Check for updates

OPEN ACCESS

EDITED BY David Warburton, Saban Research Institute, United States

REVIEWED BY Shubhada Bopegamage, Slovak Medical University, Slovakia Katja Wolthers, University of Amsterdam, Netherlands

*CORRESPONDENCE Haider Al-Hello ⊠ haider.al-hello@thl.fi

RECEIVED 17 January 2024 ACCEPTED 15 March 2024 PUBLISHED 25 March 2024

CITATION

Al-Hello H, Blomqvist S and Savolainen-Kopra C (2024) Commentary: Risk factors and early markers for echovirus type 11 associated haemorrhage-hepatitis syndrome in neonates, a retrospective cohort study. Front. Pediatr. 12:1338097. doi: 10.3389/fped.2024.1338097

COPYRIGHT

© 2024 Al-Hello, Blomqvist and Savolainen-Kopra. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Commentary: Risk factors and early markers for echovirus type 11 associated haemorrhagehepatitis syndrome in neonates, a retrospective cohort study

Haider Al-Hello^{1,2*}, Soile Blomqvist¹ and Carita Savolainen-Kopra¹

¹Expert Microbiology Unit, Department of Health Security, National Institute for Health and Welfare, Helsinki, Finland, ²College of Health and Medical Technologies, National University of Science and Technology, Thi-Qar, Iraq

KEYWORDS

mutations, RNA-dependant RNA polymerase, echovirus 11, enterovirus, fidelity

A Commentary on

Risk factors and early markers for echovirus type 11 associated haemorrhage-hepatitis syndrome in neonates, a retrospective cohort study

By Wang P, Xu Y, Liu M, Li H, Wang H, Liu Y, Wang B, Xia S, Su H, Wei M, Tao L, Chen X, Lu B, Gu X, Lyu H, Zhou W, Zhang H, Gong S (2023). Front. Pediatr. 11:1063558. doi: 10.3389/fped. 2023.1063558

Dear Editors

We have read with great interest the publication by Wang et al. (1), which reports on haemorrage-hepatitis syndrome in neonates caused by Echovirus 11 (E-11). We appreciate the article's suggestions on the diagnosis and treatment of haemorrage-hepatitis syndrome but wonder if the data and conclusions regarding the similarity of the E-11 strain isolated in 2019 GWCMC01/GZ/CHN/2019 and E-11 D207 (GenBank accession EF634316), isolated in 2002 and not in 2007, as stated in the publication above, should need further study. The reasons are as follows:

Firstly, the RNA-dependent RNA polymerase of the picornavirus family has low replication fidelity, and the absence of proofreading mechanism leads to genetic mutations. The mutation rate for RNA viruses is 10^{-3} to 10^{-5} per 10 kb of the RNA genome, which is equivalent to 0.1–10 mutations (for example, the echovirus 11 may have a mutation for each transcript). The mutation rate of polioviruses, based on several studies, is approximately 3×10^{-2} mutations/synonymous site/year in the gene encoding viral protein 1 (VP1) (2). As a result of this high error rate, each time the genome is replicated, new variants arise that produce heterogeneous virus progeny referred to as "quasispecies" (3–5). High mutation frequencies allow these viruses to rapidly adapt to changing environments. According to information obtained from GenBank, the two strains carry only four nucleotide changes throughout the genome. These mutations are located in the 5' untranslated region (5'UTR) of the genome and thus the entire capsid coding region as well as non-structural proteins coding region are identical. The high similarity between the two virus strains is challenging to

understand, considering that the time between the first and the second isolations was 18 years, and the geographical area was completely different.

