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Traditional Chinese medicine for treatment of sepsis and related multi-organ injury

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Sepsis is a common but critical illness in patients admitted to the intensive care unit and is associated with high mortality. Although there are many treatments for sepsis, specific and effective therapies are still lacking. For over 2,000 years, traditional Chinese medicine (TCM) has played a vital role in the treatment of infectious diseases in Eastern countries. Both anecdotal and scientific evidence show that diverse TCM preparations alleviate organ dysfunction caused by sepsis by inhibiting the inflammatory response, reducing oxidative stress, boosting immunity, and maintaining cellular homeostasis. This review reports on the efficacy and mechanism of action of various TCM compounds, herbal monomer extracts, and acupuncture, on the treatment of sepsis and related multi-organ injury. We hope that this information would be helpful to better understand the theoretical basis and empirical support for TCM in the treatment of sepsis.

KEYWORDS

sepsis, inflammation, traditional Chinese medicine compounds, herbal monomer extracts, acupuncture

1 Introduction

Sepsis is one of the leading causes of death from critical diseases, affecting each year more than 30 million people worldwide (Fleischmann et al., 2016). Sepsis is defined as a life-threatening organ dysfunction caused by the host's dysfunctional response to infection, and is diagnosed based on a Sequential [Sepsis-related] Organ Failure Assessment (SOFA) score ≥ 2 (Singer et al., 2016). This definition highlights the importance of the systemic inflammatory response caused by immune disturbance. Pattern recognition receptors of innate immune cells recognize highly conserved microbial pathogen-associated molecular patterns (PAMPs), which activate signaling pathways, e.g., mitogen activated protein kinase (MAPK) and nuclear factor-kB (NF-kB), thus triggering the production and secretion of pro-inflammatory cytokines such as tumor necrosis factor alpha (TNF-a) and interleukin 1 (IL-1), IL-2, IL-6, and IL-8. These cytokines promote the adhesion between neutrophils and endothelial cells, which is followed by the activation of complement and blood coagulation cascades, and eventually lead to disseminated intravascular coagulation (Boyd et al., 2014; Huang et al., 2019). According to conventional view, after this early hyperinflammatory state, a subsequent hypoinflammatory state partly from the release of anti-inflammatory cytokines such as IL-4 and IL-10, results in widely immune suppression. However, newer paradigms indicate a phase at which the pro-inflammatory and immunosuppression may occur simultaneously, and the complex interactions between host (genetics and comorbidities) and pathogen (type, virulence, and burden) is the leading factor (Cecconi et al., 2018).

Despite recent advances in anti-infective therapy and advanced life support, the high mortality rate of sepsis remains an urgent clinical challenge. High medical costs and possible sequela such as renal insufficiency and cognitive impairment make sepsis a global public health issue (Singer et al., 2016). In addition, one-sixth of survivors of sepsis frequently suffer from long-term impairments such as physical, cognitive, and organ function (Delano and Ward, 2016; Prescott and Angus, 2018). Accelerated progression of preexisting chronic conditions, residual organ damage, and impaired immune function accounts for the deterioration of health after sepsis (Prescott and Angus, 2018). Since a sustained, uncontrolled inflammatory response is regarded as key factor promoting the onset of sepsis and multiple organ damage, novel anti-inflammatory therapies are eagerly pursued for sepsis treatment. However, clinical trials aimed at blocking cytokine responses, such those testing TNF-a inhibitors and Tolllike receptor 4 (TLR4)/myeloid differentiation factor 2 (MD2) antagonists, failed to decrease the mortality rate of patients with septic (Fisher et al., 1996; Opal et al., 2013). In view of the limited treatment options, complementary and replacement therapies are increasingly investigated to develop new therapeutic measures to treat sepsis and related organ damage.

As an alternative and complementary therapy, TCM is increasingly recognized for its efficacy and safety in the treatment of diseases. For instance, in the last 2 years, TCM has been shown to improve immunity and alleviate fever and other symptoms of COVID-19 (Kang et al., 2022). Indeed, TCM offers unique advantages in the treatment of inflammatory diseases such as sepsis (classified as "exogenous fever disease" in TCM) (Li C et al., 2018; Lu Z.B. et al., 2020). According to TCM tenets, the main principles of sepsis treatment are clearing away heat and detoxifying, clearing the internal organs and expelling heat, promoting blood circulation and removing blood stasis, and strengthening the body and solidifying the detoxification. Many studies, including some clinical trials, have reported the effectiveness of TCM in suppressing inflammatory pathways, regulating the immune response, and inhibiting oxidative stress (Gao et al., 2019; Song Y et al., 2019; Xia et al., 2019; Shang et al., 2020). Evidence supports as well the potential of TCM compounds, herbal extracts, and electroacupuncture in the prevention and treatment of heart-, brain-, lung-, and intestine-related diseases (Kim et al., 2007; Liu et al., 2018; Nabavi et al., 2018; Pang et al., 2018; Zhang W et al., 2019; Zhang X et al., 2019; Liu S et al., 2020). Therefore, this review aims to summarize the efficacy and mechanism of action of TCM compounds, herbal extracts, and electroacupuncture, in the treatment of sepsis and sepsis-related multiple organ damage.

2 TCM compounds

TCM compounds are prescriptions consisting of two or more substances that provide multi-target synergistic effects (Sun et al., 2017). Through mutual compatibility of different chemical substances, components of TCM herbs react with each other, thereby lowering toxicity and adverse side effects and enhancing the therapeutic effects (Zhang R et al., 2019). Several TCM compounds, such as Xuebijing injection, Shenfu injection, Huanglian Jiedu decoction, Dachengqi decoction, and Xijiao Dihuang decoction, demonstrated efficacy in the treatment of sepsis-related organ injury.

2.1 Xuebijing injection

Xuebijing injection (XBJI) is an injectable prescription obtained from a combination of carthami flos, paeoniae radix rubra, szechuan lovage rhizome, angelicae sinensis radix, and salviae miltiorrhizae (Li et al., 2021) that shows distinct anti-inflammatory activities in several settings. XBJI decreased the expression of IL-6, TNF-a, IL-1β, and IL-12 in mouse macrophages stimulated by Pam3CSK4 (a synthetic tripalmitoylated lipopeptide mimicking bacterial lipoproteins) (Li T et al., 2020). Its pharmacology targets are the NF-KB and MAPK pathways, and its effects are manifested by inhibition of the phosphorylation of IKK α/β , I κ B α , p65 NF- κ B, and JNK (Li T et al., 2020). High-dose XBJI increased the number of T-regs, reduced the number of Th-17 T cells, downregulated the expression of inflammatory cytokines such as IL-6 and TNF- α , inhibited the infiltration of neutrophils in lung and kidney tissues, and improved survival in cecal ligation and puncture (CLP) model mice (Chen et al., 2018). In another study addressing also the mouse CLP sepsis model, XBJI administration significantly improved renal microvascular perfusion and oxygenation and inhibited renal expression of IL-1β, IL-6, TNF-a, and high mobility group box 1 (HMGB1) protein, although without affecting the survival rate (Liu J et al., 2021).

2.2 Shenfu injection

Shenfu injection (SFI) is mainly composed of ginsenosides and aconitine alkaloids (Liu et al., 2019). SFI was reported to exert antioxidant, anti-inflammatory, anti-apoptotic, and immunoregulatory effects in a rabbit model of lipopolysaccharide (LPS)-induced septic shock. SFI decreased serum levels of lactate dehydrogenase (LDH) and aminotransferase (AST), improved myocardial metabolism, and protected tissue morphology in the heart, liver, and kidney (Liu et al., 2019). SFI also suppressed inflammatory markers, such as TNF- α and IL-1 β , in serum and heart of LPS-treated rats, and disrupted inflammatory signal transduction mediated by the mitogen-activated protein kinase repalmitoylated (MEK) and extracellular regulated protein kinase (ERK) pathways by decreasing p-MEK and p-ERK expression in LPS-stimulated H9C2 cells (Chen et al., 2020). Besides, SFI upregulated the expression of B cell lymphoma-2 (Bcl-2) and lowered the expression of Bid, t-Bid, and caspase-9, thus reducing cardiomyocyte apoptosis and attenuating myocardial injury in septic rats (Xu et al., 2020). In turn, beneficial effects of SFI on patients with septic shock were manifested by increased CD4⁺ and CD8⁺ T cells in peripheral blood, upregulated expression of human leukocyte antigen DR (HLA-DR) in monocytes, and enhanced cellular immunity (Zhang N et al., 2017). Importantly, clinical trials on patients with septic showed that SFI combined with conventional treatment led to significant improvement of clinical symptoms and prognosis, without obvious adverse reactions (Wen-Ting et al., 2012; Mo et al., 2014; Zhang Q et al., 2017; Wang X et al., 2019).

2.3 Shengmai injection

Shengmai injection (SMI), consisting of extracts from panax ginseng, ophiopogon japonicas, and schisandra chinensis, is one of the most widely used TCM prescriptions. According to the basic



pathways.

theory of TCM, simple tonic drugs like SMI cannot normally be used to treat infectious disease. However, in animal models of cardiac disease, marked improvement of myocardial metabolism was observed after treatment with SMI (Li et al., 2019; Cao et al., 2020). Specifically, SMI enhanced fatty acid and glucose oxidation, promoted mitochondrial biogenesis, and inhibited apoptosis by activating the AMPK signaling pathway in cardiomyocytes rendered hypertrophic by exposure to angiotensin II (Li et al., 2019). Meanwhile, SMI upregulated the expression of PTEN-induced kinase 1 (Pink1) and Parkin RBR E3 ubiquitin-protein ligase (Parkin), therefore improving myocardial mitophagy, in a mouse model of septic cardiomyopathy (Cao et al., 2020). Furthermore, SMI was shown to improve immune function and prolong survival in mice with CLP-induced peritonitis (Yu et al., 2005).

2.4 Huanglian Jiedu decoction

Huanglian Jiedu decoction (HLJDD) consist of rhizoma coptidis, radix scutellariae, cortex phellodendri, and fructus gardeniae and has been widely used in the treatment of inflammatory diseases (Lu Z et al., 2020). The beneficial regulatory role of HLJDD in lipid homeostasis represents the key mechanism of its anti-inflammatory actions. Upon LPS-induced inflammation in zebrafish, HLJDD ameliorated lipid

imbalance mainly through the glycerophospholipid metabolism pathway. By normalizing the production of proinflammatory lipid intermediates, this effect was proposed to underlie TLR4/myeloid differentiation factor 88 (MyD88)/NF-kB pathway inhibition and reduced secretion of IL-6, IL-1β, TNF-α, and IFN-γ (Zhou et al., 2019). Notably, berberine, baicalin, and gardenin, the main components of HLJDD, were shown to play a protective role against sepsis-related multi-organ damage by binding to lipid A to neutralize LPS activity. This resulted in inhibition of IL-6, TNF-a, and IFN-y secretion, as well as reduced synthesis of pathological lipid markers (Chen G et al., 2017). In a rat model of LPS-induced gingivitis, HLJDD administration suppressed serum inflammatory cytokines, lowered malondialdehyde (MDA) and reactive oxygen species (ROS) production, and upregulated total antioxidant capacity in periodontitis lysates. These effects were correlated with inhibition of AMPK and ERK1/2 expression (Zhang F et al., 2018).

2.5 Dachengqi decoction

Dachengqi decoction (DCQD) is composed of extracts from rheum palmatum l, magnolia henryi dunn, citrus aurantium l, and natrii sulfas. In a mouse model of LPS-induced acute lung injury (ALI), DCQD treatment inhibited TLR4/NF- κ B signaling and IL-6,

TABLE 1 Characteristics of TCM compounds on sepsis.

