#### Check for updates

#### **OPEN ACCESS**

EDITED BY Chiara De Santi, Royal College of Surgeons in Ireland, Ireland

#### REVIEWED BY

Antonio Leo, University Magna Graecia of Catanzaro, Italy Imran Shair Mohammad, City of Hope National Medical Center, United States

\*CORRESPONDENCE Marta Biagioli, amarta.biagioli@unitn.it Jasmin Morandell, jasmin.morandell@unitn.it

RECEIVED 15 February 2025 ACCEPTED 02 May 2025 PUBLISHED 16 May 2025

#### CITATION

Galli V, Vinciguerra S, Biagioli M and Morandell J (2025) Circular RNAs as disease modifiers of complex neurologic disorders. *Front. Pharmacol.* 16:1577496. doi: 10.3389/fphar.2025.1577496

#### COPYRIGHT

© 2025 Galli, Vinciguerra, Biagioli and Morandell. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Circular RNAs as disease modifiers of complex neurologic disorders

Veronica Galli, Sara Vinciguerra, Marta Biagioli\* and Jasmin Morandell\*

NeuroEpigenetics Laboratory, Department of Cellular, Computational and Integrative Biology - CIBIO, University of Trento, Trento, Italy

Circular RNAs are a large class of non-coding RNA molecules, conserved across species and produced by back-splicing. While their molecular functions are still elusive, the ones primarily retained in the nucleus are usually associated to regulation of transcription and mRNA processing patterns. Instead, the majority, are transported to the cytoplasm where they elicit micro-RNA (miRNA) or RNA binding protein (RBP)-sponging functions, or could be translated. CircRNAs are abundantly expressed in brain tissue, where they do not only act as regulators of brain development and physiology, but can also contribute to complex neurological conditions. In fact, deregulated circRNA expression levels were described in neurodevelopmental and neurodegenerative disorders, like Alzheimer's disease, Parkinson's disease and Huntington's disease. Because of their described roles in pathology, these molecules may not only represent possible disease bio-markers, but they could even function as disease modifiers. As such, they could be targeted or protected in search of novel routes of therapeutic intervention. In this review, we highlight recent developments in the field, first discussing circRNAs involved in physiologic brain development and function, then reviewing studies that implicate circRNAs in neurodevelopmental and neurodegenerative disorders, with major attention to experimental studies exploring circRNA function and their role in neuropathologic processes. Such experimental strategies are mainly based on depletion or over-expression approaches and provide important insights into the modulatory potential of these molecules. They are relevant for clinical translation of basic research findings to drug development, possibly generating a positive impact for patients' quality of life.

#### KEYWORDS

circRNA, non-coding RNA, alternative splicing, neurodevelopment, neurodegeneration

## Introduction

Circular RNAs are a recently re-discovered, evolutionary conserved, large class of RNA molecules (Wang et al., 2014). Eukaryotic circRNAs are produced by a process called back-splicing, leading to the formation of a covalent bond between a pre-mRNA 3'-splice donor (down-stream) and a 5'-splice acceptor (up-stream) site, resulting in the generation of a unique junction, the back-splice sequence (BSJ) (Jeck et al., 2013; Memczak et al., 2013) (Figure 1A). Still, it is the canonical splicing machinery that generates these continuous loops of RNA while processing exons, introns, or both exon-introns of, primarily, protein coding genes. The precise details of circRNA biogenesis are not yet fully elucidated, but



#### FIGURE 1

CircRNAs as modifiers of complex brain disorders. (A) Schematic of linear splicing (top) vs back-splicing (bottom). Back-splicing leads to the formation of a covalent bond between the 3'-splice donor and a 5'-splice acceptor site of unprocessed pre-mRNAs, generating the unique back-splicing sequence and rendering the resulting circRNAs extremely resistant against degradation through exoribonucleases. (B) CircRNAs are evolutionarily conserved and especially abundant in the nervous system of a large number of different species and especially mammals. Globally, circRNA expression increases during brain development, maturation and aging (bottom). In the last decade, numerous studies have linked deregulation of a variety of circRNAs with both, neurodevelopmental disorders (NDDs) such as Autism Spectrum Disorders and Schizophrenia (left); neurodegenerative (NDG) processes such as Alzheimer's, Parkinson's and Huntington's Disease, as well as Frontotemporal dementia and Amyotropic lateral sclerosis (right); and other complex brain disorders such as Epilepsy, Depression and brain tumor development (top). Relevant circRNAs (in red) have been shown to regulate the cell cycle of neural stem cells (radial glia -RG, intermediate progenitors–IP), the migration of newborn neurons (MN), the differentiation into inhibitory (IN) and excitatory (EN) post-mitotic neurons, astrocytes (A), microglia (MG), and oligodendrocytes (O), synaptogenesis, and mechanisms involved in mature brain function, homeostasis and survival- synaptic transmission and plasticity, neuronal and dendritic morphology, as well as brain immune activation.

evidence underscores the importance of base-pairing of the two introns adjacent to the circularizing exons, with typically long length (>10 kilobases), enriched in inverted ALU repeats (Jeck et al., 2013). In other cases, specific binding of RBPs (such as FUS and QKI) to the intronic sequences adjacent to circularizing exons drives circularization (Ashwal-Fluss et al., 2014; Starke et al., 2015).

The circular conformation of these RNA molecules confers a unique resistance against degradation, since 3'-5' exonucleases

preferentially digest linear RNAs while leaving most circRNAs intact (Suzuki et al., 2006). Thus, circRNAs are much more stable than linear RNA transcripts (Hwang and Kim, 2024), with a median half-life that is at least 2.5 times longer (Enuka et al., 2016; Jeck et al., 2013). Nonetheless, most circRNAs are expressed at low levels when compared to their cognate linear counterparts (Santer et al., 2019), but in a highly tissue-, cell type-, and time point-dependent context. To date, based on large scale, bulk RNA

sequencing (RNAseq) studies, multiple databases have been constructed to help annotate circRNA expression patterns of millions of circRNAs across different cell lines, tissues and organisms (Chen et al., 2016; Dong et al., 2018; Glažar et al., 2014; Meng et al., 2019; Wu et al., 2020). Furthermore, with the advent of single cell RNA sequencing, efforts have been made to resolve the complexity of circRNA expression landscapes also at the single cell level (Fan et al., 2015; W. Wu et al., 2022; Zhong et al., 2017).

While a molecular and functional characterization remains neglected for the majority of circRNAs, some circular RNA molecules that are primarily retained in the nucleus, have been described to act on the transcription and splicing regulation of their host and/or nearby located genes (Mai et al., 2022). Others are primarily exported into the cytoplasm, where they can act as miRNA "sponges" (Hansen et al., 2013; Peng et al., 2017), or interact with RBPs (Holdt et al., 2016; Ikeda et al., 2023), thus competing with linear mRNAs. Furthermore, some circRNAs appear to be translated in a cap-independent fashion, via internal ribosomal entry sites (IRES) (Chen et al., 2021), although the peptide coding potential of circRNAs, at endogenous levels and in physiological context, remains under discussion. CircRNAs expression is especially abundant in the brain and increases during aging, independently from the host gene mRNA expression (Hanan et al., 2017; Knupp and Miura, 2018; Rybak-Wolf et al., 2015). Over the last decade, abnormal levels of circRNAs have been associated with a large number of complex human neurologic disorders, ranging from ischemic stroke, over neurodevelopmental disorders (such as epilepsy, schizophrenia and autism spectrum disorders), Alzheimer's Disease and Parkinson's Disease, as well as Huntington's Disease (Mai et al., 2022; Morandell et al., 2024; Titze-de-Almeida & Titze-de-Almeida, 2023; Wang et al., 2023).

Recent work has further identified the dedicated mechanism by which circRNAs are exported to the cytoplasm, dependent upon Ran-GTP/Exportin-2, underscoring the potential relevance of circRNA localization for cell function (Ngo et al., 2024).

While for a vast majority of the identified deregulated circRNAs associated with human neurologic disorders their biologic function and relevance for the disease pathogenesis/progression remains elusive, in the past decade a number of studies have discovered important regulatory roles for some circRNAs in neuronal development, function and homeostasis with the detailed analysis of their molecular mechanisms of action. This review will put these most recent functional studies into the spotlight and consequently discuss how deregulation of circRNAs can modify complex neurologic disorders, from neurodevelopment to neurodegeneration.

# CircRNAs in physiologic brain development and function

CircRNAs are most abundantly expressed in brain tissues (Hanan et al., 2017). The majority of neuronal circRNAs were shown to be localized both at the cell body and dendrites of neurons. Additionally, their expression pattern changes according to the developmental stage of the cell, suggesting a functional role of circRNAs during development and the mature brain (Hollensen et al., 2020; Venø et al., 2015). In line with these observations, circRNAs found in dopaminergic and pyramidal neurons are predominantly expressed from gene loci relevant for synaptic activity, underscoring their potential functional role in determining activity of these neuronal subtypes, but also the whole neuronal network (Dong et al., 2023; You et al., 2015). Single cell RNAseq efforts have furthermore shown that circRNAs display specific expression patterns in different neuronal cell types, presumably contributing to cell diversity and identity (Dong et al., 2023).

Arguably the most thoroughly studied neuronal circRNA is complementary determining region 1 antisense (Cdr1-as). Cdr1as contains >70 binding sites for the neuronal microRNA (miRNA) miR-7 and acts as miR-7 sponge (Hansen et al., 2013), as part of a complex non-coding regulatory network (Kleaveland et al., 2018; Piwecka et al., 2017). Interestingly, binding of miR-7 seems to actively mediate the circularization of Cdr1-as (Belter et al., 2022), while loss of Cdr1-as leads to miR-7 destabilization and alters excitatory synaptic transmission, driving schizophreniaassociated behaviours (Piwecka et al., 2017). Moreover, synaptic localization of Cdr1-as is required for fear-memory extinction (Zajaczkowski et al., 2023), while, in cortical neurons, Cdr1-as buffers miR-7 activity to control glutamatergic excitatory transmission and neuronal connectivity. As such, these observations highlight the possible relevance of circRNAs for memory formation and long-lasting synaptic adaptations (Cerda-Jara et al., 2024).

Human *circSLC45A4*, one of the highest expressed circRNAs in the developing human frontal cortex, was reported to be necessary in maintaining the pool of neural progenitors *in vivo*, and its knockdown promoted Cajal-Retius cell differentiation in the cortical plate during early embryonic development (Suenkel et al., 2020). Similarly important for neural progenitor differentiation and cell positioning in the developing cortex is *circFAT3*. As shown by Seeler et al., it is especially relevant for differentiation of human embryonic stem cells into rostral and caudal neural progenitors. While *circFAT3* knockdown caused minimal changes in bulk RNA expression, it had significant effects at the single-cell level in human cerebral organoids, disrupting expression patterns linked to neural differentiation and migration of telencephalic radial glial cells and mature cortical neurons (Seeler et al., 2023).

Morphology and migration of midbrain dopamine neurons (mDA), crucial for motor and cognitive behaviours and linked to various brain disorders, are critically regulated by circRmst (Rybiczka-Tešulov et al., 2024). While single-molecule fluorescence in situ hybridization (smFISH) did not reveal any specific compartmentalization of highly expressed circRNAs in the cell, in vitro knock-down of the candidate circRmst had widespread effects in in vitro primary cultures, such as increased soma size, soma-to-nucleus ratio, reduced numbers of tyrosine hydroxylase positive neurons and, in vivo, accelerated neuronal migration and lateral mislocalization of the mDA neuron pool (Rybiczka-Tešulov et al., 2024). On the other hand, circTulp4 depletion, also highly expressed in developing mDA neurons, only led to milder effects, causing a small decrease in neurite length and increase in secondary branch formation (Rybiczka-Tešulov et al., 2024). Its primary function, in fact, is thought to lie in the regulation of general brain physiology and synaptic activity,

due to its compartmentalized enrichment at the synapse. Concordantly, depletion of *circTulp4* - by the alteration of the splice acceptor site responsible for its biogenesis - modulated the strength of excitatory neurotransmission in primary cultures and sensitivity to aversive stimuli in behavioral assays (Giusti et al., 2024). While *circTulp4* molecular mechanism is unlikely to be driven by miRNA-sponging or translation from the circular molecule, its mode of action remains to be identified (Giusti et al., 2024).

