



Beyond Alkaloids: Novel Bioactive Natural Products From Lobelia Species

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In this work, we reviewed the progress in the phytochemical and biological investigations of bioactive components derived from medicinally valuable *Lobelia* species. In the last 60 years, *Lobelia* has garnered significant attention from the phytochemist from around the world, majorly due to the discovery of bioactive piperidine alkaloids (e.g., lobinaline and lobeline) in the early 1950s. Later, lobeline underwent clinical trials for several indications including the treatment of attention deficit hyperactivity disorder and a multicenter phase three trial for smoking cessation. Subsequently, several other alkaloids derived from different species of *Lobelia* were also investigated for their pharmacological characterization of the other novel chemical classes. The major shift has been noticed due to the structurally similar alkaloid components, which essentially share similar pharmacological, physicochemical, and toxicological profiles. In this review, we present an up-to-date overview of their progress with special attention to understanding the molecular mechanisms of the novel bioactive components.

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INTRODUCTION

Lobelia, named after the botanist Matthias de lobel, is a large genus of medicinally valuable flowering plants belonging to the Campanulaceae family comprising more than 450 species distributed predominantly in tropical and temperate regions of the world. Several species of *Lobelia* are traditionally used in different folk medicine to treat various diseases. Among them, *Lobelia inflata, Lobelia nicotianaefolia, Lobelia cardinalis, Lobelia chinensis, Lobelia laxiflora, Lobelia trigona* Roxb., *Lobelia siphilitica* L., *Lobelia sessilifoilia* Lamb, *Lobelia polyphylla*, and *Lobelia pyramidalis* Wall. are some of the frequently used *Lobelia* species. The phytochemical researches of these species led to the discovery of several novel bioactive secondary metabolites.

The earliest scientific report on the pharmacological use of *Lobelia* species in diseased conditions dates back to 1828 (Andrew, 1828). More than a century later (in 1950), Tondeur and Charlier reported the extraction and pharmacological activities of alkaloids from *Lobelia suavibracteata* (Haumann) and *Lobelia giberroa* (Hemsl.) (Charlier and Tondeur, 1950; Tondeur and Charlier, 1950). In the 1950s, other studies began to reveal the distinctive pharmacological properties of the alkaloid constituents derived from different *Lobelia* species (Lendle and Richter, 1950). One of the alkaloids that garnered major attention was lobeline 1 (**Figure 1**), which was primarily derived from the aerial parts of *Lobelia inflata*, (Kaczmarek and Steinegger, 1959; Saldanha et al., 1959). In the

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subsequent decades, the studies on the prospective evaluation of the use of lobeline have seen unprecedented advances, including but not limited to smoking cessation (Glover et al., 2010; Stead and Hughes, 2012), treatment of drug and alcohol abuse (Bell et al., 2009; Hart et al., 2010; Roni and Rahman, 2017), adult ADHD (Martin et al., 2018), anti-depressant (Roni and Rahman, 2015), anti-epileptic (Tamboli et al., 2012), and Parkinson's disease (Li et al., 2014).

For example, in their study, Roni and Rahman noticed that adult C57BL/6J mice when treated with once daily dose of lobeline (1 mg/kg, s.c.) in the last 14 days of their exposure to chronic unpredictable stress (CUS) for 6 weeks produced antidepressant-like effects. They proposed targeting brain nicotinic acetylcholine receptors (nAChRs), brain-derived neurotrophic factor (BDNF) expression and/or cell proliferation in the hippocampus to be the most likely cause of the observed antidepressant-like effects of lobeline (Roni and Rahman, 2015). Saline was used as the negative control. Hart et al. reported that at doses of 1.0 mg/kg and 3.0 mg/kg subcutaneous injections of lobeline attenuates self-administration of intravenous (i.v.) heroin (3,6 diacetylmorphine HCl) infusions (18 µg/kg) in male Sprague-Dawley rats suggesting their use in pharmacotherapy for opioid abuse (Hart et al., 2010). In both studies, however, they did not make a comparison of the effect of lobeline with any reference standard.

Martin et al. reported the results of a preclinical study in which they studied the effect of lobeline in adult ADHD (Martin et al., 2018). In their study, they subjected a total of nine adults (5 females, 4 males, ages 23-41 [31.11 ± 7.08 years]) to a 7-day protocol of oral administration of methylphenidate capsule (0, 15, or 30 mg, p.o.) followed by sublingual (s.l.) administration of lobeline tablets (0, 7.5, 15, or 30 mg, s.l.) after 1 h. Inactive doses of both sublingual and oral formulations were used as placebo. They observed that lobeline could improve working memory in a modest capacity (without significant improvement in their attention) in nine volunteers with ADHD.

Li et al. investigated the effect of s.c administrations of lobeline hydrochloride (1 or 3 mg/kg) on male C57BL/6J mice against methyl-4-phenyl-1,2,3,6-tetra-hydropyridine (MPTP, 30 mg/kg)-induced cell death in vivo using L-dopa (80 mg/kg) and the potent and selective dopamine reuptake inhibitor, GBR12935 (10 mg/kg) as positive controls (Li et al., 2014). The s.c. injections of lobeline, GBR12935, or L-dopa were done for 11 days 30 min prior to MPTP administration. They observed that MPTP induced locomotive effects detected in behavioral tests were significantly reduced by lobeline as compared to L-dopa and GBR12935. Further, they observed that the loss of neurotoxin-induced immunoreactivity was significantly reduced in the substantia nigra and striatum regions of the brain. The overall results indicated that lobeline may protect dopaminergic neurons and alleviate the symptoms of Parkinson's disease. Saline was used as the negative control.

Tamboli et al. investigated lobeline isolated from *Lobelia* nicotianaefolia in chemoconvulsant-induced seizures using diazepam as a reference standard (Tamboli et al., 2012). The anticonvulsant activity of the isolated lobeline was investigated at 5, 10, 20 and 30 mg/kg, intraperitoneal (i.p.)

doses in Pentylenetetrazol (PTZ) and strychnine induced seizures in Swiss albino male mice. They found that isolated lobeline induces potent anticonvulsant activity against PTZ induced seizures by enhancing the brain gamma amino butyric acid (GABA) level at 20 mg/kg i.p. within a duration of 45 min after their administration. Saline was used as the negative control.

Mechanistically, lobeline was reported binding to the a4β2 nAChR with nanomolar affinity (Brioni et al., 1997) but with less potent affinity against muscarinic and acetylcholinesterase receptors (Terry et al., 1998). The high-affinity partial agonist property of lobeline against a4B2 nAChR was later identified as the mechanism for smoking cessation (Wu et al., 2006; Billen et al., 2012). Recently, Anitha et al. shed additional light on how smoking cessation by lobeline occurs through trapping in $\alpha 4\beta 2$ receptors-containing acidic vesicles and how intracellular pH homeostasis plays a crucial role in it (Govind et al., 2017). Lobeline was also demonstrated to block the blood-brain barrier (BBB) basic amine transporter (Allen et al., 2003), and act as VMAT2 ligand (Zheng et al., 2005; Zheng et al., 2006). In another study, (-)-lobeline hydrochloride monohydrate (10 or 20 mg/kg, i.p.) was reported to reduce the DNA damage and oxidative stress induced by seizures in a pilocarpine-induced seizure model on 69 adult male CF-1 mice (da Costa E Silva et al., 2018). The administrations of pilocarpine (300 mg/kg) were performed after 30 min treatment of lobeline, diazepam (positive control) or saline (negative control).

