#### Check for updates

#### **OPEN ACCESS**

EDITED BY Jing Wu, Shandong Provincial Qianfoshan Hospital, China

REVIEWED BY Ziqing Chen, Princeton University, United States Xinshuang Yu, The First Affiliated Hospital of Shandong First Medical University, China

\*CORRESPONDENCE Kaiming Ren, i renkmcmu@163.com

SPECIALTY SECTION This article was submitted to Experimental Pharmacology and Drug Discovery, a section of the journal Frontiers in Pharmacology

RECEIVED 21 December 2022 ACCEPTED 30 January 2023 PUBLISHED 10 February 2023

#### CITATION

Yu Y, Nie Q, Wang Z, Di Y, Chen X and Ren K (2023), Targeting acetyl-CoA carboxylase 1 for cancer therapy. *Front. Pharmacol.* 14:1129010. doi: 10.3389/fphar.2023.1129010

#### COPYRIGHT

© 2023 Yu, Nie, Wang, Di, Chen and Ren. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

### Targeting acetyl-CoA carboxylase 1 for cancer therapy

Yong Yu<sup>1</sup>, Qingzhu Nie<sup>1</sup>, Ziyi Wang<sup>2</sup>, Yu Di<sup>1</sup>, Xiaolong Chen<sup>1</sup> and Kaiming Ren<sup>3</sup>\*

<sup>1</sup>Department of Ophthalmology, Shengjing Hospital of China Medical University, Shenyang, Liaoning, China, <sup>2</sup>Department of Thoracic Surgery, First Hospital of China Medical University, Shenyang, Liaoning, China, <sup>3</sup>Department of Thoracic Surgery, Shengjing Hospital of China Medical University, Shenyang, Liaoning, China

Metabolic adaptation is an emerging hallmark of tumors. De novo fatty acid synthesis is an important metabolic process to produce metabolic intermediates for energy storage, biosynthesis of membrane lipids and generation of signaling molecules. Acetyl-CoA carboxylase 1 (ACC1) is a critical enzyme in the fatty acid synthesis, which carboxylates acetyl-CoA carboxylic acid to form malonyl-CoA. The role of acetyl-CoA carboxylase 1 in fatty acid synthesis makes it a promising therapeutic target for various metabolic diseases such as non-alcoholic fatty liver disease, obesity and diabetes. Tumors have a high energy flow and a strong dependence on fatty acid synthesis. Thus, acetyl-CoA carboxylase inhibition has become a potential choice for anti-tumor therapy. In this review, we first introduced the structure and expression pattern of Acetyl-CoA carboxylase 1. We also discussed the molecular mechanisms of acetyl-CoA carboxylase 1 in the initiation and progression of various cancer types. Furthermore, acetyl-CoA carboxylase1 inhibitors has also been discussed. Collectively, we summarized the interplay between acetyl-CoA carboxylase 1 and tumorigenesis, indicating acetyl-CoA carboxylase 1 as a promising therapeutic target for tumor management.

#### KEYWORDS

metabolism, cancer, acetyl-CoA carboxylase 1, fatty acid, fatty acid synthesis

#### Introduction

Fatty acid metabolism, particularly the synthesis of fatty acids, is an essential metabolic process that produces various metabolic intermediates for energy storage, biosynthesis of membrane lipids and production of signaling molecules (Snaebjornsson et al., 2020). Thus, fatty acid synthesis, uptake, and oxidation contributes to the maintenance of biological function, and blocking these processes regulating fatty acid metabolism may affect cell survival. Alterations in fatty acid metabolism are one of the most significant metabolic changes involved in tumorigenesis. Increased fatty acid synthesis supports tumor cell survival and growth (Munir et al., 2019; Broadfield et al., 2021). Therefore, targeting fatty acid metabolism, particularly fatty acid synthesis, is gradually considered as a promising anti-tumor therapeutic strategy.

