



Hyperprogressive Disease in Cancers Treated With Immune Checkpoint Inhibitors

Pan Shen, Liang Han, Xin Ba, Kai Qin and Shenghao Tu*

Department of Integrated Traditional Chinese and Western Medicine, Tongji Hospital, Tongji Medical College of Huazhong University of Science and Technology, Wuhan, China

Immunotherapy, which takes advantage of the immune system to eliminate cancer cells, has been widely studied and applied in oncology. Immune checkpoint inhibitors (ICIs) prevent the immune system from being turned off before cancer cells are eliminated. They have proven to be among the most promising and effective immunotherapies, with significant survival benefits and durable responses in diverse tumor types. However, an increasing number of retrospective studies have found that some patients treated with ICIs experience unusual responses, including accelerated proliferation of tumor cells and rapid progression of the disease, with poor outcomes. Such unexpected adverse events are termed hyperprogressive disease (HPD), and their occurrence suggests that ICIs are detrimental to a subset of cancer patients. HPD is common, with an incidence ranging between 4 and 29% in several cancer types. However, the mechanisms of HPD remain poorly understood, and no clinical predictive factors of HPD have been identified. In this review, we summarize current findings, including retrospective studies and case reports, and focus on several key issues including the defining characteristics, predictive biomarkers, potential mechanisms of HPD, and strategies for avoiding HPD after ICI treatment.

Keywords: immune checkpoint inhibitors, hyperprogressive disease, immunotherapy, predictive biomarker, pseudoprogression

INTRODUCTION

Cancer is a prevalent disease that threatens human health worldwide (Bray et al., 2018). The immune escape of cancer cells is a significant challenge in tumor treatment. In-depth studies of the molecular interactions between tumors and immune cells have provided new prospects for tumor treatments. In recent years, nivolumab, pembrolizumab, durvalumab, and atezolizumab have been approved for the treatment of non-small-cell lung cancer (NSCLC) and head and neck squamous cell carcinoma (HNSCC) (Hodi et al., 2010; Robert et al., 2015; Herbst et al., 2016; Di Pilato et al., 2019). Immune checkpoint blockade reestablishes the anti-tumor response and prevents tumor cells from evading immune surveillance by targeting specific molecules such as programmed death receptor 1 (PD-1) or its ligand (PD-L1) and cytotoxic T lymphocyte antigen 4 (CTLA-4) (Ribas and Wolchok, 2018). Immune checkpoint inhibitors (ICIs) have been referred to as a breakthrough therapy in some cancer types, and their application has led to a new era in immunotherapy. A total of ten anti-PD-1/PD-L1 antibodies, including pembrolizumab, are in immunotherapy. A total of ten anti-PD-1/PD-L1 antibodies, carrelizumab, tilelizumab, avelumab, and cemiplimab, as well as one anti-CLTA-4 antibody, ipilimumab has been approved for the clinical application in United States and China. These agents are different from conventional cytotoxic therapies and molecular targeted drugs in

OPEN ACCESS

Edited by:

Halina Was, Military Institute of Medicine, Poland

Reviewed by:

Justine Bailleul, UCLA David Geffen School of Medicine, United States Abir Kumar Panda, National Institutes of Health (NIH), United States Gaurisankar Sa, Bose Institute, India

*Correspondence:

Shenghao Tu shtu@tjh.tjmu.edu.cn

Specialty section:

This article was submitted to Pharmacology of Anti-Cancer Drugs, a section of the journal Frontiers in Pharmacology

> Received: 09 March 2021 Accepted: 22 June 2021 Published: 05 July 2021

Citation:

Shen P, Han L, Ba X, Qin K and Tu S (2021) Hyperprogressive Disease in Cancers Treated With Immune Checkpoint Inhibitors. Front. Pharmacol. 12:678409. doi: 10.3389/fphar.2021.678409

1

TABLE 1 | Studies on HPD after ICI treatment.

References	Tumor types	Therapy of ICIs	HPD criteria	Previous therapies	HPD incidence	HPD predictors/factors	Study desigr
Champiat et al. (2017)	Multiple cancer types	PD-1/PD-L1 inhibitors	1. RECIST 1.1 progression 2. TGR ≥ 2-fold increase	Chemotherapy/ radiotherapy/targeted therapy/immunotherapy	9% (12/131)	Older age (≥65) (<i>p</i> = 0.007)	Phase I trials
Kato et al. (2017)	Multiple cancer types (Melanoma, 33%. NSCLC, 25%)	PD-1/PD-L1/CTLA-4 inhibitors, other investigational agents	 TTF < 2 months Progression pace > 2x Tumor burden increase >50% compared with baseline 	Chemotherapy/ radiotherapy/targeted therapy/immunotherapy<	3.9% (6/155)	 EGFR alteration (ρ = 0.005) MDM2/MDM4 amplification (ρ = 0.007) 	Genomic Analysis
Saâda-Bouzid et al. (2017)	R/M HNSCC	PD-1/PD-L1 inhibitors	TGK _R ≥ 2	_	29.4% (10/34)	 Locoregional recurrence (p = 0.008) Presence of cervical nodes at diagnosis 	Retrospective
Weiss et al. (2017)	Multiple cancer types (Melanoma, 22%)	PD-1/PD-L1/CTLA-4 inhibitors	-	Chemotherapy/targeted therapy/radiotherapy/ immunotherapy	10.7% (6/56)	Chromosomal instability quantification in plasma of cell- free DNA	Retrospective
Faure et al. (2018)	Anorectal malignant melanoma	PD-1 inhibitors	-	Chemotherapy	Case report	A role of monocytes	Retrospective
Ferrara et al. (2018)	NSCLC	PD-1/PD-L1 inhibitors	1. RECIST 1.1 progression 2. Disease progression with change in TGR >50%	Chemotherapy/ radiotherapy	13.8% (56/406)	Metastatic sites >2 (p = 0.006)	Retrospective
Matos et al. (2018)	Multiple cancer types	PD-1/PD-L1/CTLA-4 inhibitors	TTF <2 months and increase in measurable lesions of 10 mm plus increase of ≥40% in target tumor burden compared with baseline or increase ≥20% in target tumor burden plus multiple new lesions	_	15.4% (33/214)	_	Phase I trials
Boland et al. (2019)	Epithelial ovarian cancer	PD-1/PD-L1/CTLA-4/LAG3 inhibitors	_	_	33.7% (30/89)	1. Neutrophil-to-lymphocyte ratio (NLR) > 4 ($p = 0.017$) 2. Liver parenchymal metastases ($p = 0.001$)	Retrospective
Costantini et al. (2019)	Advanced NSCLC	PD-1 inhibitors	Patients receiving less than three injections of nivolumab	Radiotherapy	19.5% (57/292)	$PS \ge 2$, Shorter duration of treatment before nivolumab ($p < 0.0001$)	Retrospective
Ji et al. (2019)	Malignant tumors of digestive system	PD-1/PD-L1/CTLA-4 inhibitors	TGK _R ≥ 2	_	20% (5/25)	_	Retrospective
Kanjanapan et al. (2019)	Multiple cancer types	PD-1/PD-L1/CTLA-4 inhibitors	 RECIST 1.1 progression TGR ≥ twofold increase 	_	7% (12/182)	Female gender	Phase I trials
Lo Russo et al. (2019)	NSCLC	PD-1/PD-L1/CTLA-4 inhibitors	 RECIST 1.1 progression and at least 3 of: 1. TTF <2 months 2. ≥50% increase of sum of target lesions major diameters 3. At least two new lesions in an organ already involved 4. ECOG PS ≥ 2 	_	25.7% (39/152)	Clustered macrophages with epithelioid morphology and colocalization of CD163, PD-L1, and CD33 markers	Retrospective
Sasaki et al. (2019)	Advanced gastric cancer	PD-1 inhibitors	TGK ≥ 2	Chemotherapy/ radiotherapy	21.0% (13/62)	Neutrophil count increased ($p = 0.002$) Increased CRP levels ($p = 0.006$)	Retrospective
		PD-1/CTLA-4 inhibitors	TGR \geq 2-fold increase		_	Previous radiotherapy treatment (Continued on fe	Case series ollowing page)

TABLE 1 (Contir	TABLE 1 (Continued) Studies on HPD after ICI treatment.	reatment.					
References	Tumor types	Therapy of ICIs	HPD criteria	Previous therapies	HPD incidence	HPD predictors/factors	Study design
Wong et al. (2019)	hepatocellular carcinoma (HCC)			Chemotherapy/ radiotherapy			
Petrioli et al. (2020)	Urothelial transitional cancer Nivolumab (UTC), lung adenocarcinoma, and HCC	Nivolumab	Two-fold or greater increase in the TGR	Chemotherapy	6.4% (3/47)	6.4% (3/47) 1. Higher LDH serum levels 2. NLR >3	Case series
Kim C. G. et al. (2019)	NSCLC	PD-1 or PD-L1 inhibitors	Post-TGR/pre-TGR > 2 or post- TGK/pre-TGK > 2	Chemotherapy/targeted therapy	19.0% (45/237)	TIGIT + PD1+ CD8 ⁺ T cells increased	Retrospective
Lu et al. (2019)	Gastric cancer, esophageal cancer, colorectal cancer	PD-1 or PD-L1 inhibitors monotherapy or combined with CTLA-4 inhibitor	Post-TGK/pre-TGK ≥ 2	Chemotherapy/ radiotherapy	8.9% (5/56)	Lower serum MCP-1 and leukocyte inhibition factor levels	Prospective
Arasanz et al. (2020)	NSCLC	PD-1 and/or PD-L1 inhibitors	TGR ≥ 2-fold increase	Chemotherapy/ radiotherapy	17.9% (10/56)	Higher levels of CD28 ⁻ CD4 ⁺ T lymphocytes	Prospective
Refae et al. (2020)	NSCLC, head and neck cancer, melanoma, RCC, and others	PD-1 or PD-L1 inhibitors alone	Post-TGR/pre-TGR > 2	I	13.8% (11/80)	 Older age (≥70)<<i>p</i> = 0.0025) Variations with rs2282055 (PD-L1) and rs1870377 (VEGFR2) 	Retrospective
Ruiz-Patiño et al. (2020)	NSCLC	PD-1 or PD-L1 inhibitor monotherapy or combined with CTLA-4 inhibitor or chemotherapy	Two-fold increase in TGR	1	19.8% (44/222)	1	Retrospective

many respects; their effects include delayed tumor regression, long-term survival benefits, and pseudoprogression (Soria et al., 2018). However, some patients experience rapid disease progression and deterioration. This particular type of accelerated tumor progression has been described as hyperprogressive disease (HPD) (Champiat et al., 2017; Kato et al., 2017; Saâda-Bouzid et al., 2017; Weiss et al., 2017; Faure et al., 2018; Ferrara et al., 2018; Matos et al., 2018; Boland et al., 2019; Costantini et al., 2019; Ji et al., 2019; Kanjanapan et al., 2019; Kim C. G. et al., 2019; Lo Russo et al., 2019; Lu et al., 2019; Sasaki et al., 2019; Wong et al., 2019; Arasanz et al., 2020; Petrioli et al., 2020; Refae et al., 2020; Ruiz-Patiño et al., 2020) (Table 1). Thus, ICI immunotherapy may be not only ineffective in some patients but also harmful. Several doctors and investigators have studied this phenomenon in practice and found that a subset of advanced cancer patients treated with ICIs are more likely to experience a severe decline in their quality of life and poor prognosis. According to previously published studies, the incidence of HPD ranges from about 4 to 29%. In addition, only 15-40% of patients benefit from ICIs (Borghaei et al., 2015), and some experience relatively long-lasting adverse responses. The emergence of HPD poses a new challenge to the current approach used to evaluate the efficacy of ICIs. Various diagnostic tools and criteria are used to assess disease progression, including the immune response evaluation criteria in solid tumors (iRECIST) (Seymour et al., 2017), immune-related response evaluation criteria in solid tumors (irRECIST) (Nishino et al., 2013), and immune-related response criteria (irRC) (Wolchok et al., 2009).

