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*CORRESPONDENCE Lei Wei, ⊠ leiwei202109@163.com

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Exploring the potential mechanisms of Shiwei Hezi pill against nephritis based on the method of network pharmacology

Lei Wei*

Department of Pharmacy, Northwest Women's and Children's Hospital, Xi'an, China

Objective: We aimed to reveal the potential active ingredients, targets and pathways of Shiwei Hezi pill (SHP) in the treatment of nephritis based on systematic network pharmacology.

Methods: The online database was used to screen the common targets of SHP and nephritis, and the interaction between targets was analyzed. Gene Ontology (GO) functional annotation and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis were performed using the Bioinformatics website. Molecular docking was carried out to verify the correlation between core ingredients and key targets. Cytoscape 3.6.1 was applied to perform protein-protein interactions (PPT) network construction and data visualization.

Results: A total of 82 active ingredients in SHP were screened, and 140 common targets of SHP and nephritis were obtained. Our results demonstrated that TNF, AKT1 and PTGS2 might be the key targets of SHP in the treatment of nephritis. GO enrichment analysis yielded 2163 GO entries (p < 0.05), including 2,014 entries of the biological process (BP) category, 61 entries of the cell composition (CC) category and 143 entries of the molecular function (MF) category. KEGG pathway enrichment analysis produced 186 signaling pathways (p < 0.05), involving the AGE-RAGE, IL-17and TNF signaling pathways. The results of molecular docking showed that three active ingredients in SHP (quercetin, kaempferol and luteolin) could effectively bind to the TNF, AKT1 and PTGS2 targets.

Conclusion: The effective active ingredients in SHP may regulate multiple signaling pathways through multiple targets, thereby exhibiting a therapeutic effect on nephritis.

KEYWORDS

network pharmacology, mechanisms, Shiwei Hezi pill, nephritis, molecular docking

1 Introduction

Nephritis is an immunity-mediated inflammatory response that has a great impact on human health (Fotioo, 1949; Kinoshita, Hirasawa et al., 1966). It belongs to a chronic kidney disease and has emerged as a major global health concern, accounting for approximately 69 percent of the total global disease burden. The mortality of nephritis is considerable and has increased 16.1 percent from 2006 to 2016. In China, around 119.5 million patients suffered chronic kidney disease, making it the second highest chronic disease after hypertension (254 million), and the number of patients with chronic kidney disease is higher than that with diabetes (113.9 million). Due to the unclear etiology and untypical

TABLE 1 A list of name categories for SHP.

Number	Herb name		
	Chinese spelling	Latin name	
1	Hezi	Chebulae Fructus	
2	Honghua Carthami Flos		
3	Zangqiancao	Radixet Rhizoma Rubiae	
4	Shanfanye	Mountain alum leaf	
5	Zhangyacai	Swertia petiolata	
6	Zicaorong	Lithospermum erythrorhizon	
7	Daodou	Canavaliae Semen	
8	Doukou	Alpinia Katsumadai Hayat	
9	Zhaxungao Slag breaking cream		
10	Yuanbai Sabina chinensis		

early clinical symptoms of renal disease secondary to nephritis, coupled with the fact that some patients are skeptical of existing clinical treatments, research on drugs for the treatment of nephritis is urgently needed.

With the continuous development of Chinese medical practice, traditional Chinese medicine (TCM) is becoming increasingly recognized for its potential in treating chronic and complex diseases both at home and abroad (Cheung, 2011). In 2010, TCM products exported to Europe and the United States amounted US\$ 2 billion and US\$ 7.6 billion, respectively, and these figures are still growing (Cheung, 2011). Of course, TCM is also believed to play a vital role in the treatment of kidney-related diseases. For example, Ying Ding et al. have revealed the better curative effect of Tripterygium wilfordii combined with Salvia miltiorrhiza on children with allergic purpura nephritis (Ding, Zhang et al., 2019). Wan Yudang et al. have summarized the clinical characteristics of patients with heat shock nephritis, providing the most effective theoretical basis for TCM in terms of heat shock nephritis treatment (Dang, Yan et al., 2018). Overall, TCM has been studied to be effective in the treatment of kidneyrelated diseases, especially nephritis.

Shiwei Hezi pill (SHP) is a classic Tibetan medicine prescription commonly used to treat nephritis, which consists of ten species of medicinal herbs, including *Chebulae Fructus* (Hezi), *Carthami Flos* (Honghua), *Radixet Rhizoma Rubiae* (Zangqiancao), *Mountain alum leaf* (Shanfanye), *Swertia petiolata* (Zhangyacai), *Lithospermum erythrorhizon* (Zicaorong), *Canavaliae Semen* (Daodou), Alpinia Katsumadai Hayat (Doukou), *Slag breaking cream* (Zhaxungao), and *Sabina chinensis* (Yuanbai) (Table 1).



Network pharmacology flow chart of SHP in treating nephritis, including database preparation, PPI network construction, GO and KEGG pathway analyses, and molecular docking validation.

TABLE 2 The basic information list of chemical constituents in SHP.