Overall, to date, more than 610 scientifically peer-reviewed research articles carrying the pharmacological investigation of the pyridine alkaloid lobeline and its structural analogs were reported underlying its medicinal importance. However, the gastrointestinal side effects (due to nicotinic effects), the lethal toxicity due to its narrow therapeutic index, and other adverse side effects such as choking and cough limits the clinical utility of lobeline (Butler et al., 2001; Raj et al., 2005; Stead and Hughes, 2012; Heinzerling, 2013).

Beyond Lobeline: Other Bioactive Alkaloids

In the early 1950s, the German phytochemist Steinegger and team reported several key discoveries on the identification and pharmacological studies of the newer alkaloids from different Lobelia species. First, they reported the identification of a crystallized alkaloid named lurenine isolated from Lobelia urens (Steinegger and Grutter, 1950). Subsequently, they also identified two new alkaloids lophilin and lophilacrin in the alkaloid fraction of Lobelia siphilitica (Steinegger and Egger, 1952). In the 1950s, other studies began to reveal the distinctive pharmacological properties of the alkaloid constituents identified in different Lobelia species (Lendle and Richter, 1950), which has further raised tremendous interest in phytochemical research. Lobinaline 2, the only alkaloid found in the Lobelia cardinalis L. and the first binitrogenous alkaloid extracted from any Lobelia species was reported having a unique nAChR binding activity (Klosa, 1953; Kaczmarek and Steinegger, 1958; Brown et al., 2016). In addition to their equipotent α4β2-and α7-nAChRs binding profile, the purified lobinaline obtained from CHCl₃ fraction of air-dried aerial

portions of *L. cardinalis* also exhibited potent 2,2-Diphenyl-1picrylhydrazyl (DPPH) free radical scavenging properties, inhibited *in vitro* [(3)H]-dopamine uptake in adult male Sprague-Dawley rats striatal synaptosomes, and potent dopamine transporter inhibition (Brown et al., 2016). Reportedly, lobinaline (stock solution: 1 mg/ml) scavenge DPPH free radicals with an EC₅₀ of 17.98 μ M when examined in DPPH (stock solution: 600 μ M) free radical scavenging assay. In contrast, lobeline (stock solution: 1 mg/ml) demonstrated a weak DPPH free radical scavenging property (EC₅₀ = 228.8 μ M). *In vivo* electrochemical studies indicated that lobinaline in a short acting and likely competitive-capacity inhibits dopamine transporter function in the dorsal striatum of urethaneanesthetized rats. Interestingly, lobinaline (1 mM) was noticed significantly increasing ⁴⁵Ca²⁺ entry in SH-SY5Y cells.

In 1954, major alkaloids named lobelanidine 3, and lelobanidine 4 were also isolated from *Lobelia nicotianaefolia* (Gedeon and Gedeon, 1954). Lobelanidine 3 was subsequently identified in *Lobelia chinensis* exhibiting anticancer and antiemetic properties (Chen et al., 2014). In 2001, two new pyrrolidine alkaloids, radicamines A 5 and radicamines B 6 isolated from the hot water extract of *Lobelia chinensis* Lour. (Shibano et al., 2001). The basic extract solution (25 μ L) was were identified exhibiting potent alpha-glucosidase (100 μ L) inhibition (Shibano et al., 2001). Radicamines A, radicamines B and the positive control (2*R*,3*R*,4*R*,5*R*) 2,5-dihydroxymethyl-3,4-dihydroxy-pyrrolidine (DMDP) exhibited IC₅₀ of 6.7 × 10⁻⁶, 9.3 × 10⁻⁶, and 4.9 × 10⁻⁶ M, respectively.

In 2015, Paz et al. reported a new alkaloid, pentylsedinine 7, isolated from the fresh aerial parts of *Lobelia tupa* (Paz et al., 2015). When evaluated against the $\alpha 3\beta 2/\alpha 3\beta 4$ nAChR and $\alpha 7$ nAChR receptors expressed in the human neuroblastoma cell line SH-SY5Y, like lobeline, (-)-pentylsedinine acted as a non-selective partial agonist at homomeric $\alpha 7$ nAChR (EC₅₀ 0.4 \pm 0.08 μ M) and heteromultimeric $\alpha 3\beta 2/\alpha 3\beta 4$ (EC₅₀ 0.33 \pm 0.07 μ M) nAChR. Interestingly, the co-addition of lobeline and (-)-pentylsedinine desensitized $\alpha 7$ nAChR to the response from the selective $\alpha 7$ nAChR agonist choline. Apparent IC₅₀ of lobeline and (-)-pentylsedinine were 1.6 \pm 0.4 μ M and 0.37 \pm 0.08 μ M, respectively.

Lobelane 8, a defunctionalized analog of lobeline exhibited 10–15 fold more potent effects than lobeline in inhibition of the vesicular dopamine (DA) uptake by VMAT2, and inhibited (+)-methamphetamine-evoked DA release (Vartak et al., 2009; Nickell et al., 2010). It was also reported to possess a fivefold binding affinity than lobeline for the dihydrotetrabenazine binding site on VMAT2 (Miller et al., 2004; Zheng et al., 2005). In a study aimed at synthesizing lobelane analogs as therapeutic agents, replacement of the phenyl groups of lobelane with quinolyl groups resulted in a water-soluble analog called quinlobelane nine that possessed stronger VMAT2 inhibition (Vartak et al., 2010).

The phytochemical research conducted over the years suggested that alkaloids are the predominant secondary metabolites (\sim 46%) present in most of the *Lobelia* species known. These alkaloids ubiquitously contain *N*-methylpiperidine ring or piperidine ring along with one or

two substituents at the C_2 and/or C_6 position of the ring (Felpin and Lebreton, 2004; Folquitto et al., 2019).

One of the major disadvantages of the alkaloids derived from the Lobelia species has been their extreme resemblance in the chemical structures to each other, essentially sharing similar pharmacological, physicochemical, and toxicological profiles which also include cardiotoxicity (Figure 2). The alkaloids in most of the species of Lobelia (with lobeline being the most cited in the literature) are broadly known for presenting medicinal properties against drug abuse, neurological disorder, and respiratory stimulant. The limited medicinal properties of these alkaloids can be attributed to their binding properties at the nAChR receptors. In this present manuscript, we have only considered the publications where the alkaloid was biologically tested. Unfortunately, the bioactivity characterization of a large majority of those alkaloids was not yet reported and thus excluded. Moreover, to date, more than 80 alkaloids are known to be isolated from different species of Lobelia. The readers are recommended to the following review for the updates on other isolated alkaloids (Folguitto et al., 2019).