Compound	Efficacy	Composition	Animal/Cell model	Dose/Concentration	Targets	Ref
Xuebijing Injection	Regulating the balance of Tregs and Th17 cells; Decreasing inflammatory mediators and bacterial load	Honghua (Carthami tinctorii L.), Chishao (Paeonia lactiflora Pall.), Chuanxiong (Ligusticum chuanxiong Hort.), Danggui [Angelica sinensis (Oliv.) Diels], and Danshen (Salvia miltiorrhizae Bge.)	Mice: MRSA 7 × 10 ⁸ CFU, RAW264.7 cells: Pam3CSK4 100 ng/ mL (Li T et al., 2020); Mice: CLP (Chen et al., 2018); Rats: CLP (Liu J et al., 2021)	Mice: 5, 10 mL/kg, RAW264.7 cells: 3, 10, 30 μL/mL (Li T et al., 2020); Mice: 18 mL/kg (Chen et al., 2018); Rats: 4 mL/kg (Liu J et al., 2021)	NF-κB and MAPK↓; PI3K/Akt phosphorylation ↓; HMGB1 ↓	Chen et al. (2018); Li T et a (2020); Liu J et al. (2021)
Shenfu injection	Improving energy metabolism and antioxidation; Attenuating the inflammation and apoptosis	Ginsenosides and aconitine alkaloids	Rabbits: LPS .6 mg/kg (Liu et al., 2019); Rats: LPS 20 mg/kg, H9C2 cells: LPS 16 μg/mL (Chen et al., 2020); Mice: LPS 8 mg/kg (Xu et al., 2020)	Rabbits: 4.5, 6, 8 mL/kg (Liu et al., 2019); Rats: 10 mL/kg, Cells: 80 μL/mL (Chen et al., 2020); Mice: 3, 10 mL/kg (Xu et al., 2020)	p-MEK ↓, p-ERK ↓; cleaved- caspase 3 ↓, caspase 9↓, Bax↓; Bid and t-Bid ↓; Bcl-2 ↑	Chen et al. (2020); Liu et al (2019); Xu et al (2020)
Shengmai injection	Promoting myocardial mitochondrial autophagy and mitochondrial membrane potential	Panax ginseng, Ophiopogon japonicas and Schisandra chinensis	Mice: LPS 8 mg/kg, HL-1 cells: LPS 1 µg/mL	Mice: 10 mL/kg	caspase-3/Beclin-1axis ↓	Cao et al. (2020)
Huanglian Jiedu decoction	Anti-inflammatory	Coptidis Rhizoma, Scutellariae Radix, Phellodendri Chinensis Cortex, and Gardeniae Fructus	Zebrafishes: LPS 10 mg/mL	Zebrafishes: 50 µg/mL	TLR4/MyD88 ↓	Zhou et al. (2019)
Dachengqi decoction	Alleviating the release of inflammatory cytokines and regulating capillary permeability	Da Huang, Houpu, Zhishi, and Mangxia	Rats: LPS 10 mg/kg, HUVEC-5a cells: LPS 100 ng/mL	Rats: .9 g/kg, HUVEC-5a cells: 100 μg/mL	TLR4 \downarrow ; NF- κ B \downarrow	Hu et al. (2019)
Xijiao Dihuang decoction	Inhibiting aerobic glycolysis and inflammatory cytokines	Rehmannia, Peony, Cortex Moudan and Cornu Bubali	Rats: CLP, NR8383 cells: LPS 1 µg/mL (Lu et al., 2020a; Lu et al., 2020b)	Rats: 12.5, 25 g/kg, NR8383 cells: 4 mg/ mL (Lu et al., 2020b); Rats: 25 g/kg, NR8383 cells: 4 mg/mL (Lu et al., 2020a)	TLR4/HIF-1α/PKM2 ↓; NF- κB ↓; HIF-1α ↓	Lu et al. (2020a) Lu et al. (2020b
Liang-Ge-San	Inhibiting inflammatory response; Reducing infiltration of inflammatory cells; Decreasing recruitment of macrophages and neutrophils	Fructus forsythiae (Lian Qiao), Rheum officinale (Da Huang), Fructus Gardeniae (Zhi Zi), Radix Scutellariae (Huang Qin), Liquorice (Gan Cao), Mint (Bo He), Mirabilite (Mang Xiao)	Zebrafishes larvae: LPS .5 mg/mL, RAW264.7 cells: LPS 100 ng/mL	Zebrafish: 62.5, 125, 250 μg/mL, RAW264.7 cells: 25, 50, 100 μg/mL	p-JNK ↓; p-Nur77 ↓	Zhou et al. (2020)
Xuanbai Chengqi decoction	Attenuating proinflammatory cytokines release	Rheum palmatum rhizome and root (Dahuang), Gypsum Fibrosum (Shigao), Prunus armeniaca seed (Kuxingren), and <i>Trichosanthes kirilowii</i> fruit (Gualou)	Rats: LPS 8 mg/kg	Rats: 5, 20 g/kg	PI3K/mTOR/HIF-1α/VEGF↓	Zhu et al. (2021
Sini decoction	Inhibiting inflammatory cell infiltration and the production of inflammatory cytokines; Antioxidant stress	Aconite, Liquorice and Ginger Rhizome	Mice: LPS 8 mg/kg, HUVECs cells: LPS 1 µg/mL	Mice: 5 g/kg, HUVECs cells: 6, 12.5, 25 mg/mL	MAPK↓, ACE/AT1R↓; ACE2/ Ang1–7↑	Chen Q et al. (2019)
Fangji Fuling decoction	Inhibiting inflammatory reaction and apoptosis	Stephania tetrandra S.Moore (Fangji), Astragalus propinquus Schischkin (Huangqi), Cinnamomum cassia (Nees & T.Nees) J.Presl (Guizhi), Glycyrrhiza uralensis Fisch. (Gancao), and Poria cocos (Schwein.) F.A.Wolf (Fuling)	Mice: LPS 10 mg/kg, HK-2 cells: LPS 1 μg/mL	Mice: 25,50 mg/kg, HK-2 cells: 200 µg/mL	iNOS↓; NF-κB↓	Su et al. (2018)
Xuefu Zhuyu decoction	Inhibiting apoptosis; antioxidation	Prunus persica (L) Batch. (Tao Ren), Angelicae sinensis (oliv.) Diels. (Dang Gui), Ligusticumi chuangxiong Hort. (Chuang Xiong), Carthamus tinctorius L.	Mice: LPS 10 mg/kg	Mice: 3.9, 7.8, 15.6 g/kg	SOD \uparrow , Bcl-2 \uparrow ; TNF- $\alpha \downarrow$, IL-1 $\beta \downarrow$, IL-6 \downarrow ; MDA \downarrow , Bax \downarrow , Caspase-3 \downarrow	Meng F et al. (2018)

(Continued on following page)

ABLE 1 (Continued)	TABLE 1 (Continued) Characteristics of TCM compounds on sepsis.	ı sepsis.				
Compound	Efficacy	Composition	Animal/Cell model	Dose/Concentration	Targets	Ref
		(Hong Hua), Paeonia lactiflora Pall. (Chi Shao), Rehmannia glutinosa Libosch. (Di Huang), Citrus aurantium L. (Zhi Qiao), Bupleurum chinense DC. (Chai Hu), <i>Platycodon grandiflorum</i> (Jacq) A. DC. (Jie Geng), Achyranthes bidentata BL. (Niu Xi), Glycyrthiza uralensis Fisch. (Gan Cao)				
Lianhua Qingwen	Inhibiting Apoptosis	Forsythia suspensa (Thunb.) Vahl (Lianqiao, LQ), Lonicera japonica Thunb. (Jinyinhua, JYH), Ephedra sinica Stapf (Mahuang, MH), Isatis tinctoria L. (Banlangen, BLG), Pogostemon cablin (Blanco) Benth. Guanghuoxiang, GHX), Rheum palmatum L. (Dahuang, DH), <i>Glycyrrhiza</i> uralensis Fisch. (Gancao, GC), Dryopteris crassichizoma Nakai (Mianmaguarzhong, GZ), Rhodiola crenulata (Hook.f. and Thomson) H. Ohba (Hongjingtian, HJT), Houttuynia cordata Thunb. (Yuxingren, KXR), Gypsum and I. (Kuxingren, KXR), Gypsum and I-Menthol	Mice: LPS 5 mg/kg	Mice: .7, 1.4, 2.8 g/kg	Bcl-2f; BaxJ, caspase-3J, caspase-9J; p53J	Yang et al. (2021)

IL-8, and TNF- α secretion in lung tissue, and reduced pulmonary edema by upregulating aquaporin (AQP) 1/5 expression (Hu et al., 2019). Another study showed that DQCD attenuated intestinal vascular endothelial injury in rats with severe acute pancreatitis (SAP) induced by cerulein and LPS, and decreased matrix metalloproteinase 9 (MMP-9) and junctional adhesion molecule C (JAM-C) expression, while increasing AQP-1 expression, in TNF- α -treated vascular endothelial cells (Pan et al., 2017). Further *in vivo* and *in vitro* research on the SAP rat model indicated that DCQD inhibited the phosphatidylinositol 3kinase (PI3K)/protein kinase B (AKT) signaling pathway, thereby inhibiting inflammation by decreasing production of IL-1 β , IL-6, and TNF- α and promoting apoptosis of pancreatic acinar cells (Sun et al., 2020).

2.6 Xijiao Dihuang decoction

The Xijiao Dihuang decoction (XJDHD) comprises rehmannia, peony, cortex moudan, and cornu bubali and is used in China for the treatment of sepsis (Lu et al., 2020b). XJDHD administration was shown to improve the survival rate of rats subjected to CLP, as well as of cultured macrophages, by inhibiting aerobic glycolysis triggered by the TLR4/hypoxia-inducible factor 1 α (HIF-1 α)/ pyruvate kinase M2 (PKM2) pathway (Lu et al., 2020b). Additional research on the above sepsis models further indicated that prolonged survival correlated with XJDHDmediated inhibition of HIF-1 α and p65 (Lu et al., 2020a).

3 TCM monomers

3.1 Triterpenoid

3.1.1 Tanshinone IIA

Tanshinone IIA (TSA) occurs in the dried roots and rhizomes of salvia miltiorrhiza (lamiaceae), and is one of the main pharmacologically active components of hydrophilic tanshinones (Wang N et al., 2020). TSA has anti-inflammatory activity by inhibiting a variety of cytokines. In LPS-stimulated bone marrowderived macrophages (BMDMs), TSA exposure inhibited succinate dehydrogenase (SDH)-mediated IL-1β and IL-6 production and blocked BMDM polarization towards the M1 phenotype (Liu Q.Y. et al., 2021). In addition, suggesting beneficial effects against neuroinflammatory and neurotoxic insults, TSA pretreatment was shown to attenuate pro-inflammatory cytokine secretion through inhibition of TLR4, MyD88, and TNF receptor associated factor 6 (TRAF6) expression and subsequent repression of signaling through the NF-KB and MAPK pathways in LPS-treated human U87 astrocytoma cells (Jin et al., 2020). TSA treatment also reduced calcium inflow, inhibited transient receptor potential melastatin 7 (TRPM7), and suppressed the release of proinflammatory cytokines in pulmonary interstitial macrophages from rats with sepsis-induced ALI (Li J et al., 2018). Meanwhile, incubation with Salvia miltiorrhiza extract decreased LPS-induced phosphorylation of IkB-a and IKK, thus inhibiting NF-kB activity, inhibited MAPK phosphorylation, and disrupted TLR4 dimerization to prevent TLR4-MyD88 complex formation in RAW264.7 cells (Gao et al., 2017).

TABLE 2 Characteristics of TCM extracts/monomers on sepsis.

Tanshinone IIA	C ₁₉ H ₁₈ O ₃	Anti-inflammatory; Antifibrotic; Antioxidative activities	Salvia miltiorrhiza	Mice: LPS 15 mg/kg, RAW264.7 cells: LPS	Mice: 20 mg/kg, RAW264.7 cells: 10 μM(Liu Q.Y.	Sirt2 ↑ HIF-1α ↓, SDH↓; NLRP3 ↓, HK-II ↓	
Astragaloside IV	\times		Bunge	100 ng/mL (Liu Q.Y. et al., 2021); U87 cells: LPS 100 ng/mL (Jin et al., 2020)	where 20 mg ng, KAW 2017, Cents: 10 μM(Jin et al., 2021); U87 cells: 1, 5, 10 μM(Jin et al., 2020)	PKM2 ↓; TLR4/NF-ĸB/MAPKs↓	Liu Q.Y. et al. (2021) Jin et al. (2020)
	$\begin{array}{c} C_{41}H_{68}O_{14} \\ \underset{m_{0}}{\overset{m_{0}}{\leftarrow}} \\ \underset{m_{0}}{\overset{m_{0}}{\overset{m_{0}}{\leftarrow}} \\ \underset{m_{0}}{\overset{m_{0}}{\overset{m_{0}}{\overset{m_{0}}{\leftarrow}} \\ \underset{m_{0}}{\overset{m_{0}}{\overset{m_{0}}{\overset{m_{0}}{\overset{m_{0}}{\overset}} \\ \underset{m_{0}}{\overset{m_{0}}{m_{$	Anti-inflammatory; Anti-oxidative and anti- apoptotic effects; Decreasing barrier permeability; Increasing tight junction	Astragalus membranaceus (Fisch) Bge	Mice: CLP, Caco-2 cells: LPS 100 µg/mL (Xie et al., 2020); Rats: CLP (Huang and Li, 2016); Mice: LPS 1 mg/kg (Song et al., 2018)	Mice: 3 mg/kg Caco-2 cells: 200 µg/mL (Xie et al., 2020); Rats: 2.5, 5, 10 mg/kg (Huang and Li, 2016); Mice: 20, 40 mg/kg (Song et al., 2018)	RhoA/NLRP3 \downarrow PPARy \uparrow NF- $\kappa B \downarrow$	Xie et al. (2020) Huang and Li, (2016) Song et al. (2018)
Glycyrrhizin	$\begin{array}{c} C_{42}H_{62}O_{16} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\$	Suppressing proinflammatory cytokines and apoptosis	Gancao (licorice root)	Rats: CLP, NR8383 cells: LPS 1 µg/mL or HMGB1 1 µg/mL	Rats:10 mg/kg, NR8383 cells: 10,50, 100 µg/mL	HMGB1↓, RAGE/TLR4 ↓ MAPK ↓ NF-κB ↓	Zhao et al. (2017)
Triptolide	$C_{20}H_{24}O_6$	Anti-inflammatory	Tripterygium wilfordii Hook.F	HUVECs: LPS 1 µg/mL	HUVECs: 25, 50, 100 nM	NF-κB↓	Song C et al. (2019)
Artemisinin	$\begin{array}{c} C_{15}H_{22}O_5\\ \hline \\ \hline \\ \hline \\ \hline \\ \hline \\ \\ \hline \\ \\ \hline \\ \\ H \\ \\ \\ \\$	Anti-inflammatory; Improving cognitive impairments and attenuating neuronal damage and microglial activation	artemisinin	Mice: LPS 750 µg/kg, BV2 cells: LPS 100 ng/mL	Mice: 30 mg/kg, BV2 cells: 40 µM	AMPKαl ↑, NF-κΒ ↓	Lin S.P et al (2021)
Ginsenoside Rg1	$\begin{array}{c} C_{42}H_{72}O_{14} \\ H_0 & H_0 \\ H_0 & H_1 \\ H_0 & H_1 \\ H_0 & H_1 \\ H_0 & H_0 \\ H_$	Suppressing inflammation and apoptosis	ginseng	Mice: LPS 5 mg/kg, NRCMs cells: LPS 1 μg/mL	NRCMs cells: 20 µM	TLR4/NF-kB/NLRP3 ↓	Luo et al. (2020)
Apigenin	$C_{1s}H_{10}Os$	Enhancing the antioxidant ability and decreasing the production of inflammatory cytokines	parsley, chamomile and propolis	Mice: D-GalN 700 mg/kg, LPS 20 µg/kg	Mice: 100, 200 mg/kg	Nrf-2 ↑, PPARγ ↑ NF-κB ↓	Zhou et al. (2017)
Salidroside	$\begin{array}{c} C_{14}H_{20}O_7 \\ \\ \overset{HO}{\longrightarrow} \\ HO^{+} \\ \overset{HO}{\longrightarrow} \\ \\ H^{+} \\ \end{array} \\ \begin{array}{c} C_{14}H_{20}O_7 \\ \\ OH \\ \end{array} \\ \end{array}$	Suppressing myocardial lipid peroxidation and inhibiting inflammatory cytokines	Rhodiola rosea	Rats: LPS 15 mg/kg, H9C2 cells: LPS 4 µg/mL (Chen L et al., 2017); Mice: CLP, RAW264.7 cells: LPS 1 µg/mL (Lan et al., 2017); HUVECs: LPS 10 µg/mL (You et al., 2021)	Rats: 20, 40 mg/kg, H9C2 cells: 20, 40 μ M (Chen L et al., 2017) Mice:20, 40 mg/kg, RAW264.7 cells: 30, 60, 120 μ M(Lan et al., 2017) HUVECs: 50 μ M(You et al., 2021)	NF-ĸB ↓, PI3K/Akt/mTOR ↓; SIRT1 ↑, HMGB1 ↓, NLRP3↓	Chen L et al. (2017) Lan et al. (2017) You et al.

(Continued on following page)

TABLE 2 (Continued) Characteristics of TCM extracts/monomers on sepsis.

