Check for updates

OPEN ACCESS

EDITED BY Guzide Aksu, Ege University, Turkey

REVIEWED BY Mehmet Yildiz, Istanbul University Cerrahpasa, Turkey Soamarat Vilaiyuk, Mahidol University, Thailand

*CORRESPONDENCE Begüm Kocatürk bkocaturk@hacettepe.edu.tr Seza Özen sezaozen@gmail.com

SPECIALTY SECTION

This article was submitted to Pediatric Immunology, a section of the journal Frontiers in Pediatrics

RECEIVED 19 October 2022 ACCEPTED 24 November 2022 PUBLISHED 23 December 2022

CITATION

Kocatürk B, Balık Z, Pişiren G, Kalyoncu U, Özmen F and Özen S (2022) Spondyloarthritides: Theories and beyond. Front. Pediatr. 10:1074239. doi: 10.3389/fped.2022.1074239

COPYRIGHT

© 2022 Kocatürk, Balık, Pişiren, Kalyoncu, Özmen and Özen. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Spondyloarthritides: Theories and beyond

Begüm Kocatürk^{1*}, Zeynep Balık², Gaye Pişiren¹, Umut Kalyoncu³, Füsun Özmen¹ and Seza Özen^{2*}

¹Department of Basic Oncology, Hacettepe University Cancer Institute, Ankara, Turkey, ²Division of Pediatric Rheumatology, Department of Pediatrics, Hacettepe University Faculty of Medicine, Ankara, Turkey, ³Division of Rheumatology, Department of Internal Medicine, Hacettepe University Faculty of Medicine, Ankara, Turkey

Spondyloarthritides (SpA) are a family of interrelated rheumatic disorders with a typical disease onset ranging from childhood to middle age. If left untreated, they lead to a severe decrease in patients' quality of life. A succesfull treatment strategy starts with an accurate diagnosis which is achieved through careful analysis of medical symptoms. Classification criterias are used to this process and are updated on a regular basis. Although there is a lack of definite knowledge on the disease etiology of SpA, several studies have paved the way for understanding plausible risk factors and developing treatment strategies. The significant increase of HLA-B27 positivity in SpA patients makes it a strong candidate as a predisposing factor and several theories have been proposed to explain HLA-B27 driven disease progression. However, the presence of HLA-B27 negative patients underlines the presence of additional risk factors. The current treatment options for SpAs are Non-Steroidal Anti-Inflammatory Drugs (NSAIDs), TNF inhibitors (TNFis), Disease-Modifying Anti-Rheumatic Drugs (DMARDs) and physiotherapy yet there are ongoing clinical trials. Anti IL17 drugs and targeted synthetic DMARDs such as JAK inhibitors are also emerging as treatment alternatives. This review discusses the current diagnosis criteria, treatment options and gives an overview of the previous findings and theories to clarify the possible contributors to SpA pathogenesis with a focus on Ankylosing Spondylitis (AS) and enthesitis-related arthritis (ERA).

KEYWORDS

medical biology, spondyloarthritides, ankylosing spondylitis, enthesitis-related arthritis, rheumatology

Introduction

SpA is a group of immune system related disorders predominantly causing sterile inflammation at sacroiliac joints. In adults, patients often meet the definition of axial spondyloarthritis. In addition to AS, reactive arthritis, psoriatic arthritis, enteropathic arthritis, Reiter's syndrome, Inflammatory Bowel Disease (IBD)-associated arthritis and undifferentiated SpA can be included within this disease subset (1). The classification and terminology of juvenile SpA (JSpA) patients differ from the adults. Childhood onset patients are classified as ERA and juvenile onset psoriatic arthritis (JPsA) and juvenile idiopathic arthritis (JIA) is used as an umbrella term for these arthritides (2). In Europe and North America 10% of JIA patients are diagnosed as ERA (3, 4) whereas this ratio increases further to 35%-40% in Asia (5-8).

SpAs affects up to 2% of the population (1). The prevalence of the disease is highest in Europe followed by an Asian population whereas it is uncommon in Africans (9). SpA patients suffer from a significant decrease in their quality of life and may even need surgical operations as a remedy. The therapeutic agents used for the disease may cause side-effects (i.e., infection) and a certain portion of patients fail to respond to therapy (10, 11). Overall, it is clear that the development of alternative treatment strategies are necessary however, the obscure disease etiology plays a negative role in this process. Although there are studies underlining the possible contribution of HLA-B27 allele in disease pathogenesis, the presence of HLA-B27 negative SpA patients indicates the presence of extra risk factors (12).

Clinical features

Similar to other diseases the early diagnosis of SpA is crucial. Delays may result in increased disease activity, irreversible structural damage, low therapy response and limited mobility (13). Physicians and patients should work hand in hand for early diagnosis to eliminate undesirable long-term effects. Diagnosis might be stalled if patients delay visiting a doctor due to limited access or in the belief that their symptoms will disappear spontaneously. Moreover, seeing other specialists rather than a rheumatologist might not only cause a delay but also may result in misdiagnosis (14).

Thorough and distinctive analysis of the symptoms plays a fundamental role in the validity of diagnosis. SpA patients display several common clinical and laboratory findings such as arthritis (Figure 1A), psoriasis (Figure 1A), enthesitis, anterior uveitis (Figure 1B), inflammatory low back pain and family history of HLA-B27-related disease. Although the common features remain the same, the clinical phenotype differs across the ages in certain aspects with peripheral arthritis being predominant in JSpA and axial manifestations being more common in adult-onset disease (15). Inflammatory back pain is the most common complaint of SpA suggesting axial involvement. Shoulder and hip joint involvement is also more common in ERA (16). In fact the differences between childhood and adult- onset disease have been highlighted in a number of studies. Both are more common in males. Childhood cases typically present in adolescent years with arthritis in the big joints and often enthesitis (17). The most frequently involved joints are the knee (40%-50%), hip (30%-40%) and ankle (25%-40%) (16, 18). Axial disease and back pain are less than expected in adult-onset disease. The frequency of axial involvement differs between studies. In a systematic review of the literature, comparing juvenile-onset AS (JoAS) and adult-onset AS (AoAS) cohorts showed that axial disease is significantly more frequent in AoAS than JoAS cases (4.3%–74% vs. 56%–95%) (19). Again family history seems to be more common in childhood-onset disease and may suggest a higher genetic load associated with the disease. Despite these differences, the pathogenesis of these two different onsets are similar; thanks to the recent emerging data we know the main pathways involved and it may not be appropriate to classify the childhood-onset disease separately from the adult one, under the "idiopathic" term anymore.

In recent years MRI has improved the assessment and diagnosis of axial disease. Axial disease was probably underestimated before the widespread use of MRI both in children and adults. Approximately half of the patients are known to be first been diagnosed with sacroiliitis on MRI (20).

Classification criteria

Most juvenile SpA are classified as enthesitis-related arthritis or undifferentiated arthritis, depending on whether psoriasis is present in the patient or their family. International League of Associations for Rheumatology (ILAR) criteria are used for the classification of JIA subtypes including ERA (**Supplementary Table S1**) (2, 21). According to the ILAR, ERA classification criteria is arthritis plus enthesitis or arthritis or enthesitis plus two of the following: (1) Sacroiliac joint tenderness and/or inflammatory back pain, (2) HLA-B27 positivity, (3) >6 years old boy and (4) Acute anterior uveitis and (5) Family history in at least one first degree relative of HLA-B27 associated disease like ankylosing spondylitis, ERA, sacroiliitis with IBD, reactive arthritis or acute anterior uveitis.

In adults the diagnosis is based on the Assessment of SpA International Society (ASAS) classification (**Supplementary Table S1**). ASAS criteria include both imaging and clinical findings: if sacroiliitis is present on imaging [by radiographs or magnetic resonance imaging (MRI)] (**Figure 2**) only one other SpA feature is sufficent for classification. However, if imaging evidence of sacroiliitis is absent, positive HLA-B27 along with at least two other SpA features is required for the patient to be classified as having axial SpA. ASAS criteria for peripheral spondyloarthritis include peripheral arthritis and/or enthesitis and/or dactylitis plus 1 SpA feature (uveitis, psoriasis, Crohn's/colitis, preceding infection, HLA-B27, sacroiliitis on imaging) or ≥ 2 other SpA features (arthritis, enthesitis, dactylitis, inflammatory back pain, family history of SpA).

The ASAS and ILAR criteria set indeed overlap in the defined features and they share several clinical and laboratory findings for classification. However, there are also important differences between the two. Firstly, the ILAR system does not



specifically address children who have SpA by fulfilling the criteria for ankylosing spondylitis, or who have coexisting conditions such as inflammatory bowel disease (22). Reactive arthritis, IBD-related arthritis, and psoriatic arthritis are not among the diagnostic criteria in ERA. While psoriatic arthritis in children is a different subgroup of JIA, both psoriatic arthritis, reactive arthritis, and IBD are included in the SpA group in adults (23). Secondly, the ILAR classification criteria focus on the importance of extra-axial manifestations such as peripheral arthritis and enthesitis, while SpA classification pays attention to the presence of axial and spinal involvement. Finally, elevated inflammatory markers is one of the criteria in SpA, whereas that is not the case in ERA.

Indeed the ILAR criteria has important limitations the classification regarding of patients in the spondyloarthropathy group. Enthesitis-related JIA was considered an undifferentiated SpA in ILAR, whereas all the different forms of adult SpA can be found in children, with the major difference being the higher proportion of undifferentiated forms in children. Thus a new classification criteria have been proposed by researchers from PRINTO, suggesting fundamental changes, in the classification of enthesitis-associated arthritis (24). This new criteria was called "Enthesitis/spondylitis-related JIA and included the following criteria: peripheral arthritis and enthesitis, or arthritis or enthesitis, plus ≥ 3 months of inflammatory back pain and



FIGURE 2

MRI findings in SpA patients. (A) On T2-weighted fat-suppressed coronal sections, areas of bone marrow edema consistent with acute sacroiliitis are observed on the facing sides of the bilateral sacroiliac joint (white arrows). (B) On T1-weighted fat-free coronal sections, erosion and irregularities at the level of the left sacroiliac joint (white arrow) and pseudo-widening (star) of the joint space are observed. In the right sacroiliac joint, there is an appearance of fat replacement (black arrow) in the sacral region.

sacroiliitis on imaging, or arthritis or enthesitis plus 2 of the following: (1) sacroiliac joint tenderness; (2) inflammatory back pain; (3) presence of HLA-B27 antigen; (4) acute (symptomatic) anterior uveitis; and (5) history of a SpA in a first-degree relative. Of note, if peripheral arthritis is present, it should persist for at least 6 weeks.

The current PRINTO definition has been partly harmonized with the adult one, and an imaging criterion for radiographs (25) or magnetic resonance imaging (26) has been introduced. The adult definition of inflammatory back pain has been adopted. Because the term ERA could wrongly suggest the existence of a form of SpA that is specific to childhood, it was initially proposed to name this condition juvenile SpA and was later changed to enthesitis/spondylitis-related JIA. Furthermore one of the main differences of PRINTO classification criteria is that sacroiliitis on imaging was added among the list. Albeit definition of sacroiliitis on MRI for adult patients well-defined, the use of this definition of MRI findings for pediatric cases may cause false-positive results due to the physiologic bone marrow changes of growing bone. Recently, MRI definitions for active and structural sacroiliac joint lesions in juvenile cases are published (27).

Although there is a lack of substantial molecular omics studies, most pediatricians would regard ERA more like a juvenile AS, especially once sacroiliitis is detected. There is a need for more follow-up data on patients with peripheral disease -fulfilling the ERA criteria, to understand whether they constitute a separate pediatric group. Moreover, sophisticated studies are crucial to understand whether axial-ERA is truly an early onset AS or SpA.

