Check for updates

OPEN ACCESS

EDITED BY Zhiyu Zhang, Fourth Affiliated Hospital of China Medical University, China

REVIEWED BY Satoru Sasagawa, Nozaki Tokushukai Hospital, Japan Sam Rowbotham, Harvard Medical School, United States

*CORRESPONDENCE Wenjie Jiao, ⊠ jiaowj@qduhospital.cn

[†]These authors have contributed equally to this work

RECEIVED 07 February 2024 ACCEPTED 02 April 2024 PUBLISHED 12 April 2024

CITATION

Yao Z, Song P and Jiao W (2024), Pathogenic role of super-enhancers as potential therapeutic targets in lung cancer. *Front. Pharmacol.* 15:1383580. doi: 10.3389/fphar.2024.1383580

COPYRIGHT

© 2024 Yao, Song and Jiao. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Pathogenic role of super-enhancers as potential therapeutic targets in lung cancer

Zhiyuan Yao^{1†}, Peng Song^{2†} and Wenjie Jiao^{1*}

¹Department of Thoracic Surgery, The Affiliated Hospital of Qingdao University, Qingdao, China, ²Department of Thoracic Surgery, Beijing Friendship Hospital, Capital Medical University, Beijing, China

Lung cancer is still one of the deadliest malignancies today, and most patients with advanced lung cancer pass away from disease progression that is uncontrollable by medications. Super-enhancers (SEs) are large clusters of enhancers in the genome's non-coding sequences that actively trigger transcription. Although SEs have just been identified over the past 10 years, their intricate structure and crucial role in determining cell identity and promoting tumorigenesis and progression are increasingly coming to light. Here, we review the structural composition of SEs, the auto-regulatory circuits, the control mechanisms of downstream genes and pathways, and the characterization of subgroups classified according to SEs in lung cancer. Additionally, we discuss the therapeutic targets, several small-molecule inhibitors, and available treatment options for SEs in lung cancer. Combination therapies have demonstrated considerable advantages in preclinical models, and we anticipate that these drugs will soon enter clinical studies and benefit patients.

KEYWORDS

epigenetic modifications, lung cancer, subclassification, super-enhancers, therapeutic targets

1 Introduction

Lung cancer remains one of the most prevalent and severe tumors, although the mortality risk continues to decline thanks to improvements in early identification and treatment strategies (Siegel et al., 2022). Of all lung cancer histologic subtypes, the most common (85%) is non-small cell lung cancer (NSCLC), with the remaining 10%–15% being small cell lung cancer (SCLC) and other rare cancers (Siegel et al., 2020). Frustratingly, with a 5-year survival rate of just seven percent, SCLC is the most aggressive subtype of lung cancer (Frese et al., 2021). With the advance of targeted medicines and immunotherapies, the long-term prognosis of lung cancer has dramatically improved; however, it is still less than satisfactory for most advanced lung cancers (Wang et al., 2021).

Genetic mutations have been extensively studied in lung cancer, which change coding sequences (e.g., sequences directly coding for oncoproteins) and non-coding sequences (e.g., promoters and enhancers), directly or indirectly favoring clonal proliferation in cancer cells (Lengauer et al., 1998). Current targeted therapies for lung cancer are also mainly based on genetic mutation. However, disturbances in epigenetic regulation can also lead to the occurrence and metastasis of lung cancer (Dong et al., 2023; Kostyrko et al., 2023). For example, the MYC proto-oncogenes family can broadly affect cell proliferation and apoptosis through complex regulatory networks (Carroll et al., 2018). In addition to the copy number increases from genetic mutations, MYC overexpression is also inextricably linked to surrounding regulatory sequences, with a growing number of studies



demonstrating that super-enhancers (SEs) act as culprits in this process. Studies have revealed that the mechanism of SE formation involves mutations, and the uniqueness of epigenetic modifications at SEs loci is related to the progression of acute myeloid leukemia (Cao et al., 2022), colorectal cancer (Liu Q. et al., 2022), and esophageal carcinoma (Shi et al., 2022).

Hence, we introduce the structure and function of SEs and summarize the oncogenic mechanism, classifying functions, and potential therapeutic targets and drugs of SEs in lung cancer here.

2 Subsections

2.1 Identification and structural characterization of SEs

In the 1980s, a 72 bp sequence, later called "enhancer," was identified in the genome of the SV40/hemoglobin β 1 recombinant gene, accompanied by the overexpression of the rabbit β globin gene (Banerji et al., 1981). Enhancers are cis-regulatory elements that interact with promoters from different positions and distances to increase the expression of controlled genes (Blackwood and Kadonaga, 1998). Richard A. Young identified larger clusters of enhancers, known as SEs (or stretch enhancers), which drove high transcription levels of key genes and defined cell identity in pluripotent embryonic stem cells (ESCs) (Whyte et al., 2013). Researchers have created some online repositories such as dbSUPER (Khan and Zhang, 2016) (https://asntech.org/dbsuper/), SEdb (Jiang et al., 2019; Wang Y. et al., 2023) (https://bio.liclab. net/sedb/), SEanalysis (Qian et al., 2019) (https://bio.liclab.net/ SEanalysis/), and SEA (Wei et al., 2016; Chen et al., 2020) (http://sea.edbc.org/) to share SEs (and related genes) and their genetic and epigenetic annotations. The constituent enhancers of SEs are with multiple overlapping enrichments of H3K4me1 and H3K27ac of active enhancers (AEs) marks (Yoo et al., 2019; Barral and Déjardin, 2023) (Figure 1). The Rank Ordering of Super Enhancers (ROSE) algorithm is the most commonly used method to identify SEs (Lovén et al., 2013). The algorithm ranks H3K27ac chromatin immunoprecipitation sequencing (ChIP-Seq) signals and defines enhancers above the inflection point (slope >1) as SEs. However, there is still no consensus on an arbitrary cutoff of enrichment level within certain proximity for considering whether an active enhancer is a super-enhancer. As ChIP-seq has advanced, more and more SEs have been discovered to be involved in carcinogenesis, tumor immune evasion (Xu et al., 2019), and various autoimmune disorders (Parker et al., 2013; Vahedi et al., 2015; Yamagata et al., 2022). It is worth mentioning that other techniques, including 4C-seq (Jiang et al., 2020), Hi-C (Huang J. et al., 2018), ChIA-PET (Dowen et al., 2014), ChIP-STARR-seq (Barakat et al., 2018), GRO-seq (Hah et al., 2015), DNase-seq (Kang et al., 2021), and ATAC-seq (Adam et al., 2020), have also been employed to detect SEs in recent years. Lately, Jialiang Huang's team refined the hierarchical structure of SEs into hub enhancers and non-hub enhancers by hierarchical scoring (Huang J. et al., 2018). Hub enhancers outperform non-hub enhancers in the interaction with regulatory factors and effects on activating target gene transcription.

As a member of the bromodomain and extra-terminal domain (BET) family and closely associated with SE-related oncogenic transcription, bromodomain-containing protein 4 (BRD4) binds to acetylated modified histones and recruits additional proteins (such as TEFb and Mediator) to indirectly participate in the initiation of transcription and the control of transcriptional elongation (Yang et al., 2005). BET inhibitors (BETi) separate BET proteins from chromatin by competitively binding with histone-acetylated lysines of the BET bromodomain. SEs are highly sensitive to BETi (Fontanals-Cirera et al., 2017). JQ1 is the first reported BETi (Filippakopoulos et al., 2010) and has been

widely used for SE identification (Das et al., 2023; Zheng et al., 2024). JQ1 disrupts the transcriptional activity of the master regulators ETS2, HNF4A, and JUNB, suppresses the TGF-induced EMT status, and decreases tumor cell migration and invasion in NSCLC cells (Chang et al., 2016). JQ1, in combination with NSD2 knockdown, inhibits the activation of RAS-driven transcriptional programs in lung cancer cells (García-Carpizo et al., 2016). JQ1 downregulates the expression of HK2, whose promoter is specifically SEs hijacked in a dose-dependent manner in LUAD (Song et al., 2022). In addition, JQ1 exhibits this dose-dependent pharmacological profile in acute myeloid leukemia (Jang et al., 2017), uterine carcinoma (Bonazzoli et al., 2018), and diffuse large B-cell lymphoma (Trabucco et al., 2015).

2.2 Transcription factors (TFs) and core TF involved in coding SE-associated genes

Chromatin accessibility allows physical contact between cisacting elements and trans-acting factors to regulate transcription processes (Klemm et al., 2019). Breast cancer cells undergoing SE reprogramming produced the accessible chromatin structure and had a higher propensity for lung metastasis (Li K. et al., 2019). TFs are proteins that bind to specific DNA sequences (Spitz and Furlong, 2012). If abnormally enriched at oncogene promoters or enhancers, however, they can result in gene overexpression and carcinogenesis (You et al., 2018). SE-driven genes encode core TFs, which can bind to their SEs or others and interact with other core TFs to regulate the expression of additional genes and form cell- or tissue-specific core regulatory circuits (CRC) (Saint-André et al., 2016; Feng C. et al., 2023). For example, the three specifically SE-activated core TFs ELF3, EHF, and TGIF1 constituted the major component of a CRC with an interdependent pattern that tightly regulated cell migration and invasion in lung adenocarcinoma (LUAD) (Zhang T. et al., 2020).

2.3 Essential SE-associated transcriptional coactivators

Transcriptional coactivators are a variety of proteins with transcription-related functions. As mentioned above, BRD4 is a crucial part of SEs and plays an essential role in the transcriptional activation of oncogenes. Besides, MED1, Mediator, and CBP/ p300 with acetyltransferase activity bind to TFs to assist in the assembly of the transcription preinitiation complex and stabilize the transcriptional activity of RNA polymerase II (Ogryzko et al., 1996; Mao et al., 2019; Rengachari et al., 2021). Benjamin R Sabari's team proposed that transcriptional coactivators enhance transcriptional activity by forming droplets around SEs through a phase separation process (Sabari et al., 2018). However, whether SE-dependent transcriptional enhancement depends on this mechanism is still debatable (Trojanowski et al., 2022).

Overall, the effects of SEs on transcriptional activation are more forceful than that of typical enhancers (TEs) due to their higher region spanning, higher histone modifications, TFs, and RNA polymerase II enrichment, and more binding sites for transcriptional coactivators (Hnisz et al., 2013; Whyte et al., 2013).

2.4 Epigenetic modification associated with SEs

Two key epigenetic regulatory mechanisms involved in SEs are histone modifications (mostly methylation and acetylation) and DNA methylation. Modified histones in the presence of catalytic enzymes (histone lysine methyltransferases or histone acetyltransferases, etc.) can alter chromatin accessibility, affect the assembly of transcriptional complexes, and enhance or reduce the interaction of TFs to gene structures, resulting in altered levels of transcription and translation of downstream genes, which can trigger the development of cancer, autoimmune diseases, and even chronic diseases (Arrowsmith et al., 2012; Fatma et al., 2022; Yang J. et al., 2023). For instance, in a mouse model of LUAD, the deletion of KMT2D (a histone methyltransferase) caused decreased global levels of H3K4me1 and H3K27ac of SEs, leading to the suppression of SE-associated antioncogene Per2 (an antioncogene that inhibits glycolysis) and the activation of glycolytic processes, promoting the growth of cancer (Alam et al., 2020). Another study found that overexpression of NSD2, a histone methyltransferase, resulted in H3K36me2 spreading to the intergenic region of the low H3K27me3 region to exert protranscriptional effects (García-Carpizo et al., 2016). That is an example of long-range epigenetic remodeling. It is generally accepted that hypermethylation of CpG islands in the promoter region usually represses gene expression, whereas hypermethylation of gene bodies usually shows the opposite effects (Yang et al., 2014). Interestingly, a study found that highly expressed genes covered by SEs had lower levels of gene body methylation, which resulted in a significant negative correlation between gene body methylation and expression, contrary to popular belief (Pongor et al., 2022). These findings imply an intrinsic link between DNA modification, histone modification, and SEs that jointly influence gene expression.