Monomers	Molecular structure	Effects	Source	Animal/Cell model	Dose/Concentration	Targets	Ref
Baicalein	$\begin{array}{c} C_{15}H_{10}O_5\\ HO & C_{15}H_{10}O_5\\ HO & C_{15}H_{10}O_5 \end{array}$	Suppressing the ROS level and pro-inflammatory cytokines	root of Scutellaria baicalensis Georgi	BV-2 cells: LPS 0.1 µg/mL (Yan et al., 2020); THP-1 and RAW264.7 cells: LPS 1 µg/mL and ATP 5 mM (Luo et al., 2017)	BV-2 cells: .5, 1, 2, 4 μM(Yan et al., 2020) THP-1 and RAW264.7 cells: 10, 25, 50 μM(Luo et al., 2017)	ROS ↓, COX2 ↓, NF-κB ↓; TLR4/MyD88 MD- 2/TLR4 complex ↓, NLRP3 ↓	Yan et al. (2020) Luo et al. (2017)
Resveratrol	$\begin{array}{c} C_{14}H_{12}O_3 \\ \\ \overset{\text{HO}}{\underset{\text{OH}}{\longrightarrow}} \mathcal{O}_{14}^{\text{OH}}C_{14}H_{12}O_3 \end{array}$	Anti-inflammation and anti-apoptotic; Improving in vascular relaxation reactivity	mulberries, peanuts, and grape skins	Mice: CLP, MH-S cells: LPS 150 µg/mL (Yang et al., 2018); PBMC: LPS 100 ng/mL (Wang B et al., 2020); Mice: CLP (Zhang Z.S. et al., 2019)	Mice: 40 mg/kg, MH-S cells: 10 μM(Yang et al., 2018) PBMC: 40 μM(Wang B et al., 2020) Mice: 5,10 mg/kg (Zhang Z.S. et al., 2019)	VEGF-B ↑ NF-κB ↓, SphK ↓, ERK1/2 phosphorylation ↓ MyD88 ↓; Rac-1 ↓, HIF- 1α ↓	Yang et al. (2018) Wang B et al. (2020)
Paeonol	C ₉ H ₁₀ O ₃ C ₉ H ₁₀ O ₃	Inhibiting the inflammatory response; Promoting the phagocytic ability of macrophages	moutan cortex	Mice: CLP, RAW264.7 cells: LPS .2 μg/mL (Mei et al., 2019); Mice: LPS .2 mg/kg, RAW264.7 cells: LPS .2 μg/mL (Miao et al., 2020)	Mice: 120 mg/kg, RAW264.7 cells: 1 mM(Mei et al., 2019); Mice: 80 mg/kg, RAW264.7 cells: 600, 1000 nM(Miao et al., 2020)	miR-339-5p ↑, HMGB1 ↓, IKK-β↓, P53 ↓	Zhang Z.S. et al. (2019) Mei et al. (2019) Miao et al. (2020)
6-Gingerol	C ₁₇ H ₂₆ O ₄	Inhibiting inflammasome formation and pyroptosis	ginger rhizome	Mice: CLP, RAW264.7 and BMDM cells: LPS 100 ng/mL and ATP 5 mM(Zhang F.L. et al., 2020); Mice: CLP, RAW264.7 cells: LPS 1 µg/ mL and ATP 5 mM(Hong et al., 2020)	Mice: 20 mg/kg, BMDMs: 8μM, RAW264.7: 4 μM(Zhang F.L. et al., 2020) Mice: 40 mg/kg, RAW264.7 cells: 8 μM(Hong et al., 2020)	MAPK ↓, NLRP3 ↓, Nrf2 ↑	Zhang F.L. et al. (2020) Hong et al. (2020)
Berberine	C ₂₀ H ₁₈ NO ₄	Inhibiting gluconeogenesis, insulin resistance and proinflammatory molecule release; Lowering gut- vascular barrier hyperpermeability	Coptis chinensis	Rats: CLP, HepG2 and rat intestinal microvascular endothelial cells: LPS 100 ng/mL (Li Y et al., 2020); Rats: CLP, RIMECs: LPS 50 ng/mL (He et al., 2018); Mice: LPS 5 mg/kg (Xu et al., 2021); Rats: LPS 10 mg/kg (Chen et al., 2021); RAW264.7 cells: LPS 100 ng/mL (Zhang H et al., 2017)	Rats:25, 50, 100 mg/kg, HepG2 and rat intestinal microvascular endothelial cells: 5, 10, 20 μ M(Li Y et al., 2020); Rats: 25, 50 mg/kg, RIMECs: 10, 20 μ M(He et al., 2018); Mice: i.p. 1, 2 mg/kg and inh. 1, 2 mg/mL (Xu et al., 2021); Rats: 50 mg/kg (Chen et al., 2021), RAW264.7 cells: 1, 2.5, 5 μ M(Zhang H et al., 2017)	ApoM/S1P ↑ Wnt/beta-catenin ↑ TLR4/NF-ĸB ↓ JAK2/STAT3 ↓; SIRT1 ↑	Li Y et al. (2020) He et al. (2018) Xu et al. (2021) Chen et al. (2021) Zhang H et al. (2017)
Cordycepin	$\begin{array}{c} C_{10}H_{13}N_{5}O_{3} \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ H_{0} \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & $	Alleviating inflammation	Cordyceps sinensis	Mice: LPS 30 mg/kg (Qing et al., 2018); RAW264.7 cells and THP-1 cells: LPS 100 ng/ mL (Yang J et al., 2017)	Mice: 1, 10, 30 mg/kg (Qing et al., 2018); RAW264.7 cells: 6.25, 12.5, 25, 50 μmol/L (Yang J et al., 2017)	Nrf2 ↑, HO-1 ↑; NLRP3 ↓, ERK1/2 ↓, COX2 ↓	Qing et al. (2018) Yang J et al. (2017)
Emodin	$\begin{array}{c} C_{15}H_{10}O_5\\ OH\\ O\\ OH\\ O\\ OH\end{array} \begin{array}{c} C_{15}H_{10}O_5\\ C_{15}H_{10}O_5\\ OH\end{array}$	Ameliorating hypercoagulation and fibrinolytic inhibition; Inhibiting inflammatory reaction; Anti-neuroinflammatory	Radix rhizoma Rhei	Mice: LPS 40µL, 4 mg/mL inhale (Liu B et al., 2020); Microglia cells: LPS 1 µg/mL (Park et al., 2016)	Mice: 5,10, 20 mg/kg (Liu B et al., 2020); Microglia cells: 40 $\mu M({\rm Park}~et~al.,~2016)$	NF-κB and p65 DNA binding activity↓; AMPK/Nrf2 ↑; NF-κB ↓, AP-1↓, STAT ↓, MAPK ↓	Liu B et al. (2020) Park et al. (2016)

3.1.2 Astragaloside IV

Astragaloside IV (AS-IV) is a small saponin obtained from the roots of astragalus membranaceus (Xia et al., 2020). AS-IV has antiinflammatory and neuroprotective activities. Studies in the CLP mouse model showed that AS-IV treatment inhibited the secretion of inflammatory cytokines and downregulated the expression of NLRP3, apoptosis associated speck like protein containing CARD (ASC), and cleaved caspase-1 in both intestinal tissue and BALF, thus reducing multi-organ injury (Huang and Li, 2016; Xie et al., 2020). AS-IV administration was also shown to protect blood-brain barrier integrity in LPS-treated mice by upregulating zonula occludens-1 (ZO-1) and occludin and downregulating vascular cell adhesion molecule 1 (VCAM1) expression in brain micro-vessels; experimental results in cultured bEnd.3 and microglial cells attributed this protective effect to both activation of the Nrf2dependent antioxidant response and inhibition of the NF-KB/ NLRP3 inflammasome signaling pathway (Li H et al., 2018; Yang et al., 2019). Further research showed that AS-IV attenuated LPSinduced neuroinflammation in mice by upregulating the expression of PPARy and inducing phosphorylation of glycogen synthase kinase-3β (GSK3-β) in the hippocampus. AS-IV-mediated neuroprotection was further evidenced by reduced expression of inflammatory factors, in association with NF- κ B signaling suppression (Song et al., 2018). In a mouse model of acute E. coli peritoneal infection, AS-IV treatment alleviated peritonitis symptoms by promoting the influx of neutrophils to the infection site, an effect mediated by inhibition of G proteincoupled receptor kinase-2 (GRK2) expression and subsequent blockade of LPS-induced suppression of CXC motif chemokine receptor 2 (CXCR2) on neutrophils (Huang et al., 2016). Moreover, in LPS-treated rats, AS-IV administration attenuated cardiac dysfunction, reduced myocardial damage, improved mitochondrial energy metabolism, and inhibited cardiomyocyte apoptosis and autophagy by downregulating miRNA-1 expression (Wang et al., 2021).

3.1.3 Glycyrrhizin

Glycyrrhizin is a natural triterpene glycoside and the major active component of gan cao (licorice root). Glycyrrhizin treatment was associated with significantly improved survival in a rat model of CLP-induced sepsis. This effect was associated with suppression of HMGB1 expression and inhibition of downstream MAPK/NF- κ B pathways both *in vivo* and *in vitro* (Zhao et al., 2017). In turn, in LPS-treated mice, a protective role of glycyrrhizin on heart and lung was attributed to the increase in the ratio of myeloid-derived suppressor cells (MDSCs) to CD11b⁺Gr1 bone marrow cells in the blood, heart, and lungs (Seo et al., 2017). In addition, glycyrrhizin was shown to negatively regulate PI3K/mTOR signaling and inhibit the expression of inflammatory markers such as iNOS, COX-2, HMGB1, TNF- α , IL-1 β , and IL-6 in LPS-stimulated human liver macrophages (Shen et al., 2020).

3.1.4 Triptolide

Triptolide is a primary bioactive ingredient of the roots of the Chinese herb tripterygium wilfordii Hook. F. Triptolide exerted vascular anti-inflammatory effects by downregulating proinflammatory cytokine and chemokine secretion, attenuating VCAM-1 and intercellular adhesion molecule-1 (ICAM-1) expression, and inhibiting $I\kappa B\alpha$ phosphorylation and NF- κB p65 DNA binding activity in LPS-stimulated endothelial cells (Song

C et al., 2019). ZT01, a triptolide derivative, showed also antiinflammatory activity *via* reducing TNF- α and IL-6 levels, blocking the formation of the TGF- β -activated kinase1 (TKA1)/TAK1-binding protein1 (TAB1) complex, and inhibiting the phosphorylation of both mitogen-activated protein kinase kinase 4 (MKK4) and JNK in both *in vivo* and *in vitro* sepsis models (Fu et al., 2020).

3.1.5 Artemisinin

Artemisinin is a sesquiterpene lactone obtained from the sweet wormwood plant Artemisia annua. Although mainly recognized by its efficacy to treat malaria, there is growing evidence that artemisinin may be useful to treat several health conditions, including sepsis, by exerting potent anti-inflammatory and immunoregulatory effects. Artemisinin treatment was shown to significantly reduce LPSinduced cognitive impairment in mice by attenuating both neuronal damage and microglial activation in the hippocampus (Lin S.P et al., 2021). This study further showed that artemisinin reduced TNF-a, IL-6, IL-1a, IL-1β, and iNOS production and suppressed the migratory ability of LPS-stimulated BV2 microglial cells by activating the AMPKa1 pathway and inhibiting nuclear translocation of NF-KB (Lin S.P et al., 2021). Artesunate is a watersoluble derivative of artemisinin with multiple biological activities. In rats with LPS-induced ALI, artesunate treatment reduced TNF- α and IL-6 levels in BALF, decreased oxidative stress markers (i.e., MDA, SOD, and GSH-Px), and reduced apoptosis of lung cells by activating the mTOR/AKT/PI3K signaling pathway (Zhang E et al., 2020). Notably, artesunate was reported to reverse sepsis-induced immunosuppression in mice by interacting with the vitamin D receptor in an autophagy- and NF-KB-dependent manner (Shang et al., 2020).

3.1.6 Ginsenoside Rg1

Ginsenoside Rg1 (GRg1), a triterpenoid saponin, is the main bioactive component of ginseng (panax ginseng). Administration of GRg1 ameliorated LPS-induced acute myocardial injury by inhibiting the NF- κ B pathway and attenuating inflammatory responses, including NLRP3 expression (Luo et al., 2020). GRg1 treatment also alleviated lung injury and extended survival in mice with LPSinduced ALI; experiments in LPS-challenged pulmonary epithelial A549 cells further showed that GRg1 exposure inhibited ROS production, prevented apoptosis, and reduced ER stress and inflammatory cytokine expression by upregulating Sirt1 (Wang Q.L. et al., 2019). In animal models of sepsis-associated ALI and encephalopathy, the protective effect of GRg1 was shown to also depend on autophagy enhancement, *via* a mechanism related to the activation of Nrf-2 and inhibition of NF- κ B signal transduction (Li Y et al., 2017; Ji et al., 2021).

3.2 Flavonoids

3.2.1 Apigenin

Apigenin, a flavonoid found in abundance in many fruits and vegetables, has shown remarkable efficiency in controlling the inflammatory response. In LPS-treated mice, apigenin administration decreased the levels of cardiac troponin I (cTnI), cardiac myosin light chain-1 (cMLC1), and inflammatory cytokines such as TNF- α , IL-1 β , MIP-1 α , and MIP-2, an effect attributed to reduced NF- κ B nuclear translocation and enhanced peroxisome

proliferator-activated receptor gamma (PPAR γ) nuclear translocation (Li F et al., 2017; Zhou et al., 2017). In addition, apigenin ameliorated LPS-induced acute liver injury in mice *via* inhibiting oxidative stress, evidenced by upregulation of the activities of superoxide dismutase (SOD), catalase (CAT), glutathione-S-transferase (GST) and glutathione reductase (GR). This protective role was partly dependent on increased hepatic expression of nuclear factor erythroid 2-related factor 2 (Nrf-2) (Zhou et al., 2017). The potential of apigenin to counteract neuroinflammation was in turn suggested by its ability to downregulate the expression of CD68 (an M1 pro-inflammatory microglia marker), OX42 (a microglia activation marker), IL-6, and glycoprotein 130 in LPS-stimulated neuronal/glial co-cultures (Dourado et al., 2020).