Beyond Alkaloids: The Bioactive Components of Other Classes

Over the years, the isolation and characterization of secondary metabolites derived from *Lobelia* species have garnered significant attention. It is now an established fact that most species of the genus *Lobelia* contain different classes of secondary metabolites including but not limited to flavonoids, terpenoids, fatty acids, polyacetylenes, coumarins, neolignans, polysaccharides, and amides in varying concentrations. In recent years, the identifications of these newer compounds also presented the opportunities to address the limited bioactivity of alkaloids mentioned earlier. Due to this fact, growing numbers of literature are being published. In this section, we will present a current account of these newer novel bioactive natural products derived from *Lobelia* species. A special focus has been given to accounting for the molecular mechanism of these newer secondary metabolites.

Since mid-to end- 20th-century investigations into the pharmacology of components derived from *Lobelia* species were centered onto the alkaloids. In 1961, Sasaki et al. reported the isolation of a new glucofructan "sessilifolan" from *Lobelia sessilifolia* Lamb. I. (Sasaki et al., 1961; Sasaki and Mikami, 1962). It was perhaps the first report identifying other secondary metabolites than alkaloids in *Lobelia*. In 1986, Santosa et al. reported the *in vitro* as well as *in vivo* activity of the hot water extract of *Lobelia chinensis* against Ehrlich ascites tumor and Sarcoma 180 cells (Santosa et al., 1986). However, in subsequent decades, not much progress has been made on characterizing the other classes of secondary metabolites.

Subarnas et al. identified a novel antidepressant bioactive component, β -amyrin palmitate 10 in an extract of the leaves of *Lobelia inflata* (Subarnas et al., 1992). Further studies indicated that the antidepressant activity of β -amyrin palmitate 10 was related to the activation of the noradrenergic system by releasing



[3H]norepinephrine from newly synthesized pools, with mianserin-like action (Subarnas et al., 1993b; Subarnas et al., 1993c). In a forced swimming test, β -amyrin palmitate at 10 mg/kg reduced the tetrabenazine-induced (100 and 200 mg/kg) increase in the duration of the immobility of mice whereas it has shown no effect when treated with alpha-methylpara-tyrosine (500 mg/kg). Similarly, at 5 and 10 mg/kg, it has decreased the duration of immobility in mice treated with 6hydroxy-dopamine (50 µg/mouse) and desipramine (15 mg/kg) without showing any effect to the treatment of mice with nomifensine plus 6-hydroxydopamine (Subarnas et al., 1993b). Physicochemically, beta-amyrin palmitate is a water-insoluble, extremely weak basic compound belonging to the triterpenoids compound class which contains six isoprene units. Other studies reported β-amyrin palmitate exhibiting different pharmacological properties such as induction of hypoactivity by inhibiting alpha 1-adrenoceptors (Subarnas et al., 1993a), anti-diabetic and anti-hyperglycemic activity by blocking the entry of glucose from the intestine (Nair et al., 2014), and antibacterial against Gram-positive and negative bacteria and antifungal activities (Abed et al., 2016).

Recently, Wang et al. reported the identification of four novel flavonoid glycosides, lobelitin A-D 11–14 exhibiting agonistic activities against G protein-coupled receptor 35 (GPR35) (Wang et al., 2019). The compounds were isolated from *Lobelia chinensis*.

The authors noticed that lobelitin A-D triggers dosedependent positive signals (monitored for 1 h) in dynamic mass redistribution (DMR) agonist assay in varying concentration. For example, at 200 pm, lobelitin A, B, and D were recorded exhibiting the largest DMR signal whereas at about 100 pm, lobelitin C exhibited a weaker signal at the maximum concentration of 250 μ M. Based on these results, the authors suggested lobelitin A, B and D as the full agonist and lobelitin C as a partial agonist of GPR35. The structures and the apparent EC₅₀ and IC₅₀ values of lobelitin A-D are shown in **Figure 3** and **Table 1**. GPR35 has been implicated in several diseased conditions including hypertension, coronary artery disease, and cancer, and is an area of active research (Divorty et al., 2015; Wei et al., 2017; Quon et al., 2020).

The glucoside 2,6-Dideoxy-7-O-(b- D -glucopyranosyl) 2,6-imino- D -glycero- L -gulo-heptitol (7-O- β -D-glucopyranosyl-



α-homonojir-imycin, 15) isolated from the methanol extract of the whole plant of *Lobelia sessilifolia* exhibited potent inhibitory activity against various α-glucosidases (Ikeda et al., 2000). Specifically, 7-*O*-β- D -glucopyranosyl-α-homonojir-imycin exhibited an IC₅₀ of 0.25 and 0.27 μM against rice and rat intestinal isomaltase α-glucosidase, respectively. Additionally, it has also shown the inhibition of porcine kidney trehalase (IC₅₀ = 0.013 μM) and anti-diabetic property (Ikeda et al., 2000).

Kuo et al. reported the antivirus and anti-inflammatory properties of a novel compound lobechine 16 alongwith scoparone (6,7-dimethoxycoumarin) 17 isolated from the methanol extracts of *Lobelia chinensis*. Lobechine 16 inhibited elastase release with IC₅₀ of 25.01 \pm 6.95 μ M whereas scoparone 17 inhibited superoxide anion generation with IC₅₀ of 6.14 \pm 1.97 μ M (Kuo et al., 2011).

A neutral polysaccharide (LCPS) 18 was isolated from *Lobelia* chinensis lour. The monosaccharide composition of LCPS was observed as fructan composed of 2,1-linked- β -D-fructofuranosyl residue and α -D-glu-copyranosyl residue with α -D-Glcp-(1 \rightarrow [1)- β -D-Fruf-(2]₁₅ glycosidic linkage. LCPS was identified as an inulin-type fructan with 2.6 kDa molecular weight with an effective therapeutic option in the treatment of obesity. Oral administration of this inulin alleviated obesity and reduced body weight in high-fat diet-induced mice (Zhang et al., 2020). Specifically, the anti-obesity effects of LCPS evaluated *in vivo* in male Kunming mice treated with normal chow diet (D12450 J), high fat diet (D12492) and high fat diet plus LCPS (300 mg/kg/ day) for eight weeks showed 67% lower body weight gain as compared to high fat diet treatment. High fat diet plus glibenclamide (5 mg/kg/day) was used as a positive control which has shown 77% effect.

