



Pediatric Liver Disease Patients and Secondary Glycosylation Abnormalities

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Background: Isoelectric focusing (IEF) of serum transferrin (Tf) is still the method of choice for diagnosis of congenital disorders of glycosylation (CDG). An abnormal glycosylation is also a known phenomenon in adult liver disease patients. The aim of this study was to characterize glycosylation disturbances in pediatric patients with primary liver disease. However, there are no reports of this phenomenon in children.

Materials and Methods: Between 1995 and 2019, circa 2,000 serum Tf isoform analyses have been performed in children with primary liver diseases; some of them underwent subsequent analyses. We enrolled in this study 19 patients who developed an acute liver injury (ALI)/failure (ALF) or exhibited a chronic liver disease (CLD) and were evaluated and listed for liver transplantation (LTx) or had just undergone this procedure, and secondary abnormal serum Tf isoform profile.

Results: Among 12 patients with ALI/ALF, 10 had an increased percentage of asialo-, monosialo-, and disialo-Tf isoforms. All patients with CLD had an increased percentage of asialo- and monosialo-Tf isoform. Two patients diagnosed with recurrent ALF had very specific serum Tf profile with a huge increase in the asialo- and monosialo-Tf isoform. On follow-up analyses (available in some patients), serum Tf IEF profile normalized in parallel to normalization of liver function tests, spontaneously or during treatment, including glucocorticosteroids in AIH, LTx in CLD.

Conclusions: All pediatric patients with primary liver disease had increased asialo-Tf as well as monosialo-Tf isoforms. None of them had elevated percentage of trisialo-Tf isoform.

Keywords: congenital disorder of glycosylation (CDG), acute liver failure (ALF), acute liver injury (ALI), chronic liver disease (CLD), liver transplantation, serum transferrin isoelectrofocusing, autoimmune hepatitis (AIH)

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INTRODUCTION

Liver is the main site of synthesis and physiological activity of transferrin (Tf) and other serum glycoproteins (1). Isoelectric focusing (IEF) of serum Tf is still the method of choice for diagnosis of congenital disorders of glycosylation (CDG), especially N-glycosylation disorders associated with sialic acid deficiency (2, 3). Transferrin is the iron-binding serum glycoprotein which is produced in hepatocytes and contains two N-linked glycosylation sites (4, 5). An abnormal glycosylation is also a known phenomenon in liver disease patients; it has been described in alcoholic liver disease (decreased enzyme activities of mannosyltransferase and galactosyltransferase, lowered intracellular dolichol concentration, desialylation of serum Tf, *α*1-antitrypsin and ceruloplasmin; hyperfucosylation of haptoglobin and serum Tf), galactosemia and fructosemia (the pattern resembling CDG type I), chronic hepatitis B and C, non-alcoholic steatohepatitis, and bile-related liver diseases (6-21). Despite of IEF, there are also some other methods, like HPLC or MALDI-TOF but they are not available in Poland. Specifically, carbohydrate deficient transferrin (CDT) analysed using isoelectric focusing (IEF) as a sum of asialo-, monosialo-, and disialo-Tf isoforms has been widely used in recent years as a marker of chronic alcohol abuse in adults. However, the literature about serum Tf isoform profiles in children with liver disease is sparse. The aim of this study was to characterize glycosylation disturbances in pediatric patients with primary liver disease.

MATERIALS AND METHODS

Between 1995 and 2019 years, about 2,000 serum Tf isoform analyses have been performed in children with primary liver diseases hospitalized in our Institute. Some of the patients underwent subsequent analyses during follow-up. Serum Tf isoforms were analyzed by isoelectrofocusing (IEF) agarose gel electrophoresis according to the method described by Van Eijk et al. (5).

Serum Tf consists of the mixture of isoforms and has two iron binding sites, which was saturated with iron $(20 \,\mu$ l serum with $80 \,\mu$ l 0,9% NaCl, $2 \,\mu$ l 10 mM Fe(III) citrate and $2 \,\mu$ l 0.1 M NaHCO₃). Transferrin migrated through a 1% agarose gel (45 mg of agarose were added to 4.5 ml of distilled water) with 5% ampholines in a pH range of 5.0-7.0 on a Multiphore 2117 apparatus (LKB) with a modified electrode lid. Tf isoforms were visualized by immunofixation ($200 \,\mu$ l polyclonal rabbit antihuman transferrin serum) and 0.5% Coomasie Brillant Blue solution staining, as presented in **Figure 1**. The percentage of Tf fractions was assessed densitometrically and carbohydrate deficient transferrin value (% CDT) was measured as a sum of asialo-, monosialo-, and disialo-Tf isoforms.

We enrolled in this study 19 patients with primary liver disease, who developed an acute liver injury/failure or exhibited a chronic liver disease, and secondary abnormal serum Tf isoform profile. They are the totality of patients who showed glycosylation abnormalities among patients with primary liver disease. Patients presenting an abnormal serum Tf isoform profile secondary to galactosemia (galactose-1-phosphate uridyltransferase deficiency) or fructosemia (fructose 1-phosphate aldolase deficiency) were excluded from the study.

