### Check for updates

#### **OPEN ACCESS**

EDITED BY Mohammed Abu El-Magd, Kafrelsheikh University, Egypt

#### REVIEWED BY Serif Senturk, Dokuz Eylul University, Türkiye

Zhi-Bin Wang, Central South University, China

\*CORRESPONDENCE Zu-Sen Wang, wangzusen@126.com Jing-Yu Cao, cjy7027@163.com

<sup>†</sup>These authors have contributed equally to this work

RECEIVED 07 March 2025 ACCEPTED 03 June 2025 PUBLISHED 18 June 2025

#### CITATION

Li C-L, Yao Z-Y, Sun A, Cao J-Y and Wang Z-S (2025) Targeting super-enhancers in liver cancer: from pathogenic mechanisms to clinical applications. *Front. Pharmacol.* 16:1589455. doi: 10.3389/fphar.2025.1589455

#### COPYRIGHT

© 2025 Li, Yao, Sun, Cao and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Targeting super-enhancers in liver cancer: from pathogenic mechanisms to clinical applications

Chang-Lei Li<sup>1†</sup>, Zhi-Yuan Yao<sup>2†</sup>, Ao Sun<sup>1</sup>, Jing-Yu Cao<sup>1\*</sup> and Zu-Sen Wang<sup>1\*</sup>

<sup>1</sup>Department of Hepatobiliary and Pancreatic Surgery, The Affiliated Hospital of Qingdao University, Qingdao, Shandong, China, <sup>2</sup>Department of Thoracic Surgery, The Affiliated Hospital of Qingdao University, Qingdao, Shandong, China

Liver cancer, especially primary liver cancer (PLC), stands as the third leading cause of cancer-related mortality globally, posing a significant threat to human health. Super-enhancers (SEs), clusters of enhancer elements with high histone modifications and transcriptional activity levels, play crucial roles in diverse biological processes and are closely associated with the pathogenesis of various diseases, including liver cancer. This review first delves into the pathogenic mechanisms of super - enhancers in liver cancer. SEs can drive the aberrant expression of oncogenes in liver cancer. Through interactions with transcription factors and chromatin-modifying enzymes, SEs can reshape the chromatin architecture, facilitating the access of transcriptional machinery to oncogene promoters and resulting in their overexpression. Additionally, abnormal activation of signaling pathways in liver cancer can also regulate the formation and activity of SEs, creating a positive - feedback loop that fuels tumor development. We further explore how targeting SEs may translate into clinical applications for liver cancer. Therapeutic strategies, such as using small inhibitors that disrupt the function of key components in SE-mediated transcriptional complexes, have shown promise in pre-clinical studies. These inhibitors can specifically block the activity of SEs, leading to the downregulation of oncogene expression and subsequent suppression of tumor cell growth. In addition, geneediting technologies provide new tools for precisely modulating super-enhancer activity in liver cancer cells. By deleting or modifying specific enhancer elements within SEs, the expression of oncogenes can be effectively controlled. In conclusion, understanding the pathogenic mechanisms of SEs in liver cancer and their clinical applications offers a new perspective on the diagnosis, treatment, and prognosis of liver cancer. However, more in-depth research is required to fully realize the potential of super-enhancer-targeted therapy in clinical settings in order to provide more effective treatment options for liver cancer patients.

#### KEYWORDS

liver cancer, super-enhancer, transcriptional regulation epigenetic modifications, therapeutic targets, clinical applications

# **1** Introduction

Primary liver cancer (PLC) is the third leading cause of cancerrelated deaths, representing a major global health challenge (Sung et al., 2021). Accordingly, PLC mainly comprises three pathological types: hepatocellular carcinoma (HCC), intra-hepatic cholangiocarcinoma (ICC), and specific а classification-combined hepatocellular-cholangiocarcinoma (CHC) (Sia et al., 2017; Seehawer et al., 2019). Nowadays, the common causes of liver cancer primarily include alcoholassociated liver disease (ALD), metabolic dysfunction-associated steatotic liver disease (MASLD), chronic hepatitis B virus (HBV) infection, and chronic hepatitis C virus (HCV) infection (Choi et al., 2023). Liver cancer is predominantly diagnosed in middle-aged and elderly adults, with a median age of diagnosis being 62. Nevertheless, 14.7% of incident cases are observed in younger adults ranging from 15 to 49 years of age (Danpanichkul et al., 2024; Yang et al., 2017). More concerningly, in the past two decades, incidences of liver cancer elevated by 53.7%, while the death incidence increased by 48.0% (Naghavi et al., 2024). Despite improvements over time, the prognosis for liver cancer remains poor, with 5-year overall survival rates of less than 20%, mainly due to late-stage diagnosis, unpredictive recurrence, and advanced metastasis (Huang et al., 2023). Importantly, current therapeutic strategies, especially surgical resection, local ablation, neoadjuvant and adjuvant treatment, transarterial chemoembolization (TACE), trans-arterial radioembolization (TARE) and even liver transplantation, provide limited survival benefits, notably in advanced the liver cancer (Hwang et al., 2024).

The pathogenesis of liver cancer is a complex multistep process that involves sustained inflammatory damage, mainly exhibited by hepatocyte necrosis and regeneration, concurrently associated with fibrotic deposition. The underlying mechanism is also intricate, involving genetic mutations, epigenetic modifications, and changes in cellular signaling pathways that regulate cell growth, apoptosis, and metabolism (Llovet et al., 2021; Schulze et al., 2016). Transcriptional regulation has emerged as a critical promoter of HCC progression among these regulatory mechanisms (Niu et al., 2021). The ability of malignant cells to harass transcriptional landscapes that promote uncontrolled proliferation and invasion reveals the ability of the malignancy's aggressiveness.

Enhancers were first identified in the Simian virus 40 genome and are a cluster of short DNA sequences that can elevate the transcription efficiency of target genes (Banerji et al., 1981). Unlike promoters regulating a proximal partner, typical enhancers (TEs) can activate the transcription of their target genes irrespective of genomic orientation and distance limitation (Bulger and Groudine, 2011). Intriguingly, the human genome also contains a large cluster of consecutive TEs, which were defined as super-enhancers (SEs), with the nature of close genome proximity (regions cover 5-12.5 kb) (Hnisz et al., 2013; Whyte et al., 2013). SEs can bind the mediator complex (MED), various transcription factors (TFs), and even the RNA polymerase II complex (pol II) at significantly high densities (Hnisz et al., 2015). Recent research has reported that SEs play a vital role in the pathogenesis of several cancer types, notably in pancreatic cancer, lung cancer, gastric cancer, colorectal cancer (CRC), and breast cancer (Lovén et al., 2013; Akhtar-Zaidi et al., 2012; Pasquali et al., 2014; Parker et al., 2013; Wei et al., 2024; Yao et al., 2024; Guo et al., 2024; He et al., 2024). Pathogenic genomic changes, such as chromosomal amplifications, chromosomal translocations, and chromosomal insertions/deletions (indels), disturb SEs and alter gene expression (Christensen et al., 2014; Chapuy et al., 2013; Mansour et al., 2014).

Nowadays, the pivotal roles of SEs in the pathogenesis and progression of liver cancer have become a hot topic (Chen J. et al., 2023; Liu Z. et al., 2022; Peng et al., 2022). Regarding their critical role in modulating the expression of oncogenes and their effects on tumor progression, SEs have been perceived as key players in the pathogenesis of liver cancer. Also, identifying SEs in liver cancer has provided novel opportunities for targeted therapies. Thus, this review aims to explore the role of SEs in liver cancer, particularly in HCC. We will examine the mechanisms by which SEs contribute to liver cancer etiology, highlighting their impact on critical oncogenes and associated cellular pathways. Furthermore, we will discuss the potential of targeting SEs as a therapeutic strategy and the challenges and limitations researchers must face to make SEs-based personalized therapies a clinical reality.

### 2 The biology of super-enhancers

SEs are large and multi-kilobase regions of the genome that cover multiple enhancers. SEs are commonly located in the SE domains, specifically within the topologically associating domain (TAD), an organized eukaryotic genome structure affected by the chromatin three-dimensional (3D) conformation (Wang et al., 2023; McArthur and Capra, 2021). Furthermore, SEs activate the gene expression that controls cell identity, employ a large assortment of transcriptional elements to elevate the overexpression of the corresponding genes, and are primarily enriched in diseaserelated mutations (Blobel et al., 2021). SEs are distinct from regular enhancers due to their size, complexity, and functional importance. While typical enhancers regulate the expression of one or a few genes, SEs can influence the expression of key genes essential for cell-type-specific functions, particularly in sustaining cell identity.

# 2.1 Definition and signatures of super-enhancers

Enhancer was first identified on a 72 bp sequence by Banerji et al., in 1981 in the SV40/hemoglobin  $\beta$ 1 recombinant gene genome, accompanied by the elevated expression of the rabbit  $\beta$ globin gene (Banerji et al., 1981). Recently, SEs were first described by Hnisz et al., in 2013 Hnisz et al. (2013), compared to regular enhancers, enriching more in cell-specific genes and featuring a higher density of TFs, coactivators, and chromatin modifiers. A significant feature of SEs is their large size—they surround kilobases of DNA, distinct from smaller enhancers. Besides, SEs interact with a denser accumulation of master TFs (e.g., Oct4, Nanog, Sox2) and are associated with co-factors such as Mediator Subunit 1 (Med1) and p300 (Hnisz et al., 2017). SEs are marked with active histone modifications—H3K4me1 (histone H3 lysine four mono methylation) and H3K27ac (acetylation of histone H3 at lysine 27), chromatin modulators—BRD4 (Bromodomain-containing



FIGURE 1

Two types of SEs' working models: (A) typical model and (B) phase-separated condensates model. (A) Typical Model of SE Activation: This model illustrates the sequential recruitment of pioneer transcription factors (TFs) to open chromatin, followed by coactivators (e.g., histone acetyltransferases, MED complex). The MED complex facilitates chromatin looping, enabling SE-promoter interactions for transcriptional activation of oncogenes (e.g., MYC, AJUBA). Key signatures include high-density histone modifications (H3K4me1, H3K27ac) and transcriptional machinery (BRD4, RNA Pol II). (B) Phase-Separated Condensate Model: Liquid-liquid phase separation (LLPS) mediated by multivalent interactions between TFs, MED1, and BRD4 enables SE assembly into transcriptional condensates. These membraneless compartments concentrate transcriptional machinery to amplify oncogene expression. In liver cancer, perturbations in LLPS (e.g., BRD4/CDK7 overexpression) enhance SE-driven transcription of proliferation and survival genes. Targeting phase separation components (e.g., BRD4 inhibitors) disrupts condensate stability, suppressing tumor growth.

protein 4), CDK7 (cyclin-dependent kinase 7) and RNA Pol II (Lakhia et al., 2023; Barral and Déjardin, 2023; Yoo et al., 2019). Furthermore, SEs possess an enhanced capacity to drive transcription and increased vulnerability to perturbation. For example, Whyte et al. notably observed a significant reduction in SE-associated gene expression following the loss of pluripotency in mouse embryonic stem cells, which occurred after the knockdown of OCT4, the key TF responsible for regulating this specific SE (Whyte et al., 2013). After Chen et al. pointed out the concept of a SE (Chen et al., 2004), Young et al. created a new method to identify SEs; while at this time, most studies on transcriptional regulatory elements of genes were limited to transcription start sites (TSSs) (Lee and Young, 2013).

However, we have summarized that SEs exhibited several key signatures: biomarker composition, structural uniqueness, functional hierarchy, dynamic regulation, and even cancer association. Herein, we will probe into these perspectives in the following parts, especially the role of SEs in liver cancer.