Etiology

The lack of knowledge on SpA etiology has been a major concern in diagnosis and disease treatment. The diagnosis is dependent on clinical manifestations which shows heterogeneity between patients whereas the therapeutic interventions were developed based on observational studies. In this section we will discuss the possible risk factors for SpAs and theories related with them.

HLA-B27

The immune system acts as a safeguard to protect our body from the invasion of harmful intruders. These foreign entities' proteins should be presented as peptides to our immune cells to activate a potent immune response. Peptides loaded on Major histocompatibility complex (MHC) molecules located on cell surface can be recognized by T cells which in turn cause their activation. All nucleated cells have MHC class I molecules that take part in the presentation of intracellular antigens (i.e., viral, tumor) to CD8+ T lymphocytes and their heavy chains are encoded by genes at HLA-A, HLA-B and HLA-C loci (28). Antigen presenting cells (APCs) play a significant role in activating the adaptive immune system and are specialized cells. MHC class II molecules are expressed on these cells and are involved in the presentation of exogenous peptides (i.e., bacteria, parasites) to CD4+ T cells. These molecules are encoded by HLA-DR, HLA-DP and HLA-DQ (29).

The first report showing the association of the MHC class I molecule HLA-B27 with SpAs was published in 1973 (30). Since then many studies were conducted to obtain more information on disease etiology and underlying mechanisms. HLA-B27 positive population constitutes 6%–8% of the general population whereas this ratio increases to more than 80% in AS patients (31, 32) implementing its strong plausible contribution to disease etiology. HLA-B27 has different variants with aminoacid substitutions mostly in their peptide binding cleft (33). Among these variants, HLA-B*27:05, HLA-B*27:02 and HLA-B*27:04 show association with SpA whereas this is not the case for HLA-B*27:06 and HLA-B*27:09 (34, 35). HLA-B*27:05 is more common in Caucasian, HLA-B*27:04 in Chinese and HLA-B*27:02 in Mediterranean population (36).

The importance of HLA-B27 in SpA etiology was also recapitulated using animal models. Rats having high levels of HLA-B27*05 and human beta 2 microglobulin (B27-Tg) partially phenocopy the human disease with inflammatory bowel condition, inflammatory peripheral arthritis and skin lesions (37). Interestingly, genetic factors seems to play a role in the process based on the fact that SpA related symptoms are only manifested in rats having Lewis or Fischer background but not in Dark Agouti background. Mice with same genetic modifications also display spontaneous arthritis (38) and the lack of β 2-microglobulin (β 2m) or TAP1 gene does not impair the manifestation of disease related phenotype (39, 40). The background of the mice has been found to be relative for the development of the disease as well (41).

The HLA-B27 levels seems to be a pivotial factor regulating disease susceptibility. Higher levels of HLA-B27 are typically seen in the peripheral blood mononuclear cells (PBMCs) of patients compared to healthy controls positive for this allele (42). Moreover, individuals homozygote for HLA-B27 are associated with an increased risk of AS development compared to heterozygotes (43). The same phenomenon is also observed in animal models. Disease susceptibility shows a positive correlation with HLA-B27 copy number and its relative expression in lymphoid cells (44) that can be upregulated *via* pro-inflammatory stimuli. Of note, this dose dependent effect might also explain why only 2% of HLA-B27 positive patients develop the disease.

The positivity of HLA-B27 also has an influence on disease manifestation. In more than 80% of AS patients, symptoms emerge at \leq 30 years of age. Interestingly, HLA-B27 positive

AS patients show an earlier disease onset compared to negative ones (45) and have a worse prognosis with elevated disease activity and duration (46). The frequency of specific symptoms also depends on HLA-B27 status. Psoriasis and IBD are more common in HLA-B27 negative patients whereas peripheral arthiritis and uveitis are observed more frequently in HLA-B27 positive ones (47).

Overall, it is clear that HLA-B27 somehow plays a role in disease pathogenesis. Its possible contribution to disease progression and related theories are discussed below.

Arthritogenic peptide/molecular mimicry hypothesis

The mature MHC I molecule is composed of a heavy chain (HC), a ß2m light chain and a peptide, 8-10 amino acids in length. Its formation involves a series of protein assembly and disassembly within the complex. First, newly synthesized heavys chains are translocated to the endoplasmic reticulum (ER) and glycosylated. This post-translational modification acts as a signal for incomplete folding which in turn triggers HCs interactions with chaperones calnexin and calreticulin. As HCs gain the correct tertiary structure, they associate with β 2m resulting in the dissociation of calnexin (48, 49). Next, the complex further interacts with a transporter associated with antigen processing (TAP) via tapasin which is bound to ERp57 to form the peptide loading complex (48). Eventually, Erp57 and calreticulin dissociate to allow the binding of peptides to the MHC I. Although, MHC class I molecules are responsible for the presentation of the peptides to CD8+ T cells, these peptides should be trimmed before they are loaded on the complex. For that, purpose proteasome performs the initial trimming process causing the formation of peptides ~15 aa in length. These peptides enter ER through TAP transporter and furher cleaved by ERAP1 and ERAP2 to have the optimal length for the loading (50). Finally, as MHC I is loaded with the peptide, the complex is sent to the surface of nucleated cells in particular APCs to perform a successful round of peptide presentation.

The plausible contribution of APCs in disease pathogenesis has been the center of many studies. The increased abundance of macrophages in sacroiliac (51) and enthesis (52) biopsies of AS patients attain a possible role for these cells. The level of circulating CD141+ dendritic cells (DCs) show a positive correlation with BASDAI in AS patients (53). Moreover, lower levels of MHC class II expression in DCs of AS patients (54) and animal model (55) implies that distortions in antigen presentation might very well be a key factor in disease pathogenesis.

Previous studies suggested that HLA-B27 binds to a distinctive set of peptides that show similarity to self-peptides (Figure 3). Their presentation to CD8+ T lymphocytes triggers the breakdown of self tolerance which in turn activates a destructive immune response in affected sites (56). In support of this notion, HLA-B27-restricted CD8+ T cells

were detected in the synovial fluid of AS patients (57, 58). They are also found to be directed against self-peptides derived from vasoactive intestinal peptide type 1 receptor (VIP1R, aminoacids 400–408) and glucagon receptor (GR, aminoacids 412–420) (59, 60). A controversial finding pointed out that HLA-B*27:09 subtype that is not associated with the disease also presents the VIP1R-derived peptide (61). However, further investigations revealed that peptide's conformation differs from the one presented by the disease relevant variant HLA-B*27:05 (59).

The molecular mimicry between HLA-B27 and gramnegative bacteria was also suggested to be a key element in autoreactive T cell activation and autoimmune reaction. Indeed, the sequence homology between HLA-B27 and arthritogenic bacterias *Klebsiella pneumoniae* (62), *Yersinia enterocolitica, Salmonella typhimurium, Shigella flexneri* and *Shigella sonnei* was described (63). This theory is further supported by a study by Ramos et al. showing that a peptide derived from the intracytoplasmic tail of HLA-B27 shows similarity to *Chlamydia trachomatis* and acts as a ligand only for disease associated HLA-B27 variants (64).

Although presentation of unusual peptides by HLA-B27 was suggested in disease pathogenesis, the ongoing presentation of disease related symptoms in CD8+ T cell depleted B27-Tg rats or TAP1^{-/-} mice argues strongly against the importance of antigen presentation in disease etiology (65, 66) thus other theories were also developed (40, 65, 66).

Homodimeric HLA-B27 molecule

As mentioned above, the major function of a MHC class I molecule which is composed of a heavy chain, ß2m and a peptide, is to present these peptides to CD8+ T cells. Rather, HLA-B27 was reported to be recognized by Natural Killer Cells (NKs) and CD4+ T lymphocytes in the form of ß2mfree homodimers (67) which is established through an unpaired cysteine at position 67 (68) (Figure 3). Strikingly, the B27-Tg rats with functional NK cells do not present disease symptoms indicating that these cells act in concert with lymphoctes in disease manifestation (69). As a matter of fact the critical involvement of CD4+ T cells in disease progression was recapitulated in many studies. Transfer of different T lymphocyte populations to athymic nude B27-Tg mice revealed that CD4+ T cells are the major cell population inducing colitis (70). Moreover, their levels shows an increment in the lymph nodes draining the sites of inflammation in animal model (71) and in peripheral blood of SpA patients (72). Higher levels of activated CD4+ T cells is also observed in B27-Tg rats compared to healthy ones (65).

Studies aiming to understand the involvement of dimer formation in disease pathogenesis unearthed that HLA-B27 variants associated with SpA have an increased tendency for dimer formation (73). The receptors for HLA-B27 homodimers were found to be KIR3DL1, LILRB2 (74),



KIR3DL2 (75) and LIR6 (68) and expressed on lymphocytes, monocytes and Natural Killer Cells (NKs) (75). The level of KIR3DL2 positive NK and CD4+ T cells increases in the peripheral blood and synovial fluid of SpA and ERA patients. The receptor engagement also shows a survival and activation profile in NKs and T cells respectively (73, 76) whereas the dissociation between KIR3DL2 and HLA-B27 decreases the survival/proliferation of NKs and the release of disease related cytokine IL17 from the PBMCs of AS patients (77). In line with this finding, KIR3DL2+ CD4+ T cells collected from the synovial fluid of SpA patients displays enhanced levels of IL17 secretion (78).

Unfolded protein response (UPR) activation

UPR activation was suggested to be a major explanation in SpA pathogenesis. Proteins are biomolecules that orchestrate many cellular processes. To fulfill their task, they need to be folded

properly in the organelle called Endoplasmic Reticulum (ER). Proteins with folding deficits can be removed via autophagy or Endoplasmic Reticulum Associated Degradation (ERAD) (79). However, the accumulation of misfolded proteins may also take place which in turn activates a stress response machinery namely UPR. This stress is regulated through 3 transmembrane proteins located on ER: Inositol-requiring enzyme 1 (IRE1), PKR-like ER kinase (PERK) and Activating Transcription Factor 6 (ATF6). Upon its activation, IRE1 cleaves Xbp mRNA leading to the formation of an active transcription factor sXbp. This factor is responsible for the synthesis of chaperones and ERAD components to achieve cellular homeostasis whereas an increase in magnitude and duration of stress results in the activation of the apoptotic IRE1-JNK pathway. Activation of PERK leads to the phosphorylation of eIF2a which in turn put a halt to translation whereas activating selective translation of the

ATF4 transcription factor. Similar to IRE1 pathway, ATF4 is also responsible for the activation of homeostatic genes yet unresolved stress promotes the expression of pro-apoptotic CHOP. ATF6 is translocated to Golgi upon UPR and cleaved with S1P and S2P proteases. The newly formed cytosolic domain of ATF6 is a transcription factor and is involved in the transcription of chaperones (80).

HLA-B27 is unique in a way that it misfolds even in the presence of β 2m and peptides which in turn activate UPR (**Figure 3**). There are several bodies of evidence proving this aberrant feature of HLA-B27:

- Folding rate: HLA-B27 has a slow folding nature which in turn induces homodimer formation and its retention in the ER. These molecules can then activate the UPR. The B pocket which is located at the peptide binding groove of HLA-B27 seems to be crucial for this machinery. Altering residues in this region not only enhances HLA-B27's folding but also alleviates homodimer formation (81).
- ERAD: Another clue showing the misfolded nature of HLA-B27 is its enhanced predisposition to undergo ERAD (82) in which EDEM1, and HRD1 were found to be pivotal regulators (83, 84). In support of this notion, the use of ERAD blocking reagents results in an increment in the levels of HLA-B27 dimers/oligomers.
- Interaction with chaperones: The chaperones help misfolded proteins to gain a proper tertiary structure. The prolonged interaction between HLA-B27 multimers and chaperone Bip indicates the improper folding of HLA-B27 which in turn activates the stress response (85). In addition, the enhanced interaction between HLA-B27 and oxidoreductase Erp57 is also involved in dimer formation which again may turn on UPR (86).

