Check for updates

OPEN ACCESS

EDITED BY Habibu Tijjani, Bauchi State University, Nigeria

REVIEWED BY

Pinarosa Avato, University of Bari Aldo Moro, Italy Roodabeh Bahramsoltani, Tehran University of Medical Sciences, Iran Mithun Rudrapal, Vignan's Foundation for Science, Technology and Research, India

*CORRESPONDENCE Godwin Anywar, ⊠ godwinanywar@gmail.com

RECEIVED 29 May 2023 ACCEPTED 18 December 2023 PUBLISHED 10 January 2024

CITATION

Anywar G and Muhumuza E (2024), Bioactivity and toxicity of coumarins from African medicinal plants. *Front. Pharmacol.* 14:1231006. doi: 10.3389/fphar.2023.1231006

COPYRIGHT

© 2024 Anywar and Muhumuza. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Bioactivity and toxicity of coumarins from African medicinal plants

Godwin Anywar* and Emmanuel Muhumuza

Department of Plant Sciences, Microbiology and Biotechnology, College of Natural Sciences, Makerere University, Kampala, Uganda

Introduction: Coumarins are naturally occuring metabolites from plants and a few micro-organisms. They have been widely used in the food and drug industry in their natural or synthetic forms. Numerous coumarins possess several biological activities such as anti-inflammatory, anti-ulcers, anti-tumour, anti-microbial, anti-coagulant. The aim of this study was to assess the bioactivity, and toxicity of coumarins from African medicinal plants.

Methods: We searched online databases and search engines such as PubMed, Google Scholar and Web of Science for key terms such as coumarins, toxicity, bioavailability, bioactivity with appropriate Boolean operators. Only fulllength research articles published in English between 1956 to 2023 were reviewed.

Results: We recorded 22 coumarins from 15 plant species from Africa. Most of the plant species (33%) were from North Africa. These were followed by East Africa at 21%, then West, and Central Africa at 18.2% each. Most of the coumarins (21.3%) were isolated from the entire plant and the leaves (19.1%) and most of them (46.7%) had some antimicrobial activity. Five coumarins viz osthole, pseudocordatolide C & calanolide, chartreusin and esculetin had either antitumor or anticancer activity. Six coumarins had varying levels and types of toxicity ranging from inhibiting blood clotting as anticoagulants, to cytotoxic effects, causing hyperventilation, tremor, & photophobia, pulmonary haemorrhage, carcinogenic activity, severe neurotoxicity, hepato- and phototoxicity.

Conclusion: Several African medicinal plants are sources of various coumarins that possess several biological activities as well as toxicities. This calls for more research into their safety and efficacy because of their wide spread applications as therapeutic agents.

KEYWORDS

coumarins, bioavailability, safety, therapeutic, medicinal plants, drug discovery

1 Introduction

Coumarins are naturally occurring metabolites in a variety of plants, micro-organisms and in some animal species (Perone, 2016). Coumarin (1,2-benzopyrone; 2H-1-benzopyran-2-one; cis-o-coumarinic acid lactone) is also known as benzopyrone or coumarinic anhydride and tonka bean camphor. It is a white crystalline solid, belonging to the class of lactones. It consists of an aromatic ring fused to a condensed lactone ring (Venugopala et al., 2013). Coumarins usually occur in plants as glycosides and esters but mostly occur in free form. Coumaric metabolites are lactones of 2coumaric acid (2-hydroxy-Z-cinnamic acid) and are constructed by a benzene ring fused to an α -pyrone ring (Figure 1) (Matos et al., 2015). Coumarins are very complex and diverse metabolites. Naturally occurring coumarins may be categorized as coumarins, isocoumarins, furanocoumarins, pyranocoumarins, biscoumarins, and phenylcoumarins (Lacy and O'kennedy, 2004; Zhu and Jiang, 2018) and synthetic pyrone-substituted coumarins (Lacy and O'kennedy, 2004).

The chemical formula of coumarins is $C_9H_6O_2$, and they can exist in various isomeric forms due to differences in the substitution pattern on the benzene and the pyrone ring (Wu et al., 2009). The presence or absence of particular functional groups confers the coumarin its name and properties, such as simple coumarins which are the basic coumarin metabolites with no additional substituents. These include hydroxycoumarins like scopoletin and umbelliferone, methoxycoumarins like esculetin and 7-methoxycoumarin and alkyl coumarins like daphnetin and aesculetin (Wu et al., 2009). Coumarins are variably soluble in most organic solvents or their combinations (Huang et al., 2015) but are freely soluble in ethanol, chloroform, diethyl ether and oils (Jung and Oh, 2011).

Coumarin occurs naturally in many plants and confers a pleasant spice aroma akin to vanilla (Egan et al., 1990). Coumarins were first isolated from Dipteryx odorata (Aubl.) Forsyth f (tonka beans) by Vogel in 1820 (Doctor et al., 2020). They also occur in high concentrations in some essential oils, such as Cinnamomum aromaticum (cinnamon bark oil) and Cassia fistula L. (cassia leaf oil). Coumarins have been identified in various plant species from many different families including the fruits of Vaccinium myrtillus L. (bilberry), Rubus chamaemorus L (cloudberry), Camellia sinensis (L.) Kuntze (green tea), Melilotus albus Medik. and Melilotus officinalis (L.) Lam (sweet clover), Galium odoratum (L.) Scop(sweet woodruff), Vanilla planifolia Andrews (vanilla leaves and beans), Lavendula officinalis subsp. angustifolium (lavender) and Myroxylon pereirae (L.) Harms (balsam of Peru) (Kostova et al., 2011; Kicel and Wolbis, 2012). A broad spectrum of coumarins present in both free state and as glucosides have also been found in many other plant species. Several coumarins have been identified, principally as secondary metabolites in green plants, but the exact number is hard to determine due to synthetic coumarins created regularly (Kostova, 2005).



Most commercially used coumarins were synthesized from salicylaldehyde (DeGarmo and Raizman, 1967). High-grade coumarin is still isolated from tonka beans using various methods such as hot water steeping and sub critical water extraction (Doctor et al., 2020). Coumarins are extremely variable in structure leading to various pharmacological activities (Lacy and O'kennedy, 2004). The simplicity and versatility of the coumarin scaffold makes it a convenient starting-point for a wide range of applications (Stefanachi et al., 2018; Annunziata et al., 2020).

1.1 Industrial applications and uses of Coumarins

Although coumarins are widely distributed in the plant kingdom, most of the commercially used ones have been synthetically produced for many years. Coumarins have several applications such as in the perfumery, cosmetic and related industries (Givel, 2003). Large quantities of coumarin, mostly associated with vanillin, are used in the food industry for flavouring chocolates, baked goods, and in flavoured beverages (Givel, 2003). However, since 1954, synthetic vanillin use as a direct additive in tobacco products has been suspended in the United States by the FDA due to the production of hazardous compounds when burnt (Riveiro et al., 2010; Sarker and Nahar, 2017; Prusty and Kumar, 2020). Because of their unique sweet note and stability, coumarins have long been recognized as an important raw material in the fragrance industry as odourenhancers. They help to achieve a long-lasting effect when combined with natural essential oils (Doctor et al., 2020). As such, they are widely used in hand soaps, detergents, lotions and perfumes, in tobacco to enhance its natural aroma, in toothpastes, antiperspirant deodorants, creams, hair sprays, shampoos and many other household products (Perone, 2016). Large quantities of coumarins are used in rubber and plastic materials and in paints and sprays to neutralize unpleasant odours (Kirsch et al., 2016). Coumarins also have significant uses in electroplating, mostly in the automotive industry, to provide high polished quality chrome-plated steel. However, this use has been on the decline (Kirsch et al., 2016).