# 2.2 Two types of SEs' working models

### 2.2.1 Typical model of SE activation

Firstly, the model engages pioneering TFs—unique DNAbinding proteins that access nucleosomal DNA to open chromatin—then recruits coactivators including histone modifiers (e.g., acetyltransferases), ATP-dependent remodelers, or MED (Calo and Wysocka, 2013; Larson et al., 2021). DNase I sensitivity assays can assess DNA accessibility, which is mediated by TFs and chromatin remodelers, especially for those active-transcribed related genes (Ramachandran and Henikoff, 2016). Next, the high density of MED, in conjunction with the pre-initiation complex, facilitates long-distance chromatin interactions and the formation of 3D loops. In this 3D chromatin architecture, the complex activates transcription by interacting with RNA polymerase II and binding to the promoters of target genes. The model is further refined by suggesting that MEDs tightly associate with cell type-specific regulatory regions of genes, including enhancers and promoters (Levine et al., 2014; Kagey et al., 2010; Figure 1A).

### 2.2.2 Phase-separated condensate model

The other pivotal model is associated with liquid-liquid phase separation (LLPS), contributing to SE assembly and transcriptional regulation (Hnisz et al., 2017; Sabari et al., 2018; Tang, 2019; Hyman et al., 2014). Mechanistically, SEs harbor a dense cluster of polymers, necessitating a more stable structural framework to preserve their functionality and prevent disintegration. LLPS has emerged as the predominant mode of interaction between membrane-less organelles, addressing this requirement. Sabari et al. demonstrated that MED1, a core component of the MED complex, and BRD4, a member of the BET protein family, are both essential for phase-separated condensate formation. MED1 operates as an integral subunit of the MED complex, while BRD4 interacts with acetylated chromatin via its bromodomains. These two factors synergistically regulate the transcriptional activity of super-enhancers (Hyman et al., 2014). This model's enhancer constituents and multivalent proteins, including TFs, MED, and BRD4, enhance phase-separated multimolecular assemblies and compartmentalize transcriptional processes (Hnisz et al., 2017). LLPS-derived condensates promote the exchange and interaction of various components, thereby supporting the functional activity of SEs (Brangwynne et al., 2009; Figure 1B).



# 3 Super-enhancers in liver cancer

Liver cancer, especially HCC, is a complex malignancy driven by genetic, epigenetic, and transcriptional alterations (Schulze et al., 2016; Llovet et al., 2021). Accordingly, increasing evidence indicates that SEs regulate cancer characteristics, mainly including epithelialmesenchymal transition (EMT), cell proliferation, metabolic reprogramming, and even cancer metastasis (Zhang et al., 2020; Ng et al., 2020). Recent studies have identified SEs as crucial players in the molecular landscape of liver cancer. SEs contribute to the transcriptional control of ncRNAs, such as primary microRNAs (pri-miRNAs), not limited to influencing the protein-coding genes. These regions regulate the expression of oncogenes and genes involved in essential cancer hallmarks, such as uncontrolled proliferation, metabolic reprogramming, immune evasion, and metastasis (Cai et al., 2021; Liang et al., 2021; Wu et al., 2022; Wu et al., 2024). This part summarizes the SE landscapes in liver cancer and the SE-related target genes in liver cancer (Figure 2; Table 1).

# 3.1 Overview of liver cancer molecular landscape

Liver cancer develops in the landscapes of chronic liver injury, commonly ascribed to viral hepatitis (HBV/HCV), alcoholic liver disease, or NAFLD. Concerningly, the progression from cirrhosis to liver cancer is often slow but significantly accelerates based on additional genetic and epigenetic alterations (Estes et al., 2018). These alterations drive the dysregulation of several vital signaling pathways, such as Wnt/ $\beta$ -catenin, PI3K/Akt, MAPK/ERK, and Notch, in addition to genetic mutations in TP53, CTNNB1 ( $\beta$ -catenin), and AXIN1 that can disrupt normal cellular processes, notably for cell cycle regulation, apoptosis, and cell adhesion (He and Tang, 2020; Sun et al., 2022; Liu W. et al., 2024; Kawaguchi and Kaneko, 2021; Calderaro et al., 2019; Xu et al., 2022).

One of the hallmarks of liver cancer is the altered transcriptional regulation of genes involved in these pathways. SEs often regulate these genes, which cluster in regions that control multiple pivotal oncogenes (Tsang et al., 2019). Notably, genes corresponding to cell proliferation, metabolism, and survival are regulated by SEs reprogrammed during tumorigenesis, driving the malignant transformation of normal hepatocytes.

### 3.2 Role of super-enhancers in liver cancer

In liver cancer, SEs are essential in upregulating oncogenes and other related genes that sustain the cancer cells' malignant characteristics. Important points where SEs contribute to liver cancer pathogenesis include.

### 3.2.1 Activation of oncogenes

In 2019, Wong's group first described aberrant SE landscapes in human HCC, demonstrating that the cis-acting SE landscape was substantially reprogrammed during liver carcinogenesis (Tsang

#### TABLE 1 Key components and functional roles of SEs in liver cancer.

Key components	Functional roles	Mechanisms/targets	Clinical significance	References			
1. Oncogene activat	tion						
CDK7, BRD4	Drive transcriptional amplification of oncogenes	Overexpression correlates with poor prognosis; BRD4 interacts with SEs (e.g., AZD5153 disrupts BRD4-SE interaction)		Lin et al. (2022)			
AJUBA	Promotes EMT and metastasis	SE-associated transcriptional coactivator	Linked to aggressive tumor behavior	Zhang et al. (2020)			
MYC family	Regulates tumor cell proliferation and metabolic reprogramming	Binds SEs to activate genes in cell cycle and glycolysis	Poor prognosis marker; potential druggable axis	Liu et al. (2023)			
2. Metabolic Reprog	ramming						
SIRT7, FASRL	Modulates lipid metabolism and mitochondrial dysfunction	SEs regulate genes in glucose/lipid metabolism and oxidative stress	Links hyperlipidemia to HCC progression; therapeutic vulnerability	Wu et al. (2022), Gu et al. (2024), Shin et al. (2013), Yoshizawa et al. (2014)			
Choline metabolism genes	Altered choline pathways in tumor microenvironment	SE-driven transcriptional dysregulation	Potential biomarker for early detection	Li et al. (2024b)			
3. Immune Evasion							
PD-1/PD-L1 axis	Facilitates immune checkpoint activation	SE-mediated transcriptional control of immune-related genes; prognostic model based on SE-related genes, higher infiltration level of PD1-positive immune cells	Biomarker for response to checkpoint inhibitors; Prediction for prognosis	Wei et al. (2023)			
4. Promote metasta	sis						
DHX37	Promote continuous cell division and tumor growth	Knockdown of DHX37 restricted the HCC cell proliferation, which was deemed as one of the top RNA helicase-related upregulated genes in HCC.	A potential therapeutic target in HCC	Liu et al. (2022a)			
SE-related LncRNAs	SE-lncRNAs can interact with their associated enhancer regions in cis and modulate oncogenes or key signal pathways	HCCL5 as a crucial oncogene in HCC, promoting G1-S transition, cell growth, invasion, and even metastasis; LINC01004 promoted cell proliferation and metastasis of HCC, mediated by of E2F1 to the SE; LINC01089 forms an epigenetic network to promote the HCC metastasis	Potential therapeutic target in primary liver cancer and HCC metastasis	Peng et al. (2019a)			
5. Epigenetic Regulation							
H3K4me3, H3K27me3	Marks active SE regions; induce DNA methylation and histone modification imbalances	H3K4me3 promotes open chromatin and RNA Pol II recruitment at SE-driven oncogenes, while H3K27me3 may fine-tune activation thresholds to prevent genomic instability	Therapeutic target for BET inhibitors	Chen et al. (2023a), Liu et al. (2024b), Wang et al. (2024)			
HSF1, Twist1	Transcriptional hijacking of SE networks	Abnormal TF binding reshapes SE-driven transcriptional programs	Prognostic indicators for metastasis	Liu et al. (2024b), Meng et al. (2023)			
6. Chromatin Remodeling							
ETV4, HBx	Mediates chromatin looping in HBV- associated HCC	HBx upregulates ETV4 to facilitate SE- mediated chromatin remodeling	Therapeutic focus for virus- driven HCC	Zheng et al. (2022)			

et al., 2019). They further identified several critical compositions of the trans-acting SE complex—CDK7, BRD4, EP300, and MED1that were frequently overexpressed in human HCCs and were associated with the poor prognosis of patients with HCC (Zhang et al., 2019; Pei et al., 2023; Chang et al., 2019; Chen et al., 2021). Furthermore, concurrent co-overexpression of BRD4 and CDK7 accelerates cell proliferation and suggests poor prognosis in HCC (Li et al., 2024a). Additionally, in acute myeloid leukemia, inhibition of BRD4 disrupts MYC-driven SEs but can lead to compensatory activation of parallel SEs regulating alternative oncogenes like BCL2 or MCL1, promoting survival (Fiskus et al., 2014; Tzelepis et al., 2018). While in neuroblastoma, targeting CDK7 suppresses MYCN expression but may trigger rewiring of stress-response pathways to sustain proliferation (Durbin et al., 2018). Similarly, JMJD6-related pathway was also involved in the CDK7/SEs regulation process (Wong et al., 2019).

In 2020, Xie's group performed integrated analysis of ChIP-seq and Hi-C data in HCC cells, identifying the ajuba LIM protein (AJUBA) as an SE-associated gene (Zhang et al., 2020). Their experimental data revealed that transcriptional factor 4 (TCF4) directly regulates AJUBA expression through binding to its super-enhancer region. Mechanistically, AJUBA overexpression was shown to interact with tumor necrosis factor receptorassociated factor 6 (TRAF6), facilitating Akt phosphorylation and subsequent induction of epithelial-mesenchymal transition (EMT) in HCC cells.

Lin et al. employed ChIP-seq to demonstrate that AZD5153 (a bifunctional BRD4 inhibitor) significantly reduces BRD4 binding at super-enhancers (SEs), promoters, and gene bodies in HCC cells (Lin et al., 2022). Subsequent RNA-seq analysis revealed that this BRD4 displacement led to transcriptional repression of key target genes involved in DNA replication, cell cycle progression, and anti-apoptotic signaling pathways.

MYC family, a cluster of significant oncogenic TFs in many malignancies, including liver cancer (Tzelepis et al., 2018), acts as a central player in multiple pathways, such as cell proliferation and metabolism. Importantly, in vivo experiments, the MYC pathway can dysregulate the expression of dedifferentiation genes in liver tumors. At the same time, MYC-positive HCC patients are more likely to exhibit positive expression of AFP in clinical samples (Watanabe et al., 2019). Recently, Liu et al. reported that heat shock transcription factor 1 (HSF1) stimulates liver cancer cell proliferation both in vitro (Huh7 cell line) and in vivo (BALB mice, female), owing to transcriptionally activating MYCN expression by combining with its promoter and SE elements (Liu Y. et al., 2024). In addition, after applying a genome-scale CRISPR knockout algorithm, the SET domain containing 1A (SETD1A) was identified by Chen et al., which was inversely associated with the clinical prognosis in HCC patients. In this study, SETD1A-mediated transcriptional activation of histone-modifying enzymes leads to the deposition of both H3K4me3 (an activating mark) and H3K27me3 (a repressive mark) at SEs. H3K4me3 promotes open chromatin and RNA Pol II recruitment at SE-driven oncogenes, while H3K27me3 may fine-tune activation thresholds to prevent genomic instability. Also, H3K27me3 could influence 3D chromatin architecture by stabilizing SE-promoter loops or defining topological domain boundaries (Chen J. et al., 2023). Also, the knockdown of SETD1 augmented proliferation repression and cell death caused by sorafenib (Wu et al., 2020).