Number	Herbs	Mol ID	Molecule name	MW	OB (%)	DL
1	Daodou	MOL000359	sitosterol	414.79	36.91	0.75
2	Daodou	MOL000449	Stigmasterol	412.77	43.83	0.76
3 Honghua		MOL001771	poriferast-5-en-3beta-ol	414.79	36.91	0.75
4	Honghua	MOL002680	Flavoxanthin	584.96	60.41	0.56
5	Honghua	MOL002694	4-[(E)-4-(3,5-dimethoxy- 4-oxo-1- cyclohexa-2,5-dienylidene)but-2-enylidene]- 2,6-dimethoxycyclohexa-2,5- dien-1-one	356.4	48.47	0.36
6	Honghua	MOL002695	lignan	458.55	43.32	0.65
7	Honghua	MOL002698	lupeol-palmitate	665.26	33.98	0.32
8	Honghua	MOL002706	Phytoene	545.04	39.56	0.5
9	Honghua	MOL002707	phytofluene	543.02	43.18	0.5
10	Honghua	MOL002710	Pyrethrin II	372.5	48.36	0.35
11	Honghua	MOL002712	6-Hydroxykaempferol	302.25	62.13	0.22
12	Honghua	MOL002714	baicalein	270.25	33.52	0.2
13	Honghua	MOL002717	qt_carthamone	286.25	51.03	0.2
14	Honghua	MOL002719	6-Hydroxynaringenin	288.27	33.23	0.24
15	Honghua	MOL002721	quercetagetin	318.25	45.01	0.3
16	Honghua	MOL002757	7,8-dimethyl-1H-pyrimido [5,6-g]quinoxaline- 2,4-dione	242.26	45.75	0.1
17	Honghua	MOL002773	beta-carotene	536.96	37.18	0.5
18	Honghua	MOL002776	Baicalin	446.39	40.12	0.75
19	Honghua	MOL000358	beta-sitosterol	414.79	36.91	0.75
20	Honghua	MOL000422	kaempferol	286.25	41.88	0.2
21	Honghua	MOL000006	luteolin	286.25	36.16	0.2
22	Honghua	MOL000953	CLR	386.73	37.87	0.6
23	Honghua	MOL000098	quercetin	302.25	46.43	0.28
24	Honghua	MOL000449	Stigmasterol	412.77	43.83	0.70
25	Hezi	MOL001002	ellagic acid	302.2	43.06	0.43
26	Hezi	MOL002276	Sennoside E_qt	524.5	50.69	0.6
27	Hezi	MOL006376	7-Dehydrosigmasterol	414.79	37.42	0.7
28	Hezi	MOL006826	chebulic acid	356.26	72	0.3
29	Hezi	MOL009135	ellipticine	246.33	30.82	0.2
30	Hezi	MOL009136	Peraksine	310.43	82.58	0.78
31	Hezi	MOL009137	(R)-(6-methoxy-4-quinolyl)-[(2R,4R,5S)-5- vinylquinuclidin-2-yl]methanol	324.46	55.88	0.4
32	Hezi	MOL009149	Cheilanthifoline	325.39	46.51	0.72
33	Doukou	MOL000224	(4E,6E)-1,7-bis(3,4-dihydroxyphenyl)hepta- 4,6-dien-3-one	326.37	33.06	0.3
34	Doukou	MOL000228	(2R)-7-hydroxy-5-methoxy-2- phenylchroman-4-one	270.3	55.23	0.2

(Continued on following page)

TABLE 2 (Continued) The basic information list of chemical constituents in SHP.

Number	Herbs	Mol ID	Molecule name	MW	OB (%)	DL
35	Doukou	MOL000230	Pinocembrin	270.3	57.56	0.2
36	Doukou	MOL000235	1,7-diphenyl-3,5-dihydroxy-1-heptene	282.41	49.01	0.18
37	Doukou	MOL000238	1,7-diphenyl-5-hydroxy-6-hepten-3-one	280.39	32.65	0.18
38	Doukou	MOL000239	Jaranol	314.31	50.83	0.29
39	Doukou	MOL000242	7-O-Methyleriodictyol	302.3	56.56	0.27
40	Doukou	MOL000243	alpinolide peroxide	282.37	87.67	0.19
41	Doukou	MOL000258	dehydrodiisoeugenol	312.39	56.84	0.29
42	Doukou	MOL000260	5-[(2R,3R)-7-methoxy-3-methyl-5-[(E)-prop- 1-enyl]-2,3-dihydrobenzofuran-2-yl]-1,3- benzodioxole	324.4	65.55	0.4
43	Doukou	MOL000006	luteolin	286.25	36.16	0.25
44	Doukou	MOL000098	quercetin	302.25	46.43	0.28
45	Zhangyacai	MOL003137	Leucanthoside	462.44	32.12	0.78
46	Zhangyacai	MOL005530	Hydroxygenkwanin	300.28	36.47	0.27
47	Zhangyacai	MOL005573	Genkwanin	284.28	37.13	0.24
48	Zhangyacai	MOL005575	Gentiacaulein	288.27	72.82	0.27
49	Zhangyacai	MOL007957	Swertiaperennin	288.27	96.85	0.27
50	Zhangyacai	MOL007960	8-hydroxy-1,2,6-trimethoxy-xanthone	302.3	77.13	0.3
51	Zhangyacai	MOL007962	1,7- dihydroxy-3,5-dimethoxy xanthone	288.27	103.37	0.27
52	Zhangyacai	MOL007963	1-hydroxy-2,3,5-trimethoxy-xanthone	302.3	101.06	0.3
53	Zhangyacai	MOL007966	1-Hydroxy-2,3,4,7-tetramethoxyxanthone	332.33	88.86	0.37
54	Zhangyacai	MOL007967	1-hydroxy-2,3,5,7-tetramethoxyxanthone	332.33	97.52	0.37
55	Zhangyacai	MOL007968	norbellidifolin	262.23	58.82	0.22
56	Zhangyacai	MOL007970	5,8-Dimethylbellidifolin	302.3	99.75	0.3
57	Zhangyacai	MOL007972	8-hydroxypinoresinal	374.42	71.09	0.55
58	Zangqiancao	MOL006160	Alizarin	240.22	32.67	0.19
59	Zangqiancao	MOL005638	Mollugin	284.33	42.34	0.26
60	Zangqiancao	_	Rubidate	_		_
61	Zangqiancao	MOL006139	1,3-dimethoxy-2-carboxyanthraquinone	312.29	102.89	0.33
62	Zangqiancao	MOL006153	2'-hydroxymollugin	302.35	40.5	0.29
63	Shanfanye	MOL000006	luteolin	286.25	36.16	0.25
64	Shanfanye	MOL000028	α-Amyrin	426.8	39.51	0.76
65	Shanfanye	MOL000211	Mairin	456.78	55.38	0.78
66	Shanfanye	MOL000422	kaempferol	286.25	41.88	0.24
67	Shanfanye	MOL000098	quercetin	302.25	46.43	0.28
68	Zhaxungao	MOL007115	manool	304.57	45.04	0.2
69	Zhaxungao	_	HUMIC ACID	_	_	_
70	Zhaxungao	MOL002943	1-Butanol	74.14	22.02	0
71	Zicaorong		valerenic acid			_

(Continued on following page)

Number	Herbs	Mol ID	Molecule name	MW	OB (%)	DL
72	Zicaorong	_	Erythrolaccin	_	_	_
73	Zicaorong	_	deoxyerythrolaccin	_	_	_
74	Zicaorong		Aloesaponarin II	_	_	_
75	Yuanbaigao	MOL000422	kaempferol	286.25	41.88	0.24
76	Yuanbaigao	MOL013083	Skimmin (8CI)	324.31	38.35	0.32
77	Yuanbaigao	MOL000492	(+)-catechin	290.29	54.83	0.24
78	Yuanbaigao	MOL002840	Cryptopimaric acid	302.5	39.58	0.28
79	Yuanbaigao	MOL002222	sugiol	300.48	36.11	0.28
80	Yuanbaigao	MOL001951	Bergaptin	338.43	41.73	0.42
81	Yuanbaigao	MOL000392	formononetin	268.28	69.67	0.21
82	Yuanbaigao	MOL004564	Kaempferid	300.28	73.41	0.27

TABLE 2 (Continued) The basic information list of chemical constituents in SHP.