2.5 SEs formation (or translocation) caused by genetic variation in lung cancer

Several studies have elucidated that the formation (or translocation) of SEs is dependent on genetic variants such as single nucleotide variants (SNVs), insertion deletions (indels), and chromosomal structural variants (SVs) (Mansour et al., 2014; Xing et al., 2019; Wang J. et al., 2023). However, the current research discovered only gene fusion or rearrangement was responsible for SEs' formation in different tissue subtypes of lung cancer. A study found that focal amplified SEs in LUAD acted on the MYC promoter to spark off transcriptional overactivation (Zhang et al., 2016). And it subsequently demonstrated that the mutations arose from a tandem duplication of a non-coding sequence. In LUAD patients, ALK or ROS1 rearrangements triggered SEs hijacking upstream of EML4 and SLC34A2, driving expression of the new oncogenic fusion genes ALK-EML4 and ROS1-SLC34A2, respectively (Yuan et al., 2021). NUT Carcinoma of the Lung is a rare lung cancer subtype of squamous origin that is extremely aggressive and highly associated with NUT gene fusions (Travis et al., 2015). The fusion gene BRD4/3-NUT co-localized with p300, and the corresponding hyperacetylated region showed a similar SE hijacking function to increase the pro-transcriptional effects of BRD4 (Eagen and French,



2021). Adenoid cystic carcinoma (ACC) originates from the secretory glands and is usually molecularly labeled by the MYB-NFIB fusion oncogene (Persson et al., 2009). MYB rearrangements are widely observed in primary pulmonary adenoid cystic carcinoma (PACC) (Roden et al., 2015). A self-regulatory feedback loop was generated when gene rearrangements (e.g., MYB -NFIB, MYB-TGFBR3, MYB-RAD51B gene fusion) occurring in ACC caused SE translocations and overexpression of MYB proteins (Drier et al., 2016).

2.6 Carcinogenic roles and regulatory mechanisms of SEs in lung cancers

Previous studies on SE-associated activation of oncogenes and signaling pathways have mainly focused on LUAD, as shown in Figure 2. A study showed that focal amplification of the SEs 3'to MYC caused increased expression of MYC and other MYC-target genes (Zhang et al., 2016). The master transcription factor ASCL1 regulates the expression of multiple SE-associated oncogenes, such as RET and MYCL (Miyashita et al., 2018). Nuclear transcription factor estrogen-related receptor alpha (ERRα) coordinates with BRD4 to drive the expression of SE- associated glycolytic gene HK2, leading to enhanced glycolysis progress and growth of malignant cells (Song et al., 2022). Specific SE-driven PADI gene families associated with tumor proliferation, invasion, and colony formation were identified as highly expressed in Osimertinib-resistant LUAD cells (Li H. et al., 2022). SE-associated CENPO was highly expressed in LUAD and positively correlated with the tumor stage, which could promote carcinogenesis and progression by regulating the cell cycle and tumor immune microenvironment (Shi et al., 2023).

As we all know, long noncoding RNAs (LncRNAs) are a class of RNAs longer than 200 nt without encoding proteins. Superenhancer lncRNAs (SE-lncRNAs) are transcribed from superenhancer genomic regions, which activate the neighboring genes or trans-activate the distant genes through several mechanisms such as transcription factor trapping, promotion of chromatin looping, histone modification, recruitment of phosphorylating RNA polymerase II (Pol II), and isolation of the transcriptional repressor (Schaukowitch et al., 2014; Alvarez-Dominguez et al., 2017; Soibam, 2017; Lee et al., 2020). FOXA1, a driver of multiple cancers (Martínez-Jiménez et al., 2020), can maintain malignant cell survival and promote progression by sensitizing the TGF- β pathway (Wei et al., 2023), activating glycolysis (Xie et al., 2022), and inhibiting autophagy (Li J. et al., 2022). FOXA1 was

reported to stimulate LncRNA DSCAM-AS1 at its SEs region (Zhang Y. et al., 2020). DSCAM-AS1, in turn, positively activates FOXA1 expression by interacting with Y box binding protein 1 (YBX1, a DNA/RNA binding protein). Silencing of DSCAM-AS1 inhibits FOXA1 expression and cell growth. Epithelialmesenchymal transition (EMT), a process by which epithelial cells acquire a mesenchymal cell phenotype and the ability to migrate and invade, is regulated by a series of TFs (Akhmetkaliyev et al., 2023). As a mediator between CDK1 and PRDX1, SE-associated lncRNA LINC00880, activated by TFs FOXP3, leads to phosphorylation of PRDX1 and dysregulation of the PTEN/AKT pathway and metastasis of LUAD (Feng Y. et al., 2023). However, as the LUAD stage progressed, the expression of LINC00880 tended to decrease, and over-expression of LINC00880 had a significant negative prognostic value only in patients with pathological stage I. SEs and related TFs such as ETS2, HNF4A, and JUNB control the high expression of some key EMT genes and define the intermediate state of the EMT process in LUAD cells (Chang et al., 2016). Besides, EMT can be mediated by classical TGF-β/SMAD pathway (Zhang et al., 2014). As one of the receptor-regulated SMAD family members, SMAD3 readily occupies the SE site of LINC01977 and drives its expression in a high TGF-B background (Zhang et al., 2022). In response, LINC01977 boosts the interaction between SMAD3 and CBP/ P300 to form a SMAD3/CBP/P300 complex targeting and activating the ZEB1 gene, which induces EMT and causes migration, proliferation, and invasion of LUAD, especially in the early stage of LUAD due to high infiltration of TAM2 (inducing a rich TGF-\beta environment) and elevated expression of SE-related LINC01977 in this stage. Notably, in p53-deficient lung cancers, the presence of the USP7 (a deubiquitinase) maintains the adherence of the SMAD3 to its own SE elements (EN4, EN9, and EN10), thereby maintaining a positive autoregulatory loop (Huang et al., 2021). However, due to the absence of the p53/MDM2 pathway, USP7 acts as a cancer suppressor. A study showed that TP63-mediated SEdriven LINC01503 was overexpressed and oncogenic in esophageal squamous carcinoma (Xie et al., 2018). Overexpression of LINC01503 was also observed in NSCLC cells and associated with proliferation, migration, and invasion, although it is unclear whether this process is SE-dependent (Shen et al., 2020). Another analysis found that SE-associated lncRNA AC074117.1 was significantly expressed and served as an independent prognostic marker in LUAD (Li et al., 2021).

2.7 Differential landscapes of SEs between LUAD and normal tissues

Several studies comparing LUAD cell lines (or cancerous tissues from patients) with normal tissues have revealed marked differences in SEs landscapes. Enrichment analysis of the respective SE-target genes may help to explain normal physiological processes and tumor carcinogenesis and progression to some extent. LUAD-specific SEregulated genes are involved in the "lung cancer" disease class and series of oncogenic pathways and are strongly associated with transcriptional disorder. In contrast, normal tissue-specific SEregulated genes are prominent in fundamental cellular structures and activities (Zhang T. et al., 2020; Zhou et al., 2020; Yuan et al., 2021). Interestingly, from one study, only GO terms (like cell-cell adhesion) with significantly lower SEs counts were observed in cancer cells, which suggested that downregulation of "normal" genes that negatively regulated carcinogenesis could also lead to cancers even when cancer-specific SEs "did not work" (Li X. et al., 2019). A bioinformatics study revealed that cancer-specific H3K27Ac peak had a higher percentage of overlap with accessible chromatin regions to favor transcription than H3K27Ac peak of normal-specific (Jiang et al., 2022).

In conclusion, cancer-specific SEs are more susceptible to other regulators and play a crucial role in determining cancer progression.

2.8 Tumor heterogeneity in SEs helping in typing of lung cancer

Dentro's whole genome sequence analysis of 2,658 cancer samples from 38 cancer types from the PCAWG project found that genomic instability allowed tumors to evolve into distinct subclonal populations with different biological behaviors among them (Dentro et al., 2021). Epigenomic diversity and instability are widely observed in heterogeneous subclonal populations of multiple types of cancer, involving differences in chromatin accessibility, methylation levels of gene structure (such as gene bodies, promoters, and enhancers of non-coding sequences), and histone profiles (Ding et al., 2019; Pastore et al., 2019; Cejas et al., 2021; El et al., 2023). The analysis of the H3K27Ac peak can directly clarify the distribution characteristics of SEs and assist in identifying associated tumor heterogeneity with them. Figure 3 illustrates the major regulators of SEs and enrichment pathways of SE-related genes in different subgroups of lung cancer.

2.9 High inter-tumor epigenetic heterogeneity presenting in NSCLC

Based on histone modification profiles, a study divided LUAD tissues into two groups that differed significantly in clinical features (e.g., histopathologic subtypes, lymph node invasion), gene mutation patterns, and prognosis (Yuan et al., 2021). Each had its own set of specific core regulatory factors and networks. Specific SE target genes in the poor prognosis group were enriched in invasion- and metastasis-related pathways (e.g., FOXM1, E2F pathway) and dedifferentiated state. In addition, core regulatory networks might shift at early stages, leading to the transition to a terminal stage. Super-enhancer RNA (SeRNA) is known to reflect SE properties and cell specificity (Chen and Liang, 2020; Tu et al., 2021). The distinct groups based on SeRNA profiles showed significant differences in immune infiltration, glycolysis levels, and prognosis (Song et al., 2022). Specifically, tissues in the worst prognostic group had the most chromosomally unstable regions, highest glucose uptake progression, hypoxia-immunotherapy response scores, the most high-risk solid subtype component, and lymph node metastases tendency. A bioinformatics analysis distinguished two subgroups of LUAD with significantly different somatic mutations, survival outcomes, and sensitivities to chemotherapeutic agents by SEs clustering analysis (Jiang et al., 2022). The transcription factor SOX2 usually cooperates with p63 to determine classical subtypes of



squamous lung cancer (Watanabe et al., 2014). However, a highly expressed lineage-specific transcription factor Brn2 (an oncogenic factor associated with neuroendocrine carcinomas) was found to replace p63 in lung squamous cell carcinoma (LUSC) cells and to synergize with SOX2 in defining a new "neural" subtype with higher aggressiveness related to poorer prognosis (Sato et al., 2019).

2.10 Fine staging of SCLC using SEs as a hot research topic presently

Investigators have found that differential expression of the TFs ASCL1, NEUROD1, POU2F3, and YAP1 defined four molecular subtypes of SCLC, respectively, and that these TFs co-localized with SEs in the corresponding SCLC subtypes (Borromeo et al., 2016; Huang YH. et al., 2018; Rudin et al., 2019). Multiple lineage-specific TFs are also present in different subtypes of SCLC, and subtype switching occurs even in the same tumor (Ireland et al., 2020; Gay et al., 2021). Focusing on the SCLC-A subtype, Karine Pozo found that the TFs ASCL1, NKX2-1, and PROX1 were associated with SEs and that their interactions formed a regulatory network to control Notch signaling (Pozo et al., 2021). On this basis, Ranran Kong subdivided SCLC-A isoforms and discovered that collaborative NKX2-1/SOX1 regulation defined a new, unique SCLC-A α subtype (Kong et al., 2022). A recent study found that the complex network of ASCL1-regulated SE-associated miRNAs

influenced the identification of SCLC molecular subtypes (Miyakawa et al., 2022). SCLC subtypes identified by TFs differ in EMT status, immune gene expression, immunotherapy (or chemotherapy) benefit, and drug resistance (Gay et al., 2021; Qu et al., 2022). Therefore, subdividing the subtypes of SCLC step by step is an important research direction, which helps us better understand its pathogenesis and develop therapeutic approaches.

2.11 Potential therapeutic targets and clinical/preclinical trials associated with SEs in lung cancer

In recent years, large-scale, individualized molecular profiling of tumors has led to the discovery of an increasing number of targets and therapeutic options to improve the clinical prognosis of patients (Corces et al., 2018; Wu et al., 2022). Due to an inadequate understanding of etiology and pathogenesis and a lack of identified therapeutic targets, more biomarker-driven trials are required to advance therapeutic technologies (William and Glisson, 2011). Transcription of SE-driven genes is highly sensitive to BRD4 inhibitors since studies reveal BETi preferentially repress transcription of SE-related genes relative to TE-related genes (Lovén et al., 2013). The TFIIH complex subunit cyclin-dependent kinase 7 (CDK7), which controls the cell cycle and initiates transcription by Pol II, has emerged as a brand-new target

Target	Compound	Phase	Status	Cancer type	NCT number
BET/BRD4	Birabresib	1	Terminated	NSCLC, NMC	NCT02698176
		1	Completed (Lewin et al., 2018)	NSCLC, NMC	NCT02259114
	PLX-2853	1	Completed	SCLC	NCT03297424
	Mivebresib	1	Completed	NSCLC, SCLC	NCT02391480
	Molibresib	2	Withdrawn	SCLC	NCT03266159
		1	Completed (Cousin et al., 2022)	SCLC, NMC	NCT01587703
	ZEN-3694	2	Recruiting	LUSC	NCT05607108
	INCB-057643	1/2	Terminated	NSCLC	NCT02959437
CDK7	Seliciclib	2	Terminated	NSCLC	NCT00372073
	SY-5609	1	Active, not recruiting	SCLC	NCT04247126
	Roniciclib	2	Terminated (Reck et al., 2019)	SCLC	NCT02161419
		1/2	Withdrawn	NSCLC	NCT02522910
		1/2	Terminated (Cho et al., 2018)	SCLC	NCT01573338
CDK9	Dinaciclib	2	Completed (Stephenson et al., 2014)	NSCLC	NCT00732810
	Hydrochloride	1	Terminated	NSCLC, SCLC	NCT00094978
	PRT-2527	1	Active, not recruiting	NSCLC	NCT05159518

TABLE 1 SE-related potential drugs in clinical trials for lung cancer.

