

Inflammation Affects Liver Function and the Metabolism of Voriconazole to Voriconazole-N-Oxide in Adult and Elderly Patients

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OPEN ACCESS

Edited by:

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Reviewed by:

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Specialty section:

This article was submitted to Inflammation Pharmacology, a section of the journal Frontiers in Pharmacology

Received: 16 December 2021 Accepted: 22 March 2022 Published: 06 April 2022

Citation:

Liang Z, Yu M, Liu Z, Liu F, Jia C, Xiong L, Dai Q, Qin S, Cheng L and Sun F (2022) Inflammation Affects Liver Function and the Metabolism of Voriconazole to Voriconazole-N-Oxide in Adult and Elderly Patients. Front. Pharmacol. 13:835871. doi: 10.3389/fphar.2022.835871 **Background:** The inner association of inflammation with voriconazole (VCZ) metabolism has not been fully investigated. We intend to investigate the effects of inflammation on liver function, VCZ trough concentration (C_0), C_0 /dose ratio and the ratio of VCZ to VCZ-N-oxide concentration (C_0/C_N) in adult and elderly patients.

Methods: A single-center retrospective study was conducted among patients who were treated in our hospital between January 2018 and December 2021. For each eligible patient, demographic details, medical history, laboratory parameters, procalcitonin (PCT), C reactive protein (CRP), and interleukin-6 (IL-6) were collected from the medical chart. VCZ C_N , TNF- α , IL-1 β , IL-8, and IL-10 concentrations were detected in blood samples.

Results: A total of 356 patients were included in our study, with 195 patients in the adult cohort (<60 years) and 161 patients in the elderly cohort (≥60 years). In adult patients, CRP and IL-8 levels showed moderate association with VCZ C_0/C_N ratio (CRP: r = 0.512, p < 0.001; IL-8: r = 0.476, p = 0.002). IL-6 level shallowly associated with VCZ C_0/C_N ratio both in adult and elderly patients (r = 0.355, p = 0.003; r = 0.386, p = 0.001). A significantly higher VCZ C_0 , C_0 /dose ratio and C_0/C_N ratio was observed in adult patients with severe inflammation compared with patients with moderate inflammation and no to mild inflammation, as reflected by PCT levels (p < 0.05). However, there was no significant difference observed among different inflammation degrees in elderly patients. Lower albumin (AL) and higher total bilirubin (TBIL) were observed along with the degree of inflammation in both adult and elderly patients, as reflected by CRP and PCT levels (p < 0.05).

Conclusion: Inflammation may affect the metabolism of VCZ to VCZ-N-oxide both in adult and elderly patients, and decreased plasma AL levels and increased TBIL levels under inflammatory conditions may also alter VCZ metabolism.

Keywords: voriconazole, inflammation, voriconazole-N-oxide, invasive fungal infections, plasma concentration, therapeutic drug monitoring

Metabolism of Voriconazole to Voriconazole-N-Oxide

INTRODUCTION

Invasive fungal infections are common complications in immunosuppressed patients with high morbidity and mortality, such as patients with hematologic malignancy, solid organ transplant patients, and intensive care unit patients (Jenks et al., 2020). The incidence of invasive fungal infections in elderly patients is also obvious due to their decreased physiological defense function and immune function and complicated chronic diseases such as hypertension, diabetes, coronary atherosclerotic heart disease, etc. Voriconazole (VCZ) is a triazole antifungal agent widely used in patients with invasive Aspergillus or Candida infection (Patterson et al., 2016). It can be used as prophylactic or therapeutic. However, the therapy range of VCZ is narrow; in Europe, it is 1.0-5.5 µg/ml, and in China, it is 0.5-5.0 µg/ml (Chen et al., 2018; Ullmann et al., 2018). The metabolism of VCZ exhibits nonlinear pharmacokinetic characteristics highly individual variability with (Theuretzbacher et al., 2006). Many factors have been reported to contribute to the concentration variability of VCZ, including age, sex, drug-drug interactions, liver dysfunction, serum total bilirubin (TBIL), albumin (AL), cytochrome P450 (CYP) 2C19 gene polymorphism, and inflammation (Theuretzbacher et al., 2006; Bruggemann et al., 2009; Weiss et al., 2009; Veringa et al., 2017; Vreugdenhil et al., 2018; Xu et al., 2018; Shang et al., 2020).

Recently, much work has been done to explore the association of inflammation, as reflected by C reactive protein (CRP), interleukin-6 (IL-6), and IL-8, with VCZ concentration in transplantation patients, patients with blood diseases, and critically ill patients (van Wanrooy et al., 2014; Vreugdenhil et al., 2018; Gautier-Veyret et al., 2019). In

our previous study, we identified that procalcitonin (PCT) is an independent factor affecting the VCZ trough concentration (C_0) in elderly patients (Cheng et al., 2019). In the case of infection, immune cells release tumor necrosis factor (TNF- α), IL-1, IL-6, IL-8 and other signaling molecules, causing a systemic inflammatory response and stimulating different tissues (kidney, adipose tissue, lung, and liver) to secrete PCT, and their blood concentrations increase (Maruna et al., 2000). Although CRP, IL-6, and IL-8 were associated with VCZ C₀ (Vreugdenhil et al., 2018) and CRP was correlated with the metabolism of VCZ in hematological patients (major adults) (Veringa et al., 2017), the effect of IL-6 and IL-8 on VCZ metabolism in adult and elderly patients is still unknown.

