



# Opinion: An Existing Drug to Assess *In Vivo* for Potential Adjunctive Therapy of Ebola Virus Disease and Post-Ebola Syndrome

Katherine Hendricks<sup>1\*</sup>, Meredith Gilman Parrado<sup>1</sup> and John Bradley<sup>2</sup>

<sup>1</sup> Independent researcher, Atlanta, GA, United States, <sup>2</sup> Division of Infectious Diseases, Department of Pediatrics, UCSD School of Medicine, San Diego, CA, United States

## OPEN ACCESS

### Edited by:

Salvatore Salomone,  
University of Catania, Italy

### Reviewed by:

Sina Bavari,  
United States Army Medical Research  
Institute of Infectious Diseases  
(USAMRIID), United States

### \*Correspondence:

Katherine Hendricks  
Kate@HealthKAP.com

### Specialty section:

This article was submitted to  
Experimental Pharmacology  
and Drug Discovery,  
a section of the journal  
*Frontiers in Pharmacology*

Received: 31 October 2019

Accepted: 24 December 2019

Published: 30 January 2020

### Citation:

Hendricks K, Parrado MG and  
Bradley J (2020) Opinion: An Existing  
Drug to Assess *In Vivo* for Potential  
Adjunctive Therapy of Ebola Virus  
Disease and Post-Ebola Syndrome.  
*Front. Pharmacol.* 10:1691.  
doi: 10.3389/fphar.2019.01691

**Keywords:** Ebola virus disease, minocycline, drug discovery, cytokine, chemokine, hepatoprotection

We currently have no approved drugs for the treatment of Ebola virus disease (EVD) or post-Ebola syndrome (PES). A substantial proportion of patients presenting for treatment die, including healthcare workers (HCWs) with hospital-acquired infections. More than 28,000 people were suspected or confirmed with EVD and 11,000 died in the West Africa outbreak during 2014–2016. A new EVD epicenter developed in the Democratic Republic of Congo in mid-2018, and it is likely that new outbreaks will occur in the future. The low survival rate discourages patients from presenting for treatment, and the occupational risk discourages HCWs from caring for highly infectious patients. A substantial proportion of survivors complain of chronic symptoms, such as eye problems (47%) and arthralgias (64%)—a condition that has been termed PES (Wilson et al., 2018). Inflammation may play a role in both the uveitis, which can result in blindness (Shantha et al., 2017) and in arthritis (Amissah-Arthur et al., 2017). Availability of an efficacious adjunctive treatment drug would save lives, increase the number of people presenting for treatment, and increase the willingness of HCWs to care for patients. An efficacious drug for adjunctive treatment of PES could decrease morbidity suffered by survivors. To date, most research efforts have focused on vaccine for prevention and either antivirals or antibody preparations for treatment. However, given the extensive inflammatory component of EVD, adjunctive therapy to decrease inflammation—but not globally downregulate the host immune response in a manner that could be detrimental (e.g., steroids)—may hold promise for better outcomes for those infected. Although drugs such as acetylsalicylic acid, ibuprofen, indomethacin, and celecoxib are also broadly anti-inflammatory, they all inhibit cyclooxygenase and can interfere with platelet aggregation—a characteristic that would be disqualifying for use with hemorrhagic fevers.

“Cytokine storm,” a burst in production of inflammatory cytokines, is thought by many to be integral to EVD pathogenesis, and high levels of tumor necrosis factor (TNF)- $\alpha$ , interleukin (IL)-6, IL-10, IL-1 $\beta$ , macrophage inflammatory protein (MIP)-1 $\alpha$ , MIP-1 $\beta$ , and macrophage chemoattractant protein (MCP)-1 are associated with fatal infections (Ruibal et al., 2016;

