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EDITED BY

François Chassagne,
IRD UMR152 Pharmacochimie et Biologie
Pour le Développement (PHARMADEV),
France

REVIEWED BY

Smith B. Babiaka,
University of Buea, Cameroon
Latifa Bouissane,
Université Sultan Moulay Slimane,
Morocco

*CORRESPONDENCE

Gizachew Kassahun Bizuneh,
✉ gizachewkassahun4@gmail.com

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Ethnobotanical uses, phytochemistry and biological activity of the genus *Euclea*: A review

Abebe Dagne Taye¹, Gizachew Kassahun Bizuneh^{2*} and
Asmamaw Emagn Kasahun³

¹Department of Pharmacy, College of Health Sciences, Debre Markos University, Debre Markos, Ethiopia,

²Department of Pharmacognosy, School of Pharmacy, College of Medicine and Health Sciences,

University of Gondar, Gondar, Ethiopia, ³Department of Pharmaceutics, School of Pharmacy, College of
Medicine and Health Sciences, University of Gondar, Gondar, Ethiopia

Euclea (Ebenaceae) is a genus of flowering shrubs and trees widely distributed in Africa, the Comoro Islands, and Arabia. This review aimed to evaluate the ethnobotanical uses, phytochemistry, and biological activities of the genus *Euclea* on available research reports. This was achieved through PubMed, Medline, Google Scholar, Science Direct, Taylor and Francis Online, Wiley Online Library which provides access to scientific and medical research. The extensive literature survey revealed that plants that belong to this genus are used as folkloric medicine for the treatment of diabetes mellitus, toothache, diarrhea, cancer, malaria, leprosy, and genital and oral diseases in the case of HIV/AIDS-related diseases. To date, more than 40 secondary metabolites have been isolated and identified from these plants, especially from *E. natalensis* and *E. divinorum*. Among these, naphthoquinones, terpenes, and flavonoids are potential secondary metabolites with profound biological activities. *Euclea* plant extracts and their bioactive compounds possess outstanding pharmacological properties, especially antimalarial, antidiabetic, anticancer, antimicrobial, and antioxidant properties.

KEYWORDS

Euclea, naphthoquinones, phytochemistry, ethnobotanical use, pharmacological activity

Introduction

The word “*Euclea*” comes from a Greek word “eukleia”, “eu” meaning “good”, and “kleios” meaning report (Maroyi, 2017). The genus *Euclea* belongs to the family Ebenaceae and is composed of 16 accepted species (Dhayalan et al., 2015; Botha, 2016).

The genus *Euclea* is distributed in the tropical and subtropical regions of the world. However, it is most abundant in Eastern and Southern Africa (Mebe et al., 1998) and South-East Asia (Botha, 2016). *Euclea divinorum* is distributed in Botswana, South Africa, Namibia, Swaziland, Zimbabwe, Tanzania, Uganda (Shumba, 2018), Sudan, Kenya, and Ethiopia (Woldemedhin et al., 2017). *Euclea natalensis* is widely found along the eastern coast of southern Africa (Johanna, 2007). *Euclea latideus* is well presented in the lowlands of the tropical and to a lesser extent, in subtropical regions of the world (Philip et al., 2018). A versatile medicinal plant in Ethiopia from this genus is *Euclea divinorum*. Traditionally it is used for the treatment of skin inflammation, scabies, cancer, hepatitis, urinary inconsistency,

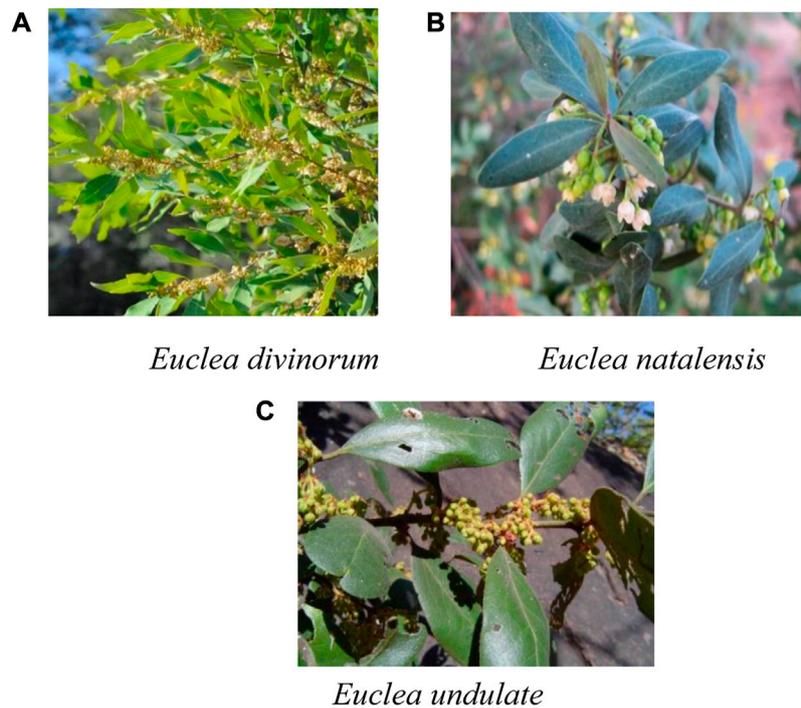


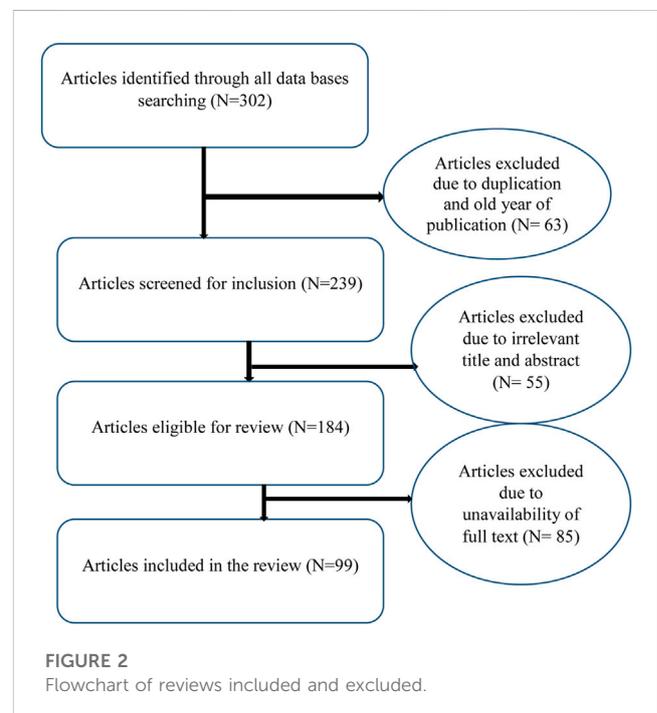
FIGURE 1

(A) *Euclea divinorum* (B) *Euclea natalensis* (C) *Euclea undulate*.

chest pain, pneumonia, gonorrhoea, constipation, edema, abdominal and chest pain (Feyissa et al., 2013; Woldemedhin et al., 2017; Mekonnen et al., 2018).

Botanical profile and taxonomy of *Euclea*

Most of the plants are trees, shrubs, and sub-shrubs, usually evergreen with alternate, opposite to sub-opposite, or in pseudo-whorls and diamond leaved (Figure 1A). Inflorescences: dioecious, axillary, or less frequently in branched pseudo-racemes, or flowers occasionally solitary (Figure 1B). Calyx: 4-5-lobed, usually polysepalous, not accrescent on fruits. Corolla: urceolate to subglobose, 5 - 8-lobed or campanulate and deeply 4-5-lobed. Stamens: 10-30; anthers dehiscing by large ellipsoidal apical pores, hairy or glabrous, oblong or lanceolate, 2-celled; filaments short, usually slender and glabrous. Staminodes: usually absent, glabrous; styles 2 (or 1, bifid), usually glabrous; stigmas bifid at apex. Ovary: ovoid or globular, hairy or glabrous, usually 4-celled; ovules 4, pendulous. Fruit: usually globose, 1-seeded berry (Halim et al., 2014), edible, spherical and one-seeded berries (Figure 1C) (Dhayalan et al., 2015). Many members of this genus are traditionally used to treat different diseases. Some are scientifically investigated for various biological activities and phytoconstituents. Previously, reviews that focus on single species, *E. undulata* Thunb (Maroyi, 2017) and *E. divinorum* Hiern (Omara et al., 2020) have been conducted. To the authors' knowledge, no study reviewed the ethnopharmacological use, phytochemistry, and biological activities of the whole genus.



Methodology

This review aims to critically evaluate available research reports on the genus and systematically organize and present the results. The review summarizes the existing knowledge on the

TABLE 1 Traditional uses of members of the genus *Euclea*.