Li et al. isolated another neutral α -glucan, named BP1 19 (Mol. weight: 9.45 kDa), from Lobelia chinensis by hot-water extraction and studied its immunomodulating activities (Li et al., 2016). The authors proposed that a-glucan BP1 activates Toll-like receptor 4 (TLR4) and exerts immunomodulating effects such as enhancement of the cell proliferation, nitric oxide production, cytokine secretion and phagocytosis in a dose-dependent manner. Additionally, the glucan also induced cytokine and nitric oxide production mediated by toll-like receptor 4 (TLR4) in RAW 264.7 cells. The backbone of BP1 consisted of alternating units of $_{6}\alpha$ - D -Glcp¹ -- $_{6,3}\alpha$ - D -Glcp¹ -- ($_{6}\alpha$ - D-Glcp¹) x--($_{6,3}\alpha$ - D -Glcp¹)y. The side chains were α -D-Glcp¹ --($_{6}\alpha$ -D- $Glcp^1)z-_4\alpha$ -D- $Glcp^1-_3\alpha$ -D- $Glcp^1-_4\alpha$ -D- $Glcp^1$ and α - D- $Glcp^1$ terminally attached to backbone O-3 of $_{6.3}\alpha$ - D -Glcp¹ (as shown in Figure 4). Specifically, BP1 demonstrated a dose-dependent induction of proliferation of RAW 264.7 cells at the concentrations of 12.5, 25, 50, and 100 µg/ml when administered together with polyB in an MTT assay and tested for 24 h.

Ishimaru et al., isolated two new polyacetylene compounds lobetyolin 20 and lobetyol 21 from the hairy roots of *Lobelia inflata* (Ishimaru et al., 1991). Recently, He et al. studied the effect of lobetyolin on colorectal cancer (He et al., 2020). They found that lobetyolin induces colon cancer cells by inhibiting the ASCT2-mediated glutamine metabolism. Briefly, they have



incubated lobetyolin in HCT-116 cells for 3, 6, 12, 18 and 24 h at varying concentrations of 10, 20 and 40 μ mol/L in an MTT assay and observed that lobetyolin induces largest inhibitory effect on survival rate at 24 h. On the other hand, the authors did not observe any significant influence in the cell viability in NCM460 cells at 10, 20 and 40 μ mol/L. Additionally, they observed that

lobetyolin at 10, 20, and 40 μ mol/L prominently suppresses the expression of ASCT2 protein and reduces the mRNA levels. In addition, lobetyolin at 40 μ mol/L also inhibited the expression of ASCT2 in HCT-116 cells. Further, lobetyolin (10, 20, and 40 mg/kg) inhibited the tumor volume in the nude (nu/nu) mice transplanted with HCT-116 cells. In another study, Shen et al.

TABLE 1 Pharmacological characteristics of active flavonoid glycosides derived
from Lobelia chinensis in HT-29 cells (Wang et al., 2019).

Compounds	EC ₅₀ (μΜ)	IC ₅₀ ^a (μM)	IC₅₀^b (μΜ) 47.63 ± 6.31	
Lobelitin A	49.92 ± 12.61	0.38 ± 0.08		
Lobelitin B	18.77 ± 2.02	0.27 ± 0.03	27.89 ± 2.33	
Lobelitin C	Weak	0.39 ± 0.21	72.74 ± 22.96	
Lobelitin D	67.65 ± 37.99	0.48 ± 0.13	64.70 ± 17.19	

^aIC₅₀ of ML145 to block the DMR of compounds.

^bIC₅₀ to desensitize the DMR of 1 μM zaprinast in HT-29 cells.

reported that lobetyol induces anti-carcinoma effect in MKN45 cells and activates MAPK pathways associated with G_1/S cell cycle arrest in a time- and dose-dependent manner (Shen et al., 2016). Specifically, the anti-proliferative effect of lobetyol in MKN45 cells was evaluated in an MTT assay at a varying concentration and time ranging from 50 µg/ml, 75 µg/ml, to 110 µg/ml and 12 h, 24 h, to 48 h, respectively. Lobetyol

demonstrated an IC₅₀ of 71.47 \pm 4.29 µg/ml at 48 h. Further, in MKN45 cells, an increasing dose of lobetyol ranging from 0, 50, and 75 µg/ml, to 110 µg/ml led to an increase in apoptotic population by 5.5%, 13.74%, 27.32%–31.57%, respectively.

Two other polyacetylene isolobetyol 22 and lobetyol 21 isolated from *Lobelia chinensis* exhibited moderate cytotoxic activities against MSTO-211H and NCI-H292 cell lines (treated for 48 h). Isolobetyol exhibited the IC₅₀ values of 12.36 and 9.31 μ M against the two cell lines; while the lobetyol showed the IC₅₀ values of 11.76 and 9.64 μ M, respectively (Yang et al., 2014). In contrast, positive control (cisplatin) showed an IC₅₀ of 4.91 and 7.89 μ M against MSTO-211H and NCI-H292 cell lines, respectively. However, the authors did not mention at which concentrations the compounds were tested.

In another report, Joshi et al. analyzed the essential oil composition of the aerial parts of *Lobelia pyramidalis* Wall and evaluated their antimicrobial activity (Joshi et al., 2011).



TABLE 2 Lobelia species us	sed in our study to investigate th	e neuroprotection mechanism.

NPASS IDs	Herb name	Family	Super kingdom	# Of natural product	References
NPO10220	Lobelia davidii	Campanulaceae	Eukaryota	3	Zhang et al. (1990)
NPO11547	Lobelia cardinalis	Campanulaceae	Eukaryota	4	Kaczmarek and Steinegger (1958), Kelley et al. (2019)
NPO11565	Lobelia salicifolia	Campanulaceae	Eukaryota	7	Steinegger and Ochsner (1956)
NPO14650	Lobelia chinensis ^a	Campanulaceae	Eukaryota	71	Shibano et al. (2001), Zhou et al. (2008), Kuo et al. (2011), Yang et al. (2014), L et al. (2016), Wang et al. (2019), Kuo et al. (2011), Ge et al. (2020)
NPO14996	Lobelia syphilitica	Campanulaceae	Eukaryota	20	Kesting et al. (2009)
NPO15709	Lobelia langeana	Campanulaceae	Eukaryota	20	
NPO16059	Lobelia polyphylla	Campanulaceae	Eukaryota	2	Weinges et al. (1972)
NPO17414	Lobelia portoricensis	Campanulaceae	Eukaryota	1	Meléndez et al. (1967)
NPO18202	Lobelia radicans ^a	Campanulaceae	Eukaryota	5	
NPO238	Lobelia st	Campanulaceae	Eukaryota	16	
NPO24373	Lobelia inflata	Campanulaceae	Eukaryota	20	Ishimaru et al. (1991), Bálványos et al. (2004), Kursinszki et al. (2008)
NPO26111	Lobelia camporum	Campanulaceae	Eukaryota	1	
NPO293	Lobelia nicotianifolia	Campanulaceae	Eukaryota	1	
NPO29542	Lobeliae chinensis herba ^a	Campanulaceae	Eukaryota	41	
NPO28802	Lobelia nummularia lam	Campanulaceae	Eukaryota	3	Ishimaru et al. (1991), Ho et al. (1995)
NPO5012	Lobelia hassleri	Campanulaceae	Eukaryota	4	
NPO5051	Lobelia urens	Campanulaceae	Eukaryota	3	
NPO5103	Lobelia berlandieri	Campanulaceae	Eukaryota	4	Williams et al. (1987)
NPO615	Lobelia exaltata	Campanulaceae	Eukaryota	1	
NP08894	Lobelia fistulosa	Campanulaceae	Eukarvota	2	