3.2.2 Salidroside

Salidroside is extracted mainly from the root and rhizome tissues of the rose (rhodiola rosea). Many studies reported that salidroside has anti-inflammatory, anti-oxidative, and antibacterial properties. In rats with myocardial or lung injury triggered by LPS-induced endotoxemia, salidroside enhanced antioxidative activity and inhibited iNOS and COX2 expression, as well as NF-KB phosphorylation, in cardiac and lung tissues (Chen L et al., 2017; Jingyan et al., 2017; Zheng et al., 2020). Salidroside also reduced lung inflammation and alleviated ALI symptoms by upregulating Sirt1 expression and inhibiting both NF-KB activity and nucleocytoplasmic translocation of HMGB1 both in vivo and in vitro (Lan et al., 2017; Qi et al., 2017). In LPS-stimulated HUVECs, salidroside exposure increased antioxidant activity, inhibited apoptosis and NLRP3 inflammasome activation, and decreased the expression of NLRP3-related proteins, including ASC and caspase-1 (You et al., 2021).

3.2.3 Baicalein

Baicalein is a flavonoid extracted from the roots of the Chinese herb scutellaria baicalensis georgi. Its anti-inflammatory and antioxidative qualities have been demonstrated in several experimental settings. Baicalein-mediated protection against neuroinflammation was exemplified by its ability to downregulate LPS-induced NO generation, inhibit the expression of inflammatory cytokines such as IL-6, TNF-a, and COX2, and suppress NF-kB and p65-MAPK signaling in BV2 microglial cells and macrophages (Luo et al., 2017; Yan et al., 2020). Baicalin alleviated LPS-induced liver injury in mice by inhibiting the expression of IL-1a, IL-1β, and gasdermin D (GSDMD) and blocking NLRP3/IL-1β signaling (Xiao et al., 2021). Moreover, baicalin prevented the development of LPS-induced ALI and alleviated colitis symptoms by blocking LPS-induced TLR4/MD-2 complex formation, inhibiting the activation of MAPK and NF- κ B signaling pathways, and reducing leukocyte infiltration and production of inflammatory mediators (Luo et al., 2017; Chen H et al., 2019).

3.3 Phenols

3.3.1 Resveratrol

Resveratrol is a non-flavonoid polyphenol with potent antioxidant properties, present in the skin of fruits such as grapes and berries. Resveratrol pretreatment protected mice against CLP-induced ALI. Research on LPS stimulated MH-S alveolar macrophages suggested

that the underlying mechanism was related to inhibition of NF-KB and MAPK pathways, as well as of LPS-induced autophagy, which depended on enhanced expression of vascular endothelial growth factor B (VEGF-B) (Yang et al., 2018). In primary monocytes, resveratrol inhibited the LPS-stimulated inflammatory response by blocking phospholipase D activity and its downstream signaling molecules SphK1, ERK1/2, and NF-KB, and protected mice against septic shock induced by CLP (Wang B et al., 2020). In rats subjected to CLP, resveratrol improved vasodilation and hemodynamic parameters by upregulating endothelial nitric oxide synthase (eNOS) and downregulating iNOS, Rac family small GTPase 1 (RAC-1), and HIF-1a expression in arterial tissue (Zhang Z.S. et al., 2019). Resveratrol was also shown to ameliorate acute kidney injury (AKI) induced by sepsis by inhibiting renal inflammation triggered by endoplasmic reticulum (ER) stress activated-IRE1/NF-ĸB pathway activation (Wang et al., 2017). In LPS-treated mice, cardioprotective effects of resveratrol, evidenced by decreased 4-hydroxynonenal and MDA levels in myocardial tissue and improved contractility and Ca²⁺ homeostasis in cardiomyocytes, were attributed to increased Nrf-2 expression and to phospholamban oligomerization leading to enhanced SERCA2a activity (Bai et al., 2016).

3.3.2 Paeonol

Paeonol, a phenolic compound found in peonies such as paeonia suffruticosa (moutan cortex), shows multiple pharmacological effects, including anti-inflammatory and anti-tumoral activities. Paeonol exposure increased the expression of miR-339-5p and downregulated the expression of inflammatory markers, such as TNF-α, IL-1β, IKK-β, and HMGB1, in LPS-stimulated RAW264.7 cells (Mei et al., 2019). In a rat model of sepsis-induced AKI, paeonol treatment showed protective effects by lowering serum levels of TNF- α and IL-1 β and suppressing NF- κ B signaling in renal tissue (Mei et al., 2019). In the advanced stage of sepsis, impaired phagocytic activity of macrophages and monocytes contributes to immune dysfunction. Interestingly, the decline in the phagocytic ability of peritoneal macrophages induced by LPS could be reversed by paeonol co-administration, an effect mediated by suppression of HMGB1 nucleocytoplasmic translocation (Miao et al., 2020).

3.3.3 6-gingerol

6-gingerol, a major polyphenol extracted from the ginger rhizome (zingiber officinale roscoe), exhibits anti-inflammatory, antioxidant, anticancer, and neuroprotective properties. Treatment with 6-gingerol suppressed systemic IL-1ß release and prolonged survival in mice with CLP-induced sepsis. Complementary in vitro studies showed that 6-gingerol pre-treatment blocked MAPKdependent NLRP3 inflammasome activation in LPS/ATP-treated BMDMs and RAW264.7 cells, which attenuated pyroptosis and therefore decreased the release of mature IL-1 β into the medium (Zhang F.L. et al., 2020). In mice with CLP-induced acute liver injury, 6-gingerol administration reduced serum levels of AST, ALT, and IL-1β and elicited antioxidant and anti-apoptotic effects by upregulating hepatic Nrf-2 and HO-1 transcription (Hong et al., 2020). Similarly, renoprotective effects of 6-gingerol, consistent with its antioxidant and anti-inflammatory properties, were reported in a rat model of sepsis-induced AKI (Rodrigues et al., 2018).

3.4 Alkaloids

3.4.1 Berberine

Berberine is potent anti-inflammatory alkaloid compound extracted from herbs such as cortex phellodendri and rhizoma coptidis. In animal studies, berberine treatment ameliorated sepsisrelated intestinal vascular barrier injury by modulating the Apoliprotein M (ApoM)/Sphingosine-1-Phosphate (S1P) axis and the Wnt/β-catenin pathway (He et al., 2018; Li Y et al., 2020). In LPS-induced acute respiratory distress syndrome (ARDS), berberine treatment prevented endothelial glycocalyx damage and hence reduced pulmonary vascular permeability by inhibiting TNF-a, IL-1β, IL-6, MMP-9, and heparanase expression, attenuating ROS production, and decreasing neutrophil infiltration in BALF (Huang et al., 2018). Further research in the ARDS model indicated that berberine's protective mechanism resulted from inhibition of TLR4/ NF-KB and JAK2/STAT3 signaling pathways (Xu et al., 2021). Cardioprotective effects of berberine, manifested by improved cardiac diastolic function and hemodynamics, were observed in rats with LPS-induced septic cardiomyopathy (Chen et al., 2021). Berberine treatment was also reported to reduce LPS-induced cognitive deficits and restore spatial learning ability in rats. These effects were correlated with increased antioxidant activity, reflected by upregulation of glutathione peroxidase (GPx), SOD, CAT, and glutathione, and decreased acetylcholinesterase (AChE), MDA, carbonyl protein, and caspase-3 activity in the hippocampus (Sadraie et al., 2019). In cultured RAW264.7 macrophages, berberine exposure inhibited LPS-induced synthesis of proinflammatory cytokines (MCP-1, IL-6, and TNF-a). This effect was mediated by reversal of LPS-induced Sirtuin1 (Sirt1) downregulation, which inhibited $I\kappa B\alpha$ degradation and IKKphosphorylation, effectively suppressing NF-kB signaling (Zhang H et al., 2017).

3.4.2 Cordycepin

Cordycepin is the main active component of the fruiting bodies of the ascomycete fungus cordyceps militaris. In a mouse model of LPSinduced ALI, cordycepin administration downregulated the expression of MPO and MDA in lung tissue and reduced TNF- α and IL-1 β levels in BALF. The underlying mechanism was found to be related to inhibition of NF- κ B activity and stimulation of Nrf-2 and HO-1 expression (Lei et al., 2018; Qing et al., 2018). Cordycepin was also reported to decrease LPS-induced pro-inflammatory cytokine production and COX-2 expression in RAW264.7 and THP-1 cells, effects attributed to the inhibition of the NLRP3 inflammasome and the ERK1/2 signaling pathway (Yang J et al., 2017).

3.5 Quinones

Emodin belongs to the Quinones class of compounds and is mainly extracted from the dry roots and rhizome of rhubarb (Hu et al., 2020). According to TCM precepts, emodin is effective in reducing accumulation, cooling blood, reducing fire, promoting blood circulation, removing blood stasis, and draining the gallbladder to relieve jaundice. Emodin was shown to possess a wide range of pharmacological properties, linked to anticancer, hepatoprotective, anti-inflammatory, antioxidant, and antimicrobial effects (Cui et al., 2020). In rats with LPS-induced ALI, emodin

suppressed IKKβ, p-IKKβ, p65, and p-p65 levels, decreased NF-κB DNA binding activity, and inhibited IL-8, IL-1β, TNF-α, and myeloperoxidase (MPO) expression in lung tissues, and increased the proportion of Gr1+/CD11b+ cells in bronchoalveolar lavage fluid (BALF) (Liu B et al., 2020). It was reported that emodin further protected against ALI by downregulating the mechanistic target of rapamycin kinase (mTOR)/HIF-1a/vascular endothelial growth factor (VEGF) signaling pathway (Li X et al., 2020). Granulocytes are the first line of defense against pathogen invasion and play a crucial role in innate immunity. Emodin could upregulate the ability of granulocytes to phagocytize bacteria and generate of neutrophil extracellular trap (NETs), meanwhile, downregulated the production of ROS expression in the LPS-stimulated granulocytes, therefore alleviating lung tissue damage (Mei et al., 2020). Anti-neuroinflammatory effects of emodin were also evidenced by decreased TNF-a, IL-6, nitric oxide (NO), prostaglandin E2 (PGE2), inducible nitric oxide synthase (iNOS), and cyclooxygenase 2 (COX-2) synthesis, inhibition of NF-KB and activation of activator protein-1 (AP-1) signaling pathways, and suppressed phosphorylation of STATs and MAPKs in LPSstimulated microglial cells (Park et al., 2016; Xie et al., 2019). In a mouse model of LPS-induced acute liver injury, emodin treatment alleviated hepatic inflammation and promoted M2 polarization of liver macrophages. Consistent with these findings, emodin exposure downregulated TLR4, MyD88, Toll/interleukin-1 receptor (TIR) domain-containing adaptor protein (TIRAP), TRAF-6, TIRdomain-containing adapter-inducing interferon-β (TRIF), interferon regulatory factor 3 (IRF-3), and AP-1 protein expression in LPS-activated RAW264.7 macrophages (Ding et al., 2018). In a mouse model of LPS-induced septic cardiomyopathy, emodin treatment inhibited expression of cardiac injury markers, i.e., LDH and creatine kinase-MB (CK-MB), and downregulated the expression of inflammatory cytokines. These protective effects were attributed to inhibition of NOD-like receptor family, pyrin domain containing 3 (NLRP3) inflammasome activation in cardiomyocytes (Dai et al., 2021).

4 Acupuncture

Acupuncture is a non-pharmacological TCM method of treating diseases that acts through mechanical stimulation (usually by needling, but also via heat or pressure) on specific skin sites (acupoints) that lie along passageways through which energy flows throughout the body (meridians). In recent years, evidence that acupuncture on specific acupoints, i. e. Zusanli (ST36) and Tianshu (ST25), can regulate immunity suggested that this procedure represents a promising alternative in clinical anti-inflammatory therapy (Kim et al., 2007; Liu S et al., 2020). As a nonpharmacological therapy, and supported by preclinical studies, acupuncture has increasingly attracted the attention of clinicians (Lai et al., 2020; Liu S et al., 2020). Electroacupuncture treatment was shown to improve sepsis-related damage of brain, heart, kidney, intestines, and other organs in animal models. Its application improved survival rate in CLP model rats, and ameliorated cognitive impairment in rats with sepsis-related encephalopathy by inhibiting hippocampal synaptic damage, neuronal loss, oxidative stress, and release of inflammatory cytokines trough activation of the Nrf-2/HO-1 pathway and hippocampal a7 nicotinic acetylcholine receptors (Han et al., 2018; Li C et al., 2020). In a rat model of sepsis,

electroacupuncture pretreatment at ST36 attenuated inflammatory responses, decreased plasma urea, creatinine, and D-lactate levels, and increased intestinal tight junction protein occludin expression and intestinal barrier permeability, thereby reducing acute kidney and intestinal injury (Zhu et al., 2015; Zhang Z et al., 2018; Harpin et al., 2020). In the rat CLP model, electroacupuncture treatment at ST 36 reduced the activity of plasma CK-MB and offered cardioprotection by reducing TNF-a, NO, MPO levels and water content in myocardial tissue by activating the cholinergic anti-inflammatory pathway (Zhang L et al., 2018). In clinical trials, electroacupuncture at both ST36 and Shangjuxu (ST37) significantly reduced procalcitonin (PCT), TNF-a, intestinal fatty acid-binding protein (I-FABP), D-lactate, citrulline, and the TCM quantitative score of intestinal dysfunction in patients with sepsis-related intestinal dysfunction and intestinal obstruction (Meng J.B. et al., 2018). In addition, a prospective randomized controlled trial found that electroacupuncture at both ST 36 and Guanyuan (RN 4) in patients with sepsis played an immunoprotective role by reducing the APACHE II score, increasing CD3⁺, CD4⁺, CD8⁺ expression and the CD4⁺/CD8⁺ ratio, and increasing HLA-DR expression in lymphocytes (Yang et al., 2016).

The above systematically summarized the research and mechanism of TCM in the treatment of sepsis (the relevant mechanisms are shown in Figure 1). Tables 1, 2 respectively summarize the relevant research of TCM compounds and monomers in the treatment of sepsis in recent years, which further prove the effectiveness of TCM.

5 Post-sepsis immune suppression and fuzheng treatment strategy of TCM

There are many concepts recognizing the critical state of patients who are survived from early death, such as chronic critical illness (CCI), compensatory anti-inflammatory response syndrome (CARS), and persistent inflammation, immune suppression, and catabolism syndrome (PICS), among which immune suppression are a common state and has been implicated as a predisposing factor for the secondary nosocomial infections and increased mortality, though the identification of these concepts remains to be further clarified (Torres et al., 2022). Immunotherapy such as granulocyte-macrophage colony stimulating factor (GM-CSF) and granulocyte-colony stimulating factor (G-CSF) aiming to promote restoration of normal lymphocyte numbers and function, and/or restore mature functional myeloid populations achieves great improvement in preventing secondary infection, but failed to demonstrate any significant improvements in 28-day mortality and long-term outcomes (Bo et al., 2011; Darden et al., 2021).