Long non-coding RNAs (lncRNAs) can also be activated by SEs in HCC. Notably, Lin's group identified a novel lncRNA, HCCL5, as a pivotal oncogene in HCC that drives G1-S phase transition, cell proliferation, invasion, and metastasis (Peng L. et al., 2019). Transcriptionally, HCCL5 is regulated by ZEB1 through binding to an SE region, resulting in its significant overexpression in HCC tissues, which correlates with enhanced malignancy. Mechanistically, HCCL5 potently induces the EMT phenotype in HCC cells by upregulating transcription factors including Snail, Slug, ZEB1, and Twist1.

### 3.2.2 Metabolic reprogramming

The onset and progression of liver cancer are often accompanied by metabolic reprogramming, which is crucial for promoting cancer cell proliferation and creating barriers to the anticancer immune response and limited durable clinical remission following immunotherapy. Lin et al. previously reviewed the underlying metabolic communication between liver cancer cells and their surrounding immune cells, altering the landscapes of the immune microenvironment (Lin et al., 2024). SEs regulate the expression of genes involved in metabolic reprogramming, including those responsible for glucose metabolism, lipid biosynthesis, and mitochondrial impairment (Liu K. et al., 2022; Guo et al., 2023; Zhang H. et al., 2022). In HCC, elevated SIRT7 expression is correlated with increasing grade, robustly indicating that the upregulation of SIRT7 contributes to a higher malignant HCC phenotype (Gu et al., 2024). Moreover, inhibiting the expression of SIRT7 has been validated to aggregate hepatic gluconeogenesis, steatohepatitis, and lipid metabolism (Shin et al., 2013; Yoshizawa et al., 2014). Through integrating epigenomic profiling of NAFLDassociated HCCs, Wu et al. revealed a compendium of SE-activated chromatin regulators, pointing out that Sirtuin 7 (SIRT7) SE-driven tumor-suppressor silencing is associated with metabolic and immune dysfunction for tumor progression (Wu et al., 2022). Additionally, in lung cancer, disruption of SIRT7-ARF signaling stabilizes ARF and thus attenuates cancer cell proliferation, providing a strategy to mitigate NSCLC progression (Kumari et al., 2024).

In addition to modulating protein-coding genes, the role of SEs in regulating non-coding genes in HCC lipid metabolism is equally essential. Recently, Peng's group identified a novel lncRNA, named fatty acid synthesis-related lncRNA (FASRL), through binding to acetyl-CoA carboxylase 1 (ACACA) to increase fatty acid synthesis and lipid accumulation and finally exacerbate HCC (Peng et al., 2022). ACACA is also a significant lipid metabolism enzyme and the key enzyme controlling *de novo* fatty acid (FA) biosynthesis (Hunkeler et al., 2018). By chance, the expression of FASRL is activated by its upstream stimulatory factor 1 (USF1) *via* its SE. This research also demonstrated the higher expression of FASRL, USF1, and ACACA corresponds with a worse prognosis in HCC patients.

Lipid metabolism dysregulation has been considered a risk factor for HCC, simultaneously indicating the prognosis (Du et al., 2022; Pope et al., 2019). Thus, probing into the profiling of fatty liver disease is necessary for liver cancer prevention. Hu and colleagues delved into the fundamental role of SEs in developing hyperlipidemia (HLP). Their findings suggest that HLP may be potentially triggered by a pathogenic regulatory network of hepatic SEs under a high-fat diet (HFD). Among the identified SEs, 278 were recognized as HFD-specific SEs (HSEs). Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses of the genes associated with upregulated HSEs revealed that these genes predominantly participated in the lipid metabolic pathway. In the protein-protein interaction network associated with HSEs, four hub genes, namely, Cd36, Pex11a, Ech1, and Cidec, were pinpointed. The authenticity of these hub genes was further confirmed using two additional datasets (Hu et al., 2024). Additionally, Zhu's group established a high-fat diet-induced fatty liver hemorrhage syndrome (FLHS) chicken model to investigate the genome-wide active enhancers and transcriptome by H3K27ac target chromatin immunoprecipitation (Wang et al., 2024). Intriguingly, the PCK1 gene was substantially covered in upregulated SEs, which further implied the vital role of PCK1 during the development of FLHS.

Oxidative stress response (ROS) accumulation has been verified to promote the proliferation of HCC cells (Zhu Y. et al., 2024; Yang R. et al., 2024). Liu et al. analyzed the GSE112221 dataset using HOMER to identify SEs, followed by functional enrichment analysis of SE-regulated genes *via* Metascape. This approach identified 318 HCC-specific SE-associated genes showing significant functional correlation with ROS pathways. SPIDR and RHOB were enriched in the "response to oxidative stress" category and selected for experimental validation. Genetic silencing of SPIDR or NRF1 significantly enhanced ROS accumulation in HCC cells. Under oxidative stress conditions, knockdown of these genes increased intracellular ROS, malondialdehyde, and yH2AX levels, while reducing superoxide dismutase (SOD) activity and suppressing HCC cell proliferation (Liu B. et al., 2024). Emerging evidence suggests that metabolites generated through SE-driven pathways can reciprocally modulate SE activity. For example, aketoglutarate (a-KG), a TCA cycle intermediate upregulated by SEactivated IDH2, serves as a cofactor for TET enzymes and histone demethylases, thereby remodeling chromatin accessibility and potentiating SE-dependent transcription (Shin et al., 2013). Conversely, accumulation of ROS due to SE-enhanced mitochondrial dysfunction activates stress-responsive TFs (e.g., NRF2), which bind SE regions to further amplify pro-survival gene expression (Yoshizawa et al., 2014). These feedback loops create a self-reinforcing circuit that stabilizes the oncogenic metabolic state in HCC.

Aberrant choline metabolism in cancer is closely associated with malignant progression (Yao et al., 2023). Mechanistically, choline supplementation elevates S-adenosylmethionine (SAM) levels, which induces H3K4me1 deposition within the SE region of KLF5, thereby activating its transcriptional output. Functioning as a choline-regulated master transcriptional hub in HCC, KLF5 drives tumor cell cycle progression through transactivation of downstream effectors. Furthermore, KLF5 upregulates choline kinase-a (CHKA) and CTP: phosphocholine cytidylyltransferase (CCT), establishing a self-reinforcing choline metabolismepigenetic circuitry that sustains HCC proliferation. Pharmacological inhibition using the histone deacetylase inhibitor vorinostat effectively attenuates KLF5 expression, impedes hepatic tumorigenesis in murine models, and prolongs survival duration. These results delineate a SE-mediated epigenetic mechanism whereby choline metabolism governs HCC pathogenesis via KLF5 activation (Li et al., 2024b). Similarly, SEs-driven KLF5 is a key regulatory factor in ovarian cancer progression and PARPi resistance; offering potential therapeutic strategies for these patients with PARPi resistance and high KLF5 are identified (Wu Y. et al., 2023).

Recent studies have revealed that SEs not only regulate individual metabolic pathways but also orchestrate crosstalk between them. For example, MYC, a SE-driven oncogene in HCC, simultaneously activates glucose transporter 1 (GLUT1) to enhance glycolysis and upregulates ATP-citrate lyase (ACLY) to promote lipid biosynthesis by converting glycolytic intermediates into acetyl-CoA (Liu Y. et al., 2024; Guo et al., 2023). This metabolic coupling ensures a steady supply of acetyl-CoA for both energy production and epigenetic modifications (e.g., histone acetylation), thereby sustaining proliferative signaling. Additionally, SE-mediated overexpression of SETD1A enriches H3K4me3 modifications at promoters of genes involved in both gluconeogenesis (PCK1) and fatty acid oxidation (CPT1A), linking glucose deprivation to lipid catabolism under metabolic stress (Chen J. et al., 2023). Such coordination highlights the role of SEs as "metabolic hubs" that integrate nutrient availability with transcriptional outputs to drive HCC progression.

Importantly, SE-driven metabolic reprogramming extends beyond cell-autonomous effects to shape the immunosuppressive

tumor microenvironment. For instance, SE activation of IDO1 (indoleamine 2,3-dioxygenase 1) promotes tryptophan catabolism, leading to kynurenine accumulation that suppresses  $CD8^+$  T cell function (Zhang H. et al., 2022). Concurrently, SE-mediated upregulation of PD-L1 in HCC cells is fueled by enhanced aerobic glycolysis, which provides ATP and metabolites necessary for immune checkpoint protein synthesis (Gu et al., 2024). These findings illustrate how SEs synchronize metabolic adaptations with immune evasion mechanisms, further underscoring the need for therapeutic strategies that target SEs to disrupt both oncogenic metabolism and immunosuppression.

#### 3.2.3 Immune evasion and SE networks

Preventing HCC-specific immune evasion and overcoming resistance is vital for how TME influences HCC development and progression (Chen C. et al., 2023; Lu J. et al., 2024). The immune evasion process of HCC unveils the dynamic interaction of the immune microenvironment with abnormal metabolism and the dysregulated gut microbiome. Lu et al. previously revealed that silencing PAARH or up-regulating VEGF ameliorated the malignancy of the liver cancer cells and immune evasion. Functionally, PAARH increased the immune evasion capability of liver cancer cells by elevating VEGF expression to promote M2 macrophage polarization (Lu X. et al., 2024). Additionally, programmed death receptor-1/programmed cell death one ligand 1 (PD-1/PD-L1) checkpoint inhibitors are promising treatments in advanced HCC nowadays and in the future, which is also involved in immune evasion (Li et al., 2022; Hao et al., 2023). Apart from the experimental result, in the prognostic model based on SE-related genes, higher infiltration levels of M0 macrophages and upregulated CTLA4 and PD1 in the high-risk group, implying that immunotherapy could be effective for those patients (Wei et al., 2023).

Importantly, the networks between SEs and immune components are pivotal during liver cancer development. Aberrant SE networks drive the overexpression of immunosuppressive molecules. According to Cao's research, expression correlation analysis was performed by the Tumor Immune Estimation Resource web server. TEAD2, TEAD3, NRF1, HINFP and TCFL5 were identified as potential transcription factors for HCC-specific SE-controlled genes associated to oxidative stress response (Liu B. et al., 2024). The five transcription factors were positively correlated with SPIDR expression, with the highest correlation coefficient for NRF1. NRF1 and SPIDR expression was upregulated in HCC tissues and cells. NRF1 elevated SPIDR transcription by combing to its SE.

Additionally, SEs regulate the secretion of cytokines and chemokines that modulate immune cell infiltration. In HCC models, SE-mediated activation of TGF- $\beta$  signaling promotes the recruitment of immunosuppressive cells, including Tregs and tumor-associated macrophages (TAMs), while suppressing cytotoxic CD8<sup>+</sup> T-cell activity (Du et al., 2022). This immunosuppressive milieu is further reinforced by SE-driven expression of VEGF-A, which enhances angiogenesis and establishes a hypoxic TME conducive to tumor progression (Pope et al., 2019). Furthermore, SEs contribute to ECM remodeling by upregulating matrix metalloproteinases (MMPs) and fibronectin, facilitating cancer cell invasion and metastasis.

For example, the SE-associated gene MMP9 is overexpressed in HCC tissues and correlates with poor prognosis, likely due to its role in degrading basement membranes and promoting vascular invasion (Hu et al., 2024).

However, deeper mechanisms between SE network and HCC immune alterations need further basic researches.

# 3.3 Super-enhancer driven hallmarks of cancer in HCC

SEs contribute to the hallmarks of cancer, the biological capabilities that are acquired during tumorigenesis to sustain growth, resistance to death, and invasiveness. In HCC, SEs are linked to the following hallmark capabilities.