Glucose metabolism interacts with fatty acid metabolism at the point of citric acid, an intermediate of the tricarboxylic acid cycle. Converting carbon from citrate to bioactive fatty acids requires several steps. These steps involve ATP citrate lyase (ACLY), acetyl-CoA carboxylase (ACC), fatty acid synthase (FASN), and acyl-CoA synthase (ACSL) (Brownsey et al., 2006; Zaidi et al., 2012; Röhrig and Schulze, 2016). ACC carboxylates acetyl-CoA to form malonyl-CoA, and is a critical enzyme in the fatty acid synthesis. Two isoforms of ACC exist in eukaryotic cells (Kim, 1997). ACC1 is highly enriched in adipogenic tissues and ACC2 exists in oxidized tissues. Since ACC1 and ACC2 are found in different tissues, they have distinct metabolic roles. The first rate-limiting

step in fatty acid synthesis is mediated by ACC1, which converts acetyl-CoA to malonyl-CoA in the cytoplasm. Malonyl-CoA is consumed by FASN for *de novo* fatty acid synthesis, while consuming large amounts of NADPH. ACC2 localizes to the outer mitochondrial membrane. Malonyl-CoA produced by ACC2 inhibits carnitine palmitoyltransferase 1 (CPT-1) and prevents fatty acids entering mitochondria to drive fatty acid oxidation. Malonyl-CoA synthesized by ACC1 is considered as a substrate for fatty acid synthesis, while malonyl-CoA synthesized by ACC2 primarily participates in the inhibition of CPT-1 and fatty acid degradation (Wang et al., 2022). The overview of fatty acid metabolism has been shown in Figure 1.

### Structure and regulation of human acetyl-CoA carboxylase 1

The critical function of ACC1 in fatty acid metabolism has sparked huge amounts of investigations on its structure. The structure of eukaryotic ACC1 is polypeptide, containing main domains including biotin carboxylase (BC), biotin containing carboxyl carrier protein (BCCP), carboxyltransferase (CT), as well as an interaction domain (BT) and a non-catalytic center domain (CD) (Tong, 2012). CD is composed of four domains: N-terminal CDN, the linking CDL, and the tandem C-terminal CDC1 and CDC2 (Hunkeler et al., 2016). Acetyl-CoA carboxylation is a two-step reaction: BCCP linked-biotin is partially carboxylated, and ATP is consumed by BC domain; Next, the generated carboxybiotin is transported to the CT domain, and the carboxyl group is exchanged to acetyl-CoA (Hunkeler et al., 2018).

It has been shown that ACC1 is regulated by protein phosphorylation, allosteric regulator binding and protein-protein interactions. Human ACC1 is inactivated by AMPK phosphorylation and cAMP-dependent protein kinase (PKA) phosphorylation (Ha et al., 1994). Inhibition of ACC1 phosphorylation by AMPK is critical for controlling fatty acid metabolism and cellular response to metabolic stress. AMPK is an essential regulator of metabolism and is activated by both AMP-dependent and AMPindependent mechanisms (Fullerton et al., 2013). Mice with knock-in mutations in both ACC1 (Ser79) and ACC2 (Ser212) exhibit increased fatty acid synthesis and inhibition of fatty acid oxidation compared with wild-type mice (Fullerton et al., 2013). Moreover, inhibition of ACC1 by AMPK is essential for maintaining NADPH levels under conditions of energy stress, which is achieved by reducing NADPH consumption in fatty acid synthesis and enhancing NADPH produced by fatty acid oxidation. ACC1 inhibition compensates AMPK activation and promotes anchor-independent growth and solid tumor formation *in vivo*, whereas ACC1 activation attenuates these processes (Fullerton et al., 2013).

In addition, the tumor suppressor BRCA1 is speculated to regulate ACC1 (Shen and Tong, 2008). BRCA1 interacts with ACC1 and participates in the regulation of lipogenesis in adipose tissues. BRCA1 binds to ACC1 through its C-terminal tandem BRCT domain, which recognizes phosphorylated Ser1263 in the CD region of ACC1 (Hunkeler et al., 2018). Binding to BRCA1 can prevent the dephosphorylation of Ser80, thereby inhibiting the activation of ACC1. Mutations in the BRCT domain could disrupt the binding of BRCA1 to ACC1 and lead to elevated adipogenesis, which is critical for tumor growth. The interaction between cytosolic ACC1 and BRCT domain of BRCA1 is mainly localized in the nucleus, but also occurs in the cytosols. The interaction between the dimerized BRCT domain and the ACC1 phosphorylation loop triggers the polymerization of the extended ACC1 dimer into an ACC1-BRCT double stranded filament with a unique CD conformation and a monomeric BC domain (Fullerton et al., 2013). Post-translational modifications of ACC1 contribute to the function of ACC1, which may provide promising therapeutic targets for anti-tumor strategies.