Acute liver injury (ALI) was characterized by an acute hepatocellular damage or death, expressed by elevation of serum transaminases, leading to rapid loss of hepatocellular function, excretory (cholestasis defined as serum conjugated bilirubin > 1,0 mg/dL according to the latest ESPGHAN recommendations) and synthetic (coagulopathy defined as INR > 1.2), in a patient without chronic liver disease (22–24). Pediatric acute liver failure (ALF) was diagnosed based on the above-mentioned criteria and one of the following: hepatic-based coagulopathy defined as INR \geq 1.5 not corrected by vitamin K in the presence of clinical hepatic encephalopathy, or INR \geq 2.0 regardless of the presence or absence of clinical hepatic encephalopathy (22, 23). The term 'recurrent ALF' (RALF) was used to describe patients with repeated episodes of acute liver injury with recovery of hepatic function between crises (25, 26).

All the patients with ALI/ALF were evaluated according to the current clinical practical guidelines (23). In some of them, liver biopsy was essential to make a diagnosis, i.e., autoimmune hepatitis (AIH). There were also patients in whom an undetermined etiology of ALI/ALF was established.

Chronic liver disease (CLD) refers to liver disease that lasted over a period of at least 6 months. In this group were enrolled patients evaluated and listed for liver transplantation (LTx) or who had just undergone this procedure (27).

The retrospective chart review of patients' medical long-term records concerning the demographics, clinical characteristics, and biochemical data, including serum Tf IEF profile, aspartate (AST) and alanine (ALT) aminotransferases, international normalized ratio (INR), total and direct serum bilirubin, gammaglutamyl transpeptidase (GGT) were collected. Ethical approval was obtained from the Children's Memorial Health Institute Bioethical Committee, num 23/KBE/2020, Warsaw, Poland.

Statistical analysis was conducted in R software and consisted of: first, calculating the mean of the observed levels of Tf isoforms and liver tests results in each patients (some of them had only one measurement, but some had even six measurements taken). Next, simple t-tests or Welsch t-tests (if the variances differed between patients groups) were performed in order to check if the three groups are different in terms of the measured level of Tf isoforms or liver test results. Since for every parameter we compare group I with group II, group II with group III, and group I with group III, we account for multiple comparisons by pvalue adjustments using Bonferroni-Holm method (28). Finally, within every group, we check if there is a correlation between Tf isoforms % and liver tests results by calculating Pearson's linear correlation coefficient r. We also check if the obtained coefficients are statistically significant, accounting for multiple comparisons using Benjamini-Hochberg procedure (29).

RESULTS

Clinical and Biochemical Results

Nineteen pediatric patients (13 males and 6 females) with primary liver disease presented secondary N-hypoglycosylation of serum Tf. The mean age at diagnosis was 3 years and 1



month. Based on the phenotype of liver disease, patients were divided into 3 groups, as following: group I—acute presentation, including ALI/ALF; group II—chronic presentation, including CLD; group III—recurrent ALF (RALF).

Detailed characteristics of clinical and biochemical features are summarized in Table 1.

Out of 19 patients, 14 presented ALI/ALF, including 4 with AIH, 1 with citrin deficiency, 1 with deoxyguanosine kinase (DGOUK) deficiency, 6 with an indeterminate etiology, and 2 with recurrent ALF (RALF). The mean age at diagnosis was 4.5 years.

In the group of 5 patients with CLD, 1 had portal hypertension due to Abernethy malformation and was listed for LTx; 1 had biliary atresia and was listed for LTx; 2 had chronic cholestatic liver disease of an unknown etiology and had undergone LTx; one was diagnosed with liver cirrhosis of unknown etiology and had undergone LTx. The mean age at diagnosis was 8 months.

We present the classification of patients with these clinical details in **Figure 1**.

Among 12 patients from group I, 10 had an increased percentage of asialo-, monosialo-, and disialo-Tf isofroms. Follow-up analyses were available only in four patients (including two with AIH); serum Tf IEF profile normalized in parallel to the normalization of liver function tests (in patients with AIH on treatment with glucocorticosteroids).

Among five patients from group II, all had an increased percentage of asialo- and monosialo-Tf isoforms while two of them had also increased disialo-Tf isoform. Follow-up analyses were available only in two patients, after LTx; serum Tf IEF profile normalized in parallel to the normalization of liver function tests.

Group III included two patients diagnosed with RALF and an increased percentage of asialo- (huge increase) and monosialo-Tf isoforms, and in one of them also disialo-Tf isoform on some of the results. Between episodes of ALF, serum Tf isoform profile was normal as well as liver function tests.

Regarding particular serum Tf isoforms, all patients had an increase in asialo-Tf as well as monosialo-Tf isoform. An elevated percentage of disialo-Tf isoform was observed in 12 out of 14 patients with an acute hepatic injury, including 10 out of 12 from group I and 2 from group III (transient elevation). None of the patients had elevated percentage of trisialo-Tf isoform.