Species	Part used	Indication	Country	References
<i>E. crispa</i>	R	cough	South Africa	Schmidt et al. (2002)
	R	constipation	South Africa	Chinsamy and Koitsiwe (2016)
			Swaziland	Long (2005)
	B, F, L, R	Diabetes	South Africa, Swaziland, Zimbabwe	Palmer and Pitman (1972) Chinsamy and Koitsiwe, 2016; Schmidt et al., 2002; Masoga, 2020
	L	Dysmenorrhoea	South Africa	Steenkamp (2003)
R	convulsions and epilepsy	Zimbabwe	Stafford et al. (2008)	
<i>E. coriacea</i>	B	Constipation, Stomach pains	South africa	Kose et al. (2015)
	R, L and S	Gonorrhea	Lesotho	Mugomeri et al., 2014
<i>E. divinorum</i>	R	HIV/AIDS-related diseases	Zambia	Chinsemu (2016)
	R	urine retention	Ethiopia	Woldemedhin et al. (2017)
	L	malaria, leprosy, gonorrhea, syphilis and tapeworm	Ethiopia	Geyid et al., 2005; Nanyingi et al., 2008
	R	induce labour	Kenya	Kaingu et al. (2012)
	F	abdominal upsets, skin, kidney and respiratory disorders	Kenya	Kigen et al. (2017)
	R	convulsions	Zimbabwe	Sobiecki (2002)
	R	schistosomiasis	South African	Sparg et al. (2000)
	<i>E. kellau</i>	B	splenic pain	Tanzania
R		Ancylostomiasis	Tanzania	Orzalesi et al. (1970)
L		snake-bite		Orzalesi et al. (1970)
<i>E. natalensis</i>	R	Diabetes	South Africa; Kenya	Deuschländer M. S. et al., 2009; Keter and Mutiso, 2012
	R	Epilepsy	South Africa	Van Wyk and Gericke (2000)
	L	Antidote for poisoning, snake bite	Malawi	Morris and Msonthi (1996)
	B	Prostate cancer	Uganda	Omara et al. (2020)
	R	Leprosy, syphilis, toothache	South Africa	Oosthuizen and Lall (2020)
	Sh and B	chest pains, bronchitis, pleurisy, and asthma	South Africa	Oosthuizen and Lall (2020)
	R	Herpes simplex virus 1	South Africa	Lall et al. (2005a)
	B	Tuberculosis	South Africa	Lall and Meyer (2001)
schistosomiasis		South Africa	Sparg et al. (2000)	
<i>E. racemosa</i>	R	Warts of the rectum, Constipation	Ethiopia	D'avigdor et al., 2014; Ayele et al., 2023
	RB	Cancer	Ethiopia	Wube et al. (2005)
	RB	toothache and malaria	Uganda	Neuwinger (2000)
	R	cancer, abdominal pain and convulsive dysmenorrhoea	Tanzania	Neuwinger (2000)
	RB, L, S, Br	Diabetes	Kenya	Keter and Mutiso (2012)
<i>E. schimper</i>	L	gonorrhea, eczema and constipation, snake biting, scabies, leprosy, Tinea capitis, acne, warts, rheumatic pain and elephantiasis	Ethiopia	Gelahun, 1989; Abebe and Ayehu, 1993; Mekonnen et al., 2018
	R	febrile disease (fever, headache and sweating)	Ethiopia	Giday et al. (2003)
	B	Dysmenorrhoea	South Africa	Steenkamp (2003)

(Continued on following page)

TABLE 1 (Continued) Traditional uses of members of the genus *Euclea*.

Species	Part used	Indication	Country	References
<i>E. undulata</i>	RB	Diabetes	South Africa	Deuschländer M. et al., 2009; Deuschländer, 2010
	B, R	Toothache	Botswana	Palmer and Pitman, 1972; Motlhanka and Nthoiwa, 2013
	L	Diarrhoea	South Africa	Deuschländer M. S. et al. (2009)
	S	Chewing stick	Zimbabwe	Joshua et al., 2013

AP, aerial part; B, bark; Br, Branch; F, flower; Fr, Fruit, L, leaf; R, root; RB, root bark; Sh, shoot; S, stem; Sd, Seed; WP, whole plant.

ethnobotanical use, phytochemistry, and pharmacological activity of species belonging to the genus *Euclea* to bring the reader up to date with the current literature. Articles on the species of the genus *Euclea* that reported ethnobotanical uses, biological activities, and isolation and identification of compounds were included. It is attempted to include articles published from 1975–2023 while some articles published before 1975 were also included by considering their importance. In this review articles where the full text was not available in the database or even after contacting the author by email were excluded (Figure 2).

This review excluded unpublished results and publications unavailable online, articles written in languages other than English, and articles whose titles and abstracts did not contain the search terms. Chemical structures of only isolated and characterized compounds were provided, while structures of compounds identified from essential oils and other chemical analyses were not. Different databases, including PubMed, Google Scholar, Scopus, and Medline, were employed to search literature using “keywords such as “*Euclea*”, “ethnobotanical use”, “phytochemistry”, and “pharmacological activity” dated up to December 2023.”

Ethno pharmacological uses

Ethnomedicinal claims on the genus *Euclea* to treat several ailments are illustrated in Table 1. The genus *Euclea* is used to treat hypnosis, toothache, headache (Bapela et al., 2008; Babula et al., 2009), chest complaints, bronchitis, pleurisy, chronic asthma, urinary tract infections, and venereal diseases (Lall & Meyer, 2000; Lall and Meyer, 2001; Weigenand et al., 2004; Kooy et al., 2006; Johanna, 2007; Bapela et al., 2008). An infusion of the roots of *E. ceispa* possesses antiepileptic activity (Dhayalan et al., 2015). The root bark of *E. undulata* is reported to be used for the management of body pains, diabetes, headache, and toothache while an infusion of its leaves is used for stomach problems or diarrhea, and leaf decoction for tonsillitis (Deuschländer M. et al., 2009; Dhayalan et al., 2015; Maroyi, 2017). This plant is a folk medicine for diabetes in the Venda area, Limpopo Province (Deuschländer M. S. et al., 2009; Babiaka et al., 2015; Maroyi, 2017). In the Western Cape, the root infusion of *E. undulata* is

used as enemata or as an ingredient of inembe (herbal medication regularly taken during pregnancy to ensure trouble-free confinement). Emesis or purgation is induced with root preparations (Deuschländer M. et al., 2009).

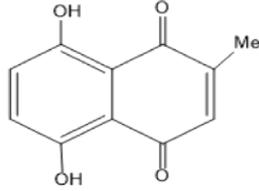
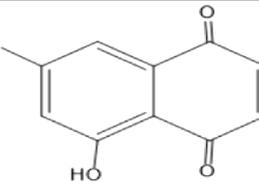
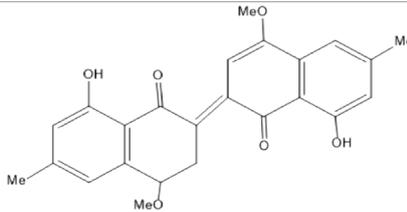
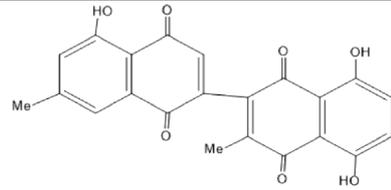
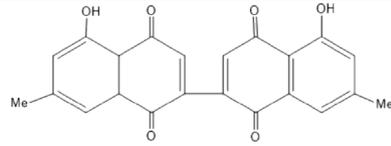
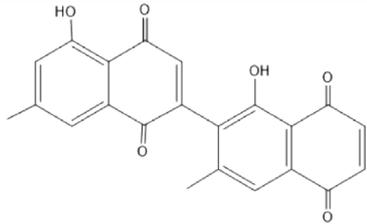
The Zulu people use *E. natalensis* as a purgative (Lall and Meyer, 2001; Weigenand et al., 2004) and for abdominal complaints in the form of infusion (Deuschländer M. S. et al., 2009; Deuschländer, 2010). Its charred and powdered root is used treat leprosy, urinary tract infections, venereal diseases, dysmenorrhea, and ancylostomiasis among Shangaan people (Lall & Meyer, 2000; Lall and Meyer, 2001); Kooy et al., 2006; Deuschländer, 2010) while its root bark infusions for sores and wounds in South Africa (Lall and Meyer, 2001). Within the Tonga people, the same part of this plant exhibits toothache and headache relief (Deuschländer M. et al., 2009; Babiaka et al., 2015; Dhayalan et al., 2015).

In Swaziland, the stem bark decoction of *E. divinorum* is a folk medicine for constipation (Amusan et al., 2007). The root bark is used for diarrhea, convulsions, cancer, and skin diseases (Mebe et al., 1998; Babiaka et al., 2015). In Kenya, the root of this plant is a remedy for chest pain, pneumonia, and internal body swelling (Woldemedhin et al., 2017). In Ethiopia, the roots and leaves of this plant are used for treating urinary retention, malaria, leprosy, gonorrhea, syphilis, and tapeworm (Feyissa et al., 2013; Woldemedhin et al., 2017). *E. schimperii* is traditionally prescribed for managing wounds, teeth infection, eye disorder, headache, gonorrhea, eczema, skin disorder, snake biting, scabies, leprosy, and elephantiasis in Ethiopia (Mekonnen et al., 2018).

Phytochemistry

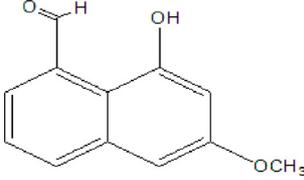
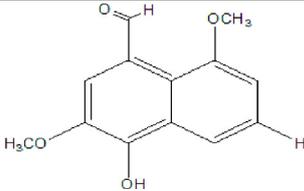
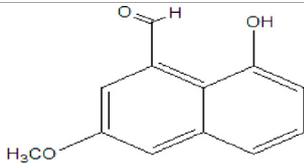
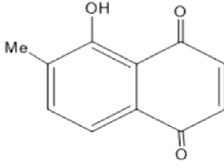
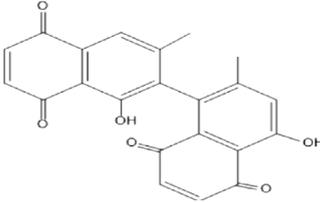
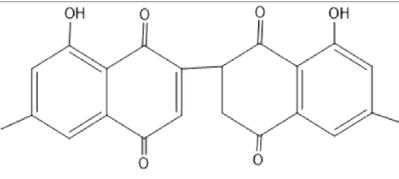
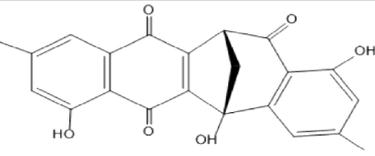
Euclea is a good source of naphthoquinones, pentacyclic triterpenes (Dagne et al., 1993; Joubert et al., 2006; Kwon et al., 2011; Dhayalan et al., 2015), flavonoids, naphthols (Dagne et al., 1993) and diosindigo (Dhayalan et al., 2015). Members of the genus *Euclea* contain primarily naphthoquinones and the root/root bark of the plant is the main source of the naphthoquinones. Phytochemical screening revealed that the leaf of *E. schimperii* contains saponins,

TABLE 2 Naphthoquinones isolated from *Euclea* species.