Macrophages residing in the peripheral joints of AS patients have increased levels of Bip compared to osteoarthritis patients (87) and mononuclear cells collected from the synovial fluid of SpA patients shows an activation state for UPR (88). In B27-Tg rats, bone marrow derived macrophages shows prominent UPR activation status which shows a positive correlation with HLA-B27 levels (89, 90). The UPR induction was also observed in B27-Tg rats' dendritic cells (55). Strikingly, ERAP1 deficient B27-Tg rats remained healthy due to the blockade of UPR activation (91).

The pathogenesis of SpA clearly involves the activation of the immune system thus UPR-driven immune modulation has also been the subject of extensive investigation. NF κ B was shown to be activated during UPR (92) that mediates Th17 differentiation *via* IL23, a cytokine that is elevated in the serum and synovial fluid of SpA patients (93, 94). The activated Th17 cells in turn produce cytokines such as IL17, TNF and IL6 (95). A strong activation for IL23/IL17 axis was detected in the colon of B27-Tg rats (96) and overexpression of IL23 causes a disease phenotype similar to AS in mice (97). In addition, Th17 cells were found to be enriched in the peripheral blood of AS patients (98). Furthermore, DCs, a major source for IL23, contribute to Th17 cells' expansion in the B27-Tg animal model (71). Although, macrophages with prominent UPR activation are destined to produce higher levels of IL23 (96), there are also studies showing that IL23 production is independent of UPR activation thus further studies are warranted (99).

Another NF κ B dependent cytokine TNF- α is also a critical component of the disease and also used as a target for therapy. Similar to IL23, overexpression of TNF results in spondyloarthritis formation in mice and this process was found to be regulated through mesenchymal cells (100). The level of Bip in the macrophages collected from the synovial fluid shows a positive correlation with TNF levels indicating that immune modulation by UPR might be the basis for elevated TNF levels in disease (87). TNF is detected in the inflamed tissues of SpA patients and also is elevated in PBMCs and serum (101).

Other susceptibility genes

Studies on families revealed that SpAs may have a heritable component. For JIA the recurrence risk in first cousins was determined to be 5.8 fold whereas the sibling relative risk is estimated to be 11.6 fold (102). AS's heritability is \geq 90% (103, 104) with a sibling recurrence risk of 8.2% (105). The prevalence increases dramatically with the presence of a first degree relative suffering from the disease (106, 107) and the concordance rate was determined to be 25%-75% and 4%-15% in monozygotic and dizygotic twins respectively (103, 104, 108). Overall, these findings strongly indicate that genetic factors are key determinants in disease pathogenesis. As mentioned above the presence of HLA-B27 showed the highest association with disease susceptibility. However, the fact that only 2% of the HLA-B27 positive population develops SpA is a strong indicator that there are additional genetic risk factors for the development of the disease (109). To understand this phenomenon better genome wide association studies (GWAS) were performed in SpA patients.

The second well established susceptibility locus for AS and ERA was found to be ERAP1 (110, 111) which is involved in the presentation of peptides with optimal length. Several studies made it apparent that defects in ERAP function might be involved in disease pathogenesis. ERAP1 variants with a loss of function shows a protective effect for disease which provides strong evidence for the involvement of atypical processing of antigenic peptides (112). Indeed, HLA-B27 was shown to bind extended peptides with protruding C-terminus (113) which in turn may activate a potent T cell response thus leading to SpA development. The deficits in peptide trimming might also decrease the level of peptide- loaded

10.3389/fped.2022.1074239

MHC molecules which in turn increase the levels of misfolded HLA-B27 molecules, UPR activation and disease progression. Elevated UPR levels might also be regulated through other mechanisms including damaged ubiquitin-ERAD machinery. Previous studies clearly show that ubiquitin conjugating enzyme UBE2J1 is involved in targeting of MHC class I molecules for ERAD (83) thus further studies aiming to unveil the link between another susceptibility gene UBE2E3 (114) and ERAD in disease etiology would be valuable.

Shaping immune response is indispensable in AS pathogenesis. As mentioned above, the IL23/IL17 axis is a crucial component of this machinery. Its importance was also verified in GWAS studies. Molecules related with this pathway (IL23R, IL12B, IL6R, IL1R1, IL1R2, TYK2, IL27A, STAT3, JAK2) are among the gene loci that shows association with AS (83, 114). This also holds true for the TNF- α pathway. Research revealed the presence of disease associated SNPs near to/in TNFRSF1A (83, 114), TNFSF15 (115) and TRADD (116). Genes related to T cell regulation, RUNX3, IL7R, EOMES, ZMIZ1, ICOSLG, SH2B3 and BACH2, are also among the AS risk loci (9). Other genes showing association with AS are GPR25, GPR65, GPR35, TBKBP1, PTGER4, BACH2, NOS2, FCGR2A, NKX2-3 (9), CARD9 (117, 118), KIF21B (119), ANTXR2 (120), ANO6 (121).

For JIA, ERAP1and IL23R are among the disease susceptibility genes for ERA and juvenile psoriatic arthritis respectively (111). The lower prevalence of JIA subtypes hampers the construction of well-powered cohorts for GWAS analysis. Thus combining all JIA subtypes rather than investigating them separately was used to detect genetic associations. PTPN2, ANGPT1, COG6 (122), CD80, JMJD1C (123), TRAF1-C5 (124), VTCN1 (125), IL2RA, IL2RB, STAT4 (126), TNFAIP3 and TRAF1/C5 (127) were found to be JIA-predisposing loci.

Gender

Many rheumatic diseases display gender predominance. Both the incidence of AS (109) and ERA (128) are higher in males. However, the male to female ratio has showed decrement over time (129). This gender predominance indicates that sexspecific factors might play a role in SpA pathogenesis. Among these factors the impact of hormones in SpA progression was extensively analyzed. Of note, the age interval for ERA patients is 6–16 and it is well known that the level of sex hormones increases with puberty thus the following observations may partially explain the male dominance of ERA.

The effect of TNF inhibitors on SpA progression underlines TNF's importance in disease progression. Interestingly, estrogen was shown to decrease inflammation in SpA patients *via* downregulating TNF alpha levels (130). Estrogen supplementation was also shown to decrease disease severity both in human (131) and animal female subjects (132) whereas there are other studies showing no evident association (133). Testosterone levels on the other hand did not show any difference between SpA patients and healthy controls and were not likely to regulate disease progression (131, 134). Sex hormones were also shown to regulate the microbiome (135) and immune system (136). The fact that both of these factors play a role in disease progression (see below) underlines the presence of a possible hormone-driven microbiome and immune system related axis however, further studies are needed.

The gut and microbiome

The gut is one of the affected sites in SpA. Inflammatory bowel disease (Crohn's disease and ulcerative colitis) occur concomitantly in up to 10% of SpA positive population (137). Patients display inflammatory lesions at intestinal mucosa (138) and the gut is also inflamed in the animal model (139) indicating that an active immune response in the gut and SpA might be interlinked. In support of this notion, macrophages expressing the CD163 scavenger receptor increases in the colonic mucosa of SpA patients (140) and IL23/IL17 axis is exacerbated in the colon of B27-Tg rats (96).

The relation between SpA development and microbiota has been the center of attention for decades. In animal models, housing of the animals in pathogen-free conditions alleviated the formation of several disease related symptoms including colitis and arthritis. However, their transfer to conventional conditions caused their manifestation. Moreover, treating B27-Tg rats with antibiotics hampered colitis formation (141) indicating that the microbiome is a key player in disease pathogenesis (39, 142). Indeed, there are studies showing the differences in microbiome of SpA patients and healthy controls (143-145) and ileal biopsies from AS patients revealed the presence of adherent and invasive bacteria which is accompanied by the decreased barrier function of the gut (146). Mucins play a major role in barrier function. Mucin-degrading Akkermansia muciniphila species was found to be elevated in B27-Tg rats indicating that SpA related dysbiosis may be involved in impaired gut barrier (147). T cells are another key player for maintaining the tolerance against commensal bacteria (148). Interestingly, CD4+ T cells isolated from B27-Tg rats produces higher levels of IFN- γ in response to antigens derived from these organisms implying that there might be a loss of tolerance for the microbiome (149, 150). Moreover, the defective stimulation of T cells by APCs might also contribute to the loss of tolerance for microbial flora (151, 152).

The link between treatment response and microbiome was also investigated. Patients receiving 3 months of anti-TNF therapy did not show a significant difference in their microbiata composition. However, having higher levels of *Burkholderiales* prior to therapy and an increment in genus Dialister after therapy was observed in responders (153).

Diet

Diet plays a crucial role in the development and progression of many diseases. Diet has also been investigated in SpA however, most studies were not replicated. A study by Haugen et al. indicated that many AS patients reported that diet plays a role in the manifestation and severity of their symptoms (154) and they follow certain diets to decrease their intensity (155).

Starch consumption was suggested to be an exacerbating factor in SpAs and a low starch diet was found to lower disease activity whereas there are also studies showing no impact (155, 156). Salt and dietary fat consumption did not show any correlation with the severity of the symptoms (155, 157). Although quitting dairy products seem to have an ameliorating role in disease (158) there are also studies showing no effect (157).

In human subjects, the impact of prebiotic uptake in SpA progression was analyzed. SpA patients with concomitant quiescent ulcerative colitis receiving *Lactobacillus acidophilus* and *Lactobacillus salivarius* displayed lower disease activity (159). In contrast, a meta-analysis by Sanchez et al. opposed this finding (160). In animal model the severity of colitis was diminished with the supplementation of diet with prebiotics (161). The constituent, fructo-oligosaccharides was found to have the greatest anti-inflammatory effect in this regard (162, 163). Fibre-rich diets also showed a remedial effect on disease by upregulating short chain fatty acids. Indeed, administration of propionate to B27-Tg animals attenuates intestinal inflammation (164).

Treatment and outcome

There are several treatment options used in clinics for SpAs. However, the current therapy options do not always result in full remission. Treatment of ERA varies according to whether the disease is axial or peripheral, the number of active joints, the presence of risk factors, and accompanying extra-articular features (Table 1). NSAIDs are used as the first-line treatment in enthesitis and sacroiliitis because of their analgesic and anti-inflammatory effects. For peripheral disease, DMARDs, especially methotrexate or salazopyrin are recommended. Sulfasalazine or methotrexate is used for enthesitis or active peripheral arthritis (165). The response to these non-biologic DMARDs varies in a wide range (166). Non-biologic DMARDs can also be used to prevent the development of anti-drug monoclonal antibodies against TNF inhibitors (TNFis) (167). Methotrexate and Salazopyrin monotherapy is not recommended in active sacroiliitis whereas they can be used as an adjunct therapy. If arthritis does not respond to non-biologic DMARDs or for patients who develop the axial disease then biologic DMARDs would be indicated, often along with the NSAID treatment. Among these, anti-TNF drugs are the first choice. Since etanercept and adalimumab are licensed for pediatric use, the present data is mainly focused on the effectiveness and safety of these two monoclonal anti-TNF drugs (168, 169).

Recently anti-IL17 has become an alternative treatment in ERA as well. A total of 86 patients (52 ERA, 34 JSpA 34 patients; median age, 14 years) were enrolled for an open-label secukinumab trial in

TABLE 1 ERA treatment algorithm.



[†]Adapted from the 2019 American College of Rheumatology/Arthritis Foundation Guideline

^{*} Methotrexate monotherapy is not recommended as a treatment for sacroiliitis. It may have utility as adjunct therapy in patients with concomitant perpheral polyarthritis or to prevent the development of anti-drug antibodies against certain monoclonal TNFis.

the first treatment period. In the second period, responders received secukinumab or placebo. Secukinumab demonstrated a significantly longer time to disease flare and a consistent safety profile similar to adults (170). Moreover, there are ongoing clinical trials for ixekizumab (NCT04527380).