Currently, there is no consensual definition of HPD. Previous studies have explored its occurrence and biomarkers and the molecular mechanisms underlying the role of ICIs in HPD. This review aims to discuss unexplored questions and mechanisms relevant to HPD and summarizes the current data on diagnostic tools and potential biomarkers for HPD.

EVIDENCE OF HYPERPROGRESSIVE DISEASE AND CONTROVERSY

In 2016, Champiat et al. first reported HPD with an incidence of 9% (12/131) in a phase 1 trial of anti-PD-1/PD-L1 inhibitors (Champiat et al., 2017). In this study, HPD was not associated with an increased tumor load at baseline or specific types of tumors but was found to occur more frequently in patients over 65 years of age. A retrospective analysis of 34 patients with HNSCC by Saada-Bouzid et al. reported a maximum incidence of 29% (Saâda-Bouzid et al., 2017). In the clinical trials CheckMate 057 (28), CheckMate 227 (Hellmann et al., 2018), and CheckMate 141 (Ferris et al., 2016) that compared ICIs with standard chemotherapy, the crossing of overall and disease-free survival curves was proposed as evidence for HPD. In recent years, a study by Lahmar et al. found that 10% of NSCLC patients developed HPD (Lahmar et al., 2016). In a cohort of 242 NSCLC patients, the incidence of HPD was 16% (Ferrara et al., 2017). Although the clinical diagnostic criteria for HPD varied between studies, tumor growth was compared

before and after the initiation of immunotherapy within a short time.

As HPD clinical studies are mostly retrospective, whether HPD is an independent pattern of post-treatment responses to ICIs remains controversial. The main dispute is over whether HPD is a natural process of tumors or an accelerated growth process that occurs after ICI treatment (Pearson and Sweis, 2019). In most trials, there is no reference data on the tumor growth rate (TGR) before initiation of ICIs. Moreover, the crossing of the two survival curves is considered to occur because chemotherapy works more efficiently than ICIs, rather than because of HPD itself. Gandara et al. reported a similar proportion of fast-progressing patients in a cohort of 850 NSCLC participants treated with docetaxel or atezolizumab; they suggested that HPD may be caused by a poor prognosis, not by the immunotherapy (Gandara et al., 2018). Moreover, despite accumulating data on HPD, its definition is not universal. Future studies should assess the optimal HPD criteria to identify patients who cannot benefit from ICI treatments and those who are most likely to benefit from these costly and potentially toxic treatments. It is important to collect imaging treatment and tumor growth kinetic data before ICI therapy and to distinguish HPD from an inherently aggressive disease or pseudoprogression.

DEFINITION AND DIAGNOSIS OF HYPERPROGRESSIVE DISEASE

In contrast to pseudoprogression, tumor growth is not caused by increased inflammation but by the specific action of ICIs as enhancers of tumor progression. It is essential to detect and distinguish progression, pseudoprogression, and HPD at an early stage. Currently, the assessment and diagnosis of HPD are mainly based on parameters related to pretreatment tumor kinetics and early changes after the start of immunotherapy, including the TGR, tumor growth kinetics (TGK), and time to treatment failure (TTF). HPD was first described in case reports and retrospective studies involving patients with accelerated tumor growth after ICI treatment. According to RECIST 1.1, HPD can be defined by a ≥2-fold increase in TGR after immunotherapy (Kanjanapan et al., 2019), whereas Ferrara et al. established a different criterion of the TGR increasing by 50% (Ferrara et al., 2018). The TGR is a ratio of the change in tumor size over a given time interval (Gomez-Roca et al., 2011; Ferté et al., 2014) and is a predictor of overall survival in clinical practice (Ten Berge et al., 2019). Assessment of the TGR is based on RECIST, which assesses changes in the sum of the largest diameter of target lesions at multiple time points and uses a natural logarithm for correction. The lack of an association between RECIST and changes in TGR before and after treatment suggests that RECIST provides limited information (Gomez-Roca et al., 2011). Therefore, RECIST 1.1 is not the most accurate way to evaluate the efficacy of immunotherapy. In fact, the use of RECIST 1.1 was found to underestimate the positive response rate in 160 NSCLC patients and indicated a 15% treatment benefit rate in 655 melanoma patients treated with ICIs.

The evaluation of TGK is similar to that of TGR: changes in the sum of the largest diameter of the target lesion are also measured, but no logarithm correction is performed. HPD is defined by a \geq 2-fold increase in the TGK of the target lesion at the time of the first evaluation compared with before ICI treatment (Saâda-Bouzid et al., 2017). According to RECIST, HPD assessment requires tumor burden data to be obtained at earlier time points for the first TGR assessment, and two computed tomography scan evaluations are needed. However, this HPD evaluation method is limited to target lesions of the tumor and does not consider the appearance and changes of nontarget lesions. In actual clinical practice, TGR data before immunotherapy cannot be obtained, and the rate cannot be distinguished by basic imaging analysis of changes in tumor size. Patients with HPD have no clinically detectable tumor growth prior to their first treatment, and the maximum tumor single diameter is unavailable; therefore TGR and TGK cannot not be assessed. TGR- and TGK-based methods cannot be used to evaluate new lesions in the assessment of tumor growth. Importantly, these methods use only radiological criteria, which may lead to misclassification of response patterns. Previous studies have used a combination of clinical and radiological criteria. A TTF of less than 2 months is also used as a surrogate indicator for evaluating HPD. Kato et al. (2017) provided an additional two criteria: an increase of more than 50% of tumor burden compared with pre-immunotherapy imaging, and a 2-fold or greater increase in progression pace. Kato et al. found that melanoma patients had longer TTF compared with patients with other tumor types; this may suggest either that patients with melanoma are less likely to develop HPD or that TTF is not an effective diagnostic marker for HPD.

Zuazo-Ibarra et al. quantified the number of circulating senescent CD4⁺ T cells (Tsens) before ICI treatment. Increased numbers of Tsens before immunotherapy indicate a response, whereas a decrease in the number of Tsens after the first treatment cycle indicates a good response. The decrease may be due to G1 phase withdrawal or tumor cell recruitment from the blood. Conversely, proliferation of Tsens suggests progression. The authors concluded that extensive validation is necessary to apply these findings in NSCLC and more widely in clinical practice (Zuazo et al., 2018). Boeri et al. (2018) identified potential prognostic value in the immune environment by analyzing microRNAs in plasma samples. In general, technologies based on liquid biopsy are expected to provide reliable methods for stratification and disease tracking for clinical applications, including future real-time applications. Combinations of several methods provide the most valuable assessments. Despite the relative inaccuracy of HPD diagnosis, the current results are still informative for further studies.

WHAT IS PSEUDOPROGRESSION?

In brain cancer, the concept of tumor pseudoprogression was first proposed in patients treated with the non-immunotherapeutic agent temozolomide. However, this phenomenon was not accompanied by real tumor progression, and the brain tumor may have grown before temozolomide treatment (Brandsma and van den Bent, 2009). Although pseudoprogression has rarely been observed in patients treated with conventional cytotoxic drugs, it occurs relatively frequently during ICI therapy. Pseudoprogression in patients treated with ICIs was first identified in a melanoma study of the anti-CTLA4 inhibitor ipilimumab (Wolchok and Saenger, 2008) and then in subsequent studies of the anti-PD-1 inhibitors pembrolizumab and nivolumab (Chiou and Burotto, 2015). Patients with pseudoprogression and HPD have completely different outcomes. Moreover, pseudoprogression can either occur within the first 12 weeks of treatment or be delayed. Given that beneficial treatments are often discontinued in initial trials because of pseudoprogression, it is essential to distinguish between pseudoprogression and HPD. Pseudoprogression is not true tumor progression but radiographic growth pathologically characterized by infiltration, edema, and necrosis of immune cells surrounding the tumor. Hodi et al. considered pseudoprogression to be an increase in the tumor burden of at least 25% with no representative progressive disease detected in subsequent assessments (Hodi et al., 2016). In addition to the inflammatory response induced by tumor immune cell invasion, delayed immune responses may also play a part in pseudoprogression, especially in patients with tumor regression after pseudoprogression. There have been several studies on the incidence of pseudoprogression when ICIs are used to treat solid tumors, although this is relatively rare. In melanoma, an unusual immune response or pseudoprogression followed by a delayed response was observed in 3.7-15.8% of patients treated with ICIs (Hodi et al., 2008; Ribas et al., 2012; Millward et al., 2013). The incidences of pseudoprogression in patients with NSCLC, urothelial carcinoma, HNSCC, and mesothelioma have been reported to be 0.6-5.8, 1.5-7.1, 1.8, and 6.9%, respectively. Although pseudoprogression mostly occurs in patients receiving single checkpoint inhibitors, it has also been observed in patients receiving dual immunotherapy. In a case report, a patient with microsatellite unstable high metastatic colorectal cancer treated with the combination of a PD-L1 antagonist and OX40 agonist showed pseudoprogression, with a 163% increase in the baseline tumor burden (Chae et al., 2017). Tumor atrophy was subsequently observed, and the patient's condition was stable. This large increase in tumor size presents a challenge in differentiating pseudoprogression from authentic progression (Wolchok et al., 2009). Therefore, clinicians should depend on other information to accurately assess tumor status. To address this issue, new criteria, including irRC, irRECIST, and iRECIST, were applied to differentiate true progression from pseudoprogression. The patient was re-examined 4 weeks after the diagnosis of an underlying progressive disease to ensure that it was not spurious progression (Seymour et al., 2017).