No.	Name of the compound	Species	Plant part	Structure	References
1.	2-methylnaphthazarin	<i>E. pseudebenus</i>	R		Ferreira et al., 1973; Dhayalan et al., 2015
2.	7-methyl-juglone	<i>E. undulata</i>	R		Deutschländer M. et al., 2009; Mbaveng & Kuete, 2014; Botha, 2016
		<i>3E.natalensis</i>	R		Mbaveng & Kuete, 2014; Deutschländer M. S. et al., 2009; Babula et al., 2009
		<i>4E. natalensis</i>	S, R and Sd		Lall et al., 2005b; Joubert et al., 2006; Johanna, 2007; Bapela et al., 2008; Babiaka et al., 2015; Kooy et al., 2006
		<i>5E. divinorum</i>	Us		Mebe et al., 1998; Babula et al., 2009
		<i>6E. divinorum</i>	R		Al-fatimi (2019)
		<i>7E. pseudebenus</i>	R		Ferreira et al. (1973)
		<i>8E. racemosa ssp. schimperi</i>	R		Wube et al. (2005)
		<i>9E. lanceolata</i>	RB		Ferreira et al. (1974)
3.	8,8'-dihydroxy-4,4'-dimethoxy-6,6'-dimethyl-2,2'-binaphthyl-1,1'-quinone	<i>E. lanceolata</i>	RB		Ferreira et al. (1974)
4.	8'-hydroxydiospyrin	<i>E. lanceolata</i>	RB		Ferreira et al. (1974)
5.	biramentaceone	<i>E. pseudebenus</i>	R		Ferreira et al. (1973)
6.	diospyrin	<i>E. undulata</i>	R		Deutschländer M. et al. (2009)
		<i>E. crispa var crispa</i>	R,Fr		Botha (2016)
		<i>E. undulata</i>	Fr		Botha (2016)
		<i>E. natalensis</i>	S, R and Sd		Lall et al., 2005a; Joubert et al., 2006; Johanna, 2007; Bapela et al., 2008; Babiaka et al., 2015
		<i>E. pseudebenus</i>	Us		Dhayalan et al. (2015)
		<i>E. divinorum</i>	R		Al-fatimi (2019)
		<i>E. natalensis</i>	Us		Deutschländer M. S. et al. (2009)

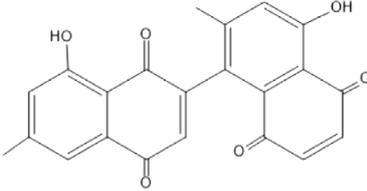
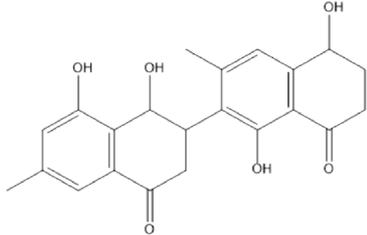
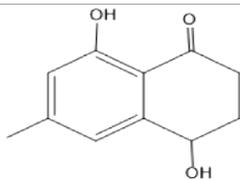
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TABLE 2 (Continued) Naphthoquinones isolated from *Euclea* species.

No.	Name of the compound	Species	Plant part	Structure	References
7.	eucleanal	<i>E. divinorum</i>	L		Ng'ang'a et al., 2012
8.	eucleanal B	<i>E. divinorum</i>	L		Mwihaki Ng'ang'a et al., 2012
9.	eucleanal A	<i>E. divinorum</i>	L		Mwihaki Ng'ang'a et al., 2012
10.	euclein (3, 6'-dimer of 7-methyljuglone.)	<i>E.pseudebenus</i>	R		Ferreira et al. (1973)
11.	isodiospyrin	<i>E. undulata</i>	Fr		Deutshländer M. et al., 2009; Botha, 2016
		<i>E. crista</i>	Fr		Botha (2016)
		<i>E. natalensis</i>	Us		Kooy et al., 2006; Wube et al., 2005
		<i>E. racemosa</i> ssp. <i>schimperi</i>	R		Wube et al., 2005; Al-fatimi, 2019
12.	mamegakinone	<i>E. natalensis</i>	Us		Kooy et al., 2006
		<i>E. natalensis</i>	Us		Deutshländer M. S. et al. (2009)
		<i>E. pseudebenus</i>	R		Ferreira et al., 197
		<i>E. racemosa</i> ssp. <i>schimperi</i>	R		Wube et al. (2005)
		<i>E. natalensis, E. crista, and E. schimperi</i>	Us		Dhayalan et al. (2015)
		<i>E. divinorum</i>	R		Al-fatimi (2019)
		<i>E. lanceolata</i>	RB		Ferreira et al. (1974)
13.	natalonone	<i>E. crista</i> subsp. <i>Crista</i>	Us		Kwon et al. (2011)

(Continued on following page)

TABLE 2 (Continued) Naphthoquinones isolated from *Euclea* species.

No.	Name of the compound	Species	Plant part	Structure	References
14.	neodiospyrin	<i>E. natalensis</i>	S, R and Sd		Joubert et al., 2006; Johanna, 2007; Bapela et al., 2008; Babiaka et al., 2015
		<i>15E. racemosa</i> ssp. <i>schimperi</i>	R		Wube et al. (2005)
15.	octahydroeuclein	<i>E. natalensis</i>	RB		Weigenand et al., 2004; Lall et al., 2006
16.	shinanolone	<i>E. natalensis</i>	RB		Weigenand et al., 2004; Kooy et al., 2006; Lall et al., 2006
		<i>18E. natalensis</i>	S, R and Sd		Joubert et al., 2006; Johanna, 2007; Bapela et al., 2008
		<i>19E. divinatorum</i>	Us		Mebe et al. (1998)
		<i>20E. racemosa</i> ssp. <i>schimperi</i>	R		Wube et al. (2005)
		<i>21E. natalensis</i>	Us		Kooy et al., 2006

B, bark; F, flower; Fr, Fruit; R, root; RB, root bark; S, shoot; Sd, Seed; Us, Unspecified.

terpenoids, tannins, steroids, polyphenols, and flavonoids after extraction with methanol and chloroform (Mekonnen et al., 2018). Aqueous and 80% methanol root extract of *E. divinatorum* had shown to contain saponins, flavonoids, glycosides, steroids, tannins, and terpenoids (Woldemedhin et al., 2017; Al-fatimi, 2019) but alkaloids and anthraquinones were absent (Woldemedhin et al., 2017). On the other hand, the root bark of this plant produces alkaloids, terpenoids, flavonoids, tannins, and saponins (Shumba, 2018). Methanol leaf and stem extracts of *E. undulata* contained alkaloids, diterpenes, glycosides, phytosterols, reducing sugars, saponins, and tannins (Maroyi, 2017). Essential oils, saponins, terpenoid derivatives, alkaloids, and flavonoids are the constituents of *E. crispa* subsp. *crispa* (Kwon et al., 2011).

Naphthoquinone

Quinones are one of the plant-derived secondary metabolites. Based on the number of benzene rings in the structural fused and skeleton, they are mainly classified as naphthoquinone, phenanthrenequinone, anthraquinone, and benzoquinone (Demir, 2020). Naphthoquinones are phenolic compounds derived from naphthalene occurring in plants (common) and fungi (Mbaveng &

Kuete, 2014; Botha, 2016). They were mainly detected from the root barks of the genus *Euclea* (Khan, 1985). Naphthoquinone isolated from the genus *Euclea* is presented in Table 2.

Flavonoids

Flavonoids are phenolic compounds having two benzene rings linked through a heterocyclic pyrane ring (Shumba, 2018). Quercetin, kaempferol (Al-fatimi, 2019), new aromadendrin-3-O-β-L-arabinopyranoside (17), and known flavonoids such as catechin (Dagne et al., 1993; Mebe et al., 1998), myricetin-3-O-α-L-rhamnopyranoside (21) and quercetin-3-O-α-L-rhamnopyranoside (22) were isolated from the extract of ethanol aerial part of *E. divinatorum* (Dagne et al., 1993), (Table 3). Acetone leaves extract of *E. racemosa* ssp. *Schimperi* yields quercetin, myricitrin, myricetin-3-O-arabinopyranoside (20) and rutin (23), (Asres et al., 2006).

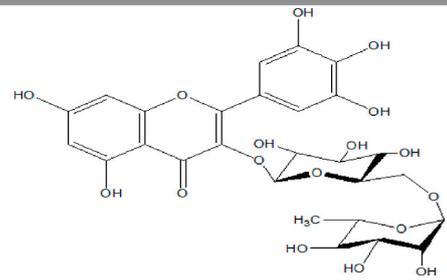
HPLC detects large amounts of myricitrin and small amounts of isoquercitrin and quercitrin in *E. schimperi* (Mueller-harvey et al., 1987). Root bark extracts of *E. undulata* (acetone) (Deutschländer, 2010; Babiaka et al., 2015; Botha, 2016), *E. divinatorum* (chloroform) and *E. undulata* (acetone) resulted in the isolation of epicatechin (19) and catechin (18) respectively (Babiaka et al., 2015).

TABLE 3 Flavonoids isolated from *Euclea* species.

No.	Name of the compound	Species	Plant part	Structure	References
17	aromadendrin-3-O- β -L-arabinopyranoside	<i>E. divinatorum</i>	A		Dagne et al., 1993
18	catechin	<i>E. divinatorum</i>	RB		Babiaka et al. (2015)
19	epicatechin	<i>E. undulate</i>	RB		Deuschländer, 2010; Babiaka et al., 2015; Botha, 2016
20	myricetin-3-O-arabinopyranoside	<i>E. racemosa</i>	L		Asres et al. (2006)
21	myricetin-3-O- α -L-rhamnopyranoside	<i>E. divinatorum</i>	A		Dagne et al., 1993
22	quercetin-3-O- α -L-rhamnopyranoside	<i>E. divinatorum</i>	A		Dagne et al., 1993

(Continued on following page)

TABLE 3 (Continued) Flavonoids isolated from *Euclea* species.

No.	Name of the compound	Species	Plant part	Structure	References
23	rutin	<i>E. racemosa</i>	L		Asres et al. (2006)

A, aerial; RB, root bark; L, leaf.

Hyperoside, quercitrin, epicatechin, catechins and gallic acid were isolated from the leaves of *E. crispa* subsp. *Crispa* (Rademana et al., 2019).