According to the basic theory of TCM, there are two strategies for the treatment of infectious diseases and followed critical state—Quxie and Fuzheng. The function of TCM on inhibiting inflammatory response during the early sepsis refers to the Quxie strategy that means blocking of factors leading to disease. In contrast, Fuzheng strategy agree with the therapy enhancing the anti-disease capacity of the body, which focus on the enhancement of anti-infectious immunity. Innate and adaptive immune cells are both the targets of TCM herbs. Dendritic cells are the main professional antigenpresenting cells, whose function drives the activation of macrophage and antigen-specific T-cells (Lin W et al., 2021). Accumulating evidence shows that the extracts or active monomers of TCM herbs, such as cordyceps sinensis, ganoderma lucidum, astragalus

mongholicus. etc. can act as the adjuvant to promote the maturation, pro-inflammatory cytokine production and function of DCs, therefore enhancing the immune responses against tumor and infection (Li et al., 2015a). It is worth recalling that astragalus, especially the astragalus polysaccharide may be the representative of TCM herbs for immunity enhancement, whose function varies from humoral to cellular immune responses (Sultan et al., 2014; Deng et al., 2022). Though there is a lack of direct evidence, studies about the immune deficiency diseases and cancer suggested the potential function of immune enhancement herbs on sepsis related immune suppression. Except for the direct inhibition of tumor, immunity-enhancing capacity plays a vital role in the anticancer activity of TCM herbs (Wang S et al., 2020; Wang Y et al., 2020) For example, shenqi fuzheng injection, mainly consist of ginsen and astragalus, could promote NK and T helper cells proliferation and function, therefore enhancing the effects of chemotherapy drugs and decreasing adverse events in cancers (Dong et al., 2010; Li et al., 2015b; Yang Y et al., 2017). Results of randomized clinical trials and real-world data reveals that integrating Fuzheng TCM herbs and anti-retroviral therapy promotes long-term reconstitution of the immune system and significantly improved the survival periods of AIDS patients (Zou et al., 2016; Tao et al., 2021; Jin et al., 2022).

6 Discussion and perspectives

Sepsis is a common severe complication of patients with infection, severe trauma, shock, burns, etc. With the development of public health policy, infection-prevention efforts reduce sepsis incidence. Diversified treatment and multiple organ support therapy contribute to the reduced mortality in past years. However, sepsis remains a major cause of health loss worldwide with high health-related burden, especially in Asia and Africa (Rudd et al., 2020). The problems facing the effective antibiotics-based combination therapies are the ever-growing antibiotic resistance (Reynolds et al., 2022) and the potential risk of drug-induced liver injury, enteric dysbacteriosis, and fungal infection (Kwon et al., 2022), which may further aggravate organ dysfunction (Meng et al., 2017). Therefore, more treatment strategy for sepsis is needed.

TCM established a complete system of diagnosis and treatment for pandemic and endemic diseases, possessing a well-documented history of treating infectious diseases and clinical practice, such as Treatise on Febrile and Miscellaneous Disease ("Shanghan Zabing Lun" in Chinese) and Detailed Analysis of Epidemic Warm Diseases ("Wenbing Tiaobian" in Chinese). Given the complexity of the chemical components of traditional Chinese herbs and the ambiguity of the effective components, research has been devoted to the separation of the active components of prescriptions and single traditional Chinese herb for cell or animal study. This review summarizes the function and mechanism of TCM compounds, active monomers (terpenoids, flavonoids, polyphenols, alkaloids), and acupuncture on sepsis treatment, which varies from antiinflammation, anti-oxidation, anti-mitochondrial dysfunction, regulating apoptosis and autophagy etc. Because of the lack of clinical evidence, an important issue that is not covered in this review is whether the potential side effects of TCM on related organs would aggravate the multi-organ injury in patients with sepsis. Indeed, a study reveals that TCM herbs and dietary supplements were the leading causes of drug-induced liver injury (DILI) in mainland China (Shen et al., 2019), which attracts much

attention. However, researchers pointed that implicated drug categories adopted in this study might seriously influence the reliability of conclusions (Cong et al., 2019). A latest study screened 94,593 DILI reports from 308 medical centers across the China mainland between 2012 and 2016, and found that TCM herbs only accounted for 4.5% of the DILI reports (Wang et al., 2022). The main challenge facing the application of TCM for the treatment of sepsis in drug safety aspect is how to identify the scattered categories of drug that are potentially harmful to the organ function, and the assessment of risk/benefit ratio.

In recent years, conventional molecular biological studies, network pharmacology prediction, molecular docking analysis, and visualization analysis reveal the widely potential targets of TCM compounds and active monomers in infectious diseases (Huang et al., 2021; Zhou et al., 2021; Li et al., 2022), and in this state lack of standardized and large-scale clinical studies counts in limiting clinical translation value. In addition, targeting the activation of the immune system and related imbalance of pro-inflammation and antiinflammation is the main mechanism that studies of sepsis focus on, while the protection of organs and prevention of sequelae such as sepsis-associated encephalopathy, ICU-acquired weakness, sepsiscardiomyopathy, induced etc., are partly neglected. Multidisciplinary research is needed to explain the scientific connotation of compound compatibility of anti-sepsis TCM recipes.

Author contributions

YS wrote the manuscript draft. WL revised the manuscript. WZ conceived the review structure.

References

Bai, T., Hu, X., Zheng, Y., Wang, S., Kong, J., and Cai, L. (2016). Resveratrol protects against lipopolysaccharide-induced cardiac dysfunction by enhancing SERCA2a activity through promoting the phospholamban oligomerization. *Am. J. physiology Heart circulatory physiology* 311, H1051-H1062. doi:10.1152/ajpheart.00296.2016

Bo, L., Wang, F., Zhu, J., Li, J., and Deng, X. (2011). Granulocyte-colony stimulating factor (G-CSF) and granulocyte-macrophage colony stimulating factor (GM-CSF) for sepsis: A meta-analysis. *Crit. care (London, Engl.* 15, R58. doi:10.1186/cc10031

Boyd, J. H., Russell, J. A., and Fjell, C. D. (2014). The meta-genome of sepsis: Host genetics, pathogens and the acute immune response. *J. innate Immun.* 6, 272–283. doi:10. 1159/000358835

Cao, Y., Han, X., Pan, H., Jiang, Y., Peng, X., Xiao, W., et al. (2020). Emerging protective roles of shengmai injection in septic cardiomyopathy in mice by inducing myocardial mitochondrial autophagy via caspase-3/Beclin-1 axis. *Inflamm. Res. official J. Eur. Histamine Res. Soc.* 69, 41–50. doi:10.1007/s00011-019-01292-2

Cecconi, M., Evans, L., Levy, M., and Rhodes, A. (2018). Sepsis and septic shock. Lancet London, Engl. 392, 75–87. doi:10.1016/S0140-6736(18)30696-2

Chen, G., Xu, Y., Jing, J., Mackie, B., Zheng, X., Zhang, X., et al. (2017). The anti-sepsis activity of the components of Huanglian Jiedu Decoction with high lipid A-binding affinity. *Int. Immunopharmacol.* 46, 87–96. doi:10.1016/j.intimp.2017.02.025

Chen, H., Liu, Q., Liu, X., and Jin, J. (2021). Berberine attenuates septic cardiomyopathy by inhibiting TLR4/NF- κ B signalling in rats. *Pharm. Biol.* 59, 121–128. doi:10.1080/13880209.2021.1877736

Chen, H., Zhang, Y., Zhang, W., Liu, H., Sun, C., Zhang, B., et al. (2019). Inhibition of myeloid differentiation factor 2 by baicalein protects against acute lung injury. *Phytomedicine Int. J. phytotherapy Phytopharm.* 63, 152997. doi:10.1016/j.phymed. 2019.152997

Chen, L., Liu, P., Feng, X., and Ma, C. (2017). Salidroside suppressing LPS-induced myocardial injury by inhibiting ROS-mediated PI3K/Akt/mTOR pathway *in vitro* and *in vivo. J. Cell. Mol. Med.* 21, 3178–3189. doi:10.1111/jcmm.12871

Chen, Q., Liu, J., Wang, W., Liu, S., Yang, X., Chen, M., et al. (2019). Sini decoction ameliorates sepsis-induced acute lung injury via regulating ACE2-Ang (1-7)-Mas axis and

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Conflict of interest

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inhibiting the MAPK signaling pathway. Biomed. Pharmacother. = Biomedecine Pharmacother. 115, 108971. doi:10.1016/j.biopha.2019.108971

Chen, R. J., Rui, Q. L., Wang, Q., Tian, F., Wu, J., and Kong, X. Q. (2020). Shenfu injection attenuates lipopolysaccharide-induced myocardial inflammation and apoptosis in rats. *Chin. J. Nat. Med.* 18, 226–233. doi:10.1016/S1875-5364(20)30025-X

Chen, X., Feng, Y., Shen, X., Pan, G., Fan, G., Gao, X., et al. (2018). Anti-sepsis protection of Xuebijing injection is mediated by differential regulation of pro- and anti-inflammatory Th17 and T regulatory cells in a murine model of polymicrobial sepsis. *J. Ethnopharmacol.* 211, 358–365. doi:10.1016/j.jep.2017.10.001

Cong, W., Xin, Q., and Gao, Y. (2019). RE: Incidence and etiology of drug-induced liver injury in mainland China. *Gastroenterology* 157, 1438–1439. doi:10.1053/j.gastro.2019. 05.076

Cui, Y., Chen, L. J., Huang, T., Ying, J. Q., and Li, J. (2020). The pharmacology, toxicology and therapeutic potential of anthraquinone derivative emodin. *Chin. J. Nat. Med.* 18, 425–435. doi:10.1016/S1875-5364(20)30050-9

Dai, S., Ye, B., Chen, L., Hong, G., Zhao, G., and Lu, Z. (2021). Emodin alleviates LPSinduced myocardial injury through inhibition of NLRP3 inflammasome activation. *Phytotherapy Res.* 35 (9), 5203–5213. doi:10.1002/ptr.7191

Darden, D. B., Kelly, L. S., Fenner, B. P., Moldawer, L. L., Mohr, A. M., Efron, P. A., et al. (2021). Chronic critical illness elicits a unique circulating leukocyte transcriptome in sepsis survivors. *J. Clin. Med.* 10, 3211. doi:10.3390/jcm10153211

Delano, M. J., and Ward, P. A. (2016). The immune system's role in sepsis progression, resolution, and long-term outcome. *Immunol. Rev.* 274, 330–353. doi:10.1111/imr.12499

Deng, G., Zhou, L., Wang, B., Sun, X., Zhang, Q., Chen, H., et al. (2022). Targeting cathepsin B by cycloastragenol enhances antitumor immunity of CD8 T cells via inhibiting MHC-I degradation. *J. Immunother. cancer* 10, e004874. doi:10.1136/jitc-2022-004874

Ding, Y., Liu, P., Chen, Z. L., Zhang, S. J., Wang, Y. Q., Cai, X., et al. (2018). Emodin attenuates lipopolysaccharide-induced acute liver injury via inhibiting the TLR4 signaling pathway *in vitro* and *in vivo*. *Front. Pharmacol.* 9, 962. doi:10.3389/fphar.2018.00962

Dong, J., Su, S. Y., Wang, M. Y., and Zhan, Z. (2010). Shenqi fuzheng, an injection concocted from Chinese medicinal herbs, combined with platinum-based chemotherapy

for advanced non-small cell lung cancer: A systematic review. J. Exp. Clin. cancer Res. CR 29, 137. doi:10.1186/1756-9966-29-137

Dourado, N. S., Souza, C. D. S., de Almeida, M. M. A., Bispo da Silva, A., Dos Santos, B. L., Silva, V. D. A., et al. (2020). Neuroimmunomodulatory and neuroprotective effects of the flavonoid apigenin in *in vitro* models of neuroinflammation associated with alzheimer's disease. *Front. aging Neurosci.* 12, 119. doi:10.3389/fnagi.2020.00119

Fisher, C. J., Jr., Agosti, J. M., Opal, S. M., Lowry, S. F., Balk, R. A., Sadoff, J. C., et al. (1996). Treatment of septic shock with the tumor necrosis factor receptor:Fc fusion protein. The Soluble TNF Receptor Sepsis Study Group. *N. Engl. J. Med.* 334, 1697–1702. doi:10.1056/NEJM199606273342603

Fleischmann, C., Scherag, A., Adhikari, N. K., Hartog, C. S., Tsaganos, T., Schlattmann, P., et al. (2016). Assessment of global incidence and mortality of hospital-treated sepsis. Current estimates and limitations. *Am. J. Respir. Crit. care Med.* 193, 259–272. doi:10. 1164/rccm.201504-0781OC

Fu, J., Zang, Y., Zhou, Y., Chen, C., Shao, S., Hu, M., et al. (2020). A novel triptolide derivative ZT01 exerts anti-inflammatory effects by targeting TAK1 to prevent macrophage polarization into pro-inflammatory phenotype. *Biomed. Pharmacother.* = *Biomedecine Pharmacother.* 126, 110084. doi:10.1016/j.biopha.2020.110084

Gao, H., Liu, X., Sun, W., Kang, N., Liu, Y., Yang, S., et al. (2017). Total tanshinones exhibits anti-inflammatory effects through blocking TLR4 dimerization via the MyD88 pathway. *Cell death Dis.* 8, e3004. doi:10.1038/cddis.2017.389

Gao, S., Li, L., Li, L., Ni, J., Guo, R., Mao, J., et al. (2019). Effects of the combination of tanshinone IIA and puerarin on cardiac function and inflammatory response in myocardial ischemia mice. *J. Mol. Cell. Cardiol.* 137, 59–70. doi:10.1016/j.yjmcc.2019. 09.012

Han, Y. G., Qin, X., Zhang, T., Lei, M., Sun, F. Y., Sun, J. J., et al. (2018). Electroacupuncture prevents cognitive impairment induced by lipopolysaccharide via inhibition of oxidative stress and neuroinflammation. *Neurosci. Lett.* 683, 190–195. doi:10. 1016/j.neulet.2018.06.003

Harpin, D., Simadibrata, C. L., Mihardja, H., and Barasila, A. C. (2020). Effect of electroacupuncture on urea and creatinine levels in the wistar sepsis model. *Med. Acupunct.* 32, 29–37. doi:10.1089/acu.2019.1369

He, Y., Yuan, X., Zuo, H., Sun, Y., and Feng, A. (2018). Berberine exerts a protective effect on gut-vascular barrier via the modulation of the wnt/beta-catenin signaling pathway during sepsis. *Cell. physiology Biochem. Int. J. Exp. Cell. physiology, Biochem. Pharmacol.* 49, 1342–1351. doi:10.1159/000493412

