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Ranu Surolia, University of Alabama at Birmingham, United States

\*CORRESPONDENCE Fei Wang, a wangfei896@163.com Quanyu Du, a quanydu@cdutcm.edu.cn Han Yang, 15882458490@163.com

<sup>†</sup>These authors share first authorship

RECEIVED 03 December 2023 ACCEPTED 31 January 2024 PUBLISHED 13 February 2024

#### CITATION

Wu X, Xiao X, Fang H, He C, Wang H, Wang M, Lan P, Wang F, Du Q and Yang H (2024), Elucidating shared biomarkers in gastroesophageal reflux disease and idiopathic pulmonary fibrosis: insights into novel therapeutic targets and the role of angelicae sinensis radix. *Front. Pharmacol.* 15:1348708. doi: 10.3389/fphar.2024.1348708

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# Elucidating shared biomarkers in gastroesophageal reflux disease and idiopathic pulmonary fibrosis: insights into novel therapeutic targets and the role of angelicae sinensis radix

Xuanyu Wu<sup>1†</sup>, Xiang Xiao<sup>1†</sup>, Hanyu Fang<sup>2,3</sup>, Cuifang He<sup>1</sup>, Hanyue Wang<sup>1</sup>, Miao Wang<sup>1</sup>, Peishu Lan<sup>1</sup>, Fei Wang<sup>1\*</sup>, Quanyu Du<sup>1\*</sup> and Han Yang<sup>1\*</sup>

<sup>1</sup>Hospital of Chengdu University of Traditional Chinese Medicine, School of Clinical Medicine, Chengdu University of Traditional Chinese Medicine, Chengdu, China, <sup>2</sup>Graduate School, Beijing University of Chinese Medicine, Beijing, China, <sup>3</sup>Department of Traditional Chinese Medicine for Pulmonary Diseases, Center of Respiratory Medicine, China-Japan Friendship Hospital, Beijing, China

**Background:** The etiological underpinnings of gastroesophageal reflux disease (GERD) and idiopathic pulmonary fibrosis (IPF) remain elusive, coupled with a scarcity of effective therapeutic interventions for IPF. Angelicae sinensis radix (ASR, also named Danggui) is a Chinese herb with potential anti-fibrotic properties, that holds promise as a therapeutic agent for IPF.

**Objective:** This study seeks to elucidate the causal interplay and potential mechanisms underlying the coexistence of GERD and IPF. Furthermore, it aims to investigate the regulatory effect of ASR on this complex relationship.

**Methods:** A two-sample Mendelian randomization (TSMR) approach was employed to delineate the causal connection between gastroesophageal reflux disease and IPF, with Phennoscanner V2 employed to mitigate confounding factors. Utilizing single nucleotide polymorphism (SNPs) and publicly available microarray data, we analyzed potential targets and mechanisms related to IPF in GERD. Network pharmacology and molecular docking were employed to explore the targets and efficacy of ASR in treating GERD-related IPF. External datasets were subsequently utilized to identify potential diagnostic biomarkers for GERD-related IPF.

**Results:** The IVW analysis demonstrated a positive causal relationship between GERD and IPF (IVW: OR = 1.002, 95%CI: 1.001, 1.003; p < 0.001). Twenty-five shared differentially expressed genes (DEGs) were identified. GO functional analysis revealed enrichment in neural, cellular, and brain development processes, concentrated in chromosomes and plasma membranes, with protein binding and activation involvement. KEGG analysis unveiled enrichment in proteoglycan, ERBB, and neuroactive ligand-receptor interaction pathways in cancer. Protein-protein interaction (PPI) analysis identified seven hub genes. Network pharmacology analysis demonstrated that 104 components of ASR targeted five hub genes (PDE4B, DRD2, ERBB4, ESR1, GRM8), with molecular docking confirming their excellent binding

efficiency. GRM8 and ESR1 emerged as potential diagnostic biomarkers for GERD-related IPF (ESR1: AUC<sub>GERD</sub> = 0.762, AUC<sub>IPF</sub> = 0.725; GRM8: AUC<sub>GERD</sub> = 0.717, AUC<sub>IPF</sub> = 0.908). GRM8 and ESR1 emerged as potential diagnostic biomarkers for GERD-related IPF, validated in external datasets.

**Conclusion:** This study establishes a causal link between GERD and IPF, identifying five key targets and two potential diagnostic biomarkers for GERD-related IPF. ASR exhibits intervention efficacy and favorable binding characteristics, positioning it as a promising candidate for treating GERD-related IPF. The potential regulatory mechanisms may involve cell responses to fibroblast growth factor stimulation and steroidal hormone-mediated signaling pathways.

KEYWORDS

idiopathic pulmonary fibrosis, gastroesophageal reflux disease, angelicae sinensis radix, mendelian randomization, network-pharmacology

### **1** Introduction

Pulmonary fibrosis (PF) represents a progressive lung ailment characterized by compromised alveolar tissue repair following injury, attributed to diverse internal and external stimuli. Its clinical manifestations encompass a persistent dry cough and progressively worsening dyspnea (Mathai and Schwartz, 2019). Idiopathic pulmonary fibrosis (IPF) is the most common type of PF, with a median survival time of 2-4 years (Ogawa et al., 2021). Given its intricate pathogenesis, irreversible disease trajectory, heightened mortality rates, and unfavorable prognosis, IPF assumes characteristics akin to a neoplastic disorder (Aggarwal et al., 2016; Ma et al., 2020). Current therapeutic strategies for IPF remain nascent, while clinically approved agents like Pirfenidone and Nintedanib offer a mitigating effect on lung function decline but fall short in halting disease progression, coupled with a plethora of adverse reactions (Ma et al., 2022). While lung transplantation is the sole curative recourse for IPF (George et al., 2019), its widespread application is curtailed by posttransplant rejection and a scarcity of donor resources.

Observational studies have unveiled gastroesophageal reflux disease (GERD) as an independent risk factor for IPF, with 90% of IPF patients exhibiting GERD and a twofold increase in the risk of GERD among IPF patients (Khor et al., 2022; Raghu et al., 2006; Baqir et al., 2021). Gastric contents, including bile and pepsin, have been identified in bronchoalveolar lavage fluid (BALF) of IPF patients (Davis et al., 2013; Lee et al., 2012; Lozo et al., 2014). Intriguingly, the infusion of gastric juice into the airway induces lung injury in animal models, evoking a local inflammatory response, abnormal myofibroblast activation, excessive deposition of extracellular matrix (ECM), and formation of PF (Davis et al., 2013; Hoppo et al., 2014). Accordingly, aggressive treatment of GERD will help prevent and improve the prognosis of IPF. However, current GERD intervention modalities remain constrained, with proton pump inhibitors offering symptomatic relief but falling short of achieving a definitive cure.

Traditional Chinese Medicine (TCM), with its millennia-long history, emerges as a pivotal avenue for disease prevention and treatment, offering a framework deeply rooted in TCM theory. Both clinical and preclinical studies underscore the positive impact of herbal medicine on intervening in IPF, contributing to enhanced life energy and improved blood circulation (Zhou et al., 2024). Angelicae Sinensis Radix (ASR), also named Danggui, a Chinese herbal medicine renowned for its blood-circulating and stasisremoving properties, has been demonstrated through modern pharmacology to mitigate inflammatory cell infiltration, reduce oxidative stress, restore the alveolar-capillary barrier, and ameliorate chronic pathological lung changes (Wei et al., 2016; Zhang et al., 2022; Guo et al., 2024). In addition, ASR has antifibrotic effects via regulation of inflammatory damage, epithelial-mesenchymal transition (EMT), ECM, and other pathways (Han et al., 2006; Gao et al., 2012; Qian et al., 2020; Zhang et al., 2020).

Despite advancements in utilizing Mendelian randomization (MR) to investigate the causal relationship between GERD and IPF (Reynolds et al., 2023; Cotton et al., 2023), existing studies have limitations, including incomplete confounding factor exclusion and exclusive focus on the causal link. Moreover, a comprehensive exploration of the genetic overlap, susceptibility gene sharing, and deep mechanisms between the two diseases is still lacking. To address these issues, this study explores the causal relationship between GERD and IPF utilizing MR and the possible internal targets and mechanisms of GERD-related IPF using single nucleotide polymorphisms (SNPs) combined with publicly available microarray data. Furthermore, the regulatory impact of ASR on GERD-related IPF is explored through network pharmacology and molecular docking, offering novel insights into ASR's mechanism of action and presenting potential avenues for complementary and replacement anti-PF therapies.

### 2 Methods

# 2.1 Identification of genetic causality between GERD and IPF

### 2.1.1 Study design

MR constitutes the foundation of this study, relying on genetic variants and employing the instrumental variable method. The methodological underpinning involves three core assumptions (Figure 1): Hypothesis 1: The genetic variant exhibits a robust association with the exposure (GERD). Hypothesis 2: The variant



remains independent of any confounding factors. Hypothesis 3: Please check whether my interpretation.

#### 2.1.2 Data sources

In defining GERD as the exposure factor and IPF as the outcome factor, we assessed genome-wide association study (GWAS) summary data of GERD (GCST90000514) (Ong et al., 2022) and IPF (GCST90018120) (Duckworth et al., 2021) from the GWAS Catalog (https://www.ebi.ac.uk/gwas/). The GERD dataset comprised 602,604 samples, including 462,753 GERD patients and 1,127,474 controls, with genotyping information for 2,324,711 SNPs. The IPF dataset included 451,025 samples, with 1369 IPF patients, 435,866 controls, and genotyping details for 16,137,102 SNPs.