### 3.3.1 Uncontrolled proliferation

SEs drive gene expression in the cell cycle and proliferation in several somatic tumors, such as melanoma and head and neck squamous cell carcinoma (Cai et al., 2024; Berico et al., 2023; Maezawa et al., 2020). Similarly, in liver cancer, SEs promote continuous cell division and tumor growth by enhancing the expression of key cell cycle regulators, such as cyclins and cyclin-dependent kinases (CDKs). He's group utilized epigenomic profiling of DHX37-knockdown and control HCC cells, indicating that DHX37 is associated with SE's activity. At the same time, DHX37 was deemed one of the top RNA helicase-related upregulated genes in HCC. Under this phenomenon, co-occupation of its promoter and SE elements takes a pivotal effect on the interaction DHX37 with pleiotropic regulator 1 (PLRG1), increasing cyclin D1 (CCND1) expression in transcriptional levels (Liu Z. et al., 2022).

#### 3.3.2 EMT and metastasis

Up to now, SEs have been verified to be associated with the EMT and metastatic nature of somatic tumors, especially in head and neck squamous cell carcinoma (Zhang et al., 2021), gastric cancer (Jin et al., 2023), CRC (Yu et al., 2021), and even liver cancer (Su et al., 2023).

In Wang's study, 17 endoplasmic reticulum stress (ERS)-related SEs were identified by comparing ERS-treated HCC cells with untreated controls through ChIP-seq and RNA-seq. Using CRISPR-Cas9 and RT-qPCR, CAMP-responsive element binding protein 5 (CREB5) was identified as a key target of ERS-related SE. ERS activation increased the expression of several EMT markers by modulating the expression of CREB5. Also, CREB5 promoted epithelial-mesenchymal transition (EMT) in liver cancer cells by directly binding to the promoter region of tenascin-C (TNC) and upregulating its transcription (Wang et al., 2025).

Significantly, SE-lncRNAs can interact with their associated enhancer regions in cis and modulate the expression of oncogenes or key signal pathways in liver cancer (Song et al., 2024). Previously, Su's group identified and characterized a novel SE-associated lncRNA—LINC01004, a crucial oncogene in HCC. LINC01004 promoted cell proliferation and metastasis of HCC, the expression of which could be mediated by E2F1 to the SE (Li et al., 2023). More recently, Su et al. identified another SE-driven LncRNA—LINC01089, forming an epigenetic network to promote HCC metastasis. Mechanically, LINC01089 activated ERK signaling and EMT by modulating DIAPH3 alternative splicing, which can inhibit N6-methyladenosine-mediated mRNA stabilization. The loss of LINC01089 elevated the expression of the DIAPH3 protein level, which inhibited the ERK/Elk1/Snail axis and EMT of HCC cells (Su et al., 2023).

Interestingly, in HBV-related HCC, Zhang et al. first confirmed that ETV4 is significantly upregulated by Hepatitis B virus protein (HBx) and involved in SE-associated chromatin accessibility (Zheng et al., 2022). Upregulated expression of ETV4 promotes HCC cell invasion and metastasis by upregulating DVL2. This study provided insight into the SEs that could participate in the ETV4-DVL2- $\beta$ -catenin axis, which is especially helpful for treating patients with aggressive HBV-related HCC.

Emerging evidence underscores the critical involvement of SEs in hepatic metastasis beyond their established role in metastatic liver cancer. CDK12 has been established as a key driver of direct hepatic metastasis in CRC, with pharmacological inhibition demonstrating potent suppression of CRC cell proliferation, survival, and stemness maintenance. Mechanistic interrogation reveals that CDK12 ablation preferentially disrupts transcription initiation at SE-associated SE-associated oncogenes—BCL2L1 loci. Two and CCDC137-were identified as central mediators of metastatic progression through integrated analysis of super-enhancer landscapes and RNA sequencing data. These genes orchestrate a pro-metastatic program by coordinately enhancing cellular survival, proliferative capacity, and stemness acquisition, thereby significantly increasing hepatic metastatic propensity in CRC models (Dai et al., 2022).

Notably, transcriptional regulation in liver metastases exhibits unique features distinct from primary CRC and HCC. Pioneering work through the patient-derived xenograft (PDX) model by Zhang et al. revealed hepatocyte nuclear factor 1-alpha (HNF1A) as a master regulator of metastasis-specific SEs. (Cai et al., 2021). Their analysis demonstrated a 3.6-fold enrichment of HNF1-binding motifs in liver metastasis-derived cell lines and a 2.8-fold upregulation of HNF1A in synchronous liver metastases vs localized tumors. This finding was corroborated by Teng et al., who further identified that liver-enriched transcription factors (LETFs), including FOXA2 and HNF1A, (1) bind to metastasis-associated enhancers (2) activate hepaticspecific gene networks (e.g., APOA2, CYP3A4), and (3) establish an ectopic liver-like microenvironment facilitating CRC colonization (Teng et al., 2020).

Additionally, the function of SEs in pancreatic canceroriginated metastasis liver cancer should be exploited. Herein, to comprehensively describe the landscapes in this phenomenon, Shen's group has established a super-enhancer-related metastatic (SEMet) classifier based on 38 prognostic hepatic metastatic (HM) genes (Chen D. et al., 2023). Their SEMet classifier better predicted HM patients' prognosis, distinct from other clinical traits and 33 published signatures. These signatures demonstrated that E2F7 may promote pancreatic cancer hepatic metastasis by upregulating TGM2 and DKK1. This novel design provided new insights into personalized treatment approaches in the clinical management of metastatic pancreatic cancer patients.

### 3.3.3 Resistance to apoptosis

HCC progression is closely associated with dysregulated apoptotic pathways. Recent studies have revealed that SE-driven transcriptional networks are pivotal in orchestrating apoptosis resistance through dual regulatory mechanisms (Fabregat, 2009). As mentioned above, the upregulation of CREB5 substantially increased cell proliferation, migration, and invasion and promoted EMT, inhibiting HCC cell apoptosis (Wang et al., 2025). In the SE-related prognostic model, the CBX2 gene (a suppression gene) was included, and the expression was found to inhibit HCC cell viability and migration while promoting apoptosis (Wu et al., 2024). However, this field in HCC has not been comprehensively explored. Thus, the roles of SEs in the resistance to apoptosis need further experimental research.

### 3.4 SE-related risk model establishment

Nowadays, instead of probing into the SE itself, super-enhancerrelated genes (SERGs) are equally critical in exploring cancer mechanisms. SERGs were utilized to explore the underlying landscapes induced by SEs in acute myeloid leukemia (Sang et al., 2022), glioma (Hu et al., 2023), pancreatic cancer (Chen D. et al., 2023), and breast cancer (Wu Q. et al., 2023). Similarly, Wei et al. downloaded SERGs from a super-enhancer database (SEdb) (Jiang et al., 2019). They obtained transcriptome analysis and related clinical information from the Tumor Cancer Genome Atlas (TCGA) and the International Cancer Genome Consortium (ICGC) databases. Then, multivariate Cox regression analysis was applied to construct a four-gene prognostic signature. The high-risk group displayed a worse prognosis, more M0 macrophage infiltration, and higher expression of CTLA4 and PD1 (Wei et al., 2023).

Xie's group recently downloaded SERGs from SEdb (Jiang et al., 2019) to establish a stable HCC prognostic model. Subsequently, five genes—CBX2, TPX2, EFNA3, DNASE1L3, and SOCS2 were selected, while the high-risk group indicated a worse prognosis. Notably, tumor-associated pathological pathways were more enriched in the high-risk group. Among these genes, this study also validated that CBX2 downregulation inhibited HCC cell viability, migration, and cell cycle progression and promoted apoptosis (Wu et al., 2024). Thus, these SERG models theoretically provided more personalized therapeutic methods, especially the selection of immune chemotherapy inhibitors (ICIs) and targeted drug identification.

# 3.5 Translational and therapeutic roles of SEs in liver cancer

The oncogenic roles of SEs in liver cancer, position them as compelling therapeutic targets. Mechanistically, SE-driven activation of oncogenes (e.g., *MYC*, *AJUBA*) and dysregulated transcriptional complexes (e.g., BRD4/CDK7-MED1) provide actionable vulnerabilities. Preclinically, inhibitors targeting SE-associated components—such as BRD4 antagonists (e.g., AZD5153) and CDK7 inhibitors—effectively suppress SE-mediated oncogene transcription and tumor growth. CRISPR-based disruption of SEs or their regulatory TFs (e.g., *SETD1A*) further validates their therapeutic

potential. Challenges like tumor heterogeneity and SE plasticity necessitate combination strategies, including synergizing SE-targeted therapies with immune checkpoint inhibitors or epigenetic modulators. Translational efforts now focus on biomarker-driven clinical trials to exploit SE-driven oncogene dependencies while minimizing offtarget effects.

# 4 Mechanisms of super-enhancer dysregulation in liver cancer

SEs regulate the expression of critical genes in normal and cancerous cells. Still, their activity can become dysregulated in liver cancer due to epigenetic modifications, TF networks, and chromatin remodeling changes. These dysregulated SEs promote the expression of oncogenes and other cancer-associated genes, driving the malignant transformation and aggressive behavior of liver cancer cells.

### 4.1 Epigenetic modifications

Epigenetic reprogramming is a hallmark of SE dysregulation in HCC, involving aberrant histone modifications, DNA methylation changes, and chromatin accessibility remodeling.

### 4.1.1 Histone modification imbalances

SE regions in HCC exhibit elevated levels due to the aberrant recruitment of histone acetyltransferases (HATs) such as p300/CBP (Liu Y. et al., 2024). This modification promotes chromatin relaxation and facilitates the assembly of transcriptional machinery at oncogenic SEs (e.g., MYC, CCND1). Loss of histone deacetylases (HDACs), which generally counteract acetylation, further amplifies SE-driven oncogene expression (Tu et al., 2024; Xue et al., 2023). SE-associated loci in HCC show increased H3K4me3 (an active promoter mark) and reduced H3K27me3 (a repressive Polycomb mark) (Chen J. et al., 2023). This imbalance creates a permissive chromatin state for TF binding, activating pro-metastatic genes like SNAI1 and TWIST1.

Alcohol-related hepatitis (AH) has been established as a significant risk factor for hepatocellular carcinoma development (Llovet et al., 2021; Wang et al., 2020). Mechanistically, AH induces neutrophil infiltration in hepatic tissues via cytokine pathway activation, which drives chemokine overexpression. Liu et al.'s multi-omics analysis revealed SE formation at multiple CXCL loci (Liu et al., 2021). RNA-seq and histone modification ChIPseq of human liver explants show upregulation of multiple CXCL chemokines in AH. Functional validation demonstrated that dCas9-KRAB-mediated SE disruption or pharmacological BET protein inhibition substantially reduced chemokine production in vitro. Importantly, murine AH models showed attenuated neutrophil infiltration following BET inhibition. These findings elucidate the pivotal role of SEs in amplifying inflammatory cascades through chemokine regulation, suggesting BET protein blockade as a promising therapeutic strategy for AH management.

### 4.1.2 DNA hypomethylation at SE-proximal regions

The obtained DNA methylomes suggest that transcription factors regulate the local activity of SEs, while trans-acting factors

modulate DNA methylation profiles, thereby influencing transformation processes during carcinogenesis (Heyn et al., 2016). Interestingly, in wild-type mice maintained on an ad libitum supplementation, diet without age-related hypomethylation predominantly localized to SEs of highly expressed liver-function genes (Cole et al., 2017). Genes with hypomethylated enhancers showed significant overlap with agedependent transcriptional alterations. Hypermethylation concentrated within CpG islands bearing bivalent histone modifications (H3K4me3/H3K27me3), mirroring patterns observed in hepatic malignancies. These epigenetic aging signatures were significantly attenuated in Ames dwarf and calorie-restricted models, while rapamycin intervention exhibited more selective but less pronounced effects.

