targets for gastrointestinal cancer immunotherapy. *Int J Biol Markers*. (2021) 36:22–32. doi: 10.1177/17246008211005473

52. Xu Z, Xia Y, Xiao Z, Jia Y, Li L, Jin Y, et al. Comprehensive profiling of JMJD3 in gastric cancer and its influence on patient survival. *Sci Rep.* (2019) 9:868. doi: 10.1038/s41598-018-37340-w

53. Zhao Y, Zhao Q, Kaboli PJ, Shen J, Li M, Wu X, et al. m1A regulated genes modulate PI3K/AKT/mTOR and ErbB pathways in gastrointestinal cancer. *Trans Oncol.* (2019) 12:1323–33. doi: 10.1016/j.tranon.2019.06.007

54. Cai Q, Zhu C, Yuan Y, Feng Q, Feng Y, Hao Y, et al. Development and validation of a prediction rule for estimating gastric cancer risk in the Chinese high-risk population: a nationwide multicentre study. *Gut.* (2019) 68:1576–87. doi: 10.1136/gutjnl-2018-317556

55. Lu Q, Liang D, Shi J, Wu S, Du X, He Y. Improved effectiveness of combined screening for multiple cancers: A government-organized population-based study in China. *Cancer Med.* (2024) 13:e70463. doi: 10.1002/cam4.v13.23