#### 2.1.3 Instrumental variables screening

Significant SNPs related to the exposure were selected based on a threshold of  $p < 5 \times 10^{-8}$ . We ensured SNPs independence by setting

the linkage disequilibrium coefficient ( $r^2 = 0.001$ ) and region width (kb = 1,000) to eliminate linkage disequilibrium and prevent bias. IPF is associated with various risk factors, including genetics, autoimmune deficiency, environmental exposure (such as smoking, diabetes, GERD), and viral infection, among others (Reynolds et al., 2023; Zhao et al., 2023). Furthermore, GERD is associated with obesity, smoking, and genetics (Maret-Ouda et al., 2020). Recognizing the influence of confounders on both exposure and outcome (Zhou et al., 2023), we identified smoking and genetics as confounders. Utilizing the PhenoScanner database, we screened SNPs associated with confounders under conditions of  $p = 5 \times 10^{-8}$ , None Proxies,  $r^2 = 0.8$ , and Build = 37. Weak instrumental variables were identified using F = $(N-2) \times \frac{R^2}{(1-R^2)}$ , where  $R^2 = 2 \times (1 - EAF) \times EAF \times (\frac{\beta}{SE \times \sqrt{N}})^2$ , with F < 10 indicating the removal of weak instrumental variables. Palindromic SNPs with intermediate allele frequencies were excluded during the harmonization of exposure and outcome data.

#### 2.1.4 MR analysis

Inverse variance weighted (IVW) served as the primary MR analysis method, supplemented by weighted median (WM), simple mode (SM), MR-Egger regression, and weighted mode (WME) to assess IVW method robustness. Cochrane's Q and leave-one-out (LOO) sensitivity tests evaluated the heterogeneity of individual genetic variation estimates. The IVW random effects model was applied in cases of heterogeneity among SNPs. Outliers were examined using MR-PRESSO, and horizontal pleiotropy was tested using MR-Egger-intercept.

### 2.1.5 Statistical analysis

R (version 4.3.0) software packages "Two-Sample-MR" and "MR-PRESSO" facilitated all data analysis and visualization. Calculated results were expressed as odds ratios (OR) and 95% confidence intervals (95%CI), considering p < 0.05 as considered significant. Sensitivity analysis with p > 0.05 indicated no significant heterogeneity of SNPs, and MR-PRESSO verified the absence of outliers with p > 0.05. MR-Egger-intercept confirmed no horizontal pleiotropy with p > 0.05.

# 2.2 The screening of shared susceptibility gene between GERD and IPF

#### 2.2.1 Microarray data acquisition and processing

SNPs from MR calculations and literature, related to genes, were collated, removing duplicates and SNPs lacking corresponding gene names (Reynolds et al., 2023). Using "Gastroesophageal reflux disease" and "idiopathic pulmonary fibrosis" as search terms in the GEO database (https://www.ncbi.nlm.nih.gov/geo/), DisGeNET database (https://www.disgenet.org), and GeneCards database (www.genecards.org), respectively. Finally, GSE65013 (GERD) and GSE10667 (IPF) were selected for obtaining differentially expressed genes (DEGs) related to IPF-related GERD. Targets from DisGeNET and GeneCards were restricted to "*Homo sapiens*" and filtered based on the median score.

### 2.2.2 Screening of shared DEGs between GERD and IPF

DEGs were analyzed using the limma package in R software, setting a threshold of "Adjusted p < 0.05 and  $\log_2(FC) > 1.5$  or  $\log_2(FC) <-1.5$ ". A heatmap was generated using the R package "pheatmap." Shared genes of GERD-related IPF were identified through a Venn plot summarizing genes from multiple sources, including the GEO database, SNPs, DisGeNET, and GeneCards database.

# 2.2.3 Functional analysis and protein-protein interaction (PPI) networks of shared DEGs

Metascape databases facilitated gene ontology (gene ontology, GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analysis using shared DEGs (https://metascape.org/gp/index.html). Sangerbox (http:// sangerbox.com/tool.html) was used for data visualization. The STRING database (https://cn.string-db.org/) was used to conduct a PPI network analysis, with a median confidence of 0.4. Cytoscape was subsequently used to visualize the results (Shannon et al., 2003). The Cytohubba plugin utilized six algorithms (MNC, EPC, DMNC, EcCentricity, Betweenness, and Degree) to identify top targets using six algorithms. A Venn plot aided in selecting potential targets.

# 2.3 Network pharmacology analysis of ASR intervention in GERD-related IPF

# 2.3.1 Drug compound-target acquisition and screening

Active compounds of ASR were collected from the TCMSP database and literature, with chemical structures obtained from PubMed. Those not found in SDF format were transformed and drawn into the corresponding SMILES structures. Since the traditional use of ASR is an oral decoction, the screening parameters included oral availability and drug likeliness. Swiss target provided corresponding targets, and PPI analysis mapped the compound-target phase. The compound-target format was imported into Cytoscape for network construction, and the Degree algorithm sorted and visualized the network.

# 2.3.2 Molecular docking of active components of ASR with hub genes

Molecular docking is widely used in drug screening and often used to predict the relationship between disease targets and drug structural ligands. The top three compounds with the highest degree value for each target were selected to screen compounds for molecular docking. Only the existing compounds were used for docking if the corresponding compounds were less than three. The 3D structures of small molecule compounds were downloaded from the PubChem database and stored in mol2 format. The 3D structures in the hub genes with a resolution lower than 2.5Å were downloaded from the PDB (https://www.rcsb.org/) database and saved in the PDB format. CB - Dock2 (https://cadd.labshare.cn/cb-dock2/), a tool that improves blind docking methods by integrating cavity detection, docking, and homology simulation fitting, was used to explore effective receptor-ligand binding sites (Liu et al., 2022a). The vina scores were finally used to calculate the binding energies between receptor-ligand molecules, with lower binding energies indicating stronger interactions. Binding energies < -5 kCal/mol represent favorable binding activity.

# 2.4 Identification and validation of diagnostic biomarkers for GERD-related IPF

Utilizing the pROC package in R, we generated ROC curves for key targets associated with GERD and IPF in GSE10667 and GSE39491 datasets. The diagnostic efficiency of each target for the two diseases was assessed by calculating the area under the curve (AUC). AUC values exceeding 0.7 indicated the identification of targets with good diagnostic efficiency. Subsequent screening and adjustment, considering the F value, removed six weak instrumental variables (rs1011407, rs11645288, rs2232423, rs3172494, rs3828917, rs569356). The

Exposure-outcome	Method	N (SNP)	Р	OR	95%CI	Cochran's Q test	MR-Egger- intercept	MR- PRESSO
GERD-IPF	MR Egger	76	0.850	1.001	0.994-1.007	0.163	0.643	0.207
	WM	76	0.001	1.002	1.001-1.003			
	IVW	76	< 0.001	1.002	1.001-1.003			
	SM	76	0.446	1.001	0.998-1.005			
	WME	76	0.372	1.002	0.998-1.005			

#### TABLE 1 Causal effect of GERD on IPF.



GeneMANIA (http://www.genemania.org) database was then employed to predict potential interacting genes and pathways associated with the identified diagnostic biomarkers, offering preliminary insights into underlying mechanisms.

### **3** Results

# 3.1 MR analysis of the causal relationship between GERD and IPF

#### 3.1.1 Instrumental variables selection

After removing the linkage disequilibrium, 90 SNPs remained. PhenoScanner eliminated four SNPs related to tobacco intake (rs6711584, rs1510719, rs10242223, rs215614) and one SNPs related to gene expression (rs13107325). Subsequently, six weak instrumental variables (rs1011407, rs11645288, rs2232423, rs3172494, rs3828917, rs569356) were removed after screening and adjustment by calculating the *F* value. The harmonization process excluded three SNPs with intermediate allele frequencies, resulting in 76 instrumental variables.

### 3.1.2 Positive causal effect of GERD on IPF

The MR analysis, employing the 76 instrumental variables (Table 1), demonstrated a positive causal relationship between GERD and IPF (IVW: OR = 1.002, 95% CI: 1.001, 1.003; p < 0.001; Figure 2A). Moreover, the scatterplot revealed that the results of WM, SM, MR-Egger regression, and weighted plurality method were consistent with IVW (Figure 2B), suggesting good robustness of the results. Cochrane's Q test yielded p = 0.16, indicating no heterogeneity of SNPs within the study. The funnel plot showed that the distribution of causal effects was symmetrical, indicating no obvious bias (Figure 2C). The MR-Egger-intercept test yielded a *p*-value of 0.643, indicating the absence of pleiotropy, suggesting that the instrumental variables did not significantly affect the outcome through pathways other than exposure. The forest plot demonstrated that the MR analysis results were consistent upon excluding individual SNPs, as indicated by the



LOO analysis (Figure 2D). MR-PRESSO test p = 0.207, no outlier SNPs were detected.

# 3.2 Shared susceptibility genes between GERD and IPF

# 3.2.1 Common targets and shared DEGs between GERD and IPF

Combining the 76 SNPs from MR analysis with those from the literature yielded 86 SNPs, and 104 corresponding genes were

obtained from the GWAS catalog (Supplementary Table S1). The Limma package identified 522 DEGs for GERD (215 upregulated and 307 downregulated, Figures 3A, B), and 3,781 DEGs for IPF (734 upregulated and 3,047 downregulated, Figures 3C, D). From DisGeNET and GeneCards databases, 1532 GERD-related genes and 5994 IPF-related genes were obtained, respectively. The Venn plot showed 25 shared DEGs for GERD-related IPF (Figure 3E).

