#### Check for updates

#### **OPEN ACCESS**

EDITED BY Wael M. Y. Mohamed, International Islamic University Malaysia, Malaysia

REVIEWED BY Jordi Bonaventura, University of Barcelona, Spain Giovanni Hernandez, McGill University, Canada

\*CORRESPONDENCE

Hojin Shin, ishin.hojin@mayo.edu Yoonbae Oh, iso oh.yoonbae@mayo.edu

RECEIVED 03 April 2023 ACCEPTED 05 June 2023 PUBLISHED 20 June 2023

#### CITATION

Yuen J, Goyal A, Rusheen AE, Kouzani AZ, Berk M, Kim JH, Tye SJ, Abulseoud OA, Oesterle TS, Blaha CD, Bennet KE, Lee KH, Oh Y and Shin H (2023), Oxycodone-induced dopaminergic and respiratory effects are modulated by deep brain stimulation. *Front. Pharmacol.* 14:1199655. doi: 10.3389/fphar.2023.1199655

#### COPYRIGHT

© 2023 Yuen, Goyal, Rusheen, Kouzani, Berk, Kim, Tye, Abulseoud, Oesterle, Blaha, Bennet, Lee, Oh and Shin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Oxycodone-induced dopaminergic and respiratory effects are modulated by deep brain stimulation

Jason Yuen<sup>1,2</sup>, Abhinav Goyal<sup>1,3</sup>, Aaron E. Rusheen<sup>1,3</sup>, Abbas Z. Kouzani<sup>4</sup>, Michael Berk<sup>2</sup>, Jee Hyun Kim<sup>2</sup>, Susannah J. Tye<sup>5,6,7,8</sup>, Osama A. Abulseoud<sup>9</sup>, Tyler S. Oesterle<sup>10</sup>, Charles D. Blaha<sup>1</sup>, Kevin E. Bennet<sup>1,11</sup>, Kendall H. Lee<sup>1,12</sup>, Yoonbae Oh<sup>1,12</sup>\* and Hojin Shin<sup>1,12</sup>\*

<sup>1</sup>Department of Neurologic Surgery, Mayo Clinic, Rochester, MN, United States, <sup>2</sup>IMPACT—The Institute for Mental and Physical Health and Clinical Translation, School of Medicine, Barwon Health, Deakin University, Geelong, VIC, Australia, <sup>3</sup>Medical Scientist Training Program, Mayo Clinic, Rochester, MN, United States, <sup>4</sup>School of Engineering, Deakin University, Geelong, VIC, Australia, <sup>5</sup>Queensland Brain Institute, The University of Queensland, St Lucia, QLD, Australia, <sup>6</sup>Department of Psychiatry and Psychology, Mayo Clinic, Rochester, MN, United States, <sup>7</sup>Department of Psychiatry and Behavioral Science, Emory University, Atlanta, GA, United States, <sup>8</sup>Department of Psychiatry, University of Minnesota, Minneapolis, MN, United States, <sup>9</sup>Department of Psychiatry, University of Minnesota, <sup>10</sup>Department of Psychiatry, Mayo Clinic, Rochester, MN, United States, <sup>11</sup>Division of Engineering, Mayo Clinic, Rochester, MN, United States, <sup>12</sup>Department of Biomedical Engineering, Mayo Clinic, Rochester, MN, United States

**Introduction:** Opioids are the leading cause of overdose death in the United States, accounting for almost 70,000 deaths in 2020. Deep brain stimulation (DBS) is a promising new treatment for substance use disorders. Here, we hypothesized that VTA DBS would modulate both the dopaminergic and respiratory effect of oxycodone.

**Methods:** Multiple-cyclic square wave voltammetry (M-CSWV) was used to investigate how deep brain stimulation (130 Hz, 0.2 ms, and 0.2 mA) of the rodent ventral segmental area (VTA), which contains abundant dopaminergic neurons, modulates the acute effects of oxycodone administration (2.5 mg/kg, i.v.) on nucleus accumbens core (NAcc) tonic extracellular dopamine levels and respiratory rate in urethane-anesthetized rats (1.5 g/kg, i.p.).

**Results:** I.V. administration of oxycodone resulted in an increase in NAcc tonic dopamine levels (296.9 ± 37.0 nM) compared to baseline (150.7 ± 15.5 nM) and saline administration (152.0 ± 16.1 nM) (296.9 ± 37.0 vs. 150.7 ± 15.5 vs. 152.0 ± 16.1, respectively, p = 0.022, n = 5). This robust oxycodone-induced increase in NAcc dopamine concentration was associated with a sharp reduction in respiratory rate (111.7 ± 2.6 min<sup>-1</sup> vs. 67.9 ± 8.3 min<sup>-1</sup>; pre- vs. postoxycodone; p < 0.001). Continuous DBS targeted at the VTA (n = 5) reduced baseline dopamine levels, attenuated the oxycodone-induced increase in

Abbreviations: AAALAC, Association for Assessment and Accreditation of Laboratory Animal Care International; CFM, carbon fiber microelectrod; DBS, deep brain stimulation; FSCV, fast scan cyclic voltammetry; IACUC, Institutional Animal Care and Use Committee; M-CSWV, multiple cyclic square wave voltammetry; NAc, nucleus accumbens; NAcc, nucleus accumbens core; VTA, ventral tegmental area.

dopamine levels to (+39.0% vs. +95%), and respiratory depression (121.5  $\pm$  6.7 min<sup>-1</sup> vs. 105.2  $\pm$  4.1 min<sup>-1</sup>; pre- vs. post-oxycodone; p = 0.072).

**Discussion:** Here we demonstrated VTA DBS alleviates oxycodone-induced increases in NAcc dopamine levels and reverses respiratory suppression. These results support the possibility of using neuromodulation technology for treatment of drug addiction.

KEYWORDS

substance use disorder, deep brain stimulation, nucleus accumbens, ventral tegmental area, oxycodone, dopamine, rodent

## Introduction

Opioids are the leading cause of overdose death in the United States, accounting for almost 70,000 deaths in 2020 (Centers for Disease Control and Prevention, 2022). The main side-effect of opioid use is dose tolerance, to which chronic users are particularly susceptible (Ballantyne and Mao, 2003). Current opioid use disorder treatments typically include replacement therapy (e.g., methadone hydrochloride and buprenorphine) and psychosocial interventions (Polydorou et al., 2017; Volkow et al., 2019). Although there has been some success with newer treatments such as buprenorphine-naloxone therapy (Weiss et al., 2011), the relapse rate remains unacceptably high. Indeed, abstinence from opioid use remains less than 30% after 10-30 years (Hser et al., 2015). Deep brain stimulation (DBS) has recently been utilized for the treatment of neuropsychiatric diseases, including depression (Mayberg et al., 2005; Yuen et al., 2021a), Tourette's syndrome (Baldermann et al., 2016), obsessive-compulsive disorder (Nuttin et al., 1999; Denys et al., 2010; McLaughlin et al., 2021), and drug addiction (Chen et al., 2019; Yuen et al., 2022a). With promising findings from recently published studies, there are ongoing clinical trials to assess the long-term effect of DBS for treating drug addiction, particularly focusing on the nucleus accumbens (NAc) as a target (Zhou et al., 2011; Valencia-Alfonso et al., 2012; Chen et al., 2019; Clinical Trials, 2019; Qu et al., 2019; Zhang et al., 2020; Mahoney et al., 2021).