We include averages of % Tf isoforms, as well as liver test results for each group, along with reference ranges (for Tf isoforms) in **Table 2**.

| Number/Sex | Age | Asjalo- | Monosjalo- <i>N</i> : 0 | Disjalo- | Trisjalo- <i>N</i> : | Tetrasjalo- <i>N</i> : 55,7–67,2 | Pentasjalo- N: 13,2–19,9 | Heksasjalo- <i>N</i> : 2,5–5,6 | AST | ALT | INR | Total/direct serum | GGT | Clinical outcome |
|------------|----------|--------------|----------------------------|----------------|-------------------------|-------------------------------------|-----------------------------|-----------------------------------|-------|-------|------|-----------------------|------|------------------------------------|
| | | <i>N</i> : 0 | | N: 1,5– 6,2 | 7,4–17,1 | N. 33,7-07,2 | 1110,2 10,0 | N. 2,3-3,0 | | | | bilirubin | | |
| Group I | | | | | | | | | | | | | | |
| 1/F | 11 m | 1,4 | 2,5 | 11,1 | 13,2 | 46,9 | 18,6 | 6,3 | 1,615 | 349 | 0,92 | 8,9/7,0 | 820 | ALI of undeterminate etiology |
| | 12 m | 4,9 | 4,1 | 6,3 | 13,7 | 45,8 | 19,1 | 6,0 | 1,081 | 425 | 0,92 | 5,0/4,6 | 675 | |
| | 14 m | 0,5 | 2,2 | 6,6 | 12,9 | 49,4 | 20,9 | 7,5 | 333 | 252 | 0,85 | 2,85/2,57 | 481 | |
| 2/M | 1 m | 0,8 | 1,9 | 3,3 | 8,7 | 61,6 | 19,0 | 4,5 | 51 | 32 | 3,3 | 7,96/1,82 | 249 | ALF due to DGUOK deficiency |
| | 3 m | 0,8 | 1,4 | 5,2 | 11,7 | 58,3 | 18,1 | 4,4 | 149 | 53 | 1,22 | 5,2/4,1 | 390 | |
| | 4 y | 0 | 0 | 5,6 | 14,2 | 58,4 | 19,0 | 2.8 | 47 | 30 | 0,96 | Normal | 12 | |
| 3/M | 7 y 4 m | 0,4 | 1,2 | 7,4 | 12,4 | 50,3 | 21,6 | 6,7 | 2,277 | 1,247 | 1,45 | 21,5/19,9 | 107 | ALI due to autoimmune hepatitis |
| | 7 y 5 m | 0,2 | 0,9 | 7,2 | 12,5 | 46,8 | 24,8 | 7,5 | 510 | 721 | 1,2 | 19,9/16,6 | 211 | |
| | 7 y 8 m | 0 | 0 | 5,2 | 12,1 | 56,8 | 21,7 | 4,2 | 30 | 39 | 1,02 | Normal | 59 | |
| 4/M | 2 y 7 m | 0,9 | 1,6 | 6,4 | 11,2 | 50,1 | 23 | 6,8 | 1,475 | 1,173 | 1,32 | 8,4/7,3 | 198 | ALI of undeterminate etiology |
| | 2 y 8 m | 1,0 | 2,0 | 7,3 | 12,5 | 46,2 | 21,4 | 6,9 | 185 | 155 | 1,05 | 15,1/12,3 | 97 | |
| 5/F | 13 y 4 m | 0,9 | 1,2 | 6,7 | 9,6 | 53 | 22,8 | 5,8 | 855 | 587 | 2,27 | 5,9/4,1 | 48 | ALF due to autoimmune hepatitis |
| 6/F | 11 y 4 m | 2,3 | 4,2 | 6,0 | 9,9 | 48,4 | 22,1 | 7,1 | 90 | 34 | 2,3 | 3,8/2,2 | n.a. | ALF of undeterminate etiology |
| | 11 y 7 m | 1,5 | 3,2 | 7,3 | 12,6 | 52,8 | 17,8 | 4,8 | 71 | 79 | 1,98 | 7,1/5,3 | 60 | |
| 7/M | 5 y 8 m | 0,6 | 1,6 | 6,3 | 12,7 | 49,4 | 21,5 | 7,9 | 2,716 | 1,678 | 1,27 | 21,1/16,8 | 68 | ALI due to autoimmune hepatitis |
| | 6 y | 0,6 | 1,1 | 7,2 | 13,6 | 46,3 | 23,1 | 8,0 | 2,094 | 1,494 | 1,23 | 21/20,2 | 75 | |
| 8/M | 2 y 6 m | 0,8 | 1,9 | 8,5 | 15,8 | 45,1 | 23,8 | 4,0 | 1,419 | 567 | 2,36 | 15,3/13,7 | 41 | ALF of undeterminate etiology |
| 9/M | 11 m | 0,9 | 1,1 | 6,2 | 17,6 | 46,6 | 22,4 | 5,1 | 2,040 | 1,470 | 1,49 | 26,1/19,5 | n.a. | ALI of undeterminate etiology |
| 10/F | 15 m | 2,4 | 1,9 | 5,8 | 7,8 | 54 | 22 | 5,1 | 1,082 | 264 | 1,69 | 6,2/5,2 | 90 | ALI of undeterminate etiology |
| | 16 m | Ν | Ν | Ν | Ν | Ν | Ν | Ν | 24 | 20 | 1,05 | Normal | 27 | |
| 11/M | 6 m | 3,8 | 5,7 | 8,0 | 9,7 | 48 | 17,3 | 7,6 | 101 | 30 | 2,01 | 9,6/4,5 | 145 | ALF due to citrin deficiency |
| 12/F | 7 y 6 m | 0 | 0 | 6.5 | 11.2 | 47.9 | 27.8 | 6.6 | 2,213 | 1,784 | 1,46 | 15,9/13,3 | 122 | ALI due to autoimmun hepatitis |
| | 7 y 9 m | 1,0 | 2,6 | 9,9 | 17,3 | 50,9 | 14,9 | 3,5 | 109 | 132 | 1,07 | 0.94/0,55 | 130 | |
| | 7 y 10 m | 0 | 0 | 6,2 | 12,5 | 56,3 | 12,5 | 6,2 | 40 | 25 | 1,08 | 0,64/0,26 | 30 | |