Hong, M. K., Hu, L. L., Zhang, Y. X., Xu, Y. L., Liu, X. Y., He, P. K., et al. (2020). 6-Gingerol ameliorates sepsis-induced liver injury through the Nrf2 pathway. *Int. Immunopharmacol.* 80, 106196. doi:10.1016/j.intimp.2020.106196

Hu, N., Liu, J., Xue, X., and Li, Y. (2020). The effect of emodin on liver disease -comprehensive advances in molecular mechanisms. *Eur. J. Pharmacol.* 882, 173269. doi:10.1016/j.ejphar.2020.173269

Hu, X., Liu, S., Zhu, J., and Ni, H. (2019). Dachengqi decoction alleviates acute lung injury and inhibits inflammatory cytokines production through TLR4/NF- κ B signaling pathway *in vivo* and *in vitro*. J. Cell. Biochem. 120, 8956–8964. doi:10.1002/jcb.27615

Huang, K., Zhang, P., Zhang, Z., Youn, J. Y., Wang, C., Zhang, H., et al. (2021). Traditional Chinese Medicine (TCM) in the treatment of COVID-19 and other viral infections: Efficacies and mechanisms. *Pharmacol. Ther.* 225, 107843. doi:10.1016/j. pharmthera.2021.107843

Huang, L., Zhang, X., Ma, X., Zhang, D., Li, D., Feng, J., et al. (2018). Berberine alleviates endothelial glycocalyx degradation and promotes glycocalyx restoration in LPS-induced ARDS. *Int. Immunopharmacol.* 65, 96–107. doi:10.1016/j.intimp.2018.10.001

Huang, M., Cai, S., and Su, J. (2019). The pathogenesis of sepsis and potential therapeutic targets. Int. J. Mol. Sci. 20, 5376. doi:10.3390/ijms20215376

Huang, P., Lu, X., Yuan, B., Liu, T., Dai, L., Liu, Y., et al. (2016). Astragaloside IV alleviates E. coli-caused peritonitis via upregulation of neutrophil influx to the site of infection. *Int. Immunopharmacol.* 39, 377–382. doi:10.1016/j.intimp.2016.08.011

Huang, R., and Li, M. (2016). Protective effect of Astragaloside IV against sepsis-induced acute lung injury in rats. *Saudi Pharm. J.* 24, 341–347. doi:10.1016/j.jsps.2016.04.014

Ji, Q., Sun, Z., Yang, Z., Zhang, W., Ren, Y., Chen, W., et al. (2021). Protective effect of ginsenoside Rg1 on LPS-induced apoptosis of lung epithelial cells. *Mol. Immunol.* 136, 168–174. doi:10.1016/j.molimm.2018.11.003

Jin, H., Peng, X., He, Y., Ruganzu, J. B., and Yang, W. (2020). Tanshinone IIA suppresses lipopolysaccharide-induced neuroinflammatory responses through NF-kB/MAPKs signaling pathways in human U87 astrocytoma cells. *Brain Res. Bull.* 164, 136–145. doi:10.1016/j.brainresbull.2020.08.019

Jin, Y., Zhang, M., Ma, Y., Sang, F., Li, P., Yang, C., et al. (2022). Effects of Chinese medicine on the survival of AIDS patients administered second-line art in rural areas of China: A retrospective cohort study based on real-world data. *Evidence-based complementary Altern. Med. eCAM* 2022, 5103768. doi:10.1155/2022/5103768

Jingyan, L., Yujuan, G., Yiming, Y., Lingpeng, Z., Tianhua, Y., and Mingxing, M. (2017). Salidroside attenuates LPS-induced acute lung injury in rats. *Inflammation* 40, 1520–1531. doi:10.1007/s10753-017-0593-6

Kang, X., Jin, D., Jiang, L., Zhang, Y., Zhang, Y., An, X., et al. (2022). Efficacy and mechanisms of traditional Chinese medicine for COVID-19: A systematic review. *Chin. Med.* 17, 30. doi:10.1186/s13020-022-00587-7

Kim, H. W., Kang, S. Y., Yoon, S. Y., Roh, D. H., Kwon, Y. B., Han, H. J., et al. (2007). Low-frequency electroacupuncture suppresses zymosan-induced peripheral inflammation via activation of sympathetic post-ganglionic neurons. *Brain Res.* 1148, 69–75. doi:10. 1016/j.brainres.2007.02.030

Kwon, J., Kong, Y., Wade, M., Williams, D. J., Creech, C. B., Evans, S., et al. (2022). Gastrointestinal microbiome disruption and antibiotic-associated diarrhea in children receiving antibiotic therapy for community-acquired pneumonia. J. Infect. Dis. 226, 1109–1119. doi:10.1093/infdis/jiac082

Lai, F., Ren, Y., Lai, C., Chen, R., Yin, X., Tan, C., et al. (2020). Acupuncture at zusanli (ST36) for experimental sepsis: A systematic review. *Evidence-based complementary Altern. Med. eCAM* 2020, 3620741. doi:10.1155/2020/3620741

Lan, K. C., Chao, S. C., Wu, H. Y., Chiang, C. L., Wang, C. C., Liu, S. H., et al. (2017). Salidroside ameliorates sepsis-induced acute lung injury and mortality via downregulating NF-κB and HMGB1 pathways through the upregulation of SIRT1. *Sci. Rep.* 7, 12026. doi:10.1038/s41598-017-12285-8

Lei, J., Wei, Y., Song, P., Li, Y., Zhang, T., Feng, Q., et al. (2018). Cordycepin inhibits LPS-induced acute lung injury by inhibiting inflammation and oxidative stress. *Eur. J. Pharmacol.* 818, 110–114. doi:10.1016/j.ejphar.2017.10.029

Li, C., Wang, P., Li, M., Zheng, R., Chen, S., Liu, S., et al. (2021). The current evidence for the treatment of sepsis with Xuebijing injection: Bioactive constituents, findings of clinical studies and potential mechanisms. *J. Ethnopharmacol.* 265, 113301. doi:10.1016/j.jep.2020. 113301

Li, C., Wang, P., Zhang, L., Li, M., Lei, X., Liu, S., et al. (2018). Efficacy and safety of xuebijing injection (a Chinese patent) for sepsis: A meta-analysis of randomized controlled trials. J. Ethnopharmacol. 224, 512–521. doi:10.1016/j.jep.2018.05.043

Li, C., Yu, T. Y., Zhang, Y., Wei, L. P., Dong, S. A., Shi, J., et al. (2020). Electroacupuncture improves cognition in rats with sepsis-associated encephalopathy. *J. Surg. Res.* 256, 258–266. doi:10.1016/j.jss.2020.06.056

Li, F., Lang, F., Zhang, H., Xu, L., Wang, Y., Zhai, C., et al. (2017). Apigenin alleviates endotoxin-induced myocardial toxicity by modulating inflammation, oxidative stress, and autophagy. *Oxidative Med. Cell. Longev.* 2017, 2302896. doi:10.1155/2017/2302896

Li, H., Wang, P., Huang, F., Jin, J., Wu, H., Zhang, B., et al. (2018). Astragaloside IV protects blood-brain barrier integrity from LPS-induced disruption via activating Nrf2 antioxidant signaling pathway in mice. *Toxicol. Appl. Pharmacol.* 340, 58–66. doi:10.1016/j.taap.2017.12.019

Li, J., Li, J., and Zhang, F. (2015a). The immunoregulatory effects of Chinese herbal medicine on the maturation and function of dendritic cells. *J. Ethnopharmacol.* 171, 184–195. doi:10.1016/j.jep.2015.05.050

Li, J., Wang, J. C., Ma, B., Gao, W., Chen, P., Sun, R., et al. (2015b). Shenqi fuzheng injection for advanced gastric cancer: A systematic review of randomized controlled trials. *Chin. J. Integr. Med.* 21, 71–79. doi:10.1007/s11655-014-1768-8

Li, J., Zheng, Y., Li, M. X., Yang, C. W., and Liu, Y. F. (2018). Tanshinone IIA alleviates lipopolysaccharide-induced acute lung injury by downregulating TRPM7 and pro-inflammatory factors. *J. Cell. Mol. Med.* 22, 646–654. doi:10.1111/jcmm.13350

Li, T., Qian, Y., Miao, Z., Zheng, P., Shi, T., Jiang, X., et al. (2020). Xuebijing injection alleviates Pam3CSK4-induced inflammatory response and protects mice from sepsis caused by methicillin-resistant *Staphylococcus aureus*. *Front. Pharmacol.* 11, 104. doi:10.3389/fphar.2020.00104

Li, X., Shan, C., Wu, Z., Yu, H., Yang, A., and Tan, B. (2020). Emodin alleviated pulmonary inflammation in rats with LPS-induced acute lung injury through inhibiting the mTOR/HIF-1a/VEGF signaling pathway. *Inflamm. Res. official J. Eur. Histamine Res. Soc. [et al]* 69, 365–373. doi:10.1007/s00011-020-01331-3

Li, X., Wei, S., Niu, S., Ma, X., Li, H., Jing, M., et al. (2022). Network pharmacology prediction and molecular docking-based strategy to explore the potential mechanism of Huanglian Jiedu Decoction against sepsis. *Computers in biology and medicine* 144, 105389. doi:10.1016/j.compbiomed.2022.105389

Li, Y., Ruan, X., Xu, X., Li, C., Qiang, T., Zhou, H., et al. (2019). Shengmai injection suppresses angiotensin II-induced cardiomyocyte hypertrophy and apoptosis via activation of the AMPK signaling pathway through energy-dependent mechanisms. *Frontiers in pharmacology* 10, 1095. doi:10.3389/fphar.2019.01095

Li, Y., Wang, F., and Luo, Y. (2017). Ginsenoside Rg1 protects against sepsis-associated encephalopathy through beclin 1-independent autophagy in mice. *The Journal of surgical research* 207, 181–189. doi:10.1016/j.jss.2016.08.080

Li, Y., Zhou, J., Qiu, J., Huang, Z., Wang, W., Wu, P., et al. (2020). Berberine reduces gutvascular barrier permeability via modulation of ApoM/S1P pathway in a model of polymicrobial sepsis. *Life sciences* 261, 118460. doi:10.1016/j.lfs.2020.118460

Lin, S. P., Wei, J. X., Hu, J. S., Bu, J. Y., Zhu, L. D., Li, Q., et al. (2021). Artemisinin improves neurocognitive deficits associated with sepsis by activating the AMPK axis in microglia. *Acta pharmacologica Sinica* 42, 1069–1079. doi:10.1038/s41401-021-00634-3

Lin, W., Shen, P., Song, Y., Huang, Y., and Tu, S. (2021). Reactive oxygen species in autoimmune cells: Function, differentiation, and metabolism. *Frontiers in immunology* 12, 635021. doi:10.3389/fimmu.2021.635021

Liu, B., Cheng, Y., Wu, Y., Zheng, X., Li, X., Yang, G., et al. (2020). Emodin improves alveolar hypercoagulation and inhibits pulmonary inflammation in LPS-provoked ARDS in mice via NF-kB inactivation. *International immunopharmacology* 88, 107020. doi:10. 1016/j.intimp.2020.107020

Liu, J., Wang, Z., Lin, J., Li, T., Guo, X., Pang, R., et al. (2021). Xuebijing injection in septic rats mitigates kidney injury, reduces cortical microcirculatory disorders, and suppresses activation of local inflammation. *Journal of ethnopharmacology* 276, 114199. doi:10.1016/j.jep.2021.114199

Liu, Q. Y., Zhuang, Y., Song, X. R., Niu, Q., Sun, Q. S., Li, X. N., et al. (2021). Tanshinone IIA prevents LPS-induced inflammatory responses in mice via inactivation of succinate dehydrogenase in macrophages. *Acta pharmacologica Sinica* 42, 987–997. doi:10.1038/s41401-020-00535-x

Liu, S., Wang, Z. F., Su, Y. S., Ray, R. S., Jing, X. H., Wang, Y. Q., et al. (2020). Somatotopic organization and intensity dependence in driving distinct NPY-expressing sympathetic pathways by electroacupuncture. *Neuron* 108, 436–450. e437. doi:10.1016/j. neuron.2020.07.015

Liu, X., Liu, R., Dai, Z., Wu, H., Lin, M., Tian, F., et al. (2019). Effect of Shenfu injection on lipopolysaccharide (LPS)-induced septic shock in rabbits. *Journal of ethnopharmacology* 234, 36–43. doi:10.1016/j.jep.2019.01.008

Liu, Y., Li, Z., Xue, X., Wang, Y., Zhang, Y., and Wang, J. (2018). Apigenin reverses lung injury and immunotoxicity in paraquat-treated mice. *International immunopharmacology* 65, 531–538. doi:10.1016/j.intimp.2018.10.046

Lu, J., Yan, J., Yan, J., Zhang, L., Chen, M., Chen, Q., et al. (2020a). Network pharmacology based research into the effect and mechanism of Xijiao Dihuang decoction against sepsis. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* 122, 109777. doi:10.1016/j.biopha.2019.109777

Lu, J., Zhang, L., Cheng, L., He, S., Zhang, Y., Yan, J., et al. (2020b). Xijiao Dihuang decoction improves prognosis of sepsis via inhibition of aerobic glycolysis. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* 129, 110501. doi:10.1016/j.biopha. 2020.110501

Lu, Z. B., Ou, J. Y., Cao, H. H., Liu, J. S., and Yu, L. Z. (2020). Heat-clearing Chinese medicines in lipopolysaccharide-induced inflammation. *Chinese journal of integrative medicine* 26, 552–559. doi:10.1007/s11655-020-3256-7

Lu, Z., Xiong, W., Xiao, S., Lin, Y., Yu, K., Yue, G., et al. (2020). Huanglian Jiedu Decoction ameliorates DSS-induced colitis in mice via the JAK2/STAT3 signalling pathway. *Chinese medicine* 15, 45. doi:10.1186/s13020-020-00327-9

Luo, M., Yan, D., Sun, Q., Tao, J., Xu, L., Sun, H., et al. (2020). Ginsenoside Rg1 attenuates cardiomyocyte apoptosis and inflammation via the TLR4/NF-kB/ NLRP3 pathway. *Journal of cellular biochemistry* 121, 2994–3004. doi:10.1002/jcb. 29556

Luo, X., Yu, Z., Deng, C., Zhang, J., Ren, G., Sun, A., et al. (2017). Baicalein ameliorates TNBS-induced colitis by suppressing TLR4/MyD88 signaling cascade and NLRP3 inflammasome activation in mice. *Scientific reports* 7, 16374. doi:10.1038/ s41598-017-12562-6