The VCZ C_0 in the elderly cohort were significantly higher than those in the adult cohort at the same normal dosage, and the proportion of VCZ $C_0 > 5.0 \,\mu\text{g/ml}$ was 2.3 fold in the elderly cohort than that in the adult cohort (35.3% vs. 15.4%) (Cheng et al., 2019). In the previous study, we also found that for patients with CYP2C19 normal metaboliser, the VCZ C₀, C₀/dose ratio and the ratio of VCZ to VCZ-N-oxide concentration (C_0/C_N) were significantly higher in the elderly cohort than those in the adult cohort (Shang et al., 2020). The adverse effects of VCZ were the main cause of drug discontinuation, and thus lead to high mortality, which was associated with its trough concentration (Chan et al., 2020; Tang et al., 2020). Infection, chronic inflammation induced by chronic disease condition, and inflammation induced by the treatment of immunosuppressants were commonly existed both in adult and elderly patients with invasive fungal infections. In addition, liver function has been generally accepted to affect the concentration of VCZ, and the liver function of elderly patients is impaired, with decreased expression of albumin (AL) and CYP enzymes and

Variable	Adult	Elderly	p value	
	cohort (<i>n</i> = 195)	cohort (<i>n</i> = 161)		
Sex			0.110	
Male [no. (%)]	116 (59.5)	109 (67.7)		
Female [no. (%)]	79 (40.5)	52 (32.3)		
Age (yr)	42.4 ± 12.0	71.9 ± 8.4	0.000	
Weight (kg)	60.35 ± 12.05	57.84 ± 11.91	1.651	
Underlying diseases				
Leukemia [no. (%)]	77 (39.5)	12 (7.4)	0.000	
Hypertension [no. (%)]	41 (21.0)	65 (40.4)	0.000	
Diabetes mellitus [no. (%)]	24 (12.3)	49 (30.4)	0.000	
Kidney disease [no. (%)]	60 (30.8)	48 (29.8)	0.757	
Renal transplant [no. (%)]	8 (4.1)	0	0.025	
Liver cancer [no. (%)]	0	4 (2.5)	0.088	
Pneumonia [no. (%)]	98 (50.3)	95 (59.0)	0.099	
Coronary atherosclerotic heart disease [no. (%)]	1 (0.5)	96 (59.6)	0.000	
Fungus category				
Aspergillus [no. (%)]	22 (11.3)	20 (12.4)	0.740	
Saccharomycetes [no. (%)]	16 (8.2)	33 (20.5)	0.001	
Monilia [no. (%)]	31 (15.9)	31 (19.3)	0.406	
Others [no. (%)]	40 (20.5)	34 (21.1)	0.997	
Negative [no. (%)]	89 (45.6)	43 (26.7)	0.000	
Route of administration			0.362	
Intravenous [no. (%)]	142 (72.8)	114 (70.8)		
Oral [no. (%)]	15 (7.7)	8 (5.0)		
Mixed [no. (%)]	38 (19.5)	39 (24.2)		
Daily voriconazole dose (mg/kg)	7.34 ± 1.36	7.33 ± 1.52	0.944	

TABLE 2	Laboratory	parameters of	of p	atients ir	h the	two	cohorts.
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Parameter	п	Adult cohort	n	Elderly cohort	p value
VCZ C ₀ (µg/ml)	298	2.90 (1.50, 4.90)	255	3.60 (2.30, 5.65)	0.002
VCZ C ₀ /dose ratio (g/ml)	217	0.42 (0.22, 0.72)	143	0.53 (0.30, 0.81)	0.036
VCZ C ₀ /C _N ratio	154	1.42 (0.67, 3.63)	130	2.05 (1.11, 3.20)	0.029
Inflammation marker					
CRP (mg/L)	89	42.90 (8.96, 119.55)	41	64.07 (14.50, 131.00)	0.106
PCT (ng/ml)	172	0.29 (0.10, 0.84)	127	0.30 (0.11, 0.91)	0.395
TNF-α (pg/ml)	53	4.59 (3.29, 7.58)	38	27.17 (4.68, 60.36)	0.000
IL-6 (ng/L)	139	26.63 (7.47, 97.30)	119	33.87 (16.08, 75.54)	0.336
IL-8 (pg/ml)	73	17.69 (16.11, 42.80)	88	16.97 (13.79, 37.08)	0.279
IL-10 (pg/ml)	46	30.41 (19.05, 58.30)	41	62.99 (20.48, 116.82)	0.094
IL-1β (pg/ml)	38	4.85 (2.53, 8.25)	33	6.88 (3.76, 16.33)	0.122
Liver function					
ALT (U/L)	193	96.0 (74.0, 151.0)	167	114.0 (79.5, 176.0)	0.056
AST (U/L)	193	18.8 (9.6, 37.5)	167	22.4 (10.85, 41.4)	0.221
γ-GT (U/L)	193	24.2 (16.1, 50.6)	167	34.2 (22.9, 56.9)	0.000
ALP (U/L)	193	73.1 (34.9, 143.9)	167	86.0 (43.8, 174.35)	0.105
TBIL (µmol/L)	193	13.9 (9.2, 24.5)	167	13.3 (9.65, 25.9)	0.777
AL (g/L)	193	32.1 (29.1, 35.5)	167	32 (29.05, 36.55)	0.711

Data are presented as median (interquartile range). VCZ, voriconazole; CRP, C-reactive protein; PCT, procalcitonin; TNF, tumor necrosis factor; IL, interleukin; ALT, alanine aminotransferase; AST, aspartate aminotransferase; γ -GT, γ -glutamyl transferase; ALP, alkaline phosphatase; TBIL, total bilirubin; AL, albumin.

increased bilirubin concentrations (Schmucker, 2005; Dong et al., 2010), which may also affect VCZ metabolism.

VCZ metabolism is mainly mediated by the drug-metabolizing enzymes CYP2C19 and CYP3A4 in the liver, and the main metabolite in plasma is VCZ-N-oxide (accounting for 72%), with almost no activity (Hyland et al., 2003). To confirm the effect of inflammation on VCZ metabolism, the ratio of VCZ C_0/C_N might provide more information. The association of VCZ C_0/C_N with inflammation was not fully investigated in elderly patients. Combined with the impaired liver function and chronic inflammation of elderly patients, we intend to investigate the influence of inflammation on liver function and VCZ metabolism in elderly patients, and take the results of adults as control, so as to provide evidence of the rational use of VCZ in the elderly.