However, electrical stimulation is inherently non-selective in what neuronal elements and terminals are activated. Thus, Nac DBS is releasing many different neurotransmitters in the Nac (e.g., glutamate, dopamine, acetylcholine) which does not address what neurotransmitter may be the most relevant in mitigating addiction. In opioid use disorder, dopamine is one of the key monoamine neurotransmitters that mediates incentive-motivation which plays an important role in drug reinforcement and compulsive drug-seeking behaviors. Opioids are known to stimulate µ-opioid receptors on GABAergic nerve terminals synapsing on dopamine neuronal cells in the ventral tegmental area (VTA) producing both dopaminedependent and dopamine-independent positive reinforcement (Ting and van der Kooy, 2012). Activation of these presynaptic µ-opioid receptors reduces GABAergic inhibition of dopamine neurons, leading to an increase in dopamine cell firing and production and release of dopamine in the NAc.

Given that opioid driven dopamine release in the NAc is considered an important factor in opioid use disorder, rather than stimulating the NAc the present study instead examined the effects of VTA DBS, which would specifically interfere with opioid-induced disinhibition of dopamine cell firing by GABA interneuronal cells in the VTA as a means of attenuating the effects of opioids. Normally, the prefrontal cortex provides negative feedback to overcome drives to perform unsafe or unwise activities. However, this feedback system becomes compromised with repeated drug administration and addiction (Kosten and George, 2002; Barrot, 2015). Repeated administration results in drug tolerance, characterized by reduced dopamine release after normally rewarding activities. This leads to dependence and compulsive drug-seeking behavior (Kosten and George, 2002). The neurocircuitry involved in the behavioral changes may be better understood by measuring dopamine accurately, particularly during therapeutic treatments. In addition, dopamine has parallel effects on respiration, generally enhancing  $CO_2$  and pH dependent respiratory drive (Lalley, 2008). To our knowledge, no studies have investigated the efficacy of DBS in attenuating opioid-induced respiratory depression.

Microdialysis is the current gold standard to measure dopamine release into the NAc extracellular space (Saigusa et al., 2021). However, newer electrochemical techniques such as multiple cyclic square wave voltammetry (M-CSWV) can measure tonic (basal) dopamine levels with higher spatio-temporal resolution and causes less trauma to the brain tissues of interest (Oh et al., 2018; Rusheen et al., 2020; Yuen et al., 2021b).

In the current proof-of-concept study, we hypothesized that VTA DBS would provide modulation of the oxycodone effect on tonic dopamine levels, as well as its ability to reduce the suppressive effect on respiration. To achieve this, we first elucidated oxycodone-induced dopaminergic changes *in vivo* using M-CSWV and simultaneously measured the respiratory rate. Then, we performed DBS of the VTA while simultaneously measuring oxycodone-induced changes in dopamine extracellular levels in the core of the nucleus accumbens (NAcc) and respiratory rate changes caused by acute intravenous oxycodone administration.

## Material and methods

#### Animal subjects

Adult male Sprague-Dawley rats (n = 10, weighing 250–300 g; Envigo, IN, United States) were used for this study. Rats were kept in group housing in an Association for Assessment and Accreditation of Laboratory Animal Care International (AAALAC) accredited vivarium following a standard 12-h light/dark cycle at constant temperature (21°C) and humidity (45%) with *ad libitum* food and water. Animal studies were approved by the Institutional Animal Care and Use Committee (IACUC), Mayo Clinic, Rochester. The NIH Guide for the Care and Use of Laboratory Animals guidelines (Department of Health and Human Services, NIH publication No. 86-23, revised 1985) were followed for all aspects of animal care.

# Dopamine sensing microelectrode fabrication

CFMs were fabricated using an established standardized CFM design at Mayo Clinic (Chang et al., 2013; Oh et al., 2016). Further details, including preparation of the Ag/AgCl counter-reference electrode can be found in our group's previous publications (Yuen et al., 2021b; Yuen et al., 2022b). CFMs were pretested in TRIS buffer prior to coating deposition with a PEDOT:Nafion deposition solution (Vreeland et al., 2015), which minimized the effect of biofouling *in vivo*.

# Implantation of recording and stimulating electrodes

Each rat was anesthetized with urethane (1.5 g/kg i.p.; Sigma-Aldrich, St Louis, MO, United States) and administered buprenorphine (0.05-0.1 mg/kg s.c., Par Pharmaceutical, Chestnut Ridge, NY, United States) for analgesia. Following anesthesia, the rat was placed in a stereotaxic frame (David Kopf Instruments, Tujunga, CA, United States). Respiratory rate (RespiRAT, Intuitive Measurement Systems, AZ, United States) and hind-paw and tail pinch were used to monitor the physiological state and depth of anesthesia, respectively. RespiRAT is a pressure monitor that only counts the peaks of pressure to mitigate interference from noise, and automatically adjusts the detection threshold with the amplitude of the incoming data. Using a standard rat brain atlas (Paxinos and Watson, 2007), three trephine holes were drilled, the first for placement of a CFM into the NAcc (all coordinates from bregma: AP 1.2 mm, ML 2.0 mm, DV 6.5-7.5 mm from dura), the second for a stimulating electrode into the VTA (twisted bipolar stimulating electrode-Plastics One, MS 303/2, Roanoke, VA, United States, with the tips separated by ~1 mm; AP -5.3 mm, ML 0.9 mm, DV 7.5-9 mm from dura), and a third for an Ag/AgCl reference electrode into the contralateral cortex (Clark et al., 2010).

#### Recordings and stimulation parameters

The depths of the stimulating electrode in the VTA and CFM in the NAcc were first adjusted to obtain a robust stimulation-evoked dopamine signal as measured by fast scan cyclic voltammetry (FSCV) (-0.4 to 1.3 V sweep; 10 Hz). Stimulation parameters were biphasic pulses at 60 Hz, 2 ms pulse width, 0.2 mA, and 2 s duration. Stimulation and FSCV were both performed using the WINCS Harmoni system (Lee et al., 2017), a wireless stimulation and neurochemical sensing system.