### 3.2.2 Functional enrichment analysis

GO functional analysis (Figure 4A) was employed to evaluate the enrichment of biological process (BP), cellular component



(CC), and molecular function (MF), respectively. The study found that the BP terms were mainly enriched in neurogenesis, cell development, telencephalon, and forebrain development, whereas CC terms were mainly enriched in the plasma membrane, chromosomal part, chromosome, and integral component of the plasma membrane. MF terms were mainly enriched in protein binding, dimerization activity, signaling receptor activity, molecular transducer activity, and protein homodimerization activity. Based on the KEGG analysis, the potential targets were enriched in pathways, including proteoglycans in cancer, erythroblastic leukemia viral oncogene homolog, neuroactive ligand-receptor interactions, and calcium signaling pathway. These findings offer avenues for deeper exploration (Figure 4B).

### 3.2.3 PPI network analysis

The STRING database facilitated PPI network analysis, and Cytoscape aided network visualization (Figure 4C). Top targets were identified using six algorithms, including MNC, EPC, DMNC, EcCentricity, Betweenness, and Degree, a Venn plot was drawn to visualize the intersection (Figure 4D), which contained seven hub genes, namely, Forkhead box p2 (FOXP2), Erb-B2 Receptor Tyrosine Kinase 4 (ERBB4), Glutamate Metabotropic Receptor 8 (GRM8), Neural Cell Adhesion Molecule 1 (NCAM1), Estrogen Receptor 1 (ESR1), Dopamine Receptor D2 (DRD2), Phosphodiesterase 4B (PDE4B).

# 3.3 Results of network pharmacology analysis of ASR intervention in GERD-related IPF

# 3.3.1 Compounds-target acquisition and screening of ASR

Utilizing TCMSP combined with a literature search (Feng et al., 2022), we amassed 360 active compounds of ASR. The small molecule structures of these compounds were retrieved from PubChem and subjected to oral availability and drug-likeness screening using SwissADME. Targets that could not be predicted in Swiss target were removed, resulting in a network of 109 nodes and 145 edges. Network pharmacology revealed that 104 ASR compounds had mapping relationships with five key targets (PDE4B, DRD2, ERBB4, ESR1, and GRM8) (Figure 5, Supplementary Table S2).



# 3.3.2 Molecular docking verification of active compounds of ASR and hub genes of GERD-related IPF

The top three compounds corresponding to the five key targets were selected for molecular docking to assess the binding efficacy of ASR's active compounds with disease targets. As there was only one corresponding active compound for ERBB4 and two for GRM8, molecular docking focused on these specific compounds. The results (Table 2) showed that the binding energies ranged from -4.5 to -8.2 kCal/mol, visually confirming excellent binding activity between ASR and the five key targets (Figure 6).

# 3.4 Screening of key targets and exploring the mechanism of GERD-related IPF

ROC curves for the five key targets were generated using the R package pROC. Diagnostic efficacy in GERD and IPF was evaluated through AUC analysis (Figures 7A,D). AUC values were as follows: ESR1 (AUC<sub>GERD</sub> = 0.762, AUC<sub>IPF</sub> = 0.725), ERBB4 (AUC<sub>GERD</sub> = 0.663, AUC<sub>IPF</sub> = 0.567), DRD2 (AUC<sub>GERD</sub> = 0.723, AUC<sub>IPF</sub> = 0.550), PDE4B (AUC<sub>GERD</sub> = 0.743, AUC<sub>IPF</sub> = 0.492), GRM8 (AUC<sub>GERD</sub> = 0.717, AUC<sub>IPF</sub> = 0.908). Of these, ESR1 and GRM8 exhibited good diagnostic efficacy for GERD and IPF. External datasets validation confirmed significant downregulation of GRM8 and ESR1 in GERD patients (Figures 7B, C), and in IPF patients, GRM8 was significantly downregulated (p = 0.0005), and ESR1 was significantly upregulated (p = 0.0397) (Figures 7E, F). ASR intervention influenced the expression of these genes, as indicated by GeneMANIA predictions (Figure 7G), suggesting potential links to RNApolymerase activity, maintenance of cell number, cellular response to fibroblast growth factor stimulus, response to fibroblast growth factor, and steroid hormone-mediated signaling pathways. RNA-polymerase activity, maintenance of cell number, cellular response to fibroblast growth factor stimulus, response to fibroblast growth factor, and steroid hormone-mediated signaling pathways.

### 4 Discussion

IPF, the prevailing manifestation of interstitial lung diseases, is burdened with a gloomy prognosis, boasting a median survival of 2–4 years after diagnosis. Clinical heterogeneity and a lack of symptom specificity often contribute to misdiagnosis or diagnostic delays (Lamas et al., 2011). In its advanced stages, IPF may progress to right heart failure coupled with respiratory failure, culminating in fatality (Sgalla et al., 2016). Timely identification and

TABLE 2 Molecular docking results of five hub genes and ASR's compounds.

Targets	PDB ID	Compounds	Cavity volume (Å <sup>3</sup> )	Affinity (kCal/mol)	Docking size			Center		
					Х	Y	Z	Х	Y	Z
ESR1	8APS	Vanillin	915	-5	17	24	25	-20	15	-10
		Senkyunolide B	915	5.9	19	19	25	-20	15	-10
		2-TRIDECANOL	915	4.5	25	25	25	-20	15	-10
DRD2	7DFP	Vanillin	1,338	5.4	17	17	26	-93	-22	214
	Senkyunolide B	1,338	7.8	19	19	26	-93	-22	214	
		2-TRIDECANOL	1,338	5.9	25	25	25	-93	-22	214
PDE4B	4MYQ	Vanillin	3,802	-6	25	27	24	-16	38	-8
		Senkyunolide B	3,802	8.2	25	27	19	-16	38	-8
		o-cresol	3,802	5.8	25	27	24	-16	38	-8
GRM8	6BSZ	(±)-Camphor	1974	-6	16	24	27	-65	-21	34
		2-Methylbutyric acid	4,898	4.9	34	28	32	-44	-32	-23
ERBB4	7PCD	Isoimperatorin	3,541	7.9	26	28	34	4	-2	18



Methylbutvric acid: (L) Docking result between ERBB4 and Isoimperatorin.

diagnosis are imperative. The pathogenesis of IPF involves unknown factors inducing repeated injury and abnormal inflammation of lung tissue, which induces crosstalk of epithelial cells, ECM, and adjacent interstitial tissue. Significant risk factors for this process include virus infection, smoke exposure, and chronic aspiration of gastric contents (Hewlett et al., 2018). A prevalent digestive tract ailment, exhibits a global incidence of 2.5%-51.2%, with a growing trend (Eusebi et al., 2018). Epidemiology has proved that GERD is an independent risk factor for IPF, which may affect more than 80% of IPF patients and promote the progression of fibrosis (Oldham and Collard, 2017). Animal studies have also confirmed that decreased gastric acid pH can aggravate lung injury (King and Nathan, 2013). Despite increased awareness of the GERD-IPF association, the molecular interplay between these conditions remains enigmatic. This study endeavors to elucidate the causal relationship between GERD and IPF, shedding light on regulatory molecules in GERDrelated IPF.

At present, MR studies have substantiated a positive causal link between GERD and IPF (Reynolds et al., 2023; Cotton et al., 2023). However, one study, while utilizing PhenoScanner for confounders comparison, failed to eliminate relevant confounders (Reynolds et al., 2023). Another multivariate MR study established a connection between high BMI and the GERD-IPF causal relationship, which waned after adjusting BMI (Cotton et al., 2023). Therefore, this study fills a gap by conducting a twosample MR analysis to demonstrate the genetic perspective of the causal relationship between GERD and IPF. Literature retrieval identified potential confounders, and PhenoScanner aided their removal. Calculation of the F-value guided the exclusion of five weak instrumental variables. IVW results (IVW: OR = 1.002, 95%CI: 1.001, 1.003) affirmed the GERD and IPF causal relationship, corroborated by the WM algorithm. Rigorous sensitivity analyses using Cochrane's Q test and LOO underscored the reliability of the findings. MR-Egger intercept and MR-PRESSO tests revealed no horizontal pleiotropy or outliers, bolstering the article's reliability.

Current studies suggest that GERD-related IPF may be due to abnormal anti-reflux barrier structure/function and local abnormal inflammation caused by the original gastric contents (gastric juice, bile, pepsin, gastric acid, chylous particles, et al.) reflux into the respiratory tract and lung (Warren et al., 2017). The BALF of patients with IPF contains gastric contents such as juice, bile, and pepsin, with esophageal sphincter relaxation due to decreased pressure (Davis et al., 2013; Lee et al., 2012; Lozo et al., 2014; Gavini et al., 2015; Soares et al., 2011). In acute IPF exacerbations, BALF has demonstrated gastrointestinal bacteria transplantation (Molyneaux et al., 2017). Continuous instillation of Hydrochloric acid (HCL) into the airway of mice can simulate gastric acid reflux and cause acute and chronic lung injury. In addition, it can also lead to the upregulation of fibrosis markers such as transforming growth factor  $\beta$  (TGF- $\beta$ ), hydroxyproline, and ECM-related proteins like elastin, tenascin C, and fibronectin. The severity of this abnormal state depends on the concentration of HCL, gender, and age of the mice (Marinova et al., 2019; Colunga Biancatelli et al., 2021; Solopov et al., 2021). It is possible that



activation of shared signal pathways could be a mechanism for the conditions. Using duodenal ligation to simulate occult reflux and microaspiration, it can be found that the GERD combined with BLM-induced PF animal model has the same increased expression

of extracellular-regulated kinases (ERK) and TGF- $\beta$ 1/Smad compared with the classical BLM animal model (Li, M., et al., 2021). The ERK1/2 signaling pathway is one of the potential shared pathways. The expression of ERK1/2 in the distal

gastroesophageal squamous mucosa of patients with GERD is abnormal. The abnormal activation of ERK1/2 in IPF can regulate cell proliferation and fibroblast differentiation by upregulating a-SMA and promoting the proliferation of type 1 collagen (Weng et al., 2019). The TGF-β/Smad pathway is another potential shared pathway. TGF- $\beta$  is the most important cytokine in the development of IPF. Inhibiting the binding of TGF-B and Smad proteins has become a key strategy of anti-PF in recent years (Xue et al., 2020). Long-term exposure to GERD can cause the upregulation of TGF-β1, Smad-3, α-SMA, and type I collagen in the esophageal epithelium of mice, promoting the expression levels of ROS, NADPH oxidase-4, and malondialdehyde (Yisireyili et al., 2019). The precise mechanism of GERD-related IPF remains elusive. Since IPF lacks a cure, a profound understanding of its pathogenesis and identification of effective intervention targets is paramount. This study explores the mechanism between GERD and IPF, incorporating conventional databases (GEO, DisGeNET, and GeneCards) and SNPs from MR analysis, providing a genetic perspective on disease localization and intervention targets (Zhang et al., 2020; Zhang et al., 2023).