MATERIALS AND METHODS

Patients and Study Design

A single center retrospective study was conducted. The patients were treated at Southwest Hospital, Chongqing, China between January 2018 and December 2021. Patients who met the following inclusion criteria were included: 1) aged \geq 18 years; 2) hospitalized patient who had at least one steady-state VCZ C₀ and CRP, PCT, or IL-6 concentration and liver function measured on the same day; and 3) not concomitantly using a strong inhibitor or inducer of CYP isoenzymes.

Data Collection

For each eligible patient, the following data were collected from the medical chart: 1) demographic details, including age, sex, and body weight; 2) medical history, including underlying disease, type of invasive fungal infections, and VCZ treatment regimen; 3) laboratory parameters, such as the liver function indices, ALP, ALT, AST, γ -GT, TBIL, and AL; and 4) inflammation markers, including PCT, CRP, and IL-6, which were measured routinely.

VCZ C₀ and VCZ C_N Determination

The VCZ C₀ was defined as the concentration obtained after 3 days of VCZ therapy with a loading dose of 6 mg/kg by intravenous administration, and maintenance dose of 4 mg/kg by intravenous administration, or sequentially taking VCZ orally (200 mg). The VCZ C₀ and VCZ C_N were measured by liquid chromatography-tandem mass spectrometry (LC–MS/MS) as previously described (Shang et al., 2020).

Cytokine Concentration Determination

Only discarded blood samples for VCZ C₀ determination between May 2021 and November 2021 were used. A total of 161 of the VCZ C₀ values matched with samples that were available for cytokine determination. IL-1 β , IL-10, and TNF- α enzyme-linked immune sorbent assay (ELISA) kits were purchased from Jianglai Biological (Shanghai, China). IL-8 ELISA kits were purchased from Elabscience Biotechnology Co., Ltd. Plasma concentrations of the cytokines were measured according to the manufacturer's instructions.

Statistical Analysis

IBM SPSS 19.0 (IBM Corp., Armonk, NY, United States) was used to perform the analysis. Categorical data were compared with the chi-squared test or Fisher's exact test. To explore the possible association of inflammation with VCZ C_0 and liver function, VCZ C_0 , C_0 /dose ratio, C_0/C_N ratio, and liver function of patients with no to mild inflammation (CRP, \leq 40 mg/L; PCT, \leq 0.25 ng/ml), with moderate inflammation (CRP, 41–200 mg/L; PCT, 0.25–0.5 ng/ml), and with severe inflammation (CRP, \geq 200 mg/L; PCT, \geq 0.5 ng/ml) were compared using a Kruskal–Wallisone test (three groups) or a Mann–Whitney U test (two groups). Pearson's correlation test was performed to assess the association of inflammation, reflected by the above inflammatory markers, with VCZ C_0 , C_0 /dose ratio, and C_0/C_N ratio.



FIGURE 1 Association of CRP, PCT, IL-6, and IL-8 plasma levels with VCZ C_0 and VCZ C_0/C_N in adult patients. (**A**,**B**) The CRP levels were poorly associated with VCZ C_0 but moderately associated with VCZ C_0/C_N ratio. (**C**,**D**) The PCT levels were poorly associated with VCZ C_0 but not associated with VCZ C_0/C_N ratio. (**E**,**F**) The IL-6 levels were poorly associated with VCZ C_0 but moderately associated with VCZ C_0 but moderately associated with VCZ C_0/C_N ratio. (**G**,**H**) The IL-8 levels were poorly associated with VCZ C_0 but moderately associated with VCZ C_0/C_N ratio.

RESULTS

Patient Characteristics

A total of 356 patients were included in our study, with 195 patients in the adult cohort (<60 years) and 161 patients in the elderly cohort (\geq 60 years). The baseline patient characteristics are depicted in **Table 1**. The main baseline diseases in the elderly cohort were hypertension, diabetes mellitus, and coronary

atherosclerotic heart disease, while the main baseline diseases in the adult cohort were leukemia and kidney disease. The main infected fungi were *Monilia*, *Saccharomycetes*, and *Aspergillus*.

VCZ C₀, C₀/Dose Ratio and C₀/C_N Ratio

The VCZ C₀, C₀/dose ratio, and C₀/C_N ratio in the elderly cohort were significantly higher than those in the adult cohort (p < 0.05, **Table 2**).



FIGURE 2 Association of CRP, PCT, IL-6, and IL-8 plasma levels with VCZ C_0 and VCZ C_0/C_N ratio in elderly patients. (**A**,**B**) The CRP levels were moderately associated with VCZ C_0 but not associated with VCZ C_0/C_N ratio. (**C**,**D**) The PCT levels were poorly associated with VCZ C_0 but not associated with VCZ C_0/C_N ratio. (**C**,**D**) The PCT levels were poorly associated with VCZ C_0 but not associated with VCZ C_0/C_N ratio. (**E**,**F**) The IL-6 levels were poorly associated with VCZ C_0/C_N ratio. (**E**,**F**) The IL-6 levels were poorly associated with VCZ C_0/C_N ratio. (**E**,**F**) The IL-6 levels were poorly associated with VCZ C_0/C_N ratio. (**C**,**D**) the vert poorly associated with VCZ C_0/C_N ratio.

Determination of Levels of Cytokines, CRP, and PCT

An overview of all measured laboratory parameters can be found in **Table 2**. The plasma levels of TNF- α and γ -GT in the elderly cohort were significantly higher than those in the adult cohort (p < 0.001).

Correlations Between Inflammatory Markers and VCZ C_0

In adult patients, CRP and PCT levels showed poor association with VCZ C₀, as well as IL-6 and IL-8 level (**Figure 1**). In elderly patients, only CRP level showed shallow association with VCZ C₀ (CRP: r = 0.370, p = 0.017, 41 observations) (**Figure 2**).



Distribution of VCZ C₀/dose ratio. (E,F) Distribution of VCZ C₀/C_N ratio. $^{\#\#}p < 0.01$.

