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# Physiologically based pharmacokinetic modeling supports investigation of potential drug-drug interactions in the preand early post-hematopoietic stem cell transplantation stages

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**Introduction:** Drug-drug interactions (DDIs) are an important issue in medication safety and a potential cause of adverse drug events in the pre- and early posthematopoietic stem cell transplantation (HSCT). This study introduced a physiologically based pharmacokinetic (PBPK) modeling platform to evaluate complex DDIs in these critical stages and to optimize dosing for personalized treatment.

**Methods:** PBPK models were developed using a bottom-up with middle-out approach and executed with PK-Sim<sup>®</sup> software. Model validation required that predicted PK values fall within a twofold range of observed data. Then, the validated model was used to simulate alternative dosing regimens to achieve target therapeutic levels.

**Results:** PBPK models were developed and evaluated for 13 drugs commonly used in HSCT, including cyclosporine, tacrolimus, sirolimus, busulfan, phenytoin, voriconazole, posaconazole, itraconazole, fluconazole, letermovir, fosaprepitant, aprepitant, and omeprazole. Simulation results indicated marked DDIs in the preand early post-HSCT phases, particularly involving cyclosporine and phenytoin. Several drugs notably increased cyclosporine concentrations, while phenytoin substantially reduced the exposure to other medications.

**Conclusion:** This PBPK modeling platform provides a robust tool for identifying and mitigating DDIs in the pre- and early post-HSCT phases. By enabling the optimization of treatment regimens, this model serves as a valuable tool for improving drug safety and therapeutic outcomes for patients with HSCT.

#### KEYWORDS

HSCT, PBPK modeling, DDIs, pharmacokinetics, personalized dosing

# 1 Introduction

Hematopoietic stem cell transplantation (HSCT) has become an important treatment option for patients with defined congenital or acquired disorders of the hematopoietic system (Gratwohl et al., 2010). During the pre- (day –6 to day –2) and early post- (day 2 to day 30) HSCT phases, patients often receive complex regimens, including chemotherapeutic,

immunosuppressive, and antimicrobial agents (Figure 1; Palmer et al., 2016; Bolaños-Meade et al., 2019; Groll et al., 2021; Ljungman et al., 2025). Certain combinations may produce drug-drug interactions (DDIs), leading to undesirable adverse outcomes and increased drug toxicity (Glotzbecker et al., 2012; Secoli et al., 2015; Pejčić et al., 2019). This concern is particularly relevant for medications with a narrow therapeutic index, such as busulfan, voriconazole, cyclosporine,

tacrolimus, and sirolimus (Table 1), which are more prone to unanticipated DDIs when co-administered with enzyme inhibitors or inducers (Palmer et al., 2016; Bolaños-Meade et al., 2019; Groll et al., 2021). In recent years, the clinical use of some new drugs (e.g., fosaprepitant and letermovir) has further increased the complexity of DDIs, so an accurate and reliable model for predicting DDIs is required for dose optimization (Patel et al., 2017; Kim, 2018).



| TABLE 1 Summary of                     | potential drug-drug- | interactions in patients | with HSCT and | therapeutic target. |
|--|----------------------|--------------------------|---------------|---------------------|
| ······································ |                      |                          |               |                     |