BET, Bromodomain and extra-terminal domain; BRD4, Bromodomain-containing protein 4; CDK, Cyclin-dependent kinase; NSCLC, Non-small cell lung cancer; SCLC, small cell lung cancer; LUSC, lung squamous cell carcinoma; NMC, Nuclear protein in testis (NUT) midline carcinoma.

Agents	Combination	Mechanisms	Cancer type	Malignant features in vivo and <i>in vitro</i> experiences	References
QCA570	Osimertinib	reduction of Mcl-1	NSCLC	decreased survival, apoptosis, inhibition of formation and colonies growth	Liu et al. (2022b)
ABBV-075	Venetoclax	disruption of Bim-Bcl2 complexes, release of Bim	SCLC	apoptosis, tumor regression	Lam et al. (2017)
JQ1	ABT-263	disruption of Bim-Bcl2 complexes, release of Bim	SCLC	apoptosis	Wang et al. (2017)
GSK525762 (I-BET762)	Talazoparib	inhibition of DNA damage response, HR-DSBR process	SCLC	reduced survival and growth	Fiorentino et al. (2020)
JQ1	Paclitaxel	downregulation of BET protein levels	NSCLC	apoptosis, inhibition of protective autophagy	Zhou et al. (2021)
JQ1, AZD5153, NHWD870	Rapamycin, Everolimus	block of TSC2-mTOR-p70S6K1-BAD pathway	SCLC	intrinsic apoptosis	Kumari et al. (2023)
JQ1	ACY-1215	upregulation of MHC II on tumor cells relies on NK cells	SCLC	apoptosis	Liu et al. (2018)
JQ1, AZD5153	AZD1775	inhibition of NHEJ activity, enhancement of DNA damage, promotion of mitotic entry and mitotic catastrophe	NSCLC	synergistic cytotoxicity, apoptosis, suppression of proliferation	Takashima et al. (2020)

TABLE 2 Combination therapeutic regimens related to SEs in pre-clinical trials for lung cancer.

NSCLC, Non-small cell lung cancer; SCLC, small cell lung cancer.

for treating cancer (Larochelle et al., 2012; Suski et al., 2021). Plus, the catalytic member of the P-TEFb complex, cyclin-dependent kinase 9 (CDK9), promotes the transcriptional elongation of Pol II (Zhou et al., 2012). CDK7 can also phosphorylate to activate CDK9 (Larochelle et al., 2012). Therefore, BET, CDK7, and CDK9 may

represent viable therapeutic targets for focusing on SE-dependent transcription. Table 1 demonstrates the clinical trials currently underway or have concluded. Besides, a series of preclinical studies of different drug combinations have laid the groundwork for more novel combination therapeutic regimens to enter clinical

trials, as shown in Table 2. We provide a brief overview of the results reported.

2.11.1 BETi

The BET protein family member BRD4 is currently the focus of much research. We previously discussed the significance of the firstgeneration BRD4 inhibitor-JQ1 in the discovery and targeted therapy of SEs. In addition to JQ1, more and more smallmolecule drugs targeting BET have been prepared and widely employed in clinical trials (Filippakopoulos et al., 2010).

Birabresib (OTX015, MK-8628), a small molecule complex, was first used in hematologic malignancies. In LUAD cells, the master transcriptional regulators ELF3, EHF, and TGIF1 are downregulated in response to JQ1 and OTX015, causing disruption of CRC structure and inhibition of cancer cells' malignant progression (Zhang T. et al., 2020).

A Phase I study was designed to evaluate the safety and efficacy of single-agent Birabresib in a range of advanced solid tumors (Lewin et al., 2018). Encouragingly, of the nine NSCLC patients with ALK rearrangement or KRAS mutation, seven developed stable disease (SD), and two developed progressive disease (PD). Though thrombocytopenia was one of the serious adverse effects, it was reversible and self-limiting. Another phase 1/2 study assessed the effectiveness and safety of incrementally applied single-agent Molibresib (Cousin et al., 2022). Of the twelve patients with evaluable SCLC, three developed SD, and nine developed PD. However, no clinically meaningful response criteria were observed, as many patients discontinued dose escalation or even reduced the dose due to drug toxicity. The most common treatmentrelated adverse events were thrombocytopenia, nausea, and decreased appetite.

The novel BRD4-targeting agent CFT-2718 showed potent antitumor activity in xenograft mice. The intensity and duration of apoptosis induction and tumor proliferation inhibition by CFT-2718 surpassed that of the CDK kinase inhibitor dinaciclib in SCLC cells (Sun et al., 2021). OTX015 showed significant anti-cancer activity in NSCLC cells with or without EML4-ALK translocation and EGFR, KRAS mutations (Riveiro et al., 2016). Meanwhile, *in vivo* assays demonstrated that OTX015 significantly inhibited the growth of EML4-ALK (+) NSCLC.

2.11.2 CDK7/9 inhibitors

THZ1 inhibits CDK7 activity in an irreversible covalent binding manner (Kwiatkowski et al., 2014). A study found that both SCLC cell lines and mouse models exhibited high sensitivity to THZ1, as evidenced by considerable tumor proliferation inhibition (Christensen et al., 2014). Genes regulated by master-regulated TFs E2F, NRF1, and CREB are preferentially repressed by THZ1 in SCLC cells, suggesting selective effects of THZ1 on SEs and transcriptional core circuits (Christensen et al., 2014), as in LUAD (Zhang T. et al., 2020), neuroblastoma (Chipumuro et al., 2014), and triple-negative breast cancer (Wang et al., 2015). On the other hand, the low sensitivity and specificity of pan-CDK inhibitors for SCLC demonstrate the significance of covalent binding mechanisms in therapeutic efficacy (Christensen et al., 2014). Since the complex formed by BRD4 binding to CDK9/P-TEFb is localized in the SEs region and stimulates the transcriptional activation of MYC, synergistic inhibition of these two targets can effectively halt the progression of cancer (Lovén et al., 2013; Lu et al., 2015).

A phase 2 study compared the difference in efficacy between dinaciclib (a CDK9 inhibitor) and erlotinib in previously treated NSCLC patients (Stephenson et al., 2014). Unfortunately, no patients in the dinaciclib group showed objective responses (OR). The main adverse effects of intravenous denazanil were myelosuppression and gastrointestinal toxicity.

YPN-005 is a novel and selective CDK7 inhibitor with super anti-cancer activity compared to conventional THZ1. YPN-005 showed powerful antiproliferative effects in chemotherapyresistant SCLC cells, patient-derived organoids, and xenograft mice (Choi et al., 2021). No significant toxic reactions were observed in the mice model. 21e is a highly effective and selective CDK9 inhibitor. After receiving 21e, the proportion of NSCLC side population cells (which indicate stem cell potential) drastically decreased (Wang et al., 2019). A reduction of the cancer stemness marker Oct4, apoptosis of cancer cells, and inhibition of proliferation were observed in xenograft mice treated with 21e.

2.11.3 Combination therapy

It is evident that existing clinical trials, although affirming the therapeutic feasibility of BETi and CDK kinase inhibitors, still suffer from ineffectiveness, inefficacy, toxicity, and resistance. Single-agent (or even partial combination) treatment in these trials does not meet the desired standard. Future studies may favor multiple combination strategies to enhance therapeutic efficacy and reduce toxicity.

Roniclib is a pan-CDK inhibitor (including both CDK7 and CDK9). A phase I/II study reported the potential feasibility of Roniclib in combination with cisplatin/carboplatin-etoposide for extensive SCLC (Cho et al., 2018). Chemotherapeutic agents did not alter the pharmacokinetics of Roniclib but reduced Roniclib exposure by more than one-third. Although none of the 46 patients receiving different doses achieved a complete response (CR), 35 achieved a partial response (PR), five achieved SD, and two developed PD. The most common adverse reactions were nausea, vomiting, anemia, and decreased neutrophil count. This study reflects the favorable efficacy and tolerability of Roniclib for SCLC in all. However, another phase II study comparing the efficacy and safety of Roniciclib in combination with platinumbased chemotherapy versus chemotherapy alone in patients with extensive SCLC did not give us encouraging results (Reck et al., 2019). Although one patient in the combination therapy group achieved CR, overall, the combination therapy group had a lower objective response rate (ORR) and more treatment-emergent events than the platinum-based chemotherapy adverse group. Common adverse reactions were similar to the previous study (Cho et al., 2018). Nausea was the primary cause of discontinuation in the combined therapy group.

Besides, BET- and CDK-targeted inhibitors demonstrate convincing anti-tumor effects in preclinical models and provide paradigms for clinical trials.

BETi can induce cancer cells toward apoptosis by regulating the balance between pro- and anti-apoptotic proteins. QCA570 is a novel BET degrader that caused the degradation of anti-apoptosis-related proteins Mcl-1 in NSCLC cells (Liu C. et al., 2022). QCA570, in combination with ositinib, induced apoptosis and inhibited the proliferation of ositinib-resistant cells in vivo and *in vitro*

experiments. Another study found that the antitumoral activity of the BET inhibitor ABBV-075 in SCLC was partially dependent on the inhibition of BCL2, an anti-apoptotic protein (Lam et al., 2017). In vivo and in vitro assays confirmed the synergistic antitumor effects of ABBV-075 with the BCL2 inhibitor venetoclax (ABT-199), particularly in tumors with a high level of BCL2 expression. Furthermore, combination treatment with JQ1 and the BCL2 inhibitors ABT-263 damaged the balance of Bim and Bcl-2, resulting in the release of the pro-apoptotic protein BIM in MYCN-amplified SCLC cell lines and exerted growth-inhibitory effects in MYCN-positive xenografts (Wang et al., 2017). Another study similarly found that BET inhibitor I-BET762 combined with the PARP inhibitor talazoparib showed higher antitumoral activity in SCLC cells with elevated MYC expression or amplification of MYCs (Fiorentino et al., 2020). The above research implies that the combined strategy boosts the inhibitory effects of BETi on the MYCrelated transcriptional pathway.

BET inhibitor treatment alone may result in compensatory alterations that can lower efficacy or develop drug resistance. Therefore, inhibiting these compensatory changes can improve efficacy. For instance, one investigation discovered that JQ1 alone compensatorily upregulated the BET protein (BRD2,3,4), triggered autophagy, and prevented apoptosis in NSCLC cells (Zhou et al., 2021). JQ1 with the chemotherapeutic agent paclitaxel reversed this upregulation and induced apoptosis. Another study found that BETi upregulate the mTOR upstream kinase RSK3, leading to resistance to apoptosis in SCLC cells (Kumari et al., 2023). The combination of BETi and mTOR inhibitors disrupted mTOR signaling and displayed superior anti-cancer activity than any single drug in vitro (JQ1/rapamycin) and in vivo (NHWD870/ everolimus) assays.

A combination of inhibitors targeting epigenetic regulators histone deacetylase (HDAC) and BET exerted synergistic antitumoral effects in pancreatic cancer (He et al., 2020), glioblastoma (Gusyatiner et al., 2021), and medulloblastoma (Kling et al., 2022). In SCLC xenograft mice, JQ1 and the histone deacetylase 6 (HDAC6) inhibitor ACY-1215 combined to exert antitumor activity dependent on the presence of NK cells (Liu et al., 2018).

Furthermore, JQ1 enhanced DNA double-strand breaks induced by the WEE1 inhibitor AZD1775 and led to mitotic catastrophe, which synergistically produced cytotoxicity and inhibited NSCLC growth in vitro and *in vivo* experiments (Takashima et al., 2020).

In conclusion, in both *in vivo* and *in vitro* studies, whether for SCLC or NSCLC, combination therapies have achieved greater success than any single agent. Combination therapies are also effective in reducing drug resistance and side effects. Based on in-depth studies of more complex carcinogenic pathways, we suppose that more clinical trials will be undertaken in the future.

2.11.4 Other drug candidates in the pancancer field

Furthermore, epigenetic modifiers CBP/p300 and HDAC dynamically regulate H3K27 acetylation levels and are potential drug targets in the pan-cancer field (Wang et al., 2022).