## Acetyl-CoA carboxylase 1 and malignancies

ACC1 has been implicated in the initiation and development of multiple malignancies. The expression levels of ACC1 have been analyzed by GEPIA database and illustrated in Figure 2. In this part, we discussed the complex and dynamic role of ACC1 in diverse cancer types.

## Acetyl-CoA carboxylase 1 and acute myeloid leukemia

Acute myeloid leukemia (AML) is a malignant tumor of myeloid stem cell precursors (Prada-Arismendy et al., 2017). ACC1 functions as a substrate for the TRIB1-COP1 complex, and its degradation is important for mediating metabolic adaptations to provide an energetic advantage to support tumor development (Ito et al., 2021). It has been shown that ACC1 binds to Tribbles through the a-helical region of its N-terminal BC domain. ACC1 stabilization impairs Trib1-COP1-induced leukemia progression by increasing ROS levels and NADPH depletion. Thus, ACC1 stabilization abrogates the self-renewal capacity of leukemic stem cells and promotes myeloid differentiation in mouse models of AML. Therefore, small molecule inhibitors targeting the ACC1-helical region are promising for AML treatment. A drug combination redeployed with bezafibrate and medroxyprogesterone acetate (designated BaP) exhibits anti-tumor activity in AML cells (Southam et al., 2015). Mass spectroscopy-based lipidomics and gene expression levels of lipogenic enzymes has indicated that ACC1, FASN, and stearoyl-CoA desaturase 1 (SCD1) were changed in AML cell lines treated with BaP (Southam et al., 2015). Supplementation with oleate, an enzyme product of SCD1, protected AML cells from

BaP-induced cell death and reduced the level of BaP-induced ROS. Precious studies indicated that monounsaturated fatty acids contribute to the tumor initiation and development (Scott et al., 2022). Mechanistically, BaP plays an anticancer role by disrupting monounsaturated fatty acid synthesis. BaP has been used in clinical trials to treat leukemia by targeting adipogenesis. The antitumor mechanism of BaP needs further study, and the combination of BaP with other antitumor agents may provide a potential strategy for the treatment of AML.

## Acetyl-CoA carboxylase 1 and breast cancer

The high invasiveness and relapse rates of breast cancer are related to obesity, and obesity-related metabolic dysfunction has become the primary risk factor for postmenopausal women to develop metastatic breast cancer (Khandekar et al., 2011). ACC1 inhibition has been identified as a molecular convergence point for epithelial mesenchymal transition and tumor invasion (Rios Garcia et al., 2017). Leptin and TGFβ could inhibit ACC1 function through TGFβ-activated kinase-AMPK signaling in breast cancer. ACC1 downregulation further led to an increased level of total acetyl CoA and protein acetylation, which acetylation activation induced of Smad2 transcription factor and epithelial mesenchymal transition to enhance migrative and invasive capability of breast cancer cells (Rios Garcia et al., 2017). Specific inhibition of ACC1 in breast tumor cells impeded palmitic acid synthesis, which further induced cell apoptosis, ROS accumulation and mitochondrial dysfunction. Small molecule inhibitors targeting ACC1 could impair cell viability of breast cancer and inhibited tumor growth in xenografts derived from cell lines and patients (Chajès et al., 2006).

## Acetyl-CoA carboxylase 1 and lung cancer

Lung cancer is the leading cause of cancer-related death worldwide because of its rapid growth and high invasiveness (Sung et al., 2021). ACC1 mRNA has been found to be upregulated in non-small cell lung cancer (NSCLC), accompanied by decreased DNA methylation on the S-shore of CpG island of ACC1 (Li et al., 2019). Besides, upregulated ACC1 level was associated with poor prognosis in NSCLC patients (Li et al., 2019). Long non-coding RNAs (LncRNAs) interact with proteins to participate in the regulation of protein function. A novel lncRNA, CTD-2245E15.3, enhanced lung tumor initiation and development by regulating ACC1 (Wang et al., 2021). CTD-245E15-3 exerted its oncogenic effect by binding to ACC1 to regulate fatty acid biosynthesis in tumor cells. Knockdown of CTD-2245E15.3 increased phosphorylation of ACC1, leading to inhibition of enzyme activity. ND-646, an allosteric inhibitor of ACC1, prevented the dimerization of ACC1 subunits to block fatty acid synthesis and thus impede lung tumor growth (Li et al., 2019). ND-646 significantly inhibited lung tumor growth in KRAS p53 and KRAS Lkb1 mouse models of NSCLC (Svensson et al., 2016).