MW: molecule weight; OB(%): oral bioavailability; DL: drug-like properties.



The effectiveness of SHP in treating nephritis has been demonstrated in many studies. For instance, in a study by Ramala, nephritis patients treated with Yishen fossil granules were recruited for the control group, and patients treated with SHP were for the study group. By comparison, Tibetan medicine SHP exhibited high therapeutic efficiency and had a positive effect on alleviating the symptoms of nephritis (Ala, 2017). However, due

to the complex formula of SHP, its specific mechanism in the treatment of nephritis remains unclear.

Nowadays, network pharmacology has been extensively adopted to elucidate the mechanism of TCM compounds and recipes in the treatment of diseases, and in other words, the mystery of TCM prescriptions in the treatment of complex diseases has been gradually unraveled. For example, Yue SJ et al. have explained

TABLE 3 A list	of basic information	about the	common	targets o	of SHP an	d
nephritis.						

Target name 1 Retinoic acid receptor RXR-alpha RXRA 2 Prostaglandin G/H synthase 1 PTGS1Prostaglandin G/H synthase 2 3 PTGS24 Urokinase-type plasminogen activator PLAU5 ESR1 Estrogen receptor Nitric oxide synthase, inducible NOS2 6 7 RAC-alpha serine/threonine-protein kinase AKT1 8 Vascular endothelial growth factor A VEGFA 9 72 kDa type IV collagenase MMP2 10 Caveolin-1 CAV1 11 Transforming growth factor beta-1 TGFB1 E-selectin SELE 12 Interleukin-6 13 IL6 Nitric oxide synthase, endothelial NOS3 14 15 Plasminogen activator inhibitor 1 SERPINE1 Collagen alpha-1(I) chain COL1A1 16 17 Cyclin-dependent kinase inhibitor 1 CDKN1A 18 Matrix metalloproteinase-9 MMP9 19 Interleukin-10 IL1020 Tumor necrosis factor TNF21 Caspase-3 CASP3Peroxisome proliferator-activated receptor 22 PPARG gamma 23 Intercellular adhesion molecule 1 ICAM124 Induced myeloid leukemia cell differentiation MCL1 protein Mcl-1 25 Interferon gamma IFNG 26 Glutathione S-transferase P GSTP1 27 CD40 ligand CD40LG 28 Hepatocyte growth factor receptor MET Apoptosis regulator Bcl-2 29 BCL2 BAX30 Apoptosis regulator BAX 31 Interleukin-1 beta IL1B C-C motif chemokine 2 CCL2 32 Vascular cell adhesion protein 1 VCAM1 33 Interleukin-8 CXCL8 34 Myeloperoxidase MPO35 36 Nuclear factor erythroid 2-related factor 2 NFE2L2 (Continued in next column) TABLE 3 (Continued) A list of basic information about the common targets of SHP and nephritis.

Number	Target name	Gene symbol
37	C-reactive protein	CRP
38	C-X-C motif chemokine 10	CXCL10
39	Osteopontin	SPP1
40	Glutathione S-transferase Mu 1	GSTM1
41	Leukocyte elastase	SERPINB1
42	Matrix metalloproteinase 1	MMP1
43	Matrix metalloproteinase 7	MMP7
44	Matrix metalloproteinase 12	MMP12
45	Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit, gamma isoform	PIK3CG
46	Carbonic anhydrase II	CA2
47	Tyrosine-protein kinase receptor UFO	AXL
48	Vascular endothelial growth factor receptor 2	KDR
49	Tyrosine-protein kinase SRC	SRC
50	Beta-glucuronidase	GUSB
51	Glutathione S-transferase A1	GSTA1
52	52 Focal adhesion kinase 1	
53	53 Androgen receptor	
54	54 Trypsin-1	
55	Dipeptidyl peptidase IV	
56	Caspase-9	CASP9
57	Mitogen-activated protein kinase 1	MAPK1
58	Cellular tumor antigen p53	TP53
59	NF-kappa-B inhibitor alpha	NFKBIA
60	Xanthine dehydrogenase/oxidase	XDH
61	DNA topoisomerase 1	TOP1
62	E3 ubiquitin-protein ligase Mdm2	MDM2
63	Proliferating cell nuclear antigen	PCNA
64	Heme oxygenase 1	HMOX1
65	Baculoviral IAP repeat-containing protein 5	BIRC5
66	Interleukin-2	IL2
67	G2/mitotic-specific cyclin-B1	CCNB1
68	Interleukin-4	IL4
69	Insulin receptor	INSR
70	Serotonin transporter	SLC6A4
71	Cytochrome P450 2C19	CYP2C19
72	Butyrylcholinesterase	BCHE
73	Cytochrome P450 17A1	CYP17A1

(Continued on following page)