| Drugs         | Major metabolism and transport pathways <sup>a</sup>     | Pharmacogenomics<br>(CPIC) <sup>b</sup> | Mechanism <sup>a</sup>   | Therapeutic<br>target                             |
|---------------|--|---|--|---|
| Busulfan      | GST  | _                                       | _  | AUC of<br>900–1,350 μM min                        |
| Phenytoin     | СҮР2С9, СҮР2С19  | СҮР2С9                                  | A strong hepatic drug-metabolizing enzymes inducer   | $C_{\rm min}$ of 10–20 $\mu g/mL$ for epilepticus |
| Voriconazole  | СҮР2С19, СҮР2С9, СҮРЗА4                                  | CYP2C19                                 | A strong CYP2C19 and CYP3A4 inhibitor  | $C_{min}$ of 1–5 µg/mL                            |
| Posaconazole  | UGT  | _                                       | A strong CYP3A and P-gp inhibitor  | $C_{min}$ of ${\geq}0.7~\mu\text{g/mL}$           |
| Itraconazole  | CYP3A4   | _                                       | A strong CYP3A4 inhibitor; a P-gp and BCRP inhibitor   | $C_{min}$ of ${\geq}0.5~\mu\text{g/mL}$           |
| Fluconazole   | Cleared primarily by renal excretion                     | _                                       | A strong CYP2C19 inhibitor; a moderate CYP2C9 and CYP3A4 inhibitor   | _   |
| Letermovir    | CYP3A, CYP2D6, P-gp, UGT1A1,<br>UGT1A3, OATP1B1, OATP1B3 | _                                       | A CYP3A, CYP2C8, OATP1B1, OATP1B3, P-gp,<br>BCRP, BSEP, and MRP inhibitor; a CYP3A, CYP2C9,<br>and CYP2C19 inducer | _   |
| Fosaprepitant | Converted to aprepitant; CYP3A4,<br>CYP1A2, CYP2C19      | _                                       | A weak CYP3A4 inhibitor  | _   |
| Aprepitant    | СҮРЗА4, СҮР1А2, СҮР2С19                                  | -                                       | A weak-to-moderate CYP3A4 inhibitor; a CYP3A4 and CYP2C9 inducer   | _   |
| Omeprazole    | СҮР2С19, СҮРЗА4  | CYP2C19                                 | A moderate CYP3A and CYP2C19 inhibitor   | _   |
| Cyclosporine  | СҮРЗА, Р-др  | _                                       | A CYP3A4 and multiple drug efflux transporters (e.g., P-gp) inhibitor  | $C_{\rm min}$ of 200–400 ng/mL                    |
| Tacrolimus    | СҮРЗА  | СҮРЗА5                                  | A CYP3A4 and CYP3A5 inhibitor  | C <sub>min</sub> of 5–15 ng/mL                    |
| Sirolimus     | CYP3A4, P-gp   | _                                       | -  | $C_{\rm min}$ of 3–8 ng/mL                        |

<sup>a</sup>from drugs.com website.

<sup>b</sup>from Clinical Pharmacogenetics Implementation Consortium (CPIC) guidelines; CYP, cytochrome P450; GST, glutathione S-transferase; UGT, UDP-glucuronosyltransferase; OATP, organic anion transporting polypeptides; P-gp, P-glycoprotein; BCRP, breast cancer resistance protein; BSEP, bile salt export pump; MRP, multidrug resistance-associated protein; AUC, the area under the curve; C<sub>min</sub>, trough concentration.

Currently, clinically relevant DDIs can be queried through online and offline interaction checkers, such as Drugs, Lexi-Interact, and ePocrates, but the available clinical dosage suggestions are limited (Storelli et al., 2018). However, model-informed precise dosing can leverage pharmacokinetic (PK) models to tailor individualized dosing (Darwich et al., 2021). Therefore, an increasing number of studies have applied physiologically based PK (PBPK) models to predict drug PK profiles by integrating drug property parameters with physiological parameters of the organism. Unlike interaction checkers, PBPK models are more prominent in describing time-variable concentrations of drugs and predicting DDI in the early stages of drug development and complex clinical scenarios, such as simultaneous inhibition and induction of enzymes and transporters, complex transporterenzyme interplay, and genetic variations of cytochrome P450 (CYP) enzymes (Vijaywargi et al., 2023; Foti, 2025). Accordingly, PBPK models have been widely used in drug discovery and development areas, including DDIs, organ injury, pediatrics, drug-gene interactions, disease impact, and food effects (Sun et al., 2024).

To date, several studies have developed PBPK models to predict the PK profiles of busulfan, phenytoin, voriconazole, posaconazole, itraconazole, fluconazole, letermovir, omeprazole, cyclosporine, tacrolimus, and sirolimus (Supplementary Table S1). However, due to the different diseases or populations of concern, DDIs caused by common drug combinations in patients with HSCT have rarely been reported, such as busulfan in combination with phenytoin, voriconazole or posaconazole in combination with cyclosporine, etc. Furthermore, no PBPK model for fosaprepitant and aprepitant has been reported. So, there is still a need for further models and more comprehensive DDIs networks.

This study aimed to develop a PBPK modeling platform to investigate the complex DDIs of 13 commonly used drugs in the preand early post-HSCT phase, including busulfan, phenytoin, voriconazole, posaconazole, itraconazole, fluconazole, letermovir, omeprazole, fosaprepitant, aprepitant, cyclosporine, tacrolimus, and sirolimus. Furthermore, dose optimization of the above drugs was predicted for personalized treatment.

### 2 Materials and methods

#### 2.1 Software

PBPK model development and simulations were performed with PK-Sim<sup>®</sup> (Version 11.2, Bayer Technology Services, Leverkusen, Germany). Clinical study data from published literature were collected with the semi-automated tool WebPlotDigitizer (Version 4.6, Ankit Rohatgi, Pacifica, CA, United States). For plot generation, GraphPad Prism 8.0.1 (GraphPad Software Inc., San Diego, CA, United States) was used.

