Check for updates

OPEN ACCESS

EDITED BY Kah Hui Wong, University of Malaya, Malaysia

REVIEWED BY Faizul Jaafar, Jeffrey Cheah School of Medicine and Health Sciences, Monash University, Malaysia Danton O'Day, University of Toronto Mississauga, Canada

*CORRESPONDENCE Hamizah Shahirah Hamezah, ⊠ hamizahshahirah@ukm.edu.my

SPECIALTY SECTION This article was submitted to Ethnopharmacology, a section of the journal Frontiers in Pharmacology

RECEIVED 02 September 2022 ACCEPTED 20 December 2022 PUBLISHED 05 January 2023

CITATION

Azlan UK, Khairul Annuar NA, Mediani A, Aizat WM, Damanhuri HA, Tong X, Yanagisawa D, Tooyama I, Wan Ngah WZ, Jantan I and Hamezah HS (2023), An insight into the neuroprotective and antineuroinflammatory effects and mechanisms of *Moringa oleifera*. *Front. Pharmacol.* 13:1035220. doi: 10.3389/fphar.2022.1035220

COPYRIGHT

© 2023 Azlan, Khairul Annuar, Mediani, Aizat, Damanhuri, Tong, Yanagisawa, Tooyama, Wan Ngah, Jantan and Hamezah. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

An insight into the neuroprotective and anti-neuroinflammatory effects and mechanisms of *Moringa oleifera*

Ummi Kalthum Azlan¹, Nur Aisyah Khairul Annuar¹, Ahmed Mediani¹, Wan Mohd Aizat¹, Hanafi Ahmad Damanhuri², Xiaohui Tong³, Daijiro Yanagisawa⁴, Ikuo Tooyama⁵, Wan Zurinah Wan Ngah⁵, Ibrahim Jantan¹ and Hamizah Shahirah Hamezah^{1*}

¹Institute of Systems Biology, Universiti Kebangsaan Malaysia, Bangi, Selangor, Malaysia, ²Department of Biochemistry, Faculty of Medicine, Universiti Kebangsaan Malaysia Medical Center, Kuala Lumpur, Malaysia, ³School of Life Sciences, Anhui University of Chinese Medicine, Hefei, China, ⁴Molecular Neuroscience Research Center, Shiga University of Medical Science, Otsu, Japan, ⁵Medical Innovation Research Center, Shiga University of Medical Science, Otsu, Japan

Neurodegenerative diseases (NDs) are sporadic maladies that affect patients' lives with progressive neurological disabilities and reduced quality of life. Neuroinflammation and oxidative reaction are among the pivotal factors for neurodegenerative conditions, contributing to the progression of NDs, such as Parkinson's disease (PD), Alzheimer's disease (AD), multiple sclerosis (MS) and Huntington's disease (HD). Management of NDs is still less than optimum due to its wide range of causative factors and influences, such as lifestyle, genetic variants, and environmental aspects. The neuroprotective and anti-neuroinflammatory activities of Moringa oleifera have been documented in numerous studies due to its richness of phytochemicals with antioxidant and anti-inflammatory properties. This review highlights up-to-date research findings on the anti-neuroinflammatory and neuroprotective effects of M. oleifera, including mechanisms against NDs. The information was gathered from databases, which include Scopus, Science Direct, Ovid-MEDLINE, Springer, and Elsevier. Neuroprotective effects of M. oleifera were mainly assessed by using the crude extracts in vitro and in vivo experiments. Isolated compounds from M. oleifera such as moringin, astragalin, and isoquercitrin, and identified compounds of M. oleifera such as phenolic acids and flavonoids (chlorogenic acid, gallic acid, ferulic acid, caffeic acid, kaempferol, quercetin, myricetin, (-)-epicatechin, and isoquercitrin) have been reported to have neuropharmacological activities. Therefore, these compounds may potentially contribute to the neuroprotective and anti-neuroinflammatory effects. More indepth studies using in vivo animal models of neurological-related disorders and extensive preclinical investigations, such as pharmacokinetics, toxicity, and bioavailability studies are necessary before clinical trials can be carried out to develop M. oleifera constituents into neuroprotective agents.

KEYWORDS

neurodegenerative, anti-neuroinflammatory, *Moringa oleifera*, neuropharmacological properties, phytochemicals, mechanisms

1 Introduction

Neurodegenerative diseases (NDs) are of serious concern as they developed surreptitiously, progressively, and often irreversible with a high social burden. In addition, the exhibited symptoms and complications such as memory and personality disturbances, personality changes, aphasia, gait disturbance, bradykinesia, and tremors derived from NDs are debilitating and progressing timely (Vöglein et al., 2021). Neurodegeneration is termly known as aggravating occurrence in the central nervous system (CNS) associated with the deterioration of neuronal cells, thus, causing neuronal cell death and cognitive impairment (Chen et al., 2016). Several factors contribute to neurodegeneration, such as aging, environmental influences, genetic variation, and inflammation. Despite the poorly described etiology, neuroinflammation has been found to contribute significantly to several neurodegenerative pathways. The pro-inflammatory cytokines liberation from the neuroinflammatory reaction is commonly observed as the pathophysiology of depression and dementia, as well as multiple sclerosis (MS), Alzheimer's disease (AD), Huntington's disease (HD) and Parkinson's disease (PD). Numerous studies on NDs have demonstrated that neuroinflammation, microglial activation and oxidative stress play a critical function in the development and pathophysiology of the disease (Chen et al., 2016; Solleiro-Villavicencio and Rivas-Arancibia, 2018; Simpson and Oliver, 2020).

The abnormal structures of neurofibrillary tangles formed from the aggregation of hyperphosphorylated tau protein and the oligomers of β -amyloid (A β) peptides have been reported as classic disease pathogenesis markers (Guzman-Martinez et al., 2019). However, increasing discovery of elevated inflammatory markers with associated functional immune risk genes has suggested the vital role of neuroinflammation in AD pathogenesis (Leng and Edison, 2021). The activated microglia as the key player of neuroinflammation display important influences on the progression of NDs markedly by the production of various immune responses with multifaceted interactions with AB, tau proteins, and the CNS (Eikelenboom et al., 2002). Co-occurrence of regulated IL-1 cytokine-activated microglia together with neurofibrillary tangles and $A\beta$ plaques have been observed in the AD pathology (Henkel et al., 2003). The activation of microglial cells has been described as a doubled-edge sword as it provides both neuroprotection and neurotoxicity in CNS (Wyss-Coray and Mucke, 2002). Microglial cells are the source of tumor necrosis factor-a (TNF-a), glutamate and oxidative stress, such as reactive oxygen and nitrogen species (RONS) (Figuera-Losada et al., 2014; Chen et al., 2016; Kabir et al., 2022). Aggregated proteins exhibited stimulation via the toll-like receptor (TLR) signaling pathway, activating the microglia and allowing the release of these substances, which can be neurotoxic at high doses (Chen et al., 2016). Therefore, understanding the mechanisms related to neuroinflammation has been of great interest as it is peculiarly attributable to NDs (Guzman-Martinez et al., 2019). Neurotropic viral infections can also trigger neuroinflammatory responses via neuroimmune activation in the NDs progression (Rock et al., 2004; Chen et al., 2016). Hence, inflammatory pathways have been suggested as a potential medicinal target for NDs (Pålsson-Mcdermott and O' Neill, 2020).

Management of NDs is still less than optimum due to its wide range of causative factors and influences, such as genetic variants, lifestyle, and environmental factors. Anti-inflammatory drugs had not revealed potential effectiveness in slowing disease progression (Jantan et al., 2015; French et al., 2017). Naproxen sodium (Aleve), celecoxib (Celebrex) and other non-steroidal anti-inflammatory drugs (NSAIDS) have been used for AD in phase III clinical trials, involving approximately 2,625 participants in 5-7 years span. Although it has been proposed to reduce the occurrence of AD by delaying or preventing the onset, as well as any associated age-related cognitive decline, no significant effect on diseases occurrence and alleviation was observed at least in the first phase of the trial (Lyketsos et al., 2007). The trial has also found contradicting results as some participants showed aggravated syndromes. Based on these observations, the search continues for other therapeutic targets. However, due to the complexity of the NDs and related complications, many of the potent agents have not shown positive results in clinical trials along with possible adverse effects. Hence, the use of herbal medicine as multi-component agents to modulate the complex immune system is of interest (Jantan et al., 2015; French et al., 2017; Durani et al., 2018). The use of herbal medicine as a multicomponent agent to modulate the complex immune system in disease prevention presents a new alternative approach (Alagan et al., 2019; Jantan et al., 2021). Among the phytoconstituents, polyphenolic compounds'role as potent neuroprotective agents has been deliberated for the contribution in mediating the inflammationrelated cell signaling pathways such as mitogen activated protein kinases (MAPK) and nuclear factor-kappa B (NF-KB) (Jantan et al., 2021). To explore the neuroprotective potential of medicinal herbs, the Neuroprotective Potential Algorithm (NPE) that consists of bioassays (e.g., oxidative stress, Aβ fibrillation, acetylcholinesterase (AChE) inhibition, neuroinflammation) was developed (Liu et al., 2016). Some of the Ayurvedic plant extracts that have been appraised using the NPE include Azadirachta indica, Cinnamomum cassia, Curcuma longa, Moringa oleifera, Phyllanthus emblica, and Punica granatum. Besides, in vivo studies on various herbal medicines, such as Nigella sativa (Babar et al., 2018), P. amarus (Alagan et al., 2019) and M. oleifera (Ekong et al., 2017) have proposed compelling potential of herbal medicine as anti-neuroinflammatory and neuroprotective agents.

Intriguingly, this review discussed specifically M. oleifera, a plant that is well known to have high polyphenolic content. In several studies, the extract was found to exert immunomodulatory effects by modulating the levels of NF-kB expression, cytokines, TNF-a, IL-1, IL-6 and nitric oxide (NO), consequently suppressing the inflammatory reaction (Jaja-Chimedza et al., 2017; Luetragoon et al., 2020). Thus, the current review aims to present published research findings on the anti-neuroinflammatory and neuroprotective properties of M. oleifera as well as its bioactive secondary metabolites, and their mechanisms of action. Articles peer-reviewed journals published in on the antineuroinflammatory and neuroprotective effects and mechanisms of M. oleifera and its constituents were gathered from databases, which include Scopus, Science Direct, Ovid-MEDLINE, Springer, and Elsevier. Specific keywords used are "Moringa", "neuroprotective", "neurodegenerative diseases", "antineuroinflammation", "in vitro studies", and "in vivo studies" were used during data collection. An insight into their antineuroinflammatory and neuroprotective activities and



FIGURE 1

Schematic illustrations of associated signal transducing pathways in LPS-induced neuroinflammation *via* TLR4 signaling pathways. The inhibition of targeted signal transducing pathways is represented by the red lines. The figure was adapted and modified from Gribar et al. (2008), Lu et al. (2008) and Jantan et al. (2021). The figure was created with Biorender com. Abbreviations: AP-1, Activator protein 1; COX-2, Cyclooxygenase-2; ERK, extracellular signal-regulated kinase; IL, interleukin; IRF3, IFN regulatory factor 3; iNOS, Nitric oxide synthase; IKKi, kinase I kappa B kinase I; IKK, inhibitor of nuclear factor-κB (NF-κB) kinase; IRAK1, interleukin-1 receptor-associated kinase 1; IRAK4, interleukin-1 receptor-associated kinase 4; JNK, c-Jun N-terminal kinase; LPS, Lipopolysaccharides; MYD88, Myeloid differentiation primary response 88; NF-κB, nuclear factor kappa Ight chain enhancer of activated B cells; P38 MAPK, P38 mitogen-activated protein kinase; RIP1, receptor-interacting serine/threonine kinase 1; TLR4, Toll-like receptor 4; TRIF, TIR-domain-containing adapter inducing interferon-β; TIRAP, Toll-interleukin 1 receptor (TIR) domain-containing adapter protein; TRAF3, TNF receptor-associated kinase 1; TNF related adaptor molecule; TRAF6, Tumor necrosis factor (TNF) receptor-associated factor 6; TAK1, Transforming growth factor beta-activated kinase 1; TNF-α, Tumor necrosis factor-alpha; TANK, TRAF family member-associated NF-kappa-B activator; TBK1, TANK Binding Kinase 1.

mechanisms of action may provide the basis for the possibility of developing the plant constituents into neuroprotective agents.

2 Neuroinflammatory pathways associated with neurodegenerative diseases and potential therapeutic targets

Neuroinflammation is one of the prominent causative factors in development of NDs (Acioglu et al., 2021). It is initiated by microglia, the resident immune cells of CNS that constituted 5%-10% of the brain cells (Leng and Edison, 2021; Sandhu and Kulka, 2021). As activated microglia can produce a wide range of neurotoxic molecules which also includes the inflammatory cytokines and reactive oxygen intermediates, it was proposed that anti-inflammatory therapies may provide new targets for the treatment of these diseases (Catorce and Gevorkian, 2020; Pålsson-Mcdermott and O' Neill, 2020). Hence, the inflammatory activation of microglia in response to neurodegenerative diseases has been intensively studied (Azam et al., 2021; Sobue et al., 2021). Inflammatory therapeutic targets can enhance the function of immunomodulatory molecules, where the endogenous immunoregulatory system involves various regulations of protein and gene expression in the TLR signaling pathway. TLRs are a

family of microbe-sensing receptors that play a crucial role in regulating the immune system. TLRs signal through the recruitment of specific adaptor molecules and lead to the activation of transcription factors NF- κ B and IRFs. There are 10 members of the TLR family (TLR1-TLR10) in humans (Kawasaki and Kawai, 2014; Frasca and Lande, 2020).

Lipopolysaccharides (LPS) is a bacterial toxin known to induce neuroinflammation by targeting the activation of TLR4 pathway through respective receptors (Boonen et al., 2018; Batista et al., 2019). LPS has been used as an important model in the study of NDs (Batista et al., 2019; Catorce and Gevorkian, 2020). (Figure 1). Subsequently, important signaling, such as TRIF-related adaptor molecule (TRAM) adapter, TIR-domain-containing adaptorinducing interferon- β (TRIF) and myeloid differentiation primary response protein 88 (MyD88) are recruited to further activate the downstream pathways (Ruckdeschel et al., 2004; Gray et al., 2011). These adapters instigated signal transduction pathways that in turn activated the NF-KB, IRFs, or MAPK associated with pro-inflammatory cytokines expression (eg; NO, TNF-a, and IL-6), chemokines, and type I interferons (IFNs) (Kawasaki and Kawai, 2014; Batista et al., 2019; Zhou et al., 2022) necessary to combat infection. However, TLR4 activation and its subsequent inflammatory pathways also contribute to glial reaction, ultimately leading to neuronal loss and damage that



resulted in cognitive impairment and shifted behavior (Batista et al., 2019).

Inflammation plays a central role in the immune system and can be destructive on several levels (Chen et al., 2018; Simpson and Oliver, 2020). Many age-related diseases related to alterations in antiinflammatory or pro-inflammatory cytokines involve TLRs (Zhong et al., 2020; Baidya et al., 2021). Hence, potent therapeutic agent that targets cell signaling pathways by mediating the inflammatory mediators may provide a compelling approach to mitigate NDs. In addition, therapeutic agent with high antioxidant properties can be a potent agent to inhibit oxidative stress along with these signaling networks predispose to neurodegeneration (Ramanan and Saykin, 2013; Tan et al., 2019; Jantan et al., 2021). Oxidative stress is described by the escalated reactive species in which the chronic state causes the alteration in redox signaling and leads to cell damage (Halliwell, 2012; Solleiro-Villavicencio and Rivas-Arancibia, 2018). These reactive species able to mediate signaling that activate astrocytes and microglia (Pawate et al., 2004). Besides, it can activate the associated signaling pathways and allows the production of proinflammatory cytokines such as TNF, IL- 6 and IL-1β (Hsieh and Yang, 2013). Oxidative stress is one of the extracellular stimuli of the MAPK signaling pathway, which is the centerpiece that converts the stimuli to various cellular activities like apoptosis, proliferation, and differentiation as well as inflammatory responses (Kim and Choi, 2010).