CBP/p300 is a histone acetyltransferase that deposits extensive H3K27ac marks and highly occupies SEs (Wang et al., 2022). A study found that the CBP/p300 inhibitor ICG-001 alone upregulated

the expression of SE-related oncogenes in glioblastomas, whereas this deleterious effect disappeared when combined with JQ1 (Wiese et al., 2020). Anti-cancer studies with SE-related CBP/ p300 inhibitors have been conducted in hematologic malignant neoplasms such as multiple myeloma, acute myeloid leukemia, and non-Hodgkin's lymphoma (Lasko et al., 2017), as well as in castration-resistant prostate cancer (Chen et al., 2022). Notably, Zhang T et al. found that the EP300 inhibitor CBP30 inhibited the expression of master TFs in LUAD cell lines (Zhang T. et al., 2020).

Studies have confirmed that HDAC inhibitors (HDACi) effectively inhibit the function of SEs (Nagaraja et al., 2017; Sanchez et al., 2018; Nguyen et al., 2020). HDACi targeting SEs exerted great anti-tumor potential in glioblastoma (Nguyen et al., 2020), rhabdomyosarcoma (Gryder et al., 2019), multiple myeloma (Alaterre et al., 2022), stem-like breast cancer (Caslini et al., 2019), ovarian cancer (Shi et al., 2019; Wu et al., 2023), and esophageal squamous cell carcinoma (Jiang et al., 2020). Numerous preclinical studies have confirmed that HDACi inhibits the proliferation of lung cancer cells and is strongly associated with drug resistance (Huang et al., 2014; Wu et al., 2020; He et al., 2022; Eichner et al., 2023). However, it is not yet clear whether this effect is dependent on SEs. Unfortunately, in clinical trials for lung cancer, HDACi are usually used in synergy with radiotherapy, immunotherapy, or epigenetic therapy, as only modest single-agent activity has been demonstrated (Bartling et al., 2005; Traynor et al., 2009; Costa et al., 2014). In any case, drug trials against more SE targets are warranted and expected to provide additional treatment options for lung cancer patients.

3 Conclusion

Since their discovery, SEs have received attention in various fields, and the specific functions they exert in different tumor types are gradually being revealed. The basic structure and function of SEs are similar in different malignant tumors, and all are closely related to cancer progression and aggressiveness. However, it is still to be demonstrated that there are many differences in the SEs characteristics of different cancer types: 1. SEs might be widespread in almost all cancer MYC regions (Zhou and Parsons, 2023), whereas studies of SE changes in the MYC region in lung cancer are limited (Zhang et al., 2016). 2. Gene rearrangements/fusions underlie the formation of cancer-specific SEs. However, compared to hematologic tumors, where highfrequency gene fusions are present, rearrangements/fusions that form SEs are less frequently observed in lung cancer (especially squamous lung cancer) (Li et al., 2016). 3. SE-lncRNA might be widespread in pan-cancer (Ropri et al., 2021; Yan et al., 2021; Chuang et al., 2022; Yang Z. et al., 2023; Hu et al., 2023) SE-IncRNA has been intensively studied in lung cancer (Xie et al., 2018; Shen et al., 2020; Zhang et al., 2022), hepatocellular carcinoma (Peng et al., 2019; Li et al., 2023; Su et al., 2023; Yuan et al., 2023), and gliomas (Bian et al., 2021; Yang Z. et al., 2023; Chen et al., 2023) but has been rarely reported in hematologic tumors (Handa et al., 2020). However, SEs still cannot be accurately identified due to the low specificity of the classical ChIP-seq technique and the technical imperfections of the Rose algorithm. In addition, the molecular mechanism of how oncogenic signaling alters super enhancerpromoter interactions is not yet understood. For example, how

10.3389/fphar.2024.1383580

do oncogenic signals promote the aggregation of phase-separated condensates near SEs and drive transcription? How do these signals reshape the three-dimensional folding of the genome, altering the otherwise insulating relationships between regulatory elements and pinpointing the timing, location, and level of transcription?

Here, we summarize the tissue specificity of SEs, mechanisms of SEs formation, oncogenic effects, therapeutic targets, and clinical trials in lung cancer.

Researchers identified subgroups of lung tumors with distinctly diverse biological traits (such as aggressiveness) and prognoses using the Rose algorithms and unsupervised clustering, indicating that these groupings have unique oncogenic signaling networks. However, to date, SEs formed by point mutations or indels have not been identified in the lung cancer genome. Moreover, it is difficult to determine the sequential relationship between the activation of oncogenic pathways and the formation of SEs. Unfortunately, only a few studies related to SEs were conducted for squamous lung cancer. It is challengeable and valuable for researchers to accurately identify SE-driven oncogenic mutations of squamous cell carcinoma, given the close connection between smoking and squamous cell carcinoma and the large number of nononcogenic passenger mutations resulting from long-term exposure to tobacco carcinogens. BETi and CDK kinase inhibitors are the predominant epigenetic agents targeting SEs. However, existing clinical trials of these drugs are less encouraging due to lack of specificity, development of resistance, and the inevitability of toxicity. Nonetheless, many studies in preclinical models have confirmed the potential benefits of combination therapies and suggested that drug combinations with multiple different mechanisms are an area to explore in the future. It is worth mentioning that the body's immunological status and the tumor microenvironment are unquestionably significant factors influencing medication action, even though the precise mechanism is still under investigation. Epigenetic therapies can remodel the immune microenvironment, and their use in conjunction with immunosuppressive agents is currently a hot topic in cancer care (Chou et al., 2020; Luo et al., 2022; Tien et al., 2023). In addition to drug therapy, gene editing techniques (e.g., CRISPR/Cas9) can specifically knock down SEs of interest to

References

Adam, R. C., Yang, H., Ge, Y., Infarinato, N. R., Gur-Cohen, S., Miao, Y., et al. (2020). NFI transcription factors provide chromatin access to maintain stem cell identity while preventing unintended lineage fate choices. *Nat. Cell Biol.* 22 (6), 640–650. doi:10.1038/s41556-020-0513-0

Akhmetkaliyev, A., Alibrahim, N., Shafiee, D., and Tulchinsky, E. (2023). EMT/MET plasticity in cancer and Go-or-Grow decisions in quiescence: the two sides of the same coin? *Mol. Cancer* 22 (1), 90. doi:10.1186/s12943-023-01793-z

Alam, H., Tang, M., Maitituoheti, M., Dhar, S. S., Kumar, M., Han, C. Y., et al. (2020). KMT2D deficiency impairs super-enhancers to confer a glycolytic vulnerability in lung cancer. *Cancer Cell* 37 (4), 599–617. doi:10.1016/j.ccell.2020.03.005

Alaterre, E., Ovejero, S., Herviou, L., de Boussac, H., Papadopoulos, G., Kulis, M., et al. (2022). Comprehensive characterization of the epigenetic landscape in Multiple Myeloma. *Theranostics* 12 (4), 1715–1729. doi:10.7150/thno.54453

Alvarez-Dominguez, J. R., Knoll, M., Gromatzky, A. A., and Lodish, H. F. (2017). The super-enhancer-derived alncRNA-EC7/bloodlinc potentiates red blood cell development in trans. *Cell Rep.* 19 (12), 2503–2514. doi:10.1016/j.celrep.2017.05.082

Arrowsmith, C. H., Bountra, C., Fish, P. V., Lee, K., and Schapira, M. (2012). Epigenetic protein families: a new frontier for drug discovery. *Nat. Rev. Drug Discov.* 11 (5), 384–400. doi:10.1038/nrd3674

prevent activation and cascade amplification of oncogenic transcripts. For instance, the knockdown of e3, one of the key components of SEs in the neighboring non-coding region of MYC, downregulates the expression of MYC and downstream targets and inhibits the clonal growth of LUAD cells (Zhang et al., 2016). Therefore, pinpointing functional components within SEs clusters may provide insight into precision medicine. In conclusion, further research into the oncogenic mechanism of SEs will aid in the development of personalized therapeutic strategies and the long-term survival of lung cancer patients.

Author contributions

ZY: Conceptualization, Writing-original draft, Writing-review and editing. PS: Conceptualization, Supervision, Writing-review and editing. WJ: Supervision, Writing-review and editing.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Banerji, J., Rusconi, S., and Schaffner, W. (1981). Expression of a beta-globin gene is enhanced by remote SV40 DNA sequences. *Cell* 27 (2 Pt 1), 299–308. doi:10.1016/0092-8674(81)90413-x

Barakat, T. S., Halbritter, F., Zhang, M., Rendeiro, A. F., Perenthaler, E., Bock, C., et al. (2018). Functional dissection of the enhancer repertoire in human embryonic stem cells. *Cell Stem Cell* 23 (2), 276–288. doi:10.1016/j.stem.2018.06.014

Barral, A., and Déjardin, J. (2023). The chromatin signatures of enhancers and their dynamic regulation. *Nucleus* 14 (1), 2160551. doi:10.1080/19491034.2022.2160551

Bartling, B., Hofmann, H. S., Boettger, T., Hansen, G., Burdach, S., Silber, R. E., et al. (2005). Comparative application of antibody and gene array for expression profiling in human squamous cell lung carcinoma. *Lung Cancer* 49 (2), 145–154. doi:10.1016/j. lungcan.2005.02.006

Bian, E., Chen, X., Cheng, L., Cheng, M., Chen, Z., Yue, X., et al. (2021). Superenhancer-associated TMEM44-AS1 aggravated glioma progression by forming a positive feedback loop with Myc. *J. Exp. Clin. Cancer Res.* 40 (1), 337. doi:10.1186/ s13046-021-02129-9

Blackwood, E. M., and Kadonaga, J. T. (1998). Going the distance: a current view of enhancer action. *Science* 281 (5373), 60–63. doi:10.1126/science.281.5373.60

Bonazzoli, E., Predolini, F., Cocco, E., Bellone, S., Altwerger, G., Menderes, G., et al. (2018). Inhibition of BET bromodomain proteins with GS-5829 and GS-626510 in uterine serous carcinoma, a biologically aggressive variant of endometrial cancer. *Clin. Cancer Res.* 24 (19), 4845–4853. doi:10.1158/1078-0432.Ccr-18-0864

Borromeo, M. D., Savage, T. K., Kollipara, R. K., He, M., Augustyn, A., Osborne, J. K., et al. (2016). ASCL1 and NEUROD1 reveal heterogeneity in pulmonary neuroendocrine tumors and regulate distinct genetic programs. *Cell Rep.* 16 (5), 1259–1272. doi:10.1016/j.celrep.2016.06.081

Cao, Z., Shu, Y., Wang, J., Wang, C., Feng, T., Yang, L., et al. (2022). Super enhancers: pathogenic roles and potential therapeutic targets for acute myeloid leukemia (AML). *Genes. Dis.* 9 (6), 1466–1477. doi:10.1016/j.gendis.2022.01.006

Carroll, P. A., Freie, B. W., Mathsyaraja, H., and Eisenman, R. N. (2018). The MYC transcription factor network: balancing metabolism, proliferation and oncogenesis. *Front. Med.* 12 (4), 412–425. doi:10.1007/s11684-018-0650-z

Caslini, C., Hong, S., Ban, Y. J., Chen, X. S., and Ince, T. A. (2019). HDAC7 regulates histone 3 lysine 27 acetylation and transcriptional activity at super-enhancer-associated genes in breast cancer stem cells. *Oncogene* 38 (39), 6599–6614. doi:10.1038/s41388-019-0897-0

Cejas, P., Xie, Y., Font-Tello, A., Lim, K., Syamala, S., Qiu, X., et al. (2021). Subtype heterogeneity and epigenetic convergence in neuroendocrine prostate cancer. *Nat. Commun.* 12 (1), 5775. doi:10.1038/s41467-021-26042-z

Chang, H., Liu, Y., Xue, M., Liu, H., Du, S., Zhang, L., et al. (2016). Synergistic action of master transcription factors controls epithelial-to-mesenchymal transition. *Nucleic Acids Res.* 44 (6), 2514–2527. doi:10.1093/nar/gkw126

Chen, C., Zhou, D., Gu, Y., Wang, C., Zhang, M., Lin, X., et al. (2020). SEA version 3.0: a comprehensive extension and update of the Super-Enhancer archive. *Nucleic Acids Res.* 48 (D1), D198-D203–d203. doi:10.1093/nar/gkz1028

Chen, H., and Liang, H. (2020). A high-resolution map of human enhancer RNA loci characterizes super-enhancer activities in cancer. *Cancer Cell* 38 (5), 701–715. doi:10. 1016/j.ccell.2020.08.020

Chen, Q., Yang, B., Liu, X., Zhang, X. D., Zhang, L., and Liu, T. (2022). Histone acetyltransferases CBP/p300 in tumorigenesis and CBP/p300 inhibitors as promising novel anticancer agents. *Theranostics* 12 (11), 4935–4948. doi:10.7150/thno.73223