(Continued)

Glycosylation Disturbances in Liver Disease

| Number/Sex | Age | Asjalo- <i>N</i> : 0 | Monosjalo- <i>N</i> : 0 | Disjalo- <i>N</i> : 1,5– 6,2 | Trisjalo- <i>N</i> : 7,4–17,1 | Tetrasjalo- <i>N</i> : 55,7–67,2 | Pentasjalo- <i>N</i> : 13,2–19,9 | Heksasjalo- <i>N</i> : 2,5–5,6 | AST | ALT | INR | Total/direct serum bilirubin | GGT | Clinical outcome |
|------------|---------------------|-------------------------|----------------------------|------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|-----------------------------------|--------------|------------|--------------|------------------------------------|-----------|---|
| Group II | | | | | | | | | | | | | | |
| 1/M | 5 m | 1,3 | 1,3 | 5,8 | 6,6 | 59,7 | 21,2 | 4,0 | 96 | 87 | 1,28 | Normal | 155 | Liver cirrhosis of unknown etiology, LTx |
| | 7 m (after LTx) | Ν | Ν | Ν | Ν | Ν | Ν | Ν | 161 | 204 | 1,12 | Normal | 183 | |
| | 14 m (after LTx) | Ν | Ν | Ν | Ν | Ν | Ν | Ν | 723 | 665 | 1,04 | Normal | 98 | |
| 2/M | 10 m | 0,8 | 1,5 | 2,9 | 9,6 | 58,5 | 21,5 | 5,2 | 157 | 95 | 1,33 | 16,6/12,8 | 41 | Biliary atresia, listed for LTx |
| | 11 m | 1,0 | 1,3 | 5,2 | 11,7 | 57,3 | 19,7 | 3,7 | 174 | 88 | 1,21 | 17,5/14,1 | 72 | |
| 3/F | 11 m | 1,6 | 2,4 | 6,5 | 10,3 | 45,6 | 25,9 | 7,8 | 418 | 229 | 1,73 | 18,8/15,5 | 85 | Chronic cholestatic live disease of unknown etiology, LTx |
| 4/M | 7 m | 1,8 | 1,3 | 5,1 | 10,9 | 48,4 | 24,3 | 8,2 | 329 | 174 | 1,43 | 15,7/13,9 | 185 | Chronic cholestatic live disease of unknown etiology, LTx |
| | 7 y (after LTx) | 0 | 0 | 4,9 | 13,4 | 59,9 | 19,1 | 2,8 | 36 | 42 | 0,98 | Normal | 28 | |
| 5/M | 9 m | 3,1 | 2,7 | 7,5 | 10,9 | 48,3 | 21,2 | 6,3 | 89 | 41 | 1,36 | Normal | 11 | CLD, portal hypertension, |
| | 14 m | 1,1 | 1,4 | 4,9 | 11,5 | 60,8 | 18,3 | 2,0 | 91 | 48 | 1,47 | Normal | n.a. | Abernethy |
| | 16 m | 0,2 | 0,3 | 3,8 | 11,9 | 59,2 | 22,1 | 2,6 | 103 | 38 | 1,38 | Normal | n.a. | malformation, listed for LTx |
| Group III | | | | | | | | | | | | | | LIX |
| 1/M | 9 m | 3,0 | 1,4 | 7,3 | 12,5 | 48,1 | 22,4 | 5,3 | 1,806 | 550 | 1,43 | 8,1/5,7 | 98 | RALF (4 episodes) |
| | 1 y 10 m | 18,3 | 5,1 | 6,5 | 11,6 | 41,6 | 13,5 | 3,3 | 1,520 | 680 | 1,53 | 8,3/6,1 | 41 | |
| | 4 y 3 m | 13,0 | 4,3 | 5,5 | 14,6 | 49,7 | 10,0 | 2,5 | 2,280 | 860 | 2,1 | 9,3/7,1 | 68 | |
| | 5 y 3 m | 0 | 0 | 5,2 | 11,5 | 57 | 21,3 | 5,0 | 26 | 26 | 0.9 | Normal | 30 | |
| | 7 y 3 m | 14,2 | 2,2 | 3,3 | 5,9 | 47,5 | 19,9 | 7,1 | 1,240 | 450 | 2,1 | 3,9/3,5 | 90 | |
| | 8 y | 0 | 0 | 2,7 | 5,9 | 67,5 | 19,5 | 4,2 | 20 | 22 | 0.9 | Normal | 15 | |
| 2/M | 5m | 6,9 | 2,8 | 5,9 | 10,9 | 42,9 | 22,6 | 8,7 | 1,350 | 580 | 1,67 | 8,2/6,9 | 60 | RALF (4 episodes) |
| | 3 y 2 m | 20,9 | 2,4 | 3,6 | 7,1 | 47,4 | 15,6 | 2,9 | 1,270 | 385 | 1,9 | 10,5/9,4 | 65 | |
| | 4 y 10 m | 0 | 0 | 2,4 | 9,2 | 65,3 | 19,2 | 3,5 | 28 | 26 | 0.95 | Normal | 30 | |
| | 5 y | 20,6 | 4,6 | 5,3 | 9,0 | 44,9 | 12,9 | 2,6 | 1,650 | 580 | 1,5 | 8,6/8,2 | 50 | |
| | 5 y 2 m | 0 | 0 | 3,2 | 9,8 | 51,8 | 28,9 | 6,4 | 30 | 20 | 0,9 | Normal | 26 | |
| | 9 y | 17,9 | 5,7 | 5,2 | 8,9 | 46,9 | 12,5 | 3,2 | 1,200 | 420 | 1,4 | 8,0/7,2 | 45 | |
| | 4 y 7 m 4 y 9 m | 0 0 | 0 | 6,3 6,3 | 14,8 14,2 | 48,1 48,6 | 25,9 25,5 | 7,8 5,4 | 315 1,128 | 143 496 | 1,03 1,05 | 0,81/0,48 0,92/0,57 | 73 489 | |
| | 4 y 9 m | 0 | 0 | 6,3 | 14,2 | 48,6 | 25,5 | 5,4 | 1,128 | 496 | 1,05 | 0,92/0,57 | 489 | |