# 2.2 PBPK models development and verification

PBPK models for 13 drugs were developed based on the bottomup with middle-out techniques (Sun et al., 2024), and the workflow adopted for model development, verification, and application was illustrated in Figure 2. Briefly, a PBPK modeling of a drug mainly consists of expression profiles, individuals, populations, compounds, formulations, administration protocols, and observed data. Protein expression of enzymes and transporters was implemented using the PK-Sim® database. Individuals or populations used to simulate the different studies were modeled according to the corresponding study reports, with age, weight, height, gender, and ethnicity (Supplementary Material). If the demographic information was not found, a 30-yearold male European was assumed, with the mean weight and height characteristics in the software database. Other building blocks required an extensive literature search to gain information about the physicochemical properties and absorption, distribution, metabolism, and excretion processes of the drug. Meanwhile, plasma concentration-time profiles of intravenous and oral administration in single or multiple doses from healthy individuals or clinical studies were digitized and used for model development and evaluation. Parameters that were not informed from the literature were optimized, fitting the model to the observed PK data in the literature.

The predictive performance of PBPK models was initially evaluated by comparing predicted plasma concentration-time profiles to observed data. The PK parameters assessed were the area under the curve (AUC) and peak plasma concentration ( $C_{max}$ ), which were quantitatively assessed based on a predefined criterion of two-fold range (0.5–2.0) of the predicted/observed ratio.

#### 2.3 DDIs network modeling and verification

In the DDIs model, there are five specified types of inhibition, including competitive, uncompetitive, non-competitive, mixed, and irreversible inhibition. Inhibition types for different drugs are based on literature reports, software databases, and optimization. Besides, drug-mediated induction of enzymes and transporters was expressed in terms of the maximum induction effect (Emax) and the concentration supporting half of Emax (EC50).

The performance of the DDI model was evaluated by comparing the predicted to observed victim drug plasma concentration-time profiles when administered alone and during coadministration. The metric to assess the DDI modeling performance was the predicted DDI ratios of AUC and  $C_{max}$  (AUC or  $C_{max}$  of victim drug during coadministration/AUC or  $C_{max}$  of victim drug alone) to the observed DDI ratios of AUC and  $C_{max}$ . A two-fold error margin (0.5–2.0) was set as the acceptance criterion.

#### 2.4 Prospective PBPK DDI scenario for dose optimization

To mimic the clinical setting, the verified model was used to simulate the exposure of drugs that need therapeutic drug monitoring (TDM), such as voriconazole, posaconazole, busulfan, cyclosporine, tacrolimus, and sirolimus. A virtual population of 100 Japanese individuals with the mean weight and height characteristics given in the software database was generated. Subsequently, the fold change in AUC or trough concentration ( $C_{min}$ ) at steady-state was calculated during coadministration to when administered alone. Voriconazole  $C_{min}$  of 1–5 µg/mL, posaconazole  $C_{min}$  of  $\geq$ 0.7 µg/mL, itraconazole  $C_{min}$  of  $\geq$ 0.5 µg/mL, busulfan AUC of 900–1,500 µM min, cyclosporine  $C_{min}$  of 200–400 ng/mL, tacrolimus  $C_{min}$  of 5–15 ng/mL, and sirolimus



 $C_{min}$  of 3–8 ng/mL are used as the therapeutic index for individualized treatment (Palmer et al., 2016; Bolaños-Meade et al., 2019; Groll et al., 2021).

# **3** Results

#### 3.1 PBPK model building and evaluation

The PBPK models of three immunosuppressants (cyclosporine, tacrolimus, and sirolimus), one chemotherapeutic agent and its anticonvulsant prophylactic agent (busulfan and phenytoin), four antifungal agents (voriconazole, posaconazole, itraconazole, and fluconazole), one antiviral agent (letermovir), two antiemetic agents (fosaprepitant and aprepitant), and omeprazole were developed and comprehensively evaluated using the blood concentration-time profiles. Among them, parameters for itraconazole, fluconazole, and omeprazole were taken directly from the PK-Sim<sup>®</sup> database, and those for voriconazole and tacrolimus were taken directly from the previously established models (Gong et al., 2023; Wang et al., 2024). The rest of drug parameters were gathered from the literature and optimized based on blood concentration-time profiles from different literature, as summarized in Supplementary Table S2.