^aare the same species with two different entries in NPASS database. (Zeng et al., 2018)

Serial doubling dilutions of the essential oil were prepared from 0.10 to 200 mg/ml and incubated in Mueller Hinton broth for the bacterial strains at 37° C for 24 h. The minimal inhibitory concentration and inhibition zone of 3.12 mg/ml and 18 mm, respectively was determined against the bacterial strain *S. aureus.* The authors suggested natural terpenoid perilla ketone could be the responsible chemical constituent. However, no further attempt was made to isolate and characterize the compound.

In a recent study, Ma and colleagues identified 208 metabolites of *Lobelia chinensis* by gas chromatography-mass spectrometer (GC/MS) and conducted a network pharmacology-based investigation of their anti-diabetic mechanism (Ge et al., 2020). Their study revealed that 5-hydroxymethylfurfural 23 acts on GSK3B, TNF, and MAPK1 and affects the insulin resistance signaling pathway as the major mechanism of action. Also, acacetin might act on INSR, DPP4, and GSK3B. No biological evaluation was performed to validate the network pharmacology approach's presented findings, which remains one major drawback of this work.

We have also recently investigated the neuroprotection mechanism of the pharmacokinetically favorable, experimentally validated active compounds derived from the 17 *Lobelia* species (**Table 2**) based on network pharmacology and molecular modeling (Zheng et al., 2020). In this work, we also studied the chemical diversity of the validated natural compounds identified in the genus of the *Lobelia* species including the major components of *Lobelia chinensis* studied by Ma and Colleagues (Ge et al., 2020).

Fascinatingly, the aglycone part of the flavonoid such as diosmetin and 18 other aglycones constituted the densest cluster indicating their profound presence in the *Lobelia* species. The representative chemical structure of the top 10 clusters is shown in **Figure 4B**. We have also not performed the biological evaluations. However, in this review, we intended to summarize the experimentally determined chemical constituents of *Lobelia* and draw a comparison in their chemical structures (as shown in **Figure 4**).

Data Sources and Study Selection

The literature search was performed using PubMed and Scifinder over the period from the beginning of the database until October 2020. Initially, all the literature associated with the search term "Lobelia" were collected. In addition, the databases were also directly searched using the botanical name (e.g., "Lobelia chinensis"). In addition, as a third search strategy, the databases of the major research journals including Science Direct, Frontiers, Wiley, American Chemical Society, MDPI, Springer-Nature, and Taylor and Francis were also searched manually. Articles that have reported only isolation and identification of chemical constituents without biological activities were excluded. No unpublished literature data or conference proceedings were included. Except for a recent systematic review by Folquitto et al. (Folquitto et al., 2019), other articles that described reviews or systematic reviews were excluded. The systematic review by Folquitto et al. was used as an additional strategy to validate the references for the phytochemical details.

CONCLUSION

Undoubtedly, *Lobelia* is an important genus with profound medicinal value. A lot of work has already been done in understanding the pharmacological role of the alkaloids derived from different *Lobelia* species. Yet, growing investigations on the newer chemical classes indicate that there is certainly an untapped opportunity to obtain bioactive compounds with diverse chemical and pharmacological properties. In this work, we tried to provide the readers the most up-to-date account of the progress made so far, especially toward understanding the molecular mechanism and pharmacological characterizations of the novel bioactive components. Considering the progress being made in the last few years, we believe that this is a timely review to provide the scientific basis for future research aimed at studying the therapeutic potential of the newer chemical compounds derived from the *Lobelia* species.

REFERENCES

- Abed, S. A., Sirat, H. M., and Taher, M. (2016). Tyrosinase inhibition, antiacetylcholinesterase, and antimicrobial activities of the phytochemicals from Gynotroches axillaris blume. *Pak. J. Pharm. Sci.* 29, 2071–2078.
- Allen, D. D., Lockman, P. R., Roder, K. E., Dwoskin, L. P., and Crooks, P. A. (2003). Active transport of high-affinity choline and nicotine analogs into the central nervous system by the blood-brain barrier choline transporter. J. Pharmacol. Exp. Ther. 304, 1268–1274. doi:10.1124/jpet.102.045856
- Andrew, J. (1828). Observations on the use of the Lobelia inflata, in asthma and other spasmodic diseases. Glasg. Med. J. 1, 177–179.
- Bálványos, I., Kursinszki, L., Bányai, P., and Szöke, É. (2004). Analysis of polyacetylenes by HPLC in hairy root cultures of *Lobelia* inflata cultivated in bioreactor. *Chromatographia* 60, S235–S238. doi:10.1365/s10337-004-0188-x
- Bell, R. L., Eiler, B. J., Cook, J. B., and Rahman, S. (2009). Nicotinic receptor ligands reduce ethanol intake by high alcohol-drinking HAD-2 rats. *Alcohol* 43, 581–592. doi:10.1016/j.alcohol.2009.09.027
- Billen, B., Spurny, R., Brams, M., van Elk, R., Valera-Kummer, S., Yakel, J. L., et al. (2012). Molecular actions of smoking cessation drugs at α4β2 nicotinic receptors defined in crystal structures of a homologous binding protein. *Proc. Natl. Acad. Sci. USA* 109, 9173–9178. doi:10.1073/pnas.1116397109
- Brioni, J. D., Decker, M. W., Sullivan, J. P., and Arneric, S. P. (1997). The pharmacology of (-)-nicotine and novel cholinergic channel modulators. *Adv. Pharmacol.* 37, 153–214. doi:10.1016/s1054-3589(08)60950-3
- Brown, D. P., Rogers, D. T., Pomerleau, F., Siripurapu, K. B., Kulshrestha, M., Gerhardt, G. A., et al. (2016). Novel multifunctional pharmacology of lobinaline, the major alkaloid from *Lobelia* cardinalis. *Fitoterapia* 111, 109–123. doi:10.1016/j.fitote.2016.04.013
- Butler, J. E., Anand, A., Crawford, M. R., Glanville, A. R., McKenzie, D. K., Paintal, A. S., et al. (2001). Changes in respiratory sensations induced by lobeline after human bilateral lung transplantation. *J. Physiol.* 534, 583–593. doi:10.1111/j. 1469-7793.2001.00583.x
- Charlier, R., and Tondeur, R. (1950). [Extraction and pharmacological study of Lobelia suavibracteata (Haumann) alkaloids]. Arch. Int. Pharmacodyn. Ther. 83, 193–195.
- Chen, M. W., Chen, W. R., Zhang, J. M., Long, X. Y., and Wang, Y. T. (2014). Lobelia chinensis: chemical constituents and anticancer activity perspective. Chin. J. Nat. Med. 12, 103–107. doi:10.1016/S1875-5364(14)60016-9
- da Costa E Silva, L. D., Pereira, P., Regner, G. G., Boaretto, F. B. M., Hoffmann, C., Pflüger, P., et al. (2018). DNA damage and oxidative stress induced by seizures are decreased by anticonvulsant and neuroprotective effects of lobeline, a candidate to treat alcoholism. *Metab. Brain Dis.* 33, 53–61. doi:10.1007/ s11011-017-0130-1