## Acetyl-CoA carboxylase 1 and liver cancer

Liver cancer leads to nearly 750,000 deaths every year, most of which are hepatocellular carcinoma (HCC) (Sung et al., 2021). ACC1 converts acetyl CoA to malonyl CoA, and malonyl CoA is the first substrate used for de novo lipogenesis (DNL). The increase of liver DNL is the key factor of HCC initiation (Liu and Dong, 2021). Fructose consumption and gene activation of ACC1 increase DNL and hepatocarcinogenesis. ND-654, a liver-specific ACC inhibitor, mimics the effect of ACC1 phosphorylation and impairs the progression of hepatic DNL and HCC (Lally et al., 2019). Single or in combination with sorafenib, a multi-kinase inhibitor, could improve the survival rate of tumor-bearing rats. Gene alteration led to the decrease of ACC phosphorylation level, which can promote the development of HCC cells. Although mutations in AMPK or ACC1 have not been described in HCC, there is sufficient literature to prove that obesity and type 2 diabetes, two major risk factors for HCC, are related to reduced AMPK activity and ACC phosphorylation level. These findings indicate that therapies that use small molecules like ND-654 to simulate the effect of AMPK phosphorylation on ACC may be valuable for the treatment of HCC.

# Acetyl-CoA carboxylase 1 and other malignancy

Lipidomic profiling of specimens from prostate tumor has indicated promising therapeutic targets involved in tumorigenesis of prostate cancer. Targeting lipid features by inhibiting ACC1 led to impaired tumor proliferation and increased cell apoptosis in patient-derived explants (Butler et al., 2021). ACC1 inhibition could impair tumor formation in colitis-associated colon cancer (Li et al., 2022). Blocking fatty acid synthesis by ACC1 inhibition in intestinal epithelial cells led to deceased epithelial crypt structures and reduced Lgr5+ intestinal epithelial stem cells by impairing the nuclear accumulation of PPAR $\delta/\beta$ -catenin (Li et al., 2022). ACC1 also participates in the rewiring of tumor metabolism in response to cetuximab treatment of head and neck squamous cell carcinoma (HNSCC). HNSCC cells with acquired cetuximab resistance display elevated expression of T172-phosphorylated AMPK, S79-phosphorylated ACC1, and ACC1 (Luo et al., 2017). ACC1 rewires HNSCC metabolism from glycolysis to lipogenesis. Cetuximab plus TOFA (an allosteric inhibitor of ACC) exhibited significantly inhibitive effect on the growth of cetuximab-resistant HNSCC xenografts (Luo et al., 2017).

#### Acetyl-CoA carboxylase 1 and nontumor diseases

ACC1 is a key regulator of fatty acid metabolism, making it promising therapeutic targets for multiple metabolic diseases. ACC1 has been regarded as attractive therapeutic targets for non-alcoholic steatohepatitis (NASH). However, the use of ACC1 inhibitors tested in clinical trials is limited by side effects such as hypertriglyceridemia (Kim et al., 2017; Goedeke et al., 2018). The anti-NASH effect of ACC1 inhibitors alone or in combination with ACC1 inhibitors (PF-05221304) and diacylglycerol acyltransferase 2 (DGAT2) inhibitors (PF-06865571) has been tested in a clinical trial (Calle et al., 2021). The results showed that ACC1 inhibitor alone had obvious inhibitory effect on NASH, but it could induce hyperlipidemia, while the combination with DGAT2 inhibitor could reduce the side effects. Therefore, the combination of PF-05221304 and PF-06865571 is a potential treatment strategy, which can eliminate the side effects of PF-05221304 single treatment and bring greater clinical benefits than PF-06865571 or PF-05221304 single treatment (Calle et al., 2021). Novel ACC1-targeted anti-NASH therapeutic strategies need to be explored to successfully address the adverse effects of hyperlipidemia while maintaining the anti-NASH efficacy. IMA-1 is a small molecule for the treatment of NASH by interrupting the interaction of arachidonic acid 12 lipoxygenase (ALOX12)-ACC1 (Zhang et al., 2021). The anti-NASH effect of IMA-1 mainly depends on its direct binding to the ALOX12 pocket near the interactive surface of ACC1, rather than inhibit the activity of ALOX12 lipoxygenase. IMA-1 treatment exhibits little side effect of inhibiting ACC1 enzyme activity. These findings have brought promising results to the indications of ACC1 inhibition in anti-NASH treatment.