Synaptogenesis and dendritic spine formation was further shown to be strictly regulated by *circRERE and circPhf21a* expression in rat hippocampal neurons. *CircRERE* depletion led to the formation of electrophysiologically silent synapses, and Kelly et al. could demonstrate *circRERE*-mediated stabilization of *miR*-*128-3p* as a novel molecular mechanism to modulate the formation of silent excitatory synaptic co-clusters (Kelly et al., 2024, 2025).

The miRNA-sponging ability of circRNAs, implies also the capability of these molecules to regulate gene expression through circRNA/miRNA/mRNA interaction networks. In line with this molecular mechanism, *circDlc1* was shown to bind to *miR-130b-5p*, controlling its synaptic localization and regulating the expression of different mRNAs in the glutamatergic circuit. Thereby it plays an important role in synaptic transmission, regulating glutamatergic signaling and plasticity in cortico-striatal pathways. In line with these findings, loss of *circDlc1* lead to hyperactivity behaviour in a knock-out mouse model (Silenzi et al., 2024).

Taken together, these studies highlight important regulatory functions of circRNAs for physiologic brain development, modulating all steps of the process, from neural progenitor positioning and neurogenesis, over migration and differentiation of newborn neurons, to dendritic spine formation, synaptogenesis, synaptic function and plasticity (Figure 1B).

# CircRNAs in neurodevelopmental disorders

Because of their enrichment in the central nervous system (CNS) and their crucial roles in neurogenesis, synaptic plasticity and neuronal identity not surprisingly, circRNAs have been associated with neurodevelopmental (NDDs) and neuropsychiatric disorders, were they are increasingly recognized as key regulators of pathological processes (Nemeth et al., 2024). According to the DSM-5, NDDs include autism spectrum disorders (ASD), intellectual disability (ID), attention deficit hyperactivity disorder (ADHD), communication and motor disorders. These diagnostic categories, defined by specific sets of symptoms, show considerable comorbidity and phenotypic overlap, complicating diagnosis and management of the disorders. In fact, genetic research supports the hypothesis that ID, ASD, ADHD and schizophrenia lie on a neurodevelopmental continuum, rather than representing independent conditions (Morris-Rosendahl and Crocq, 2020).

The phenotypic heterogeneity of ASD and the co-morbid other neurodevelopmental syndromes is mirrored by the large complexity in underlying genetic architecture and altered biologic pathways, which have been studied in detail over the last 4 decades (reviewed here (Choi and An, 2021) and here (Parenti et al., 2020)). Instead, the exploration of long non-coding (lnc) RNAs, of which circRNAs are an important subgroup, and their contribution to the etiology of NDDs is yet in its infancy, and has been, so far, primarily focused on ASD and schizophrenia, but can possibly be extrapolated to other NDDs (Ghafouri-Fard et al., 2022; Jiang et al., 2023; Stott et al., 2023). A comprehensive study utilizing previously published RNAseq data sets from total RNA (ribosomal RNA-depleted RNAs without poly(A)-selection) derived from a large collection of post mortem brain tissue of control and ASD cases (Parikshak et al., 2016; Wu et al., 2016), represents the first systematic investigation of circRNA dysregulation in ASD. In the study, the authors identified 60 circRNAs and three co-regulated modules that were perturbed in ASD. By integrating circRNA, microRNA, and mRNA dysregulation data derived from the same cortical samples, 8170 ASD-associated circRNA/miRNA/mRNA networks were identified. The 2302 putative target genes of these axes were enriched for ASD risk genes and genes encoding inhibitory postsynaptic density (PSD) proteins, but not for genes implicated in monogenic forms of other brain disorders or genes encoding excitatory PSD proteins. Functionally, only *circARID1A* was tested: upregulated in ASD brain samples and working as miR-204-3p sponge, the corresponding circRNA/miRNA/mRNA regulatory network was further characterized in vitro employing knockdown and over-expression experiments, supporting the notion that circRNAs may serve as efficient miRNA sponges, influencing the expression dynamics of ASD-associated risk genes and leading to synaptic dysfunction and abnormal neural development (Huang et al., 2023). Furthermore, the genomic study by Te-Lun Mai et al. focused on quantitative trait loci (QTL) associated with the expression of nearby circRNAs (circQTL). They integrated RNAseq and genotyping data from post mortem brain tissues of individuals with ASD, to identify circQTLs that cis-regulate nearby circRNAs expression and, in turn, trans-regulate distant gene (trans-eGenes). The researchers identified expression 3,619 circQTLs, constructing 19,804 circQTL-circRNA-transeGene regulatory axes. 291 of these circQTLs overlapped with ASD-related genetic variants identified in genome-wide association studies (GWAS), suggesting a potential link between these circQTLs, SFARI risk factor genes, and ASD pathogenesis (Mai et al., 2022). Such findings highlight the complexity of gene regulation in ASD and suggest that circRNAs may serve as key intermediaries in the genetic architecture of the disorder.

On the other hand, additional insights on the importance of circRNAs in NDDs can be derived from studies using animal models. Wang et al. identified a vast array of differentially expressed circRNAs following valproate treatment in the mouse brain. By exploring a circRNA-based competing endogenous RNA (ceRNA) network, the study revealed possible interactions between circRNAs, miRNAs, and mRNAs, highlighting the complex regulatory networks that might potentially contribute to neurodevelopmental pathology (Wang et al., 2021). Similarly, Gasparini et al. explored the differential expression of hippocampal circRNAs, such as *circCdh9*, in the BTBR mouse model of ASD (Meyza and Blanchard, 2017). Their findings suggest that these circRNAs may contribute to synaptic dysfunctions observed in ASD (Gasparini et al., 2020).

In the context of SCZ, *circHOMER1* expression levels were significantly reduced in *post mortem* orbitofrontal cortex samples of patients, and *circHOMER1* levels positively correlated with the

10.3389/fphar.2025.1577496

age of SCZ onset. CircHomer1a regulated the expression of specific isoforms of synaptic plasticity-related genes with relevance for psychiatric disorders. Knock-down of circHomer1a in the orbitofrontal cortex was sufficient to disrupt cognitive flexibility in vivo (Zimmerman et al., 2020). In SCZ, circRNAs have also been intensely studied as potential class of novel diagnostic and therapeutic biomarkers (Huang et al., 2023; Tan et al., 2021; Yao et al., 2019). For instance, circRNA expression was profiled in plasma samples of very early-onset schizophrenia (VEOS) patients and 235 circRNAs (1 upregulated, 234 downregulated) were shown to be differentially expressed. A circRNA/miRNA/ mRNA network was then constructed and identified target genes that were involved in signal transduction, the cytoskeleton and transport pathways. Thus, plasma circRNAs in this network may be potential diagnostic biomarkers for VEOS and could play an important role in the onset and development of VEOS symptoms (Huang et al., 2023).

Taken together, although with much room for further exploration, circRNA dysregulation might lead to aberrant gene expression, contributing to the development and progression of neurodevelopmental and neuropsychiatric disorders. As modulators of altered neurodevelopment, they may also represent promising therapeutic targets, offering new avenues for treating neuropsychiatric conditions or be used as diagnostic or therapeutic biomarkers in easily accessible biofluids.

# CircRNAs in the aging brain and neurodegenerative diseases

Neurodegenerative disorders, including Alzheimer's disease (AD), Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), and Huntington's disease (HD), are marked by the progressive degeneration and loss of neurons in distinct regions of the brain and spinal cord. These conditions are currently irreversible, with no effective treatments available, eventually leading to decline and death of the affected patient. Although, in certain cases, clear genetic causes can be identified, aging still represents an important risk factor for most neurodegenerative diseases (Hou et al., 2019). Thus, in line with the ever increasing life expectancy and increasing numbers of individuals with advanced age, especially in industrialized countries, neurodegenerative disorders are considered a leading cause of death by the World Health Organization (WHO).

## Alzheimer's disease

Global changes in circRNA expression levels in AD have first been explored in detail by Dube et al., demonstrating that collectively, the expression changes of ten circRNAs, in the human *post mortem* parietal cortex, had a higher predictive value for cognitive impairment and dementia, than, e.g., the number of *APOE4* alleles (Dube et al., 2019). Additionally, an association of circRNA expression levels with AD has also been found in other brain regions (Cochran et al., 2021; Lo et al., 2020), such as the frontal lobe, temporal cortex and hippocampus. Importantly, circRNAs from two AD-associated genes, *presenilin 1* (*PSEN1*) and *tau* have further been studied more in detail. *CircPSEN1* levels increase during autosomal-dominant Alzheimer's disease progression which may be related to neuro-inflammatory events and amyloid beta accumulation (Chen et al., 2022). Instead, after undergoing RNA editing, circular RNAs from the *tau* pre-mRNA encompassing the microtubule binding sites were shown to be translated into proteins that promote tau aggregation *in vitro*, and are therefore hypothesized to contribute to tauopathies more in general (Welden et al., 2022).

Maquera et al. provided initial evidence for a role of translated circRNAs in the development of AD. Through RNAseq based analysis of the mRNA and circRNA composition in *post mortem* human cortical tissues during AD progression, as characterized by the spread of neurofibrillary tangles from the entorhinal cortex (early stages) to the whole brain, they identified *circMAN2A1* expression to be correlated with AD progression. Transfection experiments indicated that RNA editing promoted its translation using start codons out of frame with linear mRNAs, which generates novel proteins (Arizaca Maquera et al., 2023).

In a study by Puri et al., RNAseq analysis of neuropathologically confirmed AD brains and controls (n = 2027) identified 4,092 deregulated circRNAs in the hippocampus and 4,912 in the cortex. The study aimed to assess the relationship between circRNA expression patterns and AD clinical and pathological outcomes (Puri et al., 2023). CircRNA expression was correlated with the Clinical Dementia Rating (CDR) and AD-related neuropathological traits, including CERAD neuritic plaque score and Braak stage. Among 48 differentially expressed circRNAs, 69% were significantly associated with all three parameters. Key circRNAs such as circSLC8A1 (linked also with Parkinson's disease) and circHOMER1, associated with synaptic function and calcium signaling, were significantly downregulated in AD patients, correlating with synaptic loss and cognitive decline. The downregulation of the validated circRNAs following treatment with oligomeric Tau (oTau) highlighted the responsiveness of AD-associated circRNAs in neuro progenitor cells (NPC). CircMAPK9 exhibited downregulation by oTau exposure, independent of its linear transcript counterpart. These observations raised the possibility that some of the changes in circRNA expression in the AD brain might reflect the response to oTau toxicity, consistent with Cervera-Carles et al., who noted similar responses of circRNAs between FTD-tau (but not FTD-TDP-43) and AD (Cervera-Carles et al., 2020).

Using the 5xFAD genetic mouse model, which exhibits early  $A\beta$  plaque deposition and progressive cognitive decline (Forner et al., 2021), Wang et al., investigated circRNA alterations at critical disease stages (5 and 7 months). A-tailing RNAseq combined with the CARP algorithm revealed 342 and 467 differentially expressed (DE) circRNAs in the cortex at these time points, with additional 264 DE circRNAs in the hippocampus (Wang et al., 2024). To determine human relevance, they compared these findings with RNAseq data from the Mount Sinai Brain Bank (MSBB), confirming that many DE circRNAs were also dysregulated in AD patient brains. One of the most significantly altered circRNAs, *circGigyf2*, was highly expressed in healthy brains but showed marked downregulation at 7 months in 5xFAD mice and in human AD cortical regions. Notably, *circGigyf2* interacts with CPSF6, a critical RNA-processing factor involved in 3'-end

cleavage and polyadenylation. Its loss correlated with increased CPSF6 availability, leading to altered poly-(A)-site (PAS) selection and disrupted gene expression in thousands of transcripts, suggesting a role in AD pathogenesis. They also observed alternative splicing events within circRNAs, affecting their interactions with miRNAs and RBPs. For example, *circCcdc50* exhibited exon 4 skipping in the 7-month 5xFAD cortex, impacting *miR-335-5p*, a miRNA linked to AD-related gene expression changes. More broadly, miRNA sequestration appeared to be dynamically regulated by circRNA clusters, where multiple low-abundance circRNAs collectively influenced miRNA activity. Although individual circRNAs within a cluster may not achieve statistical significance in DE analysis, they can collectively function as a regulatory unit.