Once the optimal electrode depths were identified, the system was switched to the M-CSWV sensing technique (Oh et al., 2018). After at least 60 min of stabilization, the rats were treated according to one of the two experiments. For Experiment 1 (N = 5), i.v. saline

(1 mL/kg) was first administered as a negative control. After 30 min, i.v. oxycodone (provided by National Institute on Drug Abuse, Bethesda, MD; 2.5 mg/kg dissolved in 0.5 mL of normal saline) was administered (infused over 2 min via a cannula inserted into the tail vein) and the rat was further observed for another 30 min.

Experiment 2 consisted of the stimulation group (separate set of animals; N = 5), VTA biphasic pulse stimulation was applied at 130 Hz (0.2 ms pulse width, 0.2 mA) continuously for 60 min. The delivered stimulation was interleaved with the M-CSWV recording to minimize artifacts. Once the signal was restabilized to a new plateau (after 30 min of stimulation), i.v. oxycodone was administered (dose and method as above) while stimulation and recording continued. After another 30 min of observation, the stimulation was turned off. The animal was observed for another 30 min post-stimulation before being euthanized using Fatalplus injection (pentobarbital 390 mg/mL i.p.; 10 mL).

Saline administration during VTA stimulation was shown to have no effect on tonic dopamine levels in our previous study and was therefore not repeated here [Figures 4B, 5B of (Yuen et al., 2023)].

### Calibration of electrodes

After experimentation, changes in dopamine release recorded from individual CFMs were calibrated *in vitro* with dopamine solutions of different known concentrations. This was performed in a similar manner to previously described procedures (Oh et al., 2018).

### Statistical analysis

For comparison between different dopamine concentrations, repeated-measures one-way (Experiment 1) and two-way (Experiment 2) ANOVA tests were performed. The levels for comparison were all calculated by averaging over 10 data points, i.e., 100 s. In Experiment 1, the levels include: "Baseline" (centered at 5 min prior to saline injection), "Saline" (same post-administration time interval as the "Oxycodone" measurement for fair comparison), and "Oxycodone (Peak)" (centered at the peak of dopamine levels after oxycodone was administered). Post-hoc two-tailed paired *t*-tests were also conducted in the Experiment 1 between sequential measurements (Baseline vs. Saline; Saline vs. Oxycodone).

In Experiment 2, two-way ANOVA test was performed across: "Baseline" (centered at 5 min prior to start of stimulation), "Stimulation" (centered at 5 min prior to administering oxycodone); "Oxycodone" (centered at the peak of dopamine levels after oxycodone was administered), and "post-stimulation" (centered at 15 min after stimulation stops).

For comparison between respiratory rates, two-tailed paired *t*-tests were also conducted. Respiratory rates are measured by calculating the moving average over 30 s. Measurements were taken approximately 1/54 s apart. The levels for comparison were calculated by averaging over 10 data points. In Experiment 1, a comparison was made between the levels post-saline (same post-administration time interval as the post-oxycodone measurement for fair comparison) and the levels post-oxycodone (centered at troughs). Likewise, in Experiment 2, a



#### FIGURE 1

Changes in NAcc tonic dopamine concentrations after saline and oxycodone. (A) Experimental design and procedure. All electrodes were implanted stereotactically into the brain of urethane-anesthetized rats and positioned at the optimal depth. Tonic dopamine levels in the NAcc were recorded throughout the experiments. Saline and oxycodone were administered i.v. accordingly. (B) Rapid increase in dopamine was seen after i.v. oxycodone administration (2.5 mg/kg) compared to i.v. saline (1 mL/kg). Arrow denotes time of oxycodone administration. (C) One-way ANOVA demonstrated significant changes among the levels (F = 12.80; p = 0.0224). *Post-hoc* analysis showed saline administration did not significantly alter tonic dopamine levels ( $+1.2 \pm 4.4$  nM, +0.8%, N = 5 rats, p = 0.1047); whereas oxycodone rapidly increased dopamine levels within ~5 min of administration (+145.0  $\pm$  41.9 nM, +95%, N = 5 rats, p = 0.0251). \* denotes p < 0.05. (D) Representative high dimensional color plots and matched voltammogram at baseline, and after saline and oxycodone administration, respectively. The circle denotes dopamine oxidation on the color plot measured by M-CSWV. Figure partly created with BioRender.com.

comparison was made between the levels post-stimulation (10 min prior to oxycodone administration) and the levels post-oxycodone (centered at troughs). Statistical analyses were performed and graphs were constructed using PRISM 8 (GraphPad). All error bars and shaded areas are represented as S.E.M. Statistical significance was set at p < 0.05. No



#### FIGURE 2

Stimulation suppresses oxycodone-associated dopamine increases. (A) Experimental design and procedure. All electrodes were implanted stereotactically into the brain of urethane-anesthetized rats and positioned at the optimal depth. Tonic dopamine levels in the NAcc were recorded throughout the experiments. VTA DBS (biphasic 130 Hz, 200  $\mu$ s pulse width, 0.2 mA) was applied (black bar). Oxycodone (2.5 mg/kg) was given intravenously. (B) Stimulation suppressed tonic dopamine levels. The levels increased after oxycodone administration but remained below the prestimulation baseline. After cessation of stimulation, the levels returned to approximately baseline. Black bars represent the stimulation period. The red arrow denotes drug administration. (C) Two-way ANOVA test demonstrated stimulation [F(1,4) = 30.48; p = 0.0053] and oxycodone [F(1,4) = 21.81; p = 0.0053] both had a statistically significant effect on tonic dopamine levels (N = 5 rats). (D) Representative high dimensional color plots and matched voltammogram corresponding to the time points. The circle and arrow denote the site of dopamine oxidation on the color plot measured by M-CSWV. Figure partly created with BioRender.com.

Bonferroni corrections to *t*-tests were applied as sequential comparisons were set up for testing a pre-planned hypothesis (sequential changes).