Bridging therapy with systemic glucocorticoids might be used during the initiation or escalation of therapy. Intraarticular glucocorticoid injections of the sacroiliac joints as an adjunct therapy are conditionally recommended (165). Physiotherapy is also a crucial element in the treatment process thus should be offered to all SpA and JIA patients.

Another important aspect of the treatment is to monitor the side-effects of the drugs. For NSAIDs, gastrointestinal problems may arise thus proton pump inhibitors might also be prescribed. On the other hand, adequate fluid intake is essential to circumvent renal injury (171). Anti-TNF drugs make patients prone to infections thus in countries where tuberculosis is still encountered, routine screening should be performed.

Disease activity has to be followed to evaluate the response to treatment. For JIA patients, Juvenile Arthritis Disease Activity Score (JADAS) and Bath Ankylosing Spondylitis Disease Activity Index (BASDAI) are screened whereas BASDAI and Ankylosing Spondylitis Disease Activity (ASDAS) are used for the assessment of therapy success in adult SpAs (172, 173). These are applied for childhood diseases with axial involvement as well. Finally, Weiss et al. have developed and validated the first disease activity assessment for JSpA through international input and consensus formation techniques: this new criterion was called the Juvenile Spondyloarthritis Disease Activity (JSpADA) Index. This outcome tool had a good performance in discriminating between subjects with active vs. inactive disease and responded well to changes in the disease activity (174).

For adults, many therapeutics have been used and published. The reader is referred to excellent reviews on the subject. The primary treatment for SpA is NSAIDs and TNFis (certolizumab, etanercept, infliximab, adalimumab, and golimumab). Of note, SpA patients have higher levels of TNF- α (101) and HLA-B27 positive patients have a better response rate to TNF therapy (175) that might be explained by higher TNF levels in these patients (176). Therefore having TNF levels above a certain threshold value may help to estimate a better response and analysis of TNF levels before treatment might be beneficial.

wIL17 inhibitors (secukinumab and ixekizumab) can also be used for patients. Anti-IL17 is not recommended in patients with IBD or recurrent uveitis. If the patient has tuberculosis or recurrent infections, sulfasalazine is preferred over secukinumab and ixekizumab. Tofacitinib (a JAK inhibitor) is a second-line option for patients with contraindications to TNFi or anti-IL17. Co-treatment with low-dose methotrexate is not generally recommended except with infliximab (177). Brodalumab (IL17RA), bimekizumab (dual inhibition of IL17A and IL17F), and upadacitinib (selective JAK1 inhibitor) demonstrated improvement in active axial SpA (178–180). Although IL23 inhibitors (tildrakizumab, risankizumab and guselkumab) are effective in the treatment of psoriatic arthritis (181), in phase 2 and phase 3 studies, the use of ustekinumab and risankizumab did not show any improvement on SpA disease activity (182, 183). Of note, discontinuation of these biologic disease-modifying drugs (DMARDs) is not recommended due to the risk of flare (177) and all these biological DMARDs may be studied in adolescent patients as well.

Conclusion

As summarized above, studies ongoing for more than 4 decades have led to the discovery of many risk factors for SpA development. Among these factors HLA-B27 seems to be the spearhead helping us to better understand the etiology of the disease. HLA-B27 driven mechanisms are thought to involve UPR activation and switching on the IL23/IL17 axis.

The fact that only a part of HLA-B27 positive people develop SpA indicates that there are additional factors contributing to disease pathogenesis. Although the threshold effect for HLA-B27 might be a possible explanation for this observation, it is most likely that further investigation of factors other than HLA-B27 is required that will also pave the way for the development of alternative therapies. The current treatment regimen involves NSAIDs, TNF inhibitors and possibly DMARDs. However, only some patients respond to the treatment which in turn causes a significant decrease in the non-responders' quality of life. Therapies targeting UPR and IL23/IL17 axis have recently gained attention but clinical trials are needed for further validation.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Acknowledgments

Figure 3 was created with Biorender.com. Anterior uveitis, photo courtesy of Sibel Kadayıfçılar, Department of Ophtalmology, Hacettepe University, Turkey.

Conflict of interest

UK has received consultancy fees and/or speaker's bureau from Abbvie, Novartis, UCB, Lilly and Pfizer. SÖ has received consultancy fees and/or speaker's bureau from Novartis and SOBI. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the

References

1. Stafford L, Youssef PP. Spondyloarthropathies: an overview. Intern Med J. (2002) 32:40-6. doi: 10.1046/j.1445-5994.2002.00132.x

2. Petty RE, Southwood TR, Manners P, Baum J, Glass DN, Goldenberg J, et al. International league of associations for rheumatology classification of juvenile idiopathic arthritis: second revision, Edmonton, 2001. *J Rheumatol.* (2004) 31:390–2.

3. Davies R, Carrasco R, Foster HE, Baildam EM, Chieng SEA, Davidson JE, et al. Treatment prescribing patterns in patients with Juvenile Idiopathic Arthritis (JIA): analysis from the UK Childhood Arthritis Prospective Study (CAPS). *Semin Arthritis Rheum.* (2016) 46:190–5. doi: 10.1016/j.semarthrit.2016.06.001

4. Guzman J, Oen K, Tucker LB, Huber AM, Shiff N, Boire G, et al. The outcomes of juvenile idiopathic arthritis in children managed with contemporary treatments: results from the ReACCh-out cohort. *Ann Rheum Dis.* (2015) 74:1854–60. doi: 10.1136/annrheumdis-2014-205372

5. Arkachaisri T, Tang SP, Daengsuwan T, Phongsamart G, Vilaiyuk S, Charuvanij S, et al. Paediatric rheumatology clinic population in Southeast Asia: are we different? *Rheumatology*. (2017) 56:390–8. doi: 10.1093/rheumatology/kew446

6. Kunjir V, Venugopalan A, Chopra A. Profile of Indian patients with juvenile onset chronic inflammatory joint disease using the ILAR classification criteria for JIA: a community-based cohort study. *J Rheumatol.* (2010) 37:1756–62. doi: 10. 3899/jrheum.090937

7. Shen CC, Yeh KW, Ou LS, Yao TC, Chen LC, Huang JL. Clinical features of children with juvenile idiopathic arthritis using the ILAR classification criteria: a community-based cohort study in Taiwan. *J Microbiol Immunol Infect.* (2013) 46:288–94. doi: 10.1016/j.jmii.2012.03.006

8. Tanya M, Teh KL, Das L, Hoh SF, Gao X, Arkachaisri T. Juvenile idiopathic arthritis in Southeast Asia: the Singapore experience over two decades. *Clin Rheumatol.* (2020) 39:3455-64. doi: 10.1007/s10067-020-05081-9

9. Cortes A, Hadler J, Pointon JP, Robinson PC, Karaderi T, Leo P, et al. Identification of multiple risk variants for ankylosing spondylitis through highdensity genotyping of immune-related loci. *Nat Genet.* (2013) 45:730–8. doi: 10. 1038/ng.2667

10. Braun J, Brandt J, Listing J, Zink A, Alten R, Golder W, et al. Treatment of active ankylosing spondylitis with infliximab: a randomised controlled multicentre trial. *Lancet.* (2002) 359:1187–93. doi: 10.1016/S0140-6736(02)08215-6

11. Minozzi S, Bonovas S, Lytras T, Pecoraro V, Gonzalez-Lorenzo M, Bastiampillai AJ, et al. Risk of infections using anti-TNF agents in rheumatoid arthritis, psoriatic arthritis, and ankylosing spondylitis: a systematic review and meta-analysis. *Expert Opin Drug Saf.* (2016) 15:11–34. doi: 10.1080/14740338. 2016.1240783

12. Chung HY, Machado P, van der Heijde D, D'Agostino MA, Dougados M. HLA-B27 positive patients differ from HLA-B27 negative patients in clinical presentation and imaging: results from the DESIR cohort of patients with recent onset axial spondyloarthritis. *Ann Rheum Dis.* (2011) 70:1930–6. doi: 10. 1136/ard.2011.152975

13. Seo MR, Baek HL, Yoon HH, Ryu HJ, Choi HJ, Baek HJ, et al. Delayed diagnosis is linked to worse outcomes and unfavourable treatment responses in patients with axial spondyloarthritis. *Clin Rheumatol.* (2015) 34:1397–405. doi: 10.1007/s10067-014-2768-y

14. Raciborski F, Klak A, Kwiatkowska B, Batko B, Sochocka-Bykowska M, Zon-Giebel A, et al. Diagnostic delays in rheumatic diseases with associated arthritis. *Reumatologia*. (2017) 55:169–76. doi: 10.5114/reum.2017.69777 editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fped. 2022.1074239/full#supplementary-material.

15. Chen HA, Chen CH, Liao HT, Lin YJ, Chen PC, Chen WS, et al. Clinical, functional, and radiographic differences among juvenile-onset, adult-onset, and late-onset ankylosing spondylitis. *J Rheumatol.* (2012) 39:1013–8. doi: 10.3899/jrheum.111031

16. Riley MJ, Ansell BM, Bywaters EG. Radiological manifestations of ankylosing spondylitis according to age at onset. *Ann Rheum Dis.* (1971) 30:138–48. doi: 10.1136/ard.30.2.138

17. Goirand M, Breton S, Chevallier F, Duong NP, Uettwiller F, Melki I, et al. Clinical features of children with enthesitis-related juvenile idiopathic arthritis/ juvenile spondyloarthritis followed in a French tertiary care pediatric rheumatology centre. *Pediatr Rheumatol Online J.* (2018) 16:21. doi: 10.1186/ s12969-018-0238-9

18. Guo R, Cao L, Kong X, Liu X, Xue H, Shen L, et al. Fever as an initial manifestation of enthesitis-related arthritis subtype of juvenile idiopathic arthritis: retrospective study. *PLoS One*. (2015) 10:e0128979. doi: 10.1371/journal.pone.0128979

19. Jadon DR, Ramanan AV, Sengupta R. Juvenile versus adult-onset ankylosing spondylitis – clinical, radiographic, and social outcomes. a systematic review. *J Rheumatol.* (2013) 40:1797–805. doi: 10.3899/jrheum.130542

20. Tsoi C, Griffith JF, Lee RKL, Wong PCH, Tam LS. Imaging of sacroiliitis: current status, limitations and pitfalls. *Quant Imaging Med Surg.* (2019) 9:318–35. doi: 10.21037/qims.2018.11.10

21. Fisher C, Ciurtin C, Leandro M, Sen D, Wedderburn LR. Similarities and differences between juvenile and adult spondyloarthropathies. *Front Med.* (2021) 8:681621. doi: 10.3389/fmed.2021.681621

22. Colbert RA. Classification of juvenile spondyloarthritis: enthesitis-related arthritis and beyond. *Nat Rev Rheumatol.* (2010) 6:477–85. doi: 10.1038/nrrheum.2010.103

23. Weiss PF, Fuhlbrigge RC, von Scheven E, Lovell DJ, Colbert RA, Brunner HI. Children with enthesitis-related arthritis and possible benefits from treatments for adults with spondyloarthritis. *Arthritis Care Res.* (2022) 74:1058–64. doi: 10.1002/acr.24529

24. Martini A, Ravelli A, Avcin T, Beresford MW, Burgos-Vargas R, Cuttica R, et al. Toward new classification criteria for juvenile idiopathic arthritis: first steps, pediatric rheumatology international trials organization international consensus. *J Rheumatol*, (2019) 46:190–7. doi: 10.3899/irheum.180168