GO functional analysis revealed that disease targets of GERDrelated IPF are mainly enriched in neural, cellular, and brain developmental processes, with chromosomal and plasma membrane fractions associated with protein binding and activation. This concurrence aligns seamlessly with our original genetic perspective, providing a comprehensive understanding of the intricate mechanisms underpinning these two diseases. The embryonic development mechanism proves that the digestive and respiratory systems have the same germ layer origin. Except for the nasal epithelium, the epithelium of the respiratory system is differentiated from the original endoderm of the digestive tract. Fibroblast growth factors (FGFs), pivotal in IPF, wield substantial influence during embryonic germ layer development. FGFs facilitate critical crosstalk connections among vital organs like the lung, brain, and intestine, orchestrating their growth spatially and temporally (El et al., 2017). FGFs also play an essential role in regulating lung homeostasis. For example, the FGFR2b ligand can inhibit lung fibrosis and promote tissue repair and regeneration (Dorry et al., 2020). GERD has been shown to increase the expression of FGFs and promote the differentiation of lung fibroblasts, mediate the upregulation of α-SMA, and activate RhoA/ROCK signaling to participate in IPF(Huang et al., 2010; Chiang et al., 2020). Additionally, GERD, arising from neurotrophin expression disorders and inflammation in the esophageal smooth muscle, triggers neuronal abnormalities and impaired esophageal motility (Chen et al., 2015). The relevance of abnormal neurotrophin signaling extends to IPF pathogenesis, where brain-derived neurotrophic factor contributes to the Twsit/snail signaling pathway, leading to abnormal EMT in IPF(Cherubini et al., 2017). Furthermore, both IPF and GERD correlate with the onset of nervous system diseases, including cognitive impairment and dementia, exhibiting overlapping pathway mechanisms (Ong et al., 2022; Adewuyi et al., 2022; Tudorache et al., 2019; Willander et al., 2011).

KEGG analysis showed that the disease targets of GERD-related IPF were mainly enriched in cancer's proteoglycan, ERBB, and neuroactive ligand-receptor interaction pathways. The proteoglycan pathway in cancer is closely associated with abnormal glycosylation metabolism. The advanced glycation end products (AGE)/receptor of advanced glycation end products (RAGE) is an important pathway for abnormal glycosylation metabolism and is associated with aging-related diseases, including IPF (Nedić et al., 2013; Machahua et al., 2016). Longterm reflux in GERD induces local inflammatory responses, and an incomplete ADP-ribosylation-dependent DNA damage response occurs (Han and Zhang, 2022). A sensitive indicator of glycosylation - the lectins (UEA-1, DBA, HPA, and PNA) are also significantly reduced (Neumann et al., 2007). In contrast, the basement membrane of IPF lung epithelial cells highly expresses RAGE, which mediates and accelerates inflammation and fibrosis (Oczypok et al., 2017; Yamaguchi et al., 2017). The ERBB family of transmembrane receptor tyrosine kinases (RTKs) consists of the epidermal growth factor receptors EGFR (ERBB1), HER2 (ERBB2), HER3 (ERBB3) and HER4 (ERBB4). Cohort studies have demonstrated that GERD is a significant risk factor for EGFRmutant lung cancer (Choi et al., 2019), and acid bile salt mimics GERD exposure and enhances ERBB2-mediated activation of immune cell/inflammatory pathways (Patankar et al., 2022). Pathological activation of ERBB/Yes-Associated Protein (YAP) in IPF patients' airway epithelium regulates epithelial-driven mesenchymal transition (Stancil et al., 2021), and EGFR-targeting drugs have a particular effect on alleviating the inflammatory and fibrotic process of IPF (Wang et al., 2022). In addition, as an autonomic nervous organ, the lung contains certain sensory nerve fibers essential in regulating cardiopulmonary function and maintaining human health (Lee and Yu, 2014). Neural activity receptor-ligand facilitates signaling between lung fibroblasts and neurons, and the abnormal expression of its receptor-ligand-related genes, such as FPR1, BDKRB2, MCHR1, NMUR1, CNR2, P2RY14, and PTGER3, is observed in IPF lung fibroblasts (Nguyen et al., 2021). In addition, several studies (Lee et al., 2018) have found that lung fibroblasts can be transformed into neurons under certain conditions to participate in tissue repair. The differentiationstimulating effect of GERD on lung-resident cells (Chiang et al., 2020) may be related to this mechanism.

As a critical TCM herb, ASR boasts many compounds and exhibits multifaceted effects on multiple targets (Hao et al., 2024). Widely utilized in clinical settings for the treatment of PF and GERD, ASR and its compounds have garnered attention in previous studies for their ability to mitigate and reverse IPF. The mechanisms involve reducing oxidative stress and inflammation, inhibiting EMT and myofibroblast activation, and improving lung function and related indicators in PF patients and rat models. Notably, ASR and its compounds have also demonstrated improvements and interventions in the gastroesophageal mucosa in animal subjects. This leads us to a logical inference: Can ASR effectively act upon and intervene in the key targets associated with GERD-related IPF? To answer this question, we embarked on a network pharmacology study, focusing on the seven hub genes (FOXP2, ERBB4, GRM8, NCAM1, ESR1, DRD2, PDE4B) of GERD and IPF.

The outcomes of the network pharmacology analysis unveiled that ASR exerts intervention effects on five hub genes (PDE4B, DRD2, ERBB4, ESR1, and GRM8). Notably, ESR1 and GRM8 exhibit good diagnostic efficacy for both GERD and IPF. PDE4B, a member of the PDE family, plays a crucial role in regulating intracellular Cyclic Adenosine Monophosphate

(cAMP) concentration, engaging in anti-fibrotic signaling. cAMP antagonizes the pro-fibrotic signaling cascade and mediates an increase in the endogenous anti-fibrotic regulator prostaglandin E2 (Chambers, 2022). Targeted inhibition of PDE4B subtype is an important direction in the current drug development for IPF (Henderson et al., 2020; Hermann et al., 2022). BI 1015550, a targeted inhibitor of PDE4B, has been shown in clinical studies to improve patient's forced vital capacity effectively and is expected to be a clinical candidate following Nintedanib and Pirfenidone (Chambers, 2022). PDE4B also affects gastritis-associated gastric cancer, and targeted inhibition of PDE4B can be involved in inhibiting the process of inflammation-associated gastric cancer (Xu et al., 2023). Dopamine receptors regulate gastrointestinal motility and gastric tone, and DRD2 receptor activation inhibits gastric tone and gastrointestinal motility (Bove et al., 2019) and stimulates pepsinogen production (Liu et al., 2022b). Moreover, dopamine receptors are related to regulating immune cell functions such as inflammation. DRD2 can inhibit the transcription of cytokines in macrophages and maximize the activation of YAP, thereby participating in the process of fibrosis (Qing et al., 2022). As one of the ERBB family genes, the role of ERBB4 in fibrosis of various organs has been emphasized (Vermeulen et al., 2017), and it has been considered a potential drug target for treating IPF (Higo et al., 2022). It is also implicated in gastric cancer mutations, apoptosis, and immune functions, although its role in GERD warrants further elucidation (Lucas et al., 2022).

ESR1, responsible for mediating hepatic metabolism, is a pivotal player in GERD, where bile acid, a constituent of gastric content, is integral. Abnormalities in estrogen genes, including ESR1, have been observed in conditions like eosinophilic esophagitis, influencing esophageal barrier function (Wheeler et al., 2019), caused by increased IL-13. Epidemiologic studies have shown (Wang et al., 2009) that men are more likely to develop IPF, with women having a lower prevalence and higher survival rates. An increasing number of studies also provide evidence for the involvement of estrogen in the development of IPF (Shim et al., 2013; Sathish et al., 2015). Estrogen E2 can regulate lung respiratory function and rhythm through the ESR1 receptor. After exogenous induction of bronchial epithelial cells with TGF- $\beta$ 1, ESR1 expression was significantly reduced (Smith et al., 2018). A significant downregulation of ESR1 expression was found in PF models exposed to multi-walled nanotubes (Smith et al., 2019) or silica (Brass et al., 2010). In the subsequent validation with external datasets, ESR1 expression was significantly lower in GERD, while ESR1 expression was significantly higher in IPF compared with the control group. The paradoxical phenomenon was also found in previous in vitro and in vivo studies (Gharaee-Kermani et al., 2005). Therefore, the specific mechanism of ESR1 in IPF, GERD, and GERD-related IPF remains to be further discovered.