Secondly, according to our article published in 2007, the E-11 D207 (EF634316) is pancreatropic. This means that E-11 D207 tends to replicate in the pancreatic islets of Langerhans, which was documented in association with diabetes in a Slovakian child. To gain new phenotypic features, the virus is supposed to acquire a new genetic characteristic represented by nucleotide mutations and amino acid substitutions. In the case of the publication Al-Hello et al. (6), the pancreatropic E-11/D207 was found to be closely related to a specific subgroup B of E-11 strains known to cause uveitis. The strains are E-11/Kust/86, E-11/Kar/87 and E11/Kh3/97 published by Lukashev et al. (7, 8). The first two strains were causative agents of uveitis outbreaks in Siberia in 1986 and 1987, while the third virus was an occasional isolate in the Russian Far East in 1997. The genetic similarity between uveitis strains and E-11 D207 (EF634316) ranges from 90% to 91%. Although they were isolated in Russia and Siberia over a similar or shorter period of time as compared to the period of time between E-11 D207 (EF634316) and GWCMC01/ GZ/CHN/2019, the genetic similarity is much less.

Thirdly, in enterovirus species B (EV-B) recombination and mutations caused by the error-prone polymerase lacking proofreading machinery have been recognized as the main mechanisms of evolution (2, 4). EV-B, to which echovirus 11 belongs, is the most abundant species of enterovirus. Recombination events have been detected mostly in species B (9). Comparison of phylogenetic trees from different genomic regions has revealed recombination events in several types, such as E-7, E-30, E-11, E-9, and CV-B (10–14). In addition, interspecies recombination has also been observed, such as what occurs in the 5'UTR in EV-A and EV-B (15, 16). This indicates that the process of recombination among enteroviruses is abundant in nature, and for this reason, the high similarity between the two strains with long time period between them is difficult, if not impossible, to understand.

For the above mentioned reasons, we would like to draw your attention to the fact that this high similarity between the two strains is highly unlikely. Also, we would like to inform you that GWCMC01/GZ/CHN/2019 virus strain has been used in several

References

1. Wang P, Xu Y, Liu M, Li H, Wang H, Liu Y, et al. Risk factors and early markers for echovirus type 11 associated haemorrhage-hepatitis syndrome in neonates, a retrospective cohort study. *Front Pediatr.* (2023) 11:1063558. doi: 10.3389/fped.2023. 1063558

2. Savolainen-Kopra C, Blomqvist S. Mechanisms of genetic variation in polioviruses. Rev Med Virol. (2010) 20(6):358-71. doi: 10.1002/rmv.663

3. Domingo E, Holland JJ. RNA Virus mutations and fitness for survival. Annu Rev Microbiol. (1997) 51:151–78. doi: 10.1146/annurev.micro.51.1.151

4. Domingo E, Perales C. Viral quasispecies. PLoS Genet. (2019) 15(10):e1008271. doi: 10.1371/journal.pgen.1008271

5. Domingo E, Martin V, Perales C, Escarmis C. Coxsackieviruses and quasispecies theory: evolution of enteroviruses. *Curr Top Microbiol Immunol.* (2008) 323:3–32. doi: 10.1007/978-3-540-75546-3_1

studies (17–19), which raises a concern about the results of those and potential new studies, if genomic data is used to draw conclusions. It is critical to note that we approached this issue from a virological perspective, and we do not take a stance on the other clinical aspects and patient care described in the paper. However, for E-11 inducing diabetes or hemorrhage-hepatitis, it is necessary to improve prevention, facilitate treatment and avoid further spread.

Sincerely,

Haider Al-Hello, Soile Blomqvist and Carita Savolainen-Kopra

Author contributions

HA-H: Supervision, Writing – original draft, Writing – review & editing. SB: Writing – review & editing, Supervision. CS-K: Writing – review & editing, Supervision.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

6. Al-Hello H, Paananen A, Eskelinen M, Ylipaasto P, Hovi T, Salmela K, et al. An enterovirus strain isolated from diabetic child belongs to a genetic subcluster of echovirus 11, but is also neutralised with monotypic antisera to coxsackievirus A9. *J Gen Virol.* (2008) 89(Pt 8):1949–59. doi: 10.1099/vir.0.83474-0