25. van der Linden S, Valkenburg HA, Cats A. Evaluation of diagnostic criteria for ankylosing spondylitis. A proposal for modification of the New York criteria. *Arthritis Rheum.* (1984) 27:361–8. doi: 10.1002/art.1780270401

26. Lambert RG, Bakker PA, van der Heijde D, Weber U, Rudwaleit M, Hermann KG, et al. Defining active sacroiliitis on MRI for classification of axial spondyloarthritis: update by the ASAS MRI working group. *Ann Rheum Dis.* (2016) 75:1958–63. doi: 10.1136/annrheumdis-2015-208642

27. Weiss PF, Brandon TG, Lambert RG, Biko DM, Chauvin NA, Francavilla ML, et al. Data-driven MRI definitions for active and structural sacroiliac joint lesions in juvenile spondyloarthritis typical of axial disease; a cross-sectional international study. *Arthritis Care Res.* (2022). doi: 10.1002/acr.25014

28. Al NH, Mawart A, Alshamsi J, Al SH, Tay GK. Major histocompatibility complex (MHC) associations with diseases in ethnic groups of the Arabian Peninsula. *Immunogenetics*. (2021) 73:131–52. doi: 10.1007/s00251-021-01204-x

29. Rock KL, Reits E, Neefjes J. Present yourself! by MHC class I and MHC class II molecules. *Trends Immunol.* (2016) 37:724–37. doi: 10.1016/j.it.2016.08.010

30. Caffrey MF, James DC. Human lymphocyte antigen association in ankylosing spondylitis. *Nature.* (1973) 242:121. doi: 10.1038/242121a0

31. Schlosstein L, Terasaki PI, Bluestone R, Pearson CM. High association of an HL-A antigen, W27, with ankylosing spondylitis. *N Engl J Med.* (1973) 288:704–6. doi: 10.1056/NEJM197304052881403

32. Sieper J, Rudwaleit M, Khan MA, Braun J. Concepts and epidemiology of spondyloarthritis. *Best Pract Res Clin Rheumatol.* (2006) 20:401–17. doi: 10. 1016/j.berh.2006.02.001

33. Lopez-Larrea C, Gonzalez-Roces S, Alvarez V. HLA-B27 structure, function, and disease association. *Curr Opin Rheumatol.* (1996) 8:296–308. doi: 10.1097/00002281-199607000-00005

34. Fiorillo MT, Meadows L, D'Amato M, Shabanowitz J, Hunt DF, Appella E, et al. Susceptibility to ankylosing spondylitis correlates with the C-terminal residue of peptides presented by various HLA-B27 subtypes. *Eur J Immunol.* (1997) 27:368–73. doi: 10.1002/eji.1830270205

35. Lopez-Larrea C, Sujirachato K, Mehra NK, Chiewsilp P, Isarangkura D, Kanga U, et al. HLA-B27 subtypes in Asian patients with ankylosing spondylitis. Evidence for new associations. *Tissue Antigens*. (1995) 45:169–76. doi: 10.1111/j.1399-0039.1995.tb02436.x

36. Khan MA. Polymorphism of HLA-B27: 105 subtypes currently known. *Curr Rheumatol Rep.* (2013) 15:362. doi: 10.1007/s11926-013-0362-y

37. Hammer RE, Maika SD, Richardson JA, Tang JP, Taurog JD. Spontaneous inflammatory disease in transgenic rats expressing HLA-B27 and human beta 2m: an animal model of HLA-B27-associated human disorders. *Cell.* (1990) 63:1099–112. doi: 10.1016/0092-8674(90)90512-D

38. Khare SD, Hansen J, Luthra HS, David CS. HLA-B27 heavy chains contribute to spontaneous inflammatory disease in B27/human beta2-microglobulin (beta2m) double transgenic mice with disrupted mouse beta2m. *J Clin Invest.* (1996) 98:2746–55. doi: 10.1172/JCI119100

39. Khare SD, Luthra HS, David CS. Spontaneous inflammatory arthritis in HLA-B27 transgenic mice lacking beta 2-microglobulin: a model of human spondyloarthropathies. *J Exp Med.* (1995) 182:1153–8. doi: 10.1084/jem.182.4.1153

40. Khare SD, Lee S, Bull MJ, Hanson J, Luthra HS, David CS. Spontaneous inflammatory disease in HLA-B27 transgenic mice does not require transporter of antigenic peptides. *Clin Immunol.* (2001) 98:364–9. doi: 10.1006/clim.2000.4984

41. Kingsbury DJ, Mear JP, Witte DP, Taurog JD, Roopenian DC, Colbert RA. Development of spontaneous arthritis in beta2-microglobulin-deficient mice without expression of HLA-B27: association with deficiency of endogenous major histocompatibility complex class I expression. *Arthritis Rheum.* (2000) 43:2290-6. doi: 10.1002/1529-0131(200010)43:10<2290::AID-ANR17>3.0.CO;2-6

42. Cauli A, Dessole G, Fiorillo MT, Vacca A, Mameli A, Bitti P, et al. Increased level of HLA-B27 expression in ankylosing spondylitis patients compared with healthy HLA-B27-positive subjects: a possible further susceptibility factor for the development of disease. *Rheumatology*. (2002) 41:1375–9. doi: 10.1093/rheumatology/41.12.1375

43. Jaakkola E, Herzberg J, Laiho K, Barnardo MC, Pointon JJ, Kauppi M, et al. Finnish HLA studies confirm the increased risk conferred by HLA-B27 homozygosity in ankylosing spondylitis. *Ann Rheum Dis.* (2006) 65:775–80. doi: 10.1136/ard.2005.041103

44. Taurog JD, Maika SD, Simmons WA, Breban M, Hammer RE. Susceptibility to inflammatory disease in HLA-B27 transgenic rat lines correlates with the level of B27 expression. *J Immunol.* (1993) 150:4168–78.

45. Feldtkeller E, Khan MA, van der Heijde D, van der Linden S, Braun J. Age at disease onset and diagnosis delay in HLA-B27 negative vs. positive patients with ankylosing spondylitis. *Rheumatol Int.* (2003) 23:61–6. doi: 10.1007/s00296-002-0237-4

46. Freeston J, Barkham N, Hensor E, Emery P, Fraser A. Ankylosing spondylitis, HLA-B27 positivity and the need for biologic therapies. *Joint Bone Spine*. (2007) 74:140–3. doi: 10.1016/j.jbspin.2006.11.003

47. Linssen A. B27+ disease versus B27- disease. Scand J Rheumatol Suppl. (1990) 87:111-8. doi: 10.3109/03009749009097070

48. Sadasivan B, Lehner PJ, Ortmann B, Spies T, Cresswell P. Roles for calreticulin and a novel glycoprotein, tapasin, in the interaction of MHC class I molecules with TAP. *Immunity*. (1996) 5:103–14. doi: 10.1016/S1074-7613(00)80487-2

49. Zhang Q, Tector M, Salter RD. Calnexin recognizes carbohydrate and protein determinants of class I major histocompatibility complex molecules. *J Biol Chem.* (1995) 270:3944–8. doi: 10.1074/jbc.270.8.3944

50. Colbert JD, Rock KL. How a tailor achieves the perfect fit. J Biol Chem. (2020) 295:7211–2. doi: 10.1074/jbc.H120.013868

51. Bollow M, Fischer T, Reisshauer H, Backhaus M, Sieper J, Hamm B, et al. Quantitative analyses of sacroiliac biopsies in spondyloarthropathies: T cells and macrophages predominate in early and active sacroilitis- cellularity correlates with the degree of enhancement detected by magnetic resonance imaging. *Ann Rheum Dis.* (2000) 59:135–40. doi: 10.1136/ard.59.2.135

52. McGonagle D, Marzo-Ortega H, O'Connor P, Gibbon W, Hawkey P, Henshaw K, et al. Histological assessment of the early enthesitis lesion in spondyloarthropathy. Ann Rheum Dis. (2002) 61:534-7. doi: 10.1136/ard.61.6.534

53. Wright PB, McEntegart A, McCarey D, McInnes IB, Siebert S, Milling SW. Ankylosing spondylitis patients display altered dendritic cell and T cell populations that implicate pathogenic roles for the IL-23 cytokine axis and intestinal inflammation. *Rheumatology.* (2016) 55:120–32. doi: 10.1093/rheumatology/kev245

54. Slobodin G, Kessel A, Kofman N, Toubi E, Rosner I, Odeh M. Phenotype of resting and activated monocyte-derived dendritic cells grown from peripheral blood of patients with ankylosing spondylitis. *Inflammation*. (2012) 35:772–5. doi: 10.1007/s10753-011-9373-x

55. Dhaenens M, Fert I, Glatigny S, Haerinck S, Poulain C, Donnadieu E, et al. Dendritic cells from spondylarthritis-prone HLA-B27-transgenic rats display altered cytoskeletal dynamics, class II major histocompatibility complex expression, and viability. *Arthritis Rheum.* (2009) 60:2622–32. doi: 10.1002/art.24780

56. Benjamin R, Parham P. Guilt by association: HLA-B27 and ankylosing spondylitis. *Immunol Today*. (1990) 11:137-42. doi: 10.1016/0167-5699(90) 90051-A

57. Atagunduz P, Appel H, Kuon W, Wu P, Thiel A, Kloetzel PM, et al. HLA-B27-restricted CD8+ T cell response to cartilage-derived self peptides in ankylosing spondylitis. *Arthritis Rheum.* (2005) 52:892–901. doi: 10.1002/art. 20948

58. Hermann E, Yu DT. Meyer zum Buschenfelde KH, Fleischer B. HLA-B27restricted CD8 T cells derived from synovial fluids of patients with reactive arthritis and ankylosing spondylitis. *Lancet.* (1993) 342:646–50. doi: 10.1016/ 0140-6736(93)91760-J

59. Hulsmeyer M, Fiorillo MT, Bettosini F, Sorrentino R, Saenger W, Ziegler A, et al. Dual, HLA-B27 subtype-dependent conformation of a self-peptide. *J Exp Med.* (2004) 199:271–81. doi: 10.1084/jem.20031690

60. Ruckert C, Fiorillo MT, Loll B, Moretti R, Biesiadka J, Saenger W, et al. Conformational dimorphism of self-peptides and molecular mimicry in a disease-associated HLA-B27 subtype. *J Biol Chem.* (2006) 281:2306–16. doi: 10. 1074/ibc.M508528200

61. Fabian H, Huser H, Narzi D, Misselwitz R, Loll B, Ziegler A, et al. HLA-B27 subtypes differentially associated with disease exhibit conformational differences in solution. *J Mol Biol.* (2008) 376:798–810. doi: 10.1016/j.jmb.2007.12.009

62. Fielder M, Pirt SJ, Tarpey I, Wilson C, Cunningham P, Ettelaie C, et al. Molecular mimicry and ankylosing spondylitis: possible role of a novel sequence in pullulanase of *Klebsiella pneumoniae*. *FEBS Lett.* (1995) 369:243–8. doi: 10.1016/0014-5793(95)00760-7

63. Lahesmaa R, Skurnik M, Toivanen P. Molecular mimicry: any role in the pathogenesis of spondyloarthropathies? *Immunol Res.* (1993) 12:193–208. doi: 10.1007/BF02918304

64. Ramos M, Alvarez I, Sesma L, Logean A, Rognan D, Lopez de Castro JA. Molecular mimicry of an HLA-B27-derived ligand of arthritis-linked subtypes with chlamydial proteins. *J Biol Chem.* (2002) 277:37573–81. doi: 10.1074/jbc. M205470200

65. May E, Dorris ML, Satumtira N, Iqbal I, Rehman MI, Lightfoot E, et al. CD8 alpha beta T cells are not essential to the pathogenesis of arthritis or colitis in HLA-B27 transgenic rats. *J Immunol.* (2003) 170:1099–105. doi: 10.4049/jimmunol.170.2.1099

66. Taurog JD, Dorris ML, Satumtira N, Tran TM, Sharma R, Dressel R, et al. Spondylarthritis in HLA-B27/human beta2-microglobulin-transgenic rats is not prevented by lack of CD8. *Arthritis Rheum*. (2009) 60:1977–84. doi: 10.1002/art. 24599

67. Boyle LH, Goodall JC, Opat SS, Gaston JS. The recognition of HLA-B27 by human CD4(+) T lymphocytes. J Immunol. (2001) 167:2619–24. doi: 10.4049/jimmunol.167.5.2619

68. Allen RL, O'Callaghan CA, McMichael AJ, Bowness P. Cutting edge: HLA-B27 can form a novel beta 2-microglobulin-free heavy chain homodimer structure. *J Immunol.* (1999) 162:5045–8.