Moreover, the analysis of cell-free circulating tumor DNA (ctDNA) levels might be used as an effective tool in distinguishing progressive disease, HPD, and pseudoprogression (Lipson et al., 2014; Cabel et al., 2018). It evaluates the alternation in ctDNA levels to separate cell-free DNA from plasma for liquid biopsy, thereby detecting the changes of tumor-specific copy number and genome instability number. Pseudoprogression showed as a decrease in genome instability in ctDNA, unlike HPD (Jensen et al., 2019). In a study of 125 patients with melanoma received ICI treatment (Lee et al., 2018), ctDNA profiles distinguished

pseudoprogression and HPD with high sensitivity (90%) and specificity (100%).

It is believed that pseudoprogression has a favorable prognosis and should be identified as soon as possible to avoid delaying the disease owing to the early interruption of treatment. At the same time, it is important to be aware of the possibility of real tumor progression (**Figure 1**).

BIOLOGICAL AND CLINICOPATHOLOGICAL FACTORS OF HYPERPROGRESSIVE DISEASE

It is essential to identify potential predictive factors of HPD. This can enable the adverse effects caused by ICIs to be avoided and has important clinical significance in prolonging the survival period and quality of life of patients. Champiat et al. (2017) and Saâda-Bouzid et al. (2017) found no association between HPD and baseline tumor load, previous treatment line, tumor histology, type of immunotherapy, or number of metastatic sites. To date, five clinical variables—aging, female sex, higher serum lactate dehydrogenase (LDH) concentration, metastasis burden, and local recurrence of the cells in the irradiation area—have been identified as potentially associated with HPD, as have specific genomic mutations including *MDM2/MDM4* amplification and *EGFR* aberrations.

In the study reported by Champiat et al., older age appeared to be related to the occurrence of HPD. Patients with HPD were significantly older than those without HPD, with 19% of patients over 65 showing HPD compared with 5% of patients younger than 64. These results indicate that older patients benefit less from ICIs than younger people. T cell immunity declines with age because of changes in T cell number, diversity, phenotype, and function (Fulop et al., 2017). In addition, T cell signaling through T cell receptors has been found to decrease with age (Solana et al., 2012; Goronzy and Weyand, 2013). However, this was not observed in two other studies. Therefore, the specific relationship between HPD and aging is not completely clear.

Saada-Bouzid et al. found that HPD was related to radiotherapy. In a previous anti-PD-1/PD-L1 study, 50% of patients with regional recurrence had HPD, whereas only 6.25% of patients without regional recurrence had HPD. Almost all cases of HPD occurred in patients with recurrence in the irradiated area, but the underlying mechanism of this phenomenon is unclear. Ferrara et al. found that HPD was more frequent in NSCLC patients with more than two metastatic sites (Weiss et al., 2017). Kanjanapan et al. found that the incidence of HPD in women was significantly higher than that in men; this was the only study in which HPD was shown to be associated with gender (Kanjanapan et al., 2019). Several studies found that regional recurrence in HNSCC was associated with a higher risk of HPD. Sasaki et al. reported that liver metastases, an early increase in neutrophil counts and C reactive protein levels, and a large sum of target lesion diameters at baseline were associated with HPD risk (Sasaki et al., 2019). The local recurrence of HNSCC and metastasis of most other types of cancer are associated with poor prognosis, because in these cases



the tumor has successfully acquired the characteristics necessary for survival at the primary site and metastasis to a distant location. Therefore, it is difficult to distinguish the relevant prognostic factors and predictors.

Kato et al. analyzed genomic mutations (155 patients) as potential genomic markers associated with immunotherapy and HPD using next-generation sequencing (Kato et al., 2017). A favorable clinical outcome (TTF \geq 2 months) was observed in patients with several genetic alternations, including mutations in *TERT* [odds ratio (OR): 0.42; p = 0.07], *PTEN* (OR: 0.28; p = 0.10), *NF1* (OR: 0.15; p = 0.07), and *NOTCH1* (OR: <0.19; p = 0.02). Conversely, EGFR (OR: 10.2; p = 0.002), MDM2/4 (OR: > 11.9; p =0.001), and DNMT3A (OR, 9.33; p = 0.03) alterations were related to worse outcomes (TTF < 2 months). Six patients with alterations exhibited an HPD phenotype, and all had a TTF < 2 months, whereas 20% of patients with EGFR alterations exhibited an HPD phenotype. Of five patients with DNMT3A alterations, only one was radiologically evaluable and did not have an HPD phenotype. In a study by Singavi et al., the incidence of HPD in patients with EGFR amplification was 50% (Singavi et al., 2017). EGFR mutations are likely to be associated with the upregulation of PD-1/PD-L1, which can activate immune escape. If the results obtained from larger cohorts are consistent with the current findings, MDM2/4 and EGFR alternations could potentially be used as reliable HPD predictive biomarkers.

Lo Russo et al. (2019) did not find significant differences in tumor-infiltrating T cells of the following types: CD4+/CD8+ lymphocytes, regulatory T cells (Tregs), peritumoral and stromal myeloperoxidase myeloid cells, and PD-1+ and PD-L1+ immune cells. However, HPD was associated with the density of myeloperoxidase myeloid cells within the tumor and inversely correlated with PD-L1 expression in tumor cells. Some investigators focused on CD8⁺ T cells in the peripheral blood to find potential predictors. Studies have found that number of effector CD8⁺ T cells (CCR7–CD45RA–) decreased (Jenkins et al., 2018), whereas number of exhausted tumor-reactive CD8⁺ T cells (TIGIT + PD-1+) increased in HPD patients with NSCLC (Huang A. C. et al., 2017). The results suggest that the exhaustion of CD8⁺ T cells is one of the potential mechanisms that triggers the acceleration of tumor growth with ICIs treatment. The severity of T cell exhaustion can be predictors for HPD. In addition, HPD patients also showed a significant upregulation in the number of CD28⁻CD4⁺ cells after immunotherapy (Arasanz et al., 2020). The number of CD62LlowCD4+ effector Th1 cells was significantly higher in the peripheral blood of patients with NSCLC before treatment with PD-1 inhibitor, while a decrease in CD62LlowCD4+ T cells was related with acquired resistance (Kagamu et al., 2020), indicating that CD4⁺ T cell immunity could be a powerful predictor in HPD.

Weiss et al. and Jensen et al. performed genome-wide sequencing of plasma/serum-derived cell-free DNA and found that genomic copy number instability (CNI) could help to determine HPD (Weiss et al., 2017; Jensen et al., 2019); however, their study included a small number of patients. Further prospective research should be conducted to determine whether CNI could be a novel marker for HPD. In an exploratory study of HPD, NSCLC patients with an increased proportion of depleted T cell subtypes and a decreased proportion of effector T cells showed a higher incidence of HPD and worse prognosis. These results suggest that T cell subtypes help predict the occurrence of HPD (Ferrara et al., 2018). Lo Russo et al. (2019) compared patient responses and histopathological and molecular expression patterns in 35 patients with advanced NSCLC after immunotherapy. The results showed that HPD was associated with the number of myeloperoxidase-positive myeloid cells in the tumor and inversely associated with PD-L1 expression in tumor cells. In patients with HPD, the presence of CD163 + CD33+PD-L1+ macrophages with an epithelioid morphology was identified more frequently than in patients without HPD, suggesting that these macrophages are important in the HPD process. In a study, infiltration of epithelioid-shaped macrophages in tumor tissues was observed in 104 NSCLC patients who experienced HPD (Lo Russo et al., 2019). Tunali et al. established a clinical prediction model by combining extensive clinical-pathological, laboratory, and imaging data to predict the occurrence of HPD in NSCLC patients (Tunali et al., 2019).

Owing to the limited size and scope of previous cohort studies, we cannot draw reliable conclusions to accurately assess risks and benefits in specific patient groups. Furthermore, the current research hypothesis needs to be further verified. Tumor types, stages, and other covariates should be controlled, especially in studies that investigate covariates and their associations with patient response or resistance to immunotherapy.

THE MAIN HYPOTHESES REGARDING THE MECHANISMS UNDERLYING HYPERPROGRESSIVE DISEASE

At present, the molecular mechanisms of HPD remain elusive. Several studies have explored multiple hypotheses or mechanisms of HPD from different perspectives (**Figure 2**; **Table 2**). These mechanisms may act independently or be complementary. It is of great significance to clarify the molecular mechanism of HPD. HPD can be caused by a variety of factors, including the characteristics of tumor cells, the status of the patient's immune system, and the patient's current or previous treatment history.

Upregulation of PD-1+ Tregs

In the context of infection or cancer, contra-suppression or immune compensation is a self-stabilizing mechanism. Immunosuppressive factors such as Tregs can maintain antiinfection or anti-tumor immunity and reduce the adverse effects of ICIs (Lehner, 2008; Barnaba and Schinzari, 2013). Studies have confirmed that inhibiting the expression of PD-1 in mice upregulates other immune checkpoints, including CTLA-4, LAG3, and TIGIT, or induces the activation and proliferation of Tregs, resulting in impaired immune killing ability (Ellestad et al., 2014; Huang R. Y. et al., 2017). FOXP3 is a classical marker of Tregs, which participate in inducing immune tolerance. Nair et al. found that pembrolizumab inhibited peripheral Treg differentiation and reduced FOXP3 expression through the mTOR pathway in vitro (Sasidharan Nair et al., 2020). A previous study analyzed gastric cancer tumor tissues before and after ICI treatment and found increased numbers of infiltrating effector Tregs (Ki67 + Tregs) in tissues from HPD patients, suggesting that these Tregs may be activated by ICI therapy (Kamada et al., 2019). Compared with patients without HPD, the number of tumor-infiltrating Ki67 + Tregs was significantly increased in HPD patients. In an in vitro gastric cancer study, it was observed that when PD-1 was knocked out or the binding of PD-1 to its ligand was inhibited in Tregs, Tregs

could proliferate and promote anti-tumor immune cells and tumor suppression. In mouse models, the results also support a similar effect of Tregs in HPD. From the above studies, PD-1 may mediate the occurrence of HPD, leading to the inhibition of anti-tumor immunity.