A significantly (p = 0.003) higher VCZ C₀ was observed in patients with severe inflammation [6.10 (IQR, 5.14–7.30) mg/L; n = 7] than in patients with moderate inflammation [3.50 (1.91–5.61) mg/L; n = 39] and no to mild inflammation [2.30 (1.25–4.10) mg/L; n = 43], as reflected by the CRP level in adult patients; there was a tendency in elderly patients (p = 0.06). These differences were shown in **Figure 3**.

A significantly (p < 0.001) higher VCZ C₀ was also observed in patients with severe inflammation [4.60 (IQR, 2.90–6.10) mg/L; n = 66] than in patients with moderate inflammation [2.70 (1.40–4.30) mg/L;

n = 28] and no to mild inflammation [2.49 (1.40–4.30) mg/L; n = 80], as reflected by the PCT level in adult patients; there was a tendency in elderly patients (p = 0.068). These differences were shown in **Figure 4**.

Correlations Between Inflammatory Markers and VCZ C₀/Dose Ratio

In adult patients, CRP and IL-6 levels showed shallow association with VCZ C₀/dose ratio (CRP: r = 0.326, p = 0.010, 61 observations; IL-6: r





= 0.370, p < 0.001, 86 observations). There was no association between inflammatory biomarkers and VCZ C_0 /dose ratio in elderly patients.

A significantly (p = 0.010) higher VCZ C₀/dose ratio was observed in patients with severe inflammation [0.76 (0.69–0.93) g/ml; n = 7] than in patients with moderate inflammation [0.47 (0.29–0.76) g/ml; n = 25] and no to mild inflammation [0.31 (IQR, 0.17–0.53) g/ml; n = 29], as reflected by the CRP level in adult patients. There was no significant difference observed among different inflammation degrees in elderly patients (**Figure 3**).

A significantly (p = 0.001) higher VCZ C₀/dose ratio was also observed in patients with severe inflammation [0.63 (IQR, 0.39–0.97)

g/ml; n = 46] than in patients with moderate inflammation [0.47 (0.25–0.71) g/ml; n = 23] and no to mild inflammation [0.35 (0.19–0.58) g/ml; n = 62], as reflected by the PCT level in adult patients. There was no significant difference observed among different inflammation degrees in elderly patients (**Figure 4**).

Correlations Between Inflammatory Markers and VCZ C₀/C_N Ratio

In adult patients, CRP and IL-8 levels showed moderate association with VCZ C_0/C_N ratio (CRP: r = 0.512, *p* < 0.001, 45 observations; IL-

Cohort	Liver function	CRP (mg/ml)				
		0–40	40–200	200-		
Adult cohort	ALP (U/L)	93.0 (78.3, 128.0)	99.5 (74.3, 153.8)	79.5 (51.0, 87.8)	0.173	
	ALT (U/L)	19.9 (15.5, 42.1)	18.7 (8.2, 43.9)	11.0 (8.4, 13.6)	0.157	
	AST (U/L)	22.4 (19.8, 43.2)	22.3 (16.7, 51.8)	16.7 (13.4, 21.3)	0.112	
	γ-GT (U/L)	79.3 (55.1, 183.9)	69.2 (35.0, 112.5)	41.6 (28.3, 76.5)	0.255	
	TBIL (µmol/L)	12.0 (9.3, 19.2)	15.5 (9.0, 22.3)	22.6 (13.1, 30.0)	0.441	
	AL (g/L)	34.8 (30.4, 37.5)	31.7 (29.5, 33.1)	30.4 (27.1, 34.7)	0.034	
Elderly cohort	ALP (U/L)	80.0 (57.0, 110.0)	97.0 (71.0, 125.0)	_	0.302	
	ALT (U/L)	15.9 (12.6, 38.2)	28.0 (14.4, 48.6)	_	0.591	
	AST (U/L)	25.8 (23.7, 57.9)	34.2 (20.9, 68.2)	_	0.983	
	γ-GT (U/L)	82.0 (31.6, 153.4)	50.0 (42.9, 114.1)	_	0.561	
	TBIL (µmol/L)	9.9 (9.1, 13.9)	13.1 (10.6, 16.8)	_	0.149	
	AL (g/L)	32.7 (30.8, 33.6)	28.0 (25.0, 32.1)	_	0.017	

TABLE 3 | Comparisons of liver function under different inflammation degrees as reflected by CRP levels in the two cohorts.

"-", not available; data are presented as median (interquartile range). ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; γ -GT, γ -glutamyl transferase; TBIL, total bilirubin; AL, albumin.

8: r = 0.476, p = 0.002, 40 observations). PCT and IL-6 had shallow association with VCZ C₀/C_N ratio (PCT: r = 0.329, p = 0.004, 75 observations; IL-6: r = 0.355, p = 0.003, 67 observations) (**Figure 1**). In elderly patients, only IL-6 had shallow association with VCZ C₀/C_N ratio (IL-6: r = 0.386, p = 0.001, 67 observations) (**Figure 2**).

In adult patients, a significantly (p = 0.025) higher VCZ C₀/C_N ratio was observed in patients with severe inflammation [4.00 (IQR, 2.80–6.90) mg/L; n = 25] than in patients with moderate inflammation [3.22 (1.29–4.70) mg/L; n = 10] and no to mild inflammation [3.27 (2.30–5.34) mg/L; n = 39], as reflected by the PCT level; however, a tendency of significant association was observed, as reflected by the CRP level (p = 0.074). These differences were shown in **Figure 4**. In elderly patients, there was no significant difference in C₀/C_N ratio under different inflammation degrees, as reflected by CRP or PCT levels.

Liver Function Results

A significantly lower AL was observed in patients with severe inflammation than in patients with moderate inflammation and no to mild inflammation among adult patients, both reflected by CRP and PCT levels (p < 0.05), and the association was obvious in elderly patients, as reflected by CRP levels (p = 0.001) (**Tables 3**, **4**; **Figure 5**). A significantly higher TBIL was observed in patients with severe inflammation than in patients with moderate inflammation and no to mild inflammation in both adult and elderly cohorts, as reflected by the PCT level (p < 0.01) (**Table 4**; **Figure 6**).