GRM8, a metabotropic glutamate receptor, intricately regulates glutamate neurotransmission and has established associations with various central nervous system (CNS) disorders, including major depression, schizophrenia, and autism (Li et al., 2016). Despite a dearth of direct studies delineating GRM8's role in both IPF and GERD, the neuromodulatory functions of GRM8 intersect with both conditions. Lung fibroblasts exhibit reverse transporter glutamate proteins linked to low glutamatergic levels in neuropsychiatric disorders (Huang et al., 2010). The antioxidant N-acetylcysteine, relevant to both PF and CNS neuropsychiatric disorders, demonstrates antifibrotic effects on lung fibroblasts. This aligns with GRM8's position as a downstream player in glutamate metabolism (Raghu et al., 2021). GRM8's involvement in central neurotransmission also extends to addiction-related behaviors where genetic variations impact consumer product intake, a significant GERD risk factor (Nilsson et al., 2004; Gast et al., 2013). The bidirectional regulation of the brain-gut axis, involving glutamate, may contribute to impaired gastrointestinal function and motility, thereby influencing GERD (Saegusa et al., 2011; Filpa et al., 2016). Although current research is limited, a more profound understanding of GRM8's mechanisms in these diseases requires extensive exploration.

To further explore the possible relationship between GRM8 and ESR1 in the intervention of ASR on GERD-related IPF, we used GeneMANIA to predict similar genes and potential mechanisms. Notably, ESR1 exhibited a richer array of potentially similar genes compared to GRM8. The PGR and POLR2 families emerged as closely associated with ESR1. The predicted pathways encompass RNA-polymerase activity, maintenance of cell number, cellular response to fibroblast growth factor stimulus, response to fibroblast growth factor, and steroid hormone-mediated processes.

Progesterone receptor (PGR) and ESR1 are steroid receptors primarily expressed in women, with high prognostic value in various diseases (Berardi et al., 2016; Hertz et al., 2016; Kriegsmann et al., 2020), including fibrotic disorders. PGR and ESR1 work together in FGF-stimulated cell proliferation, differentiation, migration, and angiogenesis, a critical pathway in forming IPF (Ka et al., 2007). Therefore, we hypothesize that the mechanism between GERDrelated IPF may linked to the above signaling pathway. Additionally, the role of steroids in this regard deserves more attention in future studies.

RNA polymerase II (POLR2) transcribes enzymes for protein gene coding and whole genome processes (Schier and Taatjes, 2020). It plays an essential role in both IPF and GERD. The POLR2 plays a crucial role in the abnormal repair of respiratory epithelial cells (Le et al., 2021). It also develops esophageal acidic microenvironment pH by binding to eotaxin-3 protein, an essential target of eosinophilic esophagitis (Zhang et al., 2012). When ASR targets GRM8 and ESR1 for GERD-related IPF, POLR2 may act as a common downstream molecule. It suggests potential for further study.

It is essential to acknowledge certain limitations in this study. Firstly, although MR enhances result reliability by mitigating confounding factors, the exclusive focus on a European population restricts the study's generalizability. Secondly, our reliance on existing data from online databases might entail missing information, and future research should explore new, undocumented chemicals or targets. Moreover, the effects of ASR on hub genes expression in GERD-related IPF warrant thorough pharmacodynamic and functional verification. Lastly, further studies are imperative to elucidate proposed mechanisms, including the involvement of ESR1/ PGR in the gender-specific effects of steroid hormones on the diseases.

### 5 Conclusion

In conclusion, this study is the first to explore the causal relationship between GERD and IPF and the mechanisms based on MR combined with other bioinformatics methods. We identified a positive causal relationship between GERD and IPF and the shared DEGs between GERD and IPF. Seven hub genes were finally selected. Five key targets were identified to be interfered with by ASR through network pharmacology and molecular docking. ESR1 and GRM8 had sound diagnostic effects in treating GERD-related IPF and are considered diagnostic biomarkers. The response of cells to FGFs stimulation and steroid hormone-mediated signaling pathway may be the potential mechanisms of ASR in GERD-related IPF.

### Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding authors.

### Author contributions

XW: Conceptualization, Visualization, Writing-original draft, Writing-review and editing. XX: Conceptualization, Visualization, Writing-original draft, Writing-review and editing. HF: Data curation, Software, Writing-review and editing. CH: Formal Analysis, Writing-review and editing. HW: Data curation, Software, Writing-original draft. MW: Formal Analysis, Writing-original draft. PL: Formal Analysis, Writing-original draft. FW: Funding acquisition, Supervision, Writing-review and editing. QD: Funding acquisition, Project administration, Writing-review and editing. HY: Conceptualization, Funding acquisition, Project administration, Writing-review and editing.

### Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This study was funded by the National Natural Science Foundation of China (Nos 82374403, 82104829, and 82174347), the General Project of Sichuan Traditional Chinese Medicine Administration (No.

### References

Adewuyi, E. O., O'Brien, E. K., Porter, T., and Laws, S. M. (2022). Relationship of cognition and alzheimer's disease with gastrointestinal tract disorders: a large-scale genetic overlap and mendelian randomisation analysis. *Int. J. Mol. Sci.* 23 (24), 16199. doi:10.3390/ijms232416199

Aggarwal, S., Mannam, P., and Zhang, J. (2016). Differential regulation of autophagy and mitophagy in pulmonary diseases. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 311 (2), L433–L452. doi:10.1152/ajplung.00128.2016

Baqir, M., Vasirreddy, A., Vu, A. N., Moua, T., Chamberlain, A. M., Frank, R. D., et al. (2021). Idiopathic pulmonary fibrosis and gastroesophageal reflux disease: a population-based, case-control study. *Respir. Med.* 178, 106309. doi:10.1016/j.rmed. 2021.106309

Berardi, R., Morgese, F., Santinelli, A., Onofri, A., Biscotti, T., Brunelli, A., et al. (2016). Hormonal receptors in lung adenocarcinoma: expression and difference in outcome by sex. *Oncotarget* 7 (50), 82648–82657. doi:10.18632/oncotarget.12244

Bove, C., Anselmi, L., and Travagli, R. A. (2019). Altered gastric tone and motility response to brain-stem dopamine in a rat model of parkinsonism. *Am. J. Physiol. Gastrointest. Liver Physiol.* 317 (1), G1–G7. doi:10.1152/ajpgi.00076.2019

Brass, D. M., McGee, S. P., Dunkel, M. K., Reilly, S. M., Tobolewski, J. M., Sabo-Attwood, T., et al. (2010). Gender influences the response to experimental silica-induced lung fibrosis in mice. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 299 (5), L664–L671. doi:10. 1152/ajplung.00389.2009 2023MS363), and the "Xinglin Scholars" Talent Youth Advanced Project for Chengdu University of Traditional Chinese Medicine 2023 (QJJJ2023006).

### Acknowledgments

We would like to acknowledge the GWAS Catalogue (GCST90000514; GCST90018120), GEO (GSE65013; GSE10667; GSE39491; GSE53845) database, and each author for providing data. We thank Biorender for improving the quality of our images (agreement number: LO26D8QBQ0). We thank Bullet Edits Limited for the linguistic editing and proofreading of the manuscript. We thank all the developers and participants of the sites mentioned in our article.

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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### Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2024.1348708/ full#supplementary-material

Chambers, R. C. (2022). Preferential PDE4B inhibition - a step toward a new treatment for idiopathic pulmonary fibrosis. N. Engl. J. Med. 386 (23), 2235–2236. doi:10.1056/NEJMe2205411

Chen, J., Hou, N., Zhang, C., Teng, Y., Cheng, X., Li, Z., et al. (2015). Smooth muscle hgs deficiency leads to impaired esophageal motility. *Int. J. Biol. Sci.* 11 (7), 794–802. doi:10.7150/ijbs.12248

Cherubini, E., Mariotta, S., Scozzi, D., Mancini, R., Osman, G., D'Ascanio, M., et al. (2017). BDNF/TrkB axis activation promotes epithelial-mesenchymal transition in idiopathic pulmonary fibrosis. *J. Transl. Med.* 15, 196. doi:10.1186/s12967-017-1298-1

Chiang, C. C., Chen, C. M., Suen, J. L., Su, H. H., Hsieh, C. C., and Cheng, C. M. (2020). Stimulatory effect of gastroesophageal reflux disease (GERD) on pulmonary fibroblast differentiation. *Dig. Liver Dis.* 52 (9), 988–994. doi:10.1016/j.dld.2020.07.010

Choi, W. I., Jeong, J., and Lee, C. W. (2019). Association between EGFR mutation and ageing, history of pneumonia and gastroesophageal reflux disease among patients with advanced lung cancer. *Eur. J. Cancer* 122, 101–108. doi:10.1016/j.ejca.2019.09.010

Colunga Biancatelli, R. M. L., Solopov, P., Dimitropoulou, C., and Catravas, J. D. (2021). Age-dependent chronic lung injury and pulmonary fibrosis following single exposure to hydrochloric acid. *Int. J. Mol. Sci.* 22 (16), 8833. doi:10.3390/ijms22168833

Cotton, C., Alton, P., Hughes, D. M., and Zhao, S. S. (2023). Genetic liability to gastroesophageal reflux disease, obesity, and risk of idiopathic pulmonary fibrosis. *Respir*. *Investig.* 61 (3), 335–338. doi:10.1016/j.resinv.2023.02.005 Davis, C. S., Mendez, B. M., Flint, D. V., Pelletiere, K., Lowery, E., Ramirez, L., et al. (2013). Pepsin concentrations are elevated in the bronchoalveolar lavage fluid of patients with idiopathic pulmonary fibrosis after lung transplantation. *J. Surg. Res.* 185, e101–e108. doi:10.1016/j.jss.2013.06.011