1.2 Medicinal applications of coumarins

Coumarins have several pharmacological properties with applications in the medical field (Skalicka-Woźniak et al., 2016), as a cardiovascular targeting drug (Najmanova et al., 2015), and as a cancer screening molecule and therapy (Lacy and O'Kennedy, 2004). Coumarins possess both immunomodulatory and antitumor activity (Riveiro et al., 2010). They have been recommended for treatment of a number of clinical conditions, including high protein oedema and brucellosis (Luszczki et al., 2009; Riveiro et al., 2010; Raja et al., 2011). Coumarins such as osthole have undergone clinical trials for treatment of lymphoedema (Farinola and Piller, 2005), following breast cancer treatment and in treatment of lung and kidney cancer and melanoma alone or in combination with cimetidine (Riveiro et al., 2010; Prusty and Kumar, 2020). Coumarins have also been

TABLE 1 Bioactivity of coumarins from African medicinal plants.

	countering from African in	icarcinal plants.		1	1	
Coumarins	Plant source & (family)	Type of extraction and compounds recovery	Part used	Country/Area of collection	Bioactivity	Measure of activity
1. Bicoumarin/ Biscoumarin	Melilotus officinalis (L.) Lam. (Fabaceae)	Ultrasonic bath and ultrasound-assisted extraction using a rotavap (Zhang et al., 2018).	L	South, North Africa. Widely distributed globally	Anticoagulant, antimicrobial	Vitamin K-blockers. Act as anticoagulants by inhibiting the cyclic inter-conversion of both vitamin K, vitamin K epoxide (Eason and Spurr, 1995; Hirsh et al., 2001; Sun et al., 2020).
2. Dicoumarol					Anti-inflammatory, Anticoagulant	Vitamin K-blockers that act as anticoagulants <i>via</i> inhibiting the cyclic inter- conversion of both vitamin K, vitamin K epoxide (Hirsh et al., 2001; Jung and Oh, 2011; Kubrak et al., 2017).
3. Psoralen	<i>Fatoua pilosa</i> Gaudich. (Moraceae) (Chiang et al., 2010).	Organic solvent extraction, column chromatography & Medium performance liquid chromatography (Hsia et al., 2019).	Wp	West, central, East Africa. Grows primarily in the wet tropical biome	Antifungal, Antimycobacterial against <i>Mycobacterium</i> <i>tuberculosis</i> H37Rv, Anti-HIV	MIC = 42 μ g/mL against <i>M.</i> <i>tuberculosis</i> H37Rv (Jadhav et al., 2017). 50 μ M taken with imperatorin inhibited HIV-1 replication in HeLa cells (Sancho et al., 2004).
4. Bergapten					Anti-tumor activity against breast cancer cells, antimycobacterial against <i>Mycobacterium</i> <i>tuberculosis</i> H37Rv, antimicrobial	Bergapten increased NF-Y nuclear translocation through p38 MAPK activation hence treatment induced a block in the G0/G1 phase and increased mRNA and protein levels of p53 and p21waf (Panno et al., 2009; Santoro et al., 2016).
5. Dihydromammea	<i>Mammea africana</i> Sabine (Guttiferae)	Methanol extracts separated by liquid–liquid extraction, & open column chromatography (Chakthong et al., 2012; Crichton and Waterman, 1978).	St/Sd	West Africa. Native range is W. Tropical Africa to SW Uganda and Angola	Antihypertensive, anti- hyperglycemic	200 mg/kg day showed anti- hypertensive activity when administered orally in rats together with 40 mg/kg day L-NAME (Nguelefack-Mbuyo et al., 2008).
6. Osthole	Ferulago galbanifera (Mill.) W.D.J.Koch (Apiaceae)	Organic solvent extraction and column chromatography (Karakaya et al., 2019).	R	Morocco, but more widely distributed in the Mediterranean	Antioxidant, antimicrobial, antifungal, antitumor, & anticonvulsant, multiple sclerosis	Doses of 253–639 mg/kg, bw in mice showed significant antioxidant activity (Liu et al., 2012; Yang et al., 2010). In convulsant studies, scores were significantly reduced in early treated EAE mice compared to untreated EAE mice (1.42 \pm 0.20 vs3.57 \pm 0.30) (Teng et al., 1994; Zhang et al., 2015).
7. Anthogenol	Aegle marmelos (L.) Corrêa (Rutaceae)	Organic solvent extraction & TLC (Chakthong et al., 2012).	Fr	Tropical Africa	Antibacterial, Anti- emetic	Effective against <i>Enterococcus</i> bacteria strains (Kubrak et al., 2017)
8. Pseudocordatolide C & calanolide	Calophyllum lanigerum Miq. (Calophyllaceae)	Organic extraction and purification using HPLC (McKee et al., 1996).	L	Central & West Africa	Anticancer, Anti-HIV, Antidermatitic	$IC_{50} = 290-351$ mol ratio/ 32 pmol TPA was effective against leukemia progression (Nahar et al., 2020), $IC_{50} =$ 120 µg/mL against breast cancer in cell lines (Sancho et al., 2004).
9. Phellodenol A	Phellodendron amurense Rupr. (Rutaceae) (Wu et al., 2003).		L	East Africa	Antimycobacterial against <i>M. tuberculosis</i>	Effective dose at 60 mg/mL in mice (Reddy et al., 2021).

(Continued on following page)

TABLE 1 (Continued) Bioactivity of coumarins from African medicinal plants.