DISCUSSION

In the present study, we examined common inflammation markers, such as CRP, PCT, and IL-6, which were detected in the clinic, and four other cytokines, both pro- and anti-inflammatory, for their correlation with VCZ C_0 , C_0 /dose ratio and VCZ C_0/C_N ratio. CRP has been mostly investigated to be associated with VCZ C_0 in hematologic malignancy and solid organ transplant patients (42–62 years) (van Wanrooy et al., 2014; Encalada Ventura et al.,

TABLE 4 | Comparisons of liver function under different inflammation degrees as reflected by PCT levels in the two cohorts.

Cohort	Liver function	PCT (ng/ml)			
		0-0.25	0.25–0.5	0.5-	
Adult cohort	ALP (U/L)	88.2 (72.5, 125.0)	93.0 (74.0, 153.0)	92.0 (68.0, 165.0)	0.616
	ALT (U/L)	22.8 (12.6, 42.4)	13.3 (8.1, 30.3)	13.1 (8.1, 35.8)	0.080
	AST (U/L)	21.9 (17.2, 36.9)	19.6 (14.3, 49.7)	21 (14.3, 71.0)	0.686
	γ-GT (U/L)	78.6 (34.7, 187.5)	75.2 (46.2, 158)	67.1 (31.9, 128.3)	0.829
	TBIL (µmol/L)	11.0 (8.1, 17.6)	13.1 (7.7, 19.3)	21.2 (12.1, 34.6)	0.001
	AL (g/L)	34.4 (29.8, 37.0)	30.6 (27.8, 33.3)	30.7 (27.6, 32.3)	0.001
Elderly cohort	ALP (U/L)	103.0 (74.0, 124.8)	156.0 (65.5, 184.8)	129.0 (82.0, 227.0)	0.140
	ALT (U/L)	22.0 (11.2, 34.8)	15.5 (10.0, 24.4)	24.6 (8.5, 72.9)	0.513
	AST (U/L)	30.6 (19.8, 46.1)	25.1 (22.7, 35.5)	38.6 (25.8, 80.6)	0.084
	γ-GT (U/L)	52.6 (33.8, 124.5)	146.0 (50.7, 188.5)	67.0 (42.5, 168.5)	0.218
	TBIL (µmol/L)	10.8 (8.3, 13.4)	15.8 (11.5, 23.8)	20.9 (13.4, 68.1)	0.000
	AL (g/L)	30.6 (27.8, 34.2)	30.9 (28.6, 32.2)	29.6 (26.3, 33.2)	0.712

Data are presented as median (interquartile range). ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; y-GT, y-glutamyl transferase; TBIL, total bilirubin; AL, albumin.



of AL according to PCT. $p^{*} < 0.05$, $p^{*} < 0.01$.

2016; Vreugdenhil et al., 2018), and CRP levels are the main risk factor for VCZ overdose in French hematological patients (Gautier-Veyret et al., 2019).

In a prospective single-centre analysis performed in adult haematology patients receiving VCZ, which included 22 patients and 143 samples, the CRP, IL-6, and IL-8 levels were significantly associated with the VCZ C_0 (CRP: r = 0.53, p < 0.0001; IL-6: r = 0.53, p < 0.0001; IL-8: r = 0.42, p < 0.0001) (Vreugdenhil et al., 2018). In a retrospective chart review of VCZ, a total of 77 samples from 63 patients were obtained, and the CRP level was significantly associated with the VCZ C_0 /dose ratio ($r^2 =$ 0.26, *p* < 0.001) (Dote et al., 2016). In our study, 195 patients with 298 samples in the adult cohort and 161 patients with 255 samples in the elderly cohort were included, the correlation coefficient of CRP with VCZ C₀ was 0.289 and 0.370 in adult and elderly patients, respectively. The low correlation coefficient may be due to 1-2 samples from one patient in our study, while 6-7 samples from one patient in the prospective study. The retrospective nature of the study and the large number of patients may contribute to the increased sample distribution widths.

Niioka et al., 2017 investigated the relationship of the CYP2C19 phenotype with the VCZ C_0/C_N ratio and demographic and clinical characteristics of Japanese patients, and found that patients with intravenous administration, higher CRP on the same day of VCZ plasma concentration measurement, CYP2C19 extensive metabolizer, and old age had higher VCZ C_0/C_N ratio. In our study, only 7.7% (adult) and 5.0% (elderly) patients had oral administration; the median CRP level was 42.90 in adult patients and 64.07 in elderly patients, respectively; the median value of C_0/C_N ratio was 1.42 in adult patients and 2.05 in elderly patients, and the VCZ C_0/C_N ratio in elderly patients was significantly higher than that in adult patients, which were consistent with previous study.

CRP concentrations was reported to significantly influence the VCZ C_0/C_N ratio even when corrected for other factors that could influence VCZ metabolism in a prospective observational study (Veringa et al., 2017). In addition to CRP level being significantly



associated with VCZ C_0/C_N ratio in adult patients, our results also showed that IL-8 plasma levels were moderately associated with VCZ C_0/C_N ratio in adult patients, and IL-6 plasma levels were shallowly associated with VCZ C0/CN ratio in adult and elderly patients, which has not been reported previously. In an inflammatory state, LPS, TNF-a, IL-1β, and IL-6 can bind to cytokine receptors and TLR4 receptors on the cell membrane, regulating the expression of transporters and drug-related metabolic enzymes by targeting nuclear receptors, liver X receptor (LXR), pregnane X receptor (PXR) and constitutive rostane receptor (CAR), through the NF-KB signaling pathway (Wu and Lin, 2019). Furthermore, IL-6 can directly target liver cells and down regulate CYP2C19 and CYP3A4 gene expression during inflammation (Li et al., 2014; Klein et al., 2015). In particular, IL-6 caused a 30%-50% reduction in CYP2C19 mRNA expression in human hepatocytes (Aitken and Morgan, 2007; Rieger et al., 2015). The mechanism by which IL-8 influences the metabolism of VCZ is not clear. In the early stage of liver injury, IL-8 recruits neutrophils to the injured tissue through its specific receptors CXCR1 and CXCR2 and adheres to liver cells through intracellular adhesion factor 1, releasing myeloperoxidase and reactive oxygen species (ROS), resulting in hepatocyte damage (Heymann and Tacke, 2016; Khanam et al., 2017; Li et al., 2017). This may explain the positive correlation between the IL-8 level and VCZ C₀/C_N ratio.