Fatty acid synthesis is also critical for platelet activation (Yoshida et al., 1992; Kelly et al., 2020). In the double knock-in (DKI) mouse model, Ser79 on ACC1 and Ser212 on ACC2, the phosphorylation site of AMPK, were mutated to block AMPK effect on ACC (Lepropre et al., 2018). Blood collected from ACC DKI mice showed an increase in thrombosis induced by vascular injury and growth of thrombus on the collagen coated surface. In collagen-stimulated platelets, the loss of AMPK-ACC1 signal is related to the expansion of thromboxane production and the secretion of dense granules. In general, AMPK-ACC signaling pathway controls thrombosis formation by specifically regulating the response of thromboxane and particle release to collagen. ACC inhibitor CP640.186 has been proved to increase tubulin acetylation and impair thrombin-mediated platelet aggregation (Octave et al., 2021).

Yu et al

### The clinical implications of targeting acetyl-CoA carboxylases

Tumor metabolic remodeling is often accompanied by changes in the expression of metabolic enzymes (Jiang et al., 2021). In addition to catalyzing intracellular metabolic reactions, metabolic enzymes also participate in the regulation of gene expression, cell cycle, DNA damage repair, cellular antioxidant capacity, proliferation, survival, apoptosis and tumor microenvironment, which further expands the metabolic dependence of tumor cells and endowing tumor cells with the ability to adapt to different environmental stimuli (Pan et al., 2021). In a study using mRNA expression data from the Cancer Genome Atlas, the expression patterns of a list of enzymes in multiple cancer types and matched normal tissues were detected to identify enzymes that were dominant in tumorigenesis. A prioritized list of new metabolic targets was developed for these cancer types with ACC1 being the most widely shared and functionally confirmed target (Marczyk et al., 2022). The expression levels of ACC1 have been analyzed by GEPIA database (Tang et al., 2017).

Soraphen A is a macrocyclic polyketide from myxomycetes, exhibiting anti-fungal activity due to its ability to bind to the dimeric interface of the BC domain of eukaryotic ACC and thereby impairing enzymatic oligomerization and blocking its activity. At nanomolar concentrations, sorafen A inhibits adipogenesis and promotes fatty acid oxidation in prostate tumor cells, while it has no cytotoxic effect on precancerous cells (Beckers et al., 2007). 5-tetracepoxy-2-furan acid (TOFA), a structural inhibitor of ACC1, exerts cytotoxic effects on lung and colon tumor cells. RNAi technology targeting ACC1 resulted in inhibition of LNCaP prostate tumor cell proliferation and caspase-induced apoptosis of highly lipogenic tumor cells (Brusselmans et al., 2005). ND-646 functions as an allosteric inhibitor of ACC1 and ACC2 by preventing ACC subunit dimerization, thus blocking fatty acid synthesis in vitro and in vivo. In several models of NSCLC, ND-646 treatment impeded fatty acid synthesis and lipid accumulation, leading to impaired tumor proliferation and enhanced cell apoptosis (Svensson et al., 2016). Collectively, there is increasing interest in the use of ACC inhibitors for anti-tumor strategies. Further evaluations of potency, pharmacokinetics, and doses of these inhibitors are required.

### References

Beckers, A., Organe, S., Timmermans, L., Scheys, K., Peeters, A., Brusselmans, K., et al. (2007). Chemical inhibition of acetyl-CoA carboxylase induces growth arrest and cytotoxicity selectively in cancer cells. *Cancer Res.* 67 (17), 8180–8187. doi:10.1158/0008-5472.CAN-07-0389

Broadfield, L. A., Pane, A. A., Talebi, A., Swinnen, J. V., and Fendt, S. M. (2021). Lipid metabolism in cancer: New perspectives and emerging mechanisms. *Dev. Cell* 56 (10), 1363–1393. Epub 2021 May 3. doi:10.1016/j.devcel.2021.04.013

Brownsey, R. W., Boone, A. N., Elliott, J. E., Kulpa, J. E., and Lee, W. M. (2006). Regulation of acetyl-CoA carboxylase. *Biochem. Soc. Trans.* 34, 223–227. doi:10.1042/ BST20060223

Brusselmans, K., De Schrijver, E., Verhoeven, G., and Swinnen, J. V. (2005). RNA interference-mediated silencing of the acetyl-CoA-carboxylase-alpha gene induces growth inhibition and apoptosis of prostate cancer cells. *Cancer Res.* 65 (15), 6719–6725. doi:10. 1158/0008-5472.CAN-05-0571