Mei, H., Tao, Y., Zhang, T., and Qi, F. (2020). Emodin alleviates LPS-induced inflammatory response in lung injury rat by affecting the function of granulocytes. *Journal of inflammation (London, England)* 17, 26. doi:10.1186/s12950-020-00252-6

Mei, L., He, M., Zhang, C., Miao, J., Wen, Q., Liu, X., et al. (2019). Paeonol attenuates inflammation by targeting HMGB1 through upregulating miR-339-5p. *Scientific reports* 9, 19370. doi:10.1038/s41598-019-55980-4

Meng, F., Lai, H., Luo, Z., Liu, Y., Huang, X., Chen, J., et al. (2018). Effect of xuefu zhuyu decoction pretreatment on myocardium in sepsis rats. *eCAM* 2018, 2939307. doi:10.1155/2018/2939307

Meng, J. B., Jiao, Y. N., Zhang, G., Xu, X. J., Ji, C. L., Hu, M. H., et al. (2018). Electroacupuncture improves intestinal dysfunction in septic patients: A randomised controlled trial. *BioMed research international* 2018, 8293594. doi:10.1155/2018/8293594

Meng, M., Klingensmith, N. J., and Coopersmith, C. M. (2017). New insights into the gut as the driver of critical illness and organ failure. *Current opinion in critical care* 23, 143–148. doi:10.1097/MCC.00000000000386

Miao, J., Ye, P., Lan, J., Ye, S., Zhong, J., Gresham, A., et al. (2020). Paeonol promotes the phagocytic ability of macrophages through confining HMGB1 to the nucleus. *International immunopharmacology* 89, 107068. doi:10.1016/j.intimp.2020.107068

Mo, Y., Liu, X., Qin, X., Huang, J., He, Z., Lin, J., et al. (2014). Evidence-based complementary and alternative medicine, 2014, 279853.Shenfu injection for intradialytic hypotension a systematic review and meta-analysiseCAM

Nabavi, S. F., Khan, H., D'Onofrio, G., Šamec, D., Shirooie, S., Dehpour, A. R., et al. (2018). Apigenin as neuroprotective agent: Of mice and men. *Pharmacological research* 128, 359–365. doi:10.1016/j.phrs.2017.10.008

Opal, S. M., Laterre, P. F., Francois, B., LaRosa, S. P., Angus, D. C., Mira, J. P., et al. (2013). Effect of eritoran, an antagonist of MD2-TLR4, on mortality in patients with severe sepsis: The ACCESS randomized trial. *Jama* 309, 1154–1162. doi:10.1001/jama.2013.2194

Pan, L. Y., Chen, Y. F., Li, H. C., Bi, L. M., Sun, W. J., Sun, G. F., et al. (2017). Dachengqi decoction attenuates intestinal vascular endothelial injury in severe acute pancreatitis in vitro and in vivo. Cellular physiology and biochemistry international journal of experimental cellular physiology, biochemistry, and pharmacology 44, 2395–2406. doi:10.1159/000486155

Pang, Q., Zhao, Y., Chen, X., Zhao, K., Zhai, Q., and Tu, F. (2018). Apigenin protects the brain against ischemia/reperfusion injury via caveolin-1/VEGF *in vitro* and *in vivo*. *Oxidative medicine and cellular longevity* 2018, 7017204. doi:10.1155/2018/7017204

Park, S. Y., Jin, M. L., Ko, M. J., Park, G., and Choi, Y. W. (2016). Antineuroinflammatory effect of emodin in LPS-stimulated microglia: Involvement of AMPK/Nrf2 activation. Neurochemical research 41, 2981-2992. doi:10.1007/s11064-016-2018-6

Prescott, H. C., and Angus, D. C. (2018). Enhancing recovery from sepsis: A review. Jama 319, 62-75. doi:10.1001/jama.2017.17687

Qi, Z., Zhang, Y., Qi, S., Ling, L., Gui, L., Yan, L., et al. (2017). Salidroside inhibits HMGB1 acetylation and release through upregulation of SirT1 during inflammation. *Oxidative medicine and cellular longevity* 2017, 9821543. doi:10.1155/2017/9821543

Qing, R., Huang, Z., Tang, Y., Xiang, Q., and Yang, F. (2018). Cordycepin alleviates lipopolysaccharide-induced acute lung injury via Nrf2/HO-1 pathway. *International immunopharmacology* 60, 18–25. doi:10.1016/j.intimp.2018.04.032

Reynolds, D., Burnham, J. P., Vazquez Guillamet, C., McCabe, M., Yuenger, V., Betthauser, K., et al. (2022). The threat of multidrug-resistant/extensively drugresistant gram-negative respiratory infections: Another pandemic. *European respiratory review an official journal of the European Respiratory Society* 31, 220068. doi:10.1183/ 16000617.0068-2022

Rodrigues, F. A. P., Santos, A., de Medeiros, P., Prata, M. M. G., Santos, T. C. S., da Silva, J. A., et al. (2018). Gingerol suppresses sepsis-induced acute kidney injury by modulating methylsulfonylmethane and dimethylamine production. *Scientific reports* 8, 12154. doi:10. 1038/s41598-018-30522-6

Rudd, K. E., Johnson, S. C., Agesa, K. M., Shackelford, K. A., Tsoi, D., Kievlan, D. R., et al. (2020). Global, regional, and national sepsis incidence and mortality, 1990-2017: Analysis for the global burden of disease study. *Lancet (London, England)* 395, 200–211. doi:10. 1016/S0140-6736(19)32989-7

Sadraie, S., Kiasalari, Z., Razavian, M., Azimi, S., Sedighnejad, L., Afshin-Majd, S., et al. (2019). Berberine ameliorates lipopolysaccharide-induced learning and memory deficit in the rat: Insights into underlying molecular mechanisms. *Metabolic Brain disease* 34, 245–255. doi:10.1007/s11011-018-0349-5

Seo, E. H., Song, G. Y., Kwak, B. O., Oh, C. S., Lee, S. H., and Kim, S. H. (2017). Effects of glycyrrhizin on the differentiation of myeloid cells of the heart and lungs in lipopolysaccharide-induced septic mice. *Shock (Augusta, Ga)* 48, 371–376. doi:10.1097/SHK.00000000000850

Shang, S., Wu, J., Li, X., Liu, X., Li, P., Zheng, C., et al. (2020). Artesunate interacts with the vitamin D receptor to reverse sepsis-induced immunosuppression in a mouse model via enhancing autophagy. *British journal of pharmacology* 177, 4147–4165. doi:10.1111/ bph.15158

Shen, C. H., Ma, Z. Y., Li, J. H., Li, R. D., Tao, Y. F., Zhang, Q. B., et al. (2020). Glycyrrhizin improves inflammation and apoptosis via suppressing HMGB1 and PI3K/ mTOR pathway in lipopolysaccharide-induced acute liver injury. *European review for medical and pharmacological sciences* 24, 7122–7130. doi:10.26355/eurrev_202006_21706

Shen, T., Liu, Y., Shang, J., Xie, Q., Li, J., Yan, M., et al. (2019). Incidence and etiology of drug-induced liver injury in mainland China. *Gastroenterology* 156, 2230–2241. e2211. doi:10.1053/j.gastro.2019.02.002

Singer, M., Deutschman, C. S., Seymour, C. W., Shankar-Hari, M., Annane, D., Bauer, M., et al. (2016). The third international consensus definitions for sepsis and septic shock (Sepsis-3). *Jama* 315, 801–810. doi:10.1001/jama.2016.0287

Song, C., Wang, Y., Cui, L., Yan, F., and Shen, S. (2019). Triptolide attenuates lipopolysaccharide-induced inflammatory responses in human endothelial cells: Involvement of NF-kB pathway. *BMC complementary and alternative medicine* 19, 198. doi:10.1186/s12096-019-2616-3

Song, M. T., Ruan, J., Zhang, R. Y., Deng, J., Ma, Z. Q., and Ma, S. P. (2018). Astragaloside IV ameliorates neuroinflammation-induced depressive-like behaviors in mice via the PPARγ/NF-κB/NLRP3 inflammasome axis. *Acta pharmacologica Sinica* 39, 1559–1570. doi:10.1038/aps.2017.208

Song, Y., Yao, C., Yao, Y., Han, H., Zhao, X., Yu, K., et al. (2019). XueBiJing injection versus placebo for critically ill patients with severe community-acquired pneumonia: A randomized controlled trial. *Critical care medicine* 47, e735–e743. doi:10.1097/CCM. 000000000003842

Su, Z., Yu, P., Sheng, L., Ye, J., and Qin, Z. (2018). Fangjifuling ameliorates lipopolysaccharide-induced renal injury via inhibition of inflammatory and apoptotic response in mice. Cellular physiology and biochemistry international journal of experimental cellular physiology, biochemistry, and pharmacology 49, 2124–2137. doi:10.1159/000493816

Sultan, M. T., Butt, M. S., Qayyum, M. M., and Suleria, H. A. (2014). Immunity: Plants as effective mediators. *Critical reviews in food science and nutrition* 54, 1298–1308. doi:10. 1080/10408398.2011.633249

Sun, J. N., Sun, W. Y., and Dong, S. F. (2017). Discussion on efficacy evaluation thought and method for innovation medicine of Chinese herbal compound formula based on clinical application characteristics. *Zhongguo Zhong yao za zhi = Zhongguo zhongyao zazhi = China journal of Chinese materia medica* 42, 852–855. doi:10.19540/j.cnki.cjcmm. 20170103.023

Sun, W., Chen, Y., Li, H., Liu, H., Li, J., Chen, J., et al. (2020). Material basis and molecular mechanisms of Dachengqi decoction in the treatment of acute pancreatitis based on network pharmacology. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* 121, 109656. doi:10.1016/j.biopha.2019.109656

Tao, Z., Huang, X. J., Liu, Y., Wang, R., Dong, J. P., Liang, B. Y., et al. (2021). Efficacy of integrated traditional Chinese medicine and anti-retroviral therapy on immunological nonresponse in patients with human immunodeficiency virus/acquired immunodeficiency syndrome: A meta-analysis of randomized controlled trial. *Journal of traditional Chinese*

medicine = Chung i tsa chih ying wen pan 41, 669–676. doi:10.19852/j.cnki.jtcm.2021. 05.002

Torres, L. K., Pickkers, P., and van der Poll, T. (2022). Sepsis-induced immunosuppression. *Annual review of physiology* 84, 157–181. doi:10.1146/annurev-physiol-061121-040214

Wang, B., Bellot, G. L., Iskandar, K., Chong, T. W., Goh, F. Y., Tai, J. J., et al. (2020). Resveratrol attenuates TLR-4 mediated inflammation and elicits therapeutic potential in models of sepsis. *Scientific reports* 10, 18837. doi:10.1038/s41598-020-74578-9

Wang, J., Song, H., Ge, F., Xiong, P., Jing, J., He, T., et al. (2022). Landscape of DILIrelated adverse drug reaction in China Mainland. *Acta Pharmaceutica Sinica B* 12, 4424–4431. doi:10.1016/j.apsb.2022.04.019

Wang, N., Mao, L., Yang, L., Zou, J., Liu, K., Liu, M., et al. (2017). Resveratrol protects against early polymicrobial sepsis-induced acute kidney injury through inhibiting endoplasmic reticulum stress-activated NF-KB pathway. *Oncotarget* 8, 36449–36461. doi:10.18632/oncotarget.16860

Wang, N., Zhang, X., Ma, Z., Niu, J., Ma, S., Wenjie, W., et al. (2020). Combination of tanshinone IIA and astragaloside IV attenuate atherosclerotic plaque vulnerability in ApoE(-/-) mice by activating PI3K/AKT signaling and suppressing TRL4/NF- κ B signaling. *Biomedicine & pharmacotherapy* = *Biomedecine & pharmacotherapie* 123, 109729. doi:10.1016/j.biopha.2019.109729

Wang, Q. L., Yang, L., Peng, Y., Gao, M., Yang, M. S., Xing, W., et al. (2019). Ginsenoside Rg1 regulates SIRT1 to ameliorate sepsis-induced lung inflammation and injury via inhibiting endoplasmic reticulum stress and inflammation. *Mediators of inflammation* 2019, 6453296. doi:10.1155/2019/6453296

Wang, Q., Yang, X., Song, Y., Sun, X., Li, W., Zhang, L., et al. (2021). Astragaloside IVtargeting miRNA-1 attenuates lipopolysaccharide-induced cardiac dysfunction in rats through inhibition of apoptosis and autophagy. *Life sciences* 275, 119414. doi:10.1016/j.lfs. 2021.119414

Wang, S., Long, S., Deng, Z., and Wu, W. (2020). Positive role of Chinese herbal medicine in cancer immune regulation. *The American journal of Chinese medicine* 48, 1577–1592. doi:10.1142/S0192415X20500780

Wang, X., Zhao, Z., Mao, J., Du, T., Chen, Y., Xu, H., et al. (2019). Randomized, doubleblinded, multicenter, placebo-controlled trial of shenfu injection for treatment of patients with chronic heart failure during the acute phase of symptom aggravation (Yang and qi deficiency syndrome). *Evidence-based complementary and alternative medicine eCAM* 2019, 9297163. doi:10.1155/2019/9297163

Wang, Y., Zhang, Q., Chen, Y., Liang, C. L., Liu, H., Qiu, F., et al. (2020). Antitumor effects of immunity-enhancing traditional Chinese medicine. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* 121, 109570. doi:10.1016/j. biopha.2019.109570

Wen-Ting, S., Fa-Feng, C., Li, X., Cheng-Ren, L., and Jian-Xun, L. (2012). Chinese medicine shenfu injection for heart failure: A systematic review and meta-analysis. Evidence-Based complementary and alternative medicine. *eCAM* 2012, 713149.