AUTHOR CONTRIBUTIONS

QZ developed the idea and the review framework, and wrote the first draft of the manuscript. YW surveyed the literature, collected and validated the data. SZ provided the supervision of the overall work, and did the final editing. All authors contributed to corrections and editing of the subsequent revisions. All authors approve and agree with the content.

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- Divorty, N., Mackenzie, A. E., Nicklin, S. A., and Milligan, G. (2015). G proteincoupled receptor 35: an emerging target in inflammatory and cardiovascular disease. *Front. Pharmacol.* 6, 41. doi:10.3389/fphar.2015.00041
- Felpin, F.-X., and Lebreton, J. (2004). History, chemistry and biology of alkaloids from *Lobelia* inflata. *Tetrahedron* 60, 10127–10153. doi:10.1016/j.tet.2004. 08.010
- Folquitto, D. G., Swiech, J. N. D., Pereira, C. B., Bobek, V. B., Halila Possagno, G. C., Farago, P. V., et al. (2019). Biological activity, phytochemistry and traditional uses of genus *Lobelia* (Campanulaceae): a systematic review. *Fitoterapia* 134, 23–38. doi:10.1016/j.fitote.2018.12.021
- Ge, Q., Chen, L., Yuan, Y., Liu, L., Feng, F., Lv, P., et al. (2020). Network pharmacology-based dissection of the anti-diabetic mechanism of *Lobelia* chinensis. *Front. Pharmacol.* 11. doi:10.3389/fphar.2020.00347
- Gedeon, J., and Gedeon, S. (1954). [Alkaloids of lobella nicotianaefolia heyne]. *Pharm. Acta Helv.* 29, 49–52.
- Glover, E. D., Rath, J. M., Sharma, E., Glover, P. N., Laflin, M., Tonnesen, P., et al. (2010). A multicenter phase 3 trial of lobeline sulfate for smoking cessation. *Am. J. Health Behav.* 34, 101–109. doi:10.5993/ajhb.34.1.12
- Govind, A. P., Vallejo, Y. F., Stolz, J. R., Yan, J. Z., Swanson, G. T., and Green, W. N. (2017). Selective and regulated trapping of nicotinic receptor weak base ligands and relevance to smoking cessation. *Elife* 6, e25651. doi:10.7554/eLife.25651
- Hart, N., Rocha, A., Miller, D. K., and Nation, J. R. (2010). Dose-dependent attenuation of heroin self-administration with lobeline. J. Psychopharmacol. 24, 51–55. doi:10.1177/0269881108092119
- He, W., Tao, W., Zhang, F., Jie, Q., He, Y., Zhu, W., et al. (2020). Lobetyolin induces apoptosis of colon cancer cells by inhibiting glutamine metabolism. J. Cell. Mol. Med. 24, 3359–3369. doi:10.1111/jcmm.15009
- Heinzerling, K. (2013). in "Chapter 48—medication development for amphetamine dependence," in *Interventions for addiction*. Editor P. M. Miller (San Diego, CA: Academic Press), 459–465. doi:10.1016/B978-0-12-398338-1.00048-8
- Ho, L. K., Ou, J. C., Sun, M. L., and Sun, C. M. (1995). Two rare alkaloids from Pratia nummularia. *Planta Med.* 61, 567–568. doi:10.1055/s-2006-959376
- Ikeda, K., Takahashi, M., Nishida, M., Miyauchi, M., Kizu, H., Kameda, Y., et al. (2000). Homonojirimycin analogues and their glucosides from *Lobelia* sessilifolia and Adenophora spp. (Campanulaceae). Carbohydr. Res. 323, 73–80. doi:10.1016/s0008-6215(99)00246-3
- Ishimaru, K., Yonemitsu, H., and Shimomura, K. (1991). Lobetyolin and lobetyol from hairy root culture of *Lobelia inflata*. *Phytochemistry* 30, 2255–2257. doi:10. 1016/0031-9422(91)83624-T
- Joshi, S., Mishra, D., Bisht, G., and Khetwal, K. S. (2011). Essential oil composition and antimicrobial activity of *Lobelia* pyramidalis Wall. *EXCLI J.* 10, 274–279.
- Kaczmarek, F., and Steinegger, E. (1958). [Chromatographic studies on the basic fractions of Lobella cardinalis, especially of lobinaline and the new cardinalis alkaloid 2]. *Pharm. Acta Helv.* 33, 852–857.