CRP, PCT, and IL-6 were routine inflammatory biomarkers detected in clinic. In the current study, we found that CRP and IL-8 levels were associated with VCZ C_0/C_N ratio in adult patients, while only IL-6 level was shallowly associated with VCZ C_0/C_N ratio in elderly patients, which indicated that IL-6 would be a better marker when evaluating the metabolism of VCZ in the elderly.

VCZ is bound to AL. Decreased AL will increase unbound fraction of VCZ. A positive relationship between VCZ plasma protein binding (PPB) and plasma AL concentrations was observed, indicating higher unbound VCZ concentrations with decreasing AL concentrations (Vanstraelen et al., 2014). Decreased AL levels were also associated with a significantly increased VCZ C₀/dose ratio (Dote et al., 2016). In patients with invasive aspergillosis, serum AL and y-GT levels were significantly correlated with the VCZ clearance rate (Chantharit et al., 2020). Association of decreased AL and increased VCZ C₀ is related to reflection of impaired liver function (changes in intrinsic clearance). In our previous study, we determined that decreased AL was significantly associated with increased VCZ C₀ in adult patients (Cheng et al., 2019). In the present study, we found that, in both adult and elderly patients, a significantly lower AL was observed in patients with severe inflammation than in patients with moderate inflammation and no to mild inflammation, as reflected by CRP levels, which indicated that inflammation induced AL decreasing also contributed to VCZ metabolism.

Apart from the plasma AL concentration, the TBIL plasma concentration also significantly influences VCZ PPB (Vanstraelen et al., 2014). The presence of a bilirubin value of >1.5 mg/dL was associated with supratherapeutic concentrations in critically ill patients (Ruiz et al., 2019). In invasive fungal infection patients with liver dysfunction, VCZ clearance was significantly associated with TBIL and platelet count (Tang et al., 2020). Our previous study also determined that

increased TBIL was significantly associated with increased VCZ C_0 in adult patients (Cheng et al., 2019). The results of the current study showed that, in both adult and elderly patients, significantly increased TBIL was observed in patients along with the severity of inflammation, as reflected by PCT levels. The decreased AL level and increased TBIL level along with the degree of inflammation provided another explanation for the increased VCZ C_0 under the inflammatory state.

This study has several limitations. We do not determine VCZ C_N in feces and urine specimens. A ratio of <2% VCZ is excreted through renal as the parent drug (Roffey et al., 2003; Geist et al., 2013; Hohmann et al., 2017). Geist et al. found that VCZ N-oxide and its conjugates excreted in urine within 12 h post dose during steady-state only accounted for 1% of the dose (Geist et al., 2013). In the current study, we also determine the C₀ and C_N of VCZ at 12 h post drug administration during steady-state, and we propose that the C_N in feces and urine specimens may be rare. We previously demonstrated that VCZ C₀ and C₀/C_N ratio were influenced by CYP2C19 polymorphisms in adult patients, but the influence was not obvious in elderly patients (Shang et al., 2020). However, the adult cohort was not stratified by CYP2C19 polymorphism status in the current analysis for its retrospective nature. Some other liver biomarkers such as ALP, ALT, AST, and y-GT have been investigated to be associated with VCZ C₀, changes of these biomarkers reflect the function of liver, thus affecting VCZ metabolism. In the current study, we focus on the effect of inflammation on liver function and VCZ metabolism, and find that ALP, ALT, AST, and y-GT are not significantly influenced by inflammation.

In conclusion, this study clearly demonstrates that inflammation, reflected by IL-6 and IL-8, may affect the metabolism of VCZ to VCZ-N-oxide in both adult and elderly patients. Furthermore, the decreased plasma AL levels and increased plasma TBIL levels under inflammatory conditions

REFERENCES

- Aitken, A. E., and Morgan, E. T. (2007). Gene-specific Effects of Inflammatory Cytokines on Cytochrome P450 2C, 2B6 and 3A4 mRNA Levels in Human Hepatocytes. *Drug Metab. Dispos* 35 (9), 1687–1693. doi:10.1124/dmd.107. 015511
- Brüggemann, R. J., Alffenaar, J. W., Blijlevens, N. M., Billaud, E. M., Kosterink, J. G., Verweij, P. E., et al. (2009). Clinical Relevance of the Pharmacokinetic Interactions of Azole Antifungal Drugs with Other Coadministered Agents. *Clin. Infect. Dis.* 48 (10), 1441–1458. doi:10.1086/598327
- Chan, S. Y., Hughes, R. M., Woo, K., Perales, M. A., Neofytos, D., and Papanicolaou, G. (2020). Reasons for Voriconazole Prophylaxis Discontinuation in Allogeneic Hematopoietic Cell Transplant Recipients: A Real-Life Paradigm. *Med. Mycol.* 58 (8), 1029–1036. doi:10.1093/mmy/ myaa008
- Chantharit, P., Tantasawat, M., Kasai, H., and Tanigawara, Y. (2020). Population Pharmacokinetics of Voriconazole in Patients with Invasive Aspergillosis: Serum Albumin Level as a Novel Marker for Clearance and Dosage Optimization. *Ther. Drug Monit.* 42 (6), 872–879. doi:10.1097/FTD. 0000000000000799
- Chen, K., Zhang, X., Ke, X., Du, G., Yang, K., and Zhai, S. (2018). Individualized Medication of Voriconazole: A Practice Guideline of the Division of Therapeutic Drug Monitoring, Chinese Pharmacological Society. *Ther. Drug Monit.* 40 (6), 663–674. doi:10.1097/FTD.00000000000561

may also alter VCZ metabolism. The inflammation state and impaired liver function should be considered when optimizing VCZ doses to decrease the risk of adverse effects in elderly.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding authors.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Ethics Committee of the First Affiliated Hospital of Army Medical University. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

LC and FS designed the study, performed the data analysis, and drafted the manuscript; ZL searched the data and did the detection; MY, ZL, QD and SQ did the detection; FL, CJ and LX searched the data. All authors approved the final version of the manuscript.