**TABLE 1 |** Comparison of the Effect of Ebola Virus and Minocycline on Selected Biomarkers.

Biological Marker	Ebola Virus			Minocycline Effect on Physical or Infectious Challenge	
	Nonsurvivors	Survivors	Animal Model <sup>§</sup>	In Vivo	In Vitro
TNF- $\alpha$	↑ (Baize et al., 2002)	↑ (Baize et al., 2002)	↑ (Geisbert et al., 2003; Mahanty et al., 2003; Rubins et al., 2007)	↓ (Ledeboer et al., 2005; Masocha et al., 2006; Suzuki et al., 2010; Wu et al., 2012; Hou et al., 2013; Ashraf et al., 2014)	↓ (Szeto et al., 2010; Tai et al., 2013)
IL-6	↑ (Baize et al., 2002; Hutchinson and Rollin, 2007; Wauquier et al., 2010)	↑ (Baize et al., 2002; Wauquier et al., 2010)	↑ (Geisbert et al., 2003; Rubins et al., 2007)	↓ (Ledeboer et al., 2005; Masocha et al., 2006; Suzuki et al., 2010)	↓ (Tai et al., 2013)
IFN- $\alpha$	↑ (Villinger et al., 1999; Gupta et al., 2012)	↑ (Villinger et al., 1999; Gupta et al., 2012)	↑ (Geisbert et al., 2003; Mahanty et al., 2003)		↓ (Drewes et al., 2014)
MIP-1 $\alpha$	↑ (Wauquier et al., 2010)	↑ (Wauquier et al., 2010)	↑ (Geisbert et al., 2003; Rubins et al., 2007)	↓ (Suzuki et al., 2010)	↓ (Tai et al., 2013)
MIP-1 $\beta$	↑ (Baize et al., 2002; Hutchinson and Rollin, 2007; Wauquier et al., 2010)	↑ (Wauquier et al., 2010)	↑ (Geisbert et al., 2003; Rubins et al., 2007)		↓ (Tai et al., 2013)
MCP-1			↑ (Geisbert et al., 2003; Mahanty et al., 2003; Rubins et al., 2007)	↓ (Suzuki et al., 2010)	↓ (Tai et al., 2013)
MMP-3			↑ (Cilloniz et al., 2011)	↓ (Masocha et al., 2006)	↓ (Fortier et al., 2010)
Markers of oxidative stress <sup>¶</sup>	↑ (Sanchez et al., 2004)		↑ (Hensley et al., 2002; Geisbert et al., 2003)	↓ (Suzuki et al., 2010); Huang et al., 2012; Ashraf et al., 2014)	
Pro-apoptotic factors/markers <sup>¶</sup>	↑ (Hutchinson and Rollin, 2007; Wauquier et al., 2010)	↑ (Hutchinson and Rollin, 2007)	↑ (Rubins et al., 2007)	↓ (Chu et al., 2005; Czerny et al., 2012; Drewes et al., 2014)	↓ (Yang et al., 2007; Tai et al., 2013)
Anti-apoptotic marker (bcl-2)			↓ (Gupta et al., 2012) ( <i>in vitro</i> )		↑ (Tang et al., 2007)
Liver function	↑ (Rollin et al., 2007)	↑ (Rollin et al., 2007)	↑ (Geisbert et al., 2003)	↓ (Chu et al., 2005; Czerny et al., 2012)	↓ (Szeto et al., 2010; Schwartz et al., 2013)

<sup>§</sup>In vitro data for Ebola can be found in references.<sup>¶</sup>iNOS (inducible nitric oxide synthase), NO (nitric oxide), nitrate, SOD (superoxide dismutase).<sup>¶</sup>TACE (tumor necrosis factor- $\alpha$  converting enzyme), TRAIL (tumor necrosis factor [TNF]-related apoptosis-inducing ligand), RANTES (regulated upon activation, normal T cell expressed and secreted), Eotaxin, Fas (Fas antigen), FasL (Fas antigen ligand), caspase-3, annexin.

IL, interleukin; MIP, macrophage inflammatory protein; MCP, macrophage chemoattractant protein; MMP, matrix metalloproteinase. ↑, significantly increased; ↓, significantly decreased.

Vernet et al., 2017). Nuclear factor of activated T cells (NFAT) is thought to be the key transcriptional regulator of inflammatory mediators (Madelain et al., 2018). Knock-out mice with dampened cytokine response (Tim-1<sup>-/-</sup>) are considerably more likely to survive Ebola virus challenge than their wild-type counterparts, despite a limited impact on viremia (Younan et al., 2017).

We questioned whether the inflammation associated with cytokine storm could be countered pharmaceutically—and having some familiarity with one antimicrobial (minocycline) with significant anti-inflammatory properties as well as documented antiviral activity—we searched PubMed for articles describing cytokine activity during EVD and during minocycline use.