Chen, Z., Tian, D., Chen, X., Cheng, M., Xie, H., Zhao, J., et al. (2023). Superenhancer-driven lncRNA LIMD1-AS1 activated by CDK7 promotes glioma progression. *Cell Death Dis.* 14 (6), 383. doi:10.1038/s41419-023-05892-z

Chipumuro, E., Marco, E., Christensen, C. L., Kwiatkowski, N., Zhang, T., Hatheway, C. M., et al. (2014). CDK7 inhibition suppresses super-enhancer-linked oncogenic transcription in MYCN-driven cancer. *Cell* 159 (5), 1126–1139. doi:10.1016/j.cell.2014.10.024

Cho, B. C., Dy, G. K., Govindan, R., Kim, D. W., Pennell, N. A., Zalcman, G., et al. (2018). Phase Ib/II study of the pan-cyclin-dependent kinase inhibitor roniciclib in combination with chemotherapy in patients with extensive-disease small-cell lung cancer. *Lung Cancer* 123, 14–21. doi:10.1016/j.lungcan.2018.04.022

Choi, Y. J., Lee, H., Kim, D. S., Kim, D. H., Kang, M. H., Cho, Y. H., et al. (2021). Discovery of a novel CDK7 inhibitor YPN-005 in small cell lung cancer. *Eur. J. Pharmacol.* 907, 174298. doi:10.1016/j.ejphar.2021.174298

Chou, J., Quigley, D. A., Robinson, T. M., Feng, F. Y., and Ashworth, A. (2020). Transcription-associated cyclin-dependent kinases as targets and biomarkers for cancer therapy. *Cancer Discov.* 10 (3), 351–370. doi:10.1158/2159-8290.Cd-19-0528

Christensen, C. L., Kwiatkowski, N., Abraham, B. J., Carretero, J., Al-Shahrour, F., Zhang, T., et al. (2014). Targeting transcriptional addictions in small cell lung cancer with a covalent CDK7 inhibitor. *Cancer Cell* 26 (6), 909–922. doi:10.1016/j.ccell.2014. 10.019

Chuang, T. D., Quintanilla, D., Boos, D., and Khorram, O. (2022). Differential expression of super-enhancer-associated long non-coding RNAs in uterine leiomyomas. *Reprod. Sci.* 29 (10), 2960–2976. doi:10.1007/s43032-022-00981-4

Corces, M. R., Granja, J. M., Shams, S., Louie, B. H., Seoane, J. A., Zhou, W., et al. (2018). The chromatin accessibility landscape of primary human cancers. *Science* 362 (6413), eaav1898. doi:10.1126/science.aav1898

Costa, C., Molina, M. A., Drozdowskyj, A., Giménez-Capitán, A., Bertran-Alamillo, J., Karachaliou, N., et al. (2014). The impact of EGFR T790M mutations and BIM mRNA expression on outcome in patients with EGFR-mutant NSCLC treated with erlotinib or chemotherapy in the randomized phase III EURTAC trial. *Clin. Cancer Res.* 20 (7), 2001–2010. doi:10.1158/1078-0432.Ccr-13-2233

Cousin, S., Blay, J. Y., Garcia, I. B., de Bono, J. S., Le Tourneau, C., Moreno, V., et al. (2022). Safety, pharmacokinetic, pharmacodynamic and clinical activity of molibresib for the treatment of nuclear protein in testis carcinoma and other cancers: results of a Phase I/II open-label, dose escalation study. *Int. J. Cancer* 150 (6), 993–1006. doi:10. 1002/ijc.33861

Das, N. D., Chang, J. C., Hon, C. C., Kelly, S. T., Ito, S., Lizio, M., et al. (2023). Defining super-enhancers by highly ranked histone H4 multi-acetylation levels identifies transcription factors associated with glioblastoma stem-like properties. *BMC Genomics* 24 (1), 574. doi:10.1186/s12864-023-09659-w

Dentro, S. C., Leshchiner, I., Haase, K., Tarabichi, M., Wintersinger, J., Deshwar, A. G., et al. (2021). Characterizing genetic intra-tumor heterogeneity across 2,658 human cancer genomes. *Cell* 184 (8), 2239–2254.e39. doi:10.1016/j.cell.2021.03.009

Ding, X., He, M., Chan, A. W. H., Song, Q. X., Sze, S. C., Chen, H., et al. (2019). Genomic and epigenomic features of primary and recurrent hepatocellular carcinomas. *Gastroenterology* 157 (6), 1630–1645. doi:10.1053/j.gastro.2019. 09.005

Dong, Y., Hu, H., Zhang, X., Zhang, Y., Sun, X., Wang, H., et al. (2023). Phosphorylation of PHF2 by AMPK releases the repressive H3K9me2 and inhibits cancer metastasis. *Signal Transduct. Target Ther.* 8 (1), 95. doi:10.1038/s41392-022-01302-6

Dowen, J. M., Fan, Z. P., Hnisz, D., Ren, G., Abraham, B. J., Zhang, L. N., et al. (2014). Control of cell identity genes occurs in insulated neighborhoods in mammalian chromosomes. *Cell* 159 (2), 374–387. doi:10.1016/j.cell.2014.09.030

Drier, Y., Cotton, M. J., Williamson, K. E., Gillespie, S. M., Ryan, R. J., Kluk, M. J., et al. (2016). An oncogenic MYB feedback loop drives alternate cell fates in adenoid cystic carcinoma. *Nat. Genet.* 48 (3), 265–272. doi:10.1038/ng.3502

Eagen, K. P., and French, C. A. (2021). Supercharging BRD4 with NUT in carcinoma. Oncogene 40 (8), 1396–1408. doi:10.1038/s41388-020-01625-0

Eichner, L. J., Curtis, S. D., Brun, S. N., McGuire, C. K., Gushterova, I., Baumgart, J. T., et al. (2023). HDAC3 is critical in tumor development and therapeutic resistance in Kras-mutant non-small cell lung cancer. *Sci. Adv.* 9 (11), eadd3243. doi:10.1126/sciadv. add3243

El, K. L. Y., Pan, X., Hlady, R. A., Wagner, R. T., Shaikh, S., Wang, L., et al. (2023). Extensive intratumor regional epigenetic heterogeneity in clear cell renal cell carcinoma targets kidney enhancers and is associated with poor outcome. *Clin. Epigenetics* 15 (1), 71. doi:10.1186/s13148-023-01471-3

Fatma, H., Maurya, S. K., and Siddique, H. R. (2022). Epigenetic modifications of c-MYC: role in cancer cell reprogramming, progression and chemoresistance. *Semin. Cancer Biol.* 83, 166–176. doi:10.1016/j.semcancer.2020.11.008

Feng, C., Song, C., Jiang, Y., Zhao, J., Zhang, J., Wang, Y., et al. (2023a). Landscape and significance of human super enhancer-driven core transcription regulatory circuitry. *Mol. Ther. Nucleic Acids* 32, 385–401. doi:10.1016/j.omtn.2023.03.014

Feng, Y., Zhang, T., Zhang, Z., Liang, Y., Wang, H., Chen, Y., et al. (2023b). The superenhancer-driven lncRNA LINC00880 acts as a scaffold between CDK1 and PRDX1 to sustain the malignance of lung adenocarcinoma. *Cell Death Dis.* 14 (8), 551. doi:10. 1038/s41419-023-06047-w

Filippakopoulos, P., Qi, J., Picaud, S., Shen, Y., Smith, W. B., Fedorov, O., et al. (2010). Selective inhibition of BET bromodomains. *Nature* 468 (7327), 1067–1073. doi:10.1038/ nature09504

Fiorentino, F. P., Marchesi, I., Schröder, C., Schmidt, R., Yokota, J., and Bagella, L. (2020). BET-inhibitor I-BET762 and PARP-inhibitor talazoparib synergy in small cell lung cancer cells. *Int. J. Mol. Sci.* 21 (24), 9595. doi:10.3390/ijms21249595

Fontanals-Cirera, B., Hasson, D., Vardabasso, C., Di Micco, R., Agrawal, P., Chowdhury, A., et al. (2017). Harnessing BET inhibitor sensitivity reveals AMIGO2 as a melanoma survival gene. *Mol. Cell* 68 (4), 731–744. doi:10.1016/j. molcel.2017.11.004

Frese, K. K., Simpson, K. L., and Dive, C. (2021). Small cell lung cancer enters the era of precision medicine. *Cancer Cell* 39 (3), 297–299. doi:10.1016/j.ccell.2021.02.002

García-Carpizo, V., Sarmentero, J., Han, B., Graña, O., Ruiz-Llorente, S., Pisano, D. G., et al. (2016). NSD2 contributes to oncogenic RAS-driven transcription in lung cancer cells through long-range epigenetic activation. *Sci. Rep.* 6, 32952. doi:10.1038/ srep32952

Gay, C. M., Stewart, C. A., Park, E. M., Diao, L., Groves, S. M., Heeke, S., et al. (2021). Patterns of transcription factor programs and immune pathway activation define four major subtypes of SCLC with distinct therapeutic vulnerabilities. *Cancer Cell* 39 (3), 346–360.e7. doi:10.1016/j.ccell.2020.12.014

Gryder, B. E., Pomella, S., Sayers, C., Wu, X. S., Song, Y., Chiarella, A. M., et al. (2019). Histone hyperacetylation disrupts core gene regulatory architecture in rhabdomyosarcoma. *Nat. Genet.* 51 (12), 1714–1722. doi:10.1038/s41588-019-0534-4

Gusyatiner, O., Bady, P., Pham, M. D. T., Lei, Y., Park, J., Daniel, R. T., et al. (2021). BET inhibitors repress expression of interferon-stimulated genes and synergize with HDAC inhibitors in glioblastoma. *Neuro Oncol.* 23 (10), 1680–1692. doi:10.1093/ neuonc/noab115

Hah, N., Benner, C., Chong, L. W., Yu, R. T., Downes, M., and Evans, R. M. (2015). Inflammation-sensitive super enhancers form domains of coordinately regulated enhancer RNAs. *Proc. Natl. Acad. Sci. U. S. A.* 112 (3), E297–E302. doi:10.1073/ pnas.1424028112

Handa, H., Honma, K., Oda, T., Kobayashi, N., Kuroda, Y., Kimura-Masuda, K., et al. (2020). Long noncoding RNA PVT1 is regulated by bromodomain protein BRD4 in multiple myeloma and is associated with disease progression. *Int. J. Mol. Sci.* 21 (19), 7121. doi:10.3390/ijms21197121

He, S., Dong, G., Li, Y., Wu, S., Wang, W., and Sheng, C. (2020). Potent dual BET/ HDAC inhibitors for efficient treatment of pancreatic cancer. *Angew. Chem. Int. Ed. Engl.* 59 (8), 3028–3032. doi:10.1002/anie.201915896

He, T., Gao, Y., Fang, Y., Zhang, Y., Zhang, S., Nan, F., et al. (2022). The HDAC inhibitor GCJ-490A suppresses c-Met expression through IKKa and overcomes gefitinib resistance in non-small cell lung cancer. *Cancer Biol. Med.* 19 (8), 1172–1192. doi:10.20892/j.issn.2095-3941.2021.0130

Hnisz, D., Abraham, B. J., Lee, T. I., Lau, A., Saint-André, V., Sigova, A. A., et al. (2013). Super-enhancers in the control of cell identity and disease. *Cell* 155 (4), 934–947. doi:10.1016/j.cell.2013.09.053

Hu, X., Wu, J., Feng, Y., Ma, H., Zhang, E., Zhang, C., et al. (2023). METTL3-stabilized super enhancers-IncRNA SUCLG2-AS1 mediates the formation of a long-range chromatin loop between enhancers and promoters of SOX2 in metastasis and radiosensitivity of nasopharyngeal carcinoma. *Clin. Transl. Med.* 13 (9), e1361. doi:10.1002/ctm2.1361

Huang, J., Li, K., Cai, W., Liu, X., Zhang, Y., Orkin, S. H., et al. (2018a). Dissecting super-enhancer hierarchy based on chromatin interactions. *Nat. Commun.* 9 (1), 943. doi:10.1038/s41467-018-03279-9

Huang, W. J., Tang, Y. A., Chen, M. Y., Wang, Y. J., Hu, F. H., Wang, T. W., et al. (2014). A histone deacetylase inhibitor YCW1 with antitumor and antimetastasis properties enhances cisplatin activity against non-small cell lung cancer in preclinical studies. *Cancer Lett.* 346 (1), 84–93. doi:10.1016/j.canlet.2013.12.016