There is a growing interest among researchers in studying TLR as a natural product target to mitigate inflammation. For example, a previous study discovered that the elicited soybean extract was able to attenuate expression of pro-inflammatory cytokines by modulating TLR3/TLR4 activation in high-fructose, high-fat diet mice (Atho'illah et al., 2021). Another study demonstrated that lauric acid; a major constituent of coconut oil has a protective role against LPS-induced inflammation in rat liver by mediating TLR4/MyD88 pathway (Khan et al., 2020). The important role of natural compounds in modulating the TLR signaling pathway, resulting in the maintenance of a healthy immune system has been emphasized in these studies. In addition, the

potential of dietary polyphenols (eg; kaempferol, quercetin, and gallic acid) in mediating multiple signaling pathways have also been previously discussed (Jantan et al., 2021). Hence, the high content of these polyphenolic compounds and rich source of antioxidants in *M. oleifera* suggests its values and neuroprotective potential.

3 Taxonomy and distribution of *Moringa* oleifera

M. oleifera is a species belonging to the Moringaceae family along with 12 other different *Moringa* species (Thapa et al., 2019). Besides being commonly known as the 'horseradish' or 'drumstick' tree, *M. oleifera* is also locally known as 'kacang kelor' in Malay or 'Murunggai' in Tamil (Adamu et al., 2021). Among the *Moringa* species, *M. oleifera* is the most notable due to its significant Ayurveda reputation and is sometimes aptly known as the miraculous tree. Each part of the tree has respective benefits and uses (Figure 2) (Leone et al., 2015).

M. oleifera plant is a tropical deciduous perennial dicotyledonous tree that is indigenous to many South Asian countries (Leone et al., 2015; Bhattacharya et al., 2018). Formerly, it was mainly found in the foothills of Himalayas, India, well spread from northeast Pakistan to northern West Bengal (Mahmood et al., 2010). Nowadays, M. oleifera has been introduced and grown naturally in many places around the globe, especially in subtropical and tropical countries, making it one of the fastest growing and naturally distributed plant species. M. oleifera is also a highly anticipated plant due to its minimal needs for plant growth making it relatively easy to cultivate. The plant can simply be propagated from direct seeding or cutting due to its high germination rate (Thapa et al., 2019). M. oleifera (syn. M. ptreygosperma Gaertn.) can easily grow in humid and dry tropical climates, of any soil type and temperature between 18 and 28 °C (Anwar et al., 2007; Mahmood et al., 2010). It can stand heavy clay, pH of 5.0-9.0, and wide-ranging rainfall with an annual requirement of approximately 250-3,000 mm. M. oleifera can

also grow up to 5–10 m in height and 45 cm in diameter (Prabu et al., 2019). Hitherto, *M. oleifera* was found in the wild and all over the grassland (Mahmood et al., 2010).

4 Phytochemistry of Moringa oleifera

M. oleifera is popularly used as a food additive and health supplement due to its abundant nutritional ingredients, such as vitamins, essential amino acids, minerals and oleic acids (Anwar et al., 2007). It also contains bioactive compounds that potentially contribute to its pharmacological properties and gives beneficial effects on humans (Padayachee and Baijnath, 2012; Luetragoon et al., 2020; Padayachee and Baijnath, 2020). M. oleifera is famously known to possess abundant phytochemicals that are mainly made up of phenolic acids (e.g., chlorogenic acid, gallic acid, and ellagic acid), flavonoids (e.g., kaempferol and quercetin), glucosinolates (GLSs), and isothiocyanates (ITCs) (Kumar et al., 2020; Lopez-Rodriguez et al., 2020; Adamu et al., 2021). The antioxidant properties of *M. oleifera* are often attributable to the presence of these phytochemicals, especially the polyphenolic compounds (phenolic acids and flavonoids). However, GLSs and ITCs have also been gaining interest as important phytochemicals of M. oleifera as it has been shown to exhibit antioxidant and anti-inflammatory activities by altering the detoxification and activation of enzymes (Maldini et al., 2014; Fahey et al., 2018). GLSs contain thioglucosidase (myrosinase) that allows hydrolytic reactions in response to aqueous contact. Consequently, different compounds including ITCs will be constructed after the structural reorder (Dinkova-kostova and Kostov, 2012). In M. oleifera, glucomoringin is the primary GLSs while moringin is the relative ITC present (Galuppo et al., 2013; Borgonovo et al., 2020). Meanwhile, $4-[(4'-O-acetyl-\alpha-L$ rhamnosyloxy) benzyl] isothiocyanate and 4-[(a-L- rhamnosyloxy) benzyl] isothiocyanate (moringin) are among the most abundant formed ITCs (Waterman et al., 2014). Other than that, nitrile glycosides like niazirin and niazirinin, as well as carotenoids and tocopherols were also found in M. oleifera (Adamu et al., 2021).

Interestingly different part of M. oleifera contains different chemical constituents. The leaves are rich in polyphenolic contents, mainly the complex glycosylated flavonoids (eg: quercetin-3-O-glucoside (isoquercitrin), kaempferol-3-O-glucoside (astragalin), quercetin-3-O-rutinoside (rutin) and phenolic acids (eg., chlorogenic acid, caffeic acid and derivatives) (Maldini et al., 2014; Lopez-Rodriguez et al., 2020). These glycosylated flavonoids through hydrolysis readily produce the aglycones, quercetin, and kaempferol (Devaraj et al., 2011; Kumar et al., 2020; Adamu et al., 2021). In addition, M. oleifera leaves contain many flavonoids, such as -(-) epicatechin, myricetin, and rutin (Zhang et al., 2011; Leone et al., 2015). Common phenolic compounds such as caffeic acid, chlorogenic acid, gallic acid, and ferulic acid are found in fruits, seeds, and roots (Singh et al., 2009). The seeds of M. oleifera are widely known for the presence of GLSs (Lopez-Rodriguez et al., 2020), while the stems and flowers contain phenolic compounds and glucosinolates (Saucedo-Pompa et al., 2018). Ironically, some compounds that are known to provide anti-nutritional potentials like tannins, saponins, oxalates, and phytates are also present sparsely in M. oleifera. However, the concentrations of these compounds are often reduced after processes such as maceration and drying (Lopez-Rodriguez et al., 2020). Plant maturity also plays a critical role in determining their nutraceutical potential as higher phytochemical contents were reported in more mature trees (Lopez-Rodriguez et al., 2020).

Flavonoids and phenolic acids are strong antioxidants in M. oleifera that may contribute to its anti-inflammatory, anti-diabetic, and neuroprotective activities (Maldini et al., 2014; Kumar et al., 2020). Besides, the GLSs and ITCs present in M. oleifera, have demonstrated anti-inflammatory and antioxidant activities, with anticancer, chemopreventive, and anti-bacterial potentials (Anwar and Bhanger, 2003; Lopez-Rodriguez et al., 2020). The alkaloids, moringinine moringine are commonly found in the bark of M. oleifera trunks and are responsible for their anti-diabetic properties (Anwar et al., 2007; Kumar et al., 2020). Numerous studies on M. *oleifera* have elucidated its significance in plant nutritional research, supporting its claim as the miracle tree. The plant also contains essential amino acids, such as methionine, and rich source of minerals including phosphorous, iron, calcium, and potassium (Kumar et al., 2020). The seeds are known to have higher lipid content than soybean, mainly palmitic acid, oleic acid and stearic acid which are considered a suitable substitution for olive oil due to the presence of many essential fatty acids. Moreover, M. oleifera leaves are a good source of phytosterols (e.g., β -sitosterol) that may exert hypolipidemic activity (Jain et al., 2010). M. oleifera leaves also consist of other important phytochemicals, such as carotenoids (βcarotene), pro-vitamin A, vitamin C, calcium, and potassium (Anwar et al., 2007; Kumar et al., 2020). In this review, the neuropharmacological potential of M. oleifera crude extract as well as its important bioactive compounds was discussed.

5 Neuroprotective and antineuroinflammatory effects of *M. oleifera* and its bioactive constituents

Traditionally, M. oleifera has been used for various purposes as the panacea for many health conditions, wastewater treatment, and food consumption (Kasolo et al., 2010; Popoola and Obembe, 2013). Although scientific evidence is currently limited, some of the Ayurveda claims on M. oleifera suggest it as an expectorant, diuretic, antispasmodic, and stimulant agent for various kinds of ailments, such as asthma, diabetes, diarrheal, fever, cough, infection, and inflammation as well as neurological disorders like epilepsy, anxiety, and paralysis (Mishra et al., 2011; Hannan et al., 2014). For neurological diseases, the fresh root, flower, and seeds have been suggested to act as stimulants, root juice as anti-epileptic, root and fruit as anti-paralytic, root and bark as anti-viral and analgesic, and lastly, root, bark, and seed as anti-inflammatory agent (Mishra et al., 2011). The beneficial compounds of M. oleifera have provided support for its potential and increased value in nutritional research as a potent pharmaceutical and nutraceutical agent. Numerous studies have suggested the potential of M. oleifera (Mishra et al., 2011; Zahirah et al., 2018; Kumar et al., 2020), and this review is particularly focused on the neuroprotective aspect.

5.1 M. oleifera extracts

Different preparations from different parts of *M. oleifera* have been used to study its neuroprotective and anti-neuroinflammatory potential. For example, the neuroprotective potential of *M. oleifera* leaf

ethanol extract has been evaluated against aluminum (Al)-induced transient cortical degeneration in albino Wistar rats (Ekong et al., 2017). Upon treatment with the extract for 28 days, the histological results revealed reduced degenerative characteristics in the cytoarchitecture of the temporal cortex. However, no significant difference was observed in serum Al for all groups. In addition, while there was also an elevated neuron-specific enolase (NSE) and glial fibrillary acidic protein (GFAP) expression in the Al group, the Moringa-treated group with Al-induced conditions also observed declined expression for both NSE and GFAP. It has been indicated that M. oleifera leaves showed protective potential against neurotoxicity in Al-induced rats. Other than that, the fruit of M. oleifera has also shown anti-AGE activities that were higher than the synthetic antiglycation agent, amino-guanidine (AG) in the bovine serum albumin (BSA)-fructose, and BSA-methylglyoxal assays (Liu et al., 2016).

Besides that, the M. oleifera leaf methanol extract has been tested in sub-chronic chlorpyrifos (CPF)-intoxicated Wistar rats for its potential neuroprotective activities (Idoga et al., 2018). CPF is a commonly used pesticide that is known to be a neurotoxicant as it stimulates oxidative damage to the tissues and leads to an increase in ROS. This has alternately affected the brain as it is most vulnerable to oxidative stress. A previous study has shown that in the CPF-induced group, the malondialdehyde concentration increased, while the activities of acetylcholinesterase (AChE), superoxide dismutase (SOD), glutathione peroxidase (GPX), and catalase (CAT) decreased with evidence of neuronal degeneration, stipulated oxidative stress. Comparatively, the pre-treated group with M. oleifera extracts showed reduced oxidative damage as increased activities were observed. The study suggested that high content of antioxidants, vitamins, and flavonoids is present in the extracts. The study also demonstrated that the neuroprotective potential was not dose-dependent for which 250 mg/kg dose showed better activities than 500 mg/kg.

The 70% ethanolic extract of M. oleifera seeds exhibited neuroprotective potential in scopolamine-induced cognitive impairment in mice (Zhou et al., 2018). In scopolamine-induced groups, impaired cognitive development was observed with reduced reactivity in the cholinergic system and neurogenesis. However, the group pre-treated with the extract showed improved cholinergic reactivity and neurogenesis. It is suggested that the neuroameliorative potential of the extract is mediated by the improved cholinergic system and hippocampal neurogenesis through Akt/ ERK1/2/CREB signaling pathways. In a study by Zeng et al. (2019), the same extract was used to evaluate the potential in acute and delayed stages of cerebral ischemic stroke, which is an injury that results in motor, sensory and cognitive dysfunctions. This study has observed the neuroprotective effect of the seed extracts in both stages of ischemic stages by the increase in animal survival rate, improved cognitive impairment, enhanced neuroplasticity, hippocampal neurogenesis, and cholinergic systems as also supported by a previous finding (Zhou et al., 2018).

Hashim et al. (2021) demonstrated that the ethanolic extracts of *Alpinia galanga* rhizomes (ARE), *Panax ginseng* leaves (PLE), *Alpinia galanga* leaves (ALE), *M. oleifera* leaves (MLE), *Vitis vinifera* seeds (VSE), and *Panax ginseng* rhizomes (PRE) were determined for their neuroprotective potential on human neuroblastoma (SHSY5Y) cells. MLE has an overall high reading of DPPH, FRAP, ROS scavenging, and nitro-blue tetrazolium (NBT) test via 2,7-

dichlorodihydrofluorescein diacetate (DCFHDA) assay. However, the neuroprotection tests on SHSY5Y cells and 3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) cytotoxicity revealed that PRE gave better neuroprotective activities, and higher cytotoxicity as compared to MLE. Thus, the authors have concluded that MLE has the most potential as a neuroprotective agent because of its high antioxidant activities and low cytotoxicity. These have been stipulated by the high content of polyphenol and antioxidant compounds from the plant. The study has also conducted LC-QTOF/MS analysis that confirmed the presence of high phenolic content of MLE.

Besides, the aqueous and ethanolic extracts of M. oleifera leaves were evaluated for neuroprotective potential against H2O2-induced oxidative stress in a PC12 cell line from transplantable rat pheochromocytoma (Kim et al., 2022). The study has found that despite all groups showing similar cell viability, the ethanolic extract was observed to reduce more oxidative stress than vitamin C (positive control group) in the malondialdehyde (MDA) assay of mouse brain homogenates. The ethanolic extract showed higher anti-oxidative activity than the aqueous extract in the 2,2'-Azino-bis (3-ethylbenzthiazoline-6-sulfonic acid) (ABTS) assay. It was suggested that higher activities of the ethanolic extract were attributable to the higher phenolic content as observed in the total phenolic content (TPC) assay as compared to the aqueous extract. The study has also found that the polyphenols, kaempferol and myricetin were the dominant phytochemicals in M. oleifera leaves as determined by highperformance liquid chromatography (HPLC) analysis. Thus, it has been postulated that the ethanol extract of M. oleifera has more potential as a neuro-protective agent due to its high phytochemical content with viable anti-oxidative activities.

5.2 Bioactive constituents

Among the phytochemicals of *M. oleifera*, moringin, astragalin and isoquercitrin have been isolated and investigated for neuroprotective effects. Table 1 shows some of the phytochemicals identified in *M. oleifera* that may potentially contribute to its neuropharmacological activities as well as the proposed mechanisms as discussed in previous studies.

5.2.1 Gallic acid

Gallic acid is a low molecular weight polyphenolic compound that is widely found in tea leaves (black tea and green tea), fruits such as grapes and strawberries as well as polyphenols-rich plants, such as M. oleifera (Badhani et al., 2015; Shabani et al., 2020.) It has been gaining interest as a potent neuroprotective agent due to its antioxidant activities and has been vastly examined for its diverse pharmacological contributions in vivo and in vitro studies (Mirshekar et al., 2018; Ko et al., 2022). Previous studies have demonstrated the activities of gallic acid against neurological disorders, such as PD, AD, ischemia and reperfusion, depression, and anxiety (Shabani et al., 2020). As the development of NDs is primarily associated with oxidative stress and neuroinflammation (Kelsey et al., 2010; Shabani et al., 2020), the high antioxidant and anti-inflammatory properties of gallic acid have suggested its potential against NDs. As such, gallic acid enhanced deoxyribose oxidation and neutralizes the free radicals

TABLE 1 Phytochemicals identified in *M. oleifera* with neuropharmacological effects and their mechanisms of action.