Coumarins	Plant source & (family)	Type of extraction and compounds recovery	Part used	Country/Area of collection	Bioactivity	Measure of activity
10. Chartreusin	Dendrobium officinale Kimura & Migo (Orchidaceae) (Zhao et al., 2020).		Fr & Sd	Tropical Africa	Cardioprotective, anti- tumor, antimicrobial	Cytotoxic against Hep3B2.1- 7 (IC ₅₀ = 18.19 μ M) & H1299 (IC ₅₀ = 19.74 μ M) cancer cell lines (Nahar et al., 2020). Antibacterial against <i>S.</i> <i>aureus</i> (IC ₅₀ = 23.25 μ M) in mice (Venugopala et al., 2013).
11. Fraxidin/Fraxin/ Fraxacin	<i>Fraxinus chinensis</i> subsp. <i>rhynchophylla</i> (Hance) A.E.Murray (Oleaceae) (Qu et al., 2021).		В	Algeria, Morrocco	Antiadipogenic, antihyperglycemic	Dosages between 30 and 90 mg/kg in mice decreased TG, CH, LDL, IL-1, IL-6, ICAM-1, NO, levels (Hirsch et al., 2002; Dabur et al., 2018)
12. Calanolide A and B	Calophyllum lanigerum Miq. (Clusiaceae)		L	Central & West Africa	Anti-HIV	IC_{50} = 130 μM in suppressing HIV1 reverse transcriptase (Buckheit et al., 1999). Highly effective against AZT- resistant G-9106 strain of HIV-1 & pyridinone- resistant A17 strain (Boyer et al., 1993).
13. Visnadine/ Visnagin	<i>Ammi daucoides</i> Gaertn. (Apiaceae)		Fl & Fr	North Africa	Muscle dilator	Pectoris, peripheral and coronary muscle vasodilator (Khalil et al., 2020).
14. Esculin	Fatoua pilosa Gaudich. (Moraceae) Fraxinus chinensis subsp. rhynchophylla (Hance) A.E.Murray (Oleaceae)		Wp	Central, West & East Africa Algeria, Morocco	Antiadipogenic Antioxidant	Concentrations of 30–90 mg/kg in mice decreased TG, CH, LDL, IL-1, IL-6, ICAM-1, NO, levels. (Hirsch et al., 2002; Venugopala et al., 2013; Dabur et al., 2018).
15. Esculetin	Bougainvillea spectabilis Willd. (Nyctaginaceae) Cichorium intybus L. (Cichoriaceae)		Sd & St	East Africa but more widely distributed in Asia and South America	Neuroprotective, antioxidant, antiadipogenic, anticancer, anti- inflammatory	Intracerebroventricularly effective, 30 min after administration at a concentration of 20 µg/mL in mice (Wang et al., 2012).
16. Scopoletin	<i>Fatoua pilosa</i> Gaudich. (Moraceae)	Organic solvent extraction and column chromatography (Marealle et al., 2023).	Wp F	Central, West & East Africa Restricted to tropical Africa	Antimycobacterial, hypotensive, muscle relaxant Anti- inflammatory	MIC readings of 58.3 (Reddy et al., 2021) and 42 µg/mL against <i>M. tuberculosis</i> H37Rv (Chiang et al., 2010).
	<i>Tetrapleura tetraptera</i> (Schumach. & Thonn.) Taub. (Fabaceae)					T. tetraptera extract (50–800 mg/kg p.o.) produced dose-related, significant reductions ($p < 0.05-0.001$) of the fresh egg albumin-induced acute inflammation of the rat hind paw oedema compared to Diclofenac (100 mg/kg p.o.) (Ojewole and Adewunmi, 2004).
17. Grandivittin	Ferulago galbanifera (Mill.) W.D.J.Koch (Apiaceae)	Organic solvent extraction and purified by vacuum liquid chromatography (Ahmadi et al., 2016).	R	East Africa	Antibacterial against Helicobacter pylori	MIC = 125 ug/mL against H. pylori (Riveiro et al., 2010).
18. Aegelinol		Organic solvent extraction and column chromatography (Basile et al., 2009).			Antibacterial against Gram-positive and negative bacteria	MIC = 16ug/mL against S. aureus, S. typhi, E. cloaca 32ug/mL against E. earogenes (Riveiro et al., 2010; Venugopala et al., 2013).



used for prevention of dental caries (Perone, 2016). Coumarins have been tested for treatment of schizophrenia, microcirculation disorders and angiopathic ulcers (Skalicka-Woźniak et al., 2016). The vitamin K epoxide reductase (VKORC1), the key enzyme of the vitamin K cycle is the molecular target of coumarins (Oldenburg et al., 2007). Thus, the objective of this study was to assess the bioactivity, and toxicity of coumarins from African medicinal plants.

2 Methods

We performed a literature search using PubMed, Google Scholar and Web of Science. We searched for terms such as coumarins, toxicity, bioavialability, bioactivity with appropriate Boolean operators. Only full-length articles published in English between 1956–2023 were reviewed. The review was limited to African medicinal plant species only. We used only original research articles or review papers and excluded results from *in silico* studies.

3 Results

We recorded 22 coumarins that were isolated from 15 plant species (Table 1). Most of the plant species (33%) were from North Africa, followed by East Africa (21%), then West, and Central Africa at 18.2% each. Most of these coumarins (21.3%) were isolated from the whole plant. These were followed by the leaves (19.1%), then seeds, fruits and roots each at 12.8%. However, only two plant species *Mammea africana* Sabine and *Tetrapleura tetraptera* (Schumach. & Thonn.) Taub. had a more restricted native range to Africa. The rest of the plant species were more widely distributed in different parts of the world. The distribution of the plant species was checked with the Kew database at https://powo.science.kew.org. All the coumarins showed multiple bioactivities.

For instance, esculin had more than six bioactivities. Ten different coumarins had antimicrobial activity including antibacterial, antiviral, antifungal or anti-parasitic activity. Bicoumarin/Biscoumarin and dicoumarol had anticoagulant activity while three coumarins: esculetin, scopoletin, dicoumarol had anti-inflammatory activity. Five coumarins: osthole, pseudocordatolide C and calanolide, chartreusin and esculetin had either antitumor or anticancer activity.

Six coumarins (dicoumarol, coumarin, osthole, esculetin, psoralen, bergapten, Figures 2A–F) had varying levels and types of toxicity ranging from excessive bleeding due to inhibition of blood clotting (anticoagulants), cytotoxic effects causing hyperventilation, tremor, & photophobia, pulmonary haemorrhage, carcinogenic activity, severe neurotoxicity, hepatotoxicity and phototoxicity (Table 2). Osthole, a natural coumarin isolated from dried fruits of *Cnidium monnieri* (L.) Cusson, had anti-allergic activity.

4 Discussion

4.1 Antioxidant properties of coumarins

Osthole, esculin, dihydroxycoumarin and many other coumarins have shown beneficial biochemical profiles in relation to pathophysiological processes dependent upon reactive oxygen species due to the possession of amine, and hydroxy groups (Kostova et al., 2011). A study by Abdizadeh et al. (2020) evaluated the photoprotective activity of some hydroxycoumarins as compared with p-aminobenzoic acid (PABA) as a model sun screen. The activity could be related to their antioxidant actions which could minimize skin photoaging (Abdizadeh et al., 2020). Another study by Mitra et al. (2013) compared the antioxidative activities of seven hydrocoumarins with those of α -tocopherol for the oxidation of tetralin and linoleic acid in a homogeneous solution. Hydrocoumarins exhibited a higher induction period than