# Results

To determine the viability of VTA DBS as a potential therapy for opioid intoxication, dopamine levels were first monitored in response to oxycodone injection. A saline injection was performed before the oxycodone injection as shown in Figure 1A. Using M-CSWV, it was demonstrated that there was a significant increase in NAcc tonic dopamine levels after oxycodone administration (Figure 1B). A one-way ANOVA demonstrated significant changes among the baseline, saline, and oxycodone (peak) dopamine levels (p = 0.0224). Post-hoc analysis showed saline did not significantly alter tonic dopamine levels ( $+1.2 \pm 4.4$  nM, +0.8%, N = 5 rats, p = 0.1047) (Figure 1C). In contrast, oxycodone rapidly increased tonic dopamine levels within 10 min from 157.7  $\pm$  41.4 nM to 296.9  $\pm$  82.8 nM (n = 5 rats, p = 0.0251).



no stimulation was applied (p < 0.001) but this was reversed when stimulation was applied (p = 0.072) (N = 5 for each group). \*\*\* denotes p < 0.001.

This is also highlighted in the high dimensional color plots, and their voltammograms in Figure 1D Higher dopamine concentrations are displayed as more intense red.

Next, we explored how VTA DBS affects tonic dopamine levels in the NAcc, including after oxycodone administration. As shown in Figure 2A, continuous VTA DBS was applied for 1 h. During the stimulation oxycodone (i.v.) was administered while monitoring changes in tonic dopamine levels. After 30 min, DBS was turned off and tonic dopamine levels were recorded for an additional 30 min. With VTA DBS, tonic dopamine levels in the NAcc decreased by 43.8%, from 135.4  $\pm$  43.8 nM to 76.1  $\pm$  32.7 nM (Figure 2B). These levels were augmented by oxycodone administration to 105.8  $\pm$ 35.0 nM (+39.0%) but the levels did not return to baseline until stimulation was switched off. After VTA DBS was turned off, tonic dopamine levels gradually returned to baseline. ANOVA analysis demonstrated stimulation [F(1,4) = 30.48; p = 0.0053] and oxycodone [F(1,4) = 21.81; p = 0.0095] both had a statistically significant effect on tonic dopamine levels (Figure 2C). Interaction between the two factors was also found to be significant [F(1,4) =130.5; p = 0.0003]. This was also reflected in the less intense red color on the color plots (Figure 2D).

One consideration was that VTA DBS may alleviate the respiration-suppressing effects of acute opioid administration.

Respiration rate was monitored and compared for saline and oxycodone treatment in the absence of VTA DBS, and oxycodone treatment during VTA DBS (Figure 3A). Oxycodone was shown to decrease the respiratory rate by 36.6% relative to baseline (p < 0.001). VTA stimulation partially alleviated respiratory suppression from 36.6% to 13.4% relative to baseline (p = 0.072) (Figure 3B). The clear trough in respiratory rate secondary to oxycodone was also less visible when stimulation was applied. Although there appears to be an increase in respiratory rates after VTA DBS was initiated (prior to oxycodone), compared to baseline just before VTA DBS, there was no statistical differences (p = 0.148).

## Discussion

The present study measured tonic dopamine levels in the NAcc and accompanying respiratory rate following i.v. oxycodone administration. The facilitatory increase in dopamine levels is consistent with previous microdialysis studies with opioids (Vander Weele et al., 2014; Saigusa et al., 2017). The rise in dopamine levels is also similar to our previous studies on the effect of i.v. cocaine on accumbal tonic dopamine

10.3389/fphar.2023.1199655

levels, and is concordant with a critical role of dopamine across addictive agents (Solinas et al., 2019; Yuen et al., 2021b). In addition, VTA DBS was also shown to significantly suppress baseline tonic dopamine levels in the NAcc and can further significantly attenuate the ability of oxycodone to elevate dopamine levels, which is consistent with our previous study with cocaine (Yuen et al., 2023). The respiratory rate depression following oxycodone administration was also partially reversed by VTA DBS.

While FSCV has been utilized in studying short-term effects of opioids on *phasic* (burst) dopamine release (Vander Weele et al., 2014), to our knowledge, no studies have used electrochemical techniques to study the effects of opioids on *tonic* dopamine concentrations.

#### Potential mechanism of action of VTA DBS

Continuous application of VTA DBS resulted in a consistent suppression of NAcc tonic dopamine levels. This also attenuated the facilitatory effects of oxycodone on tonic dopamine levels, with dopamine levels returning to baseline after cessation of DBS. The depletion of presynaptic dopamine vesicular stores may cause this reversible effect. In this case, an initial peak would be expected. Although this was not apparent in the averaged data (Figure 2B), an initial and brief peak increase was indeed seen in three out of five rats (60%) that underwent stimulation. We defined an initial peak to be one that occurs within 1 min (6 MCSWV readings) that is at least more than 2 standard deviations of the 10 averaged points used to calculate the baseline prior to stimulation. The average increase in dopamine among the three rats was 14.4 ± 3.7 mM. This is consistent with the phasic (evoked) increases of dopamine in the NAcc (measured by FSCV) that were seen with brief (2 s) stimulation of the VTA during the targeting progress. This may not be seen universally as the M-CSWV technique has a time resolution of 10 s and if the release of dopamine occurred prior to the following measurement, the peak may be much smaller. Previous studies have shown that prolonged intermittent VTA stimulation was associated with reduced dopamine levels in the NAc, likely due to depletion of accumbal presynaptic dopamine vesicular stores or autoreceptor activation (Hernandez et al., 2006; Hernandez and Shizgal, 2009). This is consistent with the phenomenon seen in the current study. Another explanation may be activation or inhibition of other inhibitory and excitatory neurotransmitters. Examples include GABA (inhibitory), dopamine (inhibitory), glutamate (excitatory), and acetylcholine (facilitatory), all of which have been found in the VTA in microdialysis studies (Westerink et al., 1996; Lester et al., 2010). It is well known that opiates increase firing activity of dopamine neurons at least in part by decreasing the inhibitory effect of GABA via activation of µ-opiate receptors on GABAergic interneuron terminals in the VTA, and glutamate transmission is required for morphine's effect on dopamine cells (Barrot, 2015; Saigusa et al., 2021). Therefore, DBS may exert its effect on extracellular dopamine levels via modulation of these other neurotransmitters.

One further possibility is that continuous electrical stimulation of the VTA may induce a relatively rapid depolarization block (DB) similar to chronic antipsychotic treatments that, in turn, decrease tonic DA levels in the NAcc (Lane and Blaha, 1987; Grace et al., 1997; Boye and Rompre, 2000; Hernandez et al., 2016). These hypotheses can be investigated by further studies using other pharmacological tests and measuring techniques such as amperometry.

### Potential clinical applications

VTA DBS is currently being used as an experimental treatment for treatment-refractory cluster headache (Akram et al., 2016). Interestingly, it has been shown that elevation of peripheral dopamine levels is implicated in cluster headache pathophysiology, suggesting cluster headaches may be a consequence of overactivity of the dopaminergic and autonomic systems (D'Andrea et al., 2006; D'Andrea et al., 2019). Therefore, the therapeutic effect of VTA DBS on cluster headache may be explained by its suppressive effect on forebrain dopamine levels.