Huang, Y. H., Klingbeil, O., He, X. Y., Wu, X. S., Arun, G., Lu, B., et al. (2018b). POU2F3 is a master regulator of a tuft cell-like variant of small cell lung cancer. *Genes. Dev.* 32 (13-14), 915–928. doi:10.1101/gad.314815.118

Huang, Y. T., Cheng, A. C., Tang, H. C., Huang, G. C., Cai, L., Lin, T. H., et al. (2021). USP7 facilitates SMAD3 autoregulation to repress cancer progression in p53-deficient lung cancer. *Cell Death Dis.* 12 (10), 880. doi:10.1038/s41419-021-04176-8

Ireland, A. S., Micinski, A. M., Kastner, D. W., Guo, B., Wait, S. J., Spainhower, K. B., et al. (2020). MYC drives temporal evolution of small cell lung cancer subtypes by reprogramming neuroendocrine fate. *Cancer Cell* 38 (1), 60–78. doi:10.1016/j.ccell. 2020.05.001

Jang, J. E., Eom, J. I., Jeung, H. K., Cheong, J. W., Lee, J. Y., Kim, J. S., et al. (2017). Targeting AMPK-ULK1-mediated autophagy for combating BET inhibitor resistance in acute myeloid leukemia stem cells. *Autophagy* 13 (4), 761–762. doi:10.1080/15548627. 2016.1278328

Jiang, X., Qin, N., Hua, T., Wei, X., Li, Y., Chen, C., et al. (2022). Functional characterization and clinical significance of super-enhancers in lung adenocarcinoma. *Mol. Carcinog.* 61 (8), 776–786. doi:10.1002/mc.23419

Jiang, Y., Qian, F., Bai, X., Liu, Y., Wang, Q., Ai, B., et al. (2019). SEdb: a comprehensive human super-enhancer database. *Nucleic Acids Res.* 47 (D1), D235-D243-d43. doi:10.1093/nar/gky1025

Jiang, Y. Y., Jiang, Y., Li, C. Q., Zhang, Y., Dakle, P., Kaur, H., et al. (2020). TP63, SOX2, and KLF5 establish a core regulatory circuitry that controls epigenetic and transcription patterns in esophageal squamous cell carcinoma cell lines. *Gastroenterology* 159 (4), 1311–1327. doi:10.1053/j.gastro.2020.06.050

Kang, Y., Kang, J., and Kim, A. (2021). Histone H3K4me1 strongly activates the DNase I hypersensitive sites in super-enhancers than those in typical enhancers. *Biosci. Rep.* 41 (7). doi:10.1042/bsr20210691

Khan, A., and Zhang, X. (2016). dbSUPER: a database of super-enhancers in mouse and human genome. *Nucleic Acids Res.* 44 (D1), D164–D171. doi:10.1093/nar/gkv1002

Klemm, S. L., Shipony, Z., and Greenleaf, W. J. (2019). Chromatin accessibility and the regulatory epigenome. *Nat. Rev. Genet.* 20 (4), 207–220. doi:10.1038/s41576-018-0089-8

Kling, M. J., Kesherwani, V., Mishra, N. K., Alexander, G., McIntyre, E. M., Ray, S., et al. (2022). A novel dual epigenetic approach targeting BET proteins and HDACs in Group 3 (MYC-driven) Medulloblastoma. *J. Exp. Clin. Cancer Res.* 41 (1), 321. doi:10. 1186/s13046-022-02530-y

Kong, R., Patel, A. S., Sato, T., Jiang, F., Yoo, S., Bao, L., et al. (2022). Transcriptional circuitry of NKX2-1 and SOX1 defines an unrecognized lineage subtype of small-cell lung cancer. Am. J. Respir. Crit. Care Med. 206 (12), 1480–1494. doi:10.1164/rccm. 202110-2358OC

Kostyrko, K., Román, M., Lee, A. G., Simpson, D. R., Dinh, P. T., Leung, S. G., et al. (2023). UHRF1 is a mediator of KRAS driven oncogenesis in lung adenocarcinoma. *Nat. Commun.* 14 (1), 3966. doi:10.1038/s41467-023-39591-2

Kumari, A., Gesumaria, L., Liu, Y. J., Hughitt, V. K., Zhang, X., Ceribelli, M., et al. (2023). mTOR inhibition overcomes RSK3-mediated resistance to BET inhibitors in small cell lung cancer. *JCI Insight* 8 (5), e156657. doi:10.1172/jci.insight.156657

Kwiatkowski, N., Zhang, T., Rahl, P. B., Abraham, B. J., Reddy, J., Ficarro, S. B., et al. (2014). Targeting transcription regulation in cancer with a covalent CDK7 inhibitor. *Nature* 511 (7511), 616–620. doi:10.1038/nature13393

Lam, L. T., Lin, X., Faivre, E. J., Yang, Z., Huang, X., Wilcox, D. M., et al. (2017). Vulnerability of small-cell lung cancer to apoptosis induced by the combination of BET bromodomain proteins and BCL2 inhibitors. *Mol. Cancer Ther.* 16 (8), 1511–1520. doi:10.1158/1535-7163.Mct-16-0459

Larochelle, S., Amat, R., Glover-Cutter, K., Sansó, M., Zhang, C., Allen, J. J., et al. (2012). Cyclin-dependent kinase control of the initiation-to-elongation switch of RNA polymerase II. *Nat. Struct. Mol. Biol.* 19 (11), 1108–1115. doi:10.1038/nsmb.2399

Lasko, L. M., Jakob, C. G., Edalji, R. P., Qiu, W., Montgomery, D., Digiammarino, E. L., et al. (2017). Discovery of a selective catalytic p300/CBP inhibitor that targets lineage-specific tumours. *Nature* 550 (7674), 128–132. doi:10.1038/nature24028

Lee, J. H., Xiong, F., and Li, W. (2020). Enhancer RNAs in cancer: regulation, mechanisms and therapeutic potential. *RNA Biol.* 17 (11), 1550–1559. doi:10.1080/15476286.2020.1712895

Lengauer, C., Kinzler, K. W., and Vogelstein, B. (1998). Genetic instabilities in human cancers. *Nature* 396 (6712), 643–649. doi:10.1038/25292

Lewin, J., Soria, J. C., Stathis, A., Delord, J. P., Peters, S., Awada, A., et al. (2018). Phase ib trial with Birabresib, a small-molecule inhibitor of bromodomain and extraterminal proteins, in patients with selected advanced solid tumors. *J. Clin. Oncol.* 36 (30), 3007–3014. doi:10.1200/jco.2018.78.2292

Li, F., Fang, Z., Zhang, J., Li, C., Liu, H., Xia, J., et al. (2016). Identification of TRA2B-DNAH5 fusion as a novel oncogenic driver in human lung squamous cell carcinoma. *Cell Res.* 26 (10), 1149–1164. doi:10.1038/cr.2016.111

Li, H., Muhetaer, G., Xie, Y., Yao, K., Ma, Q., Guan, H., et al. (2022a). Identification of super-enhancer-driven peptidyl arginine deiminases as potential biomarkers and therapeutic targets for osimertinib-resistant non-small cell lung cancer. *Front. Pharmacol.* 13, 1071365. doi:10.3389/fphar.2022.1071365

Li, J., Wang, J., Wang, Y., Zhao, X., and Su, T. (2023). E2F1 combined with LINC01004 super-enhancer to promote hepatocellular carcinoma cell proliferation and metastasis. *Clin. Epigenetics* 15 (1), 17. doi:10.1186/s13148-023-01428-6

Li, J., Zhang, Y., Wang, L., Li, M., Yang, J., Chen, P., et al. (2022b). FOXA1 prevents nutrients deprivation induced autophagic cell death through inducing loss of imprinting of IGF2 in lung adenocarcinoma. *Cell Death Dis.* 13 (8), 711. doi:10.1038/s41419-022-05150-8

Li, K., Xu, C., Du, Y., Junaid, M., Kaushik, A. C., and Wei, D. Q. (2019a). Comprehensive epigenetic analyses reveal master regulators driving lung metastasis of breast cancer. *J. Cell Mol. Med.* 23 (8), 5415–5431. doi:10.1111/jcmm.14424

Li, M., Yang, B., Li, X., Ren, H., Zhang, L., Li, L., et al. (2021). Identification of prognostic factors related to super enhancer-regulated ceRNA network in metastatic lung adenocarcinoma. *Int. J. Gen. Med.* 14, 6261–6275. doi:10.2147/ ijgm.S332317

Li, X., Lu, C., Lu, Q., Li, C., Zhu, J., Zhao, T., et al. (2019b). Differentiated superenhancers in lung cancer cells. *Sci. China Life Sci.* 62 (9), 1218–1228. doi:10.1007/ s11427-018-9319-4

Liu, C., Qian, L., Vallega, K. A., Ma, G., Zong, D., Chen, L., et al. (2022b). The novel BET degrader, QCA570, is highly active against the growth of human NSCLC cells and synergizes with osimertinib in suppressing osimertinib-resistant EGFR-mutant NSCLC cells. *Am. J. Cancer Res.* 12 (2), 779–792.

Liu, Q., Guo, L., Lou, Z., Xiang, X., and Shao, J. (2022a). Super-enhancers and novel therapeutic targets in colorectal cancer. *Cell Death Dis.* 13 (3), 228. doi:10.1038/s41419-022-04673-4

Liu, Y., Li, Y., Liu, S., Adeegbe, D. O., Christensen, C. L., Quinn, M. M., et al. (2018). NK cells mediate synergistic antitumor effects of combined inhibition of HDAC6 and BET in a SCLC preclinical model. *Cancer Res.* 78 (13), 3709–3717. doi:10.1158/0008-5472.Can-18-0161

Lovén, J., Hoke, H. A., Lin, C. Y., Lau, A., Orlando, D. A., Vakoc, C. R., et al. (2013). Selective inhibition of tumor oncogenes by disruption of super-enhancers. *Cell* 153 (2), 320–334. doi:10.1016/j.cell.2013.03.036

Lu, H., Xue, Y., Yu, G. K., Arias, C., Lin, J., Fong, S., et al. (2015). Compensatory induction of MYC expression by sustained CDK9 inhibition via a BRD4-dependent mechanism. *Elife* 4, e06535. doi:10.7554/eLife.06535

Luo, H., Shan, J., Zhang, H., Song, G., Li, Q., and Xu, C. X. (2022). Targeting the epigenetic processes to enhance antitumor immunity in small cell lung cancer. *Semin. Cancer Biol.* 86 (Pt 3), 960–970. doi:10.1016/j.semcancer.2022.02.018

Mansour, M. R., Abraham, B. J., Anders, L., Berezovskaya, A., Gutierrez, A., Durbin, A. D., et al. (2014). Oncogene regulation. An oncogenic super-enhancer formed through somatic mutation of a noncoding intergenic element. *Science* 346 (6215), 1373–1377. doi:10.1126/science.1259037

Mao, R., Wu, Y., Ming, Y., Xu, Y., Wang, S., Chen, X., et al. (2019). Enhancer RNAs: a missing regulatory layer in gene transcription. *Sci. China Life Sci.* 62 (7), 905–912. doi:10.1007/s11427-017-9370-9

Martínez-Jiménez, F., Muiños, F., Sentís, I., Deu-Pons, J., Reyes-Salazar, I., Arnedo-Pac, C., et al. (2020). A compendium of mutational cancer driver genes. *Nat. Rev. Cancer* 20 (10), 555–572. doi:10.1038/s41568-020-0290-x

Miyakawa, K., Miyashita, N., Horie, M., Terasaki, Y., Tanaka, H., Urushiyama, H., et al. (2022). ASCL1 regulates super-enhancer-associated miRNAs to define molecular subtypes of small cell lung cancer. *Cancer Sci.* 113 (11), 3932–3946. doi:10.1111/cas. 15481

Miyashita, N., Horie, M., Suzuki, H. I., Yoshihara, M., Djureinovic, D., Persson, J., et al. (2018). An integrative analysis of transcriptome and epigenome features of ASCL1-positive lung adenocarcinomas. *J. Thorac. Oncol.* 13 (11), 1676–1691. doi:10. 1016/j.jtho.2018.07.096

Nagaraja, S., Vitanza, N. A., Woo, P. J., Taylor, K. R., Liu, F., Zhang, L., et al. (2017). Transcriptional dependencies in diffuse intrinsic pontine glioma. *Cancer Cell* 31 (5), 635–652. doi:10.1016/j.ccell.2017.03.011 Nguyen, T. T. T., Zhang, Y., Shang, E., Shu, C., Torrini, C., Zhao, J., et al. (2020). HDAC inhibitors elicit metabolic reprogramming by targeting super-enhancers in glioblastoma models. J. Clin. Invest. 130 (7), 3699–3716. doi:10.1172/jci129049