Model performance was demonstrated by comparison of predicted to observed plasma concentration-time profiles in Supplementary Figures S1–S7, and predicted to observed AUC and  $C_{max}$  values was presented in Supplementary Table S3. As a result, the predicted to observed AUC and  $C_{max}$  ratios were within 0.53–1.71-fold error.

## 3.2 DDI network modeling

In this study, irreversible inhibition processes of CYP3A4 for voriconazole and tacrolimus and CYP2C19 for omeprazole were modeled using Ki (inhibition constant) and  $k_{inact}$  (maximum inactivation rate constant) values (Li et al., 2020; Loer et al., 2023);

the competitive inhibition of CYP450, organic anion-transporting peptide (OATP) 1B1/3 and P-glycoprotein (P-gp) for posaconazole, itraconazole, fluconazole, aprepitant, cyclosporine, tacrolimus, and letermovir was modeled using *Ki* values (Gertz et al., 2013; Gerner et al., 2022; Loer et al., 2023); the induction of CYP2C9, CYP2C19, CYP3A4, UDP glucuronosyltransfer (UGT) 1A4 and glutathione S-transferase (GST) A1 for phenytoin was implemented using EC50 and Emax values (Rodriguez-Vera et al., 2023).

The performance of DDI models was evaluated by comparing predicted to observed PK profiles of victim drugs with/without an enzyme or transporter perpetrator coadministration (Supplementary Figures S8–S14). In addition, the predicted to observed DDI AUC ratios and  $C_{\rm max}$  ratios of all DDI studies were shown in Supplementary Table S4, with a range of 0.54–1.86 for predicted to observed parameters.

# 3.3 Prospective PBPK DDI scenario for dose optimization

To show the utility of the model in personalized therapy, dose adaptations for different DDI scenarios were simulated. Plasma concentration-time profiles of victim drug during co-administration of perpetrator drug were simulated and compared to those without perpetrator drug (Figure 3). Notably, the coadministration of voriconazole, itraconazole, posaconazole, fluconazole, or letermovir increased cyclosporine  $C_{min}$  of approximately 2.2-, 2.1-, 2.0-, 1.6-, and 1.2-fold, respectively (Table 2). Conversely, coadministration of phenytoin decreased cyclosporine  $C_{min}$ , aprepitant AUC, fosaprepitant AUC, busulfan AUC, and voriconazole  $C_{min}$  by approximately 21.1%, 36.9%, 8.5%, 13.4%, and 51.0%, respectively (Figures 3a–d).

## 4 Discussion

Over the past decade, the use of PBPK modeling has been rising, and regulatory agencies have issued DDI guidance highlighting the



use of such modeling approaches (Sun et al., 2024; Paul et al., 2025). Commonly used PBPK software packages include SimCYP (Certara), GastroPlus (Simulations Plus), and PK-Sim (Open Systems Pharmacology), among which PK-Sim is the only opensource platform and is used for free. Alternatively, FDA reviewers endorsed the predicted results for all three platforms (Sun et al., 2024). This method using in HSCT has several advantages:

Firstly, it allows for the extrapolation of PK behavior of drugs in healthy volunteers to diseased or special populations (e.g., the elderly, children, pregnant women, and patients with hepatic and renal insufficiency). Busulfan is a widely used chemotherapic agent in pediatrics with HSCT. Given the narrow therapeutic index, it is routinely dosed with TDM (Palmer et al., 2016). In this PBPK model, age-dependent enzyme activity was tailored and evaluated with an external dataset (Supplementary Tables S3, S4). The results indicated that at the same dose (0.8 mg/kg), busulfan exposure in children (979.4  $\pm$  76.2  $\mu$ M min) was lower than that in adults (1,078.8  $\pm$  103.9  $\mu$ M min). When co-administered with phenytoin,

busulfan exposure in children (847.9  $\pm$  68.4  $\mu M$  min) was even lower (Figure 3c) and below the therapeutic window (AUC of 900–1,350  $\mu M$  min; Palmer et al., 2016); while when increasing busulfan dose to 1.0 mg/kg, the exposure (1,060.7  $\pm$  95.5  $\mu M$  min) was sufficient.