With regards to the potential applicability of DBS for substance use disorder, no human studies have been reported using VTA DBS. In preclinical studies, one study demonstrated continuous optogenetic stimulation of VTA dopaminergic neurons can reduce ethanol selfadministration in mice (Bass et al., 2013). Although optogenetic and electrical stimulation have different underlying mechanisms of action, one study demonstrated that both modalities could activate similar brain regions (Weidner et al., 2020).

In the current study, the NAc core was chosen as the main target for dopamine measurements for two main reasons. First, from microdialysis studies, extracellular dopamine in the core was higher than that in the shell, whereas norepinephrine levels (a potential electroactive interferent) were higher in the shell (McKittrick and Abercrombie, 2007). Second, these are acute experiments where the animals are exposed to the drug for the first time. It has been suggested that the NAcc underlies the phenomenon of initial acquisition of drug taking, cue-elicited drug seeking behavior, and impulsive choices (Di Chiara, 2002; Salgado and Kaplitt, 2015). Although we did not perform comparison studies using the accumbens shell subregion, it remains an interesting target for future studies using opioids in combination with VTA DBS. VTA DBS may modulate drug-seeking behavior by suppressing dopamine levels in the NAcc. This may also be applicable to hyperdopaminergic pathologies such as mania, schizophrenia, and dopamine dysregulation syndrome (Berk et al., 2007; Grace, 2016; Ashok et al., 2017).

Consistent with previous microdialysis studies (Westerink et al., 1996; Lester et al., 2010), oxycodone induced significant increases in NAcc dopamine levels, which likely account for the initiating steps in the pathophysiology of opioid use disorder. The suppression of NAcc dopamine levels appeared reversible within 60 min of DBS but this may not be the case for longer stimulation duration or repeated stimulations. Given dopamine is associated with both drug-related and pharmacological rewards, depression of dopamine levels in a normal dopaminergic state may theoretically lead to anhedonia and depression. Therefore, more studies need to be conducted before widely implementing this potential therapeutic option. In addition, the present study suggests that intermittent or continuous measurement of tonic dopamine levels may serve as a potential biomarker for closed-loop DBS for substance use disorder. However, the risks of implanting a separate sensing electrode and the ethics of such interventions should be considered (Lo et al., 2023).

The cases of opioid overdose in the United States have more than quadrupled between 2000 and 2020 and, without further intervention, is expected to double by 2029 (Humphreys et al., 2022). Therefore, it is of interest that the current study demonstrated that VTA DBS may be used to reduce the effects of oxycodone and therefore potentially prevent overdose deaths. Given the volume of tissue affected by electrical stimulation, it is highly likely that other brain structures, such as descending white matter fibers that control the motor aspect of respiratory effort, were co-stimulated, leading to the observed attenuating effect on oxycodone-associated respiratory suppression. Several studies using intracranial self-stimulation also showed that DBS can alter the physiological parameters such as the respiratory rate, demonstrating the complex interplay between brain stimulation and the autonomic nervous system (Malmo, 1963; Angyan, 1981; Burgess et al., 1993). One more possible explanation is that stimulation of the VTA would likely also stimulate the substantia nigra (SN) given its proximity to the VTA. There are known anatomical connections between the SN compacta and respiratory control centers, including a direct SNc pathway to the pre-Bötzinger complex (Zhou et al., 2011). The pre-Bötzinger complex interneurons have direct connections to caudal areas containing bulbospinal neurons that are directly involved in generating rhythm to the premotor neurons that transmit oscillatory drive to spinal respiratory motoneurons (Mahoney et al., 2021). Indeed, endogenous activation of D1Rs acts against opioid depression of the respiratory network (Qu et al., 2019). Nevertheless, to our knowledge, this is the first study that demonstrated DBS can attenuate the respiratory effect of opioids. Further optogenetic studies that focus on particular neurons such as dopamine or glutamate neurons, may be used to further discern the target that led to this effect. Also, tractography, particularly in larger animal studies, may be used to map the relationship between the volume of tissue activated by DBS and white matter tracts near the VTA.

Limited historical precedence or evidence implicates DBS as a potential procedure for respiration modulation. One study showed subthalamic nucleus stimulation may improve upper respiratory and laryngeal control in Parkinson's patients (Hammer et al., 2010), but no lower respiratory effect (intercostal muscles and diaphragm) was measured. Our results suggest that, in addition to opioid overdose, diseases characterized by respiratory insufficiency or failure, such as intractable sleep apnea, may be treated by DBS.

#### Experimental limitations and future studies

This proof-of-concept preclinical study utilized anesthetized, naïve rats as subjects rather than a chronic addiction model. Although a relatively high acute dose of oxycodone was administered that induced a clinically corollary of respiratory depression, there were no measures for addictive behaviors such as drug-seeking and compulsion. Future studies in awake animals, particularly in drug self-administration paradigms, will be required to validate the dopaminergic effects, as well as behavioral and respiratory benefits.

While dopamine plays a key role in the process of drug addiction, there are also studies which showed opioids may have

a dopamine-independent reward mechanism (Ettenberg et al., 1982; Pettit et al., 1984; Nader and van der Kooy, 1997; Hnasko et al., 2005). Therefore, how much VTA DBS may alter the behavioral changes should be further studied.

## Conclusion

The present study demonstrated that M-CSWV served as a reliable tool to measure changes in tonic extracellular dopamine levels in the NAcc following acute i.v. oxycodone administration. The observation that VTA DBS at least partially reversed the opioid-induced increase in NAcc dopamine levels and respiratory rate depression provides the basis for exploring the use of DBS to treat opioid use disorder and overdose.

# Data availability statement

The raw data supporting the conclusion of this article will be made available by the authors, without undue reservation.

# **Ethics statement**

The animal study was reviewed and approved by the Mayo Clinic Institutional Animal Care and Use Committee.

## Author contributions

KL, YO, HS, and JY conceptualized the study. JY conducted experiments and collected the data. JY, YO, and HS designed and conducted the analyses. JY drafted the first manuscript. KL, HS, and YO supervised all aspects of this work. JY, HS, and YO drafted the figures. All authors contributed to the article and approved the submitted version.

# Funding

The drug used in this study was kindly provided by the National Institute on Drug Abuse (NIDA). This research was supported by the National Institutes of Health (NIH) R01NS112176, R42NS125895-01A1, and R01NS129549 awards. Training grant funding for AR was supported by NIH F31NS115202-01A1, NIH R25GM055252-223, NIH TL1TR002380-03, and NIH T32GM065841-17. MB is supported by a NHMRC Senior Principal Research Fellowship and Leadership 3 Investigator grant (1156072 and 2017131).