FUNDING

This work was supported by the Fund for the Medical Research Project of Science and Health of Chongqing, China (2021MSXM218).

- Cheng, L., Xiang, R., Liu, F., Li, Y., Chen, H., Yao, P., et al. (2019). Therapeutic Drug Monitoring and Safety of Voriconazole in Elderly Patients. *Int. Immunopharmacol* 78, 106078. doi:10.1016/j.intimp.2019.106078
- Dong, M. H., Bettencourt, R., Barrett-Connor, E., and Loomba, R. (2010). Alanine Aminotransferase Decreases with Age: the Rancho Bernardo Study. *PLoS One* 5 (12), e14254. doi:10.1371/journal.pone.0014254
- Dote, S., Sawai, M., Nozaki, A., Naruhashi, K., Kobayashi, Y., and Nakanishi, H. (2016). A Retrospective Analysis of Patient-specific Factors on Voriconazole Clearance. J. Pharm. Health Care Sci. 2, 10. doi:10.1186/s40780-016-0044-9
- Encalada Ventura, M. A., van Wanrooy, M. J., Span, L. F., Rodgers, M. G., van den Heuvel, E. R., Uges, D. R., et al. (2016). Longitudinal Analysis of the Effect of Inflammation on Voriconazole Trough Concentrations. *Antimicrob. Agents Chemother*. 60 (5), 2727–2731. doi:10.1128/AAC.02830-15
- Gautier-Veyret, E., Truffot, A., Bailly, S., Fonrose, X., Thiebaut-Bertrand, A., Tonini, J., et al. (2019). Inflammation Is a Potential Risk Factor of Voriconazole Overdose in Hematological Patients. *Fundam. Clin. Pharmacol.* 33 (2), 232–238. doi:10.1111/fcp. 12422
- Geist, M. J., Egerer, G., Burhenne, J., Riedel, K. D., Weiss, J., and Mikus, G. (2013). Steady-state Pharmacokinetics and Metabolism of Voriconazole in Patients. J. Antimicrob. Chemother. 68 (11), 2592–2599. doi:10.1093/jac/dkt229
- Heymann, F., and Tacke, F. (2016). Immunology in the Liver-Ffrom Homeostasis to Disease. Nat. Rev. Gastroenterol. Hepatol. 13 (2), 88–110. doi:10.1038/nrgastro. 2015.200
- Hohmann, N., Kreuter, R., Blank, A., Weiss, J., Burhenne, J., Haefeli, W. E., et al. (2017). Autoinhibitory Properties of the Parent but Not of the N-Oxide Metabolite

Contribute to Infusion Rate-dependent Voriconazole Pharmacokinetics. Br. J. Clin. Pharmacol. 83 (9), 1954–1965. doi:10.1111/bcp.13297

- Hyland, R., Jones, B. C., and Smith, D. A. (2003). Identification of the Cytochrome P450 Enzymes Involved in the N-Oxidation of Voriconazole. *Drug Metab. Dispos* 31 (5), 540–547. doi:10.1124/dmd.31.5.540
- Jenks, J. D., Cornely, O. A., Chen, S. C., Thompson, G. R., 3rd, and Hoenigl, M. (2020). Breakthrough Invasive Fungal Infections: Who Is at Risk? *Mycoses* 63 (10), 1021–1032. doi:10.1111/myc.13148
- Khanam, A., Trehanpati, N., Riese, P., Rastogi, A., Guzman, C. A., and Sarin, S. K. (2017). Blockade of Neutrophil's Chemokine Receptors CXCR1/2 Abrogate Liver Damage in Acute-On-Chronic Liver Failure. *Front. Immunol.* 8, 464. doi:10.3389/ fimmu.2017.00464
- Klein, M., Thomas, M., Hofmann, U., Seehofer, D., Damm, G., and Zanger, U. M. (2015). A Systematic Comparison of the Impact of Inflammatory Signaling on Absorption, Distribution, Metabolism, and Excretion Gene Expression and Activity in Primary Human Hepatocytes and HepaRG Cells. *Drug Metab. Dispos* 43 (2), 273–283. doi:10.1124/dmd.114.060962
- Li, A. P., Yang, Q., Vermet, H., Raoust, N., Klieber, S., and Fabre, G. (2014). Evaluation of Human Hepatocytes under Prolonged Culture in a Novel Medium for the Maintenance of Hepatic Differentiation: Results with the Model Pro-inflammatory Cytokine Interleukin 6. Drug Metab. Lett. 8 (1), 12–18. doi:10.2174/ 187231280801140929155351
- Li, J. J., Ma, F. X., Wang, Y. W., Chen, F., Lu, S. H., Chi, Y., et al. (2017). Knockdown of IL-8 Provoked Premature Senescence of Placenta-Derived Mesenchymal Stem Cells. *Stem Cell Dev* 26 (12), 912–931. doi:10.1089/scd.2016.0324
- Maruna, P., Nedelníková, K., and Gürlich, R. (2000). Physiology and Genetics of Procalcitonin. *Physiol. Res.* 49 Suppl 1 (Suppl. 1), S57–S61.
- Niioka, T., Fujishima, N., Abumiya, M., Yamashita, T., Ubukawa, K., Nara, M., et al. (2017). Relationship between the CYP2C19 Phenotype Using the Voriconazole-To-Voriconazole N-Oxide Plasma Concentration Ratio and Demographic and Clinical Characteristics of Japanese Patients with Different CYP2C19 Genotypes. *Ther. Drug Monit.* 39 (5), 514–521. doi:10. 1097/FTD.0000000000000441
- Patterson, T. F., Thompson, G. R., 3rd, Denning, D. W., Fishman, J. A., Hadley, S., Herbrecht, R., et al. (2016). Executive Summary: Practice Guidelines for the Diagnosis and Management of Aspergillosis: 2016 Update by the Infectious Diseases Society of America. *Clin. Infect. Dis.* 63 (4), 433–442. doi:10.1093/cid/ciw326
- Rieger, J. K., Reutter, S., Hofmann, U., Schwab, M., and Zanger, U. M. (2015). Inflammation-associated microRNA-130b Down-Regulates Cytochrome P450 Activities and Directly Targets CYP2C9. *Drug Metab. Dispos* 43 (6), 884–888. doi:10.1124/dmd.114.062844
- Roffey, S. J., Cole, S., Comby, P., Gibson, D., Jezequel, S. G., Nedderman, A. N., et al. (2003). The Disposition of Voriconazole in Mouse, Rat, Rabbit, guinea Pig, Dog, and Human. *Drug Metab. Dispos* 31 (6), 731–741. doi:10.1124/dmd. 31.6.731
- Ruiz, J., Gordon, M., Villarreal, E., Peruccioni, M., Marqués, M. R., Poveda-Andrés, J. L., et al. (2019). Impact of Voriconazole Plasma Concentrations on Treatment Response in Critically Ill Patients. J. Clin. Pharm. Ther. 44 (4), 572–578. doi:10. 1111/jcpt.12817
- Schmucker, D. L. (2005). Age-related Changes in Liver Structure and Function: Implications for Disease ? *Exp. Gerontol.* 40 (8-9), 650–659. doi:10.1016/j.exger. 2005.06.009
- Shang, S., Cheng, L., Li, X., Xiang, R., Yu, M., Xiong, L., et al. (2020). Effect ofCYP2C19polymorphism on the Plasma Voriconazole Concentration and voriconazole-to-voriconazole-N-oxide Concentration Ratio in Elderly Patients. *Mycoses* 63, 1181–1190. doi:10.1111/myc.13105