T Cell Exhaustion

T cell dysfunction is defined as T cell exhaustion, in which the ability to recognize and eliminate antigens is weakened, and inhibitory receptors including PD-1, T cell immunoglobulin and mucin domain-containing protein 3 (TIM3), TIGIT, and LAG3 are upregulated. In mouse models and progressive cases, TIM-3 has been found to be upregulated (Koyama et al., 2016). In addition, the proportion of TIM-3+ cells was proportional to the duration of ICI treatment. No increase in TIM-3+ cells was detected in control cases. Therefore, overexpression of TIM-3 is likely to be a key mechanism of resistance to PD-1 treatment (Shayan et al., 2017). After failure of anti-PD-1 treatment, TIM-3 antibodies can provide a survival benefit. Similarly, CTLA-4 and LAG3 on cytotoxic CD8⁺ T cells were increased as a result of PD-1 blockade in a model of ovarian cancer (Huang R. Y. et al., 2017). With expression of the above compensatory receptors, CD8⁺ T cells show serious dysfunction in cytokine production, proliferation, and migration. In a mouse virus infection model, Odorizzi et al. found that cytotoxic CD8⁺ T cells were depleted, with large numbers of inhibitory receptors such as LAG3 and TIGIT on their cell surface (Odorizzi et al., 2015).

Increased Tsens

In a study of NSCLC patients with low baseline numbers of senescent Tsens treated with anti-PD-1/PD-L1 agents, those with



TABLE 2 | Potential mechanisms of HPD in different types of tumors.

Mechanisms	Tumor types		
Upregulation of PD-1 + Tregs	Multiple cancer types		
T Cell exhaustion	Metastatic ovarian cancer, Head and neck cancer		
Increased Tsens	NSCLC		
MDM2/4 amplification and EGFR mutations	Multiple cancer types		
The effects of Fc fragment of macrophages and ICIs	Multiple cancer types		
Imbalance of immunosuppressive cytokines and factors	Gastric cancer, Ovarian cancer, NSCLC		
Increase in ILC3	Breast cancer, Colon cancer		
The effects of neoantigens	Multiple cancer types		
Circulating LDH levels	UTC, Lung cancer, HCC		
Radiotherapy	HCC		

Tsens elevated by 12.4% also had HPD. Conversely, patients who showed a 14.4% decrease in Tsens experienced tumor regression (Zuazo-Ibarra et al., 2018). The results indicate that Tsens numbers in patients prior to immunotherapies could predict the risk of HPD. Tsens baseline numbers may represent a preexisting large pool of antigen-specific central and effector T cells with potential anti-tumor capacities. The decreases in Tsens following antibody administration may indicate the mobilization of Tsens from peripheral blood to secondary lymphoid organs/tumor sites.

MDM2/4 amplification and EGFR Mutations

Studies involving multiple tumor types have shown that oncogenic activation, such as that resulting from MDM2/4 amplification, is associated with the occurrence of HPD (Ferté et al., 2014). In malignant tumors, a study using secondgeneration sequencing found that the amplification rate of MDM2/4 was about 3.9% (6/155), and that the TTF of all patients with MDM2/4 amplification receiving immunotherapy was less than 2 months (Champiat et al., 2018). MDM2/4 blocks the p53 trans-activation domain and promotes proteasomal ubiquitin-dependent degradation of p53 (Wang Q. et al., 2018). IFN- γ can increase the expression of MDM2/4 and further inhibit p53 activity (Zhou et al., 2009; Zhao et al., 2014). However, loss of p53 activity is an essential driver of oncogenesis. ICIs can increase the production of IFN- γ at the tumor site, and it has been speculated that the MDM2/4-IFN- $\nu/p53$ axis may mediate the occurrence of HPD (Peng et al., 2012). In about 50% of tumors, p53 is mutated or lost and inactivated. The other 50% retain wild-type p53, the normal regulation of which is disrupted by overexpression of MDM2 (Momand et al., 1998). Anti-PD-1/PD-L1 agents can activate JAK/STAT signaling, resulting in increased expression of interferon regulator 8 (IRF-8) (Schindler et al., 2007; Waight et al., 2013). IRF-8 can bind to the MDM2/4 promoter and promote the expression of MDM2/4, but the cascade may not have a significant impact when MDM2/4 is not amplified (Zhao et al., 2014). The amplification rate of MDM2 in malignant lung, skin, and bladder tumors is lower, whereas that in sarcoma is higher, consistent with the primary resistance of sarcoma to ICIs [59]. If MDM2 amplification does cause ICI resistance or HPD, the combination of ICIs and MDM2 inhibitors may be a promising strategy to overcome resistance and HPD. In

addition to MDM2/4, the activation of other oncogenic signals may also contribute to HPD after ICI therapy. Xiong et al. found increased activity of IGF-1, ERK/MAPK, and PI3K/AKT in patients with HPD after ICI therapy, whereas tumor suppressor genes including *TSC2* and *VHL* were not expressed (Xiong et al., 2018). The activation of EGFR is usually accompanied by upregulation of PD-1/PD-L1 or CTLA-4 to promote tumor immune escape (Akbay et al., 2013). The objective response rate of anti-PD-1 treatment in patients with *EGFR* mutations is relatively low at about 3.6% (Gainor et al., 2016). This may be related to the activation of EGFR, which can promote the stability of PD-L1 and prevent it from being easily degraded (Li et al., 2016). However, the mechanisms linking *EGFR* mutations with HPD remain unclear.

The Effects of Fc Fragment of Macrophages and Immune Checkpoint Inhibitors

Immune microenvironments are closely related to HPD. NSCLC tumor tissues from patients (patient-derived xenografts, PDX) were transplanted into athymic nude or severe combined immunodeficient (SCID) mice. Nivolumab or nivolumab F (ab)2 fragments were administered to mice at the nodule. Nivolumab promoted tumor growth, whereas no substantial tumor growth or HPD was observed in mice treated with the F (ab)2 fragment of nivolumab (that is, lacking the Fc fragment) (Lo Russo et al., 2019). Further studies found that M2 macrophage aggregation only occurred in lesions after nivolumab treatment but not after the injection of nivolumab F (ab)2 fragments. In another study, compared with PDX SCID mice with wild-type EGFR, TGR and tumor cell dissemination were significantly increased in PDX SCID mice with mutated versions of EGFR after nivolumab therapy (Lo Russo et al., 2019). Similarly, when mice with EGFR-mutated PDXs were treated with nivolumab F (ab)2 fragments, there was no significant evidence of HPD or tumor dissemination. It has been suggested that the occurrence of HPD is related to the interaction between macrophages and the Fc portion of nivolumab. It is also important to consider the interaction between the different Fc regions used in various PD-1 antibodies and the distinct Fc receptor variants. In a study of malignant melanoma (Dahan et al., 2015), knocking out the Fcy receptor (FcyR) in mice enhanced the anti-tumor effect of the

anti-PD-1 antibody, directly showing a correlation between $Fc\gamma R$ and ICIs. Lo Russo et al. suggested that $Fc\gamma R$ IIb has a detrimental effect on anti-PD-1 immunotherapy efficacy in humans, resulting in HPD (Lo Russo et al., 2019). Zhang et al. designed two anti-PD-1 monoclonal antibodies with the same specificity but different Fc sequences; they found that $Fc\gamma R$ I induced immune tolerance by regulating inflammatory cytokines and played a part in the production of M2 macrophages to promote tumor growth (Zhang et al., 2018). This could be addressed by destabilizing the FcR interaction and modifying the current immunotherapy strategy.

Imbalance of Immunosuppressive Cytokines and Factors

Tumor-derived exosomes have been shown to induce PD1+ macrophages to produce IL-10 and inhibit the function of CD8+T cells (Wang F. et al., 2018). In another study, Lamichhane et al. found that inhibiting the expression of PD-1 promoted the secretion of IL-10 by dendritic cells (DCs) and further increased the expression of PD-L1 in DCs (Lamichhane et al., 2017). The analysis of tumor tissues from 104 NSCLC patients with HPD revealed a large number of M2-type PD-L1+ macrophages, which secrete IL-10 to mediate the occurrence of HPD through the depletion of the PD-1 antibody (Kanazu et al., 2018). Inhibition of PD-1 expression also increases the serum angiogenin two concentration, which can increase the number of M2 macrophages and promote tumor metastasis, angiogenesis, and immunosuppression (Wu et al., 2017). The immuneresistance of tumors can be enhanced by IFN- γ via elevated expression of PD-L1 in cancer cells (Teng et al., 2018). The activation of JAK1/STAT3 enables binding of IFN-y to its receptor IFNGR1/IFNGR2. The release of IFN-y by T cells could promote the selection pressure of cancer cells, resulting in acquired deficiency of the IFN-y pathway and acquired resistance to ICIs through loss of sensitivity to IFN-y. In addition, resistance to immunotherapy was observed with a loss of IFN- γ signaling in CD8⁺ T cells (Darvin et al., 2018). These results demonstrate the potential effects of IFN- γ in HPD.

Increase in Type 3 Innate Lymphoid Cells

Xiong et al. showed that ILC3 were specifically increased in HPD tumors (Xiong et al., 2018). ILC3s can respond to cytokine stimulation without a specific antigen. It has been reported that ILC3 can produce IL-17 and IL-22, thereby promoting cancer progression (Fung et al., 2019). Irshad et al. showed that the presence of ILC3 in the tumor microenvironment was associated with a higher risk of lymph node metastasis in breast cancer. In a mouse model of colon cancer, reduction of IL-22 production by ILC3 impaired the progression of gastrointestinal cancers (Kirchberger et al., 2013). Inhibiting the expression of PD-1 in tumors leads to increased levels of IL-6 and IL-17 in the peripheral blood; because the main function of IL-6 and IL-17 is to promote neutrophil-mediated inflammation, this may weaken the anti-tumor immune response (Fielding et al., 2008; Miossec and Kolls, 2012). This abnormal inflammatory environment is likely to be related to the poor efficacy of ICIs, but its relationship with HPD is not yet clear.

The Effects of Neoantigens

Mutations in the protein-coding regions produce truncated proteins called "neoantigens." Neoantigens result in higher heterogenicity of cells, which facilitates immune cell targeting and eliminates tumor cells. Acquired resistance to ICIs and HPD can also be predicted using neoantigens. Tumor cells alter the expression of multiple immune suppressive factors, leading to acquired resistance against ICIs (Jenkins et al., 2018). Dysfunction of neoantigens might promote metastasis and recurrence of tumors. Screening of these neoantigens has the potential to predict therapeutic resistance as well as HPD.

Other Mechanisms

As mentioned earlier, changes in tumor immune microenvironments caused by radiotherapy combined with ICIs may accelerate tumor growth. In a previous study (Saâda-Bouzid et al., 2017), almost all cases of HPD occurred in patients with at least regional recurrence in an irradiated area. Radiotherapy not only upregulates the expression of VEGF and promotes tumor angiogenesis and growth, it can also enhance the expression of TGF- β , thereby inhibiting the activation of T cells and DCs.