Secondly, the model can assess inter-individual variability in a virtual population by incorporating factors, such as age, gender, ethnicity, and genetic polymorphisms. Voriconazole demonstrates wide interpatient variability in serum concentrations due in part to variant *CYP2C19* alleles (Moriyama et al., 2017). In this PBPK model, different CYP2C19 phenotypes were integrated into the model as normal metabolizer (NM), intermediate metabolizer (IM), and poor metabolizer (PM), with reference values for *CYP2C19* expression of 0.76, 0.40, and 0.01 µmol/L, respectively (Wang et al., 2024). As shown in Figure 3d, AUC for CYP2C19 IMs and PMs were 2.3- to 4.0-fold higher than those for CYP2C19 NMs. The standard oral maintenance dose of voriconazole 200 mg twice daily would be sufficient for CYP2C19 IMs and PMs to reach the

| Victim drug (cyclosporine) | Perpetrator drug   | Predicted $C_{min}$ (ng/mL; mean $\pm$ SD) | C <sub>min</sub> ratio |
|----------------------------|--|--|------------------------|
| 100 mg po bid              | _  | 104.9 ± 32.2                               | _                      |
| 100 mg po bid              | phenytoin 100 mg po tid  | 82.7 ± 26.1                                | 0.8                    |
| 100 mg po bid              | letermovir 240 mg po qd  | 126.3 ± 49.7                               | 1.2                    |
| 100 mg po bid              | fluconazole 200 mg po qd   | 169.2 ± 67.6                               | 1.6                    |
| 100 mg po bid              | posaconazole 200 mg po tid   | 208.6 ± 113.1                              | 2.0                    |
| 100 mg po bid              | itraconazole 200 mg po qd  | 215.2 ± 119.1                              | 2.1                    |
| 100 mg po bid              | voriconazole 200 mg po bid for CYP2C19 NMs   | 233.8 ± 147.7                              | 2.2                    |
| 100 mg po bid              | voriconazole 200 mg po bid for CYP2C19 IMs   | 327.7 ± 193.3                              | 3.1                    |
| 100 mg po bid              | voriconazole 200 mg po bid for CYP2C19 PMs   | 422.7 ± 227.2                              | 4.0                    |
| 100 mg po bid              | voriconazole 200 mg po bid for CYP2C19 NMs;<br>letermovir 240 mg po qd   | 240.6 ± 124.6                              | 2.3                    |
| 100 mg po bid              | voriconazole 200 mg po bid for CYP2C19 NMs;<br>letermovir 240 mg po qd; phenytoin 100 mg po tid on<br>day -5 to -1 | 173.1 ± 76.3                               | 1.7                    |
| 150 mg po bid              | voriconazole 200 mg po bid for CYP2C19 NMs;<br>letermovir 240 mg po qd; phenytoin 100 mg po tid on<br>day –5 to –1 | 261.3 ± 109.7                              | 2.5                    |

TABLE 2 Simulated pharmacokinetic parameters of cyclosporine with different drugs.

Cmin minimum plasma concentration on day 6; SD, standard deviation; NMs, normal metabolizers; IMs, intermediate metabolizers; PMs, poor metabolizers.

tentative therapeutic range of 1.0–5.0 µg/mL for C<sub>min</sub>, while 400 mg twice daily might be more suitable for NMs. Meanwhile, the DDIs between voriconazole and three immunosuppressants were also affected by CYP2C19 genotypes (Figures 3e–g). Similarly, tacrolimus exhibited large inter-and intra-individual PK variability, partly due to genetic variations in *CYP3A5* (Birdwell et al., 2015). Accordingly, different CYP3A5 genotypes were integrated into the model as CYP3A5 expressers (\*1/\*1 or \*1/\*3) and non-expressers (\*3/\*3), with reference values of 0.68 and 0.04 µmol/L for CYP3A5 expression in liver tissues, respectively (Loer et al., 2023).

Thirdly, it enables investigation of the effect of factors, such as formulations, administration routes, and food, on drug PK parameters. In terms of posaconazole, the exposure for oral suspension increased 2.5- to 3.0-fold when it was given with a high-fat meal; whereas, exposures for tablets and capsules were not markedly affected by food and were higher than that for oral suspension (Krishna et al., 2012). During the modeling, intestinal permeability and Weibull parameters were adjusted, and the final model examined four scenarios (tablet or capsules, suspension with dietary unknown, suspension with fasted, and suspension with fed) to predict the influence of food intake and formulation on posaconazole PK profile (Supplementary Table S3; Figure 3h).