| (A) | Asiao- | Monosialo- | Disialo- | Trisialo- | Tetrasialo- | Pentasialo- | Heksasial | D- | | |
|-----------|--------|------------|----------|-----------|-------------|-------------|-----------|------|-----------------|------|
| Group I | 1.2 | 1.92 | 6.72 | 12.2 | 50.6 | 21.1 | 5.84 | | | |
| Group II | 1.23 | 1.44 | 5.35 | 10.2 | 54.7 | 22 | 5.08 | | | |
| Group III | 9.67 | 2.38 | 4.68 | 9.74 | 50.9 | 18.2 | 4.56 | | | |
| (B) | | | AST | | AL | т | | INR | Total bilirubin | GG |
| Group I | | | 952 | | 6,1 | 56 | | 1.66 | 11 | 11- |
| Group II | | | 237 | | 15 | 8 | | 1.35 | 13.1 | 80.9 |
| Group III | | 1 | 1,035 | | 38 | 3 | | 1.44 | 8.11 | 51.5 |
| 1- | | | | | | | | | | |



Data Visualization and Statistical Analysis

Serum Tf isoform results for the whole cohort of patients are depicted in **Figure 2**. Different colors are used to distinguish between the three different groups of patients. Reference ranges are visualized with gray rectangles. Some of the patients had more than one measurement performed, therefore in some places there are several values associated to one argument. Although the picture might give an idea about how Tf isoforms vary between the groups, we performed simple t-tests or Welsch t-test to check if the differences in means are statistically significant. We found that:



- Group I is different from group II in disialo-Tf (mean in group I = 6.72%, mean in group II = 5.35%; *p*-value = $0.02 \approx \alpha/3$),
- Group II differs from group III in asialo-Tf (mean in group II = 1.23%, mean in group III = 9.57%; *p*-value = 0.00016 < α/3), and pentasialo-TF (mean in group II = 22%, mean in group III = 18.2%; *p*-value = 0.01789 < α/2),
- Group I is different from group III in pentasialo-Tf (mean in group II = 21.1%, mean in group III = 18.2%; *p*-value = 0.00016< α/3)

Similar analysis was performed for liver function tests results. The visualization of data is presented in **Figure 3**. Since the reference ranges are different for different age ranges, the results falling into norms are depicted with different markers than those falling outside. *T*-test results revealed that:

• Group I is significantly different from group III in GGT results (111.2 and 51.5, *p*-value = $0.011 < \alpha/3$).

We also investigated whether there are correlations between Tfisoform profiles and liver function test results within the groups. Interestingly, we found no correlations in group I at all. In group II, we found several correlations (**Supplementary Table 1**), but only two of them were statistically significant: between INR results and monosialo-Tf, and INR and tetrasialo-Tf. In group III, since there are only two patients, and we averaged the results, we obtained only two data points for every pair Tfisoform %—liver parameter, which is insufficient for sensibly investigating correlations.