Number	Target name	Gene symbol
74	Nuclear receptor ROR-gamma	RORC
75	Fatty acid-binding protein, liver	FABP1
76	Phospholipase A2 group 1B	PLA2G1B
77	CD81 antigen	CD81
78	UDP-glucuronosyltransferase 2B7	UGT2B7
79	Mitogen-activated protein kinase 8	MAPK8
80	Signal transducer and activator of transcription 1- alpha/beta	STAT1
81	Cytochrome P450 3A4	CYP3A4
82	Cytochrome P450 1A2	CYP1A2
83	Cytochrome P450 1A1	CYP1A1
84	Matrix metalloproteinase 3	MMP3
85	Epidermal growth factor receptor	EGF
86	ETS domain-containing protein Elk-1	ELK1
87	Ornithine decarboxylase	ODC1
88	Caspase-8	CASP8
89	89 Superoxide dismutase [Cu-Zn]	
90 Protein kinase C alpha type		PRKCA
91	91 Hypoxia-inducible factor 1-alpha	
92	92 Myc proto-oncogene protein	
93	Tissue factor	
94	Protein kinase C beta type	PRKCB
95	Heat shock protein beta-1	HSPB1
96	Tissue-type plasminogen activator	PLAT
97	Thrombomodulin	THBD
98	Interleukin-1 alpha	IL1A
99	Neutrophil cytosol factor 1	NCF1
100	Poly [ADP-ribose] polymerase 1	PARP1
101	C-X-C motif chemokine 11	CXCL11
102	C-X-C motif chemokine 2	CXCL2
103	Inhibitor of nuclear factor kappa-B kinase subunit alpha	CHUK
104	Cathepsin D	CTSD
105	Interferon regulatory factor 1	IRF1
106	Receptor tyrosine-protein kinase erbB-3	ERBB3
107	Serum paraoxonase/arylesterase 1	PON1
108	Muscarinic acetylcholine receptor M3	CHRM3

TABLE 3 (*Continued*) A list of basic information about the common targets of SHP and nephritis.

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TABLE 3 (*Continued*) A list of basic information about the common targets of SHP and nephritis.

Number	Target name	Gene symbol
109	Protein kinase C delta	PRKCD
110	Heparin cofactor 2	SERPIND1
111	Transcription factor Jun	JUN
112	Cyclin-dependent kinase 1	CDK1
113	Polyunsaturated fatty acid 5-lipoxygenase	ALOX5
114	Tyrosine-protein phosphatase non-receptor type 11	PTPN11
115	Catalase	CAT
116	Mu-type opioid receptor	OPRM1
117	Solute carrier family 22 member 12	SLC22A12
118	Casein kinase II subunit alpha	CSNK2A1
119	Neutrophil elastase	ELANE
120	Tyrosine-protein kinase Lck	LCK
121	Tyrosine-protein kinase SYK	SYK
122	Transthyretin	TTR
123	123 Cystic fibrosis transmembrane conductance regulator	
124	124 ATP-dependent translocase ABCB1	
125	5 Maltase-glucoamylase	
126	Amyloid-beta precursor protein	APP
127	ADP-ribosyl cyclase/cyclic ADP-ribose hydrolase 1	CD38
128	Macrophage migration inhibitory factor	MIF
129	Vasopressin V2 receptor	AVPR2
130	Thrombin	F2
131	Insulin-like growth factor-binding protein 2	IGFBP2
132	C-X-C chemokine receptor type 1	CXCR1
133	Immunoglobulin heavy constant gamma 1	IGHG1
134	Leukotriene A-4 hydrolase	LTA4H
135	Aldo-keto reductase family 1 member B1	AKR1B1
136	Protein c-Fos	FOS
137	Cyclin-dependent kinase inhibitor 2A	CDKN2A
138	138 Phosphatidylinositol 3,4,5-trisphosphate 3- phosphatase and dual-specificity protein phosphatase PTEN	
139	Collagen alpha-1	COL3A1
140	Growth arrest-specific protein 6	GAS6





the mechanism of Danggui-Honghua in the treatment of blood stasissyndrome by the systems pharmacology approach (Yue, Xin et al., 2017); Liu J et al. have revealed the therapeutic properties of Saffron formula in treating cardiovascular diseases based on systematic pharmacology dissection (Liu, Mu et al., 2016); Pang XC et al. have employed the virtual screening and network pharmacological methods to analyze the potential efficacy of Naodesheng formula in the treatment of Alzheimer's disease (Pang, Kang et al., 2018); and similar scientific approaches were also performed by Xie W et al. to predict the anti-depressive effect of *Panax Notoginseng Saponins* (Xie, Meng et al., 2018). However, there are no in-depth studies which explore the specific mechanism of SHP in nephritis treatment by applying network pharmacological method.

This study adopted network pharmacology method to identify the potential active ingredients, key targets and pathways of SHP in the treatment of nephritis, and molecular docking was carried out to investigate the interactions between selected key targets and active compounds. As shown in Figure 1, a schematic diagram of network pharmacological strategy was generated to determine the pharmacological mechanism of SHP in treating nephritis.

2 Methods

2.1 Acquisition of SHP ingredients

The chemical composition of each herb in SHP was obtained from free public databases, including TCMSP (http://ibts.hkbu.edu.hk/LSP/ tcmsp.php) and TCMID (http://tcm.cmu.edu.tw) databases (Chen, 2011; Ru, Li et al., 2014). SHP ingredients with oral bioavailability (OB) \geq 30% and drug-likeness (DL) \geq 0.18 were then screened according to a number of relevant literature criteria. Ultimately, a data set of the potential active ingredients in SHP was constructed (Chandran, Mehendale et al., 2015; Kibble, Saarinen et al., 2015; Huang, Cheung et al., 2017).

2.2 Acquisition of SHP-related targets

The targets of the potential active ingredients in SHP were downloaded from the TCMSP database, and their duplicates were deleted. The UniProt (https://www.uniprot.org/) database was retrieved to annotate the target protein sequence, after that, the corresponding targets of the bioactive ingredients in SHP were obtained. We then established the ingredient-target data set by converting the target sites into gene names with the species limit to *"Homo sapiens"* by the DAVID database (https://david.ncifcrf. gov) (Dennis, Sherman et al., 2003).

2.3 Acquisition of nephritis-related targets

The nephritis-related targets were filtered by searching the PubMed (https://pubmed.ncbi.nlm.nih.gov/) (Amberger, Bocchini et al., 2015), OMIM (http://omim.org/) (Sayers, Agarwala et al., 2019), and GeneCard (https://www.genecards.org/) (Stelzer, Rosen et al., 2016) databases using "Nephritis" as a keyword. Afterwards, deduplication was performed, and a data set of nephritis-related targets was successfully created.

2.4 Acquisition of common targets of SHP and nephritis

The SHP and nephritis-related targets were imported into the Venny 2.1.0 online platform (https://bioinfogp.cnb.csic.es/tools/venny/) to construct a Venn diagram of common targets of SHP and nephritis.