- Kaczmarek, F., and Steinegger, E. (1959). [Paper chromatographic separation of the alkaloids of *Lobelia* inflata from lobinaline, the main alkaloid of *Lobelia* cardinalis]. *Pharm. Acta Helv.* 34, 330–333.
- Kelley, Z. D., Rogers, D. T., Littleton, J. M., and Lynn, B. C. (2019). Microfluidic capillary zone electrophoresis mass spectrometry analysis of alkaloids in *Lobelia* cardinalis transgenic and mutant plant cell cultures. *Electrophoresis* 40, 2921–2928. doi:10.1002/elps.201900220
- Kesting, J. R., Tolderlund, I. L., Pedersen, A. F., Witt, M., Jaroszewski, J. W., and Staerk, D. (2009). Piperidine and tetrahydropyridine alkaloids from *Lobelia* siphilitica and Hippobroma longiflora. *J. Nat. Prod.* 72, 312–315. doi:10.1021/ np800743w
- Klosa, J. (1953). [Lobelanine synthesis under physiological conditions in the presence of light]. *Pharmazie* 8, 1030–1032.
- Kuo, P. C., Hwang, T. L., Lin, Y. T., Kuo, Y. C., and Leu, Y. L. (2011). Chemical constituents from *Lobelia* chinensis and their anti-virus and anti-inflammatory bioactivities. *Arch. Pharm. Res.* 34, 715–722. doi:10.1007/s12272-011-0503-7
- Kursinszki, L., Ludányi, K., and Szőke, É. (2008). LC-DAD and LC-MS-MS analysis of piperidine alkaloids of *Lobelia* inflata L. (*In vitro* and *in vivo*). *Chromatographia* 68, 27. doi:10.1365/s10337-008-0628-0
- Lendle, L., and Richter, R. (1950). [Pharmacologic analysis of the emetic and asthmalytic effects of *Lobelia* tinctures]. *Klin. Wochenschr.* 28, 665–667. doi:10. 1007/BF01485598
- Li, C. Y., Zhao, L. M., Shi, X. W., and Zhang, J. D. (2014). Lobeline shows protective effects against MPTP-induced dopaminergic neuron death and attenuates behavior deficits in animals. *Exp. Ther. Med.* 7, 375–378. doi:10.3892/etm. 2013.1413
- Li, X. J., Bao, W. R., Leung, C. H., Ma, D. L., Zhang, G., Lu, A. P., et al. (2016). Chemical structure and immunomodulating activities of an α-glucan purified from *Lobelia* chinensis lour. *Molecules* 21, 779. doi:10.3390/molecules21060779
- Martin, C. A., Nuzzo, P. A., Ranseen, J. D., Kleven, M. S., Guenthner, G., Williams, Y., et al. (2018). Lobeline effects on cognitive performance in adult ADHD. J. Atten. Disord. 22, 1361–1366. doi:10.1177/1087054713497791
- Meléndez, E. N., Carreras, L., and Gijón, J. R. (1967). New alkaloid from Lobelia portoricensis urban. J. Pharm. Sci. 56, 1677–1680. doi:10.1002/jps.2600561234
- Miller, D. K., Crooks, P. A., Zheng, G., Grinevich, V. P., Norrholm, S. D., and Dwoskin, L. P. (2004). Lobeline analogs with enhanced affinity and selectivity for plasmalemma and vesicular monoamine transporters. *J. Pharmacol. Exp. Ther.* 310, 1035–1045. doi:10.1124/jpet.104.068098
- Nair, S. A., Sabulal, B., Radhika, J., Arunkumar, R., and Subramoniam, A. (2014). Promising anti-diabetes mellitus activity in rats of β -amyrin palmitate isolated from Hemidesmus indicus roots. *Eur. J. Pharmacol.* 734, 77–82. doi:10.1016/j. ejphar.2014.03.050
- Nickell, J. R., Krishnamurthy, S., Norrholm, S., Deaciuc, G., Siripurapu, K. B., Zheng, G., et al. (2010). Lobelane inhibits methamphetamine-evoked dopamine release via inhibition of the vesicular monoamine transporter-2. J. Pharmacol. Exp. Ther. 332, 612–621. doi:10.1124/jpet.109.160275
- Paz, C., Becerra, J., Silva, M., Burgosa, V., Heydenreich, M., Schmidt, B., et al. (2015). (-)-Pentylsedinine, a new alkaloid from the leaves of *Lobelia* tupa with agonist activity at nicotinic acetylcholine receptor. *Nat. Prod. Commun.* 10, 1355–1357.
- Quon, T., Lin, L. C., Ganguly, A., Tobin, A. B., and Milligan, G. (2020). Therapeutic opportunities and challenges in targeting the orphan G protein-coupled receptor GPR35. ACS Pharmacol. Transl. Sci. 3, 801–812. doi:10.1021/acsptsci.0c00079
- Raj, H., Bakshi, G. S., Tiwari, R. R., Anand, A., and Paintal, A. S. (2005). How does lobeline injected intravenously produce a cough?. *Respir. Physiol. Neurobiol.* 145, 79–90. doi:10.1016/j.resp.2004.09.001
- Roni, M. A., and Rahman, S. (2017). Lobeline attenuates ethanol abstinenceinduced depression-like behavior in mice. *Alcohol* 61, 63–70. doi:10.1016/j. alcohol.2017.01.015
- Roni, M. A., and Rahman, S. (2015). The effects of lobeline on depression-like behavior and hippocampal cell proliferation following chronic stress in mice. *Neurosci. Lett.* 584, 7–11. doi:10.1016/j.neulet.2014.10.009
- Saldanha, R. V., Cordeiro, E., Costa, E. G., and de Melo, M. E. L. O. (1959). Study of circulatory rate with lobeline (the alkaloid from *Lobelia* inflata). Arq. Bras. Cardiol. 12, 215–222.
- Sántha, E., Sperlágh, B., Zelles, T., Zsilla, G., Tóth, P. T., Lendvai, B., et al. (2000). Multiple cellular mechanisms mediate the effect of lobeline on the release of norepinephrine. J. Pharmacol. Exp. Ther. 294, 302–307.