Finally, we visualized time evolution of serum Tf-isoform % and liver function test results in patients form group III, since they had several measurements taken over time. These interesting visualizations are given in **Figures 4**, **5**.

DISCUSSION

To the best of our knowledge, this is the first study aiming to characterize pediatric liver disease patients with secondary serum Tf N-hypoglycosylation.

Fourteen out of 19 patients in the study had an acute presentation of liver disease in the form of ALI/ALF or recurrent ALF. Secondary N-hypoglycosylation of serum Tf as a mild increase in the asialo-, monosialo-, and disialo-Tf isoforms was diagnosed. However, we observed no correlation between serum Tf isoforms and liver function tests globally, i.e., in the whole cohort of patients. On the other hand, we found correlations of serum Tf isoforms and liver function tests in group II. On follow-up analyses, serum Tf IEF profile normalized in parallel to normalization of liver function tests, spontaneously or during treatment, including glucocorticosteroids in AIH. The mechanism of secondary serum Tf N-hypoglycosylation in the study group was a severe liver impairment, the exact incidence of this phenomenon is not elucidated. Due to its half-life of approximately 2 weeks, there is a lag between normalization of liver enzymes and improvement of transferrin glycosylation. This fact rules out the usage of serum transferrin glycosylation as a (secondary) marker for ALI/ALF in the absence of previous measurements.

Interestingly, two patients presenting recurrent acute liver failure (RALF) had very specific serum Tf IEF profile with a huge increase of asialo-Tf isoform during episodes of ALF. The mechanism of those abnormalities reflects a severe liver impairment and secondary Tf N-hypoglycosylation with a decrease of tetrasialo-Tf and an increase in asialo-Tf.

Between episodes of ALF, liver function recovered completely together with serum Tf IEF profile. This is the first report on







secondary serum Tf N-hypoglycosylation in the course of RALF. Pathogenic variants in *NBAS*, *RINT1*, or *LARS* genes have been identified as a cause of RALF (25, 26). Whole exome sequencing (WES) analysis did not identify the genetic background in our patients.

Besides patients with ALI/ALF, our study group included five patients with serum Tf N-hypoglycosylation secondary to chronic liver disease CLD; three of them underwent LTx. Sixty percent of patients were diagnosed with chronic cholestatic liver disease; the exact mechanism of serum Tf N-hypoglycosylation remains unknown but it could be related to an impaired excretory (and sometimes also synthetic) hepatocellular function. A chronic liver disease impairs the serum Tf glycosylation, thus a decrease of tetrasialo-Tf is observed. Cellular mechanisms of liver regeneration in the course of chronic disease need to be also taken into account.

During follow-up after LTx, the serum Tf IEF profile normalized. This is the first report on serum Tf Nhypoglycosylation in children secondary to chronic cholestatic liver disease and its normalization after LTx.

A recently published paper by Jansen et al. about secondary glycosylation defects in adult patients with liver disease comprises the biggest cohort in the literature, reported up to now (30). Out of 961 enrolled patients, including those qualified for LTx or with chronic liver disease, 247 patients (26%) had hyposialylation of serum Tf. The authors did not identify a typical CDG type 1 pattern, while the majority of patients (70%) had an

increase in the trisialo-Tf isoform. Regarding other patients, 42 (17%) had an elevated percentage of monosialo-Tf isoform and 4 (2%) had an elevated percentage of disialo-Tf isoform. None of the patients had an increase of all isoforms, a feature of most type II CDG.

The dominant isoform in healthy individuals is tetrasialo-Tf (55.7–67.2%), while asialo- and monosialo-Tf isoforms are usually not detectable (4, 5). In CDG-I, there is an increase in asialo- and disialo-Tf, and a decrease in tetrasialo-Tf, whereas in CDG-II there is an increase in asialo-, monosialo, disialo-, and trisialo-Tf (2, 3). **Figure 6** presents the exemplary Tf IEF isoform profiles for each study group in comparison to patients with CDG type I, type II, and type I/II.

An increase in the asialo-, monosialo-, and disialo-Tf isoforms could be related to endoplasmic reticulum (ER) dysregulation secondary to hepatocytes damage (1, 3). ER stress has been observed in a variety of liver diseases and since hepatocytes are enriched in both smooth and rough ER due to its metabolic functions, serum Tf isoforms could be used as a potential biomarker in pediatric liver disease (31–33). However, further studies are needed to elucidate the serum Tf isoforms in pediatric liver disease patients.

Since more convenient methods such as high performance liquid chromatography (HPLC) and ESI-Q-TOF or MALDI-TOF are available in selected laboratories, a further analytical strategy to characterize the abnormal serum Tf profile is needed.

CONCLUSIONS

All pediatric patients with primary liver disease had an increase in asialo-Tf as well as monosialo-Tf isoform. None of them had an elevated percentage of trisialo-Tf isoform.

This is the first report on secondary serum Tf N-hypoglycosylation in the course of RALF.

On follow-up analyses, serum Tf IEF profile normalized in parallel to normalization of liver function tests, spontaneously, or during treatment, including glucocorticosteroids in AIH.

The exact mechanism of serum Tf N-hypoglycosylation in pediatric liver disease patients remains unknown but it could be related to a severe hepatocellular impairment.