Additionally, Li et al. found that *circAXL* plays a key role in regulating BACE1, a protein associated with amyloid-beta production (Li et al., 2022). Over-expression of *circAXL*, which, in turn, negatively regulated *miR-328*, led to increased neuronal apoptosis, reduced neurite outgrowth, and heightened inflammation in AD models. In contrast, *miR-328* over-expression had protective effects, suggesting that *circAXL* and its modulation of BACE1 through *miR-328* may serve as potential therapeutic targets for AD.

These findings demonstrate that circRNAs are not passive byproducts of splicing but active regulators of gene expression in AD, as they affect both miRNA function and RNA-processing pathways. Their brain region-specific disruptions observed in both mice and AD patients highlight their potential as biomarkers and therapeutic targets in neurodegeneration.

### Parkinson's disease

Secondly to AD, PD is one of the most common neurodegenerative disorders with loss of dopamine production from substantia nigra dopaminergic neurons, leading to overt motor impairment and psychological decline. RNAseq studies on PD mouse models highlighted several dysregulated circRNAs, among which circSV2b which was further studied. Evidence shows that its over-expression, in PD induced mice, results in the restoration of dopamine synthesis and a normal nigro-striatal function. CircSV2b regulates the activity of mirR-5107-5p which modulates the transcription of Foxk1 mRNA and acts on the Akt1 signaling pathway, participating in PD-related oxidation. These data suggest that circSV2b may not only be investigated to understand the pathogenesis of PD, but could also be a promising candidate for PD treatment or even exploited as a molecular biomarker (Cheng et al., 2022). Furthermore, studies on PD patients revealed an overexpression of circSLC8A1, predicted to bind mir-128, in patients. Over-expression of this circRNA is strongly linked to the oxidative stress, a crucial characteristic of PD and most neurodegenerative disorders (Hanan et al., 2020).

## Frontotemporal dementia

The human microtubule-associated protein tau (MAPT) stabilizes microtubules in neurons through up to four

microtubule binding repeats, mainly formed by exons 7-12. Exon 10 - encoding the second microtubule-binding repeat - is alternatively spliced in the adult human brain leading to a mixture of tau proteins with 3 or 4 microtubule binding repeats. This ratio can be altered in disease, especially in several forms of frontotemporal dementia with parkinsonism linked to chromosome 17 (FTDP-17) (Corsi et al., 2022). Margvelani et al. investigated the impact of FTDP-17 mutations on the formation and translation of circular RNAs derived from the MAPT gene (Margvelani et al., 2023). Interestingly, five FTDP-17 mutations increased the formation of circRNA involving back-splicing from exon 12 to exon 7 (circTau (7,8,9,10,11,12)), while 3 other different mutations elevated the levels of circRNA from exon 12 to exon 10 (circTau(10,11,12)). These circRNAs undergo RNA editing through the interferon-induced ADAR1-p150 isoform which promotes the translation of circTau RNAs.

Atrian et al. further support the role of circRNAs and N6methyladenosine (m6A) modification, a key epitranscriptome mark affecting RNA stability and processing, in tau-induced neurotoxicity, by using both Drosophila models and human induced pluripotent stem cell (iPSC)-derived neurons. RNAseq analysis found, among others, *circMbl* to be significantly upregulated in tauopathy models. Depletion of circMbl reduced tau-induced cell death in Drosophila neurons, suggesting that increased *circMbl* contributes to neurotoxicity. Further experiments revealed an overall increase in m6A levels, which led to enhanced back-splicing and circRNA biogenesis in tau transgenic Drosophila. By m6A-RIP sequencing, tau-induced circMbl upregulation was confirmed to be m6A-dependent with suppression of RNA methylation factors mitigating tau-induced neurotoxicity. Taken together, these findings underscore a complex interplay between RNA editing and FTDP-17 mutations in regulating circRNA dynamics and their potential contribution to disease pathology. On the other hand, they also highlight a novel molecular mechanism where tau-driven m6A modifications might promote circRNA formation and contribute to neurotoxicity, thus opening a possible therapeutic strategy for tauopathies (Atrian et al., 2024).

### Amyotrophic lateral sclerosis

Amyotrophic lateral sclerosis (ALS) is a progressive neurodegenerative disease affecting both upper and lower motor neurons, leading to muscle atrophy, speech difficulties, and respiratory failure (Brown and Al-Chalabi, 2017). With an average life expectancy of just 30 months from symptom onset, early and accurate diagnosis is critical (Hardiman et al., 2017). Given the variability in clinical presentation, especially in patients without genetic mutations or familial history, reliable biomarkers are essential for timely detection and intervention. There are very few studies available exploring the role of circRNAs in ALS, with most focusing on their dysregulation rather than their functional significance. For instance, Dolinar et al. identified 274 upregulated and 151 downregulated circRNAs in ALS patients compared to healthy controls using microarray assays. Based on these findings and the functional relevance of their host genes in ALS, 10 circRNAs were selected for further PCR validation in a larger sample set. Of the

ones experimentally validated, two are of particular interest based on their host gene identity, namely *hsa\_circ\_0000567*, which is derived from *SETD3*, linked to muscle differentiation, and *hsa\_circ\_ 0023919*, derived from *PICALM*, which plays a role in clathrinmediated endocytosis at neuromuscular junctions and has been associated with AD. Notably, all selected circRNAs contain predicted binding sites for one or more RBPs. Further analysis revealed that *hsa\_circ\_0000567*, *hsa\_circ\_0023919*, and *hsa\_circ\_ 0088036* were negatively correlated with patient age at blood collection, while *hsa\_circ\_0000567* and *hsa\_circ\_0088036* were also negatively associated with the age of disease onset (Dolinar et al., 2019).

Aquilina-Reid et al. assessed circRNA expression levels in spinal cord tissues and explored the functional interactions between circRNAs, miRNAs and mRNAs in ALS, identifying 92 differentially expressed circRNAs (Aquilina-Reid et al., 2022). TargetScanHuman database based prediction of targeted mRNAs by selected miRNAs, then, revealed 55 genes whose targets were both significantly upregulated and differentially expressed in ALS (Friedman et al., 2009; Garcia et al., 2011; Grimson et al., 2007; Lewis et al., 2005). These genes were enriched for Gene Ontology terms such as "cell-cell signaling by Wnt" and "positive regulation of proteasomal ubiquitin-dependent protein catabolic process," both linked to ALS pathology (Chen et al., 2012).

Taken together these findings first demonstrate the potential significance of circRNA in ALS progression and for biomarker development, although follow up experiments are needed to validate these initial observations.

### Huntington's disease

Huntington's disease (HD) is a hereditary, fatal neurodegenerative disorder caused by a CAG trinucleotide expansion within exon 1 of the HTT gene. A polymorphic CAG tract up to 35 repeats is found in unaffected individuals, whereas alleles bearing 39 or more repeats lead to HD symptoms. Among the various dysfunctional mechanisms linked to the disease, RNA processing and alternative splicing (AS) alterations are emerging as crucial regulators of HD (Nguyen et al., 2025). Coherently, as the splicing machinery is also instrumental to circRNA biogenesis, different evidence is suggesting a functional role of circRNAs in HD. An initial study, using a circRNA microarray, compared expression patterns of rat cell lines expressing mutant or wildtype huntingtin (Marfil-Marin et al., 2021). A total of 23 circRNAs were differentially expressed, most of which downregulated. CircRNA/miRNA/mRNAs interactions were predicted, suggesting that changes in these regulatory networks might impact neuronal function and survival in HD. Capitalizing on accurate genetic mouse models of the disease, Ayyildiz et al. examined AS and circRNA biogenesis in HD by RNAseq approach (Ayyildiz et al., 2023). While circRNA expression was generally higher in NPCs compared to embryonic stem cells (ESC), cells expressing mutant huntingtin exhibit a mis-shaped repertoire of linear and back-spliced circular RNA isoforms. A negative correlation between CAG length and circRNA expression was noted in NPCs, suggesting a mutationdependent disruption of circularization. A significant proportion (32.7%) of differentially expressed circRNAs were found to be modified by m6A. This indicates that mutant huntingtin may impact circRNA biogenesis through m6A-mediated regulation. Interestingly, these splicing defects were most prominent in neural progenitors, with minimal changes observed in ESCs, highlighting the early vulnerability of neural cells to RNA dysregulation in HD. Interestingly, a highly conserved circRNA is also stemming from the HD gene locus itself, circHTT(2,3,4,5,6). CircHTT(2,3,4,5,6) expression significantly correlates with the length of the CAG-repeat tract in exon-1 of HTT in human and mouse HD model systems. Mouse circHtt(2,3,4,5,6), is expressed during embryogenesis and increases during nervous system development. CircHtt(2,3,4,5,6) over-expression experiments in the HD-relevant STHdh striatal cells further revealed its ability to modulate CAG expansion-driven cellular defects in cell-to-substrate adhesion, acting as novel modifier of HD pathology and potential drug target (Morandell et al., 2024). These three studies underscore the potential for targeting splicing, back-splicing and RNA modifications as therapeutic strategies in HD.

# CircRNAs in other disorders of the CNS

Deregulation of circRNAs has also been described in the context of other disorders of the central nervous system, that are not strictly of neurodevelopmental or neurodegenerative nature. These disorders range from epilepsy, depression and bipolar disorder, the inflammatory disease multiple sclerosis, to brain tumors. All these conditions are similarly complex and heterogeneous in their etiology, phenotypic composition and therapeutic management, as the typical neurodevelopmental and neurodegenerative disorders.

### Temporal lobe epilepsy

CircRNAs can regulate synaptogenesis and synaptic transmission, thus they are directly linked with neural network activity and the excitation-inhibition interplay in the brain. In line with this notion, numerous circRNAs have been investigated to provide a better understanding of the pathogenesis of mesial temporal lobe epilepsy (mTLE), the most common form of refractory focal epilepsy. Through high-throughput sequencing of mTLE patients-derived samples, thousands of circRNAs were identified; circSATB1, circSATB2 and cirLRP6 showed an interesting downregulation in the hippocampus of patients when compared to controls. Functionally, circSatb1 regulates dendritic spine morphology and function (Venø et al., 2015). The deregulated expression of circRNAs in mTLE was also confirmed by Gomes-Duarte et al., highlighting aberrant increase in *circArhgap4* and circNav3 expression from early to late stage of experimental epilepsy, likely as consequence of recurrent spontaneous seizures. Abnormal electrical brain activity, in turn, could be potentially correlated to the bioavailability of miRNA pathophysiologic drivers, binding targets of the dysregulated circRNAs with predicted miRNA-sponging activity (Gomes-Duarte et al., 2021). The hypothesis that circRNAs may interfere with the abundance and availability of miRNAs in the context of TLE is particularly relevant since different studies previously reported dysregulated miRNA levels in TLE patients' circulation, miRNAs as disease modifiers as well as

their importance for drug resistance in TLE [reviewed here (Guarnieri et al., 2024)].

#### Depression

Major depressive disorder (MDD) is a very common disorder, severely limiting psychosocial functioning and diminishing quality of life of affected individuals. The WHO ranked MDD as the third cause of burden of disease worldwide and projected that it will rank first by 2030 (Malhi and Mann, 2018). The etiology of MDD is believed to be multifactorial, with biological, genetic, environmental, and psychosocial contributing factors (Cui et al., 2024). Pathophysiologic mechanisms include neuroinflammation and alterations in neuroplasticity and epigenetics. Several long noncoding RNAs, including miRNAs and circRNAs, have been found to be deregulated in MDD, and could represent potential biomarkers for clinical diagnosis and monitoring of the therapeutic outcomes (Shi et al., 2021b). A small pilot study on 129 participants showed that individuals affected by MDD have increased plasma levels of circHIPK2 (Yu et al., 2024), which is consistent with previous findings in a mouse model system of depression (Zhang et al., 2019). In this study, the authors also speculated about a possible involvement of circHIPK2 in the gut-brain axis: Previously implicated in astrocyte activation in brain physiology (Huang et al., 2017), alterations of circHIPK2-regulated astrocyte activity in response to changes in the microbiome could modify the MDD behavioral characteristics (Hao et al., 2022; Zhang et al., 2019).