Compound	Chemical structure	Bioassay methods	Mechanisms of action	References		
Phenolic acids						
Gallic acid		Deoxyribose and hydrogen peroxidase assay	Enhanced deoxyribose oxidation and neutralizes free radicals.	Yen et al. (2002)		
	но он	In vitro assay using glioma cell	Dowregulated the PI3K/Akt and rat sarcoma virus (Ras)/MAPK signaling pathways by suppressing coding gene (ADAM17), p-Akt, and p- extracellular signal-regulated kinase (Erk) expression.	Lu et al. (2010)		
	ОН	In vitro using primary rat cortex neuronal culture	Provide anti-oxidative activities and suppressed regulation of proinflammatory cytokines.	Maya et al. (2018)		
		Glutamate-induced neurotoxicity	Malia da idaletare da marte di strata d			
		In vivo male Sprague-Dawley	astrocytes and microglia, proinflammatory	Liu et al. (2020a)		
	0 _{5 2} 0H	*LPS-induced neuroinflammation	enzymes and cytokines, apoptotic cells.			
Chlorogenic acid	HO HO	In vitro assay using primary cortical neurons	Mediation Nrf2-NF-kB pathways by the activation of Sirt1 and further reduce the apoptosis of brain neurons.	Zheng et al. (2022)		
	но н он	In vivo male Sprague Dawley rats	Suppressed hypoxia ischemia-induced			
		*Hypoxia-ischemia brain injury	injury.			
			Inhibited the expression of TNF- α , IL-1 β , and nitric oxide synthase (iNOS) in the brain tissue.			
Ferulic acid	но н	<i>In vitro</i> assay using SH-SY5Y neuroblastoma cells	Activated HO-1/Nrf2 system through carbon monoxide and bilirubin production.	Catino et al. (2016)		
			Prevented disruption of cell lines by up- regulating the HO-1/Nrf2 system.			
Caffeic acid	НО НО НО Н	<i>In vivo</i> male Swiss albino mice	Supressed production of LPS-induced TNF- α and IL-6.	Mallik et al. (2016)		
			Reducing the levels of malondialdehyde and GSH as well as inhibiting the c-Src/ERK pathway of MAPK activation to alleviate oxidative stress.			
		*LPS-induced neuroinflammation	Facilitated downregulation of NF-ĸB- dependent pro-inflammatory genes.			
Flavonoids						
Kaempferol	HO	<i>In vitro</i> assay using murine microglial BV2 cells	Alleviated LPS-induced TNF-a, IL-1b, NO, ROS production, prostaglandin E2 (PGE2), and phagocytosis.	Park et al. (2011)		
			Facilitate the downregulation of TLR4, NF- kB, p38 MAPK, c-Jun N-terminal kinase (JNK), and protein kinase B (AKT) phosphorylation.			
		In vivo rat model of neuropathic pain	Inhibited microglial activation, reduced	Chang et al.		
		Chronic constriction injury (CCI)-induced injury	cytokine production and aneviate pain	(2022)		
Myricetin	он І	<i>In vitro</i> assay using mouse macrophage RAW 264.7	Inhibited NO and pro-inflammatory cytokines.	Kim et al. (2013)		
	но строн он он	*LPS-stimulated cells	Reduce expression of cyclooxygenase-2 (COX-2) and iNOS.			

(Continued on following page)

TABLE 1 (Continued) Phytochemicals identified in M. oleifera with neuropharmacological effects and their mechanisms of action.

Compound	Chemical structure	Bioassay methods	Mechanisms of action	References			
Phenolic acids							
(–)– Epicatechin	ОН	<i>In vivo</i> male C57BL/6 mice (wild type) and Nrf2 gene knockout (KO) mice	Reduced neutrophil infiltration and oxidative stress.	Cheng et al. (2016)			
	HO HO OH	*Traumatic brain injury (TBI)-induced	Facilitate activation of Nrf2 pathway, averted expression of heme oxygenase-1 protein, and reduced iron deposition.				
Quercetin	HO, OH OH OH OH	<i>In vitro</i> assay using SK-N-MC human neuroblastoma cell line <i>In vivo</i> male Sprague-Dawley (SD) rat *Manganese (Mn)-induced neurotoxicity	Alleviated oxidative stress and inhibited cell apoptosis. - Exhibited antioxidant activity by apoptosis regulation, iNOS/NF-KB, and HO-1/ Nrf2 related pathways.	Bahar et al. (2017)			
Isoquercitrin	$HO \xrightarrow{OH}_{HO} \xrightarrow{OH}_{HO} \xrightarrow{OH}_{HO} \xrightarrow{OH}_{HO}$	<i>In vitro</i> assay using mesenchymal stem cells *ROS-Induced Damage	Exhibited ROS scavenging activities.	Li et al. (2016)			
		<i>In vivo</i> male C57BL/6J mice *1-methyl-4-phenyl-1,2,3,6- tetrahydropyridine (MPTP)-induced acute mouse model of PD	Attenuated impaired behaviors and loss of dopamine neurons, elevated expression of dopamine transporter and tyrosine hydroxylase.	Liu et al. (2021)			
			Reduced the expression of pro-apoptotic signaling molecule Bax and supressing MPTP-triggered oxidative stress.				
		In vivo male C57BL mice *LPS-induced neuroinflammation	Suppressed inflammatory responses and inhibited TLR4 and C5aR1 expression, as well as down-regulated MAPK signaling pathways.	Kim et al. (2022b)			
			Inhibited ROS generation.				
		In vitro assay using mesenchymal stem cells	Exhibited ROS scavenging activities.	Li et al. (2016)			
		*ROS-Induced Damage	-				
Astragalin		<i>In vivo</i> female C57BL/6 mice *Neuropathy model	Modulated inflammatory cascade; decreased release of INF-γ and IL-17 (pro- inflammatory cytokines) and increased for IL-10 (anti-inflammatory cytokine). Inhibited inflammatory pathway and impeded voltage-gated ion channels.	Liu et al. (2021)			
Isothiocyanates (ITCs)							
Moringin	S C N N HO O HO OH	In vivo female C57BL/6 mice	Modulated inflammatory cascade; reduced production of INF- γ and IL-17 (pro-inflammatory cytokines) and increased IL-10 (anti-inflammatory cytokine).	Giacoppo et al. (2017)			
		*Neuropathy model	Inhibited inflammatory pathway and impeded voltage-gated ion channels.				

(Continued on following page)

TABLE 1 (Continued)	Phytochemicals	identified in A	<i>I. oleifera</i> with	neuropharmacological	l effects and their	mechanisms of action

Compound	Chemical structure	Bioassay methods	Mechanisms of action	References		
Phenolic acids						
Allyl isothiocyanates (AITC)	s c n	<i>In vitro</i> assay using C6 glioma, BV2 murine microglia, and N2a mouse neuroblastoma cells	Impeded with TNF-α formation in LPS- activated microglia and activation of JNK, NF-κB.	Subedi et al. (2017)		
			Alleviated inducing anti-apoptotic proteins and pro-apoptotic proteins to avert neuronal death.			

effectively in dose-dependent manners (Yen et al., 2002). Gallic acid was also observed to mediate the constriction of ADAM17 in U87 and U251n cell lines as well as down-regulated the PI3K/Akt and Ras/MAPK pathways (Lu et al., 2010). Besides, gallic acid enhanced anti-oxidant activities and suppressed the regulation of proinflammatory cytokines in glutamate-induced neurotoxicity in rats (Maya et al., 2018). It also protects the neurons from neurotoxicity and stabilized the Ca²⁺ homeostasis and IGF-1 expression. In another investigation, it was found to significantly mitigate the elevated biomarkers of activated astrocytes and microglia, as well as the proinflammatory enzymes iNOS, apoptotic cells, and the related cytokines in LPSinduced rat brain (Liu et al., 2020).

5.2.2 Chlorogenic acid

Chlorogenic acid is a group of hydroxycinnamates found in M. oleifera. The HPLC analysis of M. oleifera leaves discovered the presence of chlorogenic acids identified in the form of its isomers which were 3-O-caffeoylquinic acid and 4-O- caffeoylquinic acid (Braham et al., 2020). In a study utilizing high-performance thinlayer chromatography (TLC) method, the contents of chlorogenic acid in M. oleifera were different for each part. It was estimated to be .014% in the root, .017% in the stem, .033% in the leaves, and .022% in the flower (Alam, Alam, Sharaf-Eldin, & Alqarni, 2020). Evidence suggests that chlorogenic acid exhibits the neuroprotective effect (Naveed et al., 2018; Liu et al., 2022). Several studies have demonstrated the benefits of chlorogenic acid in treating neurological disorders, such as ischemia, PD, AD, depression, and cognitive impairments (Kumar et al., 2019; Liu et al., 2020; Caruso et al., 2020; Hung et al., 2021). A study by Oboh et al. (2013) found that chlorogenic acid exerted neuroprotective properties on AD by blocking the activity of acetylcholinesterase (AChE) and butyrylcholinesterase (BChE) and lowering the breakdown of AChE and BChE. A study of cerebral ischemia discovered that chlorogenic acid modulated the Nrf2 pathway and enhanced the expression of Nrf2, HO-1, and NQO-1 to counteract the reperfusion-induced brain injury that caused brain ischemia (Liu et al., 2020). Shah et al. (2021) study has revealed that, chlorogenic acid suppressed oxidative stress by supressing the ROS. It also alleviated the neuronal cell death brought on by localized cerebral ischemia. In vivo and in vitro studies revealed that chlorogenic acid has anti-inflammatory effects by the activation of Sirt1 for Nrf2/NF-KB regulation and reduce the brain neurons apoptosis (Zheng et al., 2022). The previous studies have highlighted the chlorogenic acid neuroprotective and antiinflammatory properties.

5.2.3 Ferulic acid

Ferulic acid, a polyphenol found in M. oleifera, is known for possessing anti-inflammatory and neuroprotective potential. A quantitative analysis of phenolics isolated from M. oleifera has demonstrated the presence of ferulic acid with a retention time value of 12.89 min (Panwar and Mathur, 2020). Qadir et al. (2022) study has identified the phenolics in the M. oleifera leaves methanolic extract, which includes ferulic acid, by HPLC with a UV-visible detector. In NMR-based analysis, ferulic acid was one of the phytoconstituent detected among the targeted metabolites profile in the M. oleifera leaf (Managa et al., 2021). Furthermore, ferulic acid content in M. oleifera has been thoroughly documented, which was linked to its therapeutic benefits. According to previous studies, the neuroprotective properties of ferulic acid have the ability to increase the survival rate of neurons through various mechanisms, such as the inhibition of amyloid protein precursor (APP), fibril-destabilization, and β -amyloid aggregation (Sgarbossa et al., 2015; Kikugawa et al., 2016). A study found that ferulic acid can protect neurons against glutamate-induced toxicity by the increased expression of PEA-15, a phosphoprotein enriched in the astrocytes (Koh, 2012). Ferulic acid also was discovered to prevent disruption by up-regulating the Nrf2/ HO-1 system in human neuroblastoma cell line SH-SY5Y (Catino et al., 2016). In a PD rat model, ferulic acid treatment gives protection to dopaminergic neurons against rotenone-induced damage by restoring antioxidant enzymes, inhibiting lipid peroxidation, and preventing the loss of glutathione (Ojha et al., 2015). Additionally, ferulic acid regulates the β -secretase activity in the transgenic mice of the AD (Mori, et al., 2013). Moreover, Mori et al. (2013) and Mori et al. (2017) demonstrated the anti-inflammatory properties of ferulic acid by alleviating neuroinflammation in presenilin-APP mice and the production of pro-inflammatory cytokines such as TNF-a and IL-1β. Recently, more studies have been performed to discover the importance of ferulic acid in the neuroprotection of M. oleifera.

5.2.4 Caffeic acid

Caffeic acid found in *M. oleifera* has also been noted for its neuroprotective abilities. A study by Oldoni et al. (2019) has found that caffeic acid as the most abundant phenolics compound in *M. oleifera* leaves extracts through the phenolic compounds identification and quantification. In another study, HPLC analysis of *M. oleifera* seed oil has also resulted in the identification of caffeic acid (Gharsallah et al., 2021). Caffeic acid was known to possess protective properties such as neuroprotective and anti-inflammatory activities (Colonnello et al., 2020). According to recent studies, caffeic acid demonstrated neuroprotective effects by regulating oxidative stress and

neuroinflammation (Zhang et al., 2019; Colonnello et al., 2020; Raviteja et al., 2021; Salau et al., 2021). Caffeic acid exhibited protective mechanisms through Nrf2 and skn-1 pathways against 6-hydroxydopamine and quinolinic acid, a neurotoxicity model that led to the increase of ROS and excitotoxicity (Colonnello et al., 2020). Caffeic acid has been reported attenuate neuroinflammation of LPS-induced in mice (Mallik et al., 2016). Moreover, caffeic acid enhances the defense system in the brain by significantly reducing the malondialdehyde levels (Caruso et al., 2020) and increasing the glutathione levels, a major antioxidant (Davis and Vemuganti, 2022). In the AD rats model, Chang et al. (2019) showed the neuroprotective properties of caffeic acid via modulating synaptic plasticity, cerebral insulin signaling, and amyloid-ß formation. Caffeic acid was also reported to suppress the (cyclooxygenase-2) COX-2 pathway, which was directly associated with the onset of tissue damage in neurons (Bare et al., 2019). Previous studies revealed that caffeic acid can inhibit neuroinflammation via diverse mechanisms, which further proves that caffeic acid is an effective agent for neuroprotection.

5.2.5 Kaempferol

Kaempferol is another abundant and common antioxidant flavonoids present in many medicinal plants, such as the Ginkgo biloba, lime trees (Tilia spp.), Chrysanthemum spp., as well as M. oleifera (Silva et al., 2021). Kaempferol has presented a multifaceted pharmacological use with intriguing neuroprotective potential. It has been observed to hinder LPS-induced inflammatory markers, ROS, and phagocytosis by inhibiting NF-kB and TLR4 activation as well as p38 phosphorylation, MAPK, JNK, and AKT (Park et al., 2011; Park et al., 2011; Li et al., 2019a; Silva et al., 2021; Chang et al., 2022). Previous investigations have found that kaempferol contributed to significant reduction of oxidative stress by ROS generation inhibition and related free radical scavenging (Filomeni et al., 2012; Beg et al., 2018). In a study conducted against LPS-triggered TLR4 activation in a neuropathic pain rat model, it was found that kaempferol treatment alleviated neuropathic pain and reduced cytokine production suggested due to the inhibition of microglial activation (Chang et al., 2022). Besides, the same study has displayed that the kaempferol treatment is able to mitigate the TLR4/NF-KB pathways activation in LPS-induced microglial BV2 cells in vitro. In addition, the antioxidant and anti-apoptotic activities mediated by kaempferol have been suggested to result from the enhanced autophagy properties for mitochondrial turnover and consequently offer protection against mitochondrial toxins (Filomeni et al., 2012). This is important as mitochondrial toxins generated the typical signs of PD, suggesting the noteworthy potential of kaempferol as a neuroprotective agent. Another study has also found that the transgenic Drosophila expressing human Aβ-42 (common characterization of AD) exposed to kaempferol has mitigated memory impairment, delayed loss of physical ability (climbing), and decreased in oxidative stress as well as acetylcholinesterase activity (Beg et al., 2018). This has supported the potential of kaempferol as a good neuropharmacological target.

5.2.6 Myricetin

Myricetin was determined to be one of the most abundant polyphenols in *M. oleifera*, contributing to its anti-inflammatory and neuroprotective properties. In UHPLC analysis by Prabakaran et al. (2018) discovered the concentration of myricetin was among

the highest compared to other flavanols (i.e. quercetin and kaempferol), ranging from 600-1,530 µg/g. In previous studies, myricetin was discovered to exert neuroprotective effects by inhibiting the inflammatory response (Huang et al., 2018; Chen et al., 2020). Myricetin alleviated inflammatory responses in LPSinduced models by interediating with the AKT/IKK/NF- κ B pathway in mastitis to suppress the release of pro-inflammatory cytokines, such as IL-1β, IL-16, and TNF-α (Kan et al., 2019; Chen et al., 2020). In vivo and in vitro studies also revealed that myricetin have neuroprotective properties in LPS-induced models (Huang et al., 2018). In LPS administration in rats, activated microglia release the inflammatory mediators that are the principal cause of microglia-mediated neuroinflammation. It was found that myricetin treatment down-regulated the expression of proinflammatory factors, which were significantly up-regulated by the administration of LPS. Along with that, the treatment of myricetin improved altered motor behavior and prevented the loss of dopaminergic neurons (Huang et al., 2018). Moreover, in vitro study discovered that myricetin treatment prevented the death of neuronal cell SH-SY5Y neuroblastoma cell line by suppressing the neurotoxicity effects stimulated by LPS (Huang et al., 2018). In addition, myricetin decreased the activation of microglia in mice's hippocampus and cortical regions after LPS administration (Jang, Lee, Jung, & Park, 2020). Thus, the current studies provide more evidence that myricetin possesses neuroprotective properties by inhibiting neuroinflammation.