- Tang, D., Yan, M., Song, B. L., Zhao, Y. C., Xiao, Y. W., Wang, F., et al. (2020).
 Population Pharmacokinetics, Safety and Dosing Optimization of Voriconazole in Patients with Liver Dysfunction: A Prospective Observational Study. Br. J. Clin. Pharmacol. 87 (4), 1890–1902. doi:10.1111/bcp.14578
- Theuretzbacher, U., Ihle, F., and Derendorf, H. (2006). Pharmacokinetic/ pharmacodynamic Profile of Voriconazole. *Clin. Pharmacokinet.* 45 (7), 649–663. doi:10.2165/00003088-200645070-00002
- Ullmann, A. J., Aguado, J. M., Arikan-Akdagli, S., Denning, D. W., Groll, A. H., Lagrou, K., et al. (2018). Diagnosis and Management of Aspergillus Diseases: Executive Summary of the 2017 ESCMID-ECMM-ERS Guideline. *Clin. Microbiol. Infect.* 24 (Suppl. 1), e1–e38. doi:10.1016/j.cmi.2018.01.002
- van Wanrooy, M. J., Span, L. F., Rodgers, M. G., van den Heuvel, E. R., Uges, D. R., van der Werf, T. S., et al. (2014). Inflammation Is Associated with Voriconazole Trough Concentrations. Antimicrob. Agents Chemother. 58 (12), 7098–7101. doi:10.1128/AAC.03820-14
- Vanstraelen, K., Wauters, J., Vercammen, I., de Loor, H., Maertens, J., Lagrou, K., et al. (2014). Impact of Hypoalbuminemia on Voriconazole Pharmacokinetics in Critically Ill Adult Patients. *Antimicrob. Agents Chemother.* 58 (11), 6782–6789. doi:10.1128/AAC.03641-14
- Veringa, A., Ter Avest, M., Span, L. F., van den Heuvel, E. R., Touw, D. J., Zijlstra, J. G., et al. (2017). Voriconazole Metabolism Is Influenced by Severe Inflammation: a Prospective Study. J. Antimicrob. Chemother. 72 (1), 261–267. doi:10.1093/jac/dkw349
- Vreugdenhil, B., van der Velden, W. J. F. M., Feuth, T., Kox, M., Pickkers, P., van de Veerdonk, F. L., et al. (2018). Moderate Correlation between Systemic IL-6 Responses and CRP with Trough Concentrations of Voriconazole. Br. J. Clin. Pharmacol. 84 (9), 1980–1988. doi:10.1111/bcp.13627
- Weiss, J., Ten Hoevel, M. M., Burhenne, J., Walter-Sack, I., Hoffmann, M. M., Rengelshausen, J., et al. (2009). CYP2C19 Genotype Is a Major Factor Contributing to the Highly Variable Pharmacokinetics of Voriconazole. J. Clin. Pharmacol. 49 (2), 196–204. doi:10.1177/0091270008327537
- Wu, K. C., and Lin, C. J. (2019). The Regulation of Drug-Metabolizing Enzymes and Membrane Transporters by Inflammation: Evidences in Inflammatory Diseases and Age-Related Disorders. J. Food Drug Anal. 27 (1), 48–59. doi:10. 1016/j.jfda.2018.11.005
- Xu, R. A., Gu, E. M., Liu, T. H., Ou-yang, Q. G., Hu, G. X., and Cai, J. P. (2018). The Effects of Cytochrome P450 2C19 Polymorphism on the Metabolism of Voriconazole In Vitro. Infect. Drug Resist. 11, 2129–2135. doi:10.2147/idr.s179078

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