Furthermore, data from samples of MDD patients and two mouse models of depression (chronic unpredictable stress (CUS) and lipopolysaccharide (LPS) models), revealed increased circMBML1 levels and reduced expression of circFKBP8 (Shi et al., 2021a), hsa\_circRNA\_103636 (Cui et al., 2016) and circDYM(4,5,6) (Zhang et al., 2020) in peripheral blood samples. Interestingly, overexpression of circDym(4,5,6) in the mouse hippocampus, regulated immune activation by sponging miR-9, which, in turn, led to a consequent decrease of microglial activation upon CUS and LPS treatment, a hallmark of MDD, with depression-like behaviors ameliorated in these mice (Zhang et al., 2020). These findings suggest that, i) peripheral blood circDYM(4,5,6) levels may represent a promising biomarker, and ii) circDYM(4,5,6) overexpression may represent a novel treatment strategy for MDD. While these represent exciting initial observations, follow up studies in independent patient cohorts, alternative mouse models of depression and human in vitro model systems are imperative to assess the true value of these findings for clinical translation.

## Bipolar disorder

Bipolar disorder (BD) is a psychiatric disorder characterized by the alternation of depressive and maniacal phases during the lifetime of the affected individuals (Tondo et al., 2017). In recent years, studies explored the role of gene regulation and non-coding RNAs in the pathophysiology of BD. Analyzing the expression of circRNAs in peripheral blood mononuclear cells (PBMCs) from patients with BD, in comparison to samples from a non-psychiatric group, found around 33 circRNAs to be significantly dysregulated (Mahmoudi et al., 2021). Additionally, RNAseq analysis on samples from anterior cingulate cortex of individuals with BD compared to neurotypical controls, identified several dysregulated circRNAs among which, *circCCNT2* showed a significant upregulation in the BD cohort (Lin et al., 2021). Many studies focused on the function of *circHomer1* as a psychiatric disease-related circular RNA: its expression is reduced in BD as well as other diseases such as SCZ. Using knock-down mouse models, low levels of *circHomer1* were shown to be associated with changes in animal cognitive ability and behaviour, possibly explained by *circHomer1* ability to modulate the expression of genes involved in synaptic plasticity, thus affecting neuronal function (Zimmerman et al., 2020, 2024).

The differential expression of *circNEBL* and *circEPHA3* - altered in RNAseq data from *post-mortem* brains of BD patients versus healthy controls - could further provide new insights into the role of circRNAs in BD (Luykx et al., 2019). Specifically, the upregulation of *circEPHA3* may be of particular relevance since EPHA3's role in mediating axon growth (Javier-Torrent et al., 2019) could be linked to the neurodevelopmental aspects of BD.

#### Multiple sclerosis

Multiple sclerosis (MS) is a complex autoimmune disease of the CNS characterized by demyelination, chronic inflammation, neuronal loss, and axonal damage (Thompson et al., 2018). The most common form, affecting 80% of patients, is relapsing-remitting MS, characterized by attacks followed by complete or partial remissions (Lublin et al., 2014). Although the pathogenic factors underlying MS remain largely unknown, several recent studies pointed to alterations in AS and RNA processing as new molecular mechanisms potentially involved in the pathogenesis. Coherently, some AS events as well as ncRNAs have been proposed as novel biomarkers for MS (D'Ambrosio et al., 2015; Hecker et al., 2019; Paraboschi et al., 2015; Spurlock et al., 2015). While functional investigation of circRNAs has been largely neglected, circRNAs were first implicated in MS by Cardamone et al., identifying the GSMDB derived hsa\_circ\_0106803 circRNA as upregulated in PBMCs of 30 RR-MS patients compared to 30 healthy controls (Cardamone et al., 2017). Subsequently, Iparraguirre and colleagues detected 406 differentially expressed circRNAs through a microarray analysis on peripheral blood leucocytes of four RR-MS patients and four healthy controls; two downregulated circRNAs, both deriving from the ANXA2 host gene, were validated in an independent cohort of 20 RR-MS patients and 18 healthy controls (Iparraguirre et al., 2017). In addition, through an RNAseq based analysis of blood samples, a global, specific and sex-dependent deregulation of circRNA expression in MS was observed in a cohort of 30 patients and 20 controls, with candidate circRNAs further examined in a second, independent cohort of 70 M cases, revealing 6 upregulated circRNAs (e.g., circPADI4) as potential biomarkers (Iparraguirre et al., 2020). CircRNAs were furthermore shown to be enriched at MS loci, from genome wide studies, with further emerging characterization of hsa\_circ\_0043813 derived from the MSassociated STAT3 gene (Paraboschi et al., 2018). Similarly, expression analysis in another patient cohort, revealed

166 differentially expressed circRNAs in MS patients, 125 of which were downregulated. One of the top dysregulated circRNAs, *hsa\_circ\_0007990*, derives from the *PGAP3* gene, relevant for the control of autoimmune responses. Later analysis of two independent replication cohorts confirmed the downregulation of *circPGAP3* suggesting its implementation as a possible RNA-based biomarker (Cardamone et al., 2023).

### Brain tumor development

CircRNAs play a complex role in cancer development, influencing tumor progression through exosomal transfer and facilitating communication between cancer cells and the surrounding microenvironment. Certain circRNAs can contribute to metastasis by promoting pre-metastatic niches (Dai et al., 2018; Wang et al., 2019). Additionally, the failure of transcription termination and uncontrolled gene transcription into the downstream gene followed by back-splicing can give rise to readthrough circRNAs, which incorporate exons from adjacent genes (Liang et al., 2017; Vo et al., 2019). These aberrant circRNAs are associated with oncogenic processes, potentially driving tumor growth and malignant transformation.

Invasion and metastasis of tumor cells are the primary causes of mortality in patients with malignant tumors (Asif et al., 2021). Despite the growing interest in circRNAs in cancer, few studies specifically address their role in brain tumor development (Ahmed et al., 2022; Katsushima et al., 2023; Zhang et al., 2024). Among the multiple pathways implicated in circRNA-mediated tumorigenesis, such as p53, JAK/STAT, and ERK signaling, the PI3K/AKT/mTOR axis is particularly significant.

mTOR is a key downstream signaling molecule in the AMPK, MAPK, and PI3K/AKT pathways, playing a central role in cancer progression (Tee et al., 2003). PI3K activation promotes the transition from PIP2 to PIP3, a process reversed by PTEN, and subsequently leads to mTORC1 and mTORC2 activation, either directly or through Rheb regulation via TSC1/2 inhibition (Ma et al., 2005).

Xin et al. (2019) identified hsa\_circ\_0067934 as a critical modulator of glioblastoma progression through its regulation of the epithelial-mesenchymal transition (EMT) process and the PI3K/ AKT/mTOR signaling pathway. In their study, they analyzed 157 human brain samples from glioblastoma patients and healthy controls. qRT-PCR revealed that hsa\_circ\_0067934 was significantly overexpressed in glioblastoma tissues compared to adjacent noncancerous tissues. To investigate the functional role of hsa\_ circ\_0067934, siRNA-mediated knock-down experiments were conducted in vitro. The results demonstrated a significant reduction in cell proliferation of glioblastoma cell lines posttransfection, while apoptosis levels increased notably upon hsa\_ silencing. Hsa\_circ\_0067934 circ\_0067934 knock-down, furthermore, significantly impaired the migratory ability of glioblastoma cells in wound healing assays. At molecular level, hsa\_circ\_0067934 silencing resulted in a marked decrease of phosphorylated PI3K and phosphorylated AKT, while total PI3K and AKT levels remained unchanged. This suggests that hsa\_circ\_ 0067934 promotes glioblastoma progression by activating the PI3K-AKT signaling pathway (Xin et al., 2019).

Moreover, Bian et al., 2018 identified circ-CFH as a novel oncogenic circRNA, upregulated in glioma tissues. Hsa\_circ\_ 0015758 (circ-CFH) is a single exon-derived circRNA transcribed from the GRCh37/hg19 fragment of chromosome 1, sharing homology with the protein-coding gene complement factor H (CFH) and functioning as a miR-149 sponge, thereby upregulating AKT1 expression. To investigate the biological role of circ-CFH in glioma cells, siRNA-mediated knock-down was performed. Silencing circ-CFH significantly decreased cell proliferation and cells' ability to form colonies in vitro, while subcutaneous injection of circ-CFH -suppressing cells into nude mice significantly decreased tumor volume and weight. Finally, the RNA sponging activity of circ-CFH was explored, confirming miR-149 as a direct target through luciferase reporter assays, regulating miR-149-target mRNAs, such as AKT1 mRNA (Bian et al., 2018). Taken together, these studies highlight the possibility for circRNAs targeting in order to modulate the activity of the PI3K/AKTAkt/ mTOR signaling pathway in the context of brain tumor development.

# CircRNAs as biomarkers for brain disorders

Circularity renders RNA largely resistant to exonuclease cleavage and, therefore, circRNAs are extremely stable, as demonstrated by their long half lives in cells (Memczak et al., 2015). Furthermore, they show remarkable tissue specificity in their expression patterns, can be secreted into the extracellular space via extracellular vesicles, - both exosomes (30-100 nm) and microvesicles (100-1,000 nm) (Hon et al., 2019; Lasda and Parker, 2016; Zhao et al., 2024), and have previously been detected in easily accessible biofluids such as saliva, blood and urine (Hutchins et al., 2021; Memczak et al., 2015; Zhao et al., 2018). All these characteristics render circRNAs ideal candidates for being reliable biomarkers. Biomarkers can support diagnosis, provide information about disease prognosis, and progression, and can be used to monitor the efficacy and safety of drug therapy. In fact, when it comes to complex brain disorders, reliable peripheral biomarkers would be extremely useful. Disease relevant circRNAs could enter the blood via blood brain barrier leakage and be measured in biofluids. They could aid 1) diagnosis, which, for instance, is still very difficult in ASD and ALS (Hus and Segal, 2021), 2) disease progression and staging, which is critical for clinical trial design and endpoint assessment, 3) patient care, as sampling is much less invasive when compared with the collection of cerebrospinal fluid, as commonly done in HD (Pellegrini et al., 2022). In line with these requirements, recently, a study utilizing two large publicly available datasets with longitudinal clinical and blood transcriptomic data of PD patients, identified 71 circRNAs that were sufficient to differentiate between genetic PD, at riskindividuals and controls, highlighting the potential of these circRNAs to aid PD diagnosis. Importantly, five circRNAs significantly correlated with PD severity, yet the identified circRNAs had no better predictive value than linear RNA, limiting their utility (Beric et al., 2024). This example, amongst many others discussed above, highlights important considerations in using circRNAs as biomarkers for complex brain disorders. First,



well as the production of circRNA related peptides through translation. When molecular mechanisms of duction are investigated, it is of utmost importance to first determine endogenous expression levels of the involved components in the disorder of interest, to be able to draw physiologically relevant conclusions. 4. Finally, findings obtained from studying *in vivo* and *in vitro* model systems need to be translated back to the clinics in order to be useful for patients suffering from complex neurologic disorders. To that end, evaluation of a true contribution of the circRNA molecular function to the pathophysiologic processes, either on initiation or progression of the disease or modulation thereof, and/or its potential as biomarker as well as novel drug target need to be carefully assessed and tested.

most studies that are available to date and propose individual circRNAs as biomarkers are based on small sample sizes, i.e., few patients, and limited datasets, and only few have validated their primary findings in additional, independent patient cohorts. Secondly, gender-differences have not been taken into consideration in the majority of cases, which could hamper their potential for clinical translation. And third, the tissue-origin of the deregulated circRNAs proposed as biomarkers remains unexplored, thus it is unclear whether they truly represent a "window into the diseased brain" or whether their deregulation is a secondary,

unspecific result. Therefore, more standardised and carefully designed studies, will clarify their true potential as biomarkers in the future and for other neurologic disorders.