5.2.7 (-)- Epicatechin

(-)- Epicatechin (EC) is important flavonols found in many natural sources like cocoa, green tea, as well as M. oleifera with significant neuroprotective abilities. The EC has shown positive effects on cognitive function, which related to improved hippocampal function (Field et al., 2011; Chang et al., 2014). Besides, EC helps to stimulate NO production in endothelial cells which enhances the flow of the blood brain and consequently facilitate in cognitive function (Garate-carrillo et al., 2020). The in vivo pharmacokinetics studies of EC have demonstrated the ability for blood brain barrier (BBB) penetration upon intravenous and oral administration with potential effect on the neurons and supporting systems (Wu et al., 2012; Chang et al., 2014; Garate-carrillo et al., 2020). Even though the exact mechanism is still scarce, the neuroprotective activities of EC is attributable to its antioxidant properties, which gives effect on the synaptic plasticity by interfering with cascade of cell signaling mechanism (Williams and Spencer, 2012; Haskell-ramsay et al., 2018). EC positively influences the mitigation of anxiety, improved learning and memory, various linked-effects of enhanced angiogenesis, neuronal survival and functions, the upregulation mRNA of learning-associated proteins and downregulation of biomarkers for neurodegeneration in hippocampus (Stringer et al., 2015; Garate-carrillo et al., 2020). Besides, EC has displayed neuroprotective potential by the decreased in Aβ-induced apoptosis that is partially related to the p38 MAPK and JNK pathways activation (Ramiro-puig et al., 2009; Haskell-ramsay et al., 2018) as well as the activities against prooxidants and free radicals through the activation of Nrf2 signaling pathway (Ruijters et al., 2013). Meanwhile, EC when tested against TBI-induced brain injury in mice exhibited neuroprotective activity by mediating the Nrf2 pathway with the inhibition of HO-1 and decreased iron deposition (Cheng et al., 2016).

5.2.8 Quercetin

Quercetin is among the abundant flavonoids of M. oleifera (Vergara-jimenez, Almatrafi, & Fernandez, 2017). The leaves contain the highest quercetin, among other flavonoid compounds with four of the quercetin derivatives made up the majority of the flavonoids (Coppin et al., 2013; Zhu et al., 2020; Gao et al., 2022). Moreover, Makita et al. (2016) and Lin et al. (2018) have revealed that the quercetin present is in significant amounts as glycosides associated with a variety of sugar moieties. The bioactive polyphenol structure of quercetin contribute to its anti-inflammatory and neuroprotective properties (Lesjak et al., 2018; Lin et al., 2021). Based on an in vivo study, quercetin is highly permeable across the BBB (Youdim et al., 2014). Quercetin-pretreated SK-N-MC cell was observed to alleviate Mn-induced neurotoxicity by improved cell viability and reduced LDH with remarkable up-regulation of HO-1/Nrf2 and downregulation of NF-kB pathways (Bahar et al., 2017). In addition, the in vivo study has found that quercetin treatment significantly suppressed Mn-induced oxidative stress and neuroinflammation suggested by mediating antioxidant activities via apoptosis, iNOS/ NF-KB, and HO-1/Nrf2 pathways (Bahar et al., 2017). This has consequently restricted the expressions of inflammatory markers and reduced subsequent apoptotic releases, hence, provide with neuroprotective effects. In a Drosophila model of AD, quercetin exhibits neuroprotective effects against AB toxicity in the brain by regulating the protein expression of cyclin B (Kong et al., 2016). Additionally, quercetin reduced hypoxia-induced memory impairment in rats. It attenuated neurodegeneration by reducing the oxidative stress and caspase-3 expression in brain hippocampus (Prasad et al., 2013). A study by Dong et al. (2017) found that quercetin acts against neuronal cell death in mice through the activation of Nrf2 and D-galactose-induced cognitive impairment. It also showed increase of Nrf2-targeted antioxidant enzymes HO-1 and SOD. In an LPS-induced oxidative stress study, it was shown that quercetin suppressed the formation of intracellular ROS in response to LPS, as well as inhibiting NOX2 expression, IkBa degradation, and nuclear translocation of NF-KB, which reduced the levels of inflammatory factors (Sul and Ra, 2021). Quercetin also markedly improved memory impairments in okadaic acid-induced mice by suppressing Tau phosphorylation mediated by cyclin-dependent kinase 5 and reduced production of neurofibrillary tangles, indicating its potential in neuroprotection. Hence, it has been postulated that quercetin has demonstrated its ability for neuroprotection and anti-inflammation in numerous studies.

5.2.9 Isoquercitrin

Glycosylated flavonoids, isoquercitrin is a notable antioxidant and neuroprotective agent found abundantly in *M. oleifera* leaves (Shi et al., 2021; Luiza et al., 2022). Isoquercitrin has been shown to attenuate affected behaviors (MPTP)-induced acute mouse model of PD and inhibited the oxidative stress, neuronal cell death, and apoptosis instigated by the injury (Liu et al., 2021). It is suggested to alleviate the expression of pro-apoptotic signaling molecule Bax expression and subsequently inhibited the associated MPTPtriggered oxidative stress. Besides, isoquercitrin displayed protection against ROS-induced damage on mesenchymal stem cells by exhibiting ROS scavenging activities (Li et al., 2016). The neuroprotective potential of isoquercitrin has been appraised in many studies, as such, one study has found that the oral administration of isoquercitrin protected the hippocampal neurons from streptozotocin

(STZ)- induced neurotoxicity in rats (Chen et al., 2020). The treatment showed inhibition against the STZ-induced oxidative stress and apoptosis as well as an improved cognitive and behavioral impairment in rats. The potential of isoquercitrin has also been investigated on the cerebral injury resulting from inflammatory response upon ischemia and reperfusion by using neuron in vitro model (oxygen-glucose deprivation and reperfusion (OGD/R)) and rat model (middle cerebral artery occlusion and reperfusion (MCAO/R)) (Shi et al., 2021). Isoquercitrin provided neuroprotective activities by the suppression of the inflammatory responses and inhibited TLR4 and C5aR1 expression that contributed to the cAMP/PKA/ I-κB/NF-κB signaling upon brain injury. In addition, isoquercitrin has been observed to exhibit anti-neuroinflammatory activities against the LPS-activated microglia and hippocampus in mice by the downregulation of MAPK signaling pathways (Kim et al., 2022). It has also displayed inhibition of the ROS generation in microglia and radical scavenging activities.

5.2.10 Astragalin

Astragalin is a remarkable natural flavonoid and kaempferol derivative (kaempferol-3-glucoside) found in many medicinal plants including M. oleifera (Engsuwan et al., 2017). It is one of the most important compounds due to its abundant sources, and a broad spectrum of pharmacological uses. Astragalin has been massively studied for its anti-neuroinflammatory, antioxidative and neuroprotective contributions via the mediation and regulation of many molecular targets such as the transcription factors (NF-KB, TNF-a), enzymes (COX-2, PGE-2 AChE, SOD, GPX), and kinases (iNOS, COX-2, PGE2, JNK, MAPK), apoptotic proteins and inflammatory cytokines (Riaz et al., 2018). Astragalin exhibits an anti-neuroinflammatory reaction by mediating with the downregulation of MAPK signaling pathways, as well as reduced NO, iNOS, and pro-inflammatory cytokines in LPS-induced mice (Kim E. H. et al., 2022). Besides, it was found to inhibit MAPK phosphorylation by an extracellular signal-regulated kinase (ERKs), JNKs, and P38 signaling proteins in the LPS-activated microglia and hippocampus. Astragalin was also found to mitigate inflammation caused by aluminum chloride (AlCl3)/D-galactose (Gal)-induced microglia and astrocytes activation, and attenuated changes of regulating enzymes/markers of oxidative stress (Hu et al., 2022). The neuroprotective activities of astragalin are associated with the free radical scavenging ability and oxidative stress-induced influences on the brain neuronal cells (Wasik and Antkiewicz-Michaluk, 2017; Riaz et al., 2018). Astragalin and isoquercitrin isolated from M. oleifera leaves showed potent anti-oxidative effects as enhanced cell viabilities were observed in H2O2-induced oxidative stress in PC-12 cells (Gao et al., 2022). Although the associated active substances involved in the mechanism were not described, the study has found that the rate of survival for the damaged cells improved as the treatment concentration of isolated compounds increased, suggesting the effect of anti-oxidative activities. In another study of astragalin potential in rats with cerebral ischemia-reperfusion injury, the treatment has found that astragalin is able to improve the brain injury through the mediation of anti-inflammatory, anti-oxidative ability and apoptosis signaling pathway (Chen et al., 2020).

5.2.11 ITCs (Moringin and allyl ITCs)

ITCs are the metabolized products of GLSs, and *M. oleifera* were known to have ITCs with same pharmacophore (R–N=C=S)

from broccoli (e.g. sulforaphane, SF) as well as other cruciferous plants (Waterman et al., 2014). Moringin and 4-[(4'-O-acetyl-α-Lrhamnosyloxy) benzyl] are among identified ITCs in M. oleifera (Tumer et al., 2015). Moringin is a structurally unique derivative of ITCs that has been studied as a potent anti-neuroinflammatory agent in relieving MS-associated neuropathic pain. Moringin isolated from M. oleifera seeds was formulated into a 2% treatment cream for topical application on autoimmune encephalomyelitis murine (an animal model for MS) and alleviated neuropathic pain was observed, postulated by the mediation of inflammatory pathway (Giacoppo et al., 2017). It was found that the treatment cream inhibited the inflammatory pathways by instigating the reduced expression of proinflammatory cytokines (IL-17 and IFN- $\gamma)$ together with increased anti-inflammatory cytokines (IL-10). Moreover, the moringin cream was observed to suppress the voltage-gated ion channel expressions, where alterations on these channels (Nav 1.7, Nav 1.8 KV4.2, and a2d-1) may contribute to the progression of neuropathic pain. Besides that, allyl isothiocyanates (AITC) is other interesting ITCs that have been studied for its neuroprotective potential (Latronico et al., 2021; Tran et al., 2021). It has been previously established that AITC has antiinflammatory effects on LPS-stimulated cells (Wagner et al., 2012; Kamal et al., 2022). Subedi et al. (2017) has investigated the neuroprotective and anti-inflammatory abilities of AITC in LPS-stimulated BV2 murine microglia cells. The study has demonstrated that AITC alleviated NO production, regulated MAPK signaling, and significantly reduced the release of TNF-a and IL-6, presenting its strong anti-inflammatory and neuroprotective potential. Neuroblastoma cells exhibited decreased Bax and cleaved caspase-3 expressions and enhanced production of Bcl-2, as a result of AITC's neuroprotective impact against LPS (Subedi et al., 2017). These discoveries clarify the properties of AITC in neuroprotection and anti-inflammation. These studies of isolated compounds from M. oleifera have provided an interesting insight into the potential of plant-based phytochemicals in various applications, especially for better management of NDs.

6 Pharmacokinetics of M. oleifera

Pharmacokinetics studies about different M. oleifera parts have not been widely explored. However, a recent study by Li et al. (2019b) discovered the pharmacokinetics properties of gastrodigenin rhamnopyranoside (GR), a compound in the seeds of M. oleifera. According to the study on rodents, the time GR took to reach the highest concentration (Cmax) for oral administration was 10 min and 5 min for intravenous administration. After the administration, distribution of 10 mg/kg of GR in rodents plasma and different tissues was in the range of 5-30 min (Li et al., 2019a). Within a short period, GR was rapidly distributed to tissue with high blood flow, such as the spleen, heart and kidney. A very slight amount was distributed to brain, liver, and lung implying that the distribution may be influenced by the perfusion rate of the organs (Li et al., 2019b; Alia et al., 2022). Additionally, the presence of GR in the brain tissue demonstrates its potential to cross the BBB. Furthermore, within 30 min, the GR concentration in the tissues was seen to drop significantly and completely eliminated in 3 h. The half-life $(t_{1/2})$ of GR was between 20-30 min, suggesting that the GR was quickly cleared from the circulatory systems (Li et al., 2019b). *M. oleifera* was discovered to have a low bioavailability of iron because of the presence of high phytic acid (Azlan et al., 2022; Kashyap et al., 2022). In contrast, *M. oleifera* leaves have high bioavailability of folate compounds. In a study of rat models, the folates from *M. oleifera* were proven to be 81.9% more bioavailable than the synthetic folates (Saini et al., 2016; Kashyap et al., 2022). Despite that, the bioavailability of *M. oleifera* in various models may vary due to different chemical structures, solubility and their interactions with other compounds.

7 Safety and toxicology assessment of *M. oleifera*

Following the massive properties of M. oleifera, its safety and toxicity were appraised critically in many studies involving both in vivo and in vitro evaluations. Even though M. oleifera plant has been studied for its pharmacological targets due to its nutritional contents, the effect of the extraction procedures and preparation materials on the reported activities remains precarious. These added components may have likely contributed to any conflicting inhibitory, additive, or synergistic potential that were reflected in the pharmacological activities (Adamu et al., 2021). Therefore, the safety and toxicity assessment as well as the availability of a control group as a reference is important in many study designs. While many of the in vivo and in vitro studies of M. oleifera have been conducted by using different extracts, preparation methods and solvents, most human studies have used powdered leaves (Stohs and Hartman, 2015). However, studies have found that the extracts of M. oleifera have a relatively high level of safety, across the different preparation materials. To date, nearly all published studies have showed promising findings of M. oleifera with no critical safety issues or inauspicious findings reported.

In an *in vitro* study, the toxicity of *M. oleifera* was assessed against normal cell lines like peripheral blood mononuclear cells (PBMCs) and cancerous cell lines. The cytotoxicity assay across different concentrations of the *M. oleifera* leaves aqueous extract has shown that, at 20 mg/ml and above, the lactate dehydrogenase (LDH) enzymes increased proportionally, indicating its cytotoxicity as LDH was released during cell damage or lysis (Asare et al., 2012). But, the extract was still considered safe as the 20 mg/ml was unlikely achievable for oral administration (Stohs and Hartman, 2015). The ethanolic seed extract of *M. oleifera* evaluated against cancerous and non-cancerous cell lines showed that no cytotoxicity was observed up to 100 µg/ml concentrations. It was found that no inhibitory activities were observed in the non-cancerous cell, but significantly decreased cell viability for the cancerous cells indicates the extracts *in vitro* safety as well as anti-carcinogenic potential (Aldakheel et al., 2020).

Safety and toxicity have been commonly estimated in the *in vivo* studies involving experimental animals like rodents and rabbits. The studies have found that the extracts (eg; aqueous and methanol) of *M. oleifera* at 1,000 mg/kg dose of gavage treatment revealed no toxicity and mortality in the experimental rats (Asare et al., 2012; Olayemi et al., 2016) whereas, upon 2000 mg/kg, slight reduction in body weight in dose-dependent manners with no fatality was observed (Adedapo et al., 2009). In addition, there were also no significant changes observed in clinical signs, along with cross-sectional and gross pathology evaluation, suggesting the safety of the extracts at a 2000 mg/kg dose in animal studies (Okumu et al., 2016; Moodley, 2017). However, there was acute toxivity observed at \geq 3,000 mg/kg dose, which is a relatively high dose for supplementation intake (Asare et al., 2012). The typical dose for aqueous extract of *M. oleifera* in experimental rats is about 300 mg/kg which

corresponds to an approximate 3.9 g in 80 kg human (Stohs and Hartman, 2015).