2.5 Construction of protein-protein interaction (PPI) network

The data set of common targets of SHP and nephritis was imported into the STRING database Version 10.5 (https://cn.string-db.org/) with the species limit to "*H. sapiens*". The PPI was obtained and saved as a "tsv" format file. Afterwards, the file was input into Cytoscape 3.6.1 to build a PPI network.

2.6 Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses

GO functional annotation and KEGG pathway analysis were performed via the Bioinformatics website (http://bioinformatics.com. cn/). By entering a list of target gene names and restricting the species to human, all target gene names were corrected to their official gene symbols. The top 15 and top 20 entries, including biological processes (BP), cellular components (CC) and molecular functions (MF), were selected for analysis with *p*-value as a screening criterion. The histograms and bubble charts were plotted to further explore the biological significance of common targets of SHP and nephritis.



2.7 Construction of network

In order to further investigate the mechanism action of SHP in nephritis treatment, the herb-ingredient, herb-ingredient-target-disease and target-pathway networks were established and visualized using Cytoscape 3.7.1, an open source software for visualizing complex networks (Su, Morris et al., 2014). In these networks, "nodes" represented herbs, compounds, targets, diseases or pathways, and "edges" represented the interactions between them.

2.8 Molecular docking

The most important active ingredients and action targets are selected for molecular docking. The files of 3D structures of compounds were downloaded in SDF format from the PubChem database, while the files of the 3D structures of targets were obtained in PDB format from the Protein Data Bank (PDB) database (https://www.rcsb.org/) (Chen, Chen et al., 2019). Using AutoDock, water molecules,

TABLE 4 Ten important protein targets with top degree of PPI network.

Number	Protein target name	Degree
1	TNF	108
2	IL6	107
3	AKT1	100
4	VEGFA	96
5	TP53	96
6	IL1B	92
7	JUN	88
8	MMP9	86
9	PTGS2	82
10	CASP3	82

hydrogen, and charges were removed, and the PDBQT format was saved. Subsequently, AutoDock Vina was used to performed molecular docking. Optimal docking score combinations were visualized by PyMOL 2.4.

3 Results

3.1 Bioactive ingredients in SHP

Combining TCMSP and TCMID databases with literature search, 82 bioactive ingredients (Table 2) of 10 herbs in SHP were obtained. Among them, the active ingredients of 5 herbs (*Chebulae Fructus, Carthami Flos, Canavaliae Semen, Alpinia Katsumadai Hayat*, and *S. petiolata*) were obtained from the TCMSP and TCMID databases, while the active ingredients of the remaining 5 herbs were identified through the literature search. Notably, despite madder diester and n-butanol with OB < 30% and DL < 0.18, they still showed a high content and potential anti-nephritis value, making them as active ingredients. Cytoscape software was then used to construct a compositional network diagram, and 75 active ingredients were gained after removing duplicate ingredients of this herb (Figure 2).

3.2 Acquisition of common targets of SHP and nephritis

A total of 861 SHP-related targets were obtained based on the TCMSP database. And after the deduplication of the targets corresponding to





75 ingredients of SHP, we acquired 359 SHP-related targets. According to the Pubmed, OMIM and GeneCards databases, a total of 1925 targets related to nephritis were obtained. Ultimately, inputting 359 SHP targets and 1925 nephritis targets into Venny 2.1 software, a total of 140 targets common to both SHP and nephritis were found, and a Venn diagram was established (Table 3; Figure 3).

ingredients of herbs they corresponded to, and TNF, AKT1 and PTGS2 with a largest number of corresponding active ingredients were regarded as key targets in the study. Meanwhile, the top three components corresponding to abundant core targets were quercetin, kaempferol and luteolin, which were the main active components.

3.3 Construction and analysis of PPI network

The PPI network, containing 140 nodes and 2,660 edges, was constructed using the STRING database and visualized by the Cytoscape software (Figure 4). In this network, the node represented the active compound of SHP and the compoundrelated target, and the edge represented interactions among active compounds and target proteins. Additionally, the node size is proportional to the target degree value. The higher the degree value, the larger the node and the more important it is. Afterwards, targets were sorted according to degree values, and the first 10 key targets with the highest degree value were listed in Table 4. The first 10 targets were used to identify the active

3.4 Construction of herb-compoundtarget-nephritis network

As illustrated in Figure 5, an herb-compound-target-nephritis network was constructed to provide a clearing visualization of the relationship among herbs, ingredients, targets and nephritis. In this network, green denotes herbs, orange denotes ingredients, blue denotes targets, and red denotes diseases.

3.5 GO and KEGG analyses

GO functional enrichment analysis of target genes in the PPI network was performed using the Bioinformatics website, and a total



of 2,163 entries (2,014 of the BP category, 61 of the CC category, and 143 of the MF category) were provided. The top 10 significant enrichment results of each category are shown in Figure 6. The BP entries were mainly related to cell responses to chemical/oxidative stress and lipopolysaccharide. The CC entries focused on membrane rafts, membrane microdomains and membrane regions. The MF entries covered cytokine receptor binding, cytokine activity, and signaling receptor activator activity. Subsequently, the KEGG pathway enrichment analysis was conducted on 140 common targets, and 186 regulated pathways (Figure 7), such as the AGE-RAGE signaling pathway in diabetic complications, the IL-17 signaling pathway and the TNF signaling pathway, were identified. In addition, a target pathway network was established based on the first 10 paths in Figure 7, as shown in Figure 8.

3.6 Molecular docking of ingredients and targets

The binding interactions between main active ingredients (quercetin, kaempferol and luteolin) and key targets (TNF, AKT1 and PTGS2) was verified by molecular docking. The molecular docking binding performance is presented in Table 5. The interaction patterns of these three ingredients with key targets are shown in Figure 9. It is generally believed that binding energy less than -4.25 kcal/mol, -5.0 kcal/mol or -7.0 kcal/mol indicates certain, good or strong binding activity between ligand and receptor, respectively. Therefore, our results indicated a stable complex consisting of quercetin, kaempferol and luteolin, and

these three ingredients all showed a strong binding activities with TNF, AKT1 and PTGS2 (binding energy < -6.9 kcal/mol).

4 Discussion

Nephritis is characterized by various pathological forms and clinically presents with albuminuria, hematuria, hypertension and edema. If left untreated, it can lead to renal shrinkage and decreased function. Therefore, it is urgent to develop safer and more effective anti-nephritis drugs (Chadban and Atkins, 2005). In this study, the active components of SHP and its antinephritis molecular mechanism were explored by network pharmacology.