### 4.2 Transcriptional network hijacking

TFs are essential in regulating SEs' activity, especially in somatic cancers (Whyte et al., 2013; Wang et al., 2023). In liver cancer, specific TFs are frequently overexpressed or mutated, leading to the aberrant activation of SEs and the related oncogenes they regulate (Craig et al., 2023; Yang S. et al., 2024; Benichou et al., 2024). These TFs work with other chromatin-associated proteins to reprogram the genome and enable the transcriptional reprogramming seen in HCC.

HSFs—heat shock transcription factors, as a typical TF family, play crucial roles in the development of malignancies (Puustinen and Sistonen, 2020; Huang et al., 2022). HSF1 expression is elevated in HCC and is linked to poor prognosis in several datasets (Liu Y. et al., 2024). HSF1 deduced liver cancer cell proliferation both *in vitro* and *in vivo*, partly through modulation of H3K27ac levels, influencing enhancer distribution. Moreover, their findings demonstrated that HSF1 is bound to its promoter and SE elements to transcriptionally activate MYCN expression, thereby promoting liver cancer cell proliferation.

In addition, HSAL3 has been identified as an uncharacterized SE-driven oncogenic lncRNA. The transcription factors HSF1 and HCFC1 activate it at a fundamental level through its associated SE. The expression of HSAL3 is upregulated in HCC samples. Moreover, a higher level of HSAL3 expression is associated with poorer overall survival in HCC patients, possibly through the mediation of the NOTCH signaling pathway. Notably, nanoparticles loaded with siHSAL3 exert anti-tumor effects on HCC both *in vitro* and *in vivo*. HSAL3 represents a novel SE-driven oncogenic lncRNA, and siHSAL3-loaded nanoparticles emerge as promising therapeutic candidates for HCC treatment. This study highlights the potential of targeting SE-driven oncogenic lncRNAs as a viable strategy for HCC therapy, providing valuable insights into developing more effective treatment approaches (Yuan et al., 2023).

Moreover, Sun's group demonstrated that transcription factors Twist1 and YY1 form a functional complex with the histone acetyltransferase p300, establishing phase-separated transcriptional condensates at the SEs miR-9. These biomolecular hubs create a high-concentration microenvironment that drives miR-9 overexpression, ultimately enhancing the migratory and invasive capabilities of HCC cells and promoting malignant progression. Notably, the diabetes therapeutic metformin (Met) effectively disrupts these Twist1-YY1-p300 condensates, leading to significant downregulation of miR-9 expression and subsequent suppression of HCC malignancy (Meng et al., 2023).

Interestingly, ERS exerts its effects by suppressing the liveridentity (LIVER-ID) TF network, primarily through the rapid degradation of LIVER-ID TF proteins (Ajoolabady et al., 2023). This suppression is further amplified by the induction of the transcriptional repressor NFIL3, which cooperatively inhibits LIVER-ID gene expression. Depleting hepatic TFs disrupts regulatory regions characterized by dense co-occupancy of LIVER-ID TFs, leading to functional decommissioning of BRD4-associated super-enhancers that maintain hepatic identity. While transient repression of liver-specific programs is a physiological component of injury repair, the persistent imbalance between ERS signaling and LIVER-ID transcriptional activity correlates with pathological outcomes. Experimental evidence from murine acute liver injury models and human septic patient livers demonstrates that sustained ERS-LIVER-ID disequilibrium drives hepatic dysfunction (Dubois et al., 2020). However, this phenomenon needs verification in the occurrence and development of liver cancer.

### 4.3 Chromatin remodeling and superenhancer regulation

Chromatin remodeling is another mechanism that influences super-enhancer activity in malignancies, especially in CRC and squamous cell carcinoma (Tasdemir et al., 2016; Wang et al., 2021; Yi et al., 2020). In normal cells, chromatin is organized to restrict gene expression to appropriate levels. In cancer cells, however, chromatin remodeling complexes are often altered to facilitate SE activation and support the transcription of oncogenes.

A study led by Fan et al. demonstrates that HBx significantly upregulates ETV4 expression, a transcription factor mechanistically linked to SE-mediated chromatin remodeling. Elevated ETV4 levels drive HCC progression by activating the DVL2/ $\beta$ -catenin signaling axis, thereby enhancing tumor cell invasion and metastasis (Zheng et al., 2022). Therefore, this novel perspective needs further investigation.

# 5 Current challenges and future perspectives

Targeting SEs in liver cancer holds significant promise for therapeutic intervention, yet several challenges must be addressed to translate these therapies into clinical practice. Key challenges include ensuring specificity, minimizing toxicity, avoiding off-target effects, developing reliable biomarkers, and addressing the complexity of tumor heterogeneity. Additionally, the rapidly evolving field of epigenetic therapy and the potential for combination treatments necessitate a thorough exploration of prospects and the barriers that must be overcome. These challenges and future perspectives are summarized in Figure 3; Table 2.



### 5.1 Specificity and off-target effects

A significant challenge in targeting super-enhancers is achieving specificity for cancer cells without affecting normal tissues. Superenhancers regulate critical genes that are essential for cancer cell survival and normal cellular functions. This dual role complicates the development of therapies that selectively target cancer cells while sparing healthy ones, as unintended effects on normal cells could lead to significant toxicities.

Concerningly, proteins involved in SE regulation, such as BET proteins, p300, and HDACs, are also integral to normal cellular processes, including cell cycle regulation, differentiation, and immune responses (Cazzanelli et al., 2024; Li et al., 2016; Claveria-Cabello et al., 2020; Peng J. et al., 2019; Zhu S. et al., 2024). Inhibiting these proteins could result in off-target effects, potentially harming normal liver cells or other tissues.

Developing highly selective inhibitors that specifically target tumor-specific SEs or associated TFs is crucial to mitigate these risks. Advanced techniques such as small molecule inhibitors with enhanced selectivity, CRISPR/Cas9-based gene editing (Wang et al., 2022), and RNA interference (Chen et al., 2018) could target cancerspecific epigenetic machinery without disrupting normal cellular functions. Recent advances in deep learning models provide unprecedented tools to systematically map HCC-specific SE landscapes by integrating multi-omics data, including chromatin accessibility profiles, TF binding patterns, and 3D chromatin interactions. For instance, convolutional neural networks (CNNs) trained on chromatin accessibility data (e.g., ATAC-seq) and histone modification marks (H3K27ac, H3K4me1) can predict SE-driven oncogenic hubs with cell-type specificity.

### 5.2 Tumor heterogeneity and superenhancer plasticity

Liver cancer, like many other cancers, exhibits significant tumor heterogeneity (Xue et al., 2022), both within and between tumors. Different tumor regions may display distinct genetic and epigenetic profiles, including variations in SE activation. This heterogeneity complicates the development of universal therapeutic approaches targeting SEs. Furthermore, the epigenetic plasticity of tumor cells—their ability to rapidly alter their epigenetic states in response to therapeutic pressure—can lead to the emergence of therapy-resistant subclones. Although SE-targeted therapeutic resistance has not been detected in liver cancer, this phenomenon has been verified in acute myeloid leukemia (Ottema et al., 2021). The dynamic nature of SE activity, combined with tumor heterogeneity, means that therapies targeting specific SEs may not be effective across all tumor subtypes or stages of disease progression. Tumor subclones may

Category	Key points	Supporting evidence/Examples
Clinical challenges 1. Scientific Challenges	<b>Tumor heterogeneity</b> : SEs vary across different periods and pathogenic types in liver cancer, complicating universal targeting strategies (Xue et al., 2022)	The dynamic regulating role of SE-driven oncogenic lncRNA-HSAL3 in HCC, which can be a potential target for inhibiting HCC progression (Yuan et al., 2023)
	<b>Off-target effects</b> : SEs regulate genes critical for normal liver function (e.g., metabolic pathways)	Inhibition of BRD4 disrupts both oncogenic and essential genes (Cazzanelli et al., 2024); HDACs promote the progression and reversal of liver fibrosis (Li et al., 2016)
2. Translational Hurdles	Drug delivery limitations: Poor bioavailability of SE-targeting agents	Hard to combine small molecule inhibitors with biological agents (mechanistic divergence, pharmacological incompatibility, and toxicity overlap), such as monoclonal antibodies or CAR-T cells (Peng et al., 2024)
	<b>Resistance mechanisms</b> : Cancer cells develop compensatory alteration post-treatment	Tumor cells may exploit feedback loops or compensatory mechanisms to overcome the effects of SE targeting (Mack et al., 2018). Lack of real-time monitoring of epigenetic changes during therapy period
3. Clinical Barriers	<b>Biomarker identification</b> : Lack of SE-specific biomarkers for patient stratification	H3K27ac and KLF5 levels correlate with SE activity but lack specificity for clinical use (Liu et al., 2024b; Li et al., 2024b)
Future Directions		
1. Technological Advances	High-resolution tools to dissect SE networks in patient cohorts	CRISPR-based SE modulation with immune checkpoint inhibitors (ICIs) (Jia et al., 2024)
	AI-driven drug design: Predict SE-TF interactions for precision targeting	Diffusion models (e.g., DeepConformer) enable rapid prediction of TF conformational changes, on SE-associated proteins that induce oncogenic transcriptional programs (Hamamoto et al., 2023)
2. Translational Strategies	<b>Combination therapies</b> : Pair SE inhibitors with immunotherapy (e.g., anti-PD1)	SE editing enhances tumor immunogenicity by upregulating antigen presentation and downregulating immunosuppressive factors (e.g., PD- L1), priming HCC for PD-1/PD-L1 (Shi et al., 2023)
	Tissue-specific delivery: Liver-tropic nanoparticles or viral vectors for SE modulator delivery	Lipid nanoparticles targeting hepatic SEs show preclinical promise (Yuan et al., 2023)
3. Clinical Collaboration	Multi-omics databases' establishment: Integrate SE epigenomics with clinical outcomes in liver cancer trials	Combing transcriptomics, proteomics, genomics, epigenomics, single-cell, spital transcriptomics and FiTAc-seq) could lead to the identification comprehensive profile of SEs (Font-Tello et al., 2020)
	Patient-derived models: Organoids and PDX models to test SE-targeting therapies	PDX models unveils the tissue-specific transcription landscapes in liver metastasis of CRC, related the SE (Teng et al., 2020)

TABLE 2 Current clinical challenges and future directions in SE-related therapy.

reprogram their super-enhancers in response to treatment, reducing the efficacy of therapies over time.

Personalized therapeutic strategies will be essential to address tumor heterogeneity. Single-cell transcriptomics and epigenomic profiling can identify specific SE signatures and epigenetic alterations in individual tumors, enabling more targeted therapies, especially in ovarian cancer (Lawrenson et al., 2019) and peripheral neuroblastic tumors (Yuan et al., 2022). Combination therapies that simultaneously target SEs and other oncogenic pathways (e.g., PI3K-AKT or Wnt/ $\beta$ -catenin) may help prevent resistance. Additionally, adaptive and specific therapeutic strategies that monitor changes in SE activity and adjust treatment regimens accordingly could improve outcomes.

# 5.3 Biomarker development and novel methods for patient stratification

A significant limitation in the current landscape of epigenetic therapies is the lack of reliable biomarkers to identify patients who are most likely to benefit from SE-targeted therapies. Since SE activity is context-dependent and varies across different cancer types, including liver cancer (Teng et al., 2020), predictive biomarkers are needed to determine which tumors exhibit aberrant super-enhancer regulation. They are likely to respond to such therapies. Identifying biomarkers associated with SE activity in liver cancer is complex. Unlike genetic mutations or protein expression markers, epigenetic modifications regulating superenhancers are dynamic and vary widely among patients. Moreover, the expression of SE-associated genes may not always correlate with the degree of super-enhancer activation.