Terpenoids

Triterpenes are a group of natural products, derived from isoprene units. In nature, triterpenoids are often existed as tetra- or penta-cyclic structures but some acyclic, mono-, bi-, tri- and hexa acyclic. As described in Table 4, Lupeol, lupine, botulin and oleanolic acid are some examples of pentacyclic triterpenoids (Furtado et al., 2017). Triterpenoids were detected from root and stem barks of *E. natalensis* (Khan, 1985). Phytol (0.66%) and squalene (5.85%) were detected from hexane extract of *E. crispa* using GC-MS (Palanisamy & Ashafa, 2018).

Miscellaneous

The following bioactive compounds with their composition were identified from hexane extract of *E. crispa* using GC-MS: tetracosane (14.98%), dodecane (10.76%), 2-ethyl-1-decanol (8.00%), tridecane (7.53%), diphenyl vinyl phosphine (6.38%), triacontane (5.27%), 2,6-dimethylheptadecane (5.02%), docosane (3.68%), tetradecane (3.59%), 1-hepten-3-ol (2.63%), orthotolidine (2.31%), Phenyl glucuronide (2.25%), 5-tridecylbenzene-1,3-diol (1.90%), and Pentadecane (1.68%) (Palanisamy & Ashafa, 2018). Vitamin E, fatty acid methyl esters such as saturated (C₁₄, C₂₀) and unsaturated (C₁₆, C_{18:1}, C_{18:2}, and C_{18:3}) were isolated from twigs and leaves of *E. undulate* (Maroyi, 2017). VTLC identified gallic and ellagic acid esters in *E. schimperi* (Mueller-harvey et al., 1987).

Biological activities

Antimicrobial activity

The acetone and aqueous extract of *E. natalensis* inhibited the growth of *Bacillus cereus*, *B. pumilus*, *B. subtilis*, *Micrococcus kristinae*, and *Staphylococcus aureus* at concentrations ranging between 0.1 and 6.0 mg/mL (Lall and Meyer, 2000). Isolated compounds from the root extract also demonstrated a significant

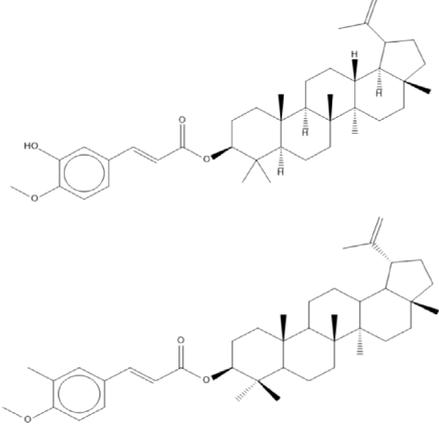
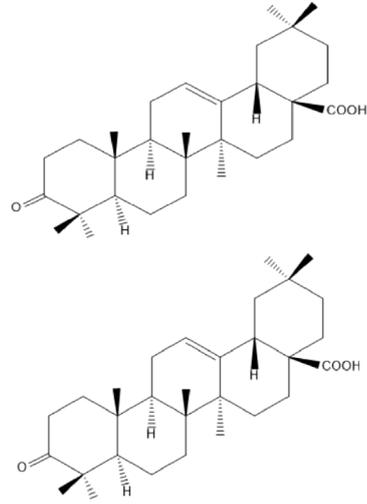
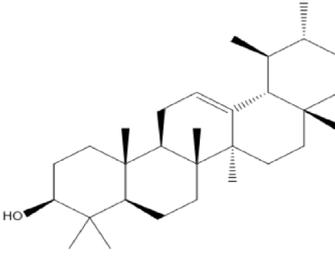
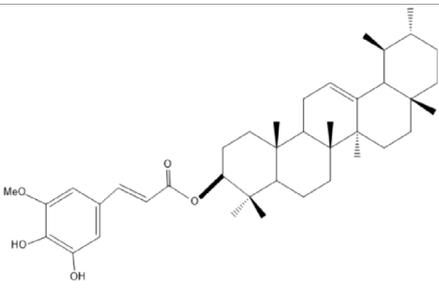
antimicrobial effect. Diospyrin and 7-methyljuglone were more effective against Gram-positive bacteria than Gram-negative bacteria.

Shinanolone, 7-methyljuglone, diospyrin, isodiospyrin and neodiospyrin in the genus *Euclea* especially *E. natalensis* are potent for the treatment of both drug-sensitive and resistant tuberculosis (Joubert et al., 2006; Johanna, 2007; Bapela et al., 2008; Babula et al., 2009; Babiaka et al., 2015). On the other hand, diospyrin, lupeol, betulin and 7-methyl juglone presented in *E. natalensis* has inhibitory activity against drug-sensitive *M. tuberculosis* at MIC of 8.0 and 0.5 mg/mL respectively (Maroyi, 2017). The intracellular and extracellular inhibition of the latter compound is greater than that of the anti-tuberculosis drugs streptomycin and ethambutol (Lall et al., 2005b; Mcgaw et al., 2008).

7-methyl juglone and mamegakinone are effective against *M. tuberculosis* (Kooy et al., 2006), *Neisseria gonorrhoeae*, *Shigella dysenteriae* and *Shigella flexneri*. Aqueous and acetone extracts of the roots of *E. natalensis* inhibited the growth of *Mycobacterium tuberculosis* at MIC value of 0.5 mg/mL while MIC values for *B. cereus*, *B. pumilus*, *B. subtilis*, *M. kristinae* and *S. aureus* ranged from 0.1–6.0 mg/mL (Lall & Meyer, 2000; Lall and Meyer, 2001). 7-methyl juglone is also effective against *Saccharomyces cerevisiae*, *M. bovis*, *M. smegmatis* and *M. fortuitum* (Mbaveng & Kuete, 2014). Due to Shinanolone, *E. natalensis* inhibits the growth of Gram-positive bacterial strains and a drug-sensitive strain of *M. tuberculosis* at a concentration of 0.1 mg/mL (Weigenand et al., 2004).

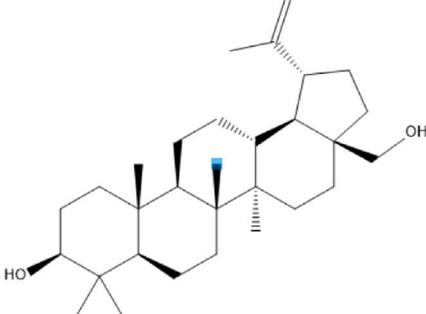
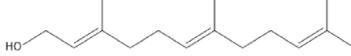
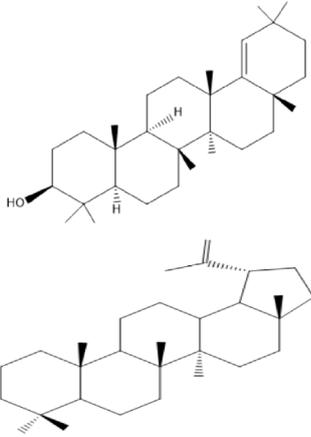
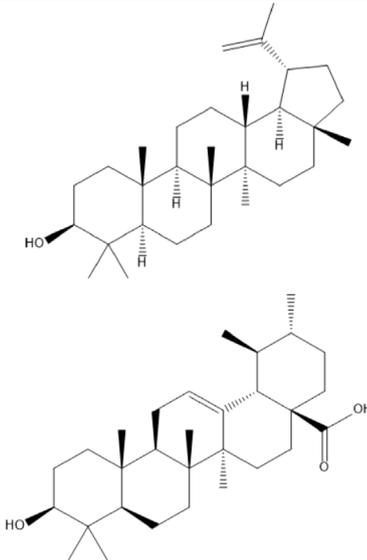
Ethanol extract of *E. crispa* leaves elicit antimicrobial activity with maximum inhibition zone against *Staphylococcus aureus*, *Streptococcus aureus*, *Escherichia coli*, *Klebsiella pneumoniae*, *Aspergillus niger* and *Aspergillus terreus* (Palanisamy et al., 2019). Previous literatures demonstrated that *E. lanceolata*, *E. undulata* and *E. multiflora* possess antifungal activity due to the presence of lawsone, juglone and 7-methyljuglone (Lall & Meyer, 2000; Lall and Meyer, 2001). *Euclea natalensis* comprises β -sitosterol (Lall et al., 2006; Moosavi et al., 2020), 20 (29)-lupine-3 β -isoferulic and shinanolone that have inhibitory activity against *Aspergillus niger* at 0.01 mg/mL. The former compound and octahydro euclein significantly show fungistatic activity against *C. cladosporioides* at 0.01 mg/mL. Besides this, octahydro euclein present in this plant is very effective for *Phytophthora* sp. at 0.1 mg/mL (Lall et al., 2006).

TABLE 4 Terpenoids isolated from *Euclea* species.