Butler, L. M., Mah, C. Y., Machiels, J., Vincent, A. D., Irani, S., Mutuku, S. M., et al. (2021). Lipidomic profiling of clinical prostate cancer reveals targetable alterations in membrane lipid composition. *Cancer Res.* 81 (19), 4981–4993. Epub 2021 Aug 6. doi:10. 1158/0008-5472.CAN-20-3863

#### Conclusion

ACC1 plays an essential role in fatty acid synthesis. For many years, the key regulatory role of ACC1 in fatty acid metabolism makes it attractive therapeutic targets for various metabolic diseases including non-alcoholic fatty liver disease, obesity and diabetes. Tumors have a high energy flow and a strong dependence on fatty acid synthesis. ACC1 inhibition has become an appealing choice for anti-tumor therapy. Some encouraging results indicated that at least some specific tumor types may respond to ACC1 inhibition. It may be one of the hot spots in future study to expand the clinical indications of ACC1 inhibitors. It is desirable to explore effective inhibitors targeting ACC1.

### Author contributions

YY wrote and elaborated the figures. QN, ZW, YD, XC, wrote and reviewed the manuscript. KR wrote and reviewed the final version. All authors contributed to the article and approved the submitted version.

### Funding

This work was supported by the National Natural Science Foundation of China (81570866).

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Calle, R. A., Amin, N. B., Carvajal-Gonzalez, S., Ross, T. T., Bergman, A., Aggarwal, S., et al. (2021). ACC inhibitor alone or co-administered with a DGAT2 inhibitor in patients with non-alcoholic fatty liver disease: Two parallel, placebo-controlled, randomized phase 2a trials. *Nat. Med.* 27 (10), 1836–1848. Epub 2021 Oct 11. doi:10.1038/s41591-021-01489-1

Chajès, V., Cambot, M., Moreau, K., Lenoir, G. M., and Joulin, V. (2006). Acetyl-CoA carboxylase alpha is essential to breast cancer cell survival. *Cancer Res.* 66 (10), 5287–5294. doi:10.1158/0008-5472.CAN-05-1489

Fullerton, M. D., Galic, S., Marcinko, K., Sikkema, S., Pulinilkunnil, T., Chen, Z. P., et al. (2013). Single phosphorylation sites in Acc1 and Acc2 regulate lipid homeostasis and the insulin-sensitizing effects of metformin. *Nat. Med.* 19 (12), 1649–1654. doi:10.1038/nm. 3372

Goedeke, L., Bates, J., Vatner, D. F., Perry, R. J., Wang, T., Ramirez, R., et al. (2018). Acetyl-CoA carboxylase inhibition reverses NAFLD and hepatic insulin resistance but promotes hypertriglyceridemia in rodents. *Hepatology* 68 (6), 2197–2211. doi:10.1002/ hep.30097

Ha, J., Daniel, S., Broyles, S. S., and Kim, K. H. (1994). Critical phosphorylation sites for acetyl-CoA carboxylase activity. *J. Biol. Chem.* 269, 22162–22168. doi:10.1016/s0021-9258(17)31770-2

Hunkeler, M., Hagmann, A., Stuttfeld, E., Chami, M., Guri, Y., Stahlberg, H., et al. (2018). Structural basis for regulation of human acetyl-CoA carboxylase. *Nature* 558 (7710), 470–474. Epub 2018 Jun 13. doi:10.1038/s41586-018-0201-4

Hunkeler, M., Stuttfeld, E., Hagmann, A., Imseng, S., and Maier, T. (2016). The dynamic organization of fungal acetyl-CoA carboxylase. *Nat. Commun.* 7, 11196. doi:10.1038/ncomms11196

Ito, H., Nakamae, I., Kato, J. Y., and Yoneda-Kato, N. (2021). Stabilization of fatty acid synthesis enzyme acetyl-CoA carboxylase 1 suppresses acute myeloid leukemia development. J. Clin. Invest. 131 (12), e141529. doi:10.1172/JCI141529

Jiang, J., Peng, L., Wang, K., and Huang, C. (2021). Moonlighting metabolic enzymes in cancer: New perspectives on the redox code. *Antioxid. Redox Signal* 34 (13), 979–1003. doi:10.1089/ars.2020.8123