Dorry, S. J., Ansbro, B. O., Ornitz, D. M., Mutlu, G. M., and Guzy, R. D. (2020). FGFR2 is required for AEC2 homeostasis and survival after bleomycin-induced lung injury. *Am. J. Respir. Cell. Mol. Biol.* 62 (5), 608–621. doi:10.1165/rcmb.2019-0079OC

Duckworth, A., Gibbons, M. A., Allen, R. J., Almond, H., Beaumont, R. N., Wood, A. R., et al. (2021). Telomere length and risk of idiopathic pulmonary fibrosis and chronic obstructive pulmonary disease: a mendelian randomisation study. *Lancet Respir. Med.* 9 (3), 285–294. doi:10.1016/S2213-2600(20)30364-7

El, A. E., Seeger, W., and Bellusci, S. (2017). Therapeutic and pathological roles of fibroblast growth factors in pulmonary diseases. *Dev. Dyn.* 246 (4), 235–244. doi:10.1002/dvdy.24468

Eusebi, L. H., Ratnakumaran, R., Yuan, Y., Solaymani-Dodaran, M., Bazzoli, F., and Ford, A. C. (2018). Global prevalence of, and risk factors for, gastro-oesophageal reflux symptoms: a meta-analysis. *Gut* 67, 430–440. doi:10.1136/gutjnl-2016-313589

Feng, H. M., Li, Y., Luo, X. D., Liang, T. T., Jia, M. T., Qiang, Z. Z., et al. (2022). Research progress on chemical constituents and pharmacological effects of danggui (radix angelicae sinensis) and prediction analysis on its quality markers. *Chin. archives* traditional Chin. Med. 40, 159–166. doi:10.13193/j.issn.1673-7717.2022.04.036

Filpa, V., Moro, E., Protasoni, M., Crema, F., Frigo, G., and Giaroni, C. (2016). Role of glutamatergic neurotransmission in the enteric nervous system and brain-gut axis in health and disease. *Neuropharmacology* 111, 14–33. doi:10.1016/j.neuropharm.2016.08.024

Gao, J., Feng, L. J., Huang, Y., Li, P., Xu, D. J., Li, J., et al. (2012). Total glucosides of Danggui Buxue Tang attenuates bleomycin-induced pulmonary fibrosis via inhibition of extracellular matrix remodelling. *J. Pharm. Pharmacol.* 64 (6), 811–820. doi:10.1111/j. 2042-7158.2012.01490.x

Gast, M. T., Tönjes, A., Keller, M., Horstmann, A., Steinle, N., Scholz, M., et al. (2013). The role of rs2237781 within GRM8 in eating behavior. *Brain Behav.* 3 (5), 495–502. doi:10.1002/brb3.151

Gavini, S., Finn, R. T., Lo, W. K., Goldberg, H. J., Burakoff, R., Feldman, N., et al. (2015). Idiopathic pulmonary fibrosis is associated with increased impedance measures of reflux compared to non-fibrotic disease among pre-lung transplant patients. *Neurogastroenterol. Motil.* 27 (9), 1326–1332. doi:10.1111/nmo.12627

George, P. M., Patterson, C. M., Reed, A. K., and Thillai, M. (2019). Lung transplantation for idiopathic pulmonary fibrosis. *Lancet Respir. Med.* 7, 271–282. doi:10.1016/S2213-2600(18)30502-2

Gharaee-Kermani, M., Hatano, K., Nozaki, Y., and Phan, S. H. (2005). Gender-based differences in bleomycin-induced pulmonary fibrosis. *Am. J. Pathol.* 166 (6), 1593–1606. doi:10.1016/S0002-9440(10)62470-4

Guo, J., Liang, J., Guo, Z., Bai, X., Zhang, H., Zhang, N., et al. (2024). Network pharmacology and transcriptomics to determine Danggui Yifei Decoction mechanism of action for the treatment of chronic lung injury. *J. Ethnopharmacol.* 318 (Pt A), 116873. doi:10.1016/j.jep.2023.116873

Han, D., and Zhang, C. (2022). The oxidative damage and inflammation mechanisms in GERD-induced Barrett's esophagus. *Front. Cell. Dev. Biol.* 10, 885537. doi:10.3389/fcell. 2022.885537

Han, G., Zhou, Y. F., Zhang, M. S., Cao, Z., Xie, C. H., Zhou, F. X., et al. (2006). Angelica sinensis down-regulates hydroxyproline and Tgfb1 and provides protection in mice with radiation-induced pulmonary fibrosis. *Radiat. Res.* 165 (5), 546–552. doi:10.1667/RR3543.1

Hao, Y., Li, J., Dan, L., Wu, X., Xiao, X., Yang, H., et al. (2024). Chinese medicine as a therapeutic option for pulmonary fibrosis: clinical efficacies and underlying mechanisms. *J. Ethnopharmacol.* 318 (Pta), 116836. doi:10.1016/j.jep.2023.116836

Henderson, N. C., Rieder, F., and Wynn, T. A. (2020). Fibrosis: from mechanisms to medicines. *Nature* 587 (7835), 555–566. doi:10.1038/s41586-020-2938-9

Herrmann, F. E., Hesslinger, C., Wollin, L., and Nickolaus, P. (2022). BI 1015550 is a PDE4B inhibitor and a clinical drug candidate for the oral treatment of idiopathic pulmonary fibrosis. *Front. Pharmacol.* 13, 838449. doi:10.3389/fphar.2022.838449

Hertz, D. L., Henry, N. L., Kidwell, K. M., Thomas, D., Goddard, A., Azzouz, F., et al. (2016). ESR1 and PGR polymorphisms are associated with estrogen and progesterone receptor expression in breast tumors. *Physiol. Genomics* 48 (9), 688–698. doi:10.1152/ physiolgenomics.00065.2016

Hewlett, J. C., Kropski, J. A., and Blackwell, T. S. (2018). Idiopathic pulmonary fibrosis: epithelial-mesenchymal interactions and emerging therapeutic targets. *Matrix Biol.* 71-72, 112–127. doi:10.1016/j.matbio.2018.03.021

Higo, H., Ohashi, K., Tomida, S., Okawa, S., Yamamoto, H., Sugimoto, S., et al. (2022). Identification of targetable kinases in idiopathic pulmonary fibrosis. *Respir. Res.* 23 (1), 20. doi:10.1186/s12931-022-01940-y

Hoppo, T., Komatsu, Y., and Jobe, B. A. (2014). Gastroesophageal reflux disease and patterns of reflux in patients with idiopathic pulmonary fibrosis using hypopharyngeal multichannel intraluminal impedance. *Dis. Esophagus* 27 (6), 530–537. doi:10.1111/j. 1442-2050.2012.01446.x

Huang, J. J., Joh, J. W., Fuentebella, J., Patel, A., Nguyen, T., Seki, S., et al. (2010). Eotaxin and FGF enhance signaling through an extracellular signal-related kinase (ERK)-dependent pathway in the pathogenesis of Eosinophilic esophagitis. Allergy Asthma Clin. Immunol. 6 (1), 25. doi:10.1186/1710-1492-6-25

Hung, C. C., Lin, C. H., and Lane, H. Y. (2021). Cystine/glutamate antiporter in schizophrenia: from molecular mechanism to novel biomarker and treatment. *Int. J. Mol. Sci.* 22 (18), 9718. doi:10.3390/ijms22189718

Ka, H., Al-Ramadan, S., Erikson, D. W., Johnson, G. A., Burghardt, R. C., Spencer, T. E., et al. (2007). Regulation of expression of fibroblast growth factor 7 in the pig uterus by progesterone and estradiol. *Biol. Reprod.* 77 (1), 172–180. doi:10.1095/biolreprod. 106.056309

Khor, Y. H., Bissell, B., Ghazipura, M., Herman, D., Hon, S. M., Hossain, T., et al. (2022). Antacid medication and antireflux surgery in patients with idiopathic pulmonary fibrosis: a systematic review and meta-analysis. *Ann. Am. Thorac. Soc.* 19, 833–844. doi:10.1513/AnnalsATS.202102-172OC

King, C., and Nathan, S. (2013). Identification and treatment of comorbidities in idiopathic pulmonary fibrosis and other fibrotic lung diseases. *Curr. Opin. Pulm. Med.* 19 (5), 466–473. doi:10.1097/MCP.0b013e328363f460

Kriegsmann, K., Zgorzelski, C., Muley, T., Christopoulos, P., Von Winterfeld, M., Herpel, E., et al. (2020). Immunohistological expression of oestrogen receptor, progesterone receptor, mammaglobin, human epidermal growth factor receptor 2 and GATA-binding protein 3 in non-small-cell lung cancer. *Histopathology* 77 (6), 900–914. doi:10.1111/his.14203

Lamas, D. J., Kawut, S. M., Bagiella, E., Philip, N., Arcasoy, S. M., and Lederer, D. J. (2011). Delayed access and survival in idiopathic pulmonary fibrosis: a cohort study. *Am. J. Respir. Crit. Care Med.* 184 (7), 842–847. doi:10.1164/rccm.201104-06680C

Le, H. Q., Hill, M. A., Kollak, I., Keck, M., Schroeder, V., Wirth, J., et al. (2021). An EZH2-dependent transcriptional complex promotes aberrant epithelial remodelling after injury. *EMBO Rep.* 22 (8), e52785. doi:10.15252/embr.202152785