# Moving forward: challenges and outstanding questions

The last decade has seen the circRNA research field explode, and experimental evidence highlighting the important regulatory

10.3389/fphar.2025.1577496

functions of these peculiar molecules for brain physiology and complex neurologic disorders has continuously piled up over the years. Nonetheless, despite the rightful excitement about the discovery of a new regulatory layer that could modulate still untreatable, severe and -in some cases- lethal disorders, measures need to be taken to validate findings derived from RNAseq studies of post mortem human, or model organism brain samples, demonstrating a clear functional role of the deregulated circRNAs in the pathogenesis, and/or disease progression by experimental scrutiny to avoid misinterpreting correlation with causation. The circRNA field thereby faces unique challenges, caused by the -mostly- low expression levels, extremely high sequence similarity with the linear RNA counterparts, as well as high stability of these molecules, in addition to issues related to a lack of standardized protocols and detection tools that limit the precision and specificity of experimental approaches. In Figure 2 we suggest a roadmap in moving the circRNA field from performing RNAseq studies, to being able to eventually translate discoveries back to the clinics in order to benefit patients' outcomes. Until very recently, the absence of a standardized circRNA nomenclature severely complicated the communication of results and replication of circRNA related data. This issue, however, was recently addressed by Chen et al. proposing guidelines for a unified circRNA nomenclature. Now the scientific community should push for the implementation of these guidelines, to benefit the circRNA research field as a whole (Chen et al., 2023). Additionally, best practice standards have been recently put forward by leading experts in the field, highlighting important pitfalls and considerations when designing experiments to ensure reproducibility of findings (Nielsen et al., 2022). Experimental strategies to identify regulatory functions are, currently, primarily based on circRNA depletion or over-expression approaches (Figure 2, step 2), which can give valuable first insights into the modulatory potential of the deregulated circRNAs. Since specific targeting is challenging when it comes to circRNAs, as their sequence is unique exclusively at the BSJ, extra care needs to be taken to avoid simultaneously altering the levels of linear RNA transcripts and splicing. When it comes to over-expression strategies, circRNAs can be either synthetically produced, or transcribed RNA or over-expression plasmids can be transfected into in vitro model systems. Next to issues relating to immune activation by the synthetic molecules, these strategies often also lead to delivery/production of linear cognate RNAs and extremely high expression levels of the circRNA of interest, far away from any physiological relevant levels. Thus, the results obtained by such overexpression approaches need to be critically evaluated and, ideally, confirmed with a second strategy.

In general, only few circRNA studies have been able to conclusively demonstrate the molecular action of function of the identified DE circRNA, but merely provide a description of the cellular effects of the modulation of their expression. In order to move the field forward, it will be necessary to thoroughly investigate the molecular mechanisms of action of the candidate circRNAs across different disorders (Figure 2, step 3) to objectively assess their true potential as druggable target, therapeutic agent or biomarker. Once these goals have been reached for a specific candidate circRNA, additional challenges need to be overcome. In case the expression of a candidate circRNA is beneficial and counteracts pathophysiological processes, then the circRNA would need to be overexpressed. Should, instead, a reduction of a candidate circRNA demonstrate desirable effects, knock-down or knock-out approaches would be needed. In either case, all the considerations for overexpression and knock-down strategies discussed above, need to be addressed for a successful translation into the clinics. Independently of a desired up-or downregulation, still, the best approach for reaching the brain needs to be identified. This requires a successful crossing of the blood-brain barrier of a therapeutic agent or small molecule, intracranial injections or intrathecal administration, the targeting of a specific brain region or cell type exclusively, while leaving surrounding areas/cells unaffected. Further limitations are arising from the potential toxicity of all viral and non viral approaches, which may either integrate into the host genome and cause insertional mutagenesis or cause inflammation in the brain. Only when all these hurdles are overcome, then, eventually, circRNAs may benefit patients affected by these complex brain disorders for real (Figure 2, step 4), a milestone that has yet to be achieved by this young research field.

## Author contributions

VG: Writing – original draft. SV: Writing – original draft. MB: Funding acquisition, Resources, Supervision, Writing – original draft. JM: Conceptualization, Funding acquisition, Project administration, Supervision, Writing – original draft.

# Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was supported by the University of Trento, the Huntington Society of Canada's NEW PATHWAYS award, the European Huntington's Disease Network (EHDN) seed fund project #1041 and the Hereditary Disease Foundation research grant #990845, the HD Human Biology Project 2018 by the Huntington's Disease Society of America (HDSA) and the MUR PNRR National Center RNA and gene Therapy Spoke 3, Bando a Cascata "RAPID" to MB. MB. was also a recipient of a Marie Skłodowska Curie reintegration fellowship (The European Union's Horizon 2020 Research and Innovation Program, GA n°706567). JM was/is supported by a EMBO postdoctoral fellowship (ALTF 897-2021) and by the European Union (under GA n°101062297- BrainCircHD).

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

## References

Ahmed, S. P., Castresana, J. S., and Shahi, M. H. (2022). Role of circular RNA in brain tumor development. *Cells* 11 (14), 2130. doi:10.3390/cells11142130

Aquilina-Reid, C., Brennan, S., Curry-Hyde, A., Teunisse, G. M., and Janitz, M. (2022). Circular RNA expression and interaction patterns are perturbed in amyotrophic lateral sclerosis. *Int. J. Mol. Sci.* 23 (23), 14665. doi:10.3390/ijms232314665

Arizaca Maquera, K. A., Welden, J. R., Margvelani, G., Miranda Sardón, S. C., Hart, S., Robil, N., et al. (2023). Alzheimer's disease pathogenetic progression is associated with changes in regulated retained introns and editing of circular RNAs. Front. Mol. Neurosci. 16, 1141079. doi:10.3389/fnmol.2023.1141079

Ashwal-Fluss, R., Meyer, M., Pamudurti, N. R., Ivanov, A., Bartok, O., Hanan, M., et al. (2014). circRNA biogenesis competes with pre-mRNA splicing. *Mol. Cell* 56 (1), 55–66. doi:10.1016/j.molcel.2014.08.019

Asif, P. J., Longobardi, C., Hahne, M., and Medema, J. P. (2021). The role of cancerassociated fibroblasts in cancer invasion and metastasis. *Cancers* 13 (18), 4720. doi:10. 3390/cancers13184720

Atrian, F., Ramirez, P., Mange, J. D., Marquez, M., Gonzalez, E. M., Minaya, M., et al. (2024). m6A-dependent circular RNA formation mediates tau-induced neurotoxicity. *bioRxiv*. doi:10.1101/2024.01.25.577211

Ayyildiz, D., Bergonzoni, G., Monziani, A., Tripathi, T., Döring, J., Kerschbamer, E., et al. (2023). CAG repeat expansion in the Huntington's disease gene shapes linear and circular RNAs biogenesis. *PLOS Genet.* 19 (10), e1010988. doi:10.1371/journal.pgen. 1010988

Belter, A., Popenda, M., Sajek, M., Woźniak, T., Naskręt-Barciszewska, M. Z., Szachniuk, M., et al. (2022). A new molecular mechanism of RNA circularization and the microRNA sponge formation. *J. Biomol. Struct. and Dyn.* 40 (7), 3038–3045. doi:10.1080/07391102.2020.1844802

Beric, A., Sun, Y., Sanchez, S., Martin, C., Powell, T., Kumar, R., et al. (2024). Circulating blood circular RNA in Parkinson's Disease; from involvement in pathology to diagnostic tools in at-risk individuals. *NPJ Parkinson's Dis.* 10 (1), 222. doi:10.1038/ s41531-024-00839-3

Bian, A., Wang, Y., Liu, J., Wang, X., Liu, D., Jiang, J., et al. (2018). Circular RNA complement factor H (CFH) promotes glioma progression by sponging miR-149 and regulating AKT1. *Med. Sci. Monit. Int. Med. J. Exp. Clin. Res.* 24, 5704–5712. doi:10. 12659/MSM.910180

Brown, R. H., and Al-Chalabi, A. (2017). Amyotrophic lateral sclerosis. N. Engl. J. Med. 377 (2), 162–172. doi:10.1056/NEJMra1603471

Cardamone, G., Paraboschi, E. M., Rimoldi, V., Duga, S., Soldà, G., and Asselta, R. (2017). The characterization of GSDMB splicing and backsplicing profiles identifies novel isoforms and a circular RNA that are dysregulated in multiple sclerosis. *Int. J. Mol. Sci.* 18 (3), 576. doi:10.3390/ijms18030576

Cardamone, G., Paraboschi, E. M., Soldà, G., Liberatore, G., Rimoldi, V., Cibella, J., et al. (2023). The circular RNA landscape in multiple sclerosis: disease-specific associated variants and exon methylation shape circular RNA expression profile. *Multiple Scler. Relat. Disord.* 69, 104426. doi:10.1016/j.msard.2022.104426

Cerda-Jara, C. A., Kim, S. J., Thomas, G., Farsi, Z., Zolotarov, G., Dube, G., et al. (2024). miR-7 controls glutamatergic transmission and neuronal connectivity in a Cdr1as-dependent manner. *EMBO Rep.* 25 (7), 3008–3039. doi:10.1038/s44319-024-00168-9

Cervera-Carles, L., Dols-Icardo, O., Molina-Porcel, L., Alcolea, D., Cervantes-Gonzalez, A., Muñoz-Llahuna, L., et al. (2020). Assessing circular RNAs in Alzheimer's disease and frontotemporal lobar degeneration. *Neurobiol. Aging* 92, 7–11. doi:10.1016/j.neurobiolaging.2020.03.017

Chen, C.-K., Cheng, R., Demeter, J., Chen, J., Weingarten-Gabbay, S., Jiang, L., et al. (2021). Structured elements drive extensive circular RNA translation. *Mol. Cell* 81 (20), 4300–4318.e13. doi:10.1016/j.molcel.2021.07.042

Chen, H.-H., Eteleeb, A., Wang, C., Fernandez, M. V., Budde, J. P., Bergmann, K., et al. (2022). Circular RNA detection identifies circPSEN1 alterations in brain specific to autosomal dominant Alzheimer's disease. *Acta Neuropathol. Commun.* 10 (1), 29. doi:10.1186/s40478-022-01328-5

Chen, L.-L., Bindereif, A., Bozzoni, I., Chang, H. Y., Matera, A. G., Gorospe, M., et al. (2023). A guide to naming eukaryotic circular RNAs. *Nat. Cell Biol.* 25 (1), 1–5. doi:10. 1038/s41556-022-01066-9

Chen, X., Han, P., Zhou, T., Guo, X., Song, X., and Li, Y. (2016). circRNADb: a comprehensive database for human circular RNAs with protein-coding annotations. *Sci. Rep.* 6 (1), 34985. doi:10.1038/srep34985

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Chen, Y., Guan, Y., Zhang, Z., Liu, H., Wang, S., Yu, L., et al. (2012). Wnt signaling pathway is involved in the pathogenesis of amyotrophic lateral sclerosis in adult transgenic mice. *Neurological Res.* 34 (4), 390–399. doi:10.1179/1743132812Y. 0000000027

Cheng, Q., Wang, J., Li, M., Fang, J., Ding, H., Meng, J., et al. (2022). CircSV2b participates in oxidative stress regulation through miR-5107-5p-Foxk1-Akt1 axis in Parkinson's disease. *Redox Biol.* 56, 102430. doi:10.1016/j.redox.2022.102430

Choi, L., and An, J.-Y. (2021). Genetic architecture of autism spectrum disorder: lessons from large-scale genomic studies. *Neurosci. Biobehav. Rev.* 128, 244–257. doi:10. 1016/j.neubiorev.2021.06.028