Kelly, K. L., Reagan, W. J., Sonnenberg, G. E., Clasquin, M., Hales, K., Asano, S., et al. (2020). De novo lipogenesis is essential for platelet production in humans. *Nat. Metab.* 2 (10), 1163–1178. doi:10.1038/s42255-020-00272-9

Khandekar, M. J., Cohen, P., and Spiegelman, B. M. (2011). Molecular mechanisms of cancer development in obesity. *Nat. Rev. Cancer* 11, 886–895. doi:10.1038/nrc3174

Kim, C. W., Addy, C., Kusunoki, J., Anderson, N. N., Deja, S., Fu, X., et al. (2017). Acetyl CoA carboxylase inhibition reduces hepatic steatosis but elevates plasma triglycerides in mice and humans: A bedside to bench investigation. *Cell Metab.* 26 (2), 394–406. doi:10. 1016/j.cmet.2017.07.009

Kim, K. H. (1997). Regulation of mammalian acetyl-coenzyme A carboxylase. Annu. Rev. Nutr. 17, 77–99. doi:10.1146/annurev.nutr.17.1.77

Lally, J. S. V., Ghoshal, S., DePeralta, D. K., Moaven, O., Wei, L., Masia, R., et al. (2019). Inhibition of acetyl-CoA carboxylase by phosphorylation or the inhibitor ND-654 suppresses lipogenesis and hepatocellular carcinoma. *Cell Metab.* 29 (1), 174–182. doi:10.1016/j.cmet.2018.08.020

Lepropre, S., Kautbally, S., Octave, M., Ginion, A., Onselaer, M. B., Steinberg, G. R., et al. (2018). AMPK-ACC signaling modulates platelet phospholipids and potentiates thrombus formation. *Blood* 132 (11), 1180–1192. doi:10.1182/blood-2018-02-831503

Li, E. Q., Zhao, W., Zhang, C., Qin, L. Z., Liu, S. J., Feng, Z. Q., et al. (2019). Synthesis and anti-cancer activity of ND-646 and its derivatives as acetyl-CoA carboxylase 1 inhibitors. *Eur. J. Pharm. Sci.* 137, 105010. doi:10.1016/j.ejps.2019.105010

Li, S., Lu, C. W., Diem, E. C., Li, W., Guderian, M., Lindenberg, M., et al. (2022). Acetyl-CoA-Carboxylase 1-mediated de novo fatty acid synthesis sustains Lgr5+ intestinal stem cell function. *Nat. Commun.* 13 (1), 3998. doi:10.1038/s41467-022-31725-2

Liu, Q., and Dong, X. (2021). Targeting de novo lipogenesis and cholesterol biosynthesis simultaneously is a novel therapeutic option for hepatocellular carcinoma. *J. Hepatocell. Carcinoma* 8, 19–21. doi:10.2147/JHC.S278517

Luo, J., Hong, Y., Lu, Y., Qiu, S., Chaganty, B. K., Zhang, L., et al. (2017). Acetyl-CoA carboxylase rewires cancer metabolism to allow cancer cells to survive inhibition of the Warburg effect by cetuximab. *Cancer Lett.* 384, 39–49. doi:10.1016/j.canlet.2016.09.020

Marczyk, M., Gunasekharan, V., Casadevall, D., Qing, T., Foldi, J., Sehgal, R., et al. (2022). Comprehensive analysis of metabolic isozyme targets in cancer. *Cancer Res.* 82 (9), 1698–1711. doi:10.1158/0008-5472.CAN-21-3983

Munir, R., Lisec, J., Swinnen, J. V., and Zaidi, N. (2019). Lipid metabolism in cancer cells under metabolic stress. *Br. J. Cancer* 120 (12), 1090–1098. doi:10.1038/s41416-019-0451-4

Octave, M., Pirotton, L., Ginion, A., Robaux, V., Lepropre, S., Ambroise, J., et al. (2021). Acetyl-CoA carboxylase inhibitor CP640.186 increases tubulin acetylation and impairs thrombin-induced platelet aggregation. *Int. J. Mol. Sci.* 22 (23), 13129. doi:10.3390/ ijms222313129 Pan, C., Li, B., and Simon, M. C. (2021). Moonlighting functions of metabolic enzymes and metabolites in cancer. *Mol. Cell* 81 (18), 3760-3774. doi:10.1016/j. molcel.2021.08.031