No	Name of the compound	Species	Plant part	Structure	References
24	20 (29)-lupene-3 α -isoferulate	<i>E. natalensis</i>	RB		Weigenand et al. (2004)
25	20 (29)-lupene-3 β -isoferulate	<i>E. natalensis</i>	RB		Lall et al. (2006)
26	3-oxo-oleanolic acid	<i>E. crispa</i> subsp. <i>crispa</i>	Us		Kwon et al. (2011)
27	3 β -(5-hydroxyferuloyl)lup-20 (30)-ene	<i>E. crispa</i> subsp. <i>crispa</i>	Us		Kwon et al. (2011)

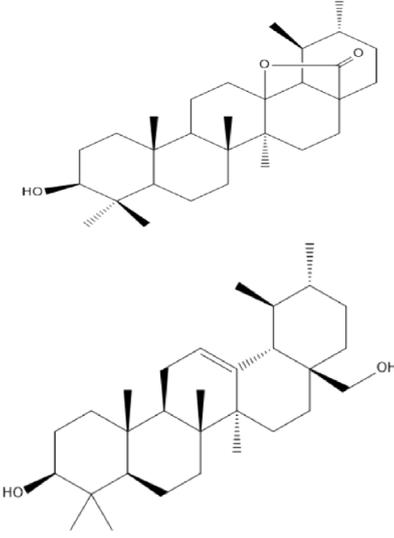
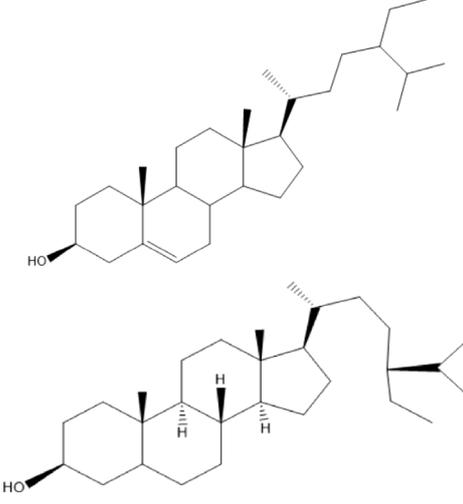
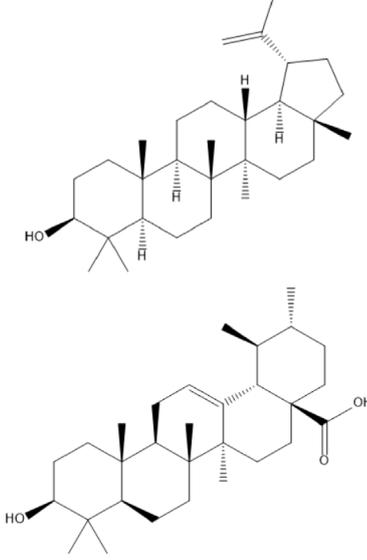
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TABLE 4 (Continued) Terpenoids isolated from *Euclea* species.

No	Name of the compound	Species	Plant part	Structure	References
28	α -amyrin	<i>E. kellaui</i>	L		Orzalesi et al. (1970)
29	α -amyrin-3O- β -(5-hydroxy) ferulic acid	<i>E. undulata</i>	RB		Deutschländer, 2010; Botha, 2016
30	betulin	<i>E. natalensis</i>	RB		Weigenand et al., 2004; Lall et al., 2005b; Lall et al., 2006
		<i>E. divinorum</i>	RB		Mebe et al., 1998; Al-fatimi, 2019
		<i>E. kellaui</i>	Br		Orzalesi et al. (1970)
		<i>E. latideus</i>	R		Philip et al. (2018)
		<i>E. undulata</i>	RB		Deutschländer, 2010
		<i>E. crispa</i> subsp. <i>crispa</i>	RB		Rademana et al. (2019)
31	farnesol	<i>E. crispa</i>	L		Palanisamy et al. (2020)

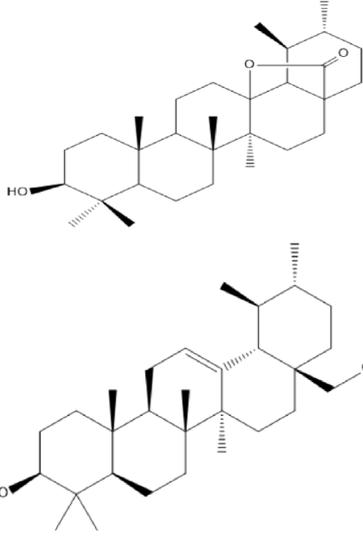
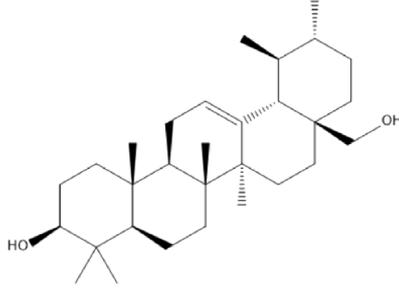
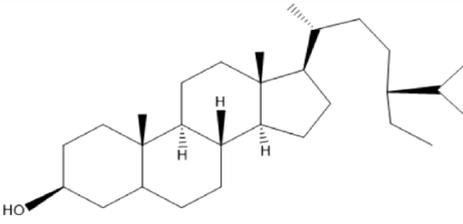
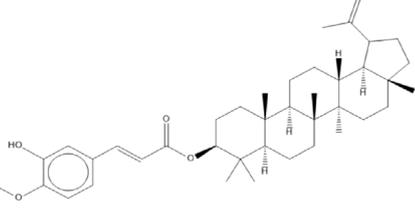
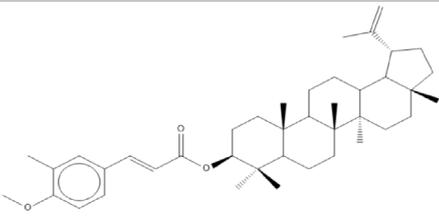
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TABLE 4 (Continued) Terpenoids isolated from *Euclea* species.

No	Name of the compound	Species	Plant part	Structure	References
32	germanicol	<i>E. divinorum</i>	RB		Kilonzo et al. (2019)
33	lupene	<i>E. divinorum</i>	RB		Mebe et al., 1998; Al-fatimi, 2019
34	lupeol	<i>E. natalensis</i>	RB		Weigenand et al., 2004; Lall et al., 2005a; Lall et al., 2006
		<i>E. divinorum</i>	RB		Mebe et al., 1998; Al-fatimi, 2019
		<i>E. kellau</i>	Br		Orzalesi et al. (1970)
		<i>E. latideus</i>	R		Philip et al. (2018)
		<i>E. undulata</i>	RB		Deutschländer, 2010; Botha, 2016

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TABLE 4 (Continued) Terpenoids isolated from *Euclea* species.

No	Name of the compound	Species	Plant part	Structure	References
35	ursolic acid	<i>E. kellaui</i>	Br, L		Orzalesi et al. (1970)
36	ursolic acid lactone	<i>E. kellaui</i>	Br, L		Orzalesi et al. (1970)
37	uvaol	<i>E. kellaui</i>	L		Orzalesi et al. (1970)
38	β -sitosterol	<i>E. natalensis</i>	RB		Lall et al., 2006; Moosavi et al., 2020
39	γ -sitosterol	<i>E. divinorum</i>	RB		Kilonzo et al. (2019)

Br, Branch; L, leaf; R, root; RB, root bark; Us, Unspecified.

Ethyl acetate root extract of *E. divinorum* has inhibitory activity against Gram-negative bacteria like *E. coli* but is ineffective for *S. aureus*. Alkaloids and terpenoids in this plant contribute to this kind of antibacterial activity (Shumba, 2018). The MIC values of the extracts of *E. divinorum* against bacterial activity for root bark ethyl acetate and leaf aqueous ranges from 0.048–0.871 mg/mL and 0.781–1.562 mg/mL respectively. The first extract is very effective against *S. typhi* followed by stem bark aqueous and root bark petroleum ether extract against *S. aureus* (Kilonzo et al., 2019).

The non-polar dichloromethane root extract of *E. divinorum* root bark has better antifungal activity than the nystatin for *Absidia corymbifera*, *Aspergillus fumigatus*, *Candida krusei*, *Microsporium gypseum*, *Mucor sp.* and *Trichophyton mentagrophytes*. This activity is maintained with lupeol, lupine, botulin, 7-methyl juglone, diospyrin, iso diospyrin and shinalone (Al-fatimi, 2019).

Antiviral activity

The acetone extract of *E. natalensis* demonstrated moderate antiviral activity against HSV-1, at concentrations of 0.1–0.02 mg ml⁻¹ (Lall et al., 2005a). In a study conducted by Tshikalange et al. (2007) 7-methyljuglone (potent), diospyrin, neodiospyrin, isodiospyrin, and 6-methyljuglone isolated from that *E. natalensis* exhibited HIV-1 reverse transcriptase activity at the concentrations ranging from 25 to 50 µg/mL. The leaf extract of *E. schimperi* showed good antiviral activity against Influenza A virus and herpes simplex virus (HSV-1) with IC₅₀ values of 6.22 6 µg/mL and 67.5 µg/mL, respectively (Gebre-Mariam et al., 2006).

Antimalarial activity

Aqueous, dichloromethane, and methanol leaf and twig extracts of *E. undulata* have shown antimalarial activity against *Plasmodium falciparum* using the parasite lactate dehydrogenase assay (Maroyi, 2017). *E. latideus* is also effective against *P. falciparum* especially for the chloroquine resistant strain of *P. falciparum* due to the presence of lupeol, betulin, and 3β-(5-hydroxy feruloyl) lup-20 (30)-ene (Philip et al., 2018). The dichloromethane and methanol (1:1) root and leaf extracts of *E. natalensis* demonstrated promising activity in a research by Clarkson et al. (2004) employing the parasite lactate dehydrogenase assay, with (IC₅₀) values of 5.1 and 5.3 mg/mL, respectively, against *P. falciparum*. A study done by Philip et al. (2018) indicated that the extracts and isolated compounds from *E. latideus* demonstrated antiplasmodial activity against chloroquine sensitive and chloroquine resistant strains of *P. falciparum*. The leaves of *E. natalensis* also showed antiplasmodial activity with an IC₅₀ of 25.6 µg/mL (Tajuddeen et al., 2022). The *in vivo* antimalarial assay of the aqueous root extract of *E. divinorum* possessed significant parasitemia suppression (Girmaw and Engidawork, 2022).

Antidiabetic activity

E. undulata containing α-amyrin-3-O-β-(5-hydroxy) ferulic acid inhibits α-glucosidase and epicatechin lowers glucose levels in the blood (Botha, 2016). Phenolic acids and flavonoids of *E. crispa* inhibit alpha amylase with IC₅₀ values of 1.001 mg/mL and 1.65 mg/mL (Tinevimbo, 2017). Lowering of blood glucose

can be achieved with acetone root bark extracts of *E. undulata* by displaying a glucose uptake of 162.2% by changing liver cells at 50 mg/mL (Maroyi, 2017). *E. coriacea* contains phytosterols that possess antidiabetic activity (Mugomeri et al., 2014). Acetone root bark extracts of *E. undulata* effectively reduced fasting blood glucose levels, raised cholesterol, and triglyceride levels to close to normal without causing weight gain in an *in vivo* model of streptozotocin-nicotinamide-induced type-2 diabetes (Deutschländer et al., 2012).