The only liver function test result that could be useful for distinguishing between the groups of patients is GGT—it is much lower for group III than for group I.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Materials**, further inquiries can be directed to the corresponding author/s.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Children's Memorial Health Institute Bioethical

REFERENCES

- Marques-da-Silva D, Dos Reis Ferreira V, Monticelli M, Janeiro P, Videira PA, Witters P, et al. Liver involvement in congenital disorders of glycosylation (CDG). A systematic review of the literature. J Inherit Metab Dis. (2017) 40:195–207. doi: 10.1007/s10545-016-0012-4
- Francisco R, Marques-da-Silva D, Brasil S, Pascoal C, Dos Reis Ferreira V, Morava E, et al. The challenge of CDG diagnosis. *Mol Genet Metab.* (2019) 126:1–5. doi: 10.1016/j.ymgme.2018.11.003
- Péanne R, de Lonlay P, Foulquier F, Kornak U, Lefeber DJ, Morava E, et al. Congenital disorders of glycosylation (CDG): Quo vadis? *Eur J Med Genet*. (2018) 61:643–63. doi: 10.1016/j.ejmg.2017.10.012
- de Jong G, van Dijk JP, van Eijk HG. The biology of transferrin. *Clin Chim Acta*. (1990) 190:1–46. doi: 10.1016/0009-8981(90)90278-Z
- Van Eijk HG, van Noort WL, Kros MJ, van der Huel C. The heterogeneity of human serum transferrin and human transferrin preparations on isoelectric focusing gels: no functional difference of the fraction *in vitro*. *Clin Chim Acta*. (1982) 121:209–16. doi: 10.1016/0009-8981(82) 90060-2
- Blomme B, Van Steenkiste C, Callewaert N, Van Vlierberghe H. Alteration of protein glycosylation in liver diseases. J Hepatol. (2009) 50:592– 603. doi: 10.1016/j.jhep.2008.12.010
- Hülsmeier AJ, Tobler M, Burda P, Hennet T. Glycosylation site occupancy in health, congenital disorder of glycosylation and fatty liver disease. *Sci Rep.* (2016) 6:33927. doi: 10.1038/srep33927
- Stibler H. Carbohydrate-deficient transferrin in serum: a new marker of potentially harmful alcohol consumption reviewed. *Clin Chem.* (1991) 37:2029–37. doi: 10.1093/clinchem/37.12.2029
- Flahaut C, Michalski JC, Danel T, Humbert MH, Klein A. The effects of ethanol on the glycosylation of human transferrin. *Glycobiology*. (2003) 13:191–8. doi: 10.1093/glycob/cwg016
- Stadheim LM, O'Brien JF, Lindor KD, Gores GJ, McGill DB. Value of determining carbohydrate-deficient transferrin isoforms

Committee, num 23/KBE/2020. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin.

AUTHOR CONTRIBUTIONS

AT-S, PL, and AB: project administration. AT-S: supervision. PL, AB, IJ, PS, and AT-S: investigation. PS-R: statistical analysis. PL and AB: writing—original draft. PL, AB, PS-R, and AT-S: writing—review and editing. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fped. 2020.613224/full#supplementary-material

Supplementary Table 1 | Pearson's correlation coefficients between % isoforms and liver function test for group II. Correlation coefficient values, for which the p-value ≤ 0.05 marked with color (reddish for positive, bluish for negative).

in the diagnosis of alcoholic liver disease. Mayo Clin Proc. (2003) 78:703-707. doi: 10.4065/78.6.703

- Clarke JD, Novak P, Lake AD, Hardwick RN, Cherrington NJ. Impaired Nlinked glycosylation of uptake and efflux transporters in human non-alcoholic fatty liver disease. *Liver Int.* (2017) 37:1074–81. doi: 10.1111/liv.13362
- Nakagawa T, Uozumi N, Nakano M, Mizuno-Horikawa Y, Okuyama N, Taguchi T, et al. Fucosylation of N-glycans regulates the secretion of hepatic glycoproteins into bile ducts. J Biol Chem. (2006) 281:29797– 806. doi: 10.1074/jbc.M605697200
- Anderson N, Pollacchi A, Hayes P, Therapondos G, Newsome P, Boyter A, et al. A preliminary evaluation of the differences in the glycosylation of alpha-1-acid glycoprotein between individual liver diseases. *Biomed Chromatogr.* (2002) 16:365–72. doi: 10.1002/bmc.167
- Sterling RK, Jeffers L, Gordon F, Sherman M, Venook AP, Reddy KR, et al. Clinical utility of AFP-L3% measurement in North American patients with HCV-related cirrhosis. *Am J Gastroenterol.* (2007) 102:2196– 205. doi: 10.1111/j.1572-0241.2007.01405.x
- Grytczuk A, Bauer A, Gruszewska E, Cylwik B, Chrostek L. Changed profile of serum transferrin isoforms in primary biliary cholangitis. J Clin Med. (2020) 9:E2894. doi: 10.3390/jcm9092894
- Gudowska M, Gruszewska E, Panasiuk A, Cylwik B, Swiderska M, Filisiak R, et al. Changed profile of serum transferrin isoforms in liver diseases. *Clin Lab.* (2017) 63:349–54. doi: 10.7754/Clin.Lab.2016.160636
- Stewart SH, Reuben A, Anton RF. Relationship of abnormal chromatographic pattern for carbohydrate-deficient transferrin with severe liver disease. *Alcohol Alcohol.* (2017) 52:24–8. doi: 10.1093/alcalc/agw069
- Gruszewska E, Wrona A, Gudowska M, Panasiuk A, Cylwik B, Lipartowska-Klimuk K, et al. The transferrin isoforms in chronic hepatitis. *Clin Biochem*. (2017) 50:1131–5. doi: 10.1016/j.clinbiochem.2017.06.013
- Arndt T, van der Meijden BB, Wielders JP. Atypical serum transferrin isoform distribution in liver cirrhosis studied by HPLC, capillary electrophoresis and transferrin genotyping. *Clin Chim Acta.* (2008) 394:42–6. doi: 10.1016/j.cca.2008.03.033