Coumarin name	Plant source & (family)	Part (s)	Country	Toxicity	Measure/comment
1. Dicoumarol	<i>Melilotus officinalis</i> (L.) Lam. (Fabaceae)	L	South Africa	Anticoagulant Inhibits blood clotting leading to severe blood loss	Inhibits hepatic synthesis of coagulation factors, acting as a vitamin K antagonist (Yarnell and Abascal, 2009).
2. Coumarin	Aegle marmelos (L.) Corrêa (Rutaceae)	Fr	Nigeria	Cytotoxic effects	$LD_{50} = 196-780$ mg/kg bw with signs of liver toxicity (Mustafa et al., 2021)
3. Osthole	Ferulago galbanifera (Mill.) W.D.J.Koch (Apiaceae) Cnidium monnieri (L.) Cusson (Apiaceae)	R	Morocco	Hyperventilation, acute neurotoxicity, tremor and photophobia	$LD_{50} = 710 \text{ mg/kg}$ bw in mice, 5–50 mg/kg in rats $ED_{50} = 253-639 \text{ mg/kg}$ and acute neurotoxicity at doses of 531–648 mg/kg (Shokoohinia et al., 2017), 5–50 mg/kg in rats (Liu et al., 2012).
4. Esculetin	Bougainvillea spectabilis Willd. (Nyctaginaceae) Cichorium intybus L. (Nyctaginaceae)	Sd & St	East & West Africa	Carcinogen, mildly cytotoxic	A 20 mg/kg bw injection was lethal to rats after 24 h (Hazleton et al., 1956). Oral LD_{50} < 2000 mg/kg bw, intraperitoneal LD_{50} = 1,450 mg/kg bw (Eason and Spurr, 1995).
5. Imperatorin	Angelica archangelica L Angelica dahurica (Hoffm.) Benth. & Hook.f. Ex Franch. & Sav. (Apiaceae)	Wp	North & South Africa	Severe neurotoxicity	TD ₅₀ = 329-443 mg/kg (Hazleton et al., 1956).
6. Psoralen	Fatoua pilosa Gaudich. (Apiaceae)	Wp	Central, West & East Africa	Hepatotoxicity	$LD_{50} = 80 \text{ mg/kg}$ bw in rats & 320 mg/kg in mice resulted into cholestatic liver injuries (Wang et al., 2013).
7. Bergapten	Fatoua pilosa Gaudich. (Apiaceae)	Wp	Central, West & East Africa	Phototoxic	Sun sensitivity to eyes and skin (Wagstaff, 1991).

TABLE 2 Toxicity of coumarins derived from African plants.

Wp = Whole plant, L = leaves, Fr = Fruit, R = root, Sd = Seeds, St = Stem, bw = body weight.

 α -tocopherol in both systems. However, the rate of oxygen absorption during the induction period for the latter was slower than that of the hydrocoumarins in both systems. In addition, 6,7-dihydroxy-4,4-dimethylhydrocoumarin showed less cytotoxicity toward human fibroblasts than 2, 6-di-t-butyl4-methylphenol (Kostova et al., 2011).

4.2 Antimicrobial activity of coumarins

Although different studies have confirmed the antimicrobial activities of various coumarins from plants (Vaou et al., 2021; Ojala et al., 1999; Karakaya et al., 2019), such studies are lacking of African medicinal plants. Some coumarin antibiotics of natural origin, although not necessarily from African plant species have been developed into modern antibiotics. These include NOVOBIOCINTM and CLOROBIOCINTM which are inhibitors of DNA gyrase. They both have a broad spectrum towards Gram-positive bacteria, including methicilin resistant strains of staphylococci species (Raja et al., 2011; Matos et al., 2012). This shows the potential of African plant species as a source of coumarin based drugs. Due to some limitations of these metabolites particularly with regard to solubility, toxicity and development of resistance, a novel series of coumarin analogues has been synthesized (Kirsch et al., 2016). The ester or carboxylic acid on the coumarin ring was found to be important for the inhibitory activity against Gram-positive and Gram-negative bacteria (Chiang et al., 2010; Annunziata et al., 2020). The presence of phenolic hydroxyl groups and or carboxylic acid was found necessary for enhanced activity against *Helicobacter pylori* (Venugopala et al., 2013).

4.3 Anti-inflammatory action of coumarins

Several coumarins like esculetin, dicoumarol isolated from plants or of synthetic origin possess significant anti-inflammatory and or analgesic activities (Kirsch et al., 2016). In a Quantitative Structure-Activity Relationship (QSAR) study for lead optimization in the design of coumarins as potent non-steroidal antiinflammatory agents, the coumarin positions C4- and C7contributed to the high activity (Poumale et al., 2013). Natural products such as esculetin, fraxetin, inhibit lipoxygenase and cyclooxygenase enzymic systems and neutrophil-dependent superoxide anion generation (Kontogiorgis and Hadjipavlou-Litina, 2005).

4.4 Antiviral properties of coumarins

This activity focuses essentially on the inhibition of HIV-1 protease (HIV-PR) and HIV-1 integrase (Sancho et al., 2004). PHENPROCOUMONTM, warfarin and substituted 4-hydroxy-2-pyrone derivatives are actually referred to as the first generation of HIV-PR inhibitors (Hassan et al., 2016). Certain coumarin dimers, particularly those containing hydrophobic moieties on the linker, display potent inhibitory activity against HIV-1 integrase (Sancho et al., 2004).

Calanolide A and B are semi-synthetic coumarins on market used as combination therapies for HIV. Psoralen was found to be a very potent anti-HIV drug during research but was discontinued for its high hepatotoxicity in mice (Sancho et al., 2004; Guo et al., 2020). Phenprocoumon emerged as a novel lead template possessing weak HIV protease inhibitory activity (Hassan et al., 2016). TRIOXSALEN[™] was then developed and marketed as chemical derivative of psoralen despite its photosensitization of the skin (Wishart et al., 2018).

4.5 Antitumor/anticancer properties of coumarins

Osthole, umbelliferon, esculetin and geiparvarin all showed anti-tumor activity. Hydroxycoumarin might be a useful adjuvant for melanoma therapy because of its selective antiproliferative agents that mediate apoptosis in renal carcinoma cells, through modulation of mitogen-activated protein kinases attributed to a nitro group in the aromatic ring (Kostova, 2005; Jung and Oh, 2011). In an attempt by Zhang et al. (2021) to develop novel antitumor candidates, a series of coumarin sulfonamides and amides derivatives were designed and synthetized. The majority of these metabolites showed good cytotoxic activity against MDA-MB-231 and kB cell lines (Zhang et al., 2021). Menezes and Diederich, (2019) conducted an extensive review on the role of natural coumarins as anticancer agents. They showed how coumarins modulate several targets such as carbonic anhydrase, Hsp90, histone deacetylase and topoisomerase enzyme in cancer cells leading to apoptosis.

4.6 Coumarins as enzyme inhibitors

Some natural and synthetic coumarins are cholinesterase inhibitors, which are a promising approach for the treatment of Alzheimer's disease and possible therapeutic applications in the treatment of Parkinsons's disease (Anand et al., 2012). A series of umbelliferon metabolites and synthetic coumarins were evaluated as inhibitors of monoaminooxigenase (MAO) and steroid 5α reductase type I in cell culture systems (Anand et al., 2012). Ether derivatives demonstrate MAO-B selectivity, and sulfonic ester derivatives demonstrate MAO-A selectivity by QSAR studies (Koyiparambath et al., 2021). Calanolide-A drugs are marketed for their ability to block activity of cytochrome P450 3A4, while ethyl biscoumacetate is a glutamine synthase inhibitor (Wishart et al., 2018; Sai et al., 1999).

4.7 Effect of coumarins on cardiovascular system

Many coumarins have different effects on blood. Most notable is warfarin, which has anti-coagulant properties (Hosseini et al., 2015). Warfarin drugs are used for the management of thromboembolism and pulmonary embolism with atrial fibrillation (Wishart et al., 2018). Ethyl biscoumacetate is a semi-synthetic compound used as anticoagulant alternative to warfarin due to fewer side effects (Sun et al., 2020). Esculin and its metabolites are used as vasoprotective agents (Kostova, 2005). Moreover, other coumarins like biscoumarin/bicoumarin have similar activity to ethyl biscoumacetate and esculin (Sun et al., 2020; Najmanova et al., 2015).