# Conflict of interest

The investigators associated with this project and Mayo Clinic have a Financial Conflict of Interest in technology used in the research and that the investigators and Mayo Clinic may stand to gain financially from the successful outcome of the research.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

### References

Akram, H., Miller, S., Lagrata, S., Hyam, J., Jahanshahi, M., Hariz, M., et al. (2016). Ventral tegmental area deep brain stimulation for refractory chronic cluster headache. *Neurology* 86, 1676–1682. doi:10.1212/WNL.000000000002632

Angyan, L. (1981). Interactions between intracranial electrical self-stimulation behavior and cardiovascular responses. J. Auton. Nerv. Syst. 4, 181–194. doi:10.1016/0165-1838(81)90094-1

Ashok, A. H., Marques, T. R., Jauhar, S., Nour, M. M., Goodwin, G. M., Young, A. H., et al. (2017). The dopamine hypothesis of bipolar affective disorder: The state of the art and implications for treatment. *Mol. Psychiatry* 22, 666–679. doi:10.1038/mp.2017.16

Baldermann, J. C., Schuller, T., Huys, D., Becker, I., Timmermann, L., Jessen, F., et al. (2016). Deep brain stimulation for tourette-syndrome: A systematic review and metaanalysis. *Brain Stimul.* 9, 296–304. doi:10.1016/j.brs.2015.11.005

Ballantyne, J. C., and Mao, J. (2003). Opioid therapy for chronic pain. N. Engl. J. Med. 349, 1943–1953. doi:10.1056/NEJMra025411

Barrot, M. (2015). Ineffective VTA disinhibition in protracted opiate withdrawal. *Trends Neurosci.* 38, 672–673. doi:10.1016/j.tins.2015.08.004

Bass, C. E., Grinevich, V. P., Gioia, D., Day-Brown, J. D., Bonin, K. D., Stuber, G. D., et al. (2013). Optogenetic stimulation of VTA dopamine neurons reveals that tonic but not phasic patterns of dopamine transmission reduce ethanol self-administration. *Front. Behav. Neurosci.* 7, 173. doi:10.3389/fnbeh.2013.00173

Berk, M., Dodd, S., Kauer-Sant'anna, M., Malhi, G. S., Bourin, M., Kapczinski, F., et al. (2007). Dopamine dysregulation syndrome: Implications for a dopamine hypothesis of bipolar disorder. *Acta Psychiatr. Scand.* 116, 41–49. doi:10.1111/j.1600-0447.2007. 01058.x

Boye, S. M., and Rompre, P. P. (2000). Behavioral evidence of depolarization block of dopamine neurons after chronic treatment with haloperidol and clozapine. *J. Neurosci.* 20, 1229–1239. doi:10.1523/JNEUROSCI.20-03-01229.2000

Burgess, M. L., Davis, J. M., Wilson, S. P., Borg, T. K., Burgess, W. A., and Buggy, J. (1993). Effects of intracranial self-stimulation on selected physiological variables in rats. *Am. J. Physiol.* 264, R149–R155. doi:10.1152/ajpregu.1993.264.1.R149

Centers for Disease Control and Prevention (2022). Death rate maps & graphs. Berlin, Germany: Springer.

Chang, S. Y., Kimble, C. J., Kim, I., Paek, S. B., Kressin, K. R., Boesche, J. B., et al. (2013). Development of the Mayo investigational neuromodulation control system: Toward a closed-loop electrochemical feedback system for deep brain stimulation. *J. Neurosurg.* 119, 1556–1565. doi:10.3171/2013.8.JNS122142

Chen, L., Li, N., Ge, S., Lozano, A. M., Lee, D. J., Yang, C., et al. (2019). Long-term results after deep brain stimulation of nucleus accumbens and the anterior limb of the internal capsule for preventing heroin relapse: An open-label pilot study. *Brain Stimul.* 12, 175–183. doi:10.1016/j.brs.2018.09.006

Clark, J. J., Sandberg, S. G., Wanat, M. J., Gan, J. O., Horne, E. A., Hart, A. S., et al. (2010). Chronic microsensors for longitudinal, subsecond dopamine detection in behaving animals. *Nat. Methods* 7, 126–129. doi:10.1038/nmeth.1412

Clinical Trials (2019). Feasibility of deep brain stimulation as a novel treatment for refractory opioid use disorder (DBS\_OUD). United States: National Institutes of Health.

D'Andrea, G., Granella, F., Perini, F., Farruggio, A., Leone, M., and Bussone, G. (2006). Platelet levels of dopamine are increased in migraine and cluster headache. *Headache* 46, 585–591. doi:10.1111/j.1526-4610.2006.00407.x

D'Andrea, G., Gucciardi, A., Perini, F., and Leon, A. (2019). Pathogenesis of cluster headache: From episodic to chronic form, the role of neurotransmitters and neuromodulators. *Headache* 59, 1665–1670. doi:10.1111/head.13673

Denys, D., Mantione, M., Figee, M., van den Munckhof, P., Koerselman, F., Westenberg, H., et al. (2010). Deep brain stimulation of the nucleus accumbens for treatment-refractory obsessive-compulsive disorder. *Arch. Gen. Psychiatry* 67, 1061–1068. doi:10.1001/archgenpsychiatry.2010.122

Di Chiara, G. (2002). Nucleus accumbens shell and core dopamine: Differential role in behavior and addiction. *Behav. Brain Res.* 137, 75–114. doi:10.1016/s0166-4328(02) 00286-3

Ettenberg, A., Pettit, H. O., Bloom, F. E., and Koob, G. F. (1982). Heroin and cocaine intravenous self-administration in rats: Mediation by separate neural systems. *Psychopharmacol. Berl.* 78, 204–209. doi:10.1007/BF00428151

Grace, A. A., Bunney, B. S., Moore, H., and Todd, C. L. (1997). Dopamine-cell depolarization block as a model for the therapeutic actions of antipsychotic drugs. *Trends Neurosci.* 20, 31–37. doi:10.1016/S0166-2236(96)10064-3

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Grace, A. A. (2016). Dysregulation of the dopamine system in the pathophysiology of schizophrenia and depression. *Nat. Rev. Neurosci.* 17, 524–532. doi:10.1038/nrn. 2016.57

Hammer, M. J., Barlow, S. M., Lyons, K. E., and Pahwa, R. (2010). Subthalamic nucleus deep brain stimulation changes speech respiratory and laryngeal control in Parkinson's disease. *J. Neurol.* 257, 1692–1702. doi:10.1007/s00415-010-5605-5