Fourthly, the model supports the optimization of rational dosing regimens by integrating therapeutics index or pharmacodynamic indicators. Following HSCT, cyclosporine is commonly administered in combination with voriconazole and letermovir, which are both CYP3A4 inhibitors, and the latter is also an inhibitor of P-gp transporter (Table 1). Thus, when co-administrated with voriconazole and letermovir, simulations showed that the steady-state  $C_{min}$  of cyclosporine on day 6 increased 2.3-fold compared with cyclosporine (100 mg) alone

(Figure 3i; Table 2). Whereas, phenytoin is commonly used as anticonvulsant prophylaxis for busulfan before HSCT. On discontinuation of phenytoin, CYP450 activity may not be fully restored for 7–10 days (Spriet et al., 2010), which may result in decreased concentrations of cyclosporine, voriconazole, and letermovir. Therefore, when phenytoin was administered 5 days before the three drugs combination, simulation showed that cyclosporine  $C_{min}$  (173.2 ± 76.3 ng/mL) on day 6 was still lower than that without phenytoin (240.6 ± 124.6 ng/mL) and also below the therapeutic window ( $C_{min}$  of 200–400 ng/mL). When increasing cyclosporine dose to 150 mg, the  $C_{min}$  (261.3 ± 109.7 ng/mL) met the requirements.

There are several limitations to the present PBPK model. Firstly, the metabolism and DDIs were based on enzyme kinetic parameters observed from in vitro experiments, while in vitro data may not accurately describe drug changes in vivo. This is a common problem with the PBPK model. To ensure the credibility of the final PBPK model, the source of the model parameters was provided, and the current PBPK models were created and refined by combining the bottom-up and middle-out approaches, that is, modifying the parameters to fit the observed data. Secondly, the population parameters for patients with HSCT are currently lacking or challenging to acquire, so this PBPK model does not take into consideration the physiological differences in patients with HSCT. Thirdly, the WHO model evaluation criteria were used in this study, i.e., the predicted-to-observed ratio within a factor of 2 (WHO, 2010). As stated by Guest et al., this strategy results in a potential bias toward successful prediction at lower interaction levels (Guest et al., 2011). For observed DDI ratios of 1 (no interaction), the 2-fold deviation would allow predicted DDI ratios between 0.5 (induction) and 2 (weak to moderate inhibition), which could overstate the DDI performance for weak interactions. Therefore, the methodology proposed by Guest et al. may be more stringent for DDI prediction assessment, accepting a 20% deviation for observed DDI ratios approaching 1. Lastly, there is a lack of clinical validation for prospective DDI scenarios for dose optimization, which needs further research.

Despite its shortcomings, the most meaningful application of PBPK is to obtain the data through prediction in the absence of clinical data and to guide clinical research. In the future, benefiting from the development of software platforms and a more comprehensive understanding of human physiological changes, especially when there are factors, such as disease, that lead to physiological changes, which means that PBPK models will be more widely applied in novel drug development and dose optimization (Sun et al., 2024).

# **5** Conclusion

In conclusion, this study introduced a PBPK modeling platform to predict potential DDIs and their impact on drug exposure in patients in the pre- and early post-HSCT. By enabling the optimization of treatment regimens, this model may serve as a valuable tool for improving drug safety and therapeutic outcomes.

# Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding author.

# **Ethics statement**

The requirement of ethical approval was waived by the ethics committee of the First Affiliated Hospital of Zhengzhou University for the studies involving humans because All human data involved in this research come from literature. The studies were conducted in accordance with the local legislation and institutional requirements. Written informed consent for participation was not required from the participants or the participants' legal guardians/next of kin because All human data involved in this research come from literature.

## References

Birdwell, K. A., Decker, B., Barbarino, J. M., Peterson, J. F., Stein, C. M., Sadee, W., et al. (2015). Clinical Pharmacogenetics Implementation Consortium (CPIC) guidelines for CYP3A5 genotype and tacrolimus dosing. *Clin. Pharmacol. Ther.* 98, 19–24. doi:10. 1002/cpt.113

Bolaños-Meade, J., Reshef, R., Fraser, R., Fei, M., Abhyankar, S., Al-Kadhimi, Z., et al. (2019). Three prophylaxis regimens (tacrolimus, mycophenolate mofetil, and cyclophosphamide; tacrolimus, methotrexate, and bortezomib; or tacrolimus, methotrexate, and maraviroc) versus tacrolimus and methotrexate for prevention of graft-versus-host disease with haemopoietic cell transplantation with reduced-intensity conditioning: a randomised phase 2 trial with a non-randomised contemporaneous control group (BMT CTN 1203). *Lancet Haematol.* 6, e132–e143. doi:10.1016/s2352-3026(18)30221-7

Darwich, A. S., Polasek, T. M., Aronson, J. K., Ogungbenro, K., Wright, D. F. B., Achour, B., et al. (2021). Model-informed precision dosing: background, requirements,

# Author contributions

PW: Funding acquisition, Methodology, Writing – original draft. JL: Writing – review and editing. JY: Conceptualization, Writing – review and editing.