Lee, J. S., Song, J. W., Wolters, P. J., Elicker, B. M., King, T. E., Jr., Kim, D. S., et al. (2012). Bronchoalveolar lavage pepsin in acute exacerbation of idiopathic pulmonary fibrosis. *Eur. Respir. J.* 39, 352–358. doi:10.1183/09031936.00050911

Lee, L. Y., and Yu, J. (2014). Sensory nerves in lung and airways. *Compr. Physiol.* 4 (1), 287–324. doi:10.1002/cphy.c130020

Lee, Y. S., Jung, W. Y., Heo, H., Park, M. G., Oh, S. H., Park, B. G., et al. (2018). Exosome-mediated ultra-effective direct conversion of human fibroblasts into neural progenitor-like cells. *ACS Nano* 12 (3), 2531–2538. doi:10.1021/acsnano.7b08297

Li, M., Zhang, J. J., Li, X. N., Zhang, J., An, Y. X., Zhang, Q. C., et al. (2021). Effects of duodenal ligation on gastroesophageal reflux and bleomycin-induced pulmonary fibrosis in rats. *Natl. Med. J. China* 101 (21), 1598–1605. doi:10.3760/cma.j. cn112137-20201017-02859

Li, W., Ju, K., Li, Z., He, K., Chen, J., Wang, Q., et al. (2016). Significant association of GRM7 and GRM8 genes with schizophrenia and major depressive disorder in the Han Chinese population. *Eur. Neuropsychopharmacol.* 26 (1), 136–146. doi:10.1016/j. euroneuro.2015.05.004

Liu, X. Y., Zheng, L. F., Fan, Y. Y., Shen, Q. Y., Qi, Y., Li, G. W., et al. (2022b). Activation of dopamine  $D_2$  receptor promotes pepsinogen secretion by suppressing somatostatin release from the mouse gastric mucosa. *Am. J. Physiol. Cell. Physiol.* 322 (3), C327–C337. doi:10.1152/ajpcell.00385.2021

Liu, Y., Yang, X., Gan, J., Chen, S., Xiao, Z. X., and Cao, Y. (2022a). CB-Dock2: improved protein-ligand blind docking by integrating cavity detection, docking and homologous template fitting. *Nucleic Acids Res.* 50 (W1), W159–W164. doi:10.1093/ nar/gkac394

Lozo, V. E., Lozo, M., Mise, K., Gudelj, I., Puljiz, Z., Jurcev-Savicevic, A., et al. (2014). Bronchoalveolar pH and inflammatory biomarkers in newly diagnosed IPF and GERD patients: a case-control study. *Med. Sci. Monit.* 15 (20), 255–261. doi:10.12659/MSM. 889800

Lucas, L. M., Dwivedi, V., Senfeld, J. I., Cullum, R. L., Mill, C. P., Piazza, J. T., et al. (2022). The yin and yang of ERBB4: tumor suppressor and oncoprotein. *Pharmacol. Rev.* 74 (1), 18–47. doi:10.1124/pharmrev.121.000381

Ma, H., Wu, X., Li, Y., and Xia, Y. (2022). Research progress in the molecular mechanisms, therapeutic targets, and drug development of idiopathic pulmonary fibrosis. *Front. Pharmacol.* 13, 963054. doi:10.3389/fphar.2022.963054

Ma, L. N., W, R., and Miao, M. S. (2020). Medication rule of Chinese materia medica in treatment of pulmonary fibrosis based on data mining. *Chin. Traditional Herb. Drugs* 51, 1406–1411. doi:10.7501/j.issn.0253-2670.2020.06.005

Machahua, C., Montes-Worboys, A., Llatjos, R., Escobar, I., Dorca, J., Molina-Molina, M., et al. (2016). Increased AGE-RAGE ratio in idiopathic pulmonary fibrosis. *Respir. Res.* 17 (1), 144–211. doi:10.1186/s12931-016-0460-2

Maret-Ouda, J., Markar, S. R., and Lagergren, J. (2020). Gastroesophageal reflux disease. JAMA 324, 2565. doi:10.1001/jama.2020.21573

Marinova, M., Solopov, P., Dimitropoulou, C., Colunga Biancatelli, R. M. L., and Catravas, J. D. (2019). Acute exposure of mice to hydrochloric acid leads to the development of chronic lung injury and pulmonary fibrosis. *Inhal. Toxicol.* 31 (4), 147–160. doi:10.1080/08958378.2019.1624895

Mathai, S. K., and Schwartz, D. A. (2019). Translational research in pulmonary fibrosis. *Transl. Res.* 209, 1–13. doi:10.1016/j.trsl.2019.02.001

Molyneaux, P. L., Cox, M. J., Wells, A. U., Kim, H. C., Ji, W., Cookson, W. O., et al. (2017). Changes in the respiratory microbiome during acute exacerbations of idiopathic pulmonary fibrosis. *Respir. Res.* 18 (1), 29. doi:10.1186/s12931-017-0511-3

Nedić, O., Rattan, S., Grune, T., and Trougakos, I. (2013). Molecular effects of advanced glycation end products on cell signalling pathways, ageing and pathophysiology. *Free Radic. Res.* 47, 28–38. doi:10.3109/10715762.2013.806798

Neumann, H., Wex, T., Vieth, M., Peitz, U., Roessner, A., and Malfertheiner, P. (2007). Gastroesophageal reflux disease leads to major alterations in lectin-binding in the columnar epithelium of the gastroesophageal junction. *Scand. J. Gastroenterol.* 42 (7), 791–798. doi:10.1080/00365520601127158

Nguyen, X. X., Renaud, L., and Feghali-Bostwick, C. (2021). Identification of impacted pathways and transcriptomic markers as potential mediators of pulmonary fibrosis in transgenic mice expressing human IGFBP5. *Int. J. Mol. Sci.* 22 (22), 12609. doi:10.3390/jims222212609

Nilsson, M., Johnsen, R., Ye, W., Hveem, K., and Lagergren, J. (2004). Lifestyle related risk factors in the aetiology of gastro-oesophageal reflux. *Gut* 53, 1730–1735. doi:10. 1136/gut.2004.043265

Oczypok, E. A., Perkins, T. N., and Oury, T. D. (2017). All the "RAGE" in lung disease: the receptor for advanced glycation endproducts (RAGE) is a major mediator of pulmonary inflammatory responses. *Paediatr. Respir. Rev.* 23, 40–49. doi:10.1016/j.prrv.2017.03.012

Ogawa, T., Shichino, S., Ueha, S., and Matsushima, K. (2021). Macrophages in lung fibrosis. Int. Immunol. 33 (12), 665–671. doi:10.1093/intimm/dxab040

Oldham, J. M., and Collard, H. R. (2017). Comorbid conditions in idiopathic pulmonary fibrosis: recognition and management. *Front. Med. (Lausanne).* 4, 123. doi:10.3389/fmed.2017.00123

Ong, J. S., An, J., Han, X., Law, M. H., Nandakumar, P., et al. (2022). Multitrait genetic association analysis identifies 50 new risk loci for gastro-oesophageal reflux, seven new loci for Barrett's oesophagus and provides insights into clinical heterogeneity in reflux diagnosis. *Gut* 71 (6), 1053–1061. doi:10.1136/gutjnl-2020-323906

Patankar, M., Li, M., Khalatbari, A., Castle, J. D., Hu, L., Zhang, C., et al. (2022). Inflammatory and proliferative pathway activation in human esophageal myofibroblasts treated with acidic bile salts. *Int. J. Mol. Sci.* 23, 10371. doi:10.3390/ijms231810371

Qian, W., Cai, X., Qian, Q., Wang, D., and Zhang, L. (2020). Angelica sinensis polysaccharide suppresses epithelial-mesenchymal transition and pulmonary fibrosis via a DANCR/AUF-1/FOXO3 regulatory Axis. *Aging Dis.* 11 (1), 17–30. doi:10.14336/AD.2019.0512

Qing, J., Ren, Y., Zhang, Y., Yan, M., Zhang, H., Wu, D., et al. (2022). Dopamine receptor D2 antagonism normalizes profibrotic macrophage-endothelial crosstalk in non-alcoholic steatohepatitis. *J. Hepatol.* 76 (2), 394–406. doi:10.1016/j.jhep.2021.09.032

Raghu, G., Berk, M., Campochiaro, P. A., Jaeschke, H., Marenzi, G., Richeldi, L., et al. (2021). The multifaceted therapeutic role of N-acetylcysteine (NAC) in disorders characterized by oxidative stress. *Curr. Neuropharmacol.* 19 (8), 1202–1224. doi:10. 2174/1570159X19666201230144109

Raghu, G., Freudenberger, T. D., Yang, S., Curtis, J. R., Spada, C., Hayes, J., et al. (2006). High prevalence of abnormal acid gastro-oesophageal reflux in idiopathic pulmonary fibrosis. *Eur. Respir. J.* 27, 136–142. doi:10.1183/09031936.06.00037005

Reynolds, C. J., Del Greco, M. F., Allen, R. J., Flores, C., Jenkins, R. G., Maher, T. M., et al. (2023). The causal relationship between gastro-oesophageal reflux disease and idiopathic pulmonary fibrosis: a bidirectional two-sample Mendelian randomisation study. *Eur. Respir. J.* 61 (5), 2201585. doi:10.1183/13993003.01585-2022

Saegusa, Y., Takeda, H., Muto, S., Oridate, N., Nakagawa, K., Sadakane, C., et al. (2011). Decreased motility of the lower esophageal sphincter in a rat model of gastroesophageal reflux disease may be mediated by reductions of serotonin and acetylcholine signaling. *Biol. Pharm. Bull.* 34 (5), 704–711. doi:10.1248/bpb.34.704