69. Taurog JD, Maika SD, Satumtira N, Dorris ML, McLean IL, Yanagisawa H, et al. Inflammatory disease in HLA-B27 transgenic rats. *Immunol Rev.* (1999) 169:209–23. doi: 10.1111/j.1600-065X.1999.tb01317.x

70. Breban M, Fernandez-Sueiro JL, Richardson JA, Hadavand RR, Maika SD, Hammer RE, et al. T cells, but not thymic exposure to HLA-B27, are required

for the inflammatory disease of HLA-B27 transgenic rats. J Immunol. (1996) 156:794-803.

71. Glatigny S, Fert I, Blaton MA, Lories RJ, Araujo LM, Chiocchia G, et al. Proinflammatory Th17 cells are expanded and induced by dendritic cells in spondylarthritis-prone HLA-B27-transgenic rats. *Arthritis Rheum.* (2012) 64:110–20. doi: 10.1002/art.33321

72. Szanto S, Aleksza M, Mihaly E, Lakos G, Szabo Z, Vegvari A, et al. Intracytoplasmic cytokine expression and T cell subset distribution in the peripheral blood of patients with ankylosing spondylitis. *J Rheumatol.* (2008) 35:2372–5. doi: 10.3899/jrheum.070839

73. Cauli A, Shaw J, Giles J, Hatano H, Rysnik O, Payeli S, et al. The arthritisassociated HLA-B*27:05 allele forms more cell surface B27 dimer and free heavy chain ligands for KIR3DL2 than HLA-B*27:09. *Rheumatology*. (2013) 52:1952–62. doi: 10.1093/rheumatology/ket219

74. Allen RL, Raine T, Haude A, Trowsdale J, Wilson MJ. Leukocyte receptor complex-encoded immunomodulatory receptors show differing specificity for alternative HLA-B27 structures. *J Immunol.* (2001) 167:5543–7. doi: 10.4049/jimmunol.167.10.5543

75. Kollnberger S, Bird L, Sun MY, Retiere C, Braud VM, McMichael A, et al. Cell-surface expression and immune receptor recognition of HLA-B27 homodimers. *Arthritis Rheum.* (2002) 46:2972–82. doi: 10.1002/art.10605

76. Chan AT, Kollnberger SD, Wedderburn LR, Bowness P. Expansion and enhanced survival of natural killer cells expressing the killer immunoglobulinlike receptor KIR3DL2 in spondylarthritis. *Arthritis Rheum*. (2005) 52:3586–95. doi: 10.1002/art.21395

77. Payeli SK, Kollnberger S, Marroquin BO, Thiel M, McHugh K, Giles J, et al. Inhibiting HLA-B27 homodimer-driven immune cell inflammation in spondylarthritis. *Arthritis Rheum.* (2012) 64:3139–49. doi: 10.1002/art.34538

78. Bowness P, Ridley A, Shaw J, Chan AT, Wong-Baeza I, Fleming M, et al. Th17 cells expressing KIR3DL2+ and responsive to HLA-B27 homodimers are increased in ankylosing spondylitis. *J Immunol.* (2011) 186:2672–80. doi: 10. 4049/jimmunol.1002653

79. Kocaturk NM, Gozuacik D. Crosstalk between mammalian autophagy and the ubiquitin-proteasome system. *Front Cell Dev Biol.* (2018) 6:128. doi: 10. 3389/fcell.2018.00128

80. Rutkowski DT, Kaufman RJ. A trip to the ER: coping with stress. Trends Cell Biol. (2004) 14:20–8. doi: 10.1016/j.tcb.2003.11.001

81. Colbert RA, DeLay ML, Layh-Schmitt G, Sowders DP. HLA-B27 misfolding and spondyloarthropathies. *Adv Exp Med Biol.* (2009) 649:217–34. doi: 10.1007/ 978-1-4419-0298-6_16

82. Mear JP, Schreiber KL, Munz C, Zhu X, Stevanovic S, Rammensee HG, et al. Misfolding of HLA-B27 as a result of its B pocket suggests a novel mechanism for its role in susceptibility to spondyloarthropathies. *J Immunol.* (1999) 163:6665–70.

83. Burr ML, Cano F, Svobodova S, Boyle LH, Boname JM, Lehner PJ. HRD1 And UBE2J1 target misfolded MHC class I heavy chains for endoplasmic reticulum-associated degradation. *Proc Natl Acad Sci.* (2011) 108:2034–9. doi: 10.1073/pnas.1016229108

84. Guiliano DB, Fussell H, Lenart I, Tsao E, Nesbeth D, Fletcher AJ, et al. Endoplasmic reticulum degradation-enhancing alpha-mannosidase-like protein 1 targets misfolded HLA-B27 dimers for endoplasmic reticulum-associated degradation. *Arthritis Rheumatol.* (2014) 66:2976–88. doi: 10.1002/art.38809

85. Tran TM, Satumtira N, Dorris ML, May E, Wang A, Furuta E, et al. HLA-B27 in transgenic rats forms disulfide-linked heavy chain oligomers and multimers that bind to the chaperone BiP. *J Immunol.* (2004) 172:5110–9. doi: 10.4049/jimmunol.172.8.5110

86. Dangoria NS, DeLay ML, Kingsbury DJ, Mear JP, Uchanska-Ziegler B, Ziegler A, et al. HLA-B27 misfolding is associated with aberrant intermolecular disulfide bond formation (dimerization) in the endoplasmic reticulum. *J Biol Chem.* (2002) 277:23459–68. doi: 10.1074/jbc.M110336200

87. Dong W, Zhang Y, Yan M, Liu H, Chen Z, Zhu P. Upregulation of 78-kDa glucose-regulated protein in macrophages in peripheral joints of active ankylosing spondylitis. *Scand J Rheumatol.* (2008) 37:427–34. doi: 10.1080/03009740802213310

88. Gu J, Rihl M, Marker-Hermann E, Baeten D, Kuipers JG, Song YW, et al. Clues to pathogenesis of spondyloarthropathy derived from synovial fluid mononuclear cell gene expression profiles. *J Rheumatol.* (2002) 29:2159–64.

89. Turner MJ, Sowders DP, DeLay ML, Mohapatra R, Bai S, Smith JA, et al. HLA-B27 misfolding in transgenic rats is associated with activation of the unfolded protein response. *J Immunol.* (2005) 175:2438–48. doi: 10.4049/jimmunol.175.4.2438

90. Turner MJ, DeLay ML, Bai S, Klenk E, Colbert RA. HLA-B27 up-regulation causes accumulation of misfolded heavy chains and correlates with the magnitude of the unfolded protein response in transgenic rats: implications for the

pathogenesis of spondylarthritis-like disease. Arthritis Rheum. (2007) 56:215-23. doi: 10.1002/art.22295

91. Tran TM, Hong S, Edwan JH, Colbert RA. ERAP1 reduces accumulation of aberrant and disulfide-linked forms of HLA-B27 on the cell surface. *Mol Immunol.* (2016) 74:10–7. doi: 10.1016/j.molimm.2016.04.002

92. Hotamisligil GS. Endoplasmic reticulum stress and the inflammatory basis of metabolic disease. *Cell.* (2010) 140:900–17. doi: 10.1016/j.cell.2010.02.034

93. Jethwa H, Bowness P. The interleukin (IL)-23/IL-17 axis in ankylosing spondylitis: new advances and potentials for treatment. *Clin Exp Immunol.* (2016) 183:30-6. doi: 10.1111/cei.12670

94. Liu T, Li S, Ying S, Tang S, Ding Y, Li Y, et al. The IL-23/IL-17 pathway in inflammatory skin diseases: from bench to bedside. *Front Immunol.* (2020) 11:594735. doi: 10.3389/fimmu.2020.594735

95. Langrish CL, Chen Y, Blumenschein WM, Mattson J, Basham B, Sedgwick JD, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J Exp Med.* (2005) 201:233–40. doi: 10.1084/jem.20041257

96. DeLay ML, Turner MJ, Klenk EI, Smith JA, Sowders DP, Colbert RA. HLA-B27 misfolding and the unfolded protein response augment interleukin-23 production and are associated with Th17 activation in transgenic rats. *Arthritis Rheum.* (2009) 60:2633–43. doi: 10.1002/art.24763

97. Sherlock JP, Joyce-Shaikh B, Turner SP, Chao CC, Sathe M, Grein J, et al. IL-23 induces spondyloarthropathy by acting on ROR-gammat+ CD3 + CD4-CD8entheseal resident T cells. *Nat Med.* (2012) 18:1069–76. doi: 10.1038/nm.2817

98. Shen H, Goodall JC, Hill Gaston JS. Frequency and phenotype of peripheral blood Th17 cells in ankylosing spondylitis and rheumatoid arthritis. *Arthritis Rheum.* (2009) 60:1647–56. doi: 10.1002/art.24568

99. Zeng L, Lindstrom MJ, Smith JA. Ankylosing spondylitis macrophage production of higher levels of interleukin-23 in response to lipopolysaccharide without induction of a significant unfolded protein response. *Arthritis Rheum.* (2011) 63:3807–17. doi: 10.1002/art.30593

100. Armaka M, Apostolaki M, Jacques P, Kontoyiannis DL, Elewaut D, Kollias G. Mesenchymal cell targeting by TNF as a common pathogenic principle in chronic inflammatory joint and intestinal diseases. *J Exp Med.* (2008) 205:331–7. doi: 10.1084/jem.20070906

101. Chen B, Huang K, Ye L, Li Y, Zhang J, Zhang J, et al. Interleukin-37 is increased in ankylosing spondylitis patients and associated with disease activity. *J Transl Med.* (2015) 13:36. doi: 10.1186/s12967-015-0394-3

102. Prahalad S, Zeft AS, Pimentel R, Clifford B, McNally B, Mineau GP, et al. Quantification of the familial contribution to juvenile idiopathic arthritis. *Arthritis Rheum*. (2010) 62:2525–9. doi: 10.1002/art.27516

103. Brown MA, Kennedy LG, MacGregor AJ, Darke C, Duncan E, Shatford JL, et al. Susceptibility to ankylosing spondylitis in twins: the role of genes, HLA, and the environment. *Arthritis Rheum.* (1997) 40:1823–8. doi: 10.1002/art.1780401015

104. Pedersen OB, Svendsen AJ, Ejstrup L, Skytthe A, Harris JR, Junker P. Ankylosing spondylitis in Danish and Norwegian twins: occurrence and the relative importance of genetic vs. environmental effectors in disease causation. *Scand J Rheumatol.* (2008) 37:120–6. doi: 10.1080/03009740701824613

105. Brown MA, Laval SH, Brophy S, Calin A. Recurrence risk modelling of the genetic susceptibility to ankylosing spondylitis. *Ann Rheum Dis.* (2000) 59:883–6. doi: 10.1136/ard.59.11.883

106. Carter N, Williamson L, Kennedy LG, Brown MA, Wordsworth BP. Susceptibility to ankylosing spondylitis. *Rheumatology*. (2000) 39:445. doi: 10. 1093/rheumatology/39.4.445