Minocycline is an FDA-approved semisynthetic tetracycline with an established safety profile that has been used for 40 years in the treatment of acne and rosacea (Cullen and Cohan, 1976; Hersle and Gisslen, 1976), and more recently, for multidrug

resistant *Acinetobacter* (Lashinsky et al., 2017). It appears to have activity against certain viral pathogens: It inhibits H7N9 replication *in vitro* (Josset et al., 2014), attenuates stimulation of interferon-related gene and TRAIL<sup>¶</sup> in human dendritic cells and PBMCs exposed to HIV or influenza virus (Drewes et al., 2014), reduces West Nile Virus titers in brain-derived cell types in a dose-dependent manner (Michaelis et al., 2007), reduces Japanese encephalitis-induced damage in neuronal cell cultures (Mishra et al., 2009), and, based on molecular dynamics, may possibly inhibit the binding of Congo Crimean hemorrhagic fever virus to host nucleoprotein during cell infection—a host protein that is believed to be pivotal to viral replication (Sharifi et al., 2017). In a randomized controlled trial of patients with dengue hemorrhagic fever, compared to patients who received standard-of-care supportive treatment, those who also received the related tetracycline class antibiotic—doxycycline—had significantly lower mortality [20.9% vs 11.2% ( $p < 0.05$ )] and lower TNF and IL6 levels on days 3, 5, and 7 ( $p < 0.05$  for all)

(Fredeking et al., 2015). **Table 1** compares the effects of Ebola virus and minocycline on selected biomarkers including important cytokines and chemokines.

As shown in our table, the anti-inflammatory activity of minocycline opposes those of many gene products of Ebola virus. It also selectively impairs NFAT-mediated transcriptional activation (Szeto et al., 2011). Due to its small size and lipophilic nature, minocycline may reach potentially therapeutic concentrations in tissue compartments for which antibiotic penetration is typically difficult, such as the eye (Abcouwer et al., 2013; Scholz et al., 2015) and joints (McEvoy, 2016). Such spaces appear to be capable of harboring Ebola virus (Varkey et al., 2015; Steptoe et al., 2017; Subissi et al., 2018) and are thought to contribute to the chronic sequelae seen in PES (Shantha et al., 2017; PREVAIL III Study Group, 2019; Heydari-Kamjani et al., 2019). However, pharmacokinetic/pharmacodynamic (PK/PD) data are lacking that would confirm minocycline penetration into such spaces. As previously mentioned, inflammation may play a role in both the potentially blinding uveitis and arthritis of PES. Although there are animal data to suggest that minocycline may have anti-inflammatory effects in the eye (Scholz et al., 2015) and human data to suggest anti-inflammatory activity in joints (Pradier et al., 2018), it is not known whether it has direct antiviral activity against Ebola virus.

Given minocycline's broad anti-inflammatory activity against cytokines/chemokines that appear to be pathologically

upregulated by Ebola virus and the safety history, relative availability, and affordability of minocycline, we feel it should be investigated as an adjunctive therapy in animal models of acute EVD. Given that it appears to cross into protected spaces where it may have anti-inflammatory activity, we feel it should also be investigated as adjunctive therapy for chronic sequelae of EVD (i.e., PES). It is likely that the anti-inflammatory benefit would be greater in certain infected populations (starting treatment early vs late in the infection), and it is also possible that, for some, downregulation of inflammation in general could impair host clearance of the virus. These important issues are amenable to investigation in animal models.

## AUTHOR CONTRIBUTIONS

KH performed the literature review and drafted the table and text. MP helped to synthesize the tabular data and edited the text. JB reviewed the information and edited the text.

## ACKNOWLEDGMENTS

We would like to thank Dr. Conrad Quinn of the Centers for Disease Control and Prevention for listening to and asking questions about the concept and offering editorial advice.