In acute, sub-acute and chronic toxicity tests, M. oleifera extracts did not reveal prominent signs of toxicity. The LD50 of M. oleifera leaves extract was discovered to be 1,585 mg/kg (Awodele et al., 2012). M. oleifera extract at 500-2,000 mg/kg was reported to be non-fatal in animals. In a study, the administration of M. oleifera extracts at 2,000 mg/kg was reported to have no mortality after 4 h of administration (Adedapo et al., 2009). It is also reported that the administration of M. oleifera seed extract in rats at 1,600 mg/kg for 21 days did not lead to significant alterations in red haemoglobin concentration, mean corpuscular haemoglobin concentration, packed cell volume, and blood cell count (Ajibade et al., 2012). In addition, experimental mice given bark extract orally at doses of 500, 1,000, and 2,000 mg/kg for 28 days showed no mortality or clinical symptoms (Reddy et al., 2013). At dosages over this level, the animals may show some toxic effects. Moreover, this assertion may not be applicable to long-term use (Awodele et al., 2012). A toxicity study in Wistar albino mice showed dullness and reduced locomotion in gavage treatment of M. oleifera aqueous extract at 3,200 and 6,400 mg/kg after 2 h (Awodele et al., 2012; Rani et al., 2018). However, there were no significant differences were observed in sperm quality, haematological, histological or biochemical characteristics of the rats (Awodele et al., 2012). Nevertheless, some studies have suggested that the toxicity is influenced by the solvents used in the M. oleifera extraction process. The LD₅₀ of ethanol extract was 39,600 mg/kg, and the aqueous extract at 16,100 mg/kg, which was within the safe range (Kasolo et al., 2012). An evaluation of sub-acute toxicity on Swiss albino rats revealed that ethanol solvent was safer as compared to aqueous (Alia et al., 2022). Sub-acute administration of M. oleifera aqueous extracts at 16,100 mg/kg demonstrated mild signs of organ toxicity, such as an increase in the concentrations of white blood cells, potassium ions (K⁺), chloride ions (Cl⁻), and calcium ions (Ca²⁺) and a rise in alkaline phosphatase, aspartate aminotransferase, total bilirubin, and alanine aminotransferase.

Most of the performed studies involving humans use the plant dried leaves powder in the study design, and minimal toxicity was generally assessed. In a previous study, it was found that no adverse effect has been observed in a regimen of nutraceutical *M. oleifera* intake for 40 days continuously (8 g/day doses in tablet form) (Kumari, 2019). Likewise, a single dose of the plant powder intake (50 g) also showed a similar safety outcome with no associated toxicity profile (William et al., 1993). To our best knowledge, no human study has been carried out using the extracts of *M. oleifera* as well as its targeted pharmaceutical compounds, resulting in ambiguous safety and toxicity profiling of the plant extracts in humans. However, despite its massive uses in various applications like cosmetic preparation, anti-bacterial agents for wastewater treatment, food supplementation intake, and nutraceuticals, no adverse effects or safety issues have ever been reported. This has thus, suggested *M. oleifera* as a safe plant in medicinal research.

8 Conclusions and future perspectives

NDs are among the concerning medical conditions that affect millions of people globally. While the statistics keep increasing over years, the natures of the diseases are still not fully understood with poor case management as no definite treatment is currently available besides treatments to facilitate associated symptoms and conditions. Diseases such as PD, AD, and HD are among the common NDs and are projected to have a very high profile of social burden and underlying

medical causalities if taken lightly. Therefore, it is very crucial to find potential alternatives to control the development of diseases. As such, supplementation intake or nutraceuticals is one of the strategic approaches, utilizing the nutritional value of medicinal plants like M. oleifera. M. oleifera is a popular nutritional plant because of its rich source of good nutrients, natural antioxidants, and other phytochemicals that are responsible for its tremendous use and benefits. This plant is highly anticipated in nutritional plant research as it is not only a good source of phytochemicals but also has a minimal need for plant growth, allowing the natural distribution and growth of these plants in many countries. The idea of employing natural products in medical research over synthetic components has recently gained interest. This will not only be beneficial medically, but it is also environmentally conscious and cost-effective. Thus, this review of the recent studies has provided insight into the potential anti-oxidative, antiinflammatory, and neuroprotective properties of M. oleifera against neurodegenerative conditions. However, as most of the studies used different extracts in mitigating diverse neurological conditions, the potential of constituents of M. oleifera as neuroprotective agents needs further investigation. In addition, the application of systems biology as an interesting approach and, the incorporation of this technology in neuropharmacology and medicinal plant research warrant future investigations. Hence, this review has presented the current research gap in the study of *M. oleifera* potential against NDs. More research is necessary to be carried out in a future perspective on the important phytochemicals contributing to the management of NDs as well as understanding its mechanism of action.

Author contributions

Conceptualization, HSH and IJ; validation, HSH, IJ, WMA, IT, DY, WZWN, and HAD; investigation, HSH, UKA, and NAKA; data curation, UKA, XT, and AM; writing initial draft, UKA and HSH; editing, UKA, HSH, IJ, NAKA, and AM; funding acquisition, HSH; supervision, HSH. All authors read and approved the final manuscript.

Funding

This work was funded by the Ministry of Education Malaysia under the Fundamental Research Grant Scheme (grant number FRGS/ 1/2021/STG02/UKM/03/1).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Acioglu, C., Li, L., and Elkabes, S. (2021). Contribution of astrocytes to neuropathology of neurodegenerative diseases. *Brain Res.* 1758 (7), 147291. doi:10.1016/j.brainres.2021. 147291

Adamu, U. M., Lawal, H., and Ramasamy, R. (2021). Immunomodulatory functions of Moringa oleifera (lam.). Malays. J. Med. Health Sci. 17 (7), 54-63.

Adedapo, A., Mogbojuri, O., and Emikpe, B. (2009). Safety evaluations of the aqueous extract of the leaves of *Moringa oleifera* in rats. *J. Med. Plants Res.* 3 (8), 586–591. doi:10. 5897/jmpr.9001097

Ajibade, T. O., Olayemi, F. O., and Arowolo, R. O. A. (2012). The haematological and biochemical effects of methanol extract of the seeds of *Moringa oleifera* in rats. *J. Med. Plants Res.* 6 (4), 615–621. doi:10.5897/jmpr11.1258

Alagan, A., Jantan, I., Kumolosasi, E., Ogawa, S., Abdullah, M. A., and Azmi, N. (2019). Protective effects of *Phyllanthus amarus* against lipopolysaccharide-induced neuroinflammation and cognitive impairment in rats. *Front. Pharmacol.* 10, 632. doi:10.3389/fphar.2019.00632

Alam, P., Alam, P., Sharaf-Eldin, M. A., and Alqarni, M. H. (2020). Simultaneous identification of rutin, chlorogenic acid and gallic acid in Moringa oleifera by densitometric high-performance thin-layer chromatography method. *J. Planar Chromatogr. - Mod. TLC.* 33 (1), 27–32. doi:10.1007/s00764-019-00002-2

Aldakheel, R. K., Rehman, S., Almessiere, M. A., Khan, F. A., Gondal, M. A., Mostafa, A., et al. (2020). Bactericidal and *in vitro* cytotoxicity of *Moringa oleifera* seed extract and its elemental analysis using laser-induced breakdown spectroscopy. *Pharmaceut* 13, 193–211. doi:10.3390/ph13080193

Alia, F., Putri, M., Anggraeni, N., and Syamsunarno, M. R. A. A. (2022). The potency of *Moringa oleifera* lam. as protective agent in cardiac damage and vascular dysfunction. *Front. Pharmacol.* 12, 724439–724518. doi:10.3389/fphar.2021.724439

Anwar, F., and Bhanger, M. I. (2003). Analytical characterization of *Moringa oleifera* seed oil grown in temperate regions of Pakistan. *J. Agric. Food Chem.* 51 (22), 6558–6563. doi:10.1021/jf0209894

Anwar, F., Latif, S., Ashraf, M., and Gilani, A. H. (2007). *Moringa oleifera*: A food plant with multiple medicinal uses. *Phytother. Res.* 25, 17–25. doi:10.1002/ptr.2023

Asare, G. A., Gyan, B., Bugyei, K., Adjei, S., Mahama, R., Addo, P., et al. (2012). Toxicity potentials of the nutraceutical *Moringa oleifera* at supra-supplementation levels. *J. Ethnopharmacol.* 139 (1), 265–272. doi:10.1016/j.jep.2011.11.009

Atho'illah, M. F., Diyah, Y., Dewi, F., Widyarti, S., Tsuboi, H., and Rifa, M. (2021). Elicited soybean extract attenuates proinflammatory cytokines expression by modulating TLR3/TLR4 activation in high-fat, high-fructose diet mice. *J. Ayurveda Integr. Med.* 12 (1), 43–51. doi:10.1016/j.jaim.2021.01.003

Awodele, O., Oreagba, I. A., Odoma, S., Teixeira Da Silva, J. A., and Osunkalu, V. O. (2012). Toxicological evaluation of the aqueous leaf extract of *Moringa oleifera* Lam. (Moringaceae). J. Ethnopharmacol. 139 (2), 330–336. doi:10.1016/j.jep.2011.10.008

Azam, S., Haque, E., Kim, I., and Choi, D. (2021). Microglial turnover in ageing-related neurodegeneration: Therapeutic Avenue to intervene in disease progression. *Cells* 10 (1), 150. doi:10.3390/cells10010150

Azlan, U. K., Mediani, A., Rohani, E. R., Tong, X., Han, R., Misnan, N. M., et al. (2022). A comprehensive review with updated future perspectives on the ethnomedicinal and pharmacological aspects of *Moringa oleifera*. *Molecules* 27 (18), 5765. doi:10.3390/molecules27185765

Babar, Z. U., Azad, A. K., Wan Sulaiman, W. M. A., Uddin, J., and Labu, Z. K. (2018). Neuroprotective properties of *Nigella sativa* (L.) seeds extract in Sprague Dawley. *Dhaka Univ. J. Pharm. Sci.* 17 (1), 113–121. doi:10.3329/dujps.v17i1.37127

Badhani, B., Sharma, N., and Kakkar, R. (2015). Gallic acid: A versatile antioxidant with promising therapeutic and industrial applications. *RSC Adv.* 5 (35), 27540–27557. doi:10. 1039/C5RA01911G

Bahar, E., Kim, J. Y., and Yoon, H. (2017). Quercetin attenuates manganeseinduced neuroinflammation by alleviating oxidative stress through regulation of apoptosis, iNoS/NF-κB and HO-1/Nrf2 pathways. *Int. J. Mol. Sci.* 18 (9), 1989. doi:10. 3390/ijms18091989

Baidya, F., Bohra, M., Datta, A., Sar, D., Shah, B., Jagtap, P., et al. (2021). Neuroimmune crosstalk and evolving pharmacotherapies in neurodegenerative diseases. *Immunol* 162 (2), 160–178. doi:10.1111/imm.13264

Bare, Y., Krisnamurti, G. C., Elizabeth, A., Rachmad, Y. T., Sari, D. R. T., and Lorenza, M. R. W. G. (2019). The potential role of caffeic acid in coffee as cyclooxygenase-2 (COX-2) inhibitor: In silico study. *Biointerface Res. Appl. Chem.* 9 (5), 4424–4427. doi:10.33263/ briac95.42442

Batista, C. R. A., Gomes, G. F., Candelario-Jalil, E., Fiebich, B. L., and de Oliveira, A. C. P. (2019). Lipopolysaccharide-induced neuroinflammation as a bridge to understand neurodegeneration. *Int. J. Mol. Sci.* 20 (9), 2293. doi:10.3390/ijms20092293

Beg, T., Jyoti, S., Naz, F., Ali, F., Ali, S. K., Reyad, A. M., et al. (2018). Protective effect of kaempferol on the transgenic Drosophila model of Alzheimer's disease. *CNS Neurol. Disord. Drug Targets.* 17, 421–429. doi:10.2174/1871527317666180508123050

Bhattacharya, A., Tiwari, P., Sahu, P. K., and Kumar, S. (2018). A review of the phytochemical and pharmacological characteristics of moringa oleifera. *Moringa Oleifera. J. Pharm. Bioallied Sci.* 10, 181–191. doi:10.4103/JPBS.JPBS_126_18

Boonen, B., Alpizar, Y. A., Sanchez, A., López-Requena, A., Voets, T., and Talavera, K. (2018). Differential effects of lipopolysaccharide on mouse sensory TRP channels. *Cell. Cal.* 73, 72–81. doi:10.1016/j.ceca.2018.04.004

Borgonovo, G., Petrocellis, L. D., Moriello, A. S., Bertoli, S., Leone, A., Battezzati, A., et al. (2020). Moringin, a stable isothiocyanate from *Moringa oleifera*, activates the somatosensory and pain receptor TRPA1 channel *in vitro*. *Molecules* 25 (976), 976–1012. doi:10.3390/molecules25040976

Braham, F., Carvalho, D. O., Almeida, C. M. R., Zaidi, F., Magalhães, J. M. C. S., Guido, L. F., et al. (2020). Online HPLC-DPPH screening method for evaluation of radical scavenging phenols extracted from Moringa oleifera leaves. S. Afr. J. Bot. 129, 146–154. doi:10.1016/j.sajb.2019.04.001

Caruso, G., Fresta, G. C., Grasso, M., Santangelo, R., Lazzarino, G., Lunte, M. S., et al. (2020). Inflammation as the common biological link between depression and cardiovascular diseases: Can carnosine exert a protective role? *Curr. Med. Chem.* 27 (11), 1782–1800. doi:10.2174/0929867326666190712091515

Catino, S., Paciello, F., Miceli, F., Rolesi, R., Troiani, D., Calabrese, V., et al. (2016). Ferulic acid regulates the Nrf2/heme oxygenase-1 system and counteracts trimethyltininduced neuronal damage in the human neuroblastoma cell line SH-SY5Y. *Front. Pharmacol.* 6, 305–312. doi:10.3389/fphar.2015.00305

Catorce, M. N., and Gevorkian, G. (2020). Evaluation of anti-inflammatory nutraceuticals in lps-induced mouse neuroinflammation model: An update. *Curr. Neuropharmacol.* 18 (7), 636–654. doi:10.2174/1570159X18666200114125628

Chang, C., Cho, S., and Wang, J. (2014). (-) Epicatechin protects hemorrhagic brain via synergistic Nrf2 pathways. Ann. Clin. Transl. Neurol. 1 (4), 258–271. doi:10.1002/acn3.54

Chang, S., Zheng, Y., Shi, H., Zhang, D., Chen, Z., Qian, G., et al. (2022). Kaempferol exerts a neuroprotective effect to reduce neuropathic pain through TLR4/NF-KB signaling pathway. *Phytother. Res.* 36, 1678–1691. doi:10.1002/ptr.7396

Hu, Y., Fang, X., Wang, J., Ren, T.-T., Zhao, Y.-Y., Dai, J.-F., et al. (2022). Astragalin attenuates AlCl3/D-galactose-induced aging-like disorders by inhibiting oxidative stress and neuroinflammation. *NeuroToxicol.* 91 (2022), 60-68. doi:10.1016/j.neuro.2022.05.003

Chang, W., Huang, D., Lo, Y. M., Tee, Q., Kuo, P., Wu, J. S., et al. (2019). Protective effect of caffeic acid against alzheimer's disease pathogenesis via modulating cerebral insulin signaling, β -Amyloid accumulation, and synaptic plasticity in hyperinsulinemic rats. *J. Agric. Food Chem.* 67 (27), 7684–7693. doi:10.1021/acs.jafc.9b02078

Che, D. N., Cho, B. O., Kim, J. S., Shin, J. Y., Kang, H. J., and Jang, S. Il. (2020). Effect of luteolin and apigenin on the production of IL-31 and IL-33 in lipopolysaccharides-activated microglia cells and their mechanism of action. *Nutrients* 12 (3), 811. doi:10.3390/nu12030811

Chen, L., Deng, H., Cui, H., Fang, J., Zuo, Z., Deng, J., et al. (2018). Inflammatory responses and inflammation-associated diseases in organs. *Oncotarget* 9 (6), 7204–7218. doi:10.18632/oncotarget.23208

Chen, L., Feng, P., Peng, A., Qiu, X., Lai, W., Zhang, L., et al. (2020b). Protective effects of isoquercitrin on streptozotocin-induced neurotoxicity. *J. Cell. Mol. Med.* 24, 10458–10467. doi:10.1111/jcmm.15658

Chen, M., Chen, Z., Huang, D., Sun, C., Xie, J., Chen, T., et al. (2020a). Myricetin inhibits TNF- α -induced inflammation in A549 cells via the SIRT1/NF- κ B pathway. *Pulm. Pharmacol.* 65 (9), 102000. doi:10.1016/j.pupt.2021.102000