Ogryzko, V. V., Schiltz, R. L., Russanova, V., Howard, B. H., and Nakatani, Y. (1996). The transcriptional coactivators p300 and CBP are histone acetyltransferases. *Cell* 87 (5), 953–959. doi:10.1016/s0092-8674(00)82001-2

Parker, S. C., Stitzel, M. L., Taylor, D. L., Orozco, J. M., Erdos, M. R., Akiyama, J. A., et al. (2013). Chromatin stretch enhancer states drive cell-specific gene regulation and harbor human disease risk variants. *Proc. Natl. Acad. Sci. U. S. A.* 110 (44), 17921–17926. doi:10.1073/pnas.1317023110

Pastore, A., Gaiti, F., Lu, S. X., Brand, R. M., Kulm, S., Chaligne, R., et al. (2019). Corrupted coordination of epigenetic modifications leads to diverging chromatin states and transcriptional heterogeneity in CLL. *Nat. Commun.* 10 (1), 1874. doi:10.1038/ s41467-019-09645-5

Peng, L., Jiang, B., Yuan, X., Qiu, Y., Peng, J., Huang, Y., et al. (2019). Super-enhancerassociated long noncoding RNA HCCL5 is activated by ZEB1 and promotes the malignancy of hepatocellular carcinoma. *Cancer Res.* 79 (3), 572–584. doi:10.1158/ 0008-5472.Can-18-0367

Persson, M., Andrén, Y., Mark, J., Horlings, H. M., Persson, F., and Stenman, G. (2009). Recurrent fusion of MYB and NFIB transcription factor genes in carcinomas of the breast and head and neck. *Proc. Natl. Acad. Sci. U. S. A.* 106 (44), 18740–18744. doi:10.1073/pnas.0909114106

Pongor, L. S., Tlemsani, C., Elloumi, F., Arakawa, Y., Jo, U., Gross, J. M., et al. (2022). Integrative epigenomic analyses of small cell lung cancer cells demonstrates the clinical translational relevance of gene body methylation. *iScience* 25 (11), 105338. doi:10.1016/ jisci.2022.105338

Pozo, K., Kollipara, R. K., Kelenis, D. P., Rodarte, K. E., Ullrich, M. S., Zhang, X., et al. (2021). ASCL1, NKX2-1, and PROX1 co-regulate subtype-specific genes in small-cell lung cancer. *iScience* 24 (9), 102953. doi:10.1016/j.isci.2021.102953

Qian, F. C., Li, X. C., Guo, J. C., Zhao, J. M., Li, Y. Y., Tang, Z. D., et al. (2019). SEanalysis: a web tool for super-enhancer associated regulatory analysis. *Nucleic Acids Res.* 47 (W1), W248-W255-w55. doi:10.1093/nar/gkz302

Qu, S., Fetsch, P., Thomas, A., Pommier, Y., Schrump, D. S., Miettinen, M. M., et al. (2022). Molecular subtypes of primary SCLC tumors and their associations with neuroendocrine and therapeutic markers. *J. Thorac. Oncol.* 17 (1), 141–153. doi:10. 1016/j.jtho.2021.08.763

Reck, M., Horn, L., Novello, S., Barlesi, F., Albert, I., Juhász, E., et al. (2019). Phase II study of roniciclib in combination with cisplatin/etoposide or carboplatin/etoposide as first-line therapy in patients with extensive-disease small cell lung cancer. *J. Thorac. Oncol.* 14 (4), 701–711. doi:10.1016/j.jtho.2019.01.010

Rengachari, S., Schilbach, S., Aibara, S., Dienemann, C., and Cramer, P. (2021). Structure of the human Mediator-RNA polymerase II pre-initiation complex. *Nature* 594 (7861), 129–133. doi:10.1038/s41586-021-03555-7

Riveiro, M. E., Astorgues-Xerri, L., Vazquez, R., Frapolli, R., Kwee, I., Rinaldi, A., et al. (2016). OTX015 (MK-8628), a novel BET inhibitor, exhibits antitumor activity in nonsmall cell and small cell lung cancer models harboring different oncogenic mutations. *Oncotarget* 7 (51), 84675–84687. doi:10.18632/oncotarget.13181

Roden, A. C., Greipp, P. T., Knutson, D. L., Kloft-Nelson, S. M., Jenkins, S. M., Marks, R. S., et al. (2015). Histopathologic and cytogenetic features of pulmonary adenoid cystic carcinoma. *J. Thorac. Oncol.* 10 (11), 1570–1575. doi:10.1097/jto.000000000000656

Ropri, A. S., DeVaux, R. S., Eng, J., Chittur, S. V., and Herschkowitz, J. I. (2021). Cisacting super-enhancer lncRNAs as biomarkers to early-stage breast cancer. *Breast Cancer Res.* 23 (1), 101. doi:10.1186/s13058-021-01479-8

Rudin, C. M., Poirier, J. T., Byers, L. A., Dive, C., Dowlati, A., George, J., et al. (2019). Molecular subtypes of small cell lung cancer: a synthesis of human and mouse model data. *Nat. Rev. Cancer* 19 (5), 289–297. doi:10.1038/s41568-019-0133-9

Sabari, B. R., Dall'Agnese, A., Boija, A., Klein, I. A., Coffey, E. L., Shrinivas, K., et al. (2018). Coactivator condensation at super-enhancers links phase separation and gene control. *Science* 361 (6400), eaar3958. doi:10.1126/science.aar3958

Saint-André, V., Federation, A. J., Lin, C. Y., Abraham, B. J., Reddy, J., Lee, T. I., et al. (2016). Models of human core transcriptional regulatory circuitries. *Genome Res.* 26 (3), 385–396. doi:10.1101/gr.197590.115

Sanchez, G. J., Richmond, P. A., Bunker, E. N., Karman, S. S., Azofeifa, J., Garnett, A. T., et al. (2018). Genome-wide dose-dependent inhibition of histone deacetylases studies reveal their roles in enhancer remodeling and suppression of oncogenic super-enhancers. *Nucleic Acids Res.* 46 (4), 1756–1776. doi:10.1093/nar/gkx1225

Sato, T., Yoo, S., Kong, R., Sinha, A., Chandramani-Shivalingappa, P., Patel, A., et al. (2019). Epigenomic profiling discovers trans-lineage SOX2 partnerships driving tumor heterogeneity in lung squamous cell carcinoma. *Cancer Res.* 79 (24), 6084–6100. doi:10. 1158/0008-5472.Can-19-2132

Schaukowitch, K., Joo, J. Y., Liu, X., Watts, J. K., Martinez, C., and Kim, T. K. (2014). Enhancer RNA facilitates NELF release from immediate early genes. *Mol. Cell* 56 (1), 29–42. doi:10.1016/j.molcel.2014.08.023

Shen, Q., Sun, Y., and Xu, S. (2020). LINC01503/miR-342-3p facilitates malignancy in non-small-cell lung cancer cells via regulating LASP1. *Respir. Res.* 21 (1), 235. doi:10. 1186/s12931-020-01464-3

Shi, K., Yin, X., Cai, M. C., Yan, Y., Jia, C., Ma, P., et al. (2019). PAX8 regulon in human ovarian cancer links lineage dependency with epigenetic vulnerability to HDAC inhibitors. *Elife* 8, e44306. doi:10.7554/eLife.44306

Shi, T., Hu, Z., Tian, L., and Yang, Y. (2023). Advances in lung adenocarcinoma: a novel perspective on prognoses and immune responses of CENPO as an oncogenic superenhancer. *Transl. Oncol.* 34, 101691. doi:10.1016/j.tranon.2023.101691

Shi, Y., Wang, M., Liu, D., Ullah, S., Ma, X., Yang, H., et al. (2022). Super-enhancers in esophageal carcinoma: transcriptional addictions and therapeutic strategies. *Front. Oncol.* 12, 1036648. doi:10.3389/fonc.2022.1036648

Siegel, R. L., Miller, K. D., Fuchs, H. E., and Jemal, A. (2022). Cancer statistics, 2022. *CA Cancer J. Clin.* 72 (1), 7–33. doi:10.3322/caac.21708

Siegel, R. L., Miller, K. D., and Jemal, A. (2020). Cancer statistics, 2020. CA Cancer J. Clin. 70 (1), 7–30. doi:10.3322/caac.21590

Soibam, B. (2017). Super-lncRNAs: identification of lncRNAs that target superenhancers via RNA:DNA:DNA triplex formation. *Rna* 23 (11), 1729–1742. doi:10.1261/ rna.061317.117

Song, X., Zhang, T., Ding, H., Feng, Y., Yang, W., Yin, X., et al. (2022). Non-genetic stratification reveals epigenetic heterogeneity and identifies vulnerabilities of glycolysis addiction in lung adenocarcinoma subtype. *Oncogenesis* 11 (1), 61. doi:10.1038/s41389-022-00436-0

Spitz, F., and Furlong, E. E. (2012). Transcription factors: from enhancer binding to developmental control. *Nat. Rev. Genet.* 13 (9), 613–626. doi:10.1038/nrg3207

Stephenson, J. J., Nemunaitis, J., Joy, A. A., Martin, J. C., Jou, Y. M., Zhang, D., et al. (2014). Randomized phase 2 study of the cyclin-dependent kinase inhibitor dinaciclib (MK-7965) versus erlotinib in patients with non-small cell lung cancer. *Lung Cancer* 83 (2), 219–223. doi:10.1016/j.lungcan.2013.11.020

Su, T., Zhang, N., Wang, T., Zeng, J., Li, W., Han, L., et al. (2023). Super enhancerregulated LncRNA LINC01089 induces alternative splicing of DIAPH3 to drive hepatocellular carcinoma metastasis. *Cancer Res.* 83 (24), 4080–4094. doi:10.1158/ 0008-5472.Can-23-0544

Sun, D., Nikonova, A. S., Zhang, P., Deneka, A. Y., Fitzgerald, M. E., Michael, R. E., et al. (2021). Evaluation of the small-molecule BRD4 degrader CFT-2718 in small-cell lung cancer and pancreatic cancer models. *Mol. Cancer Ther.* 20 (8), 1367–1377. doi:10. 1158/1535-7163.Mct-20-0831

Suski, J. M., Braun, M., Strmiska, V., and Sicinski, P. (2021). Targeting cell-cycle machinery in cancer. *Cancer Cell* 39 (6), 759–778. doi:10.1016/j.ccell.2021.03.010

Takashima, Y., Kikuchi, E., Kikuchi, J., Suzuki, M., Kikuchi, H., Maeda, M., et al. (2020). Bromodomain and extraterminal domain inhibition synergizes with WEE1inhibitor AZD1775 effect by impairing nonhomologous end joining and enhancing DNA damage in nonsmall cell lung cancer. *Int. J. Cancer* 146 (4), 1114–1124. doi:10. 1002/jic.32515

Tien, F. M., Lu, H. H., Lin, S. Y., and Tsai, H. C. (2023). Epigenetic remodeling of the immune landscape in cancer: therapeutic hurdles and opportunities. *J. Biomed. Sci.* 30 (1), 3. doi:10.1186/s12929-022-00893-0

Trabucco, S. E., Gerstein, R. M., Evens, A. M., Bradner, J. E., Shultz, L. D., Greiner, D. L., et al. (2015). Inhibition of bromodomain proteins for the treatment of human diffuse large B-cell lymphoma. *Clin. Cancer Res.* 21 (1), 113–122. doi:10.1158/1078-0432.Ccr-13-3346

Travis, W. D., Brambilla, E., Nicholson, A. G., Yatabe, Y., Austin, J. H. M., Beasley, M. B., et al. (2015). The 2015 world health organization classification of lung tumors: impact of genetic, clinical and radiologic advances since the 2004 classification. *J. Thorac. Oncol.* 10 (9), 1243–1260. doi:10.1097/jto.000000000000630

Traynor, A. M., Dubey, S., Eickhoff, J. C., Kolesar, J. M., Schell, K., Huie, M. S., et al. (2009). Vorinostat (NSC# 701852) in patients with relapsed non-small cell lung cancer: a Wisconsin Oncology Network phase II study. *J. Thorac. Oncol.* 4 (4), 522–526. doi:10. 1097/jto.0b013e3181952478

Trojanowski, J., Frank, L., Rademacher, A., Mücke, N., Grigaitis, P., and Rippe, K. (2022). Transcription activation is enhanced by multivalent interactions independent of phase separation. *Mol. Cell* 82 (10), 1878–1893.e10. doi:10.1016/j.molcel.2022.04.017