- Santosa, M. H., Herzog, R., and Voelter, W. (1986). Antitumor activity of the hot water extract of *Lobelia* chinensis. *Planta Med.* 555, 555. doi:10.1055/s-2007-969368
- Sasaki, T., and Mikami, M. (1962). [Studies on the components of Lobelia sessilifolia lamb. II. On the periodateoxidation of a new glucofructan "sessilifolan"]. Yakugaku Zasshi 82, 1520–1522.
- Sasaki, T., Mikami, M., and Ueda, H. (1961). [Studies on the components of Lobelia sessilifolia Lamb. I. Isolation of a new glucofructan]. Yakugaku Zasshi 81, 1626–1628.
- Shen, J., Lu, X., Du, W., Zhou, J., Qiu, H., Chen, J., et al. (2016). Lobetyol activate MAPK pathways associated with G1/S cell cycle arrest and apoptosis in MKN45 cells *in vitro* and *in vivo*. *Biomed. Pharmacother.* 81, 120–127. doi:10.1016/j. biopha.2016.03.046
- Shibano, M., Tsukamoto, D., Masuda, A., Tanaka, Y., and Kusano, G. (2001). Two new pyrrolidine alkaloids, radicamines A and B, as inhibitors of alphaglucosidase from *Lobelia* chinensis lour. *Chem. Pharm. Bull.* 49, 1362–1365. doi:10.1248/cpb.49.1362
- Spurny, R., Debaveye, S., Farinha, A., Veys, K., Vos, A. M., Gossas, T., et al. (2015). Molecular blueprint of allosteric binding sites in a homologue of the agonistbinding domain of the α7 nicotinic acetylcholine receptor. *Proc. Natl. Acad. Sci.* USA 112, E2543–E2552. doi:10.1073/pnas.1418289112
- Stead, L. F., and Hughes, J. R. (2012). Lobeline for smoking cessation. Cochrane Database Syst. Rev. 2012, CD000124. doi:10.1002/14651858.CD000124.pub2
- Steinegger, E., and Egger, F. (1952). [Alkaloids of Lobelia symphilitica L.]. Pharm. Acta Helv. 27, 113–120.
- Steinegger, E., and Grutter, H. (1950). [Lurenine, a new crystallized alkaloid from Lobelia urens]. Pharm. Acta Helv. 25, 276–277.
- Steinegger, E., and Ochsner, F. (1956). [Alkaloids of Lobelia salicifolia sweet. I]. Pharm. Acta Helv. 31, 89–96.
- Subarnas, A., Oshima, Y., Sidikand Ohizumi, Y. (1992). An antidepressant principle of *Lobelia inflata L.* (Campanulaceae). J. Pharm. Sci. 81, 620–621. doi:10.1002/jps.2600810705
- Subarnas, A., Tadano, T., Kisara, K., and Ohizumi, Y. (1993a). An alphaadrenoceptor-mediated mechanism of hypoactivity induced by beta-amyrin palmitate. J. Pharm. Pharmacol. 45, 1006–1008. doi:10.1111/j.2042-7158.1993. tb05649.x
- Subarnas, A., Tadano, T., Nakahata, N., Arai, Y., Kinemuchi, H., Oshima, Y., et al. (1993b). A possible mechanism of antidepressant activity of beta-amyrin palmitate isolated from *Lobelia* inflata leaves in the forced swimming test. *Life Sci.* 52, 289–296. doi:10.1016/0024-3205(93)90220-w
- Subarnas, A., Tadano, T., Oshima, Y., Kisara, K., and Ohizumi, Y. (1993c). Pharmacological properties of beta-amyrin palmitate, a novel centrally acting compound, isolated from *Lobelia* inflata leaves. *J. Pharm. Pharmacol.* 45, 545–550. doi:10.1111/j.2042-7158.1993.tb05596.x
- Tamboli, A. M., Rub, R. A., Ghosh, P., and Bodhankar, S. L. (2012). Antiepileptic activity of lobeline isolated from the leaf of *Lobelia* nicotianaefolia and its effect on brain GABA level in mice. *Asian Pac. J. Trop. Biomed.* 2, 537–542. doi:10. 1016/S2221-1691(12)60092-6
- Terry, A. V., Williamson, R., Gattu, M., Beach, J. W., McCurdy, C. R., Sparks, J. A., et al. (1998). Lobeline and structurally simplified analogs exhibit differential agonist activity and sensitivity to antagonist blockade when compared to nicotine. *Neuropharmacology* 37, 93–102. doi:10.1016/s0028-3908(97)00142-1
- Tondeur, R., and Charlier, R. (1950). [Extraction and pharmacological studies of alkaloids of Lobelia giberroa (Hemsl.)]. Arch. Int. Pharmacodyn. Ther. 83, 91–92.
- Vartak, A. P., Gabriela Deaciuc, A., Dwoskin, L. P., and Crooks, P. A. (2010). Quinlobelane: a water-soluble lobelane analogue and inhibitor of VMAT2. *Bioorg. Med. Chem. Lett.* 20, 3584–3587. doi:10.1016/j.bmcl.2010.04.117
- Vartak, A. P., Nickell, J. R., Chagkutip, J., Dwoskin, L. P., and Crooks, P. A. (2009). Pyrrolidine analogues of lobelane: relationship of affinity for the dihydrotetrabenazine binding site with function of the vesicular monoamine transporter 2 (VMAT2). J. Med. Chem. 52, 7878–7882. doi:10.1021/jm900770h
- Wang, J., Chen, L., Qu, L., Li, K., Zhao, Y., Wang, Z., et al. (2019). Isolation and bioactive evaluation of flavonoid glycosides from *Lobelia* chinensis Lour using twodimensional liquid chromatography combined with label-free cell phenotypic assays. J. Chromatogr. A. 1601, 224–231. doi:10.1016/j.chroma.2019.04.073
- Wei, L., Wang, J., Zhang, X., Wang, P., Zhao, Y., Li, J., et al. (2017). Discovery of 2H-Chromen-2-one derivatives as G protein-coupled receptor-35 agonists. J. Med. Chem. 60, 362–372. doi:10.1021/acs.jmedchem.6b01431

- Weinges, K., Bähr, W., Ebert, W., and Kloss, P. (1972). [Norlobelanidine, the main alkaloid from *Lobelia* polyphylla Hook and Arn]. *Justus Liebigs Ann. Chem.* 756, 177–180. doi:10.1002/jlac.19727560119
- Williams, H. J., Ray, A. C., and Kim, H. L. (1987). DELTA.3-Piperideine alkaloids from the toxic plant *Lobelia* berlandieri. *J. Agric. Food Chem.* 35, 19–22. doi:10. 1021/jf00073a005
- Wu, J., Liu, Q., Yu, K., Hu, J., Kuo, Y. P., Segerberg, M., et al. (2006). Roles of nicotinic acetylcholine receptor beta subunits in function of human alpha4-containing nicotinic receptors. J. Physiol. 576, 103–118. doi:10.1113/jphysiol.2006.114645
- Yang, S., Shen, T., Zhao, L., Li, C., Zhang, Y., Lou, H., et al. (2014). Chemical constituents of *Lobelia* chinensis. *Fitoterapia* 93, 168–174. doi:10.1016/j.fitote.2014.01.007
- Zeng, X., Zhang, P., He, W., Qin, C., Chen, S., Tao, L., et al. (2018). NPASS: natural product activity and species source database for natural product research, discovery and tool development. *Nucleic Acids Res.* 46, D1217–D1222. doi:10. 1093/nar/gkx1026
- Zhang, M.-Z., Wang, J.-C., and Zhou, S.-H. (1990). Alkaloids triterpenoids of *Lobelia* davidii. *Phytochemistry* 29, 1353–1354. doi:10.1016/0031-9422(90)85468-U
- Zhang, X., Hu, P., Zhang, X., and Li, X. (2020). Chemical structure elucidation of an inulintype fructan isolated from *Lobelia* chinensis lour with anti-obesity activity on dietinduced mice. *Carbohydr. Polym.* 240, 116357. doi:10.1016/j.carbpol.2020.116357
- Zheng, G., Dwoskin, L. P., and Crooks, P. A. (2006). Vesicular monoamine transporter 2: role as a novel target for drug development. AAPS J. 8, E682–E692. doi:10.1208/aapsj080478

- Zheng, G., Dwoskin, L. P., Deaciuc, A. G., Norrholm, S. D., and Crooks, P. A. (2005). Defunctionalized lobeline analogues: structure-activity of novel ligands for the vesicular monoamine transporter. *J. Med. Chem.* 48, 5551–5560. doi:10. 1021/jm0501228
- Zheng, Q., Fang, L., Huang, X., Wang, Y., and Zhang, S. (2020). Investigation of the mechanisms of neuroprotection mediated by *Lobelia* species via computational network pharmacology and molecular modeling. *Res. Sq.* doi:10.21203/rs.3.rs-61537/v1
- Zhou, Y., Wang, Y., Wang, R., Guo, F., and Yan, C. (2008). Two-dimensional liquid chromatography coupled with mass spectrometry for the analysis of *Lobelia* chinensis Lour. using an ESI/APCI multimode ion source. *J. Sep. Sci.* 31, 2388–2394. doi:10.1002/jssc.200700685

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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