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# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2025.1578643/ full#supplementary-material

validation, implementation, and forward trajectory of individualizing drug therapy. *Annu. Rev. Pharmacol. Toxicol.* 61, 225–245. doi:10.1146/annurev-pharmtox-033020-113257

Foti, R. S. (2025). Utility of physiologically based pharmacokinetic modeling in predicting and characterizing clinical drug interactions. *Drug Metab. Dispos.* 53, 100021. doi:10.1124/dmd.123.001384

Gerner, B., Aghai-Trommeschlaeger, F., Kraus, S., Grigoleit, G. U., Zimmermann, S., Kurlbaum, M., et al. (2022). A physiologically-based pharmacokinetic model of ruxolitinib and posaconazole to predict CYP3A4-mediated drug-drug interaction frequently observed in graft versus host disease patients. *Pharmaceutics* 14, 2556. doi:10.3390/pharmaceutics14122556

Gertz, M., Cartwright, C. M., Hobbs, M. J., Kenworthy, K. E., Rowland, M., Houston, J. B., et al. (2013). Cyclosporine inhibition of hepatic and intestinal CYP3A4, uptake and

efflux transporters: application of PBPK modeling in the assessment of drug-drug interaction potential. *Pharm. Res.* 30, 761–780. doi:10.1007/s11095-012-0918-y

Glotzbecker, B., Duncan, C., Iii, E. A., Campbell, B., and Soiffer, R. (2012). Important drug interactions in hematopoietic stem cell transplantation: what every physician should know. *Biol. Blood Marrow Transpl.* 18, 989–1006. doi:10.1016/j.bbmt.2011. 11.029

Gong, F., Hu, H., Ouyang, Y., Liao, Z. Z., Kong, Y., Hu, J. F., et al. (2023). Physiologicallybased pharmacokinetic modeling-guided rational combination of tacrolimus and voriconazole in patients with different CYP3A5 and CYP2C19 alleles. *Toxicol. Appl. Pharmacol.* 466, 116475. doi:10.1016/j.taap.2023.116475

Gratwohl, A., Baldomero, H., Aljurf, M., Pasquini, M. C., Bouzas, L. F., Yoshimi, A., et al. (2010). Hematopoietic stem cell transplantation: a global perspective. *J. Am. Med. Assoc.* 303, 1617–1624. doi:10.1001/jama.2010.491

Groll, A. H., Pana, D., Lanternier, F., Mesini, A., Ammann, R. A., Averbuch, D., et al. (2021). 8th European Conference on Infections in Leukaemia: 2020 guidelines for the diagnosis, prevention, and treatment of invasive fungal diseases in paediatric patients with cancer or post-haematopoietic cell transplantation. *Lancet Oncol.* 22, e254–e269. doi:10.1016/s1470-2045(20)30723-3

Guest, E. J., Aarons, L., Houston, J. B., Rostami-Hodjegan, A., and Galetin, A. (2011). Critique of the two-fold measure of prediction success for ratios: application for the assessment of drug-drug interactions. *Drug Metab. Dispos.* 39, 170–173. doi:10.1124/ dmd.110.036103

Kim, E. S. (2018). Letermovir: first global approval. Drugs 78, 147-152. doi:10.1007/ s40265-017-0860-8

Krishna, G., Ma, L., Martinho, M., and O'mara, E. (2012). Single-dose phase I study to evaluate the pharmacokinetics of posaconazole in new tablet and capsule formulations relative to oral suspension. *Antimicrob. Agents Chemother.* 56, 4196–4201. doi:10.1128/aac.00222-12

Li, X., Frechen, S., Moj, D., Lehr, T., Taubert, M., Hsin, C. H., et al. (2020). A physiologically based pharmacokinetic model of voriconazole integrating time-dependent inhibition of CYP3A4, genetic polymorphisms of CYP2C19 and predictions of drug-drug interactions. *Clin. Pharmacokinet.* 59, 781–808. doi:10.1007/s40262-019-00856-z

Ljungman, P., Alain, S., Chemaly, R. F., Einsele, H., Galaverna, F., Hirsch, H. H., et al. (2025). Recommendations from the 10th European Conference on Infections in Leukaemia for the management of cytomegalovirus in patients after allogeneic haematopoietic cell transplantation and other T-cell-engaging therapies. *Lancet Infect. Dis.* online. doi:10.1016/S1473-3099(25)00069-6