In most cases, it is common for patients to receive cytotoxic drugs before immunotherapy. However, traditional chemotherapy has been reported to reduce the anti-tumor efficacy of immunotherapy. This suggests that resistant clones are selected after chemotherapy treatment, and the immune system cannot detect them when ICIs are used. As there is currently no effective immune monitoring tool, it is impossible to predict the risk of HPD in patients before ICI treatment. Kim et al. reported that high LDH serum levels were significantly associated with HPD. High LDH levels represent hypoxia in the tumor and acidification of the extracellular environment (Kim J. Y. et al., 2019). High LDH levels and acidic environments may influence the function of antibodies and the conformation of antigens, thereby affecting the specificity and affinity of ICIs. Okeya et al. found that after 5 weeks of pembrolizumab treatment, a 66-year-old male smoker with advanced lung adenocarcinoma developed small-cell carcinoma combined with HPD and metastases (Okeya et al., 2019). In in vivo and in vitro studies, Kudo et al. observed the presence of both chemotherapy- and immunotherapy-activated cancer stem cells, which contributed to aggressive cell proliferation and drug resistance (Kudo-Saito et al., 2019). They proposed that HPD may depend on the proportions of cancer cells eliminated and dormant cells activated. In addition, ICIs can negatively affect the endocrine system and cause autoimmune diseases. Several studies have found that gut microbiota influence the effects of ICIs in melanoma patients. The microbiome is well known to be a key regulator of inflammation and immune responses and may have an important role in cancer therapy.

POSSIBLE STRATEGIES FOR AVOIDING HYPERPROGRESSIVE DISEASE

Owing to the poor prognosis of patients with HPD, there is an urgent need to develop strategies to reduce or eliminate harm to these patients. In clinical practice, it is necessary to pay more attention to changes to the patient's condition and to evaluate the efficacy of treatment with ICIs. First, patients to be treated with ICIs need to be fully informed of the risks of HPD, as the incidence is high enough that some patients may refuse the treatment owing to the unfavorable consequences (Zhang et al., 2021). In addition, current tumor evaluation methods of immunotherapy do not include TGK, which is a key to the early identification of HPD. It could enable a considerable proportion of patients who meet the definition of HPD to continue to receive the same treatment, and it will not be able to block HPD in time (Champiat et al., 2018). Existing disease monitoring and evaluation systems for cancer urgently need to be changed. In salvage therapy, early identification of HPD and timely replacement of ICIs might be the only remedies to avoid risk to patients at present. However, pseudoprogression also affects the identification of HPD; therefore, apparent progression of a tumor at the first imaging after initiation of immunotherapy does not necessarily mean that the treatment must be terminated, as patients with pseudoprogression could benefit significantly from this treatment (Tazdait et al., 2018). The assessment of MDM2/4 amplification, EGFR mutation and CNI score before treatment is useful for selecting patients who will probably develop HPD. However, the predictive value of these biomarkers has not been validated. In addition, it is not possibly to accurately predict HPD in patients with several of these features, including high serum LDH.

The antitumor activity of the treated NK cells (nuclear-trafficking property-genome modulator) was enhanced, and repeated administration of these NK cells attenuated PD-L1-positive tumor cells *in vivo*. The use of NK cells combined with ICIs could be an alternative option for patients with HPD (Teratake et al., 2020). In the Checkmate227 trial, patients treated with nivolumab combined with chemotherapy had a lower risk of progression than those receiving nivolumab combined with ipilimumab (Hellmann et al., 2019). Chemotherapy may help prevent patients from developing ICI resistance and HPD; this possibility deserves further research.

As the relevant studies are mostly retrospective analyses and case reports, the results are inherently limited by selection bias. Clinical trials do not usually include HPD based on radiological criteria as a pattern of response, making the evaluation of this phenomenon in large cohorts of patients difficult. Clinicians should realize that after failure of ICI treatment, subsequent treatment is likely to be ineffective (Schvartsman et al., 2017), and most HPD patients do not have the opportunity to receive subsequent treatment at all (Kim C. G. et al., 2019). Further studies are required to confirm more of the mechanisms underlying HPD, and to explore other accurate biomarkers for this paradoxical response to ICIs. Investigators should attempt to assess the HPD status patients to guide their management.

DISCUSSION

The application of immunotherapy has contributed to fundamental changes in the treatment of cancer. As ICIs are widely administered by physicians, better survival rates are achieved in increasing numbers of patients, especially those with solid tumors. With the rapid development of ICIs, physicians should be aware of the adverse

effects and correctly assess response to tumor therapy and the challenges in patient management. Although there are still many controversies regarding HPD, it is known to occur in multiple tumor types and almost all malignant tumors, and to be related to poor prognosis. An increasing number of retrospective studies indicate that HPD is induced by ICIs. HPD is a severe adverse reaction in ICI treatment, regardless of the specific type of ICI, but the underlying mechanism and predictive indicators are not yet clear, which limits the clinical application of ICIs. ICI-induced HPD is complex, and in some patients, the tumor size increases dramatically after checkpoint blockade. Therefore, it is imperative to explore the etiology and pathogenesis of HPD and develop predictive and detection methods to prevent the cessation of immunotherapy.

In clinical practice, the identification of HPD, disease progression and pseudo-progression is of great significance. At present, TGK and TGR are the main indicators for evaluating HPD, and rapid progression and TTF are two alternative indicators. To date, we have not yet determined the exact incidence of HPD because of the uncertainties in current testing methods and the relatively small number of patients examined. In the future, accurate diagnostic tools should be applied to evaluate tumor changes. As for predictive indicators, most existing conclusions are derived from retrospective studies that explored the specific clinical characteristics of HPD patients, but different types of studies may produce conflicting conclusions, and there is still a lack of accurate predictive indicators for HPD. To further understand HPD, more prospective, randomized and controlled studies are needed to verify whether HPD is an independent type of ICI response after treatment, identify predictive indicators and develop consistent criteria. This will allow us to efficiently predict the response of personalized treatment and develop personalized treatment plans.

In this review, we proposed that factors such as tumor microenvironments, radiation therapy, age, increased tumor burden, gut and tumor microbiomes, changes in immune cell subtypes, FcR polymorphisms, abnormal expression of tumor drivers or resistance-related genes and compensatory activation of other immune checkpoint pathways may be involved in the occurrence of HPD. Additionally, genomic analyses may help to elucidate the mechanisms of HPD and identify effective biomarkers to distinguish individuals at high-risk of HPD. Circulating tumor cell-free DNA may be used as a biomarker in the early monitoring and diagnosis of HPD. This will require large cohort studies to validate these potential detection methods.

In summary, HPD is a new phenomenon, and its underlying mechanism remains to be elucidated. In addition, the clinical relevance and predictive factors of HPD should be further investigated. HPD provides a challenge and an opportunity for the development of novel tumor biotherapies, and it will also encourage research to improve the effectiveness and safety of ICIs. It is important to study this issue in depth to protect cancer patients from the potentially harmful side effects of ICI therapies.

AUTHOR CONTRIBUTIONS

PS wrote the draft of the manuscript. All authors contributed to manuscript revision, and approved the submitted version.

REFERENCES

- Akbay, E. A., Koyama, S., Carretero, J., Altabef, A., Tchaicha, J. H., Christensen, C. L., et al. (2013). Activation of the PD-1 Pathway Contributes to Immune Escape in EGFR-Driven Lung Tumors. *Cancer Discov.* 3, 1355–1363. doi:10.1158/2159-8290.Cd-13-0310
- Arasanz, H., Zuazo, M., Bocanegra, A., Gato, M., Martínez-Aguillo, M., Morilla, I., et al. (2020). Early Detection of Hyperprogressive Disease in Non-small Cell Lung Cancer by Monitoring of Systemic T Cell Dynamics. *Cancers* 12, 344. doi:10.3390/cancers12020344
- Barnaba, V., and Schinzari, V. (2013). Induction, Control, and Plasticity of Treg Cells: the Immune Regulatory Network Revised? *Eur. J. Immunol.* 43, 318–322. doi:10.1002/eji.201243265
- Boeri, M., Milione, M., Signorelli, D., Proto, C., Lo Russo, G., Galeone, C., et al. (2018). MA04.07 Microrna-Based Liquid Biopsy Combines with PD-L1 Tumor Expression to Predict Response to Immunotherapy in advance NSCLC Patients. J. Thorac. Oncol. 13, S368–S369. doi:10.1016/ j.jtho.2018.08.345
- Boland, J. L., Zhou, Q., Martin, M., Callahan, M. K., Konner, J., O'Cearbhaill, R. E., et al. (2019). Early Disease Progression and Treatment Discontinuation in Patients with Advanced Ovarian Cancer Receiving Immune Checkpoint Blockade. *Gynecol. Oncol.* 152, 251–258. doi:10.1016/j.ygyno.2018.11.025
- Borghaei, H., Paz-Ares, L., Horn, L., Spigel, D. R., Steins, M., Ready, N. E., et al. (2015). Nivolumab versus Docetaxel in Advanced Nonsquamous Non-smallcell Lung Cancer. N. Engl. J. Med. 373, 1627–1639. doi:10.1056/ NEJMoa1507643
- Brandsma, D., and van den Bent, M. J. (2009). Pseudoprogression and Pseudoresponse in the Treatment of Gliomas. *Curr. Opin. Neurol.* 22, 633–638. doi:10.1097/WCO.0b013e328332363e
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., and Jemal, A. (2018). Global Cancer Statistics 2018: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer J. Clinicians* 68, 394–424. doi:10.3322/caac.21492
- Cabel, L., Proudhon, C., Romano, E., Girard, N., Lantz, O., Stern, M.-H., et al. (2018). Clinical Potential of Circulating Tumour DNA in Patients Receiving Anticancer Immunotherapy. *Nat. Rev. Clin. Oncol.* 15, 639–650. doi:10.1038/ s41571-018-0074-3
- Chae, Y. K., Wang, S., Nimeiri, H., Kalyan, A., and Giles, F. J. (2017). Pseudoprogression in Microsatellite Instability-High Colorectal Cancer during Treatment with Combination T Cell Mediated Immunotherapy: a Case Report and Literature Review. Oncotarget 8, 57889–57897. doi:10.18632/oncotarget.18361
- Champiat, S., Dercle, L., Ammari, S., Massard, C., Hollebecque, A., Postel-Vinay, S., et al. (2017). Hyperprogressive Disease Is a New Pattern of Progression in Cancer Patients Treated by Anti-PD-1/PD-L1. *Clin. Cancer Res.* 23, 1920–1928. doi:10.1158/1078-0432.Ccr-16-1741
- Champiat, S., Ferrara, R., Massard, C., Besse, B., Marabelle, A., Soria, J.-C., et al. (2018). Hyperprogressive Disease: Recognizing a Novel Pattern to Improve Patient Management. *Nat. Rev. Clin. Oncol.* 15, 748–762. doi:10.1038/s41571-018-0111-2
- Chiou, V. L., and Burotto, M. (2015). Pseudoprogression and Immune-Related Response in Solid Tumors. Jco 33, 3541–3543. doi:10.1200/jco.2015.61.6870
- Costantini, A., Fallet, V., Corny, J., Friard, S., Chouaid, C., Duchemann, B., et al. (2019). Nivolumab-refractory Patients with Advanced Non-small-cell Lung Cancer. Lung Cancer 130, 128–134. doi:10.1016/j.lungcan.2019.01.015
- Dahan, R., Sega, E., Engelhardt, J., Selby, M., Korman, A. J., and Ravetch, J. V. (2015). FcyRs Modulate the Anti-tumor Activity of Antibodies Targeting the PD-1/PD-L1 Axis. *Cancer Cell* 28, 543. doi:10.1016/j.ccell.2015.09.011
- Darvin, P., Toor, S. M., Sasidharan Nair, V., and Elkord, E. (2018). Immune Checkpoint Inhibitors: Recent Progress and Potential Biomarkers. *Exp. Mol. Med.* 50, 1–11. doi:10.1038/s12276-018-0191-1
- Di Pilato, M., Kim, E. Y., Cadilha, B. L., Prüßmann, J. N., Nasrallah, M. N., Seruggia, D., et al. (2019). Targeting the CBM Complex Causes Treg Cells to Prime Tumours for Immune Checkpoint Therapy. *Nature* 570, 112–116. doi:10.1038/s41586-019-1215-2
- Ellestad, K. K., Thangavelu, G., Ewen, C. L., Boon, L., and Anderson, C. C. (2014). PD-1 Is Not Required for Natural or Peripherally Induced Regulatory T Cells:

Severe Autoimmunity Despite normal Production of Regulatory T Cells. *Eur. J. Immunol.* 44, 3560–3572. doi:10.1002/eji.201444688

- Faure, M., Rochigneux, P., Olive, D., Taix, S., Brenot-Rossi, I., and Gilabert, M. (2018). Hyperprogressive Disease in Anorectal Melanoma Treated by PD-1 Inhibitors. *Front. Immunol.* 9, 797. doi:10.3389/fimmu.2018.00797
- Ferrara, R., Caramella, C., Texier, M., Audigier Valette, C., Tessonnier, L., Mezquita, L., et al. (2017). Hyperprogressive Disease (HPD) Is Frequent in Non-small Cell Lung Cancer (NSCLC) Patients (Pts) Treated with Anti PD1/PD-L1 Monoclonal Antibodies (IO). Ann. Oncol. 28, v464–v465. doi:10.1093/annonc/mdx380.009
- Ferrara, R., Mezquita, L., Texier, M., Lahmar, J., Audigier-Valette, C., Tessonnier, L., et al. (2018). Hyperprogressive Disease in Patients with Advanced Non-small Cell Lung Cancer Treated with Pd-1/pd-L1 Inhibitors or with Single-Agent Chemotherapy. JAMA Oncol. 4, 1543–1552. doi:10.1001/jamaoncol.2018.3676
- Ferris, R. L., Blumenschein, G., Jr., Fayette, J., Guigay, J., Colevas, A. D., Licitra, L., et al. (2016). Nivolumab for Recurrent Squamous-Cell Carcinoma of the Head and Neck. N. Engl. J. Med. 375, 1856–1867. doi:10.1056/NEJMoa1602252
- Ferté, C., Fernandez, M., Hollebecque, A., Koscielny, S., Levy, A., Massard, C., et al. (2014). Tumor Growth Rate Is an Early Indicator of Antitumor Drug Activity in Phase I Clinical Trials. *Clin. Cancer Res.* 20, 246–252. doi:10.1158/1078-0432.Ccr-13-2098
- Fielding, C. A., McLoughlin, R. M., McLeod, L., Colmont, C. S., Najdovska, M., Grail, D., et al. (2008). IL-6 Regulates Neutrophil Trafficking during Acute Inflammation via STAT3. J. Immunol. 181, 2189–2195. doi:10.4049/jimmunol.181.3.2189
- Fulop, T., Witkowski, J. M., Le Page, A., Fortin, C., Pawelec, G., and Larbi, A. (2017). Intracellular Signalling Pathways: Targets to Reverse Immunosenescence. *Clin. Exp. Immunol.* 187, 35–43. doi:10.1111/cei.12836
- Fung, K. Y., Nguyen, P. M., and Putoczki, T. (2019). The Expanding Role of Innate Lymphoid Cells and Their T-Cell Counterparts in Gastrointestinal Cancers. *Mol. Immunol.* 110, 48–56. doi:10.1016/j.molimm.2017.11.013
- Gainor, J. F., Shaw, A. T., Sequist, L. V., Fu, X., Azzoli, C. G., Piotrowska, Z., et al. (2016). EGFR Mutations and ALK Rearrangements Are Associated with Low Response Rates to PD-1 Pathway Blockade in Non-Small Cell Lung Cancer: A Retrospective Analysis. *Clin. Cancer Res.* 22, 4585–4593. doi:10.1158/1078-0432.Ccr-15-3101
- Gandara, D. R., Reck, M., Morris, S., Cardona, A., Mendus, D., Ballinger, M., et al. (2018). Fast Progression in Patients Treated with a Checkpoint Inhibitor (Cpi) vs Chemotherapy in OAK, a Phase III Trial of Atezolizumab (Atezo) vs Docetaxel (Doc) in 2L+ NSCLC. Ann. Oncol. 29, x39. doi:10.1093/annonc/mdy511
- Gomez-Roca, C., Koscielny, S., Ribrag, V., Dromain, C., Marzouk, I., Bidault, F., et al. (2011). Tumour Growth Rates and RECIST Criteria in Early Drug Development. *Eur. J. Cancer* 47, 2512–2516. doi:10.1016/j.ejca.2011.06.012
- Goronzy, J. J., and Weyand, C. M. (2013). Understanding Immunosenescence to Improve Responses to Vaccines. Nat. Immunol. 14, 428–436. doi:10.1038/ni.2588
- Hellmann, M. D., Ciuleanu, T.-E., Pluzanski, A., Lee, J. S., Otterson, G. A., Audigier-Valette, C., et al. (2018). Nivolumab Plus Ipilimumab in Lung Cancer with a High Tumor Mutational burden. N. Engl. J. Med. 378, 2093–2104. doi:10.1056/NEJMoa1801946
- Hellmann, M. D., Paz-Ares, L., Bernabe Caro, R., Zurawski, B., Kim, S.-W., Carcereny Costa, E., et al. (2019). Nivolumab Plus Ipilimumab in Advanced Non-Small-Cell Lung Cancer. N. Engl. J. Med. 381, 2020–2031. doi:10.1056/ NEJMoa1910231
- Herbst, R. S., Baas, P., Kim, D.-W., Felip, E., Pérez-Gracia, J. L., Han, J.-Y., et al. (2016). Pembrolizumab versus Docetaxel for Previously Treated, PD-L1-Positive, Advanced Non-small-cell Lung Cancer (KEYNOTE-010): a Randomised Controlled Trial. *The Lancet* 387, 1540–1550. doi:10.1016/ s0140-6736(15)01281-7
- Hodi, F. S., Hwu, W.-J., Kefford, R., Weber, J. S., Daud, A., Hamid, O., et al. (2016). Evaluation of Immune-Related Response Criteria and Recist v1.1 in Patients with Advanced Melanoma Treated with Pembrolizumab. J. Clin. Oncol. 34, 1510–1517. doi:10.1200/jco.2015.64.0391
- Hodi, F. S., O'Day, S. J., McDermott, D. F., Weber, R. W., Sosman, J. A., Haanen, J. B., et al. (2010). Improved Survival with Ipilimumab in Patients with Metastatic Melanoma. N. Engl. J. Med. 363, 711–723. doi:10.1056/NEJMoa1003466
- Hodi, F. S., Oble, D. A., Drappatz, J., Velazquez, E. F., Ramaiya, N., Ramakrishna, N., et al. (2008). CTLA-4 Blockade with Ipilimumab Induces Significant Clinical Benefit in a Female with Melanoma Metastases to the CNS. *Nat. Rev. Clin. Oncol.* 5, 557–561. doi:10.1038/ncponc1183