Tu, Y. H., Juan, H. F., and Huang, H. C. (2021). Identification of cell states using superenhancer RNA. *BMC Genomics* 22 (Suppl. 3), 787. doi:10.1186/s12864-021-08092-1

Vahedi, G., Kanno, Y., Furumoto, Y., Jiang, K., Parker, S. C., Erdos, M. R., et al. (2015). Super-enhancers delineate disease-associated regulatory nodes in T cells. *Nature* 520 (7548), 558–562. doi:10.1038/nature14154

Wang, H., Hong, B., Li, X., Deng, K., Li, H., Yan Lui, V. W., et al. (2017). JQ1 synergizes with the Bcl-2 inhibitor ABT-263 against MYCN-amplified small cell lung cancer. *Oncotarget* 8 (49), 86312–86324. doi:10.18632/oncotarget.21146

Wang, J., Liu, J., Li, S., Li, X., Yang, J., Dang, X., et al. (2023b). Genetic regulatory and biological implications of the 10q24.32 schizophrenia risk locus. *Brain* 146 (4), 1403–1419. doi:10.1093/brain/awac352

Wang, M., Chen, Z., and Zhang, Y. (2022). CBP/p300 and HDAC activities regulate H3K27 acetylation dynamics and zygotic genome activation in mouse preimplantation embryos. *Embo J.* 41 (22), e112012. doi:10.15252/embj.2022112012

Wang, M., Herbst, R. S., and Boshoff, C. (2021). Toward personalized treatment approaches for non-small-cell lung cancer. *Nat. Med.* 27 (8), 1345–1356. doi:10.1038/ s41591-021-01450-2

Wang, X., Yu, C., Wang, C., Ma, Y., Wang, T., Li, Y., et al. (2019). Novel cyclindependent kinase 9 (CDK9) inhibitor with suppression of cancer stemness activity against non-small-cell lung cancer. *Eur. J. Med. Chem.* 181, 111535. doi:10.1016/j. ejmech.2019.07.038

Wang, Y., Song, C., Zhao, J., Zhang, Y., Zhao, X., Feng, C., et al. (2023a). SEdb 2.0: a comprehensive super-enhancer database of human and mouse. *Nucleic Acids Res.* 51 (D1), D280–D290. doi:10.1093/nar/gkac968

Wang, Y., Zhang, T., Kwiatkowski, N., Abraham, B. J., Lee, T. I., Xie, S., et al. (2015). CDK7-dependent transcriptional addiction in triple-negative breast cancer. *Cell* 163 (1), 174–186. doi:10.1016/j.cell.2015.08.063

Watanabe, H., Ma, Q., Peng, S., Adelmant, G., Swain, D., Song, W., et al. (2014). SOX2 and p63 colocalize at genetic loci in squamous cell carcinomas. *J. Clin. Invest.* 124 (4), 1636–1645. doi:10.1172/jci71545

Wei, J., Yu, H., Liu, T., Wang, Z., Lang, C., and Pan, Y. (2023). FOXA1-induced LINC00621 promotes lung adenocarcinoma progression via activating the TGF- β signaling pathway. *Thorac. Cancer* 14 (21), 2026–2037. doi:10.1111/1759-7714.14986

Wei, Y., Zhang, S., Shang, S., Zhang, B., Li, S., Wang, X., et al. (2016). SEA: a superenhancer archive. *Nucleic Acids Res.* 44 (D1), D172–D179. doi:10.1093/nar/gkv1243

Whyte, W. A., Orlando, D. A., Hnisz, D., Abraham, B. J., Lin, C. Y., Kagey, M. H., et al. (2013). Master transcription factors and mediator establish super-enhancers at key cell identity genes. *Cell* 153 (2), 307–319. doi:10.1016/j.cell.2013.03.035

Wiese, M., Hamdan, F. H., Kubiak, K., Diederichs, C., Gielen, G. H., Nussbaumer, G., et al. (2020). Combined treatment with CBP and BET inhibitors reverses inadvertent activation of detrimental super enhancer programs in DIPG cells. *Cell Death Dis.* 11 (8), 673. doi:10.1038/s41419-020-02800-7

William, W. N., Jr., and Glisson, B. S. (2011). Novel strategies for the treatment of small-cell lung carcinoma. *Nat. Rev. Clin. Oncol.* 8 (10), 611–619. doi:10.1038/nrclinonc.2011.90

Wu, L., Yao, H., Chen, H., Wang, A., Guo, K., Gou, W., et al. (2022). Landscape of somatic alterations in large-scale solid tumors from an Asian population. *Nat. Commun.* 13 (1), 4264. doi:10.1038/s41467-022-31780-9

Wu, P. F., Gao, W. W., Sun, C. L., Ma, T., and Hao, J. Q. (2020). Suberoylanilide hydroxamic acid overcomes erlotinib-acquired resistance via phosphatase and tensin homolog deleted on chromosome 10-mediated apoptosis in non-small cell lung cancer. *Chin. Med. J. Engl.* 133 (11), 1304–1311. doi:10.1097/cm9.000000000000823

Wu, Y., Chen, S., Shao, Y., Su, Y., Li, Q., Wu, J., et al. (2023). KLF5 promotes tumor progression and parp inhibitor resistance in ovarian cancer. *Adv. Sci. (Weinh)* 10 (31), e2304638. doi:10.1002/advs.202304638

Xie, J. J., Jiang, Y. Y., Jiang, Y., Li, C. Q., Lim, M. C., An, O., et al. (2018). Superenhancer-driven long non-coding RNA LINC01503, regulated by TP63, is overexpressed and oncogenic in squamous cell carcinoma. *Gastroenterology* 154 (8), 2137–2151. doi:10.1053/j.gastro.2018.02.018

Xie, K., Feng, J., Fan, D., Wang, S., Luo, J., Ren, Z., et al. (2022). BARX2/FOXA1/ HK2 axis promotes lung adenocarcinoma progression and energy metabolism reprogramming. *Transl. Lung Cancer Res.* 11 (7), 1405–1419. doi:10.21037/tlcr-22-465

Xing, R., Zhou, Y., Yu, J., Yu, Y., Nie, Y., Luo, W., et al. (2019). Whole-genome sequencing reveals novel tandem-duplication hotspots and a prognostic mutational signature in gastric cancer. *Nat. Commun.* 10 (1), 2037. doi:10.1038/s41467-019-09644-6

Xu, Y., Wu, Y., Zhang, S., Ma, P., Jin, X., Wang, Z., et al. (2019). A tumor-specific super-enhancer drives immune evasion by guiding synchronous expression of PD-L1 and PD-L2. *Cell Rep.* 29 (11), 3435–3447. doi:10.1016/j.celrep.2019.10.093

Yamagata, K., Nakayamada, S., Zhang, T., Nguyen, A. P., Ohkubo, N., Iwata, S., et al. (2022). IL-6 production through repression of UBASH3A gene via epigenetic dysregulation of super-enhancer in CD4(+) T cells in rheumatoid arthritis. *Inflamm. Regen.* 42 (1), 46. doi:10.1186/s41232-022-00231-9

Yan, L., Chen, H., Tang, L., Jiang, P., and Yan, F. (2021). Super-enhancer-associated long noncoding RNA AC005592.2 promotes tumor progression by regulating OLFM4 in colorectal cancer. *BMC Cancer* 21 (1), 187. doi:10.1186/s12885-021-07900-x

Yang, J., Xu, J., Wang, W., Zhang, B., Yu, X., and Shi, S. (2023a). Epigenetic regulation in the tumor microenvironment: molecular mechanisms and therapeutic targets. *Signal Transduct. Target Ther.* 8 (1), 210. doi:10.1038/s41392-023-01480-x

Yang, X., Han, H., De Carvalho, D. D., Lay, F. D., Jones, P. A., and Liang, G. (2014). Gene body methylation can alter gene expression and is a therapeutic target in cancer. *Cancer Cell* 26 (4), 577–590. doi:10.1016/j.ccr.2014.07.028

Yang, Z., Yik, J. H., Chen, R., He, N., Jang, M. K., Ozato, K., et al. (2005). Recruitment of P-TEFb for stimulation of transcriptional elongation by the bromodomain protein Brd4. *Mol. Cell* 19 (4), 535–545. doi:10.1016/j.molcel.2005.06.029

Yang, Z., Zheng, Y., Wu, H., Xie, H., Zhao, J., Chen, Z., et al. (2023b). Integrative analysis of a novel super-enhancer-associated lncRNA prognostic signature and identifying LINC00945 in aggravating glioma progression. *Hum. Genomics* 17 (1), 33. doi:10.1186/s40246-023-00480-w

Yoo, K. H., Hennighausen, L., and Shin, H. Y. (2019). Dissecting tissue-specific super-enhancers by integrating genome-wide analyses and CRISPR/Cas9 genome editing. *J. Mammary Gland. Biol. Neoplasia* 24 (1), 47–59. doi:10.1007/s10911-018-9417-z

You, L., Guo, X., and Huang, Y. (2018). Correlation of cancer stem-cell markers OCT4, SOX2, and NANOG with clinicopathological features and prognosis in operative patients with rectal cancer. *Yonsei Med. J.* 59 (1), 35–42. doi:10.3349/ymj.2018.59.1.35

Yuan, C., Chen, H., Tu, S., Huang, H. Y., Pan, Y., Gui, X., et al. (2021). A systematic dissection of the epigenomic heterogeneity of lung adenocarcinoma reveals two different subclasses with distinct prognosis and core regulatory networks. *Genome Biol.* 22 (1), 156. doi:10.1186/s13059-021-02376-1

Yuan, X. Q., Zhou, N., Wang, J. P., Yang, X. Z., Wang, S., Zhang, C. Y., et al. (2023). Anchoring super-enhancer-driven oncogenic lncRNAs for anti-tumor therapy in hepatocellular carcinoma. *Mol. Ther.* 31 (6), 1756–1774. doi:10.1016/j.ymthe.2022. 11.013

Zhang, J., Zhang, X., Xie, F., Zhang, Z., van Dam, H., Zhang, L., et al. (2014). The regulation of TGF-β/SMAD signaling by protein deubiquitination. *Protein Cell* 5 (7), 503–517. doi:10.1007/s13238-014-0058-8

Zhang, T., Song, X., Zhang, Z., Mao, Q., Xia, W., Xu, L., et al. (2020a). Aberrant superenhancer landscape reveals core transcriptional regulatory circuitry in lung adenocarcinoma. *Oncogenesis* 9 (10), 92. doi:10.1038/s41389-020-00277-9

Zhang, T., Xia, W., Song, X., Mao, Q., Huang, X., Chen, B., et al. (2022). Superenhancer hijacking LINC01977 promotes malignancy of early-stage lung adenocarcinoma addicted to the canonical TGF-β/SMAD3 pathway. *J. Hematol. Oncol.* 15 (1), 114. doi:10.1186/s13045-022-01331-2

Zhang, X., Choi, P. S., Francis, J. M., Imielinski, M., Watanabe, H., Cherniack, A. D., et al. (2016). Identification of focally amplified lineage-specific super-enhancers in human epithelial cancers. *Nat. Genet.* 48 (2), 176–182. doi:10.1038/ng.3470

Zhang, Y., Huang, Y. X., Wang, D. L., Yang, B., Yan, H. Y., Lin, L. H., et al. (2020b). LncRNA DSCAM-AS1 interacts with YBX1 to promote cancer progression by forming a positive feedback loop that activates FOXA1 transcription network. *Theranostics* 10 (23), 10823–10837. doi:10.7150/thno.47830

Zheng, J., Huang, B., Xiao, L., and Wu, M. (2024). Effects of BRD4 inhibitor JQ1 on the expression profile of super-enhancer related lncRNAs and mRNAs in cervical cancer HeLa cells. *PeerJ* 12, e17035. doi:10.7717/peerj.17035

Zhou, J., Wang, D., Tang, D., and Huang, W. (2020). Abnormal activations of superenhancers enhance the carcinogenicity in lung adenocarcinoma. *Cancer Manag. Res.* 12, 8509–8518. doi:10.2147/cmar.S258497

Zhou, Q., Li, T., and Price, D. H. (2012). RNA polymerase II elongation control. Annu. Rev. Biochem. 81, 119-143. doi:10.1146/annurev-biochem-052610-095910

Zhou, R. W., and Parsons, R. E. (2023). Etiology of super-enhancer reprogramming and activation in cancer. *Epigenetics Chromatin* 16 (1), 29. doi:10.1186/s13072-023-00502-w

Zhou, X., Sun, T., Meng, Y., Luo, J., Chen, J., Xia, B., et al. (2021). BET inhibitors combined with chemotherapy synergistically inhibit the growth of NSCLC cells. *Oncol. Rep.* 45 (5), 70. doi:10.3892/or.2021.8021