Loer, H. L. H., Feick, D., Rüdesheim, S., Selzer, D., Schwab, M., Teutonico, D., et al. (2023). Physiologically based pharmacokinetic modeling of tacrolimus for food-drug and CYP3A drug-gene interaction predictions. *CPT Pharmacometrics Syst. Pharmacol.* 12, 724–738. doi:10.1002/psp4.12946

Moriyama, B., Obeng, A. O., Barbarino, J., Penzak, S. R., Henning, S. A., Scott, S. A., et al. (2017). Clinical Pharmacogenetics implementation Consortium (CPIC) guidelines for CYP2C19 and voriconazole therapy. *Clin. Pharmacol. Ther.* 102, 45–51. doi:10.1002/cpt.583

Palmer, J., Mccune, J. S., Perales, M. A., Marks, D., Bubalo, J., Mohty, M., et al. (2016). Personalizing busulfan-based conditioning: considerations from the American society for blood and marrow transplantation practice guidelines committee. *Biol. Blood Marrow Transpl.* 22, 1915–1925. doi:10.1016/j.bbmt.2016.07.013

Patel, P., Leeder, J. S., Piquette-Miller, M., and Dupuis, L. L. (2017). Aprepitant and fosaprepitant drug interactions: a systematic review. *Br. J. Clin. Pharmacol.* 83, 2148–2162. doi:10.1111/bcp.13322

Paul, P., Colin, P. J., Musuamba Tshinanu, F., Versantvoort, C., Manolis, E., and Blake, K. (2025). Current use of physiologically based pharmacokinetic modeling in new medicinal product approvals at EMA. *Clin. Pharmacol. Ther. online.* 117, 808–817. doi:10.1002/cpt.3525

Pejčić, A., Janković, S. M., Opančina, V., Babić, G., and Milosavljević, M. (2019). Drug-drug interactions in patients receiving hematopoietic stem cell transplantation. *Expert Opin. Drug Metab. Toxicol.* 15, 49–59. doi:10.1080/17425255.2019.1552256

Rodriguez-Vera, L., Yin, X., Almoslem, M., Romahn, K., Cicali, B., Lukacova, V., et al. (2023). Comprehensive physiologically based pharmacokinetic model to assess drugdrug interactions of phenytoin. *Pharmaceutics* 15, 2486. doi:10.3390/ pharmaceutics15102486

Secoli, S. R., Lima, M. H. M., Trevisan, D. D., Oliveira, H. C., and Silva, J. B. (2015). Prevalence and clinical significance of potential drug-drug interaction in hematopoietic stem cell transplantation. *Cancer Chemother. Pharmacol.* 75, 393–400. doi:10.1007/ s00280-014-2657-8

Spriet, I., Meersseman, P., Meersseman, W., De Hoon, J., and Willems, L. (2010). Increasing the dose of voriconazole compensates for enzyme induction by phenytoin. *Br. J. Clin. Pharmacol.* 69, 701–702. doi:10.1111/j.1365-2125.2010.03650.x

Storelli, F., Samer, C., Reny, J. L., Desmeules, J., and Daali, Y. (2018). Complex drugdrug-gene-disease interactions involving cytochromes P450: systematic review of published case reports and clinical perspectives. *Clin. Pharmacokinet.* 57, 1267–1293. doi:10.1007/s40262-018-0650-9

Sun, Z., Zhao, N., Zhao, X., Wang, Z., Liu, Z., and Cui, Y. (2024). Application of physiologically based pharmacokinetic modeling of novel drugs approved by the U.S. food and drug administration. *Eur. J. Pharm. Sci.* 200, 106838. doi:10.1016/j.ejps.2024. 106838

Vijaywargi, G., Kollipara, S., Ahmed, T., and Chachad, S. (2023). Predicting transporter mediated drug-drug interactions via static and dynamic physiologically based pharmacokinetic modeling: a comprehensive insight on where we are now and the way forward. *Biopharm. Drug Dispos.* 44, 195–220. doi:10.1002/bdd.2339

Wang, P., Liu, S., and Yang, J. (2024). Physiologically based pharmacokinetic modeling to investigate the disease-drug-drug interactions between voriconazole and nirmatrelvir/ritonavir in COVID-19 patients with CYP2C19 phenotypes. *Clin. Pharmacol. Ther.* 116, 363–371. doi:10.1002/cpt.3222

WHO (2010). "Characterization and application of physiologically based pharmacokinetic models in risk assessment," in *World Health Organization (WHO)*, *International Programme on Chemical Safety (IPCS)*. Available online at: https://iris. who.int/handle/10665/44495.