- Bortolotti F, De Paoli G, Tagliaro F. Carbohydrate-deficient transferrin (CDT) as a marker of alcohol abuse: a critical review of the literature 2001–2005. J Chromatogr B Analyt Technol Biomed Life Sci. (2006) 841:96– 109. doi: 10.1016/j.jchromb.2006.05.005
- Guan W, Gao Z, Huang C, Fang M, Feng H, Chen S, et al. The diagnostic value of serum DSA-TRF in hepatocellular carcinoma. *Glycoconj J.* (2020) 37:231–40. doi: 10.1007/s10719-019-09906-x
- Squires RH Jr, Shneider BL, Bucuvalas J, Alonso E, Sokol RJ, Narkewicz MR, et al. Acute liver failure in children: the first 348 patients in the pediatric acute liver failure study group. J Pediatr. (2006) 148:652– 8. doi: 10.1016/j.jpeds.2005.12.051
- Wendon, J, Cordoba J, Dhawan A, Larsen FS, Manns M, Samuel D, et al. EASL Clinical Practical Guidelines on the management of acute (fulminant) liver failure. J Hepatol. (2017) 66:1047–81. doi: 10.1016/j.jhep.2016.12.003
- 24. Fawaz R, Baumann U, Ekong U, Fischler B, Hadzic N, Mack CL, et al. Guideline for the evaluation of cholestatic jaundice in infants: joint recommendations of the North American Society for Pediatric Gastroenterology, Hepatology, and Nutrition and the European Society for Pediatric Gastroenterology, Hepatology, and Nutrition. J Pediatr Gastroenterol Nutr. (2017) 64:154–68. doi: 10.1097/MPG.000000000001334
- Staufner C, Haack TB, Köpke MG, Straub BK, Kölker S, Thiel C, et al. Recurrent acute liver failure due to NBAS deficiency: phenotypic spectrum, disease mechanisms, and therapeutic concepts. *J Inherit Metab Dis.* (2016) 39:3–16. doi: 10.1007/s10545-015-9896-7
- Cousin MA, Conboy E, Wang JS, Lenz D, Schwab TL, Williams M, et al. RINT1 Bi-allelic variations cause infantile-onset recurrent acute liver failure and skeletal abnormalities. *Am J Hum Genet.* (2019) 105:108– 21. doi: 10.1016/j.ajhg.2019.05.011
- 27. Hernaez R, Solà E, Moreau R, Ginès P. Acute-on-chronic liver failure: an update. *Gut.* (2017) 66:541–53. doi: 10.1136/gutjnl-2016-312670

- Holm, S. A simple sequentially rejective multiple test procedure. *Scand J Stat.* (1979) 6:65–70. Available online at: http://www.jstor.org/stable/4615733
- Benjamini, Y, Hochberg, Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc. Series B Stat Methodol.* (1995) 57:289–300. doi: 10.1111/j.2517-6161.1995.tb02031.x
- Jansen JC, van Hoek B, Metselaar HJ, van den Berg AP, Zijlstra F, Huijben K, et al. Screening for abnormal glycosylation in a cohort of adult liver disease patients. J Inherit Metab Dis. (2020) 43:1310–20. doi: 10.1002/jimd.12273
- Wu FL, Liu WY, Van Poucke S, Braddock M, Jin WM, Xiao J, et al. Targeting endoplasmic reticulum stress in liver disease. *Expert Rev Gastroenterol Hepatol.* (2016) 10:1041–52. doi: 10.1080/17474124.2016.1179575
- Maiers JL, Malhi H. Endoplasmic reticulum stress in metabolic liver diseases and hepatic fibrosis. Semin Liver Dis. (2019) 39:235-48. doi: 10.1055/s-0039-1681032
- Lebeaupin C, Vallée D, Hazari Y, Hetz C, Chevet E, Bailly-Maitre B. Endoplasmic reticulum stress signalling and the pathogenesis of non-alcoholic fatty liver disease. J Hepatol. (2018) 69:927–47. doi: 10.1016/j.jhep.2018. 06.008

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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