Sathish, V., Martin, Y. N., and Prakash, Y. S. (2015). Sex steroid signaling: implications for lung diseases. *Pharmacol. Ther.* 150, 94–108. doi:10.1016/j.pharmthera.2015.01.007

Schier, A. C., and Taatjes, D. J. (2020). Structure and mechanism of the RNA polymerase II transcription machinery. *Genes. Dev.* 34 (7-8), 465–488. doi:10.1101/gad. 335679.119

Sgalla, G., Biffi, A., and Richeldi, L. (2016). Idiopathic pulmonary fibrosis: diagnosis, epidemiology and natural history. *Respirology* 21 (3), 427–437. doi:10.1111/resp.12683

Shannon, P., Markiel, A., Ozier, O., Baliga, N. S., Wang, J. T., Ramage, D., et al. (2003). Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res.* 13 (11), 2498–2504. doi:10.1101/gr.1239303

Shim, B., Pacheco-Rodriguez, G., Kato, J., Darling, T. N., Vaughan, M., and Moss, J. (2013). Sex-specific lung diseases: effect of oestrogen on cultured cells and in animal models. *Eur. Respir. Rev.* 22 (129), 302–311. doi:10.1183/09059180.00002813

Smith, L. C., Moreno, S., Robertson, L., Robinson, S., Gant, K., Bryant, A. J., et al. (2018). Transforming growth factor beta1 targets estrogen receptor signaling in bronchial epithelial cells. *Respir. Res.* 19 (1), 160. doi:10.1186/s12931-018-0861-5

Smith, L. C., Moreno, S., Robinson, S., Orandle, M., Porter, D. W., Das, D., et al. (2019). Multi-walled carbon nanotubes inhibit estrogen receptor expression *in vivo* and *in vitro* through transforming growth factor beta1. *NanoImpact* 14, 100152. doi:10. 1016/j.impact.2019.100152

Soares, R. V., Forsythe, A., Hogarth, K., Sweiss, N. J., Noth, I., and Patti, M. G. (2011). Interstitial lung disease and gastroesophageal reflux disease: key role of esophageal function tests in the diagnosis and treatment. Arq. Gastroenterol. 48 (2), 91–97. doi:10. 1590/s0004-28032011000200002

Solopov, P., Colunga Biancatelli, R. M. L., Dimitropoulou, C., and Catravas, J. D. (2021). Dietary phytoestrogens ameliorate hydrochloric acid-induced chronic lung injury and pulmonary fibrosis in mice. *Nutrients* 13 (10), 3599. doi:10.3390/nu13103599

Stancil, I. T., Michalski, J. E., Davis-Hall, D., Chu, H. W., Park, J. A., Magin, C. M., et al. (2021). Pulmonary fibrosis distal airway epithelia are dynamically and structurally dysfunctional. *Nat. Commun.* 12, 4566. doi:10.1038/s41467-021-24853-8

Tudorache, V., Traila, D., Marc, M., Oancea, C., Manolescu, D., Tudorache, E., et al. (2019). Impact of moderate to severe obstructive sleep apnea on the cognition in idiopathic pulmonary fibrosis. *PLoS One* 14 (2), e0211455. doi:10.1371/journal.pone.0211455

Vermeulen, Z., Hervent, A. S., Dugaucquier, L., Vandekerckhove, L., Rombouts, M., Beyens, M., et al. (2017). Inhibitory actions of the NRG-1/ErbB4 pathway in macrophages during tissue fibrosis in the heart, skin, and lung. *Am. J. Physiol. Heart Circ. Physiol.* 313 (5), H934–H945. doi:10.1152/ajpheart.00206.2017

Wang, H. H., Liu, M., Clegg, D. J., Portincasa, P., and Wang, D. Q. (2009). New insights into the molecular mechanisms underlying effects of estrogen on cholesterol gallstone formation. *Biochim. Biophys. Acta* 1791 (11), 1037–1047. doi:10.1016/j.bbalip.2009.06.006

Wang, M., Zhang, Z., Liu, J., Song, M., Zhang, T., Chen, Y., et al. (2022). Gefitinib and fostamatinib target EGFR and SYK to attenuate silicosis: a multi-omics study with drug exploration. *Signal Transduct. Target Ther.* 7 (1), 157. doi:10.1038/s41392-022-00959-3

Warren, H. F., Louie, B. E., Farivar, A. S., Wilshire, C., and Aye, R. W. (2017). Manometric changes to the lower esophageal sphincter after magnetic sphincter augmentation in patients with chronic gastroesophageal reflux disease. *Ann. Surg.* 266, 99–104. doi:10.1097/SLA.00000000001935

Wei, D. Z., Guo, X. Y., Lin, L. N., Lin, M. X., Gong, Y. Q., Ying, B. Y., et al. (2016). Effects of angelicin on ovalbumin (OVA)-induced airway inflammation in a mouse model of asthma. *Inflammation* 39 (6), 1876–1882. doi:10.1007/s10753-016-0423-2

Weng, J., Tu, M., Wang, P., Zhou, X., Wang, C., Wan, X., et al. (2019). Amiodarone induces cell proliferation and myofibroblast differentiation via ERK1/2 and p38 MAPK signaling in fibroblasts. *Biomed. Pharmacother*. 115, 108889. doi:10.1016/j.biopha.2019.108889

Wheeler, J. C., Vanoni, S., Zeng, C., Waggoner, L., Yang, Y., Wu, D., et al. (2019). 17β-Estradiol protects the esophageal epithelium from IL-13-induced barrier dysfunction and remodeling. *J. Allergy Clin. Immunol.* 143 (6), 2131–2146. doi:10.1016/j.jaci.2018.10.070

Willander, H., Hermansson, E., Johansson, J., and Presto, J. (2011). BRICHOS domain associated with lung fibrosis, dementia and cancer-a chaperone that prevents amyloid fibril formation? *FEBS J.* 278 (20), 3893–3904. doi:10.1111/j.1742-4658.2011.08209.x

Xu, T., Xie, M., Jing, X., Jiang, H., Wu, X., Wang, X., et al. (2023). Loss of miR-26b-5p promotes gastric cancer progression via miR-26b-5p-PDE4B/CDK8-STAT3 feedback loop. *J. Transl. Med.* 21 (1), 77. doi:10.1186/s12967-023-03933-x

Xue, L., Deng, D., Zheng, S., Tang, M., Yang, Z., Pei, H., et al. (2020). Design, synthesis and discovery of 2(1H)-quinolone derivatives for the treatment of pulmonary fibrosis through inhibition of TGF- $\beta$ /smad dependent and independent pathway. *Eur. J. Med. Chem.* 197, 112259. doi:10.1016/j.ejmech.2020.112259

Yamaguchi, K., Iwamoto, H., Horimasu, Y., Ohshimo, S., Fujitaka, K., Hamada, H., et al. (2017). AGER gene polymorphisms and soluble receptor for advanced glycation end product in patients with idiopathic pulmonary fibrosis. *Respirology* 22 (5), 965–971. doi:10.1111/resp.12995

Yisireyili, M., Wulamu, W., Aili, A., Li, Y., Alimujiang, A., Aipire, A., et al. (2019). Chronic restraint stress induces esophageal fibrosis with enhanced oxidative stress in a murine model. *Exp. Ther. Med.* 18 (2), 1375–1383. doi:10.3892/etm.2019.7669

Zhang, A., Wu, H., Wang, C., and Tian, S. (2023). Integrative analysis of Mendelian randomization and gene expression profiles reveals a null causal relationship between adiponectin and diabetic retinopathy. *Adipocyte* 12 (1), 2234522. doi:10.1080/21623945. 2023.234522

Zhang, L., Yan, Q., Zhang, W., Li, X., Zhang, X., Du, S., et al. (2022). Enhancement of the functionality of attenuating acute lung injury by a microemulsion formulation with volatile oil of Angelicae Sinensis Radix and Ligusticum Chuanxiong Rhizoma encapsulated. *Biomed. Pharmacother.* 156, 113888. doi:10.1016/j.biopha.2022. 113888

Zhang, X., Cheng, E., Huo, X., Yu, C., Zhang, Q., Pham, T. H., et al. (2012). Omeprazole blocks STAT6 binding to the eotaxin-3 promoter in eosinophilic esophagitis cells. *PLoS One* 7 (11), e50037. doi:10.1371/journal.pone.0050037

Zhang, Y., Gu, L., Xia, Q., Tian, L., Qi, J., and Cao, M. (2020). Radix astragali and radix angelicae sinensis in the treatment of idiopathic pulmonary fibrosis: a systematic review and meta-analysis. *Front. Pharmacol.* 11, 415. doi:10.3389/fphar.2020.00415

Zhao, W., Wang, L., Wang, Y., Yuan, H., Zhao, M., Lian, H., et al. (2023). Injured endothelial cell: a risk factor for pulmonary fibrosis. *Int. J. Mol. Sci.* 24 (10), 8749. doi:10. 3390/ijms24108749

Zhou, C. C., Pang, Y., He, Y., and Tang, L. L. (2023). A Mendelian randomization study of depression and risk of breast cancer. *Chin. Ment. Health J.* 37 (06), 471–478. doi:10.3969/j.issn.1000-6729.2023.06.004

Zhou, Y. M., Dong, X. R., Xu, D., Tang, J., and Cui, Y. L. (2024). Therapeutic potential of traditional Chinese medicine for interstitial lung disease. *J. Ethnopharmacol.* 318, 116952. doi:10.1016/j.jep.2023.116952