In this study, quercetin, kaempferol and luteolin had higher target frequency, suggesting that they played a central role in the treatment of nephritis. Quercetin, with anti-inflammatory, antioxidant and neuroprotective propertie, is a natural flavonoid found in a wide range of fruits, herbs and vegetabless (Shen, Lin et al., 2021). A study has suggested that quercetin can help ameliorate lupus nephritis (LN)-associated renal fibrosis and inflammation (Chen, Chiang et al., 2022). Kaempferol is a dietary flavonoid existed in various plants (Wong, Chin et al., 2019) and has been explored to have protective effects on the kidneys of rats with radiation nephritis (Mostafa, Edmond et al., 2022). Luteolin is another kind of flavonoids commonly found in medicinal plants, and exhibits a strong anti-inflammatory activity both *in vitro* and *in vivo* (Aziz, Kim et al., 2018). Kin et al. had probed that luteolin may be able to mitigate kidney inflammation and interstitial fibrosis



FIGURE 8

Target-pathway network. Green represents the pathway. The purple circle is the target. Hsa04933: AGE-RAGE signaling pathway in diabetic complications; hsa05418: Fluid shear stress and atherosclerosis; hsa05417: Lipid and atherosclerosis; hsa05161: Hepatitis B; hsa04657: IL-17 signaling pathway; hsa04668: TNF signaling pathway; hsa05205: Proteoglycans in cancer; hsa04625: C-type lectin receptor signaling pathway; hsa05167: Kaposi sarcoma-associated herpesvirus infection; and hsa05142: Chagas disease.

(Kim, Kim et al., 2016). Collectively, quercetin, kaempferol and luteolin in SHP may all play a role in the treatment of nephritis.

By mapping SHP and nephritis-related targets, 140 shared genes between SHP and nephritis were detected. In order to further understand the interaction among these gene-encoded proteins, a PPI network was constructed. The results showed that TNF, AKT1 and PTGS2 were the main targets of SHP in treating nephritis, which was consistent with previous reports. The *TNF* gene encodes a multifunctional proinflammatory cytokine belonging to the tumor necrosis factor superfamily (Wu, Wen et al., 2020). Studies (Bantis, Heering et al., 2006) have revealed that the G-308A polymorphism of the *TNF*- α gene is associated with the expression of the –308A allele and the increase of TNF- α production, making TNF a risk factor for membranous glomerulonephritis (Müller, Hoppe et al., 2019). AKT encodes one of three members of the human AKT serine-threonine protein kinase family, and it is commonly referred to as the protein kinases B encoding one of three key component of many

Molecule ID	Compound	Target protein	PDB identifier	Estimated ΔG (kcal/mol)
MOL000098	Quercetin	TNF	7jra	-6.9
		AKT1	30s5	-7.5
		PTGS2	5f19	-7.1
MOL000422	Kaempferol	TNF	7jra	-7.3
		AKT1	30s5	-7.6
		PTGS2	5f19	-8.4
MOL000006	MOL000006 Luteolin		7jra	-7.4
			3os5	-6.8
		PTGS2	5f19	-9.6

TABLE 5 The binding energy values of quercetin, kaempferol and luteolin with TNF, AKT1 and PTGS2.



signaling pathways. It has been demonstrated that subcellular C5b-9 complex can induce the proliferation of glomerular mesangial cells in rat Thy-1 glomerulonephritis by activating *TRAF6*-mediated *PI3K*-dependent *AKT1* (Qiu, Zhang et al., 2012). PTGS2 (also known as *COX-2*), a prostaglandin endoperoxidase, exerts a key effect in prostaglandin biosynthesis. It has been reported that there is a certain relationship between *COX-2* inhibitors and acute interstitial nephritis (Albrecht, Giebel et al., 2017). Juan Jin et al. (Jin, Lin et al., 2018) have revealed that the over-expression of *COX-2* can lead to renal autophagy and injury. Thus, the regulation of TNF, AKT1 and PTGS2 may contribute to the treatment of nephritis.

After identifying the main targets (TNF, AKT1 and PTGS2) of SHP in nephritis treatment, we further conducted a KEGG analysis to reveal the signal pathways of these main targets. The results showed that the AGE-RAGE, IL-17 and TNF signal pathways were the important pathways of SHP acting on nephritis. Studies have shown that the accumulation of *AGE* and *RAGE* in the kidneys and other tissues of diabetic patients is related to the development of diabetic nephropathy and vascular diseases (Tanji, Markowitz et al., 2000). Additionally, a study on the pathogenesis of LN has pointed out that AGE-RAGE can regulate high nitrotyrosination in LN, thereby reducing the oxidative stress in LN (Ene, Georgescu et al., 2021). IL-17 has also been recognized as an independent risk factor for LN prognosis and an effective indicator for the clinical diagnosis, treatment and prognosis of LN (Paquissi and Abensur, 2021). Similarly, multiple evidences have suggested that recently discovered T cells (Th17 cells) that produce interleukin 17 (IL-17) are involved in the renal inflammatory cascade associated with glomerulonephritis (Ramani and Biswas, 2016). The TNF signaling pathway is also related to nephritis. For instance, Xiaoping Qing et al. have elucidated that the TNF signaling pathway plays a key role in irreversible LN kidney damage (Qing, Chinenov et al., 2018). Moreover, it has been found that TNF-a production in T lymphocytes alleviates NTN-induced kidney injury and fibrosis by inhibiting renal T helper 17 lymphocyte response and neutrophil infiltration (Wen, Rudemiller et al., 2020). In conclusion, we speculateed that SHP may ameliorate nephritis by regulating the AGE-RAGE, IL-17 and TNF signaling pathways.

To further verify the relationship between active ingredients (quercetin, kaempferol and luteolin) and key targets (TNF, AKT1 and PTGS2), we carried out molecular docking. The results showed that the binding energies of quercetin and luteolin with TNF, AKT1 and PTGS2 were lower than -5.0 kJ/mol, indicating the potential for forming an effective and stable complex between the ligand and receptor. In short, the core active compounds in SHP appeared to regulate SHP-related pathways by acting on the important genes linked to nephritis, thus offering therapeutic value for nephritis treatment.