Hernandez, G., Cossette, M. P., Shizgal, P., and Rompre, P. P. (2016). Ventral midbrain NMDA receptor blockade: From enhanced reward and dopamine inactivation. *Front. Behav. Neurosci.* 10, 161. doi:10.3389/fnbeh.2016.00161

Hernandez, G., Hamdani, S., Rajabi, H., Conover, K., Stewart, J., Arvanitogiannis, A., et al. (2006). Prolonged rewarding stimulation of the rat medial forebrain bundle: Neurochemical and behavioral consequences. *Behav. Neurosci.* 120, 888–904. doi:10. 1037/0735-7044.120.4.888

Hernandez, G., and Shizgal, P. (2009). Dynamic changes in dopamine tone during self-stimulation of the ventral tegmental area in rats. *Behav. Brain Res.* 198, 91–97. doi:10.1016/j.bbr.2008.10.017

Hnasko, T. S., Sotak, B. N., and Palmiter, R. D. (2005). Morphine reward in dopamine-deficient mice. *Nature* 438, 854–857. doi:10.1038/nature04172

Hser, Y. I., Evans, E., Grella, C., Ling, W., and Anglin, D. (2015). Long-term course of opioid addiction. *Harv Rev. Psychiatry* 23, 76–89. doi:10.1097/HRP.000000000000052

Humphreys, K., Shover, C. L., Andrews, C. M., Bohnert, A. S. B., Brandeau, M. L., Caulkins, J. P., et al. (2022). Responding to the opioid crisis in north America and beyond: Recommendations of the stanford-lancet commission. *Lancet* 399, 555–604. doi:10.1016/s0140-6736(21)02252-2

Kosten, T. R., and George, T. P. (2002). The neurobiology of opioid dependence: Implications for treatment. *Sci. Pract. Perspect.* 1, 13–20. doi:10.1151/spp021113

Lalley, P. M. (2008). Opioidergic and dopaminergic modulation of respiration. *Respir. Physiol. Neurobiol.* 164, 160–167. doi:10.1016/j.resp.2008.02.004

Lane, R. F., and Blaha, C. D. (1987). Chronic haloperidol decreases dopamine release in striatum and nucleus accumbens *in vivo*: Depolarization block as a possible mechanism of action. *Brain Res. Bull.* 18, 135–138. doi:10.1016/0361-9230(87)90042-6

Lee, K. H., Lujan, J. L., Trevathan, J. K., Ross, E. K., Bartoletta, J. J., Park, H. O., et al. (2017). WINCS Harmoni: Closed-loop dynamic neurochemical control of therapeutic interventions. *Sci. Rep.* 7, 46675. doi:10.1038/srep46675

Lester, D. B., Rogers, T. D., and Blaha, C. D. (2010). Acetylcholine-dopamine interactions in the pathophysiology and treatment of CNS disorders. *CNS Neurosci. Ther.* 16, 137–162. doi:10.1111/j.1755-5949.2010.00142.x

Lo, C., Mane, M., Kim, J. H., Berk, M., Sharp, R. R., Lee, K. H., et al. (2023). Treating addiction with deep brain stimulation: Ethical and legal considerations. *Int. J. Drug Policy* 113, 103964. doi:10.1016/j.drugpo.2023.103964

Mahoney, J. J., Haut, M. W., Hodder, S. L., Zheng, W., Lander, L. R., Berry, J. H., et al. (2021). Deep brain stimulation of the nucleus accumbens/ventral capsule for severe and intractable opioid and benzodiazepine use disorder. *Exp. Clin. Psychopharmacol.* 29, 210–215. doi:10.1037/pha0000453

Malmo, R. B. (1963). On central and autonomic nervous system mechanisms in conditioning, learning, and performance. *Can. J. Psychol.* 17, 1–36. doi:10.1037/h0083269

Mayberg, H. S., Lozano, A. M., Voon, V., McNeely, H. E., Seminowicz, D., Hamani, C., et al. (2005). Deep brain stimulation for treatment-resistant depression. *Neuron* 45, 651–660. doi:10.1016/j.neuron.2005.02.014

McKittrick, C. R., and Abercrombie, E. D. (2007). Catecholamine mapping within nucleus accumbens: Differences in basal and amphetamine-stimulated efflux of norepinephrine and dopamine in shell and core. *J. Neurochem.* 100, 1247–1256. doi:10.1111/j.1471-4159.2006.04300.x

McLaughlin, N. C. R., Dougherty, D. D., Eskandar, E., Ward, H., Foote, K. D., Malone, D. A., et al. (2021). Double blind randomized controlled trial of deep brain stimulation for obsessive-compulsive disorder: Clinical trial design. *Contemp. Clin. Trials Commun.* 22, 100785. doi:10.1016/j.conctc.2021.100785

Nader, K., and van der Kooy, D. (1997). Deprivation state switches the neurobiological substrates mediating opiate reward in the ventral tegmental area. *J. Neurosci.* 17, 383–390. doi:10.1523/JNEUROSCI.17-01-00383.1997

Nuttin, B., Cosyns, P., Demeulemeester, H., Gybels, J., and Meyerson, B. (1999). Electrical stimulation in anterior limbs of internal capsules in patients with obsessive-compulsive disorder. *Lancet* 354, 1526. doi:10.1016/S0140-6736(99)02376-4

Oh, Y., Heien, M. L., Park, C., Kang, Y. M., Kim, J., Boschen, S. L., et al. (2018). Tracking tonic dopamine levels *in vivo* using multiple cyclic square wave voltammetry. *Biosens. Bioelectron.* 121, 174–182. doi:10.1016/j.bios.2018.08.034

Oh, Y., Park, C., Kim, D. H., Shin, H., Kang, Y. M., DeWaele, M., et al. (2016). Monitoring *in vivo* changes in tonic extracellular dopamine level by charge-balancing multiple waveform fast-scan cyclic voltammetry. *Anal. Chem.* 88, 10962–10970. doi:10. 1021/acs.analchem.6b02605

Paxinos, G., and Watson, C. (2007). The rat brain in stereotaxic coordinates. Amsterdam, Boston: Academic Press/Elsevier.