107. Stolwijk C, van Onna M, Boonen A, van Tubergen A. Global prevalence of spondyloarthritis: a systematic review and meta-regression analysis. *Arthritis Care Res.* (2016) 68:1320–31. doi: 10.1002/acr.22831

108. Jarvinen P. Occurrence of ankylosing spondylitis in a nationwide series of twins. Arthritis Rheum. (1995) 38:381–3. doi: 10.1002/art.1780380313

109. Zhu W, He X, Cheng K, Zhang L, Chen D, Wang X, et al. Ankylosing spondylitis: etiology, pathogenesis, and treatments. *Bone Res.* (2019) 7:22. doi: 10.1038/s41413-019-0057-8

110. Burton PR, Clayton DG, Cardon LR, Craddock N, Deloukas P, Duncanson A, et al. Association scan of 14,500 nonsynonymous SNPs in four diseases identifies autoimmunity variants. *Nat Genet.* (2007) 39:1329–37. doi: 10.1038/ng.2007.17

111. Hinks A, Martin P, Flynn E, Eyre S, Packham J, Barton A, et al. Subtype specific genetic associations for juvenile idiopathic arthritis: ERAP1 with the enthesitis related arthritis subtype and IL23R with juvenile psoriatic arthritis. *Arthritis Res Ther.* (2011) 13:R12. doi: 10.1186/ar3235

112. Evans DM, Spencer CC, Pointon JJ, Su Z, Harvey D, Kochan G, et al. Interaction between ERAP1 and HLA-B27 in ankylosing spondylitis implicates peptide handling in the mechanism for HLA-B27 in disease susceptibility. *Nat Genet.* (2011) 43:761–7. doi: 10.1038/ng.873

113. Collins EJ, Garboczi DN, Wiley DC. Three-dimensional structure of a peptide extending from one end of a class I MHC binding site. *Nature.* (1994) 371:626–9. doi: 10.1038/371626a0

114. Davidson SI, Liu Y, Danoy PA, Wu X, Thomas GP, Jiang L, et al. Association of STAT3 and TNFRSF1A with ankylosing spondylitis in Han Chinese. *Ann Rheum Dis.* (2011) 70:289–92. doi: 10.1136/ard.2010.133322

115. Zinovieva E, Bourgain C, Kadi A, Letourneur F, Izac B, Said-Nahal R, et al. Comprehensive linkage and association analyses identify haplotype, near to the TNFSF15 gene, significantly associated with spondyloarthritis. *PLoS Genet.* (2009) 5:e1000528. doi: 10.1371/journal.pgen.1000528

116. Pointon JJ, Harvey D, Karaderi T, Appleton LH, Farrar C, Stone MA, et al. The chromosome 16q region associated with ankylosing spondylitis includes the candidate gene tumour necrosis factor receptor type 1-associated death domain (TRADD). *Ann Rheum Dis.* (2010) 69:1243–6. doi: 10.1136/ard.2009.115147

117. Baeten D, Breban M, Lories R, Schett G, Sieper J. Are spondylarthritides related but distinct conditions or a single disease with a heterogeneous phenotype? *Arthritis Rheum.* (2013) 65:12–20. doi: 10.1002/art.37829

118. Ellinghaus D, Jostins L, Spain SL, Cortes A, Bethune J, Han B, et al. Analysis of five chronic inflammatory diseases identifies 27 new associations and highlights disease-specific patterns at shared loci. *Nat Genet.* (2016) 48:510–8. doi: 10.1038/ng.3528

119. Yang X, Li M, Wang L, Hu Z, Zhang Y, Yang Q. Association of KIF21B genetic polymorphisms with ankylosing spondylitis in a Chinese Han population of Shandong Province. *Clin Rheumatol.* (2015) 34:1729–36. doi: 10. 1007/s10067-014-2761-5

120. Ou Y. Anthrax toxin receptor 2 gene (ANTXR2) rs4333130 is associated with ankylosing spondylitis. Int J Clin Exp Med. (2015) 8:7679–83.

121. Lin Z, Bei JX, Shen M, Li Q, Liao Z, Zhang Y, et al. A genome-wide association study in Han Chinese identifies new susceptibility loci for ankylosing spondylitis. *Nat Genet.* (2011) 44:73–7. doi: 10.1038/ng.1005

122. Thompson SD, Sudman M, Ramos PS, Marion MC, Ryan M, Tsoras M, et al. The susceptibility loci juvenile idiopathic arthritis shares with other autoimmune diseases extend to PTPN2, COG6, and ANGPT1. Arthritis Rheum. (2010) 62:3265-76. doi: 10.1002/art.27688

123. Thompson SD, Marion MC, Sudman M, Ryan M, Tsoras M, Howard TD, et al. Genome-wide association analysis of juvenile idiopathic arthritis identifies a new susceptibility locus at chromosomal region 3q13. *Arthritis Rheum.* (2012) 64:2781–91. doi: 10.1002/art.34429

124. Behrens EM, Finkel TH, Bradfield JP, Kim CE, Linton L, Casalunovo T, et al. Association of the TRAF1-C5 locus on chromosome 9 with juvenile idiopathic arthritis. *Arthritis Rheum.* (2008) 58:2206–7. doi: 10.1002/art.23603

125. Hinks A, Barton A, Shephard N, Eyre S, Bowes J, Cargill M, et al. Identification of a novel susceptibility locus for juvenile idiopathic arthritis by genome-wide association analysis. *Arthritis Rheum.* (2009) 60:258–63. doi: 10.1002/art.24179

126. Hinks A, Cobb J, Marion MC, Prahalad S, Sudman M, Bowes J, et al. Dense genotyping of immune-related disease regions identifies 14 new susceptibility loci for juvenile idiopathic arthritis. *Nat Genet.* (2013) 45:664–9. doi: 10.1038/ng.2614

127. Albers HM, Kurreeman FA, Houwing-Duistermaat JJ, Brinkman DM, Kamphuis SS, Girschick HJ, et al. The TRAF1/C5 region is a risk factor for polyarthritis in juvenile idiopathic arthritis. *Ann Rheum Dis.* (2008) 67:1578–80. doi: 10.1136/ard.2008.089060

128. Mistry RR, Patro P, Agarwal V, Misra DP. Enthesitis-related arthritis: current perspectives. *Open Access Rheumatol.* (2019) 11:19–31. doi: 10.2147/OARRR.S163677

129. Rusman T, van Bentum RE, van der Horst-Bruinsma IE. Sex and gender differences in axial spondyloarthritis: myths and truths. *Rheumatology.* (2020) 59:iv38–46. doi: 10.1093/rheumatology/keaa543

130. Fish EN. The X-files in immunity: sex-based differences predispose immune responses. Nat Rev Immunol. (2008) 8:737-44. doi: 10.1038/nri2394

131. Jimenez-Balderas FJ, Tapia-Serrano R, Madero-Cervera JI, Murrieta S, Mintz G. Ovarian function studies in active ankylosing spondylitis in women. Clinical response to estrogen therapy. *J Rheumatol.* (1990) 17:497–502.

132. Jeong H, Bae EK, Kim H, Eun YH, Kim IY, Kim H, et al. Estrogen attenuates the spondyloarthritis manifestations of the SKG arthritis model. *Arthritis Res Ther.* (2017) 19:198. doi: 10.1186/s13075-017-1407-9

133. Mahendira D, Thavaneswaran A, Carty A, Haroon N, Anton A, Passalent L, et al. Analysis of the effect of the oral contraceptive pill on clinical outcomes in

women with ankylosing spondylitis. J Rheumatol. (2014) 41:1344-8. doi: 10.3899/ jrheum.130996

134. Giltay EJ, Popp-Snijders C, van Schaardenburg D, Dekker-Saeys BJ, Gooren LJ, Dijkmans BA. Serum testosterone levels are not elevated in patients with ankylosing spondylitis. *J Rheumatol.* (1998) 25:2389–94.

135. Yoon K, Kim N. Roles of sex hormones and gender in the gut microbiota. J Neurogastroenterol Motil. (2021) 27:314–25. doi: 10.5056/jnm20208

136. Cutolo M, Sulli A, Capellino S, Villaggio B, Montagna P, Seriolo B, et al. Sex hormones influence on the immune system: basic and clinical aspects in autoimmunity. *Lupus*. (2004) 13:635–8. doi: 10.1191/0961203304lu1094oa

137. Molodecky NA, Soon IS, Rabi DM, Ghali WA, Ferris M, Chernoff G, et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology.* (2012) 142:46–54. doi: 10.1053/j. gastro.2011.10.001

138. Mielants H, Veys EM. Inflammation of the ileum in patients with B27-positive reactive arthritis. *Lancet.* (1984) 1:288. doi: 10.1016/S0140-6736(84)90163-6

139. Gill T, Asquith M, Brooks SR, Rosenbaum JT, Colbert RA. Effects of HLA-B27 on gut microbiota in experimental spondyloarthritis implicate an ecological model of dysbiosis. *Arthritis Rheumatol.* (2018) 70:555–65. doi: 10.1002/art.40405

140. Baeten D, Demetter P, Cuvelier CA, Kruithof E, Van DN, De VM, et al. Macrophages expressing the scavenger receptor CD163: a link between immune alterations of the gut and synovial inflammation in spondyloarthropathy. *J Pathol.* (2002) 196:343–50. doi: 10.1002/path.1044

141. Rath HC, Schultz M, Freitag R, Dieleman LA, Li F, Linde HJ, et al. Different subsets of enteric bacteria induce and perpetuate experimental colitis in rats and mice. *Infect Immun.* (2001) 69:2277–85. doi: 10.1128/IAI.69.4.2277-2285.2001

142. Taurog JD, Richardson JA, Croft JT, Simmons WA, Zhou M, Fernandez-Sueiro JL, et al. The germfree state prevents development of gut and joint inflammatory disease in HLA-B27 transgenic rats. *J Exp Med.* (1994) 180:2359–64. doi: 10.1084/jem.180.6.2359

143. Costello ME, Ciccia F, Willner D, Warrington N, Robinson PC, Gardiner B, et al. Brief report: intestinal dysbiosis in ankylosing spondylitis. *Arthritis Rheumatol.* (2015) 67:686–91. doi: 10.1002/art.38967

144. Stoll ML, Kumar R, Morrow CD, Lefkowitz EJ, Cui X, Genin A, et al. Altered microbiota associated with abnormal humoral immune responses to commensal organisms in enthesitis-related arthritis. *Arthritis Res Ther.* (2014) 16:486. doi: 10.1186/s13075-014-0486-0

145. Stoll ML, Weiss PF, Weiss JE, Nigrovic PA, Edelheit BS, Bridges Jr. SL, et al. Age and fecal microbial strain-specific differences in patients with spondyloarthritis. *Arthritis Res Ther.* (2018) 20:14. doi: 10.1186/s13075-018-1510-6

146. Ciccia F, Guggino G, Rizzo A, Alessandro R, Luchetti MM, Milling S, et al. Dysbiosis and zonulin upregulation alter gut epithelial and vascular barriers in patients with ankylosing spondylitis. *Ann Rheum Dis.* (2017) 76:1123–32. doi: 10.1136/annrheumdis-2016-210000

147. Asquith MJ, Stauffer P, Davin S, Mitchell C, Lin P, Rosenbaum JT. Perturbed mucosal immunity and dysbiosis accompany clinical disease in a rat model of spondyloarthritis. *Arthritis Rheumatol.* (2016) 68:2151–62. doi: 10. 1002/art.39681

148. Chai JN, Zhou YW, Hsieh CS. T cells and intestinal commensal bacteriaignorance, rejection, and acceptance. *FEBS Lett.* (2014) 588:4167–75. doi: 10.1016/ j.febslet.2014.06.040