## REFERENCES

- Abcouwer, S. F., Lin, C. M., Shanmugam, S., Muthusamy, A., Barber, A. J., and Antonetti, D. A. (2013). Minocycline prevents retinal inflammation and vascular permeability following ischemia-reperfusion injury. *J. Neuroinflammation* 10, 149. doi: 10.1186/1742-2094-10-149
- Amissah-Arthur, M. B., Poller, B., Tunbridge, A., and Adebajo, A. (2017). Musculoskeletal manifestations of Ebola virus. *Rheumatology* 57 (1), 28–31. doi: 10.1093/rheumatology/kex082
- Ashraf, T., Jiang, W., Hoque, M. T., Henderson, J., Wu, C., and Bendayan, R. (2014). Role of anti-inflammatory compounds in human immunodeficiency virus-1 glycoprotein120-mediated brain inflammation. *J. Neuroinflammation* 11, 91. doi: 10.1186/1742-2094-11-91
- Baize, S., Leroy, E. M., Georges, A. J., Georges-Courbot, M. C., Capron, M., Bedjabaga, I., et al. (2002). Inflammatory responses in Ebola virus-infected patients. *Clin. Exp. Immunol.* 128 (1), 163–168. doi: 10.1046/j.1365-2249.2002.01800.x
- Chu, H. C., Lin, Y. L., Sytwu, H. K., Lin, S. H., Liao, C. L., and Chao, Y. C. (2005). Effects of minocycline on Fas-mediated fulminant hepatitis in mice. *Br. J. Pharmacol.* 144 (2), 275–282. doi: 10.1038/sj.bjp.0706079
- Cilloniz, C., Ebihara, H., Ni, C., Neumann, G., Korth, M. J., Kelly, S. M., et al. (2011). Functional genomics reveals the induction of inflammatory response and metalloproteinase gene expression during lethal Ebola virus infection. *J. Virol.* 85 (17), 9060–9068. doi: 10.1128/jvi.00659-11
- Cullen, S. I., and Cohan, R. H. (1976). Minocycline therapy in acne vulgaris. *Cutis* 17 (6), 1208–1210. 14.
- Czerny, C., Khomukhamedov, A., Theruvath, T. P., Maldonado, E. N., Ramshesh, V. K., Lehnert, M., et al. (2012). Minocycline decreases liver injury after hemorrhagic shock and resuscitation in mice. *HPB Surg. World J. Hepatic Pancreatic Biliary Surg.* 2012, 259512. doi: 10.1155/2012/259512
- Drewes, J. L., Szeto, G. L., Engle, E. L., Liao, Z., Shearer, G. M., Zink, M. C., et al. (2014). Attenuation of pathogenic immune responses during infection with human and simian immunodeficiency virus (HIV/SIV) by the tetracycline derivative minocycline. *PLoS One* 9 (4), e94375. doi: 10.1371/journal.pone.0094375
- Fortier, L. A., Motta, T., Greenwald, R. A., Divers, T. J., and Mayr, K. G. (2010). Synoviocytes are more sensitive than cartilage to the effects of minocycline and doxycycline on IL-1alpha and MMP-13-induced catabolic gene responses. *J. Orthopaedic Res. Off. Publ. Orthopaedic Res. Soc.* 28 (4), 522–528. doi: 10.1002/jor.21006
- Fredeking, T. M., Zavala-Castro, J. E., Gonzalez-Martinez, P., Moguel-Rodriguez, W., Sanchez, E. C., Foster, M. J., et al. (2015). Dengue patients treated with doxycycline showed lower mortality associated to a reduction in IL-6 and TNF levels. Recent patents on anti-infective drug discovery. *Recent Pat. Antiinfect. Drug Discov.* 10, 1, 51–58. doi: 10.2174/1574891x10666150410153839
- Geisbert, T. W., Hensley, L. E., Larsen, T., Young, H. A., Reed, D. S., Geisbert, J. B., et al. (2003). Pathogenesis of Ebola hemorrhagic fever in cynomolgus macaques: evidence that dendritic cells are early and sustained targets of infection. *Am. J. Pathol.* 163 (6), 2347–2370. doi: 10.1016/s0002-9440(10)63591-2
- Gupta, M., MacNeil, A., Reed, Z. D., Rollin, P. E., and Spiropoulou, C. F. (2012). Serology and cytokine profiles in patients infected with the newly discovered Bundibugyo ebolavirus. *Virology* 423 (2), 119–124. doi: 10.1016/j.virol.2011.11.027
- Hensley, L. E., Young, H. A., Jahrling, P. B., and Geisbert, T. W. (2002). Proinflammatory response during Ebola virus infection of primate models: possible involvement of the tumor necrosis factor receptor superfamily. *Immunol. Lett.* 80 (3), 169–179. doi: 10.1016/s0165-2478(01)00327-3
- Hersle, K., and Gisslen, H. (1976). Minocycline in acne vulgaris: a double-blind study. *Curr. Ther. Res. Clin. Exp.* 19 (3), 339–342.
- Heydari-Kamjani, M., Demory Beckler, M., and Kesselman, M. M. (2019). Reconsidering the use of minocycline in the preliminary treatment regime of rheumatoid arthritis. *Cureus* 11 (8), e5351. doi: 10.7759/cureus.5351
- Hou, Y., Ryu, C. H., Park, K. Y., Kim, S. M., Jeong, C. H., and Jeun, S. S. (2013). Effective combination of human bone marrow mesenchymal stem cells and