4.8 Toxicity of coumarins

There is a large body of data on the toxicity of coumarin in experimental laboratory animals. Coumarins have hepatotoxic and carcinogenic properties in vivo in rodents and other mammalian species. These included adenomas and carcinomas of the liver, lungs and bile ducts and adenomas of the kidney and liver in rats and mice respectively. Carcinomas were found only at doses higher than 100 mg/kg body weight per day (Lake, 1999; EFSA, 2004). In humans, coumarins were approved for clinical use since the 1970s for treating various venous and lymphatic oedemas, as well as tumours including renal cell carcinoma (Marshall et al., 1994). However, their use was eventually discontinued and they were recalled from the market following the development of severe hepatotoxicity in patients after treatment (Cox et al., 1989; WHO, 1995). Coumarins are rapidly absorbed from the gastrointestinal tract after oral administration and extensively metabolized by the liver in the first pass, with only 2%-6% reaching the systemic circulation intact (Guo et al., 2020). Coumarins such as furanocoumarins are toxic to humans. Methoxypsoralen derivatives are potent photosensitizers. They can be activated by near-UV light becoming mutagenic, phototoxic and carcinogenic (Ojala et al., 1999; Ceska et al., 1997). Osthole had an acute intraperitoneal LD₅₀ in mice of 710 mg/kg bw manifesting toxicity as hyperventilation, tremors, and photophobia. In subchronic studies, orally administered osthole in Wistar rats at 5-50 mg/kg bw for 45 days, caused pulmonary haemorrhage and mild reno and hepatoinflammation (Shokoohinia et al., 2017). Esculetin showed a low acute toxicity with an oral LD₅₀ of over 2000 mg/kg bw, the intraperitoneal LD_{50} being 1,450 mg/kg bw (Tubaro et al., 1988). Orally administered psoralen in rats (80 mg/kg bw) caused cholestatic liver injuries (Wang et al., 2019).

5 Conclusion

Several African medicinal plants are sources of various coumarins which possess several biological activities. Coumarins generally have several therapeutic applications. Most of the coumarins in this review (46.7%) had some antimicrobial activity. Six coumarins: osthole, imperatorin/ umbelliferon, pseudocordatolide C & calanolide, chartreusin and esculetin had either antitumor or anticancer activity. They had varying levels and types of toxicity ranging from inhibiting

blood clotting as anticoagulants, cytotoxic effects, causing hyperventilation, tremor, and photophobia. Such plants need to be conserved because of their potential as sources of coumarins and other drugs. This calls for more research into their safety and efficacy because of their wide spread applications as therapeutic agents.

Author contributions

AG conceptualised the study. EM collected data and drafted the first manuscript. AG conducted data analysis and drafted the final manuscript. All authors contributed to the article and approved the submitted version..

References

Abdizadeh, R., Hadizadeh, F., and Abdizadeh, T. (2020). QSAR analysis of coumarin-based benzamides as histone deacetylase inhibitors using CoMFA, CoMSIA and HQSAR methods. *J. Mol. Struct.* 1199, 126961. doi:10.1016/j. molstruc.2019.126961

Ahmadi, F. S. V., Sajjadi, S. E., Shokoohinia, Y., Azizian, H., and Taheripak, G. (2016). Grandivittin as a natural minor groove binder extracted from ferulago macrocarpa to ct-DNA, experimental and *in silico* analysis. *Chemico-Biological Interact.* 258, 89–101. doi:10.1016/j.cbi.2016.08.020

Anand, P., Singh, B., and Singh, N. (2012). A review on coumarins as acetylcholinesterase inhibitors for Alzheimer's disease. *Bioorg. Med. Chem.* 20 (3), 1175–1180. doi:10.1016/j.bmc.2011.12.042

Annunziata, F., Pinna, C., Dallavalle, S., Tamborini, L., and Pinto, A. (2020). An overview of coumarin as a versatile and readily accessible scaffold with broad-ranging biological activities. *Int. J. Mol. Sci.* 21 (13), 4618. doi:10.3390/ijms21134618

Basile, A., Sorbo, S., Spadaro, V., Bruno, M., Maggio, A., Faraone, N., et al. (2009). Antimicrobial and antioxidant activities of coumarins from the roots of *Ferulago campestris* (Apiaceae). *Mol.* (*Basel, Switz.* 14 (3), 939–952. doi:10.3390/ molecules14030939

Boyer, P. L., Currens, M., McMahon, J., Boyd, M., and Hughes, S. (1993). Analysis of nonnucleoside drug-resistant variants of human immunodeficiency virus type 1 reverse transcriptase. *J. Virology* 67 (4), 2412–2420. doi:10.1128/JVI. 67.4.2412-2420.1993

Buckheit, R. W., Jr, White, E. L., Fliakas-Boltz, V., Russell, J., Stup, T. L., Kinjerski, T. L., et al. (1999). Unique anti-human immunodeficiency virus activities of the nonnucleoside reverse transcriptase inhibitors calanolide A, costatolide, and dihydrocostatolide. *Antimicrob. agents Chemother.* 43 (8), 1827–1834. doi:10.1128/AAC.43.8.1827

Chakthong, S., Weaaryee, P., Puangphet, P., Mahabusarakam, W., Plodpai, P., Voravuthikunchai, S. P., et al. (2012). Alkaloid and coumarins from the green fruits of *Aegle marmelos. Phytochemistry*, 75, 108–113. doi:10.1016/j.phytochem.2011. 11.018

Chiang, C. C., Cheng, M. J., Peng, C. F., Huang, H. Y., and Chen, I. S. (2010). A novel dimeric coumarin analog and antimycobacterial constituents from Fatoua pilosa. *Chem. Biodivers.* 7 (7), 1728–1736. doi:10.1002/cbdv.200900326

Cox, D., O'kennedy, R., and Thornes, R. D. (1989). The rarity of liver toxicity in patients treated with coumarin (1, 2-benzopyrone). *Hum. Toxicol.* 8 (6), 501-506. doi:10.1177/096032718900800612

Crichton, E. G., and Waterman, P. G. (1978). Dihydromammea c/ob: a New Coumarin from the seed of Mammea africana. *Phytochemistry* 17 (10), 1783–1786. doi:10.1016/s0031-9422(00)88695-1

Dabur, R., Sharma, B., and Mittal, A. (2018). Mechanistic approach of anti-diabetic compounds identified from natural sources. *Chem. Biol. Lett.* 5 (2), 63–99.

DeGarmo, O., and Raizman, P. (1967). Coumarin. Encycl. Chem. Technol. 2, 425-433.