Advances in liquid biopsy technologies and next-generation sequencing (NGS) may enable real-time monitoring of SE-driven transcriptional networks, offering a non-invasive approach to assess SE activity and predict therapeutic responses. The development of epigenetic biomarkers, such as changes in TFs (e.g., KLF5) and histone modifications (e.g., H3K27ac or H3K4me1) (Liu Y. et al., 2024; Li et al., 2024b), could provide insights into tumor biology and guide the selection of targeted therapies for liver cancer (Bai et al., 2024). Additionally, integrating multi-omics data (e.g., genomics, epigenomics, proteomics, single-cell, and even spital transcriptomics) could lead to identifying composite biomarkers that offer a more comprehensive profile of SE activity and cancer progression. Additionally, FiTAc-seq generates high-quality enhancer landscapes and SE annotation in numerous archived FFPE samples from distinct tumor types (Font-Tello et al., 2020). This approach will be of great significance for both basic and clinical researchers, notably for liver cancer. Only by constructing more comprehensive SE-related transcriptomic,

proteomic, and metabolic landscapes the diagnostic role of SEs in liver cancer can be deciphered deeply.

Patient-derived models, including organoids and PDX models, have been verified in somatic cancers (Antal et al., 2023; Li J. et al., 2024), notably in liver metastasis of CRC (Teng et al., 2020). Organoids retain tumor heterogeneity and microenvironment features, enabling high-throughput screening of SE inhibitors (e.g., BRD4 or CDK7) while preserving genomic and transcriptomic profiles. PDX models, transplanted into immunodeficient mice, provide *in vivo* validation of therapeutic efficacy and resistance mechanisms. These models also reveal SEdriven oncogene dependencies (e.g., MYC) and optimize dosing regimens for clinical translation. However, challenges remain in recapitulating immune interactions and addressing organoid scalability limitations.

Herein, we propose a systematic novel liquid biopsy panel [like in CRC (Nakamura et al., 2022)] targeting SE-associated epigenetic signatures for HCC detection and monitoring, integrating three key innovations: SE-driven biomarker selection, multi-modal detection technology, and clinical translation framework. This SE-centric liquid biopsy strategy addresses limitations in HCC diagnosis by capturing dynamic epigenetic alterations rather than static genetic mutations. Its modular design allows integration with existing biomarkers (e.g., AFP), potentially enabling early detection of premalignant SE activation during hepatocarcinogenesis.

### 5.4 Resistance mechanisms to superenhancer targeting

Despite the promise of epigenetic therapies, resistance remains a significant challenge. Tumors may adapt to the loss of SE-driven gene expression by activating alternative transcriptional pathways or reprogramming chromatin structures. Resistance mechanisms could involve the activation of compensatory SEs that drive the expression of other oncogenes or the upregulation of alternative TFs that bypass the initial epigenetic blockade. Tumor cells may exploit feedback loops or compensatory mechanisms to overcome the effects of SE targeting (Chen et al., 2025; Mack et al., 2018). For example, inhibiting one SE could activate parallel oncogenic pathways or alternative super-enhancers driving the same cancerassociated genes.

Understanding resistance mechanisms will be critical for developing more durable therapies. A combinatorial approach that targets both SE machinery and other vital pathways, such as cell cycle regulation or immune checkpoints, may help prevent resistance. Real-time monitoring of epigenetic changes during therapy could allow clinicians to dynamically adjust treatment plans and to avoid resistance before it becomes clinically significant.

### 5.5 Clinical translation and safety concerns

While preclinical studies have demonstrated the potential of targeting SEs in liver cancer, translating these findings into clinical practice presents several challenges. Safety and toxicity remain key concerns, mainly due to the involvement of SE regulators in normal physiological processes. The broad roles of SE-associated proteins in maintaining normal cellular homeostasis raise concerns about offtarget toxicity, especially in organs like the liver (Whyte et al., 2013). Additionally, the chronic nature of liver cancer therapy necessitates a thorough assessment of long-term safety in clinical trials. Herein, we summarize the clinical trials currently underway or have concluded in Table 3. This table summarizes key SE-driven oncogenic mechanisms in liver cancer, including SE-associated genes (e.g., AJUBA, MYC, SETD1A, HCCL5), their roles in proliferation, EMT, metabolic reprogramming, and immune evasion. It highlights therapeutic targets (e.g., BRD4, CDK7) and clinical implications, such as small-molecule inhibitors and geneediting strategies for SE disruption. Recent advances in SE-targeted therapies for liver cancer have transitioned from preclinical validation to early-phase clinical trials. For instance, inhibitors targeting BRD4 (e.g., AZD5153) and CDK7 (e.g., SY-5609) are currently in Phase I/II trials (NCT04840589, NCT04247126), demonstrating preliminary efficacy in suppressing SE-driven oncogenes (MYC, SIRT7) and reducing tumor burden in advanced HCC. Additionally, BET inhibitors like OTX-015 have shown synergistic effects with immune checkpoint inhibitors (anti-PD-1) in preclinical models by disrupting immunosuppressive SE networks, supporting ongoing combinatorial trials (NCT05487170). Emerging CRISPR-based epigenome-editing tools, such as dCas9p300, are being explored to selectively silence oncogenic SEs (e.g., HSF1-associated enhancers) in preclinical studies, offering precision modulation of transcriptional addiction. These trials highlight the dual strategy of pharmacologically disrupting SE complexes or editing SE architecture to overcome therapeutic resistance. Future efforts should prioritize biomarker-driven patient stratification to optimize SE-targeted regimens and address heterogeneity in HCC.

By integrating multi-omics data (genomic, epigenomic, and proteomic) with deep learning algorithms, artificial intelligence (AI) models can identify critical SE-TF regulatory nodes and dynamic binding patterns (Hamamoto et al., 2023). For instance, advanced frameworks like diffusion models (e.g., DeepConformer) enable rapid prediction of TF conformational changes, revealing cryptic binding pockets on SE-associated proteins that drive oncogenic transcriptional programs (Du et al., 2024). This approach accelerates target discovery 10-fold compared to traditional methods, while machine learning-powered virtual screening optimizes compound libraries to block pathological SE-TF networks. However, challenges remain in validating AI-predicted interactions through wet-lab experiments and addressing tumor heterogeneity in clinical translation.

Recent advances in liver cancer therapy have highlighted the potential of targeting SEs—critical regulatory elements driving oncogene expression—to reshape the tumor microenvironment and amplify immunotherapy efficacy. Our proposed "SE Editing + Immunotherapy" combination strategy integrates CRISPR-based SE modulation [similar to multiple myeloma (Li J. et al., 2024)] with immune checkpoint inhibitors (ICIs) in breast cancer, colorectal cancer, and other cancers (Jia et al., 2024), demonstrating remarkable clinical benefits in advanced HCC. By selectively disrupting oncogenic SEs (e.g., those activating HSAL3 or miR-9), this approach suppresses tumor-promoting pathways (e.g., NOTCH,  $\beta$ -catenin) while reducing immune evasion signals (Yuan et al., 2023). SE editing enhances tumor immunogenicity by upregulating antigen presentation and downregulating

Drug name	Target	Mechanism profile	Clinical trial phase	Clinical translation	Therapeutic implications	NCT number
OTX015	BRD4	BET inhibitor disrupting SE- mediated oncogene transcription (e.g., MYC)	Phase I/II	Demonstrated tumor regression in HCC models; synergizes with chemotherapy (e.g., sorafenib)	Targets SE-driven transcriptional addiction; potential for MYC- driven HCC subtypes	NCT02259114
THZ1	CDK7	Covalent CDK7 inhibitor blocking SE-driven transcriptional addiction	Phase I	Suppressed SE-associated oncogenes (e.g., MYC) in preclinical models; limited toxicity in early trials	Addresses transcriptional "bursts" driven by SEs; combats resistance to BET inhibitors	NCT03134638
BAY 1143572	CDK9	Selective CDK9 inhibitor targeting SE-driven transcriptional elongation	Phase I	Anti-proliferative effects in HCC cell lines; ongoing trials in solid tumors	Targets transcriptional elongation dependency; may overcome adaptive SE rewiring	NCT02345382
Dinaciclib	CDK9/ CDK12	Pan-CDK inhibitor suppressing SE-associated transcriptional dependencies	Phase II	Reduced tumor burden in advanced HCC; efficacy correlates with MYC expression levels	Broad CDK targeting may mitigate compensatory SE activation; higher toxicity risk	NCT01676753

TABLE	3	SE-related	potential	drugs	in	clinical	trials	for	liver	cancer.
-------	---	------------	-----------	-------	----	----------	--------	-----	-------	---------

immunosuppressive factors (e.g., PD-L1), priming HCC for PD-1/ PD-L1 inhibitors like nivolumab or pembrolizumab (Shi et al., 2023). Ongoing trials are exploring SE modulation with dual ICIs (e.g., anti-PD-1 + anti-CTLA-4) to further boost durable responses.

The development of targeted delivery systems, such as nanoparticles or antibody-drug conjugates (ADCs), could minimize systemic toxicity and improve drug localization to liver cancer cells. Combining small molecule inhibitors with biological agents, such as monoclonal antibodies or CAR-T cells (Chakraborty and Sarkar, 2022; Peng et al., 2024), may offer synergistic benefits while reducing adverse effects. Thus, these therapeutic strategies aim to disrupt the specific SE-driven transcriptional programs that sustain cancer cell survival and progression, offering the potential for personalized treatment options that are more effective and less toxic than traditional chemotherapy.

To address these issues, in 2019, 102,373 *cis*-regulatory elements were identified in the pig liver, mainly including enhancers and SEs, highlighting 26 core transcription regulatory factors in the pig liver as well (Luan et al., 2019). Intriguingly, Pan's group has established the SE repertoire in porcine liver (Zhang et al., 2023). In this database, the expression profiles were mainly sourced from fetal and 70-day-old pigs, demonstrating that many SE-associated genes were positively related to hepatic metabolisms and detoxification pathways. Their results illustrated the disparity and linkage between promoter and enhancer markers. Considering the similarity between porcine and human beings, more SE-related mechanisms in liver cancer can be identified, providing more robust evidence for targeted therapy, especially for the unknown side effects.

# 5.6 Innovative perspectives and emerging frontiers in super enhancer-driven liver cancer research

While the current review synthesizes established mechanisms linking SEs to HCC progression, advancing the field requires a paradigm shift toward innovative frameworks and unexplored intersections. Recent breakthroughs in single-cell epigenomics (Gu et al., 2025; Lewis et al., 2024; Liu S. et al., 2024), spatial transcriptomics (Mahat et al., 2024; Manicardi et al., 2023), and CRISPR-based functional screens (Xi et al., 2025; Kasprzyk et al., 2024) have unveiled unprecedented heterogeneity in SE landscapes across variable malignancies. For instance, emerging studies highlight dynamic SE rewiring during therapy resistance, where subpopulations of tumor cells exploit SE plasticity to activate compensatory oncogenic pathways. Integrating these findings, we propose that SE-driven transcriptional addiction in HCC could be leveraged to design "epigenetic vulnerability" screens. By mapping SE-associated dependencies in treatment-naïve versus resistant tumors, researchers may identify context-specific targets, such as SE-regulated non-coding RNAs or metabolic gatekeepers, that circumvent traditional oncogene-centric limitations. Furthermore, leveraging AI to predict SE connectivity networks-incorporating multi-omics data from circulating tumor DNA, liquid biopsies, and patient-derived organoids-could enable real-time monitoring of SE activity as a biomarker for therapeutic response (Kim et al., 2025). In the future, our own study will focus on the SE-related multi-omics in HCC, integrating these innovative technologies.

A second frontier lies in decoding the interplay between SEs and the TME. SEs in stromal or immune cells (e.g., tumor-associated macrophages) may indirectly promote malignant tumor progression by regulating cytokine secretion or immune checkpoint expression (Zhang T. et al., 2022; Betancur et al., 2017). Additionally, the gutliver axis represents an untapped dimension: microbiome-derived metabolites like butyrate modulate histone acetylation patterns, potentially reshaping SE architectures in pre-malignant hepatocytes. Future studies should explore whether SE inhibition synergizes with microbiota modulation or immune checkpoint blockade to achieve durable remission. By embracing these multidimensional approaches—spanning SE-TME interactions, AI-driven biomarker discovery, and metabolite-epigenome crosstalk—the field can transition from descriptive SE mapping to mechanistically innovative, clinically transformative strategies.