- minocycline in experimental autoimmune encephalomyelitis mice. *Stem Cell Res. Ther.* 4 (4), 77. doi: 10.1186/scrt228
- Huang, C., Li, R., Zeng, Q., Ding, Y., Zou, Y., Mao, X., et al. (2012). Effect of minocycline postconditioning and ischemic postconditioning on myocardial ischemia-reperfusion injury in atherosclerosis rabbits. *J. Huazhong University Sci. Technol. Med. Sci. = Hua zhong ke ji da xue xue bao Yi xue Ying De wen ban = Huazhong keji daxue xuebao Yixue Yingdewen ban* 32 (4), 524–529. doi: 10.1007/s11596-012-0090-y
- Hutchinson, K. L., and Rollin, P. E. (2007). Cytokine and chemokine expression in humans infected with Sudan Ebola virus. *J. Infect. Dis.* 196 Suppl 2, S357–S363. doi: 10.1086/520611
- Josset, L., Zeng, H., Kelly, S. M., Tumpey, T. M., and Katze, M. G. (2014). Transcriptomic characterization of the novel avian-origin influenza A (H7N9) virus: specific host response and responses intermediate between avian (H5N1 and H7N7) and human (H3N2) viruses and implications for treatment options. *mBio* 5 (1), e01102–e01113. doi: 10.1128/mBio.01102-13
- Lashinsky, J. N., Henig, O., Pogue, J. M., and Kaye, K. S. (2017). Minocycline for the treatment of multidrug and extensively drug-resistant *a. baumannii*: A review. *Infect. Dis. Ther.* 6 (2), 199–211. doi: 10.1007/s40121-017-0153-2
- Ledeboer, A., Sloane, E. M., Milligan, E. D., Frank, M. G., Mahony, J. H., Maier, S. F., et al. (2005). Minocycline attenuates mechanical allodynia and proinflammatory cytokine expression in rat models of pain facilitation. *Pain* 115 (1-2), 71–83. doi: 10.1016/j.pain.2005.02.009
- Madelain, V., Baize, S., Jacquot, F., Reynard, S., Fizet, A., Barron, S., et al. (2018). Ebola viral dynamics in nonhuman primates provides insights into virus immuno-pathogenesis and antiviral strategies. *Nat. Commun.* 9 (1), 4013. doi: 10.1038/s41467-018-06215-z
- Mahanty, S., Gupta, M., Paragas, J., Bray, M., Ahmed, R., and Rollin, P. E. (2003). Protection from lethal infection is determined by innate immune responses in a mouse model of Ebola virus infection. *Virology* 312 (2), 415–424. doi: 10.1016/s0042-6822(03)00233-2
- Masocha, W., Rottenberg, M. E., and Kristensson, K. (2006). Minocycline impedes African trypanosome invasion of the brain in a murine model. *Antimicrob. Agents Chemother.* 50 (5), 1798–1804. doi: 10.1128/aac.50.5.1798-1804.2006
- McEvoy, T. (2016). Minocycline: rheumatoid arthritis. *Hospital Pharm.* 51 (7), 535–538. doi: 10.1310/hpj5107-535
- Michaelis, M., Kleinschmidt, M. C., Doerr, H. W., and Cinatl, J. Jr. (2007). Minocycline inhibits West Nile virus replication and apoptosis in human neuronal cells. *J. Antimicrob. Chemother.* 60 (5), 981–986. doi: 10.1093/jac/dkm307
- Mishra, M. K., Ghosh, D., Duseja, R., and Basu, A. (2009). Antioxidant potential of minocycline in Japanese encephalitis virus infection in murine neuroblastoma cells: correlation with membrane fluidity and cell death. *Neurochem. Int.* 54 (7), 464–470. doi: 10.1016/j.neuint.2009.01.022
- Pradier, M., Robineau, O., Boucher, A., Titecat, M., Blondiaux, N., Valette, M., et al. (2018). Suppressive antibiotic therapy with oral tetracyclines for prosthetic joint infections: a retrospective study of 78 patients. *Rheumatology* 46 (1), 39–47. doi: 10.1007/s15010-017-1077-1
- PREVAIL III Study Group (2019). A longitudinal study of ebola sequelae in Liberia. *N. Engl. J. Med.* 380, 10, 924–934. doi: 10.1056/NEJMoa1805435