Chen, W., Zhang, X., and Huang, W. J. (2016). Role of neuroinflammation in neurodegenerative diseases (Review). *Mol. Med. Rep.* 13, 3391–3396. doi:10.3892/mmr. 2016.4948

Chen, X., Cheng, C., Zuo, X., and Huang, W. (2020c). Astragalin alleviates cerebral ischemia-reperfusion injury by improving anti-oxidant and anti-inflammatory activities and inhibiting apoptosis pathway in rats. *BMC Complement. Med. Ther.* 20, 120. doi:10. 1186/s12906-020-02902-x

Cheng, T., Wang, W., Li, Q., Han, X., Xing, J., Qi, C., et al. (2016). Cerebroprotection of flavanol (-)-epicatechin after traumatic brain injury via Nrf2-dependent and -independent pathways. *Free Radic. Biol. Med.* 92 (3), 15–28. doi:10.1016/j. freeradbiomed.2015.12.027

Colonnello, A., Aguilera-Portillo, G., Rubio-López, L. C., Robles-Bañuelos, B., Rangel-López, E., Cortez-Núñez, S., et al. (2020). Correction to: Comparing the neuroprotective effects of caffeic acid in rat cortical slices and *Caenorhabditis elegans*: Involvement of Nrf2 and SKN-1 signaling pathways. *Neurotox. Res.* 37 (3), 779. doi:10.1007/s12640-019-00153-4

Coppin, J. P., Xu, Y., Chen, H., Pan, M. H., Ho, C. T., Juliani, R., et al. (2013). Determination of flavonoids by LC/MS and anti-inflammatory activity in *Moringa* oleifera. J. Funct. Foods. 5 (4), 1892–1899. doi:10.1016/j.jff.2013.09.010

Davis, C. K., and Vemuganti, R. (2022). Antioxidant therapies in traumatic brain injury. *Neurochem. Int.* 152, 105255. doi:10.1016/j.neuint.2021.105255

Devaraj, V. C., Krishna, B. G., and Viswanatha, G. L. (2011). Simultaneous determination of quercetin, rutin and kaempferol in the leaf extracts of *Moringa oleifera* Lam. and *raphinus sativus* Linn, by liquid chromatography-tandem mass spectrometry. *J. Chin. Integr. Med.* 9, 1022–1030. doi:10.3736/jcim20110914

Dinkova-kostova, A. T., and Kostov, R. V. (2012). Glucosinolates and isothiocyanates in health and disease. *Trends Mol. Med.* 18 (6), 337–347. doi:10.1016/j.molmed.2012.04.003

Dong, F., Wang, S., Wang, Y., Yang, X., Jiang, J., Wu, D., et al. (2017). Quercetin ameliorates learning and memory via the Nrf2-ARE signaling pathway in D-galactose-induced neurotoxicity in mice. *Biochem. Biophys. Res. Commun.* 491 (3), 636–641. doi:10. 1016/j.bbrc.2017.07.151

Durani, L. W., Hamezah, H. S., Ibrahim, N. F., Yanagisawa, D., Nasaruddin, M. L., Mori, M., et al. (2018). Tocotrienol-rich fraction of palm oil improves behavioral impairments and regulates metabolic pathways in AβPP/PS1 Mice. *J. Alzheimer's Dis.* 64 (1), 249–267. doi:10.3233/JAD-170880

Eikelenboom, P., Bate, C., Van Gool, W., Hoozemans, J., Rozemuller, J., Veerhuis, R., et al. (2002). Neuroinflammation in Alzheimer's disease and prion disease. *Glia* 40 (2), 232–239. doi:10.1002/glia.10146

Ekong, M. B., Ekpo, M. M., Akpanyung, E. O., and Nwaokonko, D. U. (2017). Neuroprotective effect of *Moringa oleifera* leaf extract on aluminium-induced temporal cortical degeneration. *Metabol. Brain Dis.* 32 (5), 1437–1447. doi:10.1007/s11011-017-0011-7

Engsuwan, J., Waranuch, N., Limpeanchob, N., and Ingkaninan, K. (2017). HPLC methods for quality control of *Moringa oleifera* extract using isothiocyanates and astragalin as bioactive markers. *Sci. Asia.* 43 (3), 169–174. doi:10.2306/scienceasia1513-1874.2017.43.169

Fahey, J. W., Olson, M. E., Stephenson, K. K., Wade, K. L., Chodur, G. M., Odee, D., et al. (2018). The diversity of chemoprotective glucosinolates in *Moringaceae* (Moringa spp.). *Sci. Rep.* 8 (1), 7994–8014. doi:10.1038/s41598-018-26058-4

Field, D. T., Williams, C. M., and Butler, L. T. (2011). Consumption of cocoa flavanols results in an acute improvement in visual and cognitive functions. *Physiol. Behav.* 103 (3-4), 255–260. doi:10.1016/j.physbeh.2011.02.013

Figuera-Losada, M., Rojas, C., and Slusher, B. S. (2014). Inhibition of microglia activation as a phenotypic assay in early drug discovery. *J. Biomol. Screen.* 19 (1), 17–31. doi:10.1177/1087057113499406

Filomeni, G., Graziani, I., Zio, D. D., Dini, L., Centonze, D., Rotilio, G., et al. (2012). Neuroprotection of kaempferol by autophagy in models of rotenone-mediated acute toxicity: Possible implications for Parkinson's disease. *Neurobiol. Aging.* 33 (4), 767–785. doi:10.1016/j.neurobiolaging.2010.05.021

Frasca, L., and Lande, R. (2020). Toll-like receptors in mediating pathogenesis in systemic sclerosis. *Clin. Exp. Immunol.* 201 (1), 14–24. doi:10.1111/cei.13426

French, J. A., Koepp, M., Naegelin, Y., Vigevano, F., Rho, J. M., Rosenberg, E., et al. (2017). Clinical studies and anti-inflammatory mechanisms of treatments. *Epilepsia* 58 (3), 69–82. doi:10.1111/epi.13779

Galuppo, M., Nicola, G. R. D., Iori, R., Dell'Utri, P., Bramanti, P., and Mazzon, E. (2013). Antibacterial activity of glucomoringin bioactivated with myrosinase against two important pathogens affecting the health of long-term patients in hospitals. *Molecules* 18, 14340–14348. doi:10.3390/molecules181114340

Gao, Q., Wei, Z., Liu, Y., Wang, F., Zhang, S., Serrano, C., et al. (2022). Characterization, large-scale HSCCC separation and neuroprotective effects of polyphenols from *Moringa oleifera* leaves. *Molecules* 27 (3), 678–702. doi:10.3390/molecules27030678

Garate-carrillo, A., Rodriguez, A., Ceballos, G., Calzada-mendoza, C., Hogan, M. C., Villarreal, F., et al. (2020). Effects of (-)-epicatechin on neuroinflammation and hyperphosphorylation of tau in the hippocampus of aged mice. *Food Funct*. 11 (12), 10351–10361. doi:10.1039/d0f602438d

Gharsallah, K., Rezig, L., Msaada, K., Chalh, A., and Soltani, T. (2021). Chemical compositionand profile characterization of *Moringa oleifera* seed oil. S. Afr. J. Bot. 137, 475–482. doi:10.1016/j.sajb.2020.11.014

Giacoppo, S., Iori, R., Bramanti, P., and Mazzon, E. (2017). Topical moringin cream relieves neuropathic pain by suppression of inflammatory pathway and voltage-gated ion channels in murine model of multiple sclerosis. *Mol. Pain.* 13, 1744806917724318–17. doi:10.1177/1744806917724318

Gray, P., Dagvadorj, J., Michelsen, K. S., Brikos, C., Rentsendorj, A., Town, T., et al. (2011). MD-2 interacts with Lyn kinase and is tyrosine phosphorylated following LPSinduced activation of the Toll-like receptor 4 signaling pathway. *J. Immunol.* 187 (8), 4331-4337. doi:10.4049/jimmunol.1100890

Gribar, S. C., Richardson, W. M., Sodhi, C. P., and Hackam, D. J. (2008). No longer an innocent bystander: Epithelial toll-like receptor signaling in the development of mucosal inflammation. *Mol. Med.* 14, 645–659. doi:10.2119/2008-00035.Gribar

Guzman-Martinez, L., Maccioni, R. B., Andrade, V., Navarrete, L. P., Pastor, M. G., and Ramos-Escobar, N. (2019). Neuroinflammation as a common feature of neurodegenerative disorders. *Front. Pharmacol.* 10 (9), 1008–17. doi:10.3389/fphar.2019.01008

Halliwell, B. (2012). Free radicals and antioxidants: Updating a personal view. *Nutr. Rev.* 70, 257–65. doi:10.1111/j.1753-4887.2012.00476.x

Hannan, A., Kang, J., Hong, Y., Lee, H., Choi, J., Soon, I., et al. (2014). Moringa oleifera with promising neuronal survival and neurite outgrowth promoting potentials. J. Ethnopharmacol. 152 (1), 142–50. doi:10.1016/j.jep.2013.12.036

Hashim, F. J., Vichitphan, S., Boonsiri, P., and Vichitphan, K. (2021). Neuroprotective assessment of *Moringa oleifera* leaves extract against oxidative-stress-induced cytotoxicity in SHSY5Y neuroblastoma cells. *Plants* 10 (5), 889–903. doi:10.3390/plants10050889

Haskell-ramsay, C. F., Schmitt, J., and Actis-Goretta, L. (2018). The impact of epicatechin on human cognition: The role of cerebral blood flow. *Nutrients* 10, 986. doi:10.3390/nu10080986

Henkel, J. S., Engelhardt, J. I., Kim, S. H., Pan, T., Goodman, J. C., Siddique, T., et al. (2003). Presence of dendritic cells, MCP-1, and activated microglia/macrophages in amyotrophic lateral sclerosis spinal cord tissue. Am. Neurol. Assoc. 55 (2), 221–235. doi:10.1002/ana.10805

Hsieh, H., and Yang, C. (2013). Role of redox signaling in neuroinflammation and neurodegenerative diseases. *Biomed. Res. Int.* 2013, 484613. doi:10.1155/2013/484613

Huang, B., Liu, J., Ma, D., Chen, G., Wang, W., and Fu, S. (2018). Myricetin prevents dopaminergic neurons from undergoing neuroinflammation-mediated degeneration in a lipopolysaccharide-induced Parkinson's disease model. *J. Funct. Foods.* 45, 452–461. doi:10.1016/j.jff.2018.04.018

Hung, Y. C., Kuo, Y. H., Hsieh, P. W., Hsieh, T. Y., Kuo, J. R., and Wang, S. J. (2021). Chlorogenic acid decreases glutamate release from rat cortical nerve terminals by p/q-type Ca2+ channel suppression: A possible neuroprotective mechanism. *Int. J. Mol. Sci.* 22 (21), 11447. doi:10.3390/ijms222111447

Idoga, E. S., Ambali, S. F., Ayo, J. O., and Mohammed, A. (2018). Assessment of antioxidant and neuroprotective activities of methanol extract of *Moringa oleifera* Lam. leaves in subchronic chlorpyrifos-intoxicated rats. *Comp. Clin. Path.* 27, 917–925. doi:10. 1007/s00580-018-2682-9

Jain, P. G., Patil, S. D., Haswani, N. G., Girase, M. V., and Surana, S. J. (2010). Hypolipidemic activity of *Moringa oleifera Lam.*, Moringaceae, on high fat diet induced hyperlipidemia in albino rats. *Braz. J. Pharmacog.* 20 (6), 969–973. doi:10.1590/S0102-695X2010005000038

Jaja-chimedza, A., Graf, B. L., Simmler, C., Kim, Y., Kuhn, P., Pauli, G. F., et al. (2017). Biochemical characterization and anti-inflammatory properties of an isothiocyanateenriched moringa (*Moringa oleifera*) seed extract. *Plos ONE* 12 (8), e0182658. doi:10. 1371/journal.pone.0182658

Jang, J., Lee, S. H., Jung, K., and Park, G. (2020). Inhibitory effects of myricetin on lipopolysaccharide-induced neuroinflammation. *Brain Sci.* 10 (32), 32–10. doi:10.3390/ brainsci10010032

Jantan, I., Ahmad, W., and Bukhari, S. N. A. (2015). Plant-derived immunomodulators: An insight on their preclinical evaluation and clinical trials. *Front. Plant Sci.* 6 (8), 655–18. doi:10.3389/fpls.2015.00655

Jantan, I., Haque, M. A., Arshad, L., Harikrishnan, H., Septama, A. W., and Mohamed-Hussein, Z. A. (2021). Dietary polyphenols suppress chronic inflammation by modulation of multiple inflammation-associated cell signaling pathways. *J. Nutr. Biochem.* 93, 108634. doi:10.1016/j.jnutbio.2021.108634

Kabir, M. T., Rahman, M. H., Shah, M., Jamiruddin, M. R., Basak, D., Al-Harrasi, A., et al. (2022). Therapeutic promise of carotenoids as antioxidants and anti-inflammatory agents in neurodegenerative disorders. *Biomed. Pharmacother.* 146, 112610. doi:10.1016/j. biopha.2021.112610

Kamal, R. M., Razis, A. F. A., Sukri, N. S. M., Perimal, E. K., Ahmad, H., Patrick, R., et al. (2022). Beneficial health effects of glucosinolates-derived isothiocyanates on cardiovascular and neurodegenerative diseases. *Molecules* 27 (3), 624–49. doi:10.3390/ molecules27030624

Kan, X., Liu, B., Guo, W., Wei, L., Lin, Y., Guo, Y., et al. (2019). Myricetin relieves LPSinduced mastitis by inhibiting inflammatory response and repairing the blood-milk barrier. J. Cell. Physiol. 234 (9), 16252-16262. doi:10.1002/jcp.28288

Kashyap, A. S., Manzar, N., Nebapure, S. M., Rajawat, M. V. S., Deo, M. M., Singh, J. P., et al. (2022). Unraveling microbial volatile elicitors using a transparent methodology for induction of systemic resistance and regulation of antioxidant genes at expression levels in chili against bacterial wilt disease. *Antioxidants* 11 (2), 404–37. doi:10.3390/ antiox11020404

Kasolo, J. N., Bimenya, G. S., Ojok, L., Ochieng, J., and Ogwal-Okeng, J. W. (2010). Phytochemicals and uses of *Moringa oleifera* leaves in Ugandan rural communities. *J. Med. Plants Res.* 4 (9), 753–757. doi:10.5897/JMPR10.492

Kasolo, J. N., Bimenya, G. S., Ojok, L., and Ogwal-Okeng, J. W. (2012). Sub-acute toxicity evaluation of *Moringa oleifera* leaves aqueous and ethanol extracts in Swiss Albino rats. *Int. J. Med. Plant Res.* 1 (6), 75–81.