Doctor, N., Parker, G., Vang, K., Smith, M., Kayan, B., and Yang, Y. (2020). Stability and extraction of vanillin and coumarin under subcritical water conditions. *Molecules* 25 (5), 1061. doi:10.3390/molecules25051061

Eason, C., and Spurr, E. (1995). Review of the toxicity and impacts of brodifacoum on non-target wildlife in New Zealand. *N. Z. J. zoology* 22 (4), 371–379. doi:10.1080/03014223.1995.9518055

EFSA (2004). Opinion of the scientific panel on food additives, flavourings, processing aids and materials in contacts with food (AFC) on a request from the commission related to. *EFSA J.* 104, 1–36. Coumarin; adopted on 6 October 2004.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Egan, D., O'Kennedy, R., Moran, E., Cox, D., Prosser, E., and Thornes, R. D. (1990). The pharmacology, metabolism, analysis, and applications of coumarin and coumarin-related compounds. *Drug Metab. Rev.* 22, 503–529. doi:10.3109/03602539008991449

Farinola, N., and Piller, N. (2005). Pharmacogenomics: its role in re-establishing coumarin as treatment for lymphedema. *Lymphatic Res. Biol.* 3 (2), 81–86. doi:10.1089/ lrb.2005.3.81

Givel, M. (2003). A comparison of US and Norwegian regulation of coumarin in tobacco products. *Tob. Control* 12 (4), 401–405. doi:10.1136/tc.12.4.401

Guo, P.-J., Lin, Z.-J., Zhang, X.-M., Zou, L.-N., Guo, F.-F., and Zhang, B. (2020). Toxicological research and safety consideration of coumarins. *Zhongguo Zhong Yao Za Zhi.* 45 (3), 518–522. doi:10.19540/j.cnki.cjcmm.20190827.401

Hassan, M. Z., Osman, H., Ali, M. A., and Ahsan, M. J. (2016). Therapeutic potential of coumarins as antiviral agents. *Eur. J. Med. Chem.* 123, 236–255. doi:10.1016/j.ejmech. 2016.07.056

Hazleton, L., Tusing, T., Zeitlin, B., Thiessen, R., and Murer, H. (1956). Toxicity of coumarin. J. Pharmacol. Exp. Ther. 118 (3), 348–358.

Hirsch, A.-M., Longeon, A., and Guyot, M. (2002). Fraxin and esculin: two coumarins specific to Actinidia chinensis and A. deliciosa (kiwifruit). *Biochem. Syst. Ecol.* 30 (1), 55–60. doi:10.1016/s0305-1978(01)00064-3

Hirsh, J., Dalen, J. E., Anderson, D. R., Poller, L., Bussey, H., Ansell, J., et al. (2001). Oral anticoagulants: mechanism of action, clinical effectiveness, and optimal therapeutic range. *Chest* 119 (1), 8S–21S. doi:10.1378/chest.119.1_suppl.8s

Hosseini, M., Hosseinzadeh, A., Raufian, K., and Hedjazi, A. (2015). Nontraumatic retroperitoneal hematoma after warfarin administration: fatal case report and review of the literature. *Am. J. forensic Med. pathology* 36 (4), 245–248. doi:10.1097/PAF. 000000000000190

Hsia, C. W., Lin, K. C., Lee, T. Y., Hsia, C. H., Chou, D. S., Jayakumar, T., et al. (2019). Esculetin, a coumarin derivative, prevents thrombosis: inhibitory signaling on PLC γ 2-PKC-AKT activation in human platelets. *Int. J. Mol. Sci.* 20 (11), 2731. doi:10.3390/ ijms20112731

Huang, X., Wang, J., Hao, H., Ouyang, J., Gao, Y., Bao, Y., et al. (2015). Determination and correlation of solubility and solution thermodynamics of coumarin in different pure solvents. *Fluid Phase Equilibria* 394, 148–155. doi:10.1016/j.fluid.2015.03.022

Jadhav, S. B., Fatema, S., Patel, M. A. I., and Farooqui, M. (2017). Novel pyrazoles: M. Tuberculosis growth inhibition and synergistic study. *Chem. Biol. Interface* 7 (2).

Jung, J.-C., and Oh, S. (2011). Practical synthesis of hydroxychromenes and evaluation of their biological activity. *Molecules* 17 (1), 240–247. doi:10.3390/molecules17010240

Karakaya, S., Şimşek, D., Özbek, H., Güvenalp, Z., Altanlar, N., Cavit, C., et al. (2019). Antimicrobial activities of extracts and isolated coumarins from the roots of four ferulago species growing in Turkey. *Iran. J. Pharm. Res. IJPR* 18 (3), 1516–1529. doi:10. 22037/ijpr.2019.1100718

Khalil, N., Bishr, M., Desouky, S., and Salama, O. (2020). Ammi visnaga L., a potential medicinal plant: a review. *Molecules* 25 (2), 301. doi:10.3390/molecules25020301

Kicel, A., and Wolbis, M. (2012). Coumarins from the flowers of Trifolium repens. Chem. Nat. Compd. 48 (1), 130-132. doi:10.1007/s10600-012-0179-3

Kirsch, G., Abdelwahab, A. B., and Chaimbault, P. (2016). Natural and synthetic coumarins with effects on inflammation. *Molecules* 21 (10), 1322. doi:10.3390/molecules21101322

Kontogiorgis, C. A., and Hadjipavlou-Litina, D. J. (2005). Synthesis and antiinflammatory activity of coumarin derivatives. *J. Med. Chem.* 48 (20), 6400–6408. doi:10.1021/jm0580149

Kostova, I. (2005). Synthetic and natural coumarins as cytotoxic agents. Curr. Med. Chemistry-Anti-Cancer Agents 5 (1), 29–46. doi:10.2174/1568011053352550

Kostova, I., Bhatia, S., Grigorov, P., Balkansky, S., S Parmar, V., K Prasad, A., et al. (2011). Coumarins as antioxidants. *Curr. Med. Chem.* 18 (25), 3929–3951. doi:10.2174/092986711803414395

Koyiparambath, V. P., Prayaga Rajappan, K., Rangarajan, T., Al-Sehemi, A. G., Pannipara, M., Bhaskar, V., et al. (2021). Deciphering the detailed structure-activity relationship of coumarins as Monoamine oxidase enzyme inhibitors—an updated review. *Chem. Biol. Drug Des.* 98 (4), 655–673. doi:10.1111/cbdd.13919

Kubrak, T., Podgórski, R., and Stompor, M. (2017). Natural and synthetic coumarins and their pharmacological activity.

Lacy, A., and O'Kennedy, R. (2004). Studies on coumarins and coumarin-related compounds to determine their therapeutic role in the treatment of cancer. *Curr. Pharm. Des.* 10 (30), 3797–3811. doi:10.2174/1381612043382693

Lake, B. (1999). Coumarin metabolism, toxicity and carcinogenicity: relevance for human risk assessment. *Food Chem. Toxicol.* 37 (4), 423–453. doi:10.1016/s0278-6915(99)00010-1

Liu, C. W., Chien, M. W., Su, C. Y., Chen, H. Y., Li, L. J., and Lai, C. C. (2012). Analysis of flavonoids by graphene-based surface-assisted laser desorption/ionization time-of-flight mass spectrometry. *Analyst* 137 (24), 5809–5816.