- Huang, A. C., Postow, M. A., Orlowski, R. J., Mick, R., Bengsch, B., Manne, S., et al. (2017). T-cell Invigoration to Tumour burden Ratio Associated with Anti-PD-1 Response. *Nature* 545, 60–65. doi:10.1038/nature22079
- Huang, R.-Y., Francois, A., McGray, A. R., Miliotto, A., and Odunsi, K. (2017). Compensatory Upregulation of PD-1, LAG-3, and CTLA-4 Limits the Efficacy of Single-Agent Checkpoint Blockade in Metastatic Ovarian Cancer. Oncoimmunology 6, e1249561. doi:10.1080/2162402x.2016.1249561
- Jenkins, R. W., Barbie, D. A., and Flaherty, K. T. (2018). Mechanisms of Resistance to Immune Checkpoint Inhibitors. Br. J. Cancer 118, 9–16. doi:10.1038/bjc.2017.434
- Jensen, T. J., Goodman, A. M., Kato, S., Ellison, C. K., Daniels, G. A., Kim, L., et al. (2019). Genome-wide Sequencing of Cell-free Dna Identifies Copy-Number Alterations that Can Be Used for Monitoring Response to Immunotherapy in Cancer Patients. *Mol. Cancer Ther.* 18, 448–458. doi:10.1158/1535-7163.Mct-18-0535
- Ji, Z., Peng, Z., Gong, J., Zhang, X., Li, J., Lu, M., et al. (2019). Hyperprogression after Immunotherapy in Patients with Malignant Tumors of Digestive System. *BMC Cancer* 19, 705. doi:10.1186/s12885-019-5921-9
- Kagamu, H., Kitano, S., Yamaguchi, O., Yoshimura, K., Horimoto, K., Kitazawa, M., et al. (2020). CD4+ T-Cell Immunity in the Peripheral Blood Correlates with Response to Anti-PD-1 Therapy. *Cancer Immunol. Res.* 8, 334–344. doi:10.1158/2326-6066.Cir-19-0574
- Kamada, T., Togashi, Y., Tay, C., Ha, D., Sasaki, A., Nakamura, Y., et al. (2019). PD-1+ Regulatory T Cells Amplified by PD-1 Blockade Promote Hyperprogression of Cancer. *Proc. Natl. Acad. Sci. USA* 116, 9999–10008. doi:10.1073/pnas.1822001116
- Kanazu, M., Edahiro, R., Krebe, H., Nishida, K., Ishijima, M., Uenami, T., et al. (2018). Hyperprogressive Disease in Patients with Non-small Cell Lung Cancer Treated with Nivolumab: A Case Series. *Thorac. Cancer* 9, 1782–1787. doi:10.1111/1759-7714.12894
- Kanjanapan, Y., Day, D., Wang, L., Al-Sawaihey, H., Abbas, E., Namini, A., et al. (2019). Hyperprogressive Disease in Early-phase Immunotherapy Trials: Clinical Predictors and Association with Immune-related Toxicities. *Cancer* 125, 1341–1349. doi:10.1002/cncr.31999
- Kato, S., Goodman, A., Walavalkar, V., Barkauskas, D. A., Sharabi, A., and Kurzrock, R. (2017). Hyperprogressors after Immunotherapy: Analysis of Genomic Alterations Associated with Accelerated Growth Rate. *Clin. Cancer Res.* 23, 4242–4250. doi:10.1158/1078-0432.Ccr-16-3133
- Kim, C. G., Kim, K. H., Pyo, K.-H., Xin, C.-F., Hong, M. H., Ahn, B.-C., et al. (2019). Hyperprogressive Disease during PD-1/PD-L1 Blockade in Patients with Nonsmall-cell Lung Cancer. Ann. Oncol. 30, 1104–1113. doi:10.1093/annonc/mdz123
- Kim, J. Y., Lee, K. H., Kang, J., Borcoman, E., Saada-Bouzid, E., Kronbichler, A., et al. (2019). Hyperprogressive Disease during Anti-PD-1 (PDCD1)/PD-L1 (CD274) Therapy: A Systematic Review and Meta-Analysis. *Cancers* 11, 1699. doi:10.3390/cancers11111699
- Kirchberger, S., Royston, D. J., Boulard, O., Thornton, E., Franchini, F., Szabady, R. L., et al. (2013). Innate Lymphoid Cells Sustain colon Cancer through Production of Interleukin-22 in a Mouse Model. J. Exp. Med. 210, 917–931. doi:10.1084/jem.20122308
- Koyama, S., Akbay, E. A., Li, Y. Y., Herter-Sprie, G. S., Buczkowski, K. A., Richards, W. G., et al. (2016). Adaptive Resistance to Therapeutic PD-1 Blockade Is Associated with Upregulation of Alternative Immune Checkpoints. *Nat. Commun.* 7, 10501. doi:10.1038/ncomms10501
- Kudo-Saito, C., Ozaki, Y., Kinowaki, K., Kawabata, H., and Ogiwara, Y. (2019). Abstract 3751: A Mode of Treatment Resistance of Metastatic Tumor Cells: Propagation Exceeds Elimination in Number. J. Cancer Res. 79, 3751. doi:10.1158/1538-7445.AM2019-3751
- Lahmar, J., Mezquita, L., Koscielny, S., Facchinetti, F., Bluthgen, M. V., Adam, J., et al. (2016). Immune Checkpoint Inhibitors (IC) Induce Paradoxical Progression in a Subset of Non-small Cell Lung Cancer (NSCLC). Ann. Oncol. 27, vi423. doi:10.1093/annonc/mdw383.22
- Lamichhane, P., Karyampudi, L., Shreeder, B., Krempski, J., Bahr, D., Daum, J., et al. (2017). IL10 Release upon PD-1 Blockade Sustains Immunosuppression in Ovarian Cancer. *Cancer Res.* 77, 6667–6678. doi:10.1158/0008-5472.Can-17-0740
- Lee, J. H., Long, G. V., Menzies, A. M., Lo, S., Guminski, A., Whitbourne, K., et al. (2018). Association between Circulating Tumor Dna and Pseudoprogression in Patients with Metastatic Melanoma Treated with Anti-programmed Cell Death 1 Antibodies. JAMA Oncol. 4, 717–721. doi:10.1001/jamaoncol.2017.5332

- Lehner, T. (2008). Special Regulatory T Cell Review: The Resurgence of the Concept of Contrasuppression in Immunoregulation. *Immunology* 123, 40–44. doi:10.1111/j.1365-2567.2007.02780.x
- Li, C.-W., Lim, S.-O., Xia, W., Lee, H.-H., Chan, L.-C., Kuo, C.-W., et al. (2016). Glycosylation and Stabilization of Programmed Death Ligand-1 Suppresses T-Cell Activity. *Nat. Commun.* 7, 12632. doi:10.1038/ncomms12632
- Lipson, E. J., Velculescu, V. E., Pritchard, T. S., Sausen, M., Pardoll, D. M., Topalian, S. L., et al. (2014). Circulating Tumor DNA Analysis as a Real-Time Method for Monitoring Tumor burden in Melanoma Patients Undergoing Treatment with Immune Checkpoint Blockade. J. Immunotherapy Cancer 2, 42. doi:10.1186/s40425-014-0042-0
- Lo Russo, G., Moro, M., Sommariva, M., Cancila, V., Boeri, M., Centonze, G., et al. (2019). Antibody-Fc/fcr Interaction on Macrophages as a Mechanism for Hyperprogressive Disease in Non-small Cell Lung Cancer Subsequent to PD-1/PD-L1 Blockade. *Clin. Cancer Res.* 25, 989–999. doi:10.1158/1078-0432.Ccr-18-1390
- Lu, Z., Zou, J., Hu, Y., Li, S., Zhou, T., Gong, J., et al. (2019). Serological Markers Associated with Response to Immune Checkpoint Blockade in Metastatic Gastrointestinal Tract Cancer. JAMA Netw. Open 2, e197621. doi:10.1001/ jamanetworkopen.2019.7621
- Matos, I., Martin-Liberal, J., Hierro, C., Ochoa De Olza, M., Viaplana, C., Costa, M., et al. (2018). Incidence and Clinical Implications of a New Definition of Hyperprogression (HPD) with Immune Checkpoint Inhibitors (ICIs) in Patients Treated in Phase 1 (Ph1) Trials. J. Clin. Oncol. 36, 3032. doi:10.1200/JCO.2018.36.15_suppl.3032
- Millward, M., Underhill, C., Lobb, S., McBurnie, J., Meech, S. J., Gomez-Navarro, J., et al. (2013). Phase I Study of Tremelimumab (CP-675 206) Plus PF-3512676 (CPG 7909) in Patients with Melanoma or Advanced Solid Tumours. *Br. J. Cancer* 108, 1998–2004. doi:10.1038/bjc.2013.227
- Miossec, P., and Kolls, J. K. (2012). Targeting IL-17 and TH17 Cells in Chronic Inflammation. Nat. Rev. Drug Discov. 11, 763–776. doi:10.1038/nrd3794
- Momand, J., Jung, D., Wilczynski, S., and Niland, J. (1998). The MDM2 Gene Amplification Database. Nucleic Acids Res. 26, 3453–3459. doi:10.1093/nar/26.15.3453
- Nishino, M., Giobbie-Hurder, A., Gargano, M., Suda, M., Ramaiya, N. H., and Hodi, F. S. (2013). Developing a Common Language for Tumor Response to Immunotherapy: Immune-Related Response Criteria Using Unidimensional Measurements. *Clin. Cancer Res.* 19, 3936–3943. doi:10.1158/1078-0432.Ccr-13-0895
- Odorizzi, P. M., Pauken, K. E., Paley, M. A., Sharpe, A., and Wherry, E. J. (2015). Genetic Absence of PD-1 Promotes Accumulation of Terminally Differentiated Exhausted CD8+ T Cells. *J. Exp. Med.* 212, 1125–1137. doi:10.1084/jem.20142237
- Okeya, K., Kawagishi, Y., Muranaka, E., Izumida, T., Tsuji, H., and Takeda, S. (2019). Hyperprogressive Disease in Lung Cancer with Transformation of Adenocarcinoma to Small-Cell Carcinoma during Pembrolizumab Therapy. *Intern. Med.* 58, 3295–3298. doi:10.2169/internalmedicine.2892-19
- Pearson, A. T., and Sweis, R. F. (2019). Hyperprogression-immunotherapy-related Phenomenon vs Intrinsic Natural History of Cancer. JAMA Oncol. 5, 743. doi:10.1001/jamaoncol.2019.0130
- Peng, W., Liu, C., Xu, C., Lou, Y., Chen, J., Yang, Y., et al. (2012). PD-1 Blockade Enhances T-Cell Migration to Tumors by Elevating IFN-γ Inducible Chemokines. *Cancer Res.* 72, 5209–5218. doi:10.1158/0008-5472.Can-12-1187
- Petrioli, R., Mazzei, M. A., Giorgi, S., Cesqui, E., Gentili, F., Francini, G., et al. (2020). Hyperprogressive Disease in Advanced Cancer Patients Treated with Nivolumab: a Case Series Study. *Anticancer Drugs* 31, 190–195. doi:10.1097/ cad.000000000000864
- Refae, S., Gal, J., Brest, P., Giacchero, D., Borchiellini, D., Ebran, N., et al. (2020). Hyperprogression under Immune Checkpoint Inhibitor: a Potential Role for Germinal Immunogenetics. *Sci. Rep.* 10, 3565. doi:10.1038/s41598-020-60437-0
- Ribas, A., Chesney, J. A., Gordon, M. S., Abernethy, A. P., Logan, T. F., Lawson, D. H., et al. (2012). Safety Profile and Pharmacokinetic Analyses of the Anti-CTLA4 Antibody Tremelimumab Administered as a One Hour Infusion. *J. Transl. Med.* 10, 236. doi:10.1186/1479-5876-10-236
- Ribas, A., and Wolchok, J. D. (2018). Cancer Immunotherapy Using Checkpoint Blockade. *Science* 359, 1350–1355. doi:10.1126/science.aar4060
- Robert, C., Long, G. V., Brady, B., Dutriaux, C., Maio, M., Mortier, L., et al. (2015). Nivolumab in Previously Untreated Melanoma withoutBRAFMutation. N. Engl. J. Med. 372, 320–330. doi:10.1056/NEJMoa1412082