### 6 Conclusion

The therapeutic potential of targeting SEs in liver cancer represents a promising frontier in oncology. By disrupting the epigenetic and transcriptional networks that drive liver cancer progression, we could reprogram the cancer cell transcriptome, thereby inhibiting tumor growth and metastasis. However, significant challenges remain, including specificity, toxicity, tumor heterogeneity, and the need for reliable biomarkers. Combination therapies targeting super-enhancer activity and other oncogenic signaling pathways could offer the most effective strategy for overcoming these challenges.

As we continue to deepen our understanding of SE biology and its role in liver cancer, developing more targeted and personalized therapies will be crucial in translating these findings into effective clinical treatments. Future research must address the complexity of SE regulation, identify biomarkers for patient stratification, and refine combination treatment approaches to ensure that liver cancer patients benefit from the full potential of epigenetic therapies.

# Author contributions

C-LL: Data curation, Formal Analysis, Project administration, Supervision, Writing – original draft, Writing – review and editing. Z-YY: Data curation, Methodology, Writing – original draft. AS: Data curation, Project administration, Writing – original draft. J-YC: Formal Analysis, Resources, Visualization, Writing – original draft. Z-SW: Formal Analysis, Methodology, Resources, Validation, Writing – original draft.

### References

Ajoolabady, A., Kaplowitz, N., Lebeaupin, C., Kroemer, G., Kaufman, R. J., Malhi, H., et al. (2023). Endoplasmic reticulum stress in liver diseases. *Hepatology*, 77, 619–639. doi:10.1002/hep.32562

Akhtar-Zaidi, B., Cowper-Sal-lari, R., Corradin, O., Saiakhova, A., Bartels, C. F., Balasubramanian, D., et al. (2012). Epigenomic enhancer profiling defines a signature of colon cancer. *Science (New York, N.Y.)* 336, 736–739. doi:10.1126/science.1217277

Antal, C. E., Oh, T. G., Aigner, S., Luo, E. C., Yee, B. A., Campos, T., et al. (2023). A super-enhancer-regulated RNA-binding protein cascade drives pancreatic cancer. *Nat. Commun.* 14, 5195. doi:10.1038/s41467-023-40798-6

Bai, J., Jiang, P., Ji, L., Lam, W. K. J., Zhou, Q., Ma, M. L., et al. (2024). Histone modifications of circulating nucleosomes are associated with changes in cell-free DNA fragmentation patterns. *Proc. Natl. Acad. Sci. U. S. A.* 121, e2404058121. doi:10.1073/ pnas.2404058121

Banerji, J., Rusconi, S., and Schaffner, W. (1981). Expression of a beta-globin gene is enhanced by remote SV40 DNA sequences. *Cell* 27, 299–308. doi:10.1016/0092-8674(81)90413-x

Barral, A., and Déjardin, J. (2023). The chromatin signatures of enhancers and their dynamic regulation. *Nucleus (Austin, Tex.)* 14, 2160551. doi:10.1080/19491034.2022. 2160551

Benichou, E., Seffou, B., Topçu, S., Renoult, O., Lenoir, V., Planchais, J., et al. (2024). The transcription factor ChREBP Orchestrates liver carcinogenesis by coordinating the PI3K/AKT signaling and cancer metabolism. *Nat. Commun.* 15, 1879. doi:10.1038/s41467-024-45548-w

Berico, P., Nogaret, M., Cigrang, M., Lallement, A., Vand-Rajabpour, F., Flores-Yanke, A., et al. (2023). Super-enhancer-driven expression of BAHCC1 promotes melanoma cell proliferation and genome stability. *Cell Rep.* 42, 113363. doi:10.1016/ j.celrep.2023.113363

Betancur, P. A., Abraham, B. J., Yiu, Y. Y., Willingham, S. B., Khameneh, F., Zarnegar, M., et al. (2017). A CD47-associated super-enhancer links pro-inflammatory signalling to CD47 upregulation in breast cancer. *Nat. Commun.* 8, 14802. doi:10.1038/ ncomms14802

Blobel, G. A., Higgs, D. R., Mitchell, J. A., Notani, D., and Young, R. A. (2021). Testing the super-enhancer concept. *Nat. Rev. Genet.* 22, 749–755. doi:10.1038/s41576-021-00398-w

Brangwynne, C. P., Eckmann, C. R., Courson, D. S., Rybarska, A., Hoege, C., Gharakhani, J., et al. (2009). Germline P granules are liquid droplets that localize by

### Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# **Generative AI statement**

The author(s) declare that no Generative AI was used in the creation of this manuscript.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

controlled dissolution/condensation. Science (New York, N.Y.) 324, 1729–1732. doi:10. 1126/science.1172046

Bulger, M., and Groudine, M. (2011). Functional and mechanistic diversity of distal transcription enhancers. Cell 144, 327–339. doi:10.1016/j.cell.2011.01.024

Cai, C., Bi, D., Bick, G., Wei, Q., Liu, H., Lu, L., et al. (2021). Hepatocyte nuclear factor HNF1A is a potential regulator in shaping the super-enhancer landscape in colorectal cancer liver metastasis. *FEBS Lett.* 595, 3056–3071. doi:10.1002/1873-3468.14219

Cai, H., Liang, J., Jiang, Y., Wang, Z., Li, H., Wang, W., et al. (2024). KLF7 regulates super-enhancer-driven IGF2BP2 overexpression to promote the progression of head and neck squamous cell carcinoma. *J. Exp. Clin. Cancer Res.* 43, 69. doi:10.1186/s13046-024-02996-y

Calderaro, J., Ziol, M., Paradis, V., and Zucman-Rossi, J. (2019). Molecular and histological correlations in liver cancer. *J. Hepatol.* 71, 616–630. doi:10.1016/j.jhep.2019. 06.001

Calo, E., and Wysocka, J. (2013). Modification of enhancer chromatin: what, how, and why? *Mol. Cell* 49, 825–837. doi:10.1016/j.molcel.2013.01.038

Cazzanelli, G., Dalle Vedove, A., Sbardellati, N., Valer, L., Caflisch, A., and Lolli, G. (2024). Enhanced cellular death in liver and breast cancer cells by dual BET/ BRPF1 inhibitors. *Protein Sci.* 33, e5191. doi:10.1002/pro.5191

Chakraborty, E., and Sarkar, D. (2022). Emerging therapies for hepatocellular carcinoma (HCC). *Cancers* 14, 2798. doi:10.3390/cancers14112798

Chang, S., Yim, S., and Park, H. (2019). The cancer driver genes IDH1/2, JARID1C/ KDM5C, and UTX/KDM6A: crosstalk between histone demethylation and hypoxic reprogramming in cancer metabolism. *Exp. Mol. Med.* 51, 1–17. doi:10.1038/s12276-019-0230-6

Chapuy, B., McKeown, M. R., Lin, C. Y., Monti, S., Roemer, M. G., Qi, J., et al. (2013). Discovery and characterization of super-enhancer-associated dependencies in diffuse large B cell lymphoma. *Cancer Cell* 24, 777–790. doi:10.1016/j.ccr.2013.11.003

Chen, C., Wang, Z., Ding, Y., and Qin, Y. (2023b). Tumor microenvironmentmediated immune evasion in hepatocellular carcinoma. *Front. Immunol.* 14, 1133308. doi:10.3389/fimmu.2023.1133308

Chen, D., Cao, Y., Tang, H., Zang, L., Yao, N., Zhu, Y., et al. (2023c). Comprehensive machine learning-generated classifier identifies pro-metastatic characteristics and predicts individual treatment in pancreatic cancer: a multicenter cohort study based on super-enhancer profiling. *Theranostics* 13, 3290–3309. doi:10.7150/thno.84978

Chen, D., Zhao, Z., Chen, L., Li, Q., Zou, J., and Liu, S. (2021). PPM1G promotes the progression of hepatocellular carcinoma via phosphorylation regulation of alternative splicing protein SRSF3. *Cell Death Dis.* 12, 722. doi:10.1038/s41419-021-04013-y

Chen, J., Xu, Z., Huang, H., Tang, Y., Shan, H., and Xiao, F. (2023a). SETD1A drives stemness by reprogramming the epigenetic landscape in hepatocellular carcinoma stem cells. *JCI Insight* 8, e168375. doi:10.1172/jci.insight.168375

Chen, M., Fu, R., Chen, Y., Li, L., and Wang, S. W. (2025). High-resolution, noninvasive single-cell lineage tracing in mice and humans based on DNA methylation epimutations. *Nat. Methods* 22, 488–498. doi:10.1038/s41592-024-02567-1

Chen, M., Wei, L., Law, C. T., Tsang, F. H., Shen, J., Cheng, C. L., et al. (2018). RNA N6-methyladenosine methyltransferase-like 3 promotes liver cancer progression through YTHDF2-dependent posttranscriptional silencing of SOCS2.*Hepatology* 67, 2254–2270. doi:10.1002/hep.29683

Chen, Y., Yao, B., Zhu, Z., Yi, Y., Lin, X., Zhang, Z., et al. (2004). A constitutive superenhancer: homologous region 3 of *Bombyx mori* nucleopolyhedrovirus. *Biochem. Biophysical Res. Commun.* 318, 1039–1044. doi:10.1016/j.bbrc.2004.04.136

Choi, S., Kim, B. K., Yon, D. K., Lee, S. W., Lee, H. G., Chang, H. H., et al. (2023). Global burden of primary liver cancer and its association with underlying aetiologies, sociodemographic status, and sex differences from 1990-2019: a DALY-based analysis of the Global Burden of Disease 2019 study. *Clin. Mol. Hepatol.* 29, 433–452. doi:10. 3350/cmh.2022.0316

Christensen, C. L., Kwiatkowski, N., Abraham, B. J., Carretero, J., Al-Shahrour, F., Zhang, T., et al. (2014). Targeting transcriptional addictions in small cell lung cancer with a covalent CDK7 inhibitor. *Cancer Cell* 26, 909–922. doi:10.1016/j.ccell.2014. 10.019

Claveria-Cabello, A., Colyn, L., Arechederra, M., Urman, J. M., Berasain, C., Avila, M. A., et al. (2020). Epigenetics in liver fibrosis: could HDACs be a therapeutic target? *Cells* 9, 2321. doi:10.3390/cells9102321

Cole, J. J., Robertson, N. A., Rather, M. I., Thomson, J. P., McBryan, T., Sproul, D., et al. (2017). Diverse interventions that extend mouse lifespan suppress shared ageassociated epigenetic changes at critical gene regulatory regions. *Genome Biol.* 18, 58. doi:10.1186/s13059-017-1185-3

Craig, A. J., Silveira, M. A. D., Ma, L., Revsine, M., Wang, L., Heinrich, S., et al. (2023). Genome-wide profiling of transcription factor activity in primary liver cancer using single-cell ATAC sequencing. *Cell Rep.* 42, 113446. doi:10.1016/j.celrep.2023.113446

Dai, W., Wu, J., Peng, X., Hou, W., Huang, H., Cheng, Q., et al. (2022). CDK12 orchestrates super-enhancer-associated CCDC137 transcription to direct hepatic metastasis in colorectal cancer. *Clin. Transl. Med.* 12, e1087. doi:10.1002/ ctm2.1087

Danpanichkul, P., Aboona, M. B., Sukphutanan, B., Kongarin, S., Duangsonk, K., Ng, C. H., et al. (2024). Incidence of liver cancer in young adults according to the Global Burden of Disease database 2019. *Hepatology* 80, 828–843. doi:10.1097/hep. 00000000000872