Luszczki, J. J., Wojda, E., Andres-Mach, M., Cisowski, W., Glensk, M., Glowniak, K., et al. (2009). Anticonvulsant and acute neurotoxic effects of imperatorin, osthole and valproate in the maximal electroshock seizure and chimney tests in mice: a comparative study. *Epilepsy Res.* 85 (2-3), 293–299. doi:10.1016/j. eplepsyres.2009.03.027

Marealle, A. I., Moyo, A. A., Machumi, F., Qwarse, M., Chenyambuga, Y. M., Heydenreich, M., et al. (2023). Antimycobacterial activity of scopoletin from ethanolic extract of *Hymenodictyon floribundum* (Hochst. & Steud.) B.L.Rob. Stem bark. *Sci. Afr.*, 21, e01778. doi:10.1016/j.sciaf.2023.e01778

Marshall, M. E., Kervin, K., Benefield, C., Umerani, A., Albainy-Jenei, S., Zhao, Q., et al. (1994). Growth-inhibitory effects of coumarin (1, 2-benzopyrone) and 7-hydroxycoumarin on human malignant cell lines *in vitro. J. cancer Res. Clin. Oncol.* 120, S3–S10. doi:10.1007/BF01377114

Matos, J. M., Vazquez-Rodriguez, S., Santana, L., Uriarte, E., Fuentes-Edfuf, C., Santos, Y., et al. (2012). Looking for new targets: simple coumarins as antibacterial agents. *Med. Chem.* 8 (6), 1140–1145. doi:10.2174/1573406411208061140

Matos, M. J., Santana, L., Uriarte, E., Abreu, O. A., Molina, E., and Yordi, E. G. (2015). "Coumarins—an important class of phytochemicals," in *Phytochemicals - isolation, characterisation and role in human health.* Editors A. Venket Rao and L. G. Rao (InTech).

McKee, T. C., Fuller, R. W., Covington, C. D., Cardellina, J. H., Gulakowski, R. J., Krepps, B. L., et al. (1996). New pyranocoumarins isolated from *Calophyllum lanigerum* and *Calophyllum teysmannii. J. Nat. Prod.* 59 (8), 754–758. doi:10.1021/np9603784

Menezes, J. C., and Diederich, M. (2019). Translational role of natural coumarins and their derivatives as anticancer agents. *Future Med. Chem.* 11 (9), 1057–1082. doi:10. 4155/fmc-2018-0375

Mitra, I., Saha, A., and Roy, K. (2013). Quantification of contributions of different molecular fragments for antioxidant activity of coumarin derivatives based on QSAR analyses. *Can. J. Chem.* 91 (6), 428–441. doi:10.1139/cjc-2012-0527

Mustafa, Y., Mohammed, E., and Khalil, R. (2021). Synthesis, characterization, and anticoagulant activity of new functionalized biscoumarins. *Egypt. J. Chem.* 64 (8), 4461–4468.

Nahar, L., Talukdar, A. D., Nath, D., Nath, S., Mehan, A., Ismail, F. M., et al. (2020). Naturally occurring calanolides: occurrence, biosynthesis, and pharmacological properties including therapeutic potential. *Molecules* 25 (21), 4983. doi:10.3390/ molecules25214983

Najmanova, I., Dosedel, M., Hrdina, R., Anzenbacher, P., Filipsky, T., Riha, M., et al. (2015). Cardiovascular effects of coumarins besides their antioxidant activity. *Curr. Med. Chem.* 15 (9), 830-849. doi:10.2174/1568026615666150220112437

Nguelefack-Mbuyo, P., Nguelefack, T., Dongmo, A., Afkir, S., Azebaze, A., Dimo, T., et al. (2008). Anti-hypertensive effects of the methanol/methylene chloride stem bark extract of Mammea africana in L-NAME-induced hypertensive rats. *J. Ethnopharmacol.* 117 (3), 446–450. doi:10.1016/j.jep.2008.02.028

Ojala, T., Vuorela, P., Kiviranta, J., Vuorela, H., and Hiltunen, R. (1999). A bioassay using *Artemia salina* for detecting phototoxicity of plant coumarins. *Planta Medica* 65 (08), 715–718. doi:10.1055/s-1999-14049

Ojewole, J. A., and Adewunmi, C. O. (2004). Anti-inflammatory and hypoglycaemic effects of *Tetrapleura tetraptera* (Taub)[fabaceae] fruit aqueous extract in rats. *J. Ethnopharmacol.* 95 (2-3), 177–182. doi:10.1016/j.jep.2004.06.026

Oldenburg, J., Watzka, M., Rost, S., and Müller, C. R. (2007). VKORC1: molecular target of coumarins. *J. Thrombosis Haemostasis* 5, 1–6. doi:10.1111/j.1538-7836.2007. 02549.x

Panno, M., Giordano, F., Palma, M., Bartella, V., Rago, V., Maggiolini, M., et al. (2009). Evidence that bergapten, independently of its photoactivation, enhances p53 gene expression and induces apoptosis in human breast cancer cells. *Curr. cancer drug targets* 9 (4), 469–481. doi:10.2174/156800909788486786

Perone, V. B. (2016). in *The natural occurrence and uses of toxic coumarins. Microbial Toxins*. Editors S. Kadis, A. Ciegler, and S. J. Ajl, 67–88.

Poumale, H. M. P., Hamm, R., Zang, Y., Shiono, Y., and Kuete, V. (2013). "Coumarins and related compounds from the medicinal plants of Africa," in *Medicinal plant research in Africa* (Elsevier), 261–300.

Prusty, J. S., and Kumar, A. (2020). Coumarins: antifungal effectiveness and future therapeutic scope. *Mol. Divers.* 24 (4), 1367–1383. doi:10.1007/s11030-019-09992-x

Qu, L., Li, J., Wang, R., Wang, X., Zhao, T., Chen, Y., et al. (2021). Metabolic and transcriptional profiling of *Fraxinus chinensis* var. *rhynchophylla* unravels possible constitutive resistance against *Agrilus planipennis*. *Forests* 12 (10), 1373. doi:10.3390/f12101373

Raja, S. B., Murali, M. R., Roopa, K., and Devaraj, S. N. (2011). Imperatorin a furocoumarin inhibits periplasmic Cu-Zn SOD of *Shigella dysenteriae* their by modulates its resistance towards phagocytosis during host pathogen interaction. *Biomed. Pharmacother.* 65 (8), 560–568. doi:10.1016/j.biopha.2010.10.010

Reddy, D. S., Kongot, M., and Kumar, A. (2021). Coumarin hybrid derivatives as promising leads to treat tuberculosis: recent developments and critical aspects of structural design to exhibit anti-tubercular activity. *Tuberculosis* 127, 102050. doi:10. 1016/j.tube.2020.102050

Riveiro, M. E., De Kimpe, N., Moglioni, A., Vazquez, R., Monczor, F., Shayo, C., et al. (2010). Coumarins: old compounds with novel promising therapeutic perspectives. *Curr. Med. Chem.* 17 (13), 1325–1338. doi:10.2174/092986710790936284

Sai, Y., Yang, T. J., Krausz, K. W., Gonzalez, F. J., and Gelboin, H. V. (1999). An inhibitory monoclonal antibody to human cytochrome P450 2A6 defines its role in the metabolism of coumarin, 7-ethoxycoumarin and 4-nitroanisole in human liver. *Pharmacogenetics Genomics* 9 (2), 229–237.