Kawasaki, T., and Kawai, T. (2014). Toll-like receptor signaling pathways. Front. Immunol. 5 (461), 461-8. doi:10.3389/fimmu.2014.00461

Kelsey, N. A., Wilkins, H. M., and Linseman, D. A. (2010). Nutraceutical antioxidants as novel neuroprotective agents. *Molecules* 15 (11), 7792–7814. doi:10.3390/ molecules15117792

Khan, H. U., Aamir, K., Jusuf, P. R., Sethi, G., Sisinthy, S. P., Ghildyal, R., et al. (2020). Lauric acid ameliorates lipopolysaccharide (LPS)-induced liver inflammation by mediating TLR4/MyD88 pathway in Sprague Dawley (SD) rats. *Life Sci.* 15 (265), 118750. doi:10.1016/j.lfs.2020.118750

Kikugawa, M., Tsutsuki, H., Ida, T., Nakajima, H., Ihara, H., and Sakamoto, T. (2016). Water-soluble ferulic acid derivatives improve amyloid-β-induced neuronal cell death and dysmnesia through inhibition of amyloid-β aggregation. *Biosci. Biotechnol. Biochem.* 80 (3), 547–553. doi:10.1080/09168451.2015.1107463

Kim, E. H., Shim, Y. Y., Lee, H. I., Lee, S., Reaney, M. J. T., and Chung, M. J. (2022b). Astragalin and isoquercitrin isolated from *Aster scaber* suppress LPS-induced neuroinflammatory responses in microglia. *Foods* 11, 1505. doi:10.3390/foods11101505

Kim, E. K., and Choi, E. J. (2010). Pathological roles of MAPK signaling pathways in human diseases. *Biochim. Biophy. Acta Mol. Basis Dis.* 1802 (4), 396–405. doi:10.1016/j. bbadis.2009.12.009

Kim, H. H., Kim, D. H., Kim, M. H., Oh, M. H., Kim, S. R., Park, K. J., et al. (2013). Flavonoid constituents in the leaves of *Myrica rubra sieb. et zucc.* with anti-inflammatory activity. *Arch. Pharm. Res.* 36 (12), 1533–1540. doi:10.1007/s12272-013-0147-x

Kim, H. S., Jeong, C., and Lee, J. (2022a). Neuroprotective effects of *Moringa oleifera* leaf extracts. *Food Life* 2022 (1), 19–26. doi:10.5851/fl.2022.e1

Ko, E. B., Jang, Y. G., Kim, C. W., Go, R. E., Lee, H. K., and Choi, K. C. (2022). Gallic acid hindered lung cancer progression by inducing cell cycle arrest and apoptosis in a549 lung cancer cells via PI3K/Akt pathway. *Biomol. Ther.* 30 (2), 151–161. doi:10.4062/biomolther.2021.074

Koh, P. O. (2012). Ferulic acid prevents the cerebral ischemic injury-induced decreases of astrocytic phosphoprotein PEA-15 and its two phosphorylated forms. *Neurosci. Lett.* 511 (2), 101–105. doi:10.1016/j.neulet.2012.01.049

Kong, Y., Li, K., Fu, T., Wan, C., Zhang, D., Song, H., et al. (2016). Quercetin ameliorates A β toxicity in Drosophila AD model by modulating cell cycle-related protein expression. *Oncotarget* 7 (42), 67716–67731. doi:10.18632/ONCOTARGET.11963

Kumar, A., Kumar, H., Tshabalala, T., Kumar, R., Gupta, A., Ndhlala, A. R., et al. (2020). Phytochemical, nutraceutical and pharmacological attributes of a functional crop *Moringa oleifera Lam*: An overview. S. *Afr. J. Bot.* 129, 209–20. doi:10.1016/j.sajb.2019.06.017

Kumar, G., Mukherjee, S., Paliwal, P., Singh, S. S., Birla, H., Singh, S. P., et al. (2019). Neuroprotective effect of chlorogenic acid in global cerebral ischemia-reperfusion rat model. *Naunyn-Schmiedeb. Arch. Pharmacol.* 392 (10), 1293–1309. doi:10.1007/s00210-019-01670-x

Kumari, D. (2019). Hypoglycaemic effect of *Moringa oleifera* and *Azadirachta indica* in type 2 diabetes mellitus. *Bioscan* 5, 211–4.

Latronico, T., Larocca, M., Milella, S., Fasano, A., Rossano, R., and Liuzzi, G. M. (2021). Neuroprotective potential of isothiocyanates in an *in vitro* model of neuroinflammation. *Inflammopharmacol* 29 (2), 561–571. doi:10.1007/s10787-020-00772-w

Leng, F., and Edison, P. (2021). Reviews- neuroinflammation and microglial activation in alzheimer disease: Where do we go from here? *Nat. Rev. Neurol.* 17 (3), 157–72. doi:10. 1038/s41582-020-00435-y

Leone, A., Spada, A., Battezzati, A., Schiraldi, A., Aristil, J., and Bertoli, S. (2015). Cultivation, genetic, ethnopharmacology, phytochemistry and pharmacology of *Moringa oleifera* leaves: An overview. *Int. J. Mol. Sci.* 16 (6), 12791–835. doi:10.3390/ijms160612791

Lesjak, M., Beara, I., Simin, N., Pintać, D., Majkić, T., Bekvalac, K., et al. (2018). Antioxidant and anti-inflammatory activities of quercetin and its derivatives. *J. Funct. Foods.* 40, 68–75. doi:10.1016/j.jff.2017.10.047

Li, W., Cheng, X., Yang, Y., Liu, M., Zhang, S., Wang, Y., et al. (2019a). Kaempferol attenuates neuroinflammation and blood brain barrier dysfunction to improve neurological deficits in cerebral ischemia/reperfusion rats. *Brain Res.* 1722, 146361. doi:10.1016/j.brainres.2019.146361

Li, W., Sun, C., Deng, W., Liu, Y., Adu-Frimpong, M., Yu, J., et al. (2019b). Pharmacokinetic of gastrodigenin rhamnopyranoside from *Moringa* seeds in rodents. *Fitoterapia* 138, 104348. doi:10.1016/j.fitote.2019.104348

Li, X., Jiang, Q., Wang, T., Liu, J., and Chen, D. (2016). Comparison of the antioxidant effects of quercitrin and isoquercitrin: Understanding the role of the 6"-OH group. *Molecules* 21 (9), 1246. doi:10.3390/molecules21091246

Lin, M. C., Liu, C. C., Liao, C. S., and Ro, J. H. (2021). Neuroprotective effect of quercetin during cerebral ischemic injury involves regulation of essential elements, transition metals, cu/zn ratio, and antioxidant activity. *Molecules* 26 (20), 6128. doi:10.3390/molecules26206128

Lin, M., Zhang, J., and Chen, X. (2018). Bioactive flavonoids in Moringa oleifera and their health-promoting properties. *J. Funct. Foods* 47, 469–479. doi:10.1016/j.jff.2018. 06.011

Liu, C., Wang, W., Li, H., Liu, J., Zhang, P., Cheng, Y., et al. (2021). The neuroprotective effects of isoquercitrin purified from apple pomace by high-speed countercurrent chromatography in the MPTP acute mouse model of Parkinson's disease. *Food Funct.* 12 (13), 6091–101. doi:10.1039/d1fo00843a

Liu, D., Wang, H., Zhang, Y., and Zhang, Z. (2020b). Protective effects of chlorogenic acid on cerebral ischemia/reperfusion injury rats by regulating oxidative stress-related nrf2 pathway. *Drug Des. devel. Ther.* 14, 51–60. doi:10.2147/DDDT.S228751

Liu, W., Ma, H., Dasilva, N. A., Rose, K. N., Johnson, S. L., Zhang, L., et al. (2016). Development of a neuroprotective potential algorithm for medicinal plants. *Neurochem. Int.* 100, 164–77. doi:10.1016/j.neuint.2016.09.014

Liu, Y. L., Hsu, C. C., Huang, H. J., Chang, C. J., Sun, S. H., and Lin, A. M. Y. (2020a). Gallic acid attenuated LPS-induced neuroinflammation: Protein aggregation and necroptosis. *Mol. Neurobiol.* 57 (1), 96–104. doi:10.1007/s12035-019-01759-7

Liu, Y., Wang, F., Li, Z., Mu, Y., Yong, V. W., and Xue, M. (2022). Neuroprotective effects of chlorogenic acid in a mouse model of intracerebral hemorrhage associated with reduced extracellular matrix metalloproteinase inducer. *Biomolecules* 12 (8), 1020. doi:10. 3390/biom12081020

Lopez-Rodriguez, N. A., Gaytán-Martínez, M., de la Luz Reyes-Vega, M., and Loarca-Piña, G. (2020). Glucosinolates and isothiocyanates from *Moringa oleifera*: Chemical and biological approaches. *Plant Foods Hum. Nutr.* 75 (4), 447–57. doi:10.1007/s11130-020-00851-x

Lu, Y., Jiang, F., Jiang, H., Wu, K., Zheng, X., Cai, Y., et al. (2010). Gallic acid suppresses cell viability, proliferation, invasion and angiogenesis in human glioma cells. *Eur. J.Pharmacol.* 641 (2–3), 102–7. doi:10.1016/j.ejphar.2010.05.043

Lu, Y., Yeh, W., and Ohashi, P. S. (2008). LPS/TLR4 signal transduction pathway. Cytokine 42, 145–151. doi:10.1016/j.cyto.2008.01.006

Luetragoon, T., Sranujit, R. P., Noysang, C., Thongsri, Y., Potup, P., Suphrom, N., et al. (2020). Bioactive compounds in *Moringa oleifera* Lam. leaves inhibit the proinflammatory mediators in lipopolysaccharide-induced human monocyte-derived macrophages. *Molecules* 25 (1), 191–207. doi:10.3390/molecules25010191

Luiza, T., Oldoni, C., Mitterer-daltoe, M. L., Henrique, L., Pizone, I., and Lima, V. A. D. (2022). *Moringa oleifera* leaves from Brazil: Influence of seasonality, regrowth age and, region in biochemical markers and antioxidant potential. *Arab. J. Chem.* 15, 104206. doi:10.1016/j.arabjc.2022.104206

Lyketsos, C. G., Breitner, J. C. S., Green, R. C., Martin, B. K., Meinert, C., Piantadosi, S., et al. (2007). Naproxen and celecoxib do not prevent AD in early results from a randomized controlled trial. *Neurology* 68 (21), 1800–8. doi:10.1212/01.wnl. 0000260269.93245.d2

Mahmood, K. T., Mugal, T., and Ul Haq, I. (2010). Moringa oleifera: A natural gift-A review. J. Pharm. Sci. Res. 2 (11), 775-81.

Makita, C., Chimuka, L., Steenkamp, P., Cukrowska, E., and Madala, E. (2016). Comparative analyses of flavonoid content in *Moringa oleifera* and *Moringa ovalifolia* with the aid of UHPLC-qTOF-MS fingerprinting. S. Afr. J. Bot. 105, 116–122. doi:10.1016/j.sajb.2015.12.007

Maldini, M., Maksoud, S. A., Natella, F., Montoro, P., Petretto, G. L., Foddai, M., et al. (2014). *Moringa oleifera*: Study of phenolics and glucosinolates by mass spectrometry. *J. Mass Spectrom.* 49 (9), 900–10. doi:10.1002/jms.3437

Mallik, S. B., Mudgal, J., Nampoothiri, M., Hall, S., Dukie, S. A., Grant, G., et al. (2016). Caffeic acid attenuates lipopolysaccharide-induced sickness behaviour and neuroinflammation in mice. *Neurosci. Lett.* 632, 218–223. doi:10.1016/j.neulet.2016.08.044

Managa, L. R., du Toit, E. S., and Prinsloo, G. (2021). NMR-based metabolomic analysis to identify changes in the metabolite profile of three *M. oleifera* genotypes grown in an open hydroponic system. *Acta Hortic*. 1306, 35–42. doi:10.17660/ActaHortic.2021.1306.5

Maya, S., Prakash, T., and Das, K. (2018). Assessment of neuroprotective effects of Gallic acid against glutamate-induced neurotoxicity in primary rat cortex neuronal culture. *Neurochem. Int.* 121, 50–58. doi:10.1016/j.neuint.2018.10.011

Mirshekar, M. A., Sarkaki, A., Farbood, Y., Gharib, M. K., Badavi, M., Mansouri, M. T., et al. (2018). Neuroprotective effects of gallic acid in a rat model of traumatic brain injury: Behavioral, electrophysiological, and molecular studies. *Iran. J. Bas. Med. Sci.* 21 (12), 1056–1063. doi:10.22038/JJBMS.2018.29639.7165

Mishra, G., Singh, P., Verma, R., Kumar, S., Srivastav, S., Jha, K. K., et al. (2011). Traditional uses, phytochemistry and pharmacological properties of *Moringa oleifera plant*: An overview. *Der Pharm. Lett.* 3 (2), 141–64.

Moodley, I. (2017). Acute toxicity of *Moringa oleifera* leaf powder in rats. J. Med. Plants Stud. 5 (5), 180-5.

Mori, T., Koyama, N., Guillot-Sestier, M. V., Tan, J., and Town, T. (2013). Ferulic acid is a nutraceutical β -secretase modulator that improves behavioral impairment and Alzheimer-like pathology in transgenic mice. *PLoS ONE* 8 (2), e55774. doi:10.1371/journal.pone.0055774

Mori, T., Koyama, N., Tan, J., Segawa, T., Maeda, M., and Town, T. (2017). Combination therapy with octyl gallate and ferulic acid improves cognition and neurodegeneration in a transgenic mouse model of Alzheimer's disease. *J. Biol. Chem.* 292 (27), 11310–11325. doi:10.1074/jbc.M116.762658

Naveed, M., Hejazi, V., Abbas, M., Kamboh, A. A., Khan, G. J., Shumzaid, M., et al. (2018). Chlorogenic acid (cga): A pharmacological review and call for further research. *Biomed. Pharmacother.* 97, 67–74. doi:10.1016/j.biopha.2017.10.064

Oboh, G., Agunloye, O. M., Akinyemi, A. J., Ademiluyi, A. O., and Adefegha, S. A. (2013). Comparative study on the inhibitory effect of caffeic and chlorogenic acids on key enzymes linked to Alzheimer's disease and some pro-oxidant induced oxidative stress in rats' brain-*in vitro. Neurochem. Res.* 38 (2), 413–419. doi:10.1007/s11064-012-0935-6

Ojha, S., Javed, H., Azimullah, S., Khair, S. B. A., and Haque, M. E. (2015). Neuroprotective potential of ferulic acid in the rotenone model of Parkinson's disease. *Drug Des. devel. Ther.* 9, 5499–5510. doi:10.2147/DDDT.S90616

Okumu, M. O., Mbaria, J. M., Kanja, L. W., Gakuya, D. W., Kiama, S. G., Ochola, F. O., et al. (2016). Acute toxicity of the aqueous-methanolic *Moringa oleifera* (Lam) leaf extract on female Wistar albino rats. *Int. J. Basic Clin. Pharmacol.* 5 (5), 1856–1861. doi:10.18203/2319-2003.ijbcp20163153

Olayemi, A. T., Olanrewaju, M. J., and Oloruntoba, A. C. (2016). Toxicological evaluation of *Moringa oleifera* Lam seeds and leaves in Wistar rats. *Pharmacogn. Commun.* 6 (2), 100–11. doi:10.5530/pc.2016.2.8

Oldoni, T. L. C., Merlin, N., Karling, M., Carpes, S. T., Alencar, S. M. de, Morales, R. G. F., et al. (2019). Bioguided extraction of phenolic compounds and UHPLC-ESI-Q-TOF-MS/MS characterization of extracts of *Moringa oleifera* leaves collected in Brazil. *Food Res. Int.* 125, 108647. doi:10.1016/j.foodres.2019.108647

Padayachee, B., and Baijnath, H. (2012). An overview of the medicinal importance of. Moringaceae. J. Med. Plants Res. 6 (48), 5831–9. doi:10.5897/JMPR12.1187

Padayachee, B., and Baijnath, H. (2020). An updated comprehensive review of the medicinal, phytochemical and pharmacological properties of Moringa oleifera. *Moringa Oleifera. S. Afr. J. Bot.* 129, 304–16. doi:10.1016/j.sajb.2019.08.021

Pålsson-Mcdermott, E. M., and O' Neill, L. A. J. (2020). Targeting immunometabolism as an anti-inflammatory strategy. *Cell. Res.* 30 (4), 300–314. doi:10.1038/s41422-020-0291-z

Panwar, A., and Mathur, J. (2020). Genetic and biochemical variability among *Moringa* oleifera lam. Accessions collected from different agro-ecological zones. *Genome* 63 (3), 169–177. doi:10.1139/gen-2019-0102

Park, S. E., Sapkota, K., Kim, S., Kim, H., and Kim, S. J. (2011). Kaempferol acts through mitogen-activated protein kinases and protein kinase B/AKT to elicit protection in a model of neuroinflammation in BV2 microglial cells. *Br. J. Pharmacol.* 164 (3), 1008–1025. doi:10.1111/j.1476-5381.2011.01389.x

Pawate, S., Shen, Q., Fan, F., and Bhat, N. R. (2004). Redox regulation of glial inflammatory response to lipopolysaccharide and interferon γ . J. Neurosci. Res. 77 (4), 540–551. doi:10.1002/jnr.20180

Popoola, J. O., and Obembe, O. O. (2013). Local knowledge, use pattern and geographical distribution of *Moringa oleifera* Lam. (*Moringaceae*) in Nigeria. *J. Ethnopharmacol.* 150 (2), 682–91. doi:10.1016/j.jep.2013.09.043

Prabakaran, M., Kim, S. H., Sasireka, A., Chandrasekaran, M., and Chung, I. M. (2018). Polyphenol composition and antimicrobial activity of various solvent extracts from different plant parts of *Moringa oleifera*. *Food Biosci*. 26, 23–29. doi:10.1016/j.fbio. 2018.09.003

Prabu, S. L., Umamaheswari, A., and Puratchikody, A. (2019). Phytopharmacological potential of the natural gift *Moringa oleifera* Lam and its therapeutic application: An overview. *Asian pac. J. Trop. Med.* 12 (11), 485–98. doi:10.4103/1995-7645.271288

Prasad, J., Baitharu, I., Sharma, A. K., Dutta, R., Prasad, D., and Singh, S. B. (2013). Quercetin reverses hypobaric hypoxia-induced hippocampal neurodegeneration and improves memory function in the rat. *High. Alt. Med. Biol.* 14 (4), 383–394. doi:10. 1089/ham.2013.1014

Qadir, R., Anwar, F., Bashir, K., Tahir, M. H., Alhumade, H., and Mehmood, T. (2022). Variation in nutritional and antioxidant attributes of *Moringa oleifera* L. leaves at different maturity stages. *Front. Energy Res.* 10, 1–7. doi:10.3389/fenrg.2022.888355

Ramanan, V. K., and Saykin, A. J. (2013). Pathways to neurodegeneration: Mechanistic insights from GWAS in Alzheimer's disease, Parkinson's disease, and related disorders. *Am. J. Neuro Dis.* 2 (3), 145–175.