7. Lukashev AN, Lashkevich VA, Koroleva GA, Karganova GG. Phylogenetic and serological characterization of echovirus 11 and echovirus 19 strains causing uveitis. *Arch Virol.* (2002) 147(1):131-42. doi: 10.1007/s705-002-8307-2

8. Lukashev AN, Lashkevich VA, Koroleva GA, Ilonen J, Karganova GG, Reznik VI, et al. Molecular epidemiology of enteroviruses causing uveitis and multisystem hemorrhagic disease of infants. *Virology.* (2003) 307(1):45–53. doi: 10.1016/s0042-6822(02)00058-2

^{9.} Simmonds P, Welch J. Frequency and dynamics of recombination within different species of human enteroviruses. *J Virol.* (2006) 80(1):483–93. doi: 10.1128/JVI.80.1. 483-493.2006

10. Oprisan G, Combiescu M, Guillot S, Caro V, Combiescu A, Delpeyroux F, et al. Natural genetic recombination between co-circulating heterotypic enteroviruses. *J Gen Virol.* (2002) 83(Pt 9):2193–200. doi: 10.1099/0022-1317-83-9-2193

11. Lukashev AN, Lashkevich VA, Ivanova OE, Koroleva GA, Hinkkanen AE, Ilonen J. Recombination in circulating enteroviruses. J Virol. (2003) 77 (19):10423–31. doi: 10.1128/jvi.77.19.10423-10431.2003

12. Kottaridi C, Bolanaki E, Kyriakopoulou Z, Dedepsidis E, Pratti A, Markoulatos P. Possible recombination and gene adaptation exchanges among clinical echovirus strains: crossing the temporal and topological barriers. *Diagn Microbiol Infect Dis.* (2007) 58(4):407–12. doi: 10.1016/j.diagmicrobio. 2007.03.007

13. Bolanaki E, Kottaridi C, Markoulatos P, Kyriakopoulou Z, Margaritis L, Katsorchis T. Partial 3D gene sequences of coxsackie viruses reveal interspecies exchanges. *Virus Genes.* (2007) 35(2):129-40. doi: 10.1007/s11262-007-0083-2

14. Nikolaidis M, Mimouli K, Kyriakopoulou Z, Tsimpidis M, Tsakogiannis D, Markoulatos P, et al. Large-scale genomic analysis reveals recurrent patterns of

intertypic recombination in human enteroviruses. Virology. (2019) 526:72-80. doi: 10.1016/j.virol.2018.10.006

15. Santti J, Hyypiä T, Kinnunen L, Salminen M. Evidence of recombination among enteroviruses. J Virol. (1999) 73(10):8741–9. doi: 10.1128/JVI.73.10.8741-8749.1999

16. Kyriakopoulou Z, Pliaka V, Amoutzias GD, Markoulatos P. Recombination among human non-polio enteroviruses: implications for epidemiology and evolution. *Virus Genes.* (2015) 50(2):177-88. doi: 10.1007/s11262-014-1152-y

17. Zhu G, Wu C, Wang Q, Deng D, Lin B, Hu X, et al. Antiviral activity of the HSP90 inhibitor VER-50589 against enterovirus 71. *Antiviral Res.* (2023) 211:105553. doi: 10.1016/j.antiviral.2023.105553

18. Wang C, Yang R, Yang F, Han Y, Ren Y, Xiong X, et al. Echovirus 11 infection induces pyroptotic cell death by facilitating NLRP3 inflammasome activation. *PLoS Pathog.* (2022) 18(8):e1010787. doi: 10.1371/journal.ppat.1010787

19. Quan J, Zhang X, Ding Y, Li S, Qiu Y, Wang R, et al. Cucurbit[7]uril as a broad-Spectrum antiviral agent against diverse RNA viruses. *Virol Sin.* (2021) 36(5):1165–76. doi: 10.1007/s12250-021-00404-9