149. Dieleman LA, Hoentjen F, Qian BF, Sprengers D, Tjwa E, Torres MF, et al. Reduced ratio of protective versus proinflammatory cytokine responses to commensal bacteria in HLA-B27 transgenic rats. *Clin Exp Immunol.* (2004) 136:30–9. doi: 10.1111/j.1365-2249.2004.02410.x

150. Qian BF, Tonkonogy SL, Balfour SR. Luminal bacterial antigen-specific CD4+ T-cell responses in HLA-B27 transgenic rats with chronic colitis are mediated by both major histocompatibility class II and HLA-B27 molecules. *Immunology.* (2006) 117:319–28. doi: 10.1111/j.1365-2567.2005.02303.x

151. Hacquard-Bouder C, Chimenti MS, Giquel B, Donnadieu E, Fert I, Schmitt A, et al. Alteration of antigen-independent immunologic synapse formation between dendritic cells from HLA-B27-transgenic rats and CD4+ T cells: selective impairment of costimulatory molecule engagement by mature HLA-B27. *Arthritis Rheum.* (2007) 56:1478–89. doi: 10.1002/art.22572

152. Talpin A, Costantino F, Bonilla N, Leboime A, Letourneur F, Jacques S, et al. Monocyte-derived dendritic cells from HLA-B27+ axial spondyloarthritis (SpA) patients display altered functional capacity and deregulated gene expression. *Arthritis Res Ther.* (2014) 16:417. doi: 10.1186/ s13075-014-0417-0

153. Bazin T, Hooks KB, Barnetche T, Truchetet ME, Enaud R, Richez C, et al. Microbiota composition may predict anti-Tnf alpha response in spondyloarthritis patients: an exploratory study. Sci Rep. (2018) 8:5446. doi: 10.1038/s41598-018-23571-4

154. Haugen M, Kjeldsen-Kragh J, Nordvag BY, Forre O. Diet and disease symptoms in rheumatic diseases-results of a questionnaire based survey. *Clin Rheumatol.* (1991) 10:401-7. doi: 10.1007/BF02206660

155. Macfarlane TV, Abbood HM, Pathan E, Gordon K, Hinz J, Macfarlane GJ. Relationship between diet and ankylosing spondylitis: a systematic review. *Eur J Rheumatol.* (2018) 5:45–52. doi: 10.5152/eurjrheum.2017.16103

156. Rashid T, Wilson C, Ebringer A. Raised incidence of ankylosing spondylitis among Inuit populations could be due to high HLA-B27 association and starch consumption. *Rheumatol Int.* (2015) 35:945–51. doi: 10.1007/s00296-014-3164-2

157. Sundstrom B, Johansson G, Johansson I, Wallberg-Jonsson S. Modifiable cardiovascular risk factors in patients with ankylosing spondylitis. *Clin Rheumatol.* (2014) 33:111-7. doi: 10.1007/s10067-013-2410-4

158. Appelboom T, Durez P. Effect of milk product deprivation on spondyloarthropathy. Ann Rheum Dis. (1994) 53:481–2. doi: 10.1136/ard.53.7.481

159. Sanges M, Valente G, Rea M, Della GR, De FG, Sollazzo R, et al. Probiotics in spondyloarthropathy associated with ulcerative colitis: a pilot study. *Eur Rev Med Pharmacol Sci.* (2009) 13:233–4.

160. Sanchez P, Letarouilly JG, Nguyen Y, Sigaux J, Barnetche T, Czernichow S, et al. Efficacy of probiotics in rheumatoid arthritis and spondyloarthritis: a systematic review and meta-analysis of randomized controlled trials. *Nutrients*. (2022) 14:354. doi: 10.3390/nu14020354

161. Hoentjen F, Welling GW, Harmsen HJ, Zhang X, Snart J, Tannock GW, et al. Reduction of colitis by prebiotics in HLA-B27 transgenic rats is associated with microflora changes and immunomodulation. *Inflamm Bowel Dis.* (2005) 11:977–85. doi: 10.1097/01.MIB.0000183421.02316.d5

162. Koleva P, Ketabi A, Valcheva R, Ganzle MG, Dieleman LA. Chemically defined diet alters the protective properties of fructo-oligosaccharides and isomalto-oligosaccharides in HLA-B27 transgenic rats. *PLoS One.* (2014) 9: e111717. doi: 10.1371/journal.pone.0111717

163. Koleva PT, Valcheva RS, Sun X, Ganzle MG, Dieleman LA. Inulin and fructo-oligosaccharides have divergent effects on colitis and commensal microbiota in HLA-B27 transgenic rats. *Br J Nutr.* (2012) 108:1633–43. doi: 10. 1017/S0007114511007203

164. Asquith M, Davin S, Stauffer P, Michell C, Janowitz C, Lin P, et al. Intestinal metabolites are profoundly altered in the context of HLA-B27 expression and functionally modulate disease in a rat model of spondyloarthritis. *Arthritis Rheumatol.* (2017) 69:1984–95. doi: 10.1002/art.40183

165. Ringold S, Angeles-Han ST, Beukelman T, Lovell D, Cuello CA, Becker ML, et al. 2019 American College of Rheumatology/Arthritis Foundation Guideline for the treatment of juvenile idiopathic arthritis: therapeutic approaches for non-systemic polyarthritis, sacrollitis, and enthesitis. *Arthritis Rheumatol.* (2019) 71:846–63. doi: 10.1002/art.40884

166. Smith JA, Burgos-Vargas R. Outcomes in juvenile-onset spondyloarthritis. *Front Med.* (2021) 8:680916. doi: 10.3389/fmed.2021.680916

167. Jani M, Barton A, Warren RB, Griffiths CE, Chinoy H. The role of DMARDs in reducing the immunogenicity of TNF inhibitors in chronic inflammatory diseases. *Rheumatology*. (2014) 53:213–22. doi: 10.1093/rheumatology/ket260

168. Burgos-Vargas R, Tse SM, Horneff G, Pangan AL, Kalabic J, Goss S, et al. A randomized, double-blind, placebo-controlled multicenter study of Adalimumab in pediatric patients with enthesitis-related arthritis. *Arthritis Care Res.* (2015) 67:1503–12. doi: 10.1002/acr.22657

169. Foeldvari I, Constantin T, Vojinovic J, Horneff G, Chasnyk V, Dehoorne J, et al. Etanercept treatment for extended oligoarticular juvenile idiopathic arthritis, enthesitis-related arthritis, or psoriatic arthritis: 6-year efficacy and safety data from an open-label trial. *Arthritis Res Ther.* (2019) 21:125. doi: 10.1186/s13075-019-1916-9

170. Brunner HI, Foeldvari I, Alexeeva E, Ayaz NA, Calvo P I, Kasapcopur O, et al. Secukinumab in enthesitis-related arthritis and juvenile psoriatic arthritis: a

randomised, double-blind, placebo-controlled, treatment withdrawal, phase 3 trial. Ann Rheum Dis. (2022). doi: 10.1136/ard-2022-222849

171. Bindu S, Mazumder S, Bandyopadhyay U. Non-steroidal antiinflammatory drugs (NSAIDs) and organ damage: a current perspective. *Biochem Pharmacol.* (2020) 180:114147. doi: 10.1016/j.bcp.2020.114147

172. Consolaro A, Ruperto N, Bazso A, Pistorio A, Magni-Manzoni S, Filocamo G, et al. Development and validation of a composite disease activity score for juvenile idiopathic arthritis. *Arthritis Rheum.* (2009) 61:658–66. doi: 10.1002/art. 24516

173. Zochling J. Measures of symptoms and disease status in ankylosing spondylitis: Ankylosing Spondylitis Disease Activity Score (ASDAS), Ankylosing Spondylitis Quality of Life Scale (ASQoL), Bath Ankylosing Spondylitis Disease Activity Index (BASDAI), Bath Ankylosing Spondylitis Flobal Score (BAS-G), Bath Ankylosing Spondylitis Global Score (BAS-G), Bath Ankylosing Spondylitis Metrology Index (BASMI), Dougados Functional Index (DFI), and Health Assessment Questionnaire for the Spondylarthropathies (HAQ-S). Arthritis Care Res. (2011) 63(Suppl 11):S47–58. doi: 10.1002/acr.20575

174. Weiss PF, Colbert RA, Xiao R, Feudtner C, Beukelman T, DeWitt EM, et al. Development and retrospective validation of the juvenile spondyloarthritis disease activity index. *Arthritis Care Res.* (2014) 66:1775–82. doi: 10.1002/acr.22411

175. Rudwaleit M, Listing J, Brandt J, Braun J, Sieper J. Prediction of a major clinical response (BASDAI 50) to tumour necrosis factor alpha blockers in ankylosing spondylitis. *Ann Rheum Dis.* (2004) 63:665–70. doi: 10.1136/ard. 2003.016386

176. Perez-Guijo V, Santos-Lacomba M, Sanchez-Hernandez M, Castro-Villegas MC, Gallardo-Galera JM, Collantes-Estevez E. Tumour necrosis factor-alpha levels in aqueous humour and serum from patients with uveitis: the involvement of HLA-B27. *Curr Med Res Opin*. (2004) 20:155–7. doi: 10.1185/030079903125002847

177. Ward MM, Deodhar A, Gensler LS, Dubreuil M, Yu D, Khan MA, et al. 2019 Update of the American college of Rheumatology/Spondylitis Association of America/spondyloarthritis research and treatment network recommendations for the treatment of ankylosing spondylitis and nonradiographic axial spondyloarthritis. *Arthritis Rheumatol.* (2019) 71:1599–613. doi: 10.1002/art.41042

178. van der Heijde D, Song IH, Pangan AL, Deodhar A, van den Bosch F, Maksymowych WP, et al. Efficacy and safety of upadacitinib in patients with active ankylosing spondylitis (SELECT-AXIS 1): a multicentre, randomised, double-blind, placebo-controlled, phase 2/3 trial. *Lancet.* (2019) 394:2108–17. doi: 10.1016/S0140-6736(19)32534-6

179. van der Heijde D, Gensler LS, Deodhar A, Baraliakos X, Poddubnyy D, Kivitz A, et al. Dual neutralisation of interleukin-17A and interleukin-17F with bimekizumab in patients with active ankylosing spondylitis: results from a 48-week phase IIb, randomised, double-blind, placebo-controlled, doseranging study. *Ann Rheum Dis.* (2020) 79:595–604. doi: 10.1136/annrheumdis-2020-216980

180. Wei JC, Kim TH, Kishimoto M, Ogusu N, Jeong H, Kobayashi S. Efficacy and safety of brodalumab, an anti-IL17RA monoclonal antibody, in patients with axial spondyloarthritis: 16-week results from a randomised, placebo-controlled, phase 3 trial. *Ann Rheum Dis.* (2021) 80:1014–21. doi: 10.1136/annrheumdis-2020-219406

181. Yang K, Oak ASW, Elewski BE. Use of IL-23 inhibitors for the treatment of plaque psoriasis and psoriatic arthritis: a comprehensive review. *Am J Clin Dermatol.* (2021) 22:173–92. doi: 10.1007/s40257-020-00578-0

182. Baeten D, Ostergaard M, Wei JC, Sieper J, Jarvinen P, Tam LS, et al. Risankizumab, an IL-23 inhibitor, for ankylosing spondylitis: results of a randomised, double-blind, placebo-controlled, proof-of-concept, dose-finding phase 2 study. *Ann Rheum Dis.* (2018) 77:1295–302. doi: 10.1136/annrheumdis-2018-213328

183. Deodhar A, Gensler LS, Sieper J, Clark M, Calderon C, Wang Y, et al. Three multicenter, randomized, double-blind, placebo-controlled studies evaluating the efficacy and safety of ustekinumab in axial spondyloarthritis. *Arthritis Rheumatol.* (2019) 71:258–70. doi: 10.1002/art.40728