Pettit, H. O., Ettenberg, A., Bloom, F. E., and Koob, G. F. (1984). Destruction of dopamine in the nucleus accumbens selectively attenuates cocaine but not heroin self-administration in rats. *Psychopharmacol. Berl.* 84, 167–173. doi:10.1007/BF00427441

Polydorou, S., Ross, S., Coleman, P., Duncan, L., Roxas, N., Thomas, A., et al. (2017). Integrating buprenorphine into an opioid treatment program: Tailoring care for patients with opioid use disorders. *Psychiatr. Serv.* 68, 295–298. doi:10.1176/appi.ps.201500501

Qu, L., Ge, S., Li, N., Wang, W., Yang, K., Wu, P., et al. (2019). Clinical evaluation of deep brain stimulation of nucleus accumbens/anterior limb of internal capsule for opioid relapse prevention: Protocol of a multicentre, prospective and double-blinded study. *BMJ Open* 9, e023516. doi:10.1136/bmjopen-2018-023516

Rusheen, A. E., Gee, T. A., Jang, D. P., Blaha, C. D., Bennet, K. E., Lee, K. H., et al. (2020). Evaluation of electrochemical methods for tonic dopamine detection *in vivo*. *Trends Anal. Chem.* 132, 116049. doi:10.1016/j.trac.2020.116049

Saigusa, T., Aono, Y., and Waddington, J. L. (2021). Integrative opioid-GABAergic neuronal mechanisms regulating dopamine efflux in the nucleus accumbens of freely moving animals. *Pharmacol. Rep.* 73, 971–983. doi:10.1007/s43440-021-00249-9

Saigusa, T., Aono, Y., and Waddington, J. L. (2017). Mechanisms underlying deltaand mu-opioid receptor agonist-induced increases in extracellular dopamine level in the nucleus accumbens of freely moving rats. *J. Oral Sci.* 59, 195–200. doi:10.2334/josnusd. 16-0874

Salgado, S., and Kaplitt, M. G. (2015). The nucleus accumbens: A comprehensive review. *Stereotact. Funct. Neurosurg.* 93, 75–93. doi:10.1159/000368279

Solinas, M., Belujon, P., Fernagut, P. O., Jaber, M., and Thiriet, N. (2019). Dopamine and addiction: What have we learned from 40 years of research. *J. Neural Transm.* (*Vienna*) 126, 481–516. doi:10.1007/s00702-018-1957-2

Ting, A. K. R., and van der Kooy, D. (2012). The neurobiology of opiate motivation. Cold Spring Harb. Perspect. Med. 2, a012096. doi:10.1101/cshperspect.a012096

Valencia-Alfonso, C. E., Luigjes, J., Smolders, R., Cohen, M. X., Levar, N., Mazaheri, A., et al. (2012). Effective deep brain stimulation in heroin addiction: A case report with complementary intracranial electroencephalogram. *Biol. Psychiatry* 71, e35–e37. doi:10. 1016/j.biopsych.2011.12.013

Vander Weele, C. M., Porter-Stransky, K. A., Mabrouk, O. S., Lovic, V., Singer, B. F., Kennedy, R. T., et al. (2014). Rapid dopamine transmission within the nucleus accumbens: Dramatic difference between morphine and oxycodone delivery. *Eur. J. Neurosci.* 40, 3041–3054. doi:10.1111/ejn.12709

Volkow, N. D., Jones, E. B., Einstein, E. B., and Wargo, E. M. (2019). Prevention and treatment of opioid misuse and addiction: A review. *JAMA Psychiatry* 76, 208–216. doi:10.1001/jamapsychiatry.2018.3126

Vreeland, R. F., Atcherley, C. W., Russell, W. S., Xie, J. Y., Lu, D., Laude, N. D., et al. (2015). Biocompatible PEDOT:Nafion composite electrode coatings for selective detection of neurotransmitters *in vivo. Anal. Chem.* 87, 2600–2607. doi:10.1021/ac502165f

Weidner, T. C., Vincenz, D., Brocka, M., Tegtmeier, J., Oelschlegel, A. M., Ohl, F. W., et al. (2020). Matching stimulation paradigms resolve apparent differences between optogenetic and electrical VTA stimulation. *Brain Stimul.* 13, 363–371. doi:10.1016/j. brs.2019.11.005

Weiss, R. D., Potter, J. S., Fiellin, D. A., Byrne, M., Connery, H. S., Dickinson, W., et al. (2011). Adjunctive counseling during brief and extended buprenorphine-naloxone treatment for prescription opioid dependence: A 2-phase randomized controlled trial. *Arch. Gen. Psychiatry* 68, 1238–1246. doi:10.1001/archgenpsychiatry.2011.121

Westerink, B. H., Kwint, H. F., and deVries, J. B. (1996). The pharmacology of mesolimbic dopamine neurons: A dual-probe microdialysis study in the ventral tegmental area and nucleus accumbens of the rat brain. *J. Neurosci.* 16, 2605–2611. doi:10.1523/JNEUROSCI.16-08-02605.1996

Yuen, J., Goyal, A., Rusheen, A. E., Kouzani, A. Z., Berk, M., Kim, J. H., et al. (2021b). Cocaine-induced changes in tonic dopamine concentrations measured using multiplecyclic square wave voltammetry *in vivo. Front. Pharmacol.* 12, 705254. doi:10.3389/ fphar.2021.705254

Yuen, J., Goyal, A., Rusheen, A. E., Kouzani, A. Z., Berk, M., Kim, J. H., et al. (2022b). Cocaine increases stimulation-evoked serotonin efflux in the nucleus accumbens. J. Neurophysiol. 127, 714–724. doi:10.1152/jn.00420.2021

Yuen, J., Goyal, A., Rusheen, A. E., Kouzani, A. Z., Berk, M., Kim, J. H., et al. (2023). High frequency deep brain stimulation can mitigate the acute effects of cocaine administration on tonic dopamine levels in the rat nucleus accumbens. *Front. Neurosci.* 17, 1061578. doi:10.3389/fnins.2023.1061578

Yuen, J., Kouzani, A. Z., Berk, M., Tye, S. J., Rusheen, A. E., Blaha, C. D., et al. (2022a). Deep brain stimulation for addictive disorders-where are we now? *Neurotherapeutics* 19, 1193–1215. doi:10.1007/s13311-022-01229-4

Yuen, J., Rusheen, A. E., Price, J. B., Barath, A. S., Shin, H., Kouzani, A. Z., et al. (2021a). Biomarkers for deep brain stimulation in animal models of depression. *Neuromodulation* 25, 161–170. doi:10.1111/ner.13483

Zhang, C., Li, J., Li, D., and Sun, B. (2020). Deep brain stimulation removal after successful treatment for heroin addiction. *Aust. N. Z. J. Psychiatry* 54, 543–544. doi:10. 1177/0004867419890671

Zhou, H., Xu, J., and Jiang, J. (2011). Deep brain stimulation of nucleus accumbens on heroin-seeking behaviors: A case report. *Biol. Psychiatry* 69, e41–e42. doi:10.1016/j. biopsych.2011.02.012