- Ruiz-Patiño, A., Arrieta, O., Cardona, A. F., Martín, C., Raez, L. E., Zatarain-Barrón, Z. L., et al. (2020). Immunotherapy at Any Line of Treatment Improves Survival in Patients with Advanced Metastatic Non-small Cell Lung Cancer (NSCLC) Compared with Chemotherapy (Quijote-CLICaP). *Thorac. Cancer* 11, 353–361. doi:10.1111/1759-7714.13272
- Saáda-Bouzid, E., Defaucheux, C., Karabajakian, A., Coloma, V. P., Servois, V., Paoletti, X., et al. (2017). Hyperprogression during Anti-PD-1/PD-L1 Therapy in Patients with Recurrent And/or Metastatic Head and Neck Squamous Cell Carcinoma. Ann. Oncol. 28, 1605–1611. doi:10.1093/ annonc/mdx178
- Sasaki, A., Nakamura, Y., Mishima, S., Kawazoe, A., Kuboki, Y., Bando, H., et al. (2019). Predictive Factors for Hyperprogressive Disease during Nivolumab as Anti-PD1 Treatment in Patients with Advanced Gastric Cancer. *Gastric Cancer* 22, 793–802. doi:10.1007/s10120-018-00922-8
- Sasidharan Nair, V., Toor, S. M., Taouk, G., Pfister, G., Ouararhni, K., Alajez, N. M., et al. (2020). Pembrolizumab Interferes with the Differentiation of Human FOXP3+-Induced T Regulatory Cells, but Not with FOXP3 Stability, through Activation of mTOR. J. Immunol. 204, 199–211. doi:10.4049/ jimmunol.1900575
- Schindler, C., Levy, D. E., and Decker, T. (2007). JAK-STAT Signaling: from Interferons to Cytokines. J. Biol. Chem. 282, 20059–20063. doi:10.1074/ jbc.R700016200
- Schvartsman, G., Peng, S. A., Bis, G., Lee, J. J., Benveniste, M. F. K., Zhang, J., et al. (2017). Response Rates to Single-Agent Chemotherapy after Exposure to Immune Checkpoint Inhibitors in Advanced Non-small Cell Lung Cancer. Lung Cancer 112, 90–95. doi:10.1016/j.lungcan.2017.07.034
- Seymour, L., Bogaerts, J., Perrone, A., Ford, R., Schwartz, L. H., Mandrekar, S., et al. (2017). iRECIST: Guidelines for Response Criteria for Use in Trials Testing Immunotherapeutics. *Lancet Oncol.* 18, e143–e152. doi:10.1016/s1470-2045(17)30074-8
- Shayan, G., Srivastava, R., Li, J., Schmitt, N., Kane, L. P., and Ferris, R. L. (2017). Adaptive Resistance to Anti-PD1 Therapy by Tim-3 Upregulation Is Mediated by the PI3K-Akt Pathway in Head and Neck Cancer. Oncoimmunology 6, e1261779. doi:10.1080/2162402x.2016.1261779
- Singavi, A. K., Menon, S., Kilari, D., Alqwasmi, A., Ritch, P. S., Thomas, J. P., et al. (2017). Predictive Biomarkers for Hyper-Progression (HP) in Response to Immune Checkpoint Inhibitors (ICI) – Analysis of Somatic Alterations (SAs). *Ann. Oncol.* 28, v405. doi:10.1093/annonc/mdx376.006
- Solana, R., Tarazona, R., Gayoso, I., Lesur, O., Dupuis, G., and Fulop, T. (2012). Innate Immunosenescence: Effect of Aging on Cells and Receptors of the Innate Immune System in Humans. *Semin. Immunol.* 24, 331–341. doi:10.1016/ j.smim.2012.04.008
- Soria, F., Beleni, A. I., D'Andrea, D., Resch, I., Gust, K. M., Gontero, P., et al. (2018). Pseudoprogression and Hyperprogression during Immune Checkpoint Inhibitor Therapy for Urothelial and Kidney Cancer. World J. Urol. 36, 1703–1709. doi:10.1007/s00345-018-2264-0
- Tazdait, M., Mezquita, L., Lahmar, J., Ferrara, R., Bidault, F., Ammari, S., et al. (2018). Patterns of Responses in Metastatic NSCLC during PD-1 or PDL-1 Inhibitor Therapy: Comparison of RECIST 1.1, irRECIST and iRECIST Criteria. *Eur. J. Cancer* 88, 38–47. doi:10.1016/j.ejca.2017.10.017
- Ten Berge, D. M. H. J., Hurkmans, D. P., den Besten, I., Kloover, J. S., Mathijssen, R. H. J., Debets, R. J. E. M. A., et al. (2019). Tumour Growth Rate as a Tool for Response Evaluation during PD-1 Treatment for Non-small Cell Lung Cancer: a Retrospective Analysis. *ERJ Open Res.* 5, 00179–02019. doi:10.1183/23120541.00179-2019
- Teng, F., Meng, X., Kong, L., and Yu, J. (2018). Progress and Challenges of Predictive Biomarkers of Anti PD-1/PD-L1 Immunotherapy: A Systematic Review. *Cancer Lett.* 414, 166–173. doi:10.1016/j.canlet.2017.11.014
- Teratake, Y., Takashina, T., Iijima, K., Sakuma, T., Yamamoto, T., and Ishizaka, Y. (2020). Development of a Protein-Based System for Transient Epigenetic Repression of Immune Checkpoint Molecule and Enhancement of Antitumour Activity of Natural Killer Cells. Br. J. Cancer 122, 823–834. doi:10.1038/s41416-019-0708-y
- Tunali, I., Gray, J. E., Qi, J., Abdalah, M., Jeong, D. K., Guvenis, A., et al. (2019). Novel Clinical and Radiomic Predictors of Rapid Disease Progression Phenotypes Among Lung Cancer Patients Treated with Immunotherapy: An Early Report. *Lung Cancer* 129, 75–79. doi:10.1016/ j.lungcan.2019.01.010

- Waight, J. D., Netherby, C., Hensen, M. L., Miller, A., Hu, Q., Liu, S., et al. (2013). Myeloid-derived Suppressor Cell Development Is Regulated by a STAT/IRF-8 axis. J. Clin. Invest. 123, 4464–4478. doi:10.1172/jci68189
- Wang, F., Li, B., Wei, Y., Zhao, Y., Wang, L., Zhang, P., et al. (2018). Tumor-derived Exosomes Induce PD1+ Macrophage Population in Human Gastric Cancer that Promotes Disease Progression. *Oncogenesis* 7, 41. doi:10.1038/s41389-018-0049-3
- Wang, Q., Gao, J., and Wu, X. (2018). Pseudoprogression and Hyperprogression after Checkpoint Blockade. Int. Immunopharmacol. 58, 125–135. doi:10.1016/ j.intimp.2018.03.018
- Weiss, G. J., Beck, J., Braun, D. P., Bornemann-Kolatzki, K., Barilla, H., Cubello, R., et al. (2017). Tumor Cell-free DNA Copy Number Instability Predicts Therapeutic Response to Immunotherapy. *Clin. Cancer Res.* 23, 5074–5081. doi:10.1158/1078-0432.Ccr-17-0231
- Wolchok, J. D., Hoos, A., O'Day, S., Weber, J. S., Hamid, O., Lebbé, C., et al. (2009). Guidelines for the Evaluation of Immune Therapy Activity in Solid Tumors: Immune-Related Response Criteria. *Clin. Cancer Res.* 15, 7412–7420. doi:10.1158/1078-0432.Ccr-09-1624
- Wolchok, J. D., and Saenger, Y. (2008). The Mechanism of Anti-CTLA-4 Activity and the Negative Regulation of T-Cell Activation. Oncol. 13 (Suppl. 4), 2–9. doi:10.1634/theoncologist.13-S4-2
- Wong, D. J., Lee, J., Choo, S. P., Thng, C. H., and Hennedige, T. (2019). Hyperprogressive Disease in Hepatocellular Carcinoma with Immune Checkpoint Inhibitor Use: a Case Series. *Immunotherapy* 11, 167–175. doi:10.2217/imt-2018-0126
- Wu, X., Giobbie-Hurder, A., Liao, X., Connelly, C., Connolly, E. M., Li, J., et al. (2017). Angiopoietin-2 as a Biomarker and Target for Immune Checkpoint Therapy. *Cancer Immunol. Res.* 5, 17–28. doi:10.1158/2326-6066.Cir-16-0206
- Xiong, D., Wang, Y., Singavi, A. K., Mackinnon, A. C., George, B., and You, M. (2018). Immunogenomic Landscape Contributes to Hyperprogressive Disease after Anti-PD-1 Immunotherapy for Cancer. *iScience* 9, 258–277. doi:10.1016/ j.isci.2018.10.021
- Zhang, H., Fang, X., Li, D., Yang, M., Yu, L., Ding, Y., et al. (2021). Hyperprogressive Disease in Patients Receiving Immune Checkpoint Inhibitors. *Curr. Probl. Cancer* 45, 100688. doi:10.1016/j.currproblcancer.2020.100688
- Zhang, T., Song, X., Xu, L., Ma, J., Zhang, Y., Gong, W., et al. (2018). The Binding of an Anti-PD-1 Antibody to FcyRI Has a Profound Impact on its Biological Functions. *Cancer Immunol. Immunother*. 67, 1079–1090. doi:10.1007/s00262-018-2160-x
- Zhao, Y., Yu, H., and Hu, W. (2014). The Regulation of MDM2 Oncogene and its Impact on Human Cancers. Acta Biochim. Biophys. Sinica 46, 180–189. doi:10.1093/abbs/gmt147
- Zhou, J. X., Lee, C. H., Qi, C. F., Wang, H., Naghashfar, Z., Abbasi, S., et al. (2009). IFN Regulatory Factor 8 Regulates MDM2 in Germinal center B Cells. J. Immunol. 183, 3188–3194. doi:10.4049/jimmunol.0803693
- Zuazo, M., Arasanz, H., Gato-Cañas, M., Fernández-Hinojal, G., Hernández-Marín, B., Martínez-Aguillo, M., et al. (2018). Pre-treatment CD4 Senescent T Cells Accurately Predicts Lack of Response to PD-L1/PD-1 Immune Checkpoint Blockade in Non-small Cell Lung Cancer and Correlates with Risk of Hyperprogression. Ann. Oncol. 29, viii15. doi:10.1093/annonc/ mdy269.052
- Zuazo-Ibarra, M., Arasanz, H., Fernández-Hinojal, G., Gato-Cañas, M., Hernández-Marín, B., Martínez-Aguillo, M., et al. (2018). Senescent CD4 T Cells Unequivocally Identify Primary Resistance and Risk of Hyperprogression to PD-L1/PD-1 Immune Checkpoint Blockade in Lung Cancer. J. Biorxiv, 320176. doi:10.1101/320176

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Shen, Han, Ba, Qin and Tu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.