Ramiro-puig, E., Lee, H., Mcshea, A., Smith, M. A., and Perry, G. (2009). Neuroprotective effect of cocoa flavonoids on *in vitro* oxidative stress. *Eu. J. nutri.* 48 (1), 54–61. doi:10.1007/s00394-008-0761-4

Rani, N. Z. A., Husain, K., and Kumolosasi, E. (2018). *Moringa* genus: A review of phytochemistry and pharmacology. *Front. Pharmacol.* 9, 108–26. doi:10.3389/fphar.2018. 00108

Raviteja, S., Manandhar, S., Priya, K., Hari, G., and Pai, K. S. R. (2021). Protective role of caffeic acid in cognitive dysfunction and oxidative stress induced by colchicine in rats. *Indian J. Pharm. Educ. Res.* 55 (2), S457–S467. doi:10.5530/ijper.55.2s.117

Reddy, Y. R. R., Lokanatha, O., Ratnam, K. S. V. P., Reddy, C. S., Raju, I. N., and Reddy, C. D. (2013). Acute and sub acute toxicity of *Moringa oleifera* stem bark. *Int. J. Life Sci. Biotechnol. Pharma Res.* 2 (4), 73-82.

Riaz, A., Rasul, A., Hussain, G., Zahoor, M. K., Jabeen, F., Subhani, Z., et al. (2018). Astragalin: A bioactive phytochemical with potential therapeutic activities. *Adv. Pharmacol. Sci.* 2, 9794625. doi:10.1155/2018/9794625

Rock, R. B., Gekker, G., Hu, S., Sheng, W. S., Cheeran, M., Lokensgard, J. R., et al. (2004). Role of microglia in central nervous system infections. *Clin. Microbiol. Rev.* 17 (4), 942–64. doi:10.1128/CMR.17.4.942-964.2004

Ruckdeschel, K., Pfaffinger, G., Haase, R., Sing, A., Weighardt, H., Häcker, G., et al. (2004). Signaling of apoptosis through TLRs critically involves Toll/IL-1 receptor domain-containing adapter inducing IFN- β , but not MyD88, in bacteria-infected murine macrophages. *J. Immunol.* 173 (5), 3320–8. doi:10.4049/jimmunol.173.5.3320

Ruijters, E. J. B., Weseler, A. R., Kicken, C., Haenen, G. R. M. M., and Bast, A. (2013). The flavanol (-)-epicatechin and its metabolites protect against oxidative stress in primary endothelial cells via a direct antioxidant effect. *Eu. J. Pharmacol.* 715 (1–3), 147–153. doi:10.1016/j.ejphar.2013.05.029

Saini, R. K., Sivanesan, I., and Keum, Y. S. (2016). Phytochemicals of *Moringa oleifera*: A review of their nutritional, therapeutic and industrial significance. *3 Biotech*. 6 (2), 203–14. doi:10.1007/s13205-016-0526-3

Salau, V. F., Erukainure, O. L., Bharuth, V., and Islam, M. S. (2021). Caffeic acid improves glucose utilization and maintains tissue ultrastructural morphology while modulating metabolic activities implicated in neurodegenerative disorders in isolated rat brains. J. Biochem. Mol. Toxicol. 35 (1), e22610–11. doi:10.1002/jbt.22610

Sandhu, J. K., and Kulka, M. (2021). Decoding mast cell-microglia communication in neurodegenerative diseases. *Int. J. Mol.* 22 (3), 1093. doi:10.3390/ijms22031093

Saucedo-Pompa, S., Torres-Castillo, J. A., Castro-López, C., Rojas, R., Sánchez-Alejo, E. J., Ngangyo-Heya, M., et al. (2018). *Moringa* plants: Bioactive compounds and promising applications in food products. *Food Res. Int.* 111, 438–50. doi:10.1016/j.foodres.2018. 05.062

Sgarbossa, A., Giacomazza, D., and Di Carlo, M. (2015). Ferulic acid: A hope for alzheimer's disease therapy from plants. *Nutrients* 7 (7), 5764–5782. doi:10.3390/ nu7075246

Shabani, S., Rabiei, Z., and Amini-Khoei, H. (2020). Exploring the multifaceted neuroprotective actions of gallic acid: A review. *Int. J. Food Proper.* 23 (1), 736–752. doi:10.1080/10942912.2020.1753769

Shah, M. A., Kang, J. B., Park, D. J., Kim, M. O., and Koh, P. O. (2021). Chlorogenic acid alleviates neurobehavioral disorders and brain damage in focal ischemia animal models. *Neurosci. Lett.* 760, 136085. doi:10.1016/j.neulet.2021.136085

Shi, Y., Chen, X., Liu, J., Fan, X., Jin, Y., Gu, J., et al. (2021). Isoquercetin improves inflammatory response in rats following ischemic stroke. *Front. Neurosci.* 15, 555543–14. doi:10.3389/fnins.2021.555543

Silva, J., Pedro, J., Cirino, G., and Carvalho, P. D. O. (2021). The pharmacological action of kaempferol in central nervous system diseases: A review. *Front. Pharmacol.* 11 (565700), 565700–15. doi:10.3389/fphar.2020.565700

Simpson, D. S. A., and Oliver, P. L. (2020). Ros generation in microglia: Understanding oxidative stress and inflammation in neurodegenerative disease. *Antioxidants* 9 (8), 743–27. doi:10.3390/antiox9080743

Singh, B. N., Singh, B. R., Singh, R. L., Prakash, D., Dhakarey, R., Upadhyay, G., et al. (2009). Oxidative DNA damage protective activity, antioxidant and anti-quorum sensing potentials of *Moringa oleifera*. *Food Chem. Toxicol.* 47 (6), 1109–16. doi:10.1016/j.fct.2009. 01.034

Sobue, A., Komine, O., Hara, Y., Endo, F., Mizoguchi, H., Watanabe, S., et al. (2021). Microglial gene signature reveals loss of homeostatic microglia associated with neurodegeneration of Alzheimer's disease. *Acta Neuropathol. Commun.* 9 (1), 1–17. doi:10.1186/s40478-020-01099-x

Solleiro-Villavicencio, H., and Rivas-Arancibia, S. (2018). Effect of chronic oxidative stress on neuroinflammatory response mediated by CD4+T cells in neurodegenerative diseases. *Front. Cell. Neurosci.* 12 (4), 114–13. doi:10.3389/fncel.2018.00114

Stohs, S. J., and Hartman, M. J. (2015). Review of the safety and efficacy of Moringa oleifera. Phytother. Res. 804 (29), 796-804. doi:10.1002/ptr.5325

Stringer, T., Guerrieri, D., Vivar, C., and Praag, H. V. (2015). Plant-derived flavanol (–) epicatechin mitigates anxiety in association with elevated hippocampal monoamine and BDNF levels but does not influence pattern separation in mice. *Trans. Psychiat.* 5, e493. doi:10.1038/tp.2014.135

Subedi, L., Venkatesan, R., and Kim, S. Y. (2017). Neuroprotective and anti-inflammatory activities of allyl isothiocyanate through attenuation of JNK/NF- κ B/TNF- α signaling. *Int. J. Mol. Sci.* 18 (7), 1423–16. doi:10.3390/ijms18071423

Tan, S. H., Karri, V., Tay, N. W. R., Chang, K. H., Ah, H. Y., Ng, P. Q., et al. (2019). Emerging pathways to neurodegeneration: Dissecting the critical molecular mechanisms in Alzheimer's disease, Parkinson's disease. *Biomed. Pharmacother*. 111, 765–777. doi:10. 1016/j.biopha.2018.12.101

Sul, O. J., and Ra, S. W. (2021). Quercetin prevents lps-induced oxidative stress and inflammation by modulating NOX2/ROS/NF-KB in lung epithelial cells. *Molecules* 26 (22), 6949. doi:10.3390/molecules26226949

Thapa, K., Poudel, M., and Adhikari, P. (2019). *Moringa oleifera*: A review article on nutritional properties and its prospect in the context of Nepal. *Acta Sci. Agric.* 3 (11), 47–54. doi:10.31080/ASAG.2019.03.0683

Tran, H. T. T., Stetter, R., Herz, C., Spöttel, J., Krell, M., Hanschen, F. S., et al. (2021). Allyl isothiocyanate: A TAS2R38 receptor-dependent immune modulator at the interface between personalized medicine and nutrition. *Front. Immunol.* 12, 669005–13. doi:10. 3389/fmmu.2021.669005

Vergara-Jimenez, M., Almatrafi, M. M., and Fernandez, M. L. (2017). Bioactive components in *Moringa oleifera* leaves protect against chronic disease. *Antioxidants* 6 (4), 91–13. doi:10.3390/antiox6040091

Tumer, T. B., Rojas-silva, P., Poulev, A., Raskin, I., and Waterman, C. (2015). Direct and indirect antioxidant activity of polyphenol- and isothiocyanate-enriched fractions from Moringa oleifera. *J. Agric. Food Chem.* 63, 1505–1513. doi:10.1021/jf505014n

Vöglein, J., Kostova, I., Arzberger, T., Roeber, S., Schmitz, P., Simons, M., et al. (2021). First symptom guides diagnosis and prognosis in neurodegenerative diseases—A retrospective study of autopsy proven cases. *Eur. Neurol.* 28 (6), 1801–1811. doi:10. 1111/ene.14800

Wagner, A. E., Boesch-Saadatmandi, C., Dose, J., Schultheiss, G., and Rimbach, G. (2012). Anti-inflammatory potential of allyl-isothiocyanate - role of nrf2, nf-xB and microrna-155. J. Cell. Mol. Med. 16 (4), 836–843. doi:10.1111/j.1582-4934.2011.01367.x

Wasik, A., and Antkiewicz-Michaluk, L. (2017). The mechanism of neuroprotective action of natural compounds. *Pharmacol. Rep.* 69 (5), 851–860. doi:10.1016/j.pharep.2017. 03.018

Waterman, C., Cheng, D. M., Rojas-silva, P., Poulev, A., Dreifus, J., Lila, M. A., et al. (2014). Stable, water extractable isothiocyanates from *Moringa oleifera* leaves attenuate inflammation *in vitro*. *Phytochemistry* 23 (1), 114–122. doi:10.1016/j.phytochem.2014. 03.028

William, F., Lakshminarayanan, I. S., and Chegu, H. (1993). Effect of some Indian vegetables on the glucose and insulin response in diabetic subjects. J. Food Sci. Nutr. 44, 191–195. doi:10.3109/09637489309017439

Williams, R. J., and Spencer, J. P. E. (2012). Flavonoids, cognition, and dementia: Actions, mechanisms, and potential therapeutic utility for Alzheimer disease. *Free Rad. Biol. Med.* 52 (1), 35–45. doi:10.1016/j.freeradbiomed.2011.09.010

Wu, L., Zhang, Q., Zhang, X., Lv, C., Li, J., Yuan, Y., et al. (2012). Pharmacokinetics and blood – brain barrier penetration of (+)-catechin and (–)-epicatechin in rats by microdialysis sampling coupled to high-performance liquid chromatography with chemiluminescence detection. J. Agric. Food Chem. 60, 9377–9383. doi:10.1021/ jf301787f

Wyss-Coray, T., and Mucke, L. (2002). Inflammation in neurodegenerative disease — A double-edged sword. *Neuron* 35 (3), 419–32. doi:10.1016/s0896-6273(02)00794-8

Yen, G. C., Duh, P. Der, and Tsai, H. L. (2002). Antioxidant and pro-oxidant properties of ascorbic acid and gallic acid. *Food Chem.* 79 (3), 307–313. doi:10.1016/S0308-8146(02) 00145-0

Youdim, K. A., Qaiser, M. Z., Begley, D. J., Rice-Evans, C. A., and Abbott, N. J. (2014). Flavonoid permeability across an *in-situ* model of the blood-brain barrier. *Free Radic. Biol. Med.* 36 (5), 592–604. doi:10.1016/j.freeradbiomed.2003.11.023

Zahirah, N., Rani, A., Husain, K., and Kumolosasi, E. (2018). *Moringa* genus: A review of phytochemistry and pharmacology. *Front. Pharmacol.* 9 (108), 108–26. doi:10.3389/fphar.2018. 00108

Zeng, K., Li, Y., Yang, W., Ge, Y., Xu, L., Ren, T., et al. (2019). *Moringa oleifera* seed extract protects against brain damage in both the acute and delayed stages of ischemic stroke. *Exp. Gerontol.* 122, 99–108. doi:10.1016/j.exger.2019.04.014

Zhang, M., Hettiarachchy, N. S., Horax, R., Kannan, A., D, A. P. M., Muhundan, A., et al. (2011). Phytochemicals, antioxidant and antimicrobial activity of *Hibiscus sabdariffa*, Centella asiatica, Moringa oleifera and Murraya koenigii leaves. J. Med. Plants Res. 5 (30), 6672–6680. doi:10.5897/JMPR11.621

Zhang, Y., Wu, Q., Zhang, L., Wang, Q., Yang, Z., Liu, J., et al. (2019). Caffeic acid reduces A53T α -synuclein by activating JNK/Bcl-2-mediated autophagy *in vitro* and improves behaviour and protects dopaminergic neurons in a mouse model of Parkinson's disease. *Pharmacol. Res.* 150, 104538. doi:10.1016/j.phrs.2019.104538

Zheng, Y., Li, L., Chen, B., Fang, Y., Lin, W., Zhang, T., et al. (2022). Chlorogenic acid exerts neuroprotective effect against hypoxia-ischemia brain injury in neonatal rats by activating Sirt1 to regulate the Nrf2-NF-kB signaling pathway. *Cell. Commun. Signal.* 20 (1), 84–20. doi:10.1186/s12964-022-00860-0

Zhong, Q., Zou, Y., Liu, H., Chen, T., Zheng, F., Huang, Y., et al. (2020). Toll-like receptor 4 deficiency ameliorates β 2-microglobulin induced age-related cognition decline due to neuroinflammation in mice. *Mol. Brain.* 13 (1), 20–15. doi:10.1186/s13041-020-0559-8

Zhou, C., Lu, M., Cheng, J., Rohani, E. R., Hamezah, H. S., Han, R., et al. (2022). Review on the pharmacological properties of phillyrin. *Phillyrin. Mol.* 27 (12), 3670. doi:10.3390/molecules27123670

Zhou, J., Yang, W., Suo, D., Li, Y., Peng, L., Xu, L., et al. (2018). *Moringa oleifera* seed extract alleviates scopolamine-induced learning and memory impairment in mice. *Front. Pharmacol.* 9 (389), 389–11. doi:10.3389/fphar.2018.00389

Zhu, Y., Yin, Q., and Yang, Y. (2020). Comprehensive investigation of *Moringa oleifera* from different regions by simultaneous. *Molecules* 25 (1), 676. doi:10.3390/molecules25030676