Cochran, K. R., Veeraraghavan, K., Kundu, G., Mazan-Mamczarz, K., Coletta, C., Thambisetty, M., et al. (2021). Systematic identification of circRNAs in Alzheimer's disease. *Genes* 12 (8), 1258. doi:10.3390/genes12081258

Corsi, A., Bombieri, C., Valenti, M. T., and Romanelli, M. G. (2022). Tau isoforms: gaining insight into MAPT alternative splicing. *Int. J. Mol. Sci.* 23 (23), 15383. doi:10. 3390/ijms232315383

Cui, L., Li, S., Wang, S., Wu, X., Liu, Y., Yu, W., et al. (2024). Major depressive disorder: hypothesis, mechanism, prevention and treatment. *Signal Transduct. Target. Ther.* 9 (1), 30–32. doi:10.1038/s41392-024-01738-y

Cui, X., Niu, W., Kong, L., He, M., Jiang, K., Chen, S., et al. (2016). hsa\_circRNA\_ 103636: potential novel diagnostic and therapeutic biomarker in Major depressive disorder. *Biomarkers Med.* 10 (9), 943–952. doi:10.2217/bmm-2016-0130

Dai, X., Chen, C., Yang, Q., Xue, J., Chen, X., Sun, B., et al. (2018). Exosomal circRNA\_100284 from arsenite-transformed cells, via microRNA-217 regulation of EZH2, is involved in the malignant transformation of human hepatic cells by accelerating the cell cycle and promoting cell proliferation. *Cell Death and Dis.* 9 (5), 454. doi:10.1038/s41419-018-0485-1

D'Ambrosio, A., Pontecorvo, S., Colasanti, T., Zamboni, S., Francia, A., and Margutti, P. (2015). Peripheral blood biomarkers in multiple sclerosis. *Autoimmun. Rev.* 14 (12), 1097–1110. doi:10.1016/j.autrev.2015.07.014

Dolinar, A., Koritnik, B., Glavač, D., and Ravnik-Glavač, M. (2019). Circular RNAs as potential blood biomarkers in amyotrophic lateral sclerosis. *Mol. Neurobiol.* 56 (12), 8052–8062. doi:10.1007/s12035-019-1627-x

Dong, R., Ma, X.-K., Li, G.-W., and Yang, L. (2018). CIRCpedia v2: an updated database for comprehensive circular RNA annotation and expression comparison. *Genomics, Proteomics and Bioinforma.* 16 (4), 226–233. doi:10.1016/j.gpb.2018.08.001

Dong, X., Bai, Y., Liao, Z., Gritsch, D., Liu, X., Wang, T., et al. (2023). Circular RNAs in the human brain are tailored to neuron identity and neuropsychiatric disease. *Nat. Commun.* 14 (1), 5327. doi:10.1038/s41467-023-40348-0

Dube, U., Del-Aguila, J. L., Li, Z., Budde, J. P., Jiang, S., Hsu, S., et al. (2019). An atlas of cortical circular RNA expression in Alzheimer disease brains demonstrates clinical and pathological associations. *Nat. Neurosci.* 22 (11), 1903–1912. doi:10.1038/s41593-019-0501-5

Enuka, Y., Lauriola, M., Feldman, M. E., Sas-Chen, A., Ulitsky, I., and Yarden, Y. (2016). Circular RNAs are long-lived and display only minimal early alterations in response to a growth factor. *Nucleic Acids Res.* 44 (3), 1370–1383. doi:10.1093/nar/gkv1367

Fan, X., Zhang, X., Wu, X., Guo, H., Hu, Y., Tang, F., et al. (2015). Single-cell RNA-seq transcriptome analysis of linear and circular RNAs in mouse preimplantation embryos. *Genome Biol.* 16 (1), 148. doi:10.1186/s13059-015-0706-1

Forner, S., Kawauchi, S., Balderrama-Gutierrez, G., Kramár, E. A., Matheos, D. P., Phan, J., et al. (2021). Systematic phenotyping and characterization of the 5xFAD mouse model of Alzheimer's disease. *Sci. Data* 8 (1), 270. doi:10.1038/s41597-021-01054-y

Friedman, R. C., Farh, K. K.-H., Burge, C. B., and Bartel, D. P. (2009). Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res.* 19 (1), 92–105. doi:10.1101/gr.082701.108

Garcia, D. M., Baek, D., Shin, C., Bell, G. W., Grimson, A., and Bartel, D. P. (2011). Weak seed-pairing stability and high target-site abundance decrease the proficiency of lsy-6 and other microRNAs. *Nat. Struct. and Mol. Biol.* 18 (10), 1139–1146. doi:10.1038/ nsmb.2115

Gasparini, S., Del Vecchio, G., Gioiosa, S., Flati, T., Castrignano, T., Legnini, I., et al. (2020). Differential expression of hippocampal circular RNAs in the BTBR mouse model for autism spectrum disorder. *Mol. Neurobiol.* 57 (5), 2301–2313. doi:10.1007/s12035-020-01878-6

Ghafouri-Fard, S., Noroozi, R., Brand, S., Hussen, B. M., Eghtedarian, R., Taheri, M., et al. (2022). Emerging role of non-coding RNAs in autism spectrum disorder. *J. Mol. Neurosci.* 72 (2), 201–216. doi:10.1007/s12031-021-01934-3

Giusti, S. A., Pino, N. S., Pannunzio, C., Ogando, M. B., Armando, N. G., Garrett, L., et al. (2024). A brain-enriched circular RNA controls excitatory neurotransmission and restricts sensitivity to aversive stimuli. *Sci. Adv.* 10 (21), eadj8769. doi:10.1126/sciadv. adj8769

Glažar, P., Papavasileiou, P., and Rajewsky, N. (2014). circBase: a database for circular RNAs. RNA (New York, N.Y.) 20 (11), 1666–1670. doi:10.1261/rna.043687.113

Gomes-Duarte, A., Bauer, S., Venø, M. T., Norwood, B. A., Henshall, D. C., Kjems, J., et al. (2021). Enrichment of circular RNA expression deregulation at the transition to recurrent spontaneous seizures in experimental temporal lobe epilepsy. *Front. Genet.* 12, 627907. doi:10.3389/fgene.2021.627907

Grimson, A., Farh, K. K.-H., Johnston, W. K., Garrett-Engele, P., Lim, L. P., and Bartel, D. P. (2007). MicroRNA targeting specificity in mammals: determinants beyond seed pairing. *Mol. Cell* 27 (1), 91–105. doi:10.1016/j.molcel.2007.06.017

Guarnieri, L., Amodio, N., Bosco, F., Carpi, S., Tallarico, M., Gallelli, L., et al. (2024). Circulating miRNAs as novel clinical biomarkers in temporal lobe epilepsy. *Non-Coding RNA* 10 (2), 18. doi:10.3390/ncrna10020018

Hanan, M., Simchovitz, A., Yayon, N., Vaknine, S., Cohen-Fultheim, R., Karmon, M., et al. (2020). A Parkinson's disease CirCRNAs Resource reveals a link between cirCSLC8A1 and oxidative stress. *EMBO Mol. Med.* 12 (9), e11942. doi:10.15252/ emmm.201911942

Hanan, M., Soreq, H., and Kadener, S. (2017). CircRNAs in the brain. RNA Biol. 14 (8), 1028–1034. doi:10.1080/15476286.2016.1255398

Hansen, T. B., Jensen, T. I., Clausen, B. H., Bramsen, J. B., Finsen, B., Damgaard, C. K., et al. (2013). Natural RNA circles function as efficient microRNA sponges. *Nature* 495 (7441), 384–388. doi:10.1038/nature11993

Hao, W.-Z., Chen, Q., Wang, L., Tao, G., Gan, H., Deng, L.-J., et al. (2022). Emerging roles of long non-coding RNA in depression. *Prog. Neuro-Psychopharmacology Biol. Psychiatry* 115, 110515. doi:10.1016/j.pnpbp.2022.110515

Hardiman, O., Al-Chalabi, A., Brayne, C., Beghi, E., van den Berg, L. H., Chio, A., et al. (2017). The changing picture of amyotrophic lateral sclerosis: lessons from European registers. *J. Neurology, Neurosurg. Psychiatry* 88 (7), 557–563. doi:10.1136/jnnp-2016-314495

Hecker, M., Rüge, A., Putscher, E., Boxberger, N., Rommer, P. S., Fitzner, B., et al. (2019). Aberrant expression of alternative splicing variants in multiple sclerosis—a systematic review. *Autoimmun. Rev.* 18 (7), 721–732. doi:10.1016/j.autrev.2019.05.010

Holdt, L. M., Stahringer, A., Sass, K., Pichler, G., Kulak, N. A., Wilfert, W., et al. (2016). Circular non-coding RNA ANRIL modulates ribosomal RNA maturation and atherosclerosis in humans. *Nat. Commun.* 7 (1), 12429. doi:10.1038/ncomms12429

Hollensen, A. K., Thomsen, H. S., Lloret-Llinares, M., Kamstrup, A. B., Jensen, J. M., Luckmann, M., et al. (2020). circZNF827 nucleates a transcription inhibitory complex to balance neuronal differentiation. *eLife* 9, e58478. doi:10.7554/eLife.58478

Hon, K. W., Ab-Mutalib, N. S., Abdullah, N. M. A., Jamal, R., and Abu, N. (2019). Extracellular Vesicle-derived circular RNAs confers chemoresistance in Colorectal cancer. *Sci. Rep.* 9 (1), 16497. doi:10.1038/s41598-019-53063-y

Hou, Y., Dan, X., Babbar, M., Wei, Y., Hasselbalch, S. G., Croteau, D. L., et al. (2019). Ageing as a risk factor for neurodegenerative disease. *Nat. Rev. Neurol.* 15 (10), 565–581. doi:10.1038/s41582-019-0244-7

Huang, H., Luo, J., Qi, Y., Wu, Y., Qi, J., Yan, X., et al. (2023). Comprehensive analysis of circRNA expression profile and circRNA-miRNA-mRNA network susceptibility to very early-onset schizophrenia. *Schizophrenia* 9 (1), 70. doi:10.1038/s41537-023-00399-0

Huang, R., Zhang, Y., Han, B., Bai, Y., Zhou, R., Gan, G., et al. (2017). Circular RNA HIPK2 regulates astrocyte activation via cooperation of autophagy and ER stress by targeting MIR124–2HG. *Autophagy* 13 (10), 1722–1741. doi:10.1080/15548627.2017. 1356975

Hus, Y., and Segal, O. (2021). Challenges surrounding the diagnosis of autism in children. *Neuropsychiatric Dis. Treat.* 17, 3509–3529. doi:10.2147/NDT.S282569

Hutchins, E., Reiman, R., Winarta, J., Beecroft, T., Richholt, R., De Both, M., et al. (2021). Extracellular circular RNA profiles in plasma and urine of healthy, male college athletes. *Sci. Data* 8 (1), 276. doi:10.1038/s41597-021-01056-w

Hwang, H. J., and Kim, Y. K. (2024). Molecular mechanisms of circular RNA translation. *Exp. and Mol. Med.* 56 (6), 1272–1280. doi:10.1038/s12276-024-01220-3

Ikeda, Y., Morikawa, S., Nakashima, M., Yoshikawa, S., Taniguchi, K., Sawamura, H., et al. (2023). CircRNAs and RNA-binding proteins involved in the pathogenesis of cancers or central nervous system disorders. *Non-Coding RNA* 9 (2), 23. doi:10.3390/ncrna9020023

Iparraguirre, L., Alberro, A., Sepúlveda, L., Osorio-Querejeta, I., Moles, L., Castillo-Triviño, T., et al. (2020). RNA-Seq profiling of leukocytes reveals a sex-dependent global circular RNA upregulation in multiple sclerosis and 6 candidate biomarkers. *Hum. Mol. Genet.* 29 (20), 3361–3372. doi:10.1093/hmg/ddaa219

Iparraguirre, L., Muñoz-Culla, M., Prada-Luengo, I., Castillo-Triviño, T., Olascoaga, J., and Otaegui, D. (2017). Circular RNA profiling reveals that circular RNAs from

ANXA2 can be used as new biomarkers for multiple sclerosis. *Hum. Mol. Genet.* 26 (18), 3564–3572. doi:10.1093/hmg/ddx243