Antioxidant activity

Ethanol root bark and leaf extracts of *E. crispa* have radical scavenging activity because of flavonoids, phenolics (Tinevimbo, 2017) and (6E, 10E)-2, 6, 24-trimethylpentane cosa-2, 6, 10-triene isolated from the leaves of *E. crispa* exhibited potent antioxidant activity (Palanisamy et al., 2019). The leaves of *E. crispa* were tested for antioxidant activity and showed IC₅₀ values of 113.79, 109.59, and 116.65 µg/mL for DPPH, hydroxyl and nitric oxide radical scavenging assays. Farnesol contributes to such activity (Palanisamy et al., 2020). At a 2000 mg/mL concentration, *E. divinorum* inhibits DPPH by 82.5%, 74.5% and 62.5% for the methanol fraction, aqueous fraction and crude extract, respectively (Feyissa et al., 2013). Fatty acids, flavonoids, and phenolics of *E. undulata* showed antioxidant activity using the DPPH, ABTS and FRAP assays (Maroyi, 2017). The free radical scavenging effect of methanol and chloroform leaf extracts of *E. schimperi* was demonstrated. The methanol and chloroform extracts were able to scavenge the DPPH radical with a percentage scavenging activity of 85.4% and 58.5% at the concentration of 40 µg/mL, respectively (Mekonnen et al., 2018).

Anticancer activity

The leaves of *E. crispa* subsp. *crispa* extract exhibited anti-proliferative activity on human breast adenocarcinoma (MCF-7) and human epidermoid carcinoma (A431) cell lines with IC₅₀ values of 45.7 µg/mL and 41.8 µg/mL, respectively (Rademana et al., 2019). 7-methyl juglone and 3β-(5-hydroxy feruloyl) lup-20 (30)-ene, which are the main constituents of *E. divinorum*, showed anticancer effects against human breast cancer, colon cancer, fibrosarcoma, nasopharyngeal carcinoma, lung cancer, and human melanoma (Mebe et al., 1998). Diterpenes isolated from *E. coriacea* has been reported to possess an anticancer effect in human cells (Mugomeri et al., 2014). 7-Methyl juglone isolated from *E. racemosa* ssp. *schimperi* has been described to possess significant cytotoxic properties against human colon carcinoma cells (Wube et al., 2005). *Euclea natalensis* also contains this compound that has anticancer activity on several cancer cell lines, such as KB, Lu1, and LNCaP (Mbaveng & Kuete, 2014).

Other activities

E. coriacea contains phytosterols that possess anti-inflammatory and anti-pain activity (Mugomeri et al., 2014). A study showed that *E. natalensis* shoot extract has *in vivo* hepatoprotective activity by reducing the level of alanine

transaminase liver enzyme by 15% (50 mg/kg) and 40% (100 mg/kg). This plant also provides an immunomodulatory activity by increasing T-helper 1 cell cytokines such as Interleukin 2, Interleukin 12, and Interferon α by 12 fold and decreasing the T-helper 2 cell cytokine, interleukin 10 by 4 fold when compared to baseline cytokine production (Lall et al., 2016). The *in vivo* evaluation of the antidiuretic activity of *E. divinorum* revealed that the aqueous and methanol root extract of the plant possessed a significant diuretic activity by increasing urine volume and electrolyte excretion (Woldemedhin et al., 2017). Feyissa et al. (2013) demonstrated that the crude extract and solvent fractions of *E. divinorum* leaves restored gentamicin-induced nephrotoxicity by decreasing tubular necrosis, serum and oxidant markers and by increasing in antioxidant molecules. The methanol fraction provided the most renoprotection, implying that semi-polar antioxidant principles may be involved.

Acute toxicity, gentotoxicity and cytotoxicity

Acute toxicity studies of the crude and methanolic extract of *E. divinorum* leaves indicated that it was safe when administered orally at 2000 mg/kg (Feyissa et al., 2013; Woldemedhin et al., 2017). After a period of 72 h, the animals tolerated the administered dose, and there were no appreciable changes in behavior such as motor activity, diarrhoea, breathing, alertness, restlessness, convulsions, coma and appearance. Since no mortality was recorded within 14 days, the lethal dose (LD₅₀) was indicated to be more than 2000 mg/kg. Shauli, (2023) evaluated the acute and sub-acute oral toxicity of *E. natalensis* and the results demonstrate that no treatment related deaths or toxic signs were observed. Another study done by Ayele et al. (2023) revealed that *E. racemosa* was safe after oral toxicity study with LD₅₀ greater than 2000 mg/kg. *E. latideus* is considered as a non-toxic plant since acute toxicity studies showed that crude extracts had LD₅₀ > 5,000 mg/kg (Kodi et al., 2018).

Taylor et al. (2003) investigated genotoxicity in human peripheral blood lymphocytes of South African medicinal plants. The results reported that the dichloromethane root extract of *E. divinorum* induced DNA damage (more cells with high tail DNA content), which was however lower than that of the positive control (1 mM potassium bichromate). However, the bark extract of *E. natalensis* showed positive results for genotoxicity in the micronucleus test.

References

- Abebe, D., and Ayele, A. (1993). "Medicinal plants and enigmatic health practices of Northern Ethiopia," in *Addis ababa, Ethiopia: B. S. P. E.*, 511.
- Ahvazi, M., Charkchiyan, M., Khalighi-Sigaroodi, F., Mojab, F., Mozaffarian, A., and Zakeri, H. (2012). Introduction of medicinal plants species with the most traditional usage in Alamut Region. *Iran. J. Pharm. Res.* 11 (1), 185–194.
- Al-fatimi, M. (2019). Antifungal activity of *Euclea divinorum* root and study of its ethnobotany and phytopharmacology. *Processes* 7, 680. doi:10.3390/pr7100680
- Amusan, O., Sukati, N., Dlamini, P., and Sibandze, F. (2007). Some Swazi phytomedicines and their constituents. *Afr. J. Biotechnol.* 6 (3), 267–272.
- Asres, K., Gibbons, S., and Bucar, F. (2006). Radical scavenging compounds from Ethiopian medicinal plants. *Ethiop. Pharm. J.* 24, 23–30. doi:10.4314/epj.v24i1.35095
- Ayele, A. G., Mulugeta, B., and Wondmkun, Y. T. (2023). Evaluations of the *in vivo* laxative effects of aqueous root extracts of *Euclea racemosa* L. in mice. *Metab. Open* 17, 100222. doi:10.1016/j.metop.2022.100222
- Babiaka, S., Ntie-kang, F., Ndingkokhar, B., Mbah, J., Sippl, W., and Yong, J. (2015). The chemistry and bioactivity of southern african flora II: Flavonoids, quinones and minor compound classes. *RSC Adv.* 5, 57704–57720. doi:10.1039/c5ra05524e
- Babula, P., Adam, V., Havel, L., and Kizek, R. (2009). Noteworthy secondary metabolites naphthoquinones – their occurrence, pharmacological properties and analysis. *Curr. Pharm. Anal.* 5, 47–68. doi:10.2174/157341209787314936
- Bapela, J., Kuete, V., Toit, E., Meyer, M., and Lall, N. (2008). Fertilization-induced changes in growth parameters and antimicrobial activity of *Euclea natalensis* (Ebenaceae). *S. A. J. Bot.* 74, 244–250. doi:10.1016/j.sajb.2007.11.011
- Bapela, M. J. (2008). *Variation of active constituents in Euclea natalensis based on seedling stages, seasons, and fertilizers* (Doctoral dissertation, University of Pretoria).
- Botha, L. E. (2016). *Investigating the production of secondary metabolites effective in lowering blood glucose levels in Euclea Undulata Thunb. Var Myrtina* (Ebenaceae) (Doctoral dissertation).

Conclusion

The genus *Euclea* is well known for its use in the treatment of diabetic and body pain manifestations. The traditional claims were justified by different biological evaluations. The genus *Euclea* is known to be a source of biologically active compounds. More than 40 compounds were isolated from the genus and naphthoquinones, pentacyclic triterpenes and flavonoids are the most abundant bioactive secondary metabolites which are responsible for the observed biological activity. Most of these secondary metabolites are found in the roots and root bark while some in fruit, seeds, leaves and shoots. According to the present review, it has been noted that the potential uses of the species in the treatment of viral infections and nerve-related diseases have not been scientifically explored. We believe the scientific community researching on the genus will benefit from the material compiled in this review.

Author contributions

AT designed the study, conducted the literature review, extracted relevant information to the study, write the manuscript. AK and GB contributed in writing, editing and revising the manuscript. All authors read and approved the manuscript.

Conflict of interest

The authors declare that they no conflict of financial interests or personal relationships that could have appeared to influence the work reported in this review.