Prada-Arismendy, J., Arroyave, J. C., and Röthlisberger, S. (2017). Molecular biomarkers in acute myeloid leukemia. *Blood Rev.* 31 (1), 63-76. doi:10.1016/j. blre.2016.08.005

Rios Garcia, M., Steinbauer, B., Srivastava, K., Singhal, M., Mattijssen, F., Maida, A., et al. (2017). Acetyl-CoA carboxylase 1-dependent protein acetylation controls breast cancer metastasis and recurrence. *Cell Metab.* 26 (6), 842–855. doi:10.1016/j. cmet.2017.09.018

Röhrig, F., and Schulze, A. (2016). The multifaceted roles of fatty acid synthesis in cancer. *Nat. Rev. Cancer* 16 (11), 732–749. doi:10.1038/nrc.2016.89

Scott, J. S., Nassar, Z. D., Swinnen, J. V., and Butler, L. M. (2022). Monounsaturated fatty acids: Key regulators of cell viability and intracellular signaling in cancer. *Mol. Cancer Res.* 20 (9), 1354–1364. doi:10.1158/1541-7786.MCR-21-1069

Shen, Y., and Tong, L. (2008). Structural evidence for direct interactions between the BRCT domains of human BRCA1 and a phospho-peptide from human ACC1. *Biochemistry* 47 (21), 5767–5773. doi:10.1021/bi800314m

Snaebjornsson, M. T., Janaki-Raman, S., and Schulze, A. (2020). Greasing the wheels of the cancer machine: The role of lipid metabolism in cancer. *Cell Metab.* 31 (1), 62–76. doi:10.1016/j.cmet.2019.11.010

Southam, A. D., Khanim, F. L., Hayden, R. E., Constantinou, J. K., Koczula, K. M., Michell, R. H., et al. (2015). Drug redeployment to kill leukemia and lymphoma cells by disrupting SCD1-mediated synthesis of monounsaturated fatty acids. *Cancer Res.* 75 (12), 2530–2540. Epub 2015 May 5. doi:10.1158/0008-5472.CAN-15-0202

Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., et al. (2021). Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 71 (3), 209–249. doi:10.3322/caac.21660

Svensson, R. U., Parker, S. J., Eichner, L. J., Kolar, M. J., Wallace, M., Brun, S. N., et al. (2016). Inhibition of acetyl-CoA carboxylase suppresses fatty acid synthesis and tumor growth of non-small-cell lung cancer in preclinical models. *Nat. Med.* 22 (10), 1108–1119. doi:10.1038/nm.4181

Tang, Z., Li, C., Kang, B., Gao, G., Li, C., and Zhang, Z. (2017). GEPIA: A web server for cancer and normal gene expression profiling and interactive analyses. *Nucleic Acids Res.* 45 (W1), W98–W102. doi:10.1093/nar/gkx247

Tong, L. (2012). Structure and function of biotin-dependent carboxylases. Cell Mol. Life Sci. 70, 863–891. doi:10.1007/s00018-012-1096-0

Wang, C., Meng, X., Zhou, Y., Yu, J., Li, Q., Liao, Z., et al. (2021). Long noncoding RNA CTD-2245e15.3 promotes anabolic enzymes ACC1 and PC to support nonsmall cell lung cancer growth. *Cancer Res.* 81 (13), 3509–3524. doi:10.1158/0008-5472.CAN-19-3806

Wang, Y., Yu, W., Li, S., Guo, D., He, J., and Wang, Y. (2022). Acetyl-CoA carboxylases and diseases. *Front. Oncol.* 12, 836058. doi:10.3389/fonc.2022.836058

Yoshida, K., Asaoka, Y., and Nishizuka, Y. (1992). Platelet activation by simultaneous actions of diacylglycerol and unsaturated fatty acids. *Proc. Natl. Acad. Sci. U. S. A.* 89 (14), 6443–6446. doi:10.1073/pnas.89.14.6443

Zaidi, N., Swinnen, J. V., and Smans, K. (2012). ATP-Citrate lyase: A key player in cancer metabolism. *Cancer Res.* 72, 3709–3714. doi:10.1158/0008-5472.CAN-11-4112

Zhang, X., Ji, Y., Cheng, X., Cheng, Y., Yang, H., Wang, J., et al. (2021). A small molecule targeting ALOX12-ACC1 ameliorates nonalcoholic steatohepatitis in mice and macaques. *Sci. Transl. Med.* 13 (624), eabg8116. doi:10.1126/scitranslmed. abg8116