Javier-Torrent, M., Marco, S., Rocandio, D., Pons-Vizcarra, M., Janes, P. W., Lackmann, M., et al. (2019). Presenilin/γ-secretase-dependent EphA3 processing mediates axon elongation through non-muscle myosin IIA. *eLife* 8, e43646. doi:10. 7554/eLife.43646

Jeck, W. R., Sorrentino, J. A., Wang, K., Slevin, M. K., Burd, C. E., Liu, J., et al. (2013). Circular RNAs are abundant, conserved, and associated with ALU repeats. RNA (New York, N.Y.) 19 (2), 141–157. doi:10.1261/rna.035667.112

Jiang, M., Wang, Z., Lu, T., Li, X., Yang, K., Zhao, L., et al. (2023). Integrative analysis of long noncoding RNAs dysregulation and synapse-associated ceRNA regulatory axes in autism. *Transl. Psychiatry* 13 (1), 375. doi:10.1038/s41398-023-02662-5

Katsushima, K., Joshi, K., and Perera, R. J. (2023). Diagnostic and therapeutic potential of circular RNA in brain tumors. *Neuro-Oncology Adv.* 5 (1), vdad063. doi:10.1093/noajnl/vdad063

Kelly, D., Bicker, S., Winterer, J., Nanda, P., Germain, P.-L., Dieterich, C., et al. (2024). A functional screen uncovers circular RNAs regulating excitatory synaptogenesis in hippocampal neurons. *bioRxiv*. doi:10.1101/2024.04.09.588686

Kelly, D., Bicker, S., Winterer, J., Nanda, P., Germain, P.-L., Dieterich, C., et al. (2025). A functional screen uncovers circular RNAs regulating excitatory synaptogenesis in hippocampal neurons. *Nat. Commun.* 16 (1), 3040. doi:10.1038/s41467-025-58070-4

Kleaveland, B., Shi, C. Y., Stefano, J., and Bartel, D. P. (2018). A network of noncoding regulatory RNAs acts in the mammalian brain. *Cell* 174 (2), 350–362.e17. doi:10.1016/j. cell.2018.05.022

Knupp, D., and Miura, P. (2018). CircRNA accumulation: a new hallmark of aging? Mech. Ageing Dev. 173, 71–79. doi:10.1016/j.mad.2018.05.001

Lasda, E., and Parker, R. (2016). Circular RNAs Co-precipitate with extracellular vesicles: a possible mechanism for circRNA clearance. *PLOS ONE* 11 (2), e0148407. doi:10.1371/journal.pone.0148407

Lewis, B. P., Burge, C. B., and Bartel, D. P. (2005). Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 120 (1), 15–20. doi:10.1016/j.cell.2004.12.035

Li, Y., Han, X., Fan, H., Sun, J., Ni, M., Zhang, L., et al. (2022). Circular RNA AXL increases neuron injury and inflammation through targeting microRNA-328 mediated BACE1 in Alzheimer's disease. *Neurosci. Lett.* 776, 136531. doi:10.1016/j.neulet.2022. 136531

Liang, D., Tatomer, D. C., Luo, Z., Wu, H., Yang, L., Chen, L.-L., et al. (2017). The output of protein-coding genes shifts to circular RNAs when the pre-mRNA processing machinery is limiting. *Mol. Cell* 68 (5), 940–954.e3. doi:10.1016/j.molcel.2017.10.034

Lin, R., Lopez, J. P., Cruceanu, C., Pierotti, C., Fiori, L. M., Squassina, A., et al. (2021). Circular RNA circCCNT2 is upregulated in the anterior cingulate cortex of individuals with bipolar disorder. *Transl. Psychiatry* 11 (1), 629–638. doi:10.1038/s41398-021-01746-4

Lo, Ij., Hill, J., Vilhjálmsson, B. J., and Kjems, J. (2020). Linking the association between circRNAs and Alzheimer's disease progression by multi-tissue circular RNA characterization. *RNA Biol.* 17 (12), 1789–1797. doi:10.1080/15476286.2020.1783487

Lublin, F. D., Reingold, S. C., Cohen, J. A., Cutter, G. R., Sørensen, P. S., Thompson, A. J., et al. (2014). Defining the clinical course of multiple sclerosis: the 2013 revisions. *Neurology* 83 (3), 278–286. doi:10.1212/WNL.00000000000560

Luykx, J. J., Giuliani, F., Giuliani, G., and Veldink, J. (2019). Coding and noncoding RNA abnormalities in bipolar disorder. *Genes* 10 (11), 946. doi:10.3390/ genes10110946

Ma, L., Chen, Z., Erdjument-Bromage, H., Tempst, P., and Pandolfi, P. P. (2005). Phosphorylation and functional inactivation of TSC2 by Erk implications for tuberous sclerosis and cancer pathogenesis. *Cell* 121 (2), 179–193. doi:10.1016/j. cell.2005.02.031

Mahmoudi, E., Green, M. J., and Cairns, M. J. (2021). Dysregulation of circRNA expression in the peripheral blood of individuals with schizophrenia and bipolar disorder. J. Mol. Med. 99 (7), 981–991. doi:10.1007/s00109-021-02070-6

Mai, T.-L., Chen, C.-Y., Chen, Y.-C., Chiang, T.-W., and Chuang, T.-J. (2022). Transgenetic effects of circular RNA expression quantitative trait loci and potential causal mechanisms in autism. *Mol. Psychiatry* 27 (11), 4695–4706. doi:10.1038/s41380-022-01714-4

Malhi, G. S., and Mann, J. J. (2018). Depression. Lancet London, Engl. 392 (10161), 2299–2312. doi:10.1016/S0140-6736(18)31948-2

Marfil-Marin, E., Santamaría-Olmedo, M., PerezGrovas-Saltijeral, A., Valdes-Flores, M., Ochoa-Morales, A., Jara-Prado, A., et al. (2021). circRNA regulates dopaminergic synapse, MAPK, and long-term depression pathways in Huntington disease. *Mol. Neurobiol.* 58 (12), 6222–6231. doi:10.1007/s12035-021-02536-1

Margvelani, G., Welden, J. R., Maquera, A. A., Eyk, J. E. V., Murray, C., Sardon, S. C. M., et al. (2023). Influence of FTDP-17 mutants on circular Tau RNAs. *bioRxiv*. doi:10. 1101/2023.09.08.556913

Memczak, S., Jens, M., Elefsinioti, A., Torti, F., Krueger, J., Rybak, A., et al. (2013). Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* 495 (7441), 333–338. doi:10.1038/nature11928 Memczak, S., Papavasileiou, P., Peters, O., and Rajewsky, N. (2015). Identification and characterization of circular RNAs as a new class of putative biomarkers in human blood. *PloS One* 10 (10), e0141214. doi:10.1371/journal.pone.0141214

Meng, X., Hu, D., Zhang, P., Chen, Q., and Chen, M. (2019). CircFunBase: a database for functional circular RNAs. *Database* 2019, baz003. doi:10.1093/database/baz003

Meyza, K. Z., and Blanchard, D. C. (2017). The BTBR mouse model of idiopathic autism—current view on mechanisms. *Neurosci. Biobehav. Rev.* 76 (Pt A), 99–110. doi:10.1016/j.neubiorev.2016.12.037

Morandell, J., Monziani, A., Lazioli, M., Donzel, D., Döring, J., Oss Pegorar, C., et al. (2024). *CircHTT*(2,3,4,5,6)- co-evolving with the *HTT* CAG-repeat tract - modulates Huntington's disease phenotypes. *Nucleic acids.* 35 (3), 102234. doi:10.1016/j.omtn. 2024.102234

Morris-Rosendahl, D. J., and Crocq, M.-A. (2020). Neurodevelopmental disordersthe history and future of a diagnostic concept. *Dialogues Clin. Neurosci.* 22 (1), 65–72. doi:10.31887/DCNS.2020.22.1/macrocq

Nemeth, K., Bayraktar, R., Ferracin, M., and Calin, G. A. (2024). Non-coding RNAs in disease: from mechanisms to therapeutics. *Nat. Rev. Genet.* 25 (3), 211–232. doi:10. 1038/s41576-023-00662-1

Ngo, L. H., Bert, A. G., Dredge, B. K., Williams, T., Murphy, V., Li, W., et al. (2024). Nuclear export of circular RNA. *Nature* 627 (8002), 212–220. doi:10.1038/s41586-024-07060-5

Nguyen, T. B., Miramontes, R., Chillon-Marinas, C., Maimon, R., Vazquez-Sanchez, S., Lau, A. L., et al. (2025). Aberrant splicing in Huntington's disease accompanies disrupted TDP-43 activity and altered m6A RNA modification. *Nat. Neurosci.* 28 (2), 280–292. doi:10.1038/s41593-024-01850-w

Nielsen, A. F., Bindereif, A., Bozzoni, I., Hanan, M., Hansen, T. B., Irimia, M., et al. (2022). Best practice standards for circular RNA research. *Nat. Methods* 19 (10), 1208–1220. doi:10.1038/s41592-022-01487-2

Paraboschi, E. M., Cardamone, G., Rimoldi, V., Gemmati, D., Spreafico, M., Duga, S., et al. (2015). Meta-analysis of multiple sclerosis microarray data reveals dysregulation in RNA splicing regulatory genes. *Int. J. Mol. Sci.* 16 (10), 23463–23481. doi:10.3390/ ijms161023463

Paraboschi, E. M., Cardamone, G., Soldà, G., Duga, S., and Asselta, R. (2018). Interpreting non-coding genetic variation in multiple sclerosis genome-wide associated regions. *Front. Genet.* 9, 647. doi:10.3389/fgene.2018.00647

Parenti, I., Rabaneda, L. G., Schoen, H., and Novarino, G. (2020). Neurodevelopmental disorders: from genetics to functional pathways. *Trends Neurosci.* 43 (8), 608–621. doi:10.1016/j.tins.2020.05.004

Parikshak, N. N., Swarup, V., Belgard, T. G., Irimia, M., Ramaswami, G., Gandal, M. J., et al. (2016). Genome-wide changes in lncRNA, splicing, and regional gene expression patterns in autism. *Nature* 540 (7633), 423–427. doi:10.1038/nature20612

Pellegrini, M., Bergonzoni, G., Perrone, F., Squitieri, F., and Biagioli, M. (2022). Current diagnostic methods and non-coding RNAs as possible biomarkers in Huntington's disease. *Genes* 13 (11), 2017. doi:10.3390/genes13112017

Peng, L., Chen, G., Zhu, Z., Shen, Z., Du, C., Zang, R., et al. (2017). Circular RNA ZNF609 functions as a competitive endogenous RNA to regulate AKT3 expression by sponging miR-150-5p in Hirschsprung's disease. *Oncotarget* 8 (1), 808–818. doi:10. 18632/oncotarget.13656

Piwecka, M., Glažar, P., Hernandez-Miranda, L. R., Memczak, S., Wolf, S. A., Rybak-Wolf, A., et al. (2017). Loss of a mammalian circular RNA locus causes miRNA deregulation and affects brain function. *Sci. (New York, N.Y.)* 357 (6357), eaam8526. doi:10.1126/science.aam8526

Puri, S., Hu, J., Sun, Z., Lin, M., Stein, T. D., Farrer, L. A., et al. (2023). Identification of circRNAs linked to Alzheimer's disease and related dementias. *Alzheimer's and Dementia J. Alzheimer's Assoc.* 19 (8), 3389–3405. doi:10.1002/alz.12960

Rybak-Wolf, A., Stottmeister, C., Glažar, P., Jens, M., Pino, N., Giusti, S., et al. (2015). Circular RNAs in the mammalian brain are highly abundant, conserved, and dynamically expressed. *Mol. Cell* 58 (5), 870–885. doi:10.1016/j.molcel.2015.03.027

Rybiczka-Tešulov, M., Garritsen, O., Venø, M. T., Wieg, L., Dijk, R., Rahimi, K., et al. (2024). Circular RNAs regulate neuron size and migration of midbrain dopamine neurons during development. *Nat. Commun.* 15 (1), 6773. doi:10.1038/s41467-024-51041-1