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- Chinsamy, M., and Koitsiwe, M. (2016). Traditional knowledge of medicinal and food plant uses for sustainable community livelihoods: A case of batswana communities in South Africa. *J. Soc. Sci.* 46 (2), 146–154. doi:10.1080/09718923.2016.11893522
- Chinsembu, K. C. (2016). Ethnobotanical study of plants used in the management of HIV/AIDS-related diseases in Livingstone, Southern Province, Zambia. *Evid. Based Complement. Altern. Med.* 2016, 4238625. Article ID 4238625. doi:10.1155/2016/4238625
- Clarkson, C., Maharaj, V. J., Crouch, N. R., Grace, O. M., Pillay, P., Matsabisa, M. G., et al. (2004). *In vitro* antiparasitic activity of medicinal plants native to or naturalised in South Africa. *J. Ethnopharmacol.* 92 (2–3), 177–191. doi:10.1016/j.jep.2004.02.011
- Dagne, E., Alemu, M., and Serner, O. (1993). Flavonoids from *Euclea divinorum*. *Bull. Chem. Soc. Ethiop.* 7 (2), 87–92.
- D'avigdor, E., Wohlmut, H., Asfaw, Z., and Awas, T. (2014). The current status of knowledge of herbal medicine and medicinal plants in Fiche, Ethiopia. *J. Ethnobiol. Ethnomedicine* 10 (1), 38–33. doi:10.1186/1746-4269-10-38
- Demir, Y. (2020). Naphthoquinones, benzoquinones and anthraquinones: Molecular docking, ADME and inhibition studies on human serum paraoxonase-1 associated with cardiovascular diseases. *Drug Dev. Res.* 81 (5), 628–636. doi:10.1002/ddr.21667
- Deuschländer, M., Lall, N., and Venter, M. (2009). Plant species used in the treatment of diabetes by South African traditional healers: An inventory. *Pharm. Biol.* 47 (4), 348–365. doi:10.1080/13880200902752959
- Deuschländer, M. S., Van de Venter, M., Roux, S., Louw, J., and Lall, N. (2009). Hypoglycaemic activity of four plant extracts traditionally used in South Africa for diabetes. *J. Ethnopharmacol.* 124 (3), 619–624. doi:10.1016/j.jep.2009.04.052
- Dhayalan, M., Jegadeeshwari, A., and Gandhi, N. (2015). Biological activity sources from traditionally used Tribe and herbal plants material. *Asian J. Pharm. Clin. Res.* 8 (6), 11–23.
- Ferreira, A., Lopes, M., Costa, M., and Alves, C. (1974). Eucleolatin: A dimeric methyl naphthazarin from *Euclea lanceolata*. *Phytochemistry* 13, 499–501. doi:10.1016/s0031-9422(00)91243-3
- Ferreira, M., Costa, M., Alves, A., and Lopes, M. (1973). Euclein: A new naphthoquinone from *Euclea pseudebenus*. *Phytochemistry* 12, 433–435. doi:10.1016/0031-9422(73)80035-4
- Feyissa, T., Asres, K., and Engidawork, E. (2013). Renoprotective effects of the crude extract and solvent fractions of the leaves of *Euclea divinorum* Hierns against gentamicin-induced nephrotoxicity in rats. *J. Ethnopharmacol.* 145 (3), 758–766. doi:10.1016/j.jep.2012.12.006
- Furtado, N., Pirson, L., Edelberg, H., Miranda, L., Loira-Pastoriza, C., Preat, V., et al. (2017). Pentacyclic triterpene bioavailability: An overview of *in vitro* and *in vivo* studies. *Molecules* 22 (3), 1–24.
- Gebre-Mariam, T., Neubert, R., Schmidt, P. C., Wutzler, P., and Schmidtke, M. (2006). Antiviral activities of some Ethiopian medicinal plants used for the treatment of dermatological disorders. *J. Ethnopharmacol.* 104 (1–2), 182–187. doi:10.1016/j.jep.2005.08.071
- Gelahun, A. (1989). "Etse debdabe, Ethiopian traditional medicine," in *Biology department, science faculty, addis ababa university*. Editor D. Sebsebe, 64–123.
- Geyid, A., Abebe, D., Debella, A., Makonnen, Z., Aberra, F., Tekla, F., et al. (2005). Screening of some medicinal plants of Ethiopia for their anti-microbial properties and chemical profiles. *J. Ethnopharmacol.* 97 (3), 421–427. doi:10.1016/j.jep.2004.08.021
- Gayid, M., Asfaw, Z., Elmquist, T., and Woldu, Z. (2003). An ethnobotanical study of medicinal plants used by the Zay people in Ethiopia. *J. Ethnopharmacol.* 85 (1), 43–52. doi:10.1016/s0378-8741(02)00359-8
- Girmaw, F., and Engidawork, E. (2022). *In vivo* anti-malarial activity of the aqueous root extract of *Euclea divinorum* hiern (Ebenaceae) against *Plasmodium berghei* ANKA. *Evid. Based Complement. Altern. Med.* 2022, 2640648. Article ID 2640648. doi:10.1155/2022/2640648
- Gowdhani, T., Rajalakshmi, A. K., and Sugumar, N. (2015). Pharmacognostical and preliminary phytochemical screening of the leaf extract of *Jasminum auriculatum* Vahl. *Int. Lett. Nat. Sci.* 43, 69–75. doi:10.56431/p-23h92l
- Halim, A., Mohamed, A., Habeeb, R., Azer, A., Safwat, R., and Hafeez, A. (2014). Taxonomic revision of Ebenaceae in Egypt. *Curr. Sci. Int.* 3 (4), 414–425.
- Jackson, P. W., and Miller, J. (2015). Developing a world flora online - a 2020 challenge to the world's botanists from the international community. *Rodriguésia* 66 (4), 939–946. doi:10.1590/2175-7860201566402
- Joubert, A., Kooy, F., Meyer, J. M., and Lall, N. (2006). HPLC in the comparative study of the content of naphthoquinones (quinonoid constituents) in *Euclea* species of South Africa. *Chromatographia* 64 (7/8), 399–403. doi:10.1365/s10337-006-0055-z
- Kaingu, C. K., Oduma, J. A., and Kanui, T. (2012). Preliminary investigation of contractile activity of *Ricinus communis* and *Euclea divinorum* extracts on isolated rabbit uterine strips. *J. Ethnopharmacol.* 142 (2), 496–502. doi:10.1016/j.jep.2012.05.026
- Karunamoorthi, K., Jagajeevanam, K., Vijayalakshmi, J., and Mengistie, E. (2013). Traditional medicinal plants: A source of phytotherapeutic modality in resource-constrained health care settings. *J. Evid. Based Complement. Altern. Med.* 18 (1), 67–74. doi:10.1177/2156587212460241
- Keskin, C. (2018). Medicinal plants and their traditional uses. *J. Adv. plant Biol.* 1 (2), 8–12. doi:10.14302/issn.2638-4469.japb-18-2423
- Keter, L. K., and Mutiso, P. C. (2012). Ethnobotanical studies of medicinal plants used by traditional health practitioners in the management of diabetes in lower eastern Province, Kenya. *J. Ethnopharmacol.* 139 (1), 74–80. doi:10.1016/j.jep.2011.10.014
- Kigen, G., Kipkore, W., Wanjohi, B., Haruki, B., and Kemboi, J. (2017). Medicinal plants used by traditional healers in sangurur, elgeyo marakwet county, Kenya. *Pharmacogn. Res.* 9 (4), 333–347. doi:10.4103/pr.pr_42_17
- Kilonzo, M., Rubanza, C., Richard, U., and Sangiwa, G. (2019). Antimicrobial activities and phytochemical analysis of extracts from *Ormocarpum trichocarpum* and *Euclea divinorum* used as traditional medicines in Tanzania. *Tanzan J. Health Res.* 21 (2), 1–12. doi:10.4314/thrb.v21i2.6
- Kodi, P., Mwangi, M. E., Cheplogoi, P. K., Langat, M., and Hoseah, M. A. (2018). *In vitro* antiplasmodial and toxicity activities of crude extracts and compounds from *Euclea latideus* (Ebenaceae). *Int. J. Biochem. Res. Rev.* 21 (1), 1–22. doi:10.9734/ijbcr/2018/39603
- Kose, L. S., Moteetee, A., and Van Vuuren, S. (2015). Ethnobotanical survey of medicinal plants used in the Maseru district of Lesotho. *J. Ethnopharmacol.* 170, 184–200. doi:10.1016/j.jep.2015.04.047
- Kwon, H., Cha, J., Park, J., Chun, Y., Moodley, N., Maharaj, V., et al. (2011). Rapid identification of bioactive compounds reducing the production of amyloid β -Peptide (A β) from South African plants using an automated HPLC/SPE/HPLC coupling system. *Biomol. Ther.* 19 (1), 90–96. doi:10.4062/biomolther.2011.19.1.090
- Lall, N., Kumar, V., Meyer, D., Gasa, N., Hamilton, C., Matsabisa, M., et al. (2016). *In vitro* and *in vivo* antimycobacterial, hepatoprotective and immunomodulatory activity of *Euclea natalensis* and its mode of action. *J. Ethnopharmacol.* 194, 740–748. doi:10.1016/j.jep.2016.10.060
- Lall, N., and Meyer, J. J. M. (2001). Inhibition of drug-sensitive and drug-resistant strains of *Mycobacterium tuberculosis* by diospyrin, isolated from *Euclea natalensis*. *J. Ethnopharmacol.* 78 (2–3), 213–216. doi:10.1016/s0378-8741(01)00356-7
- Lall, N., Meyer, J. J. M., Taylor, M. B., and van Staden, J. (2005b). Anti-HSV-1 activity of *Euclea natalensis*. *S. Afr. J. Bot.* 71 (3–4), 444–446. doi:10.1016/s0254-6299(15)30118-6
- Lall, N., and Meyer, J. M. (2000). Antibacterial activity of water and acetone extracts of the roots of *Euclea natalensis*. *J. Ethnopharmacol.* 72, 313–316. doi:10.1016/s0378-8741(00)00231-2
- Lall, N., Meyer, J., Wang, Y., Bapela, N., Rensburg, C., Fourie, B., et al. (2005a). Characterization of intracellular activity of antitubercular constituents from the roots of *Euclea natalensis*. *Pharm. Biol.* 43 (4), 353–357. doi:10.1080/13880200590951829
- Lall, N., Weiganand, O., Hussein, A., and Meyer, J. (2006). Antifungal activity of naphthoquinones and triterpenes isolated from the root bark of *Euclea natalensis*. *Euclea natalensis* S. Afr. J. Bot. 72, 579–583. doi:10.1016/j.sajb.2006.03.005
- Long, C. (2005). *Swaziland's flora: SiSwati names and uses*. Swaziland National Trust Commission. [Last accessed on 2021 June 15] Available at: <http://www.sntc.org.sz/index.asp>.
- Maroyi, A. (2017). *Euclea undulata* thumb: Review of its botany, ethnomedicinal uses, phytochemistry and biological activities. *Asian pac. J. Trop. Med.*, 1–7.
- Masoga, M. A. (2020). "Critical reflections on selected local narratives of contextual South African indigenous knowledge," in *African studies: Breakthroughs in Research and practice* (IGI Global), 295–316.
- Mbanga, J., Ncube, M., and Magumura, A. (2013). Antimicrobial activity of *Euclea undulata*, *Euclea divinorum* and *Diospyros lycioides* extracts on multi-drug resistant *Streptococcus mutans*. *J. Med. Plant Res.* 7 (37), 2741–2746.
- Mbaveng, A., and Kuete, V. (2014). Review of the chemistry and pharmacology of 7-Methyl jugulone. *Afr. Health Sci.* 14 (1), 201–205. doi:10.4314/ahs.v14i1.31
- McGaw, L., Lall, N., Hlokw, T., Michel, A., Meyer, J. M., and Eloff, J. (2008). Purified compounds and extracts from *Euclea* species with antimycobacterial activity against *Mycobacterium bovis* and fast-growing *Mycobacteria*. *Biol. Pharm. Bull.* 31 (7), 1429–1433. doi:10.1248/bpb.31.1429
- Mebe, P., Cordell, G., and Pezzuto, J. (1998). Pentacyclic triterpenes and naphthoquinones from *Euclea divinorum*. *Phytochemistry* 47 (2), 311–313. doi:10.1016/s0031-9422(97)00398-1
- Mekonnen, A., Atabachew, M., Kassie, B., and Brien, J. A. E. (2018). Investigation of antioxidant and antimicrobial activities of *Euclea schimperii* leaf extracts. *Chem. Biol. Technol. Agric.* 5 (1), 1–24. doi:10.1007/s40801-017-0125-6
- Moosavi, B., Liu, S., Wang, N., Zhu, X., and Yang, G. (2020). The anti-fungal β -sitosterol targets the yeast oxysterol-binding protein Osh4. *Pest Manag. Sci.* 76 (2), 704–711. doi:10.1002/ps.5568
- Morris, B., and Msonthi, J. D. (1996). *Chewa medical botany: A study of herbalism in southern Malawi*, 2. LIT Verlag Münster.
- Motlhanka, D. M. T., and Nthoiwa, G. P. (2013). Ethnobotanical survey of medicinal plants of tswapong north, in eastern Botswana: A case of plants from mosweu and seolwane villages. *Eur. J. Med. Plants.* 3 (1), 10–24. doi:10.9734/ejmp/2013/1871