However, our research has some limitations. Bioactive ingredients of SHP were screened only from existing public databases and literatures, rather than using mass spectrometry and other methods. Additionally, there is a lack of animal experiments and clinical trials to verify our findings. To further improve our research, more animal experiments and clinical trials will be conducted in the future.

5 Conclusion

In this study, we combined network pharmacology and molecular docking to explore the mechanism by which SHP exerts its anti-nephritis effects. We found that quercetin, kaempferol and luteolin are likely the main active compounds of SHP responsible for its therapeutic effects against nephritis. Moreover, SHP can target the expression of TNF, AKT1 and PTGS2 via the AGE-RAGE, IL-17 and TNF signaling pathways. Overall, although more researches are needed to clarify the exact mechanism, this study provides a valuable insight into the application of SHP for nephritis treatment and the potential for future anti-nephritis drug development.

References

Aziz, N., Kim, M. Y., and Cho, J. Y. (2018). Anti-inflammatory effects of luteolin: A review of *in vitro*, *in vivo*, and *in silico* studies. *J. Ethnopharmacol.* 225, 342–358. doi:10. 1016/j.jep.2018.05.019

Ala, L. (2017). Clinical observation on treatment of nephritis with Shiwei Hezi pill. World Latest Med. Inf. 17 (78), 147.

Albrecht, B., Giebel, S., McCarron, M., and Prasad, B. (2017). Cyclooxygenase-2 inhibitor-induced acute interstitial nephritis. *BMJ Case Rep.* 2017, bcr2017219973. doi:10.1136/bcr-2017-219973

Amberger, J. S., Bocchini, C. A., Schiettecatte, F., Scott, A. F., and Hamosh, A. (2015). OMIM.org: Online Mendelian Inheritance in Man (OMIM[®]), an online catalog of human genes and genetic disorders. *Nucleic Acids Res.* 43, D789–D798. doi:10.1093/nar/gku1205

Bantis, C., Heering, P. J., Aker, S., Siekierka, M., Kuhr, N., Grabensee, B., et al. (2006). Tumor necrosis factor-alpha gene G-308A polymorphism is a risk factor for the development of membranous glomerulonephritis. *Am. J. Nephrol.* 26 (1), 12–15. doi:10.1159/000090706

Chadban, S. J., and Atkins, R. C. (2005). Glomerulonephritis. Lancet 365 (9473), 1797-1806. doi:10.1016/S0140-6736(05)66583-X

Chandran, U., Mehendale, N., Tillu, G., and Patwardhan, B. (2015). Network pharmacology of ayurveda formulation triphala with special reference to anti-cancer property. *Comb. Chem. High. Throughput Screen* 18 (9), 846–854. doi:10.2174/1386207318666151019093606

Chen, C. Y. (2011). TCM Database@Taiwan: The world's largest traditional Chinese medicine database for drug screening *in silico. PLoS One* 6 (1), e15939. doi:10.1371/journal.pone.0015939

Chen, H. Y., Chiang, Y. F., Hong, Y. H., Shieh, T. M., Huang, T. C., Ali, M., et al. (2022). Quercetin ameliorates renal injury and pyroptosis in lupus nephritis through inhibiting IL-33/ST2 pathway *in vitro* and *in vivo*. *Antioxidants (Basel)* 11 (11), 2238. doi:10.3390/antiox11112238

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/supplementary material.

Author contributions

The author confirms being the sole contributor of this work and has approved it for publication.

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Conflict of interest

The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Chen, Y., Chen, D., Liu, S., Yuan, T., Guo, J., Fang, L., et al. (2019). Systematic elucidation of the mechanism of genistein against pulmonary hypertension via network pharmacology approach. *Int. J. Mol. Sci.* 20 (22), 5569. doi:10.3390/ijms20225569

Cheung, F. (2011). Tcm: Made in China. Nature 480 (7378), S82–S83. doi:10.1038/ 480S82a

Dang, W., Yan, H., Wu, X., Zhen, X., and Hou, L. (2018). Correlation between traditional Chinese medicine symptom patterns and the renal function, immunologic function index, and blood coagulation index in patients with henoch-schönlein purpura nephritis. *Evid. Based Complement. Altern. Med.* 2018, 1972527. doi:10.1155/2018/1972527

Dennis, G., Jr., Sherman, B. T., Hosack, D. A., Yang, J., Gao, W., Lane, H. C., et al. (2003). David: Database for annotation, visualization, and integrated discovery. *Genome Biol.* 4 (5), P3. doi:10.1186/gb-2003-4-5-p3

Ding, Y., Zhang, X., Ren, X., Zhai, W., He, L., Liu, J., et al. (2019). Traditional Chinese medicine versus regular therapy in henoch-schönlein purpura nephritis in children: Study protocol for a randomized controlled trial. *Trials* 20 (1), 538. doi:10.1186/s13063-019-3484-3

Ene, C. D., Georgescu, S. R., Tampa, M., Matei, C., Mitran, C. I., Mitran, M. I., et al. (2021). Cellular response against oxidative stress, a novel insight into lupus nephritis pathogenesis. *J. Pers. Med.* 11 (8), 693. doi:10.3390/jpm11080693

Fotioo, G. J. (1949). Nephritides. Med. Bull. U. S. Army Eur. 6 (10), 33-41.

Huang, J., Cheung, F., Tan, H. Y., Hong, M., Wang, N., Yang, J., et al. (2017). Identification of the active compounds and significant pathways of yinchenhao decoction based on network pharmacology. *Mol. Med. Rep.* 16 (4), 4583–4592. doi:10.3892/mmr.2017.7149

Jin, L., Lin, X., Yang, L., Fan, X., Wang, W., Li, S., et al. (2018). AK098656, a novel vascular smooth muscle cell-dominant long noncoding RNA, promotes hypertension. *Hypertension* 71 (2), 262–272. doi:10.1161/HYPERTENSIONAHA.117.09651

Kibble, M., Saarinen, N., Tang, J., Wennerberg, K., Mäkelä, S., and Aittokallio, T. (2015). Network pharmacology applications to map the unexplored target space and therapeutic potential of natural products. *Nat. Prod. Rep.* 32 (8), 1249–1266. doi:10.1039/c5np00005j

Kim, T. W., Kim, Y. J., Seo, C. S., Kim, H. T., Park, S. R., Lee, M. Y., et al. (2016). Elsholtzia ciliata (Thunb) Hylander attenuates renal inflammation and interstitial fibrosis via regulation of TGF-ß and Smad3 expression on unilateral ureteral obstruction rat model. *Phytomedicine* 23 (4), 331–339. doi:10.1016/j.phymed.2016.01.013

Kinoshita, Y., Hirasawa, Y., and Morita, T. (1966). Nephritis (including subacute nephritis). *Naika* 18 (6), 1023–1029.