Sancho, R. o., Márquez, N., Gómez-Gonzalo, M., Calzado, M. A., Bettoni, G., Coiras, M. T., et al. (2004). Imperatorin inhibits HIV-1 replication through an Sp1-dependent pathway. *J. Biol. Chem.* 279 (36), 37349–37359. doi:10.1074/jbc. M401993200

Santoro, M., Guido, C., De Amicis, F., Sisci, D., Cione, E., Dolce, V., et al. (2016). Bergapten induces metabolic reprogramming in breast cancer cells. *Oncol. Rep.* 35 (1), 568–576. doi:10.3892/or.2015.4327

Sarker, S. D., and Nahar, L. (2017). Progress in the chemistry of naturally occurring coumarins. *Prog. Chem. Nat. Occur. coumarins* 106, 241–304. doi:10.1007/978-3-319-59542-9_3

Shokoohinia, Y., Bazargan, S., Miraghaee, S., Javadirad, E., Farahani, F., and Hosseinzadeh, L. (2017). Safety assessment of osthole isolated from Prangos ferulacea: acute and subchronic toxicities and modulation of cytochrome P450. *Jundishapur J. Nat. Pharm. Prod.* 12 (3). doi:10.5812/jjnpp.63764

Skalicka-Woźniak, K., Orhan, I. E., Cordell, G. A., Nabavi, S. M., and Budzyńska, B. (2016). Implication of coumarins towards central nervous system disorders. *Pharmacol. Res.* 103, 188–203. doi:10.1016/j.phrs.2015.11.023

Stefanachi, A., Leonetti, F., Pisani, L., Catto, M., and Carotti, A. (2018). Coumarin: a natural, privileged and versatile scaffold for bioactive compounds. *Molecules* 23 (2), 250. doi:10.3390/molecules23020250

Sun, C., Zhao, W., Wang, X., Sun, Y., and Chen, X. (2020). A pharmacological review of dicoumarol: an old natural anticoagulant agent. *Pharmacol. Res.* 160, 105193. doi:10. 1016/j.phrs.2020.105193

Teng, C.-M., Lin, C.-H., Ko, F.-N., Wu, T.-S., and Huang, T.-F. (1994). The relaxant action of osthole isolated from Angelica pubescens in Guinea-pig trachea. *Naunyn-Schmiedeberg's archives Pharmacol.* 349, 202–208. doi:10.1007/BF00169838

Tubaro, A., Del Negro, P., Ragazzi, E., Zampiron, S., and Della Loggia, R. (1988). Antiinflammatory and peripheral analgesic activity of esculetin *in vivo. Pharmacol. Res. Commun.* 20, 83–85. doi:10.1016/S0031-6989(88)80847-6

Ulrich, M., Lüer, P., and Kreusch, D. (1999). "1.24 - biosynthesis of coumarins." In: *Comprehensive natural products chemistry meth-cohn*,Eds: edited by B. Sir Derek, N. Koji, and B. T. Otto - 623–637. Oxford, Pergamon: Elsevier Science. doi:10.1016/ B978-0-08-091283-7.00026-6

Vaou, N., Stavropoulou, E., Voidarou, C., Tsigalou, C., and Bezirtzoglou, E. (2021). Towards advances in medicinal plant antimicrobial activity: A review study on challenges and future perspectives. *Microorganisms* 9 (10), 2041. doi:10.3390/ microorganisms9102041

Venugopala, K. N., Rashmi, V., and Odhav, B. (2013). Review on natural coumarin lead compounds for their pharmacological activity. *BioMed Res. Int.* 2013, 963248. doi:10.1155/2013/963248

Wagstaff, D. J. (1991). Dietary exposure to furocoumarins. Regul. Toxicol. Pharmacol. 14 (3), 261-272. doi:10.1016/0273-2300(91)90029-u

Wang, C., Pei, A., Chen, J., Yu, H., Sun, M.-L., Liu, C.-F., et al. (2012). A natural coumarin derivative esculetin offers neuroprotection on cerebral ischemia/

reperfusion injury in mice. J. Neurochem. 121 (6), 1007-1013. doi:10.1111/j.1471-4159.2012.07744.x

Wang, Y., Zhang, H., Jiang, J. M., Zheng, D., Chen, Y. Y., Wan, S. J., et al. (2019). Hepatotoxicity induced by psoralen and isopsoralen from fructus psoraleae: wistar rats are more vulnerable than ICR mice. *Food Chem. Toxicol.* 125, 133–140. doi:10.1016/J. FCT.2018.12.047

Wang, Y.-H., Avula, B., Nanayakkara, N. D., Zhao, J., and Khan, I. A. (2013). Cassia cinnamon as a source of coumarin in cinnamon-flavored food and food supplements in the United States. *J. Agric. food Chem.* 61 (18), 4470–4476. doi:10. 1021/jf4005862

Who, (1995). Coumarin: a strong association with hepatotoxicity. WHO Drug Inf. 9, 159.

Wishart, D. S., Feunang, Y. D., Guo, A. C., Lo, E. J., Marcu, A., Grant, J. R., et al. (2018). DrugBank 5.0: a major update to the DrugBank database for 2018. *Nucleic acids Res.* 46 (D1), D1074–D1082. doi:10.1093/nar/gkx1037

Wu, L., Wang, X., Xu, W., Farzaneh, F., and Xu, R. (2009). The structure and pharmacological functions of coumarins and their derivatives. *Curr. Med. Chem.* 16 (32), 4236–4260. doi:10.2174/092986709789578187

Wu, T. S., Hsu, M. Y., Kuo, P. C., Sreenivasulu, B., Damu, A. G., Su, C. R., et al. (2003). Constituents from the leaves of *Phellodendron amurense* var. wilsonii and their bioactivity. *J. Nat. Prod.* 66 (9), 1207–1211. doi:10.1021/np030034v

Yang, D., Gu, T., Wang, T., Tang, Q., and Ma, C. (2010). Effects of osthole on migration and invasion in breast cancer cells. *Biosci. Biotechnol. Biochem.* 74 (7), 1430–1434. doi:10.1271/bbb.100110

Yarnell, E., and Abascal, K. (2009). Plant coumarins: myths and realities. Altern. Complementary Ther. 15 (1), 24-30. doi:10.1089/act.2009.15104

Zhang, J., Di, H., Luo, K., Jahufer, Z., Wu, F., Duan, Z., et al. (2018). Coumarin content, morphological variation, and molecular phylogenetics of Melilotus. *Mol.* 23 (4), 810. doi:10.3390/molecules23040810

Zhang, J., Tan, Y., Li, G., Chen, L., Nie, M., Wang, Z., et al. (2021). Coumarin sulfonamides and amides derivatives: design, synthesis, and antitumor activity *in vitro*. *Molecules* 26 (4), 786. doi:10.3390/molecules26040786

Zhang, Z.-R., Leung, W. N., Cheung, H. Y., and Chan, C. W. (2015). Osthole: a review on its bioactivities, pharmacological properties, and potential as alternative medicine. *Evidence-Based Complementary Altern. Med.* 2015, 919616. doi:10.1155/2015/919616

Zhao, H., Chen, X., Chen, X., Zhu, Y., Kong, Y., Zhang, S., et al. (2020). New peptidendrocins and anticancer chartreusin from an endophytic bacterium of *Dendrobium officinale. Ann. Transl. Med.* 8 (7), 455. doi:10.21037/atm.2020.03.227

Zhu, J. J., and Jiang, J. G. (2018). Pharmacological and nutritional effects of natural coumarins and their structure-activity relationships. *Mol. Nutr. food Res.* 62 (14), 1701073. doi:10.1002/mnfr.201701073