- Mueller-harvey, I., Reed, J., and Hartley, R. (1987). Characterisation of phenolic compounds, including flavonoids and tannins, of ten Ethiopian browse species by high performance liquid chromatography. *J. Sci. Food Agri.* 39, 1–14. doi:10.1002/jfsa.2740390102
- Mugomeri, E., Chatanga, P., Hlapisi, S., and RahaL (2014). Phytochemical characterisation of selected herbal products in Lesotho. *Lesotho Med. Assoc. J.* 12 (1), 38–47.
- Nanyingi, M. O., Mbaria, J. M., Lanyasunya, A. L., Wagate, C. G., Koros, K. B., Kaburia, H. F., et al. (2008). Ethnopharmacological survey of Samburu district, Kenya. *J. Ethnobiol. Ethnomed.* 4 (1), 14–12. doi:10.1186/1746-4269-4-14
- Neuwinger, H. D. (2000). *African traditional medicine: A dictionary of plant use and applications. With supplement: Search system for diseases.* Stuttgart, Germany: Medpharm. Scientific Publ., 213.
- Nordeng, H., and Havnen, G. C. (2005). Impact of socio-demographic factors, knowledge and attitude on the use of herbal drugs in pregnancy. *Acta Obstetrica Gynecol. Scand.* 84 (1), 26–33. doi:10.1111/j.0001-6349.2005.00648.x
- Omara, T., Kiprop, A. K., Ramkat, R. C., Cherutoi, J., Kagoya, S., Moraa Nyangena, D., et al. (2020). Medicinal plants used in traditional management of cancer in Uganda: A review of ethnobotanical surveys, phytochemistry, and anticancer studies. *Evid. Based Complement. Altern. Med.* 2020, 1–26. Article ID 3529081. doi:10.1155/2020/3529081
- Oosthuizen, C. B., and Lall, N. (2020). “*Euclea natalensis*,” in *Underexplored medicinal plants from sub-saharan Africa* (Academic Press), 111–116.
- Orzalesi, G., Mezzetti, T., Rossi, C., and Bellavita, V. (1970). Pentacyclic triterpenoids from *Euclea kellaui*. *Planta Medica* 19 (1), 30–36. doi:10.1055/s-0028-1099801
- Palanisamy, C., and Ashafa, T. (2018). Analysis of novel C-X-C chemokine receptor type 4 (CXCR4) inhibitors from hexane extract of *Euclea crispa* (Thunb.) leaves by chemical fingerprint identification and molecular docking analysis. *J. Young Pharm.* 10 (2), 173–177. doi:10.5530/jyp.2018.10.39
- Palanisamy, C., Cui, B., Zhang, H., Trung, N., Tran, H., Khanh, T., et al. (2020). Characterization of (2E,6E)-3,7,11-trimethyldodecane-2,6,10-trien-1-ol with antioxidant and antimicrobial potentials from *Euclea crispa* (Thunb.) leaves. *Int. Lett. Nat. Sci.* 80, 51–63. doi:10.56431/p-v34u92
- Palanisamy, C., Selvarajan, R., Balogun, F., Kanakasabapathy, D., and Ashafa, A. (2019). Antioxidant and antimicrobial activities of (6E, 10E)-2, 6, 24-trimethyl pentacosane-2, 6, 10-triene from *Euclea crispa* leaves. *S. Afr. J. Bot.* 124, 311–319. doi:10.1016/j.sajb.2019.03.019
- Palmer, E., and Pitman, N. (1972). *Trees of southern Africa: Covering all known indigenous species in the republic of South Africa, south-west Africa, Botswana, Lesotho & Swaziland, volumes 1 & 2.* Trees of southern Africa: Covering all known indigenous species in the Republic of South Africa, south-west Africa, Botswana, Lesotho & Swaziland. Volumes 1 & 2.
- Philip, K., Elizabeth, M., Cheaplogoi, P., Langat, M., and Hoseah, M. (2018). *In vitro* antiparasitoid and toxicity activities of crude extracts and compounds from *Euclea latideus* (Ebenaceae). *Int. J. Biochem. Res. Rev.* 21 (1), 1–22. doi:10.9734/ijbcr/2018/39603
- Rademana, S., Anantharaju, P., Madhunapantula, S., and Lalla, N. (2019). The antiproliferative and antioxidant activity of four indigenous South African plants. *Afr. J. Tradit. Complement. Altern. Med.* 16 (1), 13–23. doi:10.21010/ajtcam.v16i1.2
- Schmidt, E., Lotter, M., and McClelland, W. (2002). *Trees and shrubs of Mpumalanga and Kruger national park.* Jacana Media.
- Shauli, M. (2023). Acute toxicity and 28-day oral administration of *Euclea natalensis* extract in Swiss albino mice. *J. Med. Lab. Sci. Technol. S. Afr.* 5 (1), 1–10.
- Shumba, L. (2018). *Phytochemical composition and bacterial activity of Euclea divinorum.*
- Sobiecki, J. F. (2002). A preliminary inventory of plants used for psychoactive purposes in southern African healing traditions. *Trans. R. Soc. S. Afr.* 57 (1-2), 1–24. doi:10.1080/00359190209520523
- Sparg, S. G., Van Staden, J., and Jäger, A. K. (2000). Efficiency of traditionally used South African plants against schistosomiasis. *J. Ethnopharmacol.* 73 (1-2), 209–214. doi:10.1016/s0378-8741(00)00310-x
- Stafford, G. I., Pedersen, M. E., van Staden, J., and Jäger, A. K. (2008). Review on plants with CNS-effects used in traditional South African medicine against mental diseases. *J. Ethnopharmacol.* 119 (3), 513–537. doi:10.1016/j.jep.2008.08.010
- Steenkamp, V. (2003). Traditional herbal remedies used by South African women for gynaecological complaints. *J. Ethnopharmacol.* 86 (1), 97–108. doi:10.1016/s0378-8741(03)00053-9
- Taylor, J. L., Elgorashi, E. E., Maes, A., Van Gorp, U., De Kimpe, N., Van Staden, J., et al. (2003). Investigating the safety of plants used in South African traditional medicine: Testing for genotoxicity in the micronucleus and alkaline comet assays. *Environ. Mol. Mutagen* 42 (3), 144–154. doi:10.1002/em.10184
- Tinevimbo, C. (2017). *Inhibition of α -amylase by Euclea crispa flavonoids and phenolics: Implication for herbal management of Diabetes Mellitus.*
- Tshikalange, T., Lall, N., Meyer, J., and Mahapatra, A. (2007). *In vitro* HIV-1 reverse transcriptase inhibitory activity of naphthoquinones and derivatives from *Euclea natalensis*. *S. Afr. J. Bot.* 73 (2), 339. doi:10.1016/j.sajb.2007.02.200
- Van Wyk, B. E., and Gericke, N. (2000). *People's plants: A guide to useful plants of southern Africa.* Briza publications.
- Weigenand, O., Hussein, A., Lall, N., and Meyer, J. (2004). Antibacterial activity of naphthoquinones and triterpenoids from *Euclea natalensis* root bark. *J. Nat. Prod.* 67 (11), 1936–1938. doi:10.1021/np030465d
- WHO (2019). *WHO Global report on traditional and complementary medicine 2019.* Available at: <https://www.who.int/news/item-the-who-global-report-on-traditional-and-complementary-medicine> (accessed September, 2022).
- Woldemedhin, B., Nedi, T., Shibeshi, W., and Shiferaw, M. (2017). Evaluation of the diuretic activity of the aqueous and 80% methanol extracts of the root of *Euclea divinorum* Hiern (Ebenaceae) in sprague dawley rats. *J. Ethnopharmacol.* 1, 114–121. doi:10.1016/j.jep.2017.01.015
- Wube, A. A., Streit, B., Gibbons, S., Asres, K., and Bucar, F. (2005). *In vitro* 12 (S)-HETE inhibitory activities of naphthoquinones isolated from the root bark of *Euclea racemosa* ssp. schimperi. *J. Ethnopharmacol.* 102 (2), 191–196. doi:10.1016/j.jep.2005.06.009
- Yuan, H., Ma, Q., Ye, L., and Piao, G. (2016). The traditional medicine and modern medicine from natural products. *Molecules* 21 (5), 1–18. doi:10.3390/molecules21050559