Liu, J., Mu, J., Zheng, C., Chen, X., Guo, Z., Huang, C., et al. (2016). Systemspharmacology dissection of traditional Chinese medicine compound Saffron formula reveals multi-scale treatment strategy for cardiovascular diseases. *Sci. Rep.* 6, 19809. doi:10.1038/srep19809

Mostafa, N. M., Edmond, M. P., El-Shazly, M., Fahmy, H. A., Sherif, N. H., and Singab, A. N. B. (2022). Phytoconstituents and renoprotective effect of Polyalthia longifolia leaves extract on radiation-induced nephritis in rats via TGF- β /smad pathway. *Nat. Prod. Res.* 36 (16), 4187–4192. doi:10.1080/14786419.2021.1961252

Müller, M. B., Hoppe, J. M., Bideak, A., Lux, M., Lindenmeyer, M. T., Müller, S., et al. (2019). Exclusive expression of transmembrane TNF aggravates acute glomerulonephritis despite reduced leukocyte infiltration and inflammation. *Kidney Int.* 95 (1), 75–93. doi:10.1016/j.kint.2018.08.012

Pang, X. C., Kang, D., Fang, J. S., Zhao, Y., Xu, L. J., Lian, W. W., et al. (2018). Network pharmacology-based analysis of Chinese herbal Naodesheng formula for application to Alzheimer's disease. *Chin. J. Nat. Med.* 16 (1), 53–62. doi:10.1016/S1875-5364(18)30029-3

Paquissi, F. C., and Abensur, H. (2021). The Th17/IL-17 Axis and kidney diseases, with focus on lupus nephritis. *Front. Med. (Lausanne)* 8, 654912. doi:10.3389/fmed.2021.654912

Qing, X., Chinenov, Y., Redecha, P., Madaio, M., Roelofs, J. J., Farber, G., et al. (2018). iRhom2 promotes lupus nephritis through TNF-α and EGFR signaling. *J. Clin. Invest.* 128 (4), 1397–1412. doi:10.1172/JCI97650

Qiu, W., Zhang, Y., Liu, X., Zhou, J., Li, Y., Zhou, Y., et al. (2012). Sublytic C5b-9 complexes induce proliferative changes of glomerular mesangial cells in rat Thy-1 nephritis through TRAF6-mediated PI3K-dependent Akt1 activation. *J. Pathol.* 226 (4), 619–632. doi:10.1002/path.3011

Ramani, K., and Biswas, P. S. (2016). Emerging roles of the Th17/IL-17-axis in glomerulonephritis. *Cytokine* 77, 238-244. doi:10.1016/j.cyto.2015.07.029

Ru, J., Li, P., Wang, J., Zhou, W., Li, B., Huang, C., et al. (2014). Tcmsp: A database of systems pharmacology for drug discovery from herbal medicines. *J. Cheminform* 6, 13. doi:10.1186/1758-2946-6-13

Sayers, E. W., Agarwala, R., Bolton, E. E., Brister, J. R., Canese, K., Clark, K., et al. (2019). Database resources of the national center for Biotechnology information. *Nucleic Acids Res.* 47 (D1), D23–d28. doi:10.1093/nar/gky1069

Shen, P., Lin, W., Deng, X., Ba, X., Han, L., Chen, Z., et al. (2021). Potential implications of quercetin in autoimmune diseases. *Front. Immunol.* 12, 689044. doi:10.3389/fimmu.2021.689044

Stelzer, G., Rosen, N., Plaschkes, I., Zimmerman, S., Twik, M., Fishilevich, S., et al. (2016). The GeneCards suite: From gene data mining to disease genome sequence analyses. *Curr. Protoc. Bioinforma.* 54, 1. doi:10.1002/cpbi.5-31.30.33

Su, G., Morris, J. H., Demchak, B., and Bader, G. D. (2014). Biological network exploration with Cytoscape 3. *Curr. Protoc. Bioinforma.* 47 (13), 11–24. doi:10.1002/0471250953.bi0813s47

Tanji, N., Markowitz, G. S., Fu, C., Kislinger, T., Taguchi, A., Pischetsrieder, M., et al. (2000). Expression of advanced glycation end products and their cellular receptor RAGE in diabetic nephropathy and nondiabetic renal disease. *J. Am. Soc. Nephrol.* 11 (9), 1656–1666. doi:10.1681/ASN.V1191656

Wen, Y., Rudemiller, N. P., Zhang, J., Robinette, T., Lu, X., Ren, J., et al. (2020). TNF- α in T lymphocytes attenuates renal injury and fibrosis during nephrotoxic nephritis. *Am. J. Physiol. Ren. Physiol.* 318 (1), F107–f116. doi:10.1152/ajprenal.00347.2019

Wong, S. K., Chin, K. Y., and Ima-Nirwana, S. (2019). The osteoprotective effects of kaempferol: The evidence from *in vivo* and *in vitro* studies. *Drug Des. Devel Ther.* 13, 3497–3514. doi:10.2147/DDDT.S227738

Wu, H., Wen, Y., Yue, C., Li, X., and Gao, R. (2020). Serum TNF- α level is associated with disease severity in adult patients with immunoglobulin A vasculitis nephritis. *Dis. Markers* 2020, 5514145. doi:10.1155/2020/5514145

Xie, W., Meng, X., Zhai, Y., Zhou, P., Ye, T., Wang, Z., et al. (2018). Panax Notoginseng Saponins: A review of its mechanisms of antidepressant or anxiolytic effects and network analysis on phytochemistry and pharmacology. *Molecules* 23 (4), 940. doi:10.3390/molecules23040940

Yue, S. J., Xin, L. T., Fan, Y. C., Li, S. J., Tang, Y. P., Duan, J. A., et al. (2017). Herb pair danggui-honghua: Mechanisms underlying blood stasis syndrome by system pharmacology approach. *Sci. Rep.* 7, 40318. doi:10.1038/srep40318