- Rollin, P. E., Bausch, D. G., and Sanchez, A. (2007). Blood chemistry measurements and D-Dimer levels associated with fatal and nonfatal outcomes in humans infected with Sudan Ebola virus. *J. Infect. Dis.* 196 Suppl 2, S364–S371. doi: 10.1086/520613
- Rubins, K. H., Hensley, L. E., Wahl-Jensen, V., Daddario DiCaprio, K. M., Young, H. A., Reed, D. S., et al. (2007). The temporal program of peripheral blood gene expression in the response of nonhuman primates to Ebola hemorrhagic fever. *Genome Biol.* 8 (8), R174. doi: 10.1186/gb-2007-8-8-r174
- Ruibal, P., Oestereich, L., Lutdkte, A., Becker-Ziaja, B., Wozniak, D. M., Kerber, R., et al. (2016). Unique human immune signature of Ebola virus disease in Guinea. *Nature* 533 (7601), 100–104. doi: 10.1038/nature17949
- Sanchez, A., Lukwya, M., Bausch, D., Mahanty, S., Sanchez, A. J., Wagoner, K. D., et al. (2004). Analysis of human peripheral blood samples from fatal and nonfatal cases of Ebola (Sudan) hemorrhagic fever: cellular responses, virus load, and nitric oxide levels. *J. Virol.* 78 (19), 10370–10377. doi: 10.1128/jvi.78.19.10370-10377.2004
- Scholz, R., Sobotka, M., Caramoy, A., Stempfl, T., Moehle, C., and Langmann, T. (2015). Minocycline counter-regulates pro-inflammatory microglia responses in the retina and protects from degeneration. *J. Neuroinflammation* 12, 209. doi: 10.1186/s12974-015-0431-4
- Schwartz, J., Holmuhamedov, E., Zhang, X., Lovelace, G. L., Smith, C. D., and Lemasters, J. J. (2013). Minocycline and doxycycline, but not other tetracycline-derived compounds, protect liver cells from chemical hypoxia and ischemia/reperfusion injury by inhibition of the mitochondrial calcium uniporter. *Toxicol. Appl. Pharmacol.* 273 (1), 172–179. doi: 10.1016/j.taap.2013.08.027
- Shantha, J. G., Crozier, I., and Yeh, S. (2017). An update on ocular complications of Ebola virus disease. *Curr. Opin. Ophthalmol.* 28 (6), 600–606. doi: 10.1097/icu.0000000000000426
- Sharifi, A., Amanlou, A., Moosavi-Movahedi, F., Golestanian, S., and Amanlou, M. (2017). Tetracyclines as a potential antiviral therapy against Crimean Congo hemorrhagic fever virus: Docking and molecular dynamic studies. *Comput. Biol. Chem.* 70, 1–6. doi: 10.1016/j.combiolchem.2017.06.003
- Steptoe, P. J., Scott, J. T., Baxter, J. M., Parkes, C. K., Dwivedi, R., Czanner, G., et al. (2017). Novel retinal lesion in ebola survivors, sierra leone, 2016. *Emerging Infect. Dis.* 23 (7), 1102–1109. doi: 10.3201/eid2307.161608
- Subissi, L., Keita, M., Mesfin, S., Rezza, G., Diallo, B., Van Gucht, S., et al. (2018). Ebola virus transmission caused by persistently infected survivors of the 2014–2016 outbreak in west africa. *J. Infect. Dis.* 218 (suppl\_5), S287–Ss91. doi: 10.1093/infdis/jiy280
- Suzuki, H., Sugimura, Y., Iwama, S., Suzuki, H., Nobuaki, O., Nagasaki, H., et al. (2010). Minocycline prevents osmotic demyelination syndrome by inhibiting the activation of microglia. *J. Am. Soc. Nephrol. JASN* 21 (12), 2090–2098. doi: 10.1681/asn.2010040438
- Szeto, G. L., Brice, A. K., Yang, H. C., Barber, S. A., Siliciano, R. F., and Clements, J. E. (2010). Minocycline attenuates HIV infection and reactivation by suppressing cellular activation in human CD4+ T cells. *J. Infect. Dis.* 201 (8), 1132–1140. doi: 10.1086/651277