Santer, L., Bär, C., and Thum, T. (2019). Circular RNAs: a novel class of functional RNA molecules with a therapeutic perspective. *Mol. Ther. J. Am. Soc. Gene Ther.* 27 (8), 1350–1363. doi:10.1016/j.ymthe.2019.07.001

Seeler, S., Andersen, M. S., Sztanka-Toth, T., Rybiczka-Tešulov, M., Van Den Munkhof, M. H., Chang, C.-C., et al. (2023). A circular RNA expressed from the FAT3 locus regulates neural development. *Mol. Neurobiol.* 60 (6), 3239–3260. doi:10. 1007/s12035-023-03253-7

Shi, Y., Song, R., Wang, Z., Zhang, H., Zhu, J., Yue, Y., et al. (2021a). Potential clinical value of circular RNAs as peripheral biomarkers for the diagnosis and treatment of major depressive disorder. *eBioMedicine* 66, 103337. doi:10.1016/j.ebiom.2021.103337

Shi, Y., Wang, Q., Song, R., Kong, Y., and Zhang, Z. (2021b). Non-coding RNAs in depression: promising diagnostic and therapeutic biomarkers. *EBioMedicine* 71, 103569. doi:10.1016/j.ebiom.2021.103569

Silenzi, V., D'Ambra, E., Santini, T., D'Uva, S., Setti, A., Salvi, N., et al. (2024). A tripartite circRNA/mRNA/miRNA interaction regulates glutamatergic signaling in the mouse brain. *Cell Rep.* 43 (10), 114766. doi:10.1016/j.celrep.2024.114766

Spurlock, C. F., Tossberg, J. T., Guo, Y., Sriram, S., Crooke, P. S., and Aune, T. M. (2015). Defective structural RNA processing in relapsing-remitting multiple sclerosis. *Genome Biol.* 16 (1), 58. doi:10.1186/s13059-015-0629-x

Starke, S., Jost, I., Rossbach, O., Schneider, T., Schreiner, S., Hung, L.-H., et al. (2015). Exon circularization requires canonical splice signals. *Cell Rep.* 10 (1), 103–111. doi:10. 1016/j.celrep.2014.12.002

Stott, J., Wright, T., Holmes, J., Wilson, J., Griffiths-Jones, S., Foster, D., et al. (2023). A systematic review of non-coding RNA genes with differential expression profiles associated with autism spectrum disorders. *PloS One* 18 (6), e0287131. doi:10.1371/journal.pone.0287131

Suenkel, C., Cavalli, D., Massalini, S., Calegari, F., and Rajewsky, N. (2020). A highly conserved circular RNA is required to keep neural cells in a progenitor state in the mammalian brain. *Cell Rep.* 30 (7), 2170–2179.e5. doi:10.1016/j.celrep.2020.01.083

Suzuki, H., Zuo, Y., Wang, J., Zhang, M. Q., Malhotra, A., and Mayeda, A. (2006). Characterization of RNase R-digested cellular RNA source that consists of lariat and circular RNAs from pre-mRNA splicing. *Nucleic Acids Res.* 34 (8), e63. doi:10.1093/nar/ gkl151

Tan, G., Wang, L., Liu, Y., Zhang, H., Feng, W., and Liu, Z. (2021). The alterations of circular RNA expression in plasma exosomes from patients with schizophrenia. *J. Cell. Physiology* 236 (1), 458–467. doi:10.1002/jcp.29873

Tee, A. R., Manning, B. D., Roux, P. P., Cantley, L. C., and Blenis, J. (2003). Tuberous sclerosis complex gene products, Tuberin and Hamartin, control mTOR signaling by acting as a GTPase-activating protein complex toward Rheb. *Curr. Biol. CB* 13 (15), 1259–1268. doi:10.1016/s0960-9822(03)00506-2

Thompson, A. J., Baranzini, S. E., Geurts, J., Hemmer, B., and Ciccarelli, O. (2018). Multiple sclerosis. *Lancet London, Engl.* 391 (10130), 1622–1636. doi:10.1016/S0140-6736(18)30481-1

Titze-de-Almeida, S. S., and Titze-de-Almeida, R. (2023). Progress in circRNAtargeted Therapy in experimental Parkinson's disease. *Pharmaceutics* 15 (8), 2035. doi:10.3390/pharmaceutics15082035

Tondo, L., Vázquez, G. H., and Baldessarini, R. J. (2017). Depression and mania in bipolar disorder. *Curr. Neuropharmacol.* 15 (3), 353–358. doi:10.2174/1570159X14666160606210811

Venø, M. T., Hansen, T. B., Venø, S. T., Clausen, B. H., Grebing, M., Finsen, B., et al. (2015). Spatio-temporal regulation of circular RNA expression during porcine embryonic brain development. *Genome Biol.* 16, 245. doi:10.1186/s13059-015-0801-3

Vo, J. N., Cieslik, M., Zhang, Y., Shukla, S., Xiao, L., Zhang, Y., et al. (2019). The landscape of circular RNA in cancer. *Cell* 176 (4), 869–881.e13. doi:10.1016/j.cell.2018. 12.021

Wang, F., Li, Y., Shen, H., Martinez-Feduchi, P., Ji, X., Teng, P., et al. (2024). Identification of pathological pathways centered on circRNA dysregulation in association with irreversible progression of Alzheimer's disease. *Genome Med.* 16 (1), 129. doi:10.1186/s13073-024-01404-6

Wang, J., Yang, Z., Chen, C., Xu, Y., Wang, H., Liu, B., et al. (2021). Comprehensive circRNA expression profile and construction of circRNAsrelated ceRNA network in a mouse model of autism. *Front. Genet.* 11, 623584. doi:10.3389/fgene.2020.623584

Wang, P. L., Bao, Y., Yee, M.-C., Barrett, S. P., Hogan, G. J., Olsen, M. N., et al. (2014). Circular RNA is expressed across the eukaryotic tree of life. *PLOS ONE* 9 (3), e90859. doi:10.1371/journal.pone.0090859

Wang, X., Xie, J., Tan, L., Lu, Y., Shen, N., Li, J., et al. (2023). N6-methyladenosinemodified circRIMS2 mediates synaptic and memory impairments by activating GluN2B ubiquitination in Alzheimer's disease. *Transl. Neurodegener.* 12 (1), 53. doi:10.1186/ s40035-023-00386-6

Wang, Y., Liu, J., Ma, J., Sun, T., Zhou, Q., Wang, W., et al. (2019). Exosomal circRNAs: biogenesis, effect and application in human diseases. *Mol. Cancer* 18 (1), 116. doi:10.1186/s12943-019-1041-z

Welden, J. R., Margvelani, G., Arizaca Maquera, K. A., Gudlavalleti, B., Miranda Sardón, S. C., Campos, A. R., et al. (2022). RNA editing of microtubule-associated protein tau circular RNAs promotes their translation and tau tangle formation. *Nucleic Acids Res.* 50 (22), 12979–12996. doi:10.1093/nar/gkac1129

Wu, W., Ji, P., and Zhao, F. (2020). CircAtlas: an integrated resource of one million highly accurate circular RNAs from 1070 vertebrate transcriptomes. *Genome Biol.* 21 (1), 101. doi:10.1186/s13059-020-02018-y

Wu, W., Zhang, J., Cao, X., Cai, Z., and Zhao, F. (2022). Exploring the cellular landscape of circular RNAs using full-length single-cell RNA sequencing. *Nat. Commun.* 13 (1), 3242. doi:10.1038/s41467-022-30963-8

Wu, Y. E., Parikshak, N. N., Belgard, T. G., and Geschwind, D. H. (2016). Genomewide, integrative analysis implicates microRNA dysregulation in autism spectrum disorder. *Nat. Neurosci.* 19 (11), 1463–1476. doi:10.1038/nn.4373

Xin, J., Zhang, X.-Y., Sun, D.-K., Tian, L.-Q., and Xu, P. (2019). Up-regulated circular RNA hsa\_circ\_0067934 contributes to glioblastoma progression through activating

PI3K-AKT pathway. Eur. Rev. Med. Pharmacol. Sci. 23 (8), 3447-3454. doi:10.26355/ eurrev\_201904\_17709

Yao, G., Niu, W., Zhu, X., He, M., Kong, L., Chen, S., et al. (2019). hsa\_circRNA\_ 104597: a novel potential diagnostic and therapeutic biomarker for schizophrenia. *Biomarkers Med.* 13 (5), 331–340. doi:10.2217/bmm-2018-0447

You, X., Vlatkovic, I., Babic, A., Will, T., Epstein, I., Tushev, G., et al. (2015). Neural circular RNAs are derived from synaptic genes and regulated by development and plasticity. *Nat. Neurosci.* 18 (4), 603–610. doi:10.1038/nn.3975

Yu, X., Fan, Z., Yang, T., Li, H., Shi, Y., Ye, L., et al. (2024). Plasma circRNA HIPK2 as a putative biomarker for the diagnosis and prediction of therapeutic effects in major depressive disorder. *Clin. Chim. Acta* 552, 117694. doi:10.1016/j.cca.2023.117694

Zajaczkowski, E. L., Zhao, Q., Liau, W.-S., Gong, H., Madugalle, S. U., Periyakaruppiah, A., et al. (2023). Localised Cdr1as activity is required for fear extinction memory. *Neurobiol. Learn. Mem.* 203, 107777. doi:10.1016/j.nlm.2023.107777

Zhang, B., Zhang, H., Wang, Z., Cao, H., Zhang, N., Dai, Z., et al. (2024). The regulatory role and clinical application prospects of circRNA in the occurrence and development of CNS tumors. *CNS Neurosci. and Ther.* 30 (4), e14500. doi:10.1111/cns. 14500

Zhang, Y., Du, L., Bai, Y., Han, B., He, C., Gong, L., et al. (2020). CircDYM ameliorates depressive-like behavior by targeting miR-9 to regulate microglial activation via HSP90 ubiquitination. *Mol. Psychiatry* 25 (6), 1175–1190. doi:10.1038/s41380-018-0285-0

Zhang, Y., Huang, R., Cheng, M., Wang, L., Chao, J., Li, J., et al. (2019). Gut microbiota from NLRP3-deficient mice ameliorates depressive-like behaviors by regulating astrocyte dysfunction via circHIPK2. *Microbiome* 7 (1), 116. doi:10.1186/ s40168-019-0733-3

Zhao, J., Li, Q., Hu, J., Yu, H., Shen, Y., Lai, H., et al. (2024). Circular RNA landscape in extracellular vesicles from human biofluids. *Genome Med.* 16 (1), 126. doi:10.1186/s13073-024-01400-w

Zhao, S.-Y., Wang, J., Ouyang, S.-B., Huang, Z.-K., and Liao, L. (2018). Salivary circular RNAs Hsa\_Circ\_0001874 and Hsa\_Circ\_0001971 as novel biomarkers for the diagnosis of oral squamous cell carcinoma. *Cell. Physiology Biochem. Int. J. Exp. Cell. Physiology, Biochem. Pharmacol.* 47 (6), 2511–2521. doi:10.1159/000491624

Zhong, C., Yu, S., Han, M., Chen, J., and Ning, K. (2017). Heterogeneous circRNA expression profiles and regulatory functions among HEK293T single cells. *Sci. Rep.* 7 (1), 14393. doi:10.1038/s41598-017-14807-w

Zimmerman, A. J., Hafez, A. K., Amoah, S. K., Rodriguez, B. A., Dell'Orco, M., Lozano, E., et al. (2020). A psychiatric disease-related circular RNA controls synaptic gene expression and cognition. *Mol. Psychiatry* 25 (11), 2712–2727. doi:10.1038/ s41380-020-0653-4

Zimmerman, A. J., Weick, J. P., Papageorgiou, G., Mellios, N., and Brigman, J. L. (2024). Aberrant encoding of event saliency in the orbitofrontal cortex following loss of the psychiatric-associated circular RNA, circHomer1. *Transl. Psychiatry* 14 (1), 480–514. doi:10.1038/s41398-024-03188-0