Xia, M. L., Xie, X. H., Ding, J. H., Du, R. H., and Hu, G. (2020). Astragaloside IV inhibits astrocyte senescence: Implication in Parkinson's disease. *Journal of neuroinflammation* 17, 105. doi:10.1186/s12974-020-01791-8

Xia, S., Ni, Y., Zhou, Q., Liu, H., Xiang, H., Sui, H., et al. (2019). Emodin attenuates severe acute pancreatitis via antioxidant and anti-inflammatory activity. *Inflammation* 42, 2129–2138. doi:10.1007/s10753-019-01077-z

Xiao, T., Cui, Y., Ji, H., Yan, L., Pei, D., and Qu, S. (2021). Baicalein attenuates acute liver injury by blocking NLRP3 inflammasome. *Biochemical and biophysical research communications* 534, 212–218. doi:10.1016/j.bbrc.2020.11.109

Xie, R., Liu, M., and Li, S. (2019). Emodin weakens liver inflammatory injury triggered by lipopolysaccharide through elevating microRNA-145 *in vitro* and *in vivo*. *Artificial cells, nanomedicine, and biotechnology* 47, 1877–1887. doi:10.1080/21691401.2019.1614015

Xie, S., Yang, T., Wang, Z., Li, M., Ding, L., Hu, X., et al. (2020). Astragaloside IV attenuates sepsis-induced intestinal barrier dysfunction via suppressing RhoA/ NLRP3 inflammasome signaling. *International immunopharmacology* 78, 106066. doi:10.1016/j.intimp.2019.106066

Xu, G., Wan, H., Yi, L., Chen, W., Luo, Y., Huang, Y., et al. (2021). Berberine administrated with different routes attenuates inhaled LPS-induced acute respiratory distress syndrome through TLR4/NF- κ B and JAK2/STAT3 inhibition. *European journal of pharmacology* 908, 174349. doi:10.1016/j.ejphar.2021.174349

Xu, P., Zhang, W. Q., Xie, J., Wen, Y. S., Zhang, G. X., and Lu, S. Q. (2020). Shenfu injection prevents sepsis-induced myocardial injury by inhibiting mitochondrial apoptosis. *Journal of ethnopharmacology* 261, 113068. doi:10.1016/j.jep.2020.113068

Yan, J. J., Du, G. H., Qin, X. M., and Gao, L. (2020). Baicalein attenuates the neuroinflammation in LPS-activated BV-2 microglial cells through suppression of proinflammatory cytokines, COX2/NF-κB expressions and regulation of metabolic abnormality. *International immunopharmacology* 79, 106092. doi:10.1016/j.intimp. 2019.106092

Yang, C., Mo, Y., Xu, E., Wen, H., Wei, R., Li, S., et al. (2019). Astragaloside IV ameliorates motor deficits and dopaminergic neuron degeneration via inhibiting neuroinflammation and oxidative stress in a Parkinson's disease mouse model. *International immunopharmacology* 75, 105651. doi:10.1016/j.intimp.2019.05.036

Yang, G., Hu, R. Y., Deng, A. J., Huang, Y., and Li, J. (2016). Effects of electroacupuncture at zusanli, guanyuan for sepsis patients and its mechanism through immune regulation. Chinese journal of integrative medicine 22, 219-224. doi:10.1007/s11655-016-2462-9

Yang, J., Li, Y. Z., Hylemon, P. B., Zhang, L. Y., and Zhou, H. P. (2017). Cordycepin inhibits LPS-induced inflammatory responses by modulating NOD-Like Receptor Protein 3 inflammasome activation. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* 95, 1777–1788. doi:10.1016/j.biopha.2017.09.103

Yang, L., Zhang, Z., Zhuo, Y., Cui, L., Li, C., Li, D., et al. (2018). Resveratrol alleviates sepsis-induced acute lung injury by suppressing inflammation and apoptosis of alveolar macrophage cells. *American journal of translational research* 10, 1961–1975.

Yang, R., Yang, H., Wei, J., Li, W., Yue, F., Song, Y., et al. (2021). Mechanisms underlying the effects of lianhua qingwen on sepsis-induced acute lung injury: A network pharmacology approach. *Frontiers in pharmacology* 12, 717652. doi:10.3389/fphar. 2021.717652

Yang, Y., Ting, W., Xiao, L., Shufei, F., Wangxiao, T., Xiaoying, W., et al. (2017). Immunoregulation of shenqi fuzheng injection combined with chemotherapy in cancer patients: A systematic review and meta-analysis. *eCAM* 2017, 5121538. doi:10.1155/2017/ 5121538

You, L., Zhang, D., Geng, H., Sun, F., and Lei, M. (2021). Salidroside protects endothelial cells against LPS-induced inflammatory injury by inhibiting NLRP3 and enhancing autophagy. *BMC complementary medicine and therapies* 21, 146. doi:10.1186/s12906-021-03307-0

Yu, Y. H., Cui, N. Q., Fu, Q., and Li, J. (2005). Change of TH1/TH2 cytokine equilibrium in rats with severe sepsis and therapeutic effect of recombinant interleukin-12 and Shenmai injection. *Chinese journal of integrative medicine* 11, 136–141. doi:10.1007/BF02836471

Zhang, E., Wang, J., Chen, Q., Wang, Z., Li, D., Jiang, N., et al. (2020). Artesunate ameliorates sepsis-induced acute lung injury by activating the mTOR/AKT/PI3K axis. *Gene* 759, 144969. doi:10.1016/j.gene.2020.144969

Zhang, F., Geng, Y., Zhao, H., Wang, H., Zhang, Y., Li, D., et al. (2018). Effects of huanglian jiedu decoration in rat gingivitis. *eCAM* 2018, 8249013. doi:10.1155/2018/8249013

Zhang, F. L., Zhou, B. W., Yan, Z. Z., Zhao, J., Zhao, B. C., Liu, W. F., et al. (2020). 6-Gingerol attenuates macrophages pyroptosis via the inhibition of MAPK signaling pathways and predicts a good prognosis in sepsis. *Cytokine* 125, 154854. doi:10.1016/j. cyto.2019.154854

Zhang, H., Shan, Y., Wu, Y., Xu, C., Yu, X., Zhao, J., et al. (2017). Berberine suppresses LPS-induced inflammation through modulating Sirt1/NF-κB signaling pathway in RAW264.7 cells. *International immunopharmacology* 52, 93–100. doi:10.1016/j.intimp. 2017.08.032

Zhang, L., Huang, Z., Shi, X., Hu, S., Litscher, D., Wang, L., et al. (2018). Protective Effect of Electroacupuncture A. T. Zusanli on Myocardial Injury in Septic RatseCAM. *Evidencebased complementary and alternative medicine*, 2018, 6509650.

Zhang, N., Liu, J., Qiu, Z., Ye, Y., Zhang, J., and Lou, T. (2017). Shenfu injection for improving cellular immunity and clinical outcome in patients with sepsis or septic shock. *The American journal of emergency medicine* 35, 1–6. doi:10.1016/j.ajem.2016. 09.008

Zhang, Q., Li, C., Shao, F., Zhao, L., Wang, M., and Fang, Y. (2017). Efficacy and safety of combination therapy of shenfu injection and postresuscitation bundle in patients with return of spontaneous circulation after in-hospital cardiac arrest: A randomized, assessorblinded, controlled trial. *Critical care medicine* 45, 1587–1595. doi:10.1097/CCM. 000000000002570

Zhang, R., Zhu, X., Bai, H., and Ning, K. (2019). Network pharmacology databases for traditional Chinese medicine: Review and assessment. *Frontiers in pharmacology* 10, 123. doi:10.3389/fphar.2019.00123

Zhang, W., Xu, J. H., Yu, T., and Chen, Q. K. (2019). Effects of berberine and metformin on intestinal inflammation and gut microbiome composition in db/db mice. *Biomedicine* & pharmacotherapy = Biomedecine & pharmacotherapie 118, 109131. doi:10.1016/j. biopha.2019.109131

Zhang, X., Wang, Q., Wang, X., Chen, X., Shao, M., Zhang, Q., et al. (2019). Tanshinone IIA protects against heart failure post-myocardial infarction via AMPKs/mTORdependent autophagy pathway. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* 112, 108599. doi:10.1016/j.biopha.2019.108599

Zhang, Z., Shi, Y., Cai, D., Jin, S., Zhu, C., Shen, Y., et al. (2018). Effect of electroacupuncture at ST36 on the intestinal mucosal mechanical barrier and expression of occludin in a rat model of sepsis. *Acupuncture in medicine journal of the British Medical Acupuncture Society* 36, 333–338. doi:10.1136/acupmed-2016-011187

Zhang, Z. S., Zhao, H. L., Yang, G. M., Zang, J. T., Zheng, D. Y., Duan, C. Y., et al. (2019). Role of resveratrol in protecting vasodilatation function in septic shock rats and its mechanism. *The journal of trauma and acute care surgery* 87, 1336–1345. doi:10.1097/TA. 000000000002466

Zhao, F., Fang, Y., Deng, S., Li, X., Zhou, Y., Gong, Y., et al. (2017). Glycyrrhizin protects rats from sepsis by blocking HMGB1 signaling. *BioMed research international* 2017, 9719647. doi:10.1155/2017/9719647

Zheng, L., Su, J., Zhang, Z., Jiang, L., Wei, J., Xu, X., et al. (2020). Salidroside regulates inflammatory pathway of alveolar macrophages by influencing the secretion of miRNA-146a exosomes by lung epithelial cells. *Scientific reports* 10, 20750. doi:10.1038/s41598-020-77448-6 Zhou, H., Cao, H., Zheng, Y., Lu, Z., Chen, Y., Liu, D., et al. (2020). Liang-Ge-San, a classic traditional Chinese medicine formula, attenuates acute inflammation in zebrafish and RAW 264.7 cells. *Journal of ethnopharmacology* 249, 112427. doi:10.1016/j.jep.2019. 112427

Zhou, J., Gu, X., Fan, X., Zhou, Y., Wang, H., Si, N., et al. (2019). Anti-inflammatory and regulatory effects of huanglian jiedu decoction on lipid homeostasis and the TLR4/ MyD88 signaling pathway in LPS-induced zebrafish. *Frontiers in physiology* 10, 1241. doi:10.3389/fphys.2019.01241

Zhou, R. J., Ye, H., Wang, F., Wang, J. L., and Xie, M. L. (2017). Apigenin inhibits d-galactosamine/LPS-induced liver injury through upregulation of hepatic Nrf-2 and PPARy expressions in mice. *Biochemical and biophysical research communications* 493, 625–630. doi:10.1016/j.bbrc.2017.08.141

Zhou, W., Lai, X., Wang, X., Yao, X., Wang, W., and Li, S. (2021). Network pharmacology to explore the anti-inflammatory mechanism of Xuebijing in the

treatment of sepsis. Phytomedicine international journal of phytotherapy and phytopharmacology 85, 153543. doi:10.1016/j.phymed.2021.153543

Zhu, H., Wang, S., Shan, C., Li, X., Tan, B., Chen, Q., et al. (2021). Mechanism of protective effect of xuan-Bai-cheng-qi decoction on LPS-induced acute lung injury based on an integrated network pharmacology and RNA-sequencing approach. *Respiratory research* 22, 188. doi:10.1186/s12931-021-01781-1

Zhu, M. F., Xing, X., Lei, S., Wu, J. N., Wang, L. C., Huang, L. Q., et al. (2015). Electroacupuncture at bilateral zusanli points (ST36) protects intestinal mucosal immune barrier in sepsis. *Evidence-based complementary and alternative medicine eCAM* 2015, 639412. doi:10.1155/2015/639412

Zou, W., Wang, J., and Liu, Y. (2016). Effect of traditional Chinese medicine for treating human immunodeficiency virus infections and acquired immune deficiency syndrome: Boosting immune and alleviating symptoms. *Chinese journal of integrative medicine* 22, 3–8. doi:10.1007/s11655-015-2122-5

Glossary

AChE Acetylcholinesterase **AKI** Acute kidney injury AKT Protein kinase B ALI Acute lung injury AP-1 Activator protein-1 ApoM Apoliprotein M AQP Aquaporin ARDS Acute respiratory distress syndrome ASC Apoptosis associated speck like protein containing CARD **AST** Aminotransferase BALF Bronchoalveolar lavage fluid Bcl-2 B cell lymphoma-2 BMDMs Bone marrow-derived macrophages CARS Compensatory anti-inflammatory response syndrome **CAT** Catalase CCI Chronic critical illness CK-MB Creatine kinase-MB CLP Cecal ligation and puncture cMLC1 cardiac myosin light chain-1 CNKI China National Knowledge Infrastructure COX-2 Cyclooxygenase 2 cTnI cardiac troponin I CXCR2 CXC motif chemokine receptor 2 DCQD dachengqi decoction eNOS endothelial nitric oxide synthase ER endoplasmic reticulum ERK extracellular regulated protein kinase GPx glutathione peroxidase GR glutathione reductase GRK2 G protein-coupled receptor kinase-2 GSDMD gasdermin D **GSK3-\beta** glycogen synthase kinase-3 β GST glutathione-S-transferase HIF-1a hypoxia-inducible factor 1a HLA-DR human leukocyte antigen DR HLJDD huanglian jiedu decoction HMGB1 high mobility group box 1 ICAM-1 intercellular adhesion molecule-1 I-FABP intestinal fatty acid-binding protein IL-1 interleukin 1 iNOS inducible nitric oxide synthase IRF-3 interferon regulatory factor 3 JAK Janus kinase JAM-C junctional adhesion molecule C LDH lactate dehydrogenase LPS lipopolysaccharide MAPK mitogen activated protein kinase

MD2 myeloid differentiation factor 2 MDA malondialdehyde MDSCs myeloid-derived suppressor cells MEK mitogen-activated protein kinase repalmitoylated MKK4 mitogen-activated protein kinase kinase 4 MMP-9 matrix metalloproteinase 9 MPO myeloperoxidase mTOR mechanistic target of rapamycin kinase MyD88 myeloid differentiation factor 88 NETs neutrophil extracellular trap NF-κB nuclear factor-κB NLRP3 NOD-like receptor family pyrin domain containing 3 NO nitric oxide Nrf-2 nuclear factor erythroid 2-related factor 2 PAMPs pathogen-associated molecular patterns PCT procalcitonin PGE2 prostaglandin E2 PI3K phosphatidylinositol 3-kinase Pink1 PTEN-induced kinase 1 PKM2 pyruvate kinase M2 PPARy peroxisome proliferator-activated receptor gamma RAC-1 Rac family small GTPase 1 ROS reactive oxygen species S1P Sphingosine-1-Phosphate SAP severe acute pancreatitis SDH succinate dehydrogenase SFI Shenfu injection Sirt1 Sirtuin1 SMI shengmai injection SOD superoxide dismutase SOFA sequential organ failure assessment STAT signal transducer and activator of transcription TAB1 TAK1-binding protein1 TCM traditional Chinese medicine TIR interleukin-1 receptor TIRAP TIR domain-containing adaptor protein TKA1 TGF-β-activated kinase1 TLR4 Toll-like receptor 4 TNF-a tumor necrosis factor alpha TRAF6 TNF receptor associated factor 6 TRIF TIR-domain-containing adapter-inducing interferon- β TRPM7 transient receptor potential melastatin 7 TSA Tanshinone IIA VCAM1 vascular cell adhesion molecule 1 VEGF vascular endothelial growth factor VEGF-B vascular endothelial growth factor B **XBJI** xuebijing injection XJDHD xijiao dihuang decoction ZO-1 zonula occludens-1