- Szeto, G. L., Pomerantz, J. L., Graham, D. R., and Clements, J. E. (2011). Minocycline suppresses activation of nuclear factor of activated T cells 1 (NFAT1) in human CD4+ T cells. *J. Biol. Chem.* 286 (13), 11275–11282. doi: 10.1074/jbc.M110.210518
- Tai, K., Iwasaki, H., Ikegaya, S., and Ueda, T. (2013). Minocycline modulates cytokine and chemokine production in lipopolysaccharide-stimulated THP-1 monocytic cells by inhibiting IkappaB kinase alpha/beta phosphorylation. *Transl. Res. J. Lab. Clin. Med.* 161 (2), 99–109. doi: 10.1016/j.trsl.2012.10.001
- Tang, X. N., Wang, Q., Koike, M. A., Cheng, D., Goris, M. L., Blankenberg, F. G., et al. (2007). Monitoring the protective effects of minocycline treatment with radiolabeled annexin V in an experimental model of focal cerebral ischemia. *J. Nucl. Med. Off. Publ. Soc. Nucl. Med.* 48 (11), 1822–1828. doi: 10.2967/jnumed.107.041335
- Varkey, J. B., Shantha, J. G., Crozier, I., Kraft, C. S., Lyon, G. M., Mehta, A. K., et al. (2015). Persistence of ebola virus in ocular fluid during convalescence. *N. Engl. J. Med.* 372 (25), 2423–2427. doi: 10.1056/NEJMoa1500306
- Vernet, M. A., Reynard, S., Fizet, A., Schaeffer, J., Pannetier, D., Guedj, J., et al. (2017). Clinical, virological, and biological parameters associated with outcomes of Ebola virus infection in Macenta, Guinea. *JCI Insight* 2 (6), e88864. doi: 10.1172/jci.insight.88864
- Villinger, F., Rollin, P. E., Brar, S. S., Chikkala, N. F., Winter, J., Sundstrom, J. B., et al. (1999). Markedly elevated levels of interferon (IFN)-gamma, IFN-alpha, interleukin (IL)-2, IL-10, and tumor necrosis factor-alpha associated with fatal Ebola virus infection. *J. Infect. Dis.* 179 Suppl 1, S188–S191. doi: 10.1086/514283
- Wauquier, N., Becquart, P., Padilla, C., Baize, S., and Leroy, E. M. (2010). Human fatal zaire ebola virus infection is associated with an aberrant innate immunity and with massive lymphocyte apoptosis. *PLoS Negl. Trop. Dis.* 4 (10), e837. doi: 10.1371/journal.pntd.0000837
- Wilson, H. W., Amo-Addae, M., Kenu, E., Ilesanmi, O. S., Ameme, D. K., and Sackey, S. O. (2018). Post-ebola syndrome among ebola virus disease survivors in montserrado county, liberia 2016. *BioMed Res. Int.* 2018, 1909410. doi: 10.1155/2018/1909410
- Wu, K. L., Chan, S. H., and Chan, J. Y. (2012). Neuroinflammation and oxidative stress in rostral ventrolateral medulla contribute to neurogenic hypertension induced by systemic inflammation. *J. Neuroinflammation* 9, 212. doi: 10.1186/1742-2094-9-212

- Yang, L. P., Zhu, X. A., and Tso, M. O. (2007). Minocycline and sulforaphane inhibited lipopolysaccharide-mediated retinal microglial activation. *Mol. Vision* 13, 1083–1093.
- Younan, P., Iampietro, M., Nishida, A., Ramanathan, P., Santos, R. I., Dutta, M., et al. (2017). Ebola virus binding to Tim-1 on T lymphocytes induces a cytokine storm. *mBio* 8 (5), e00845–17. doi: 10.1128/mBio.00845-17

**Disclaimers:** Use of trade names and commercial sources is for identification only and does not imply endorsement. This is an opinion piece based on literature review. No experiments have been conducted or data collected at this time for the potential adjunct treatment of Ebola virus disease or post-Ebola syndrome.

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Hendricks, Parrado and Bradley. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.