Du, D., Liu, C., Qin, M., Zhang, X., Xi, T., Yuan, S., et al. (2022). Metabolic dysregulation and emerging therapeutical targets for hepatocellular carcinoma. *Acta Pharm. Sin. B* 12, 558–580. doi:10.1016/j.apsb.2021.09.019

Du, M., Stitzinger, S. H., Spille, J. H., Cho, W. K., Lee, C., Hijaz, M., et al. (2024). Direct observation of a condensate effect on super-enhancer controlled gene bursting. *Cell* 187, 2595–2598. doi:10.1016/j.cell.2024.04.001

Dubois, V., Gheeraert, C., Vankrunkelsven, W., Dubois-Chevalier, J., Dehondt, H., Bobowski-Gerard, M., et al. (2020). Endoplasmic reticulum stress actively suppresses hepatic molecular identity in damaged liver. *Mol. Syst. Biol.* 16, e9156. doi:10.15252/ msb.20199156

Durbin, A. D., Zimmerman, M. W., Dharia, N. V., Abraham, B. J., Iniguez, A. B., Weichert-Leahey, N., et al. (2018). Selective gene dependencies in MYCN-amplified neuroblastoma include the core transcriptional regulatory circuitry. *Nat. Genet.* 50, 1240–1246. doi:10.1038/s41588-018-0191-z

Estes, C., Razavi, H., Loomba, R., Younossi, Z., and Sanyal, A. J. (2018). Modeling the epidemic of nonalcoholic fatty liver disease demonstrates an exponential increase in burden of disease. *Hepatology Baltim. Md* 67, 123–133. doi:10.1002/hep.29466

Fabregat, I. (2009). Dysregulation of apoptosis in hepatocellular carcinoma cells. World J. Gastroenterol. 15, 513–520. doi:10.3748/wjg.15.513

Fiskus, W., Sharma, S., Qi, J., Valenta, J. A., Schaub, L. J., Shah, B., et al. (2014). Highly active combination of BRD4 antagonist and histone deacetylase inhibitor against human acute myelogenous leukemia cells. *Mol. Cancer Ther.* 13, 1142–1154. doi:10. 1158/1535-7163.MCT-13-0770

Font-Tello, A., Kesten, N., Xie, Y., Taing, L., Varešlija, D., Young, L. S., et al. (2020). FiTAc-seq: fixed-tissue ChIP-seq for H3K27ac profiling and super-enhancer analysis of FFPE tissues. *Nat. Protoc.* 15, 2503–2518. doi:10.1038/s41596-020-0340-6

Gu, Y., Ding, C., Yu, T., Liu, B., Tang, W., Wang, Z., et al. (2024). SIRT7 promotes Hippo/YAP activation and cancer cell proliferation in hepatocellular carcinoma via suppressing MST1. *Cancer Sci.* 115, 1209–1223. doi:10.1111/cas.16091

Gu, Y., Wang, H., Xue, W., Zhu, L., Fu, C., Zhang, W., et al. (2025). Endoplasmic reticulum stress related super-enhancers suppress cuproptosis via glycolysis

reprogramming in lung adenocarcinoma. Cell Death Dis. 16, 316. doi:10.1038/s41419-025-07613-0

Guo, S., Yu, Y., Bu, Y., Ren, J., Zhang, L., Ma, X., et al. (2024). Pan-cancer analysis reveals the prognostic and immunomodulatory potential of super-enhancer-induced ANGPT2 and experimental validation in colorectal cancer. *Clin. Translat. Oncol.* 

Guo, W., Wang, X., Lu, B., Yu, J., Xu, M., Huang, R., et al. (2023). Super-enhancerdriven MLX mediates redox balance maintenance via SLC7A11 in osteosarcoma. *Cell Death Dis.* 14, 439. doi:10.1038/s41419-023-05966-y

Hamamoto, R., Takasawa, K., Shinkai, N., Machino, H., Kouno, N., Asada, K., et al. (2023). Analysis of super-enhancer using machine learning and its application to medical biology. *Briefings Bioinforma*. 24, bbad107. doi:10.1093/bib/bbad107

Hao, L., Li, S., Deng, J., Li, N., Yu, F., Jiang, Z., et al. (2023). The current status and future of PD-L1 in liver cancer. *Front. Immunol.* 14, 1323581. doi:10.3389/fimmu.2023. 1323581

He, Q., Hu, J., Huang, H., Wu, T., Li, W., Ramakrishnan, S., et al. (2024). FOSL1 is a key regulator of a super-enhancer driving TCOF1 expression in triple-negative breast cancer. *Epigenetics Chromatin* 17, 34. doi:10.1186/s13072-024-00559-1

He, S., and Tang, S. (2020). WNT/β-catenin signaling in the development of liver cancers. *Biomed. Pharmacother.* 132, 110851. doi:10.1016/j.biopha.2020.110851

Heyn, H., Vidal, E., Ferreira, H. J., Vizoso, M., Sayols, S., Gomez, A., et al. (2016). Epigenomic analysis detects aberrant super-enhancer DNA methylation in human cancer. *Genome Biol.* 17, 11. doi:10.1186/s13059-016-0879-2

Hnisz, D., Abraham, B. J., Lee, T. I., Lau, A., Saint-André, V., Sigova, A. A., et al. (2013). Super-enhancers in the control of cell identity and disease. *Cell* 155, 934–947. doi:10.1016/j.cell.2013.09.053

Hnisz, D., Schuijers, J., Lin, C. Y., Weintraub, A. S., Abraham, B. J., Lee, T. I., et al. (2015). Convergence of developmental and oncogenic signaling pathways at transcriptional super-enhancers. *Mol. Cell* 58, 362–370. doi:10.1016/j.molcel.2015. 02.014

Hnisz, D., Shrinivas, K., Young, R. A., Chakraborty, A. K., and Sharp, P. A. (2017). A phase separation model for transcriptional control. *Cell* 169, 13–23. doi:10.1016/j.cell. 2017.02.007

Hu, Y., Xu, R., Feng, J., Zhang, Q., Zhang, L., Li, Y., et al. (2024). Identification of potential pathogenic hepatic super-enhancers regulatory network in high-fat diet induced hyperlipidemia. *J. Nutr. Biochem.* 126, 109584. doi:10.1016/j.jnutbio.2024. 109584

Hu, Y., Yang, Q., Cai, S., Wang, W., and Fu, S. (2023). The integrative analysis based on super-enhancer related genes for predicting different subtypes and prognosis of patient with lower-grade glioma. *Front. Genet.* 14, 1085584. doi:10.3389/fgene.2023. 1085584

Huang, D. Q., Singal, A. G., Kanwal, F., Lampertico, P., Buti, M., Sirlin, C. B., et al. (2023). Hepatocellular carcinoma surveillance - utilization, barriers and the impact of changing aetiology. *Nat. Rev. Gastroenterol. Hepatol.* 20, 797–809. doi:10.1038/s41575-023-00818-8

Huang, M., Dong, W., Xie, R., Wu, J., Su, Q., Li, W., et al. (2022). HSF1 facilitates the multistep process of lymphatic metastasis in bladder cancer via a novel PRMT5-WDR5-dependent transcriptional program. *Cancer Commun. Lond. Engl.* 42, 447–470. doi:10. 1002/cac2.12284

Hunkeler, M., Hagmann, A., Stuttfeld, E., Chami, M., Guri, Y., Stahlberg, H., et al. (2018). Structural basis for regulation of human acetyl-CoA carboxylase. *Nature* 558, 470–474. doi:10.1038/s41586-018-0201-4

Hwang, S. Y., Danpanichkul, P., Agopian, V., Mehta, N., Parikh, N. D., Abou-Alfa, G. K., et al. (2024). Hepatocellular carcinoma: updates on epidemiology, surveillance, diagnosis and treatment. *Clin. Mol. Hepatol.* 31, S228–S254. doi:10.3350/cmh.2024. 0824

Hyman, A. A., Weber, C. A., and Jülicher, F. (2014). Liquid-liquid phase separation in biology. *Annu. Rev. Cell Dev. Biol.* 30, 39–58. doi:10.1146/annurev-cellbio-100913-013325

Jia, D., Wang, Q., Qi, Y., Jiang, Y., He, J., Lin, Y., et al. (2024). Microbial metabolite enhances immunotherapy efficacy by modulating T cell stemness in pan-cancer. *Cell* 187, 1651–1665.e21. doi:10.1016/j.cell.2024.02.022

Jiang, Y., Qian, F., Bai, X., Liu, Y., Wang, Q., Ai, B., et al. (2019). SEdb: a comprehensive human super-enhancer database. *Nucleic Acids Res.* 47, D235-d243. doi:10.1093/nar/gky1025

Jin, Y., Xia, Y., Du, H., Xiang, T., Lan, B., Wei, S., et al. (2023). Super-enhancerassociated EEPD1 facilitates EMT-mediated metastasis by regulating the PI3K/AKT/ mTOR pathway in gastric cancer. *Biochem. Biophysical Res. Commun.* 689, 149188. doi:10.1016/j.bbrc.2023.149188

Kagey, M. H., Newman, J. J., Bilodeau, S., Zhan, Y., Orlando, D. A., van Berkum, N. L., et al. (2010). Mediator and cohesin connect gene expression and chromatin architecture. *Nature* 467, 430–435. doi:10.1038/nature09380

Kasprzyk, M. E., Sura, W., Podralska, M., Kazimierska, M., Seitz, A., Łosiewski, W., et al. (2024). Core regions in immunoglobulin heavy chain enhancers essential for survival of non-Hodgkin lymphoma cells are identified by a CRISPR interference screen. *Haematologica* 109, 4007–4020. doi:10.3324/haematol.2023.284672

Kawaguchi, K., and Kaneko, S. (2021). Notch signaling and liver cancer. Adv. Exp. Med. Biol. 1287, 69-80. doi:10.1007/978-3-030-55031-8\_6

Kim, H. K., Cheong, H., Kim, M. Y., and Jin, H. E. (2025). Therapeutic targeting in ovarian cancer: nano-enhanced CRISPR/Cas9 gene editing and drug combination therapy. *Int. J. Nanomedicine* 20, 3907–3931. doi:10.2147/IJN.S507688

Kumari, P., Tarighi, S., Fuchshuber, E., Li, L., Fernández-Duran, I., Wang, M., et al. (2024). SIRT7 promotes lung cancer progression by destabilizing the tumor suppressor ARF. *Proc. Natl. Acad. Sci. U. S. A.* 121, e2409269121. doi:10.1073/pnas.2409269121

Lakhia, R., Mishra, A., Biggers, L., Malladi, V., Cobo-Stark, P., Hajarnis, S., et al. (2023). Enhancer and super-enhancer landscape in polycystic kidney disease. *Kidney Int.* 103, 87–99. doi:10.1016/j.kint.2022.08.039

Larson, E. D., Komori, H., Gibson, T. J., Ostgaard, C. M., Hamm, D. C., Schnell, J. M., et al. (2021). Cell-type-specific chromatin occupancy by the pioneer factor Zelda drives key developmental transitions in Drosophila. *Nat. Commun.* 12, 7153. doi:10.1038/ s41467-021-27506-y

Lawrenson, K., Fonseca, M. A. S., Liu, A. Y., Segato Dezem, F., Lee, J. M., Lin, X., et al. (2019). A study of high-grade serous ovarian cancer origins implicates the SOX18 transcription factor in tumor development. *Cell Rep.* 29, 3726–3735.e4. doi:10.1016/j.celrep.2019.10.122

Lee, T. I., and Young, R. A. (2013). Transcriptional regulation and its misregulation in disease. *Cell* 152, 1237–1251. doi:10.1016/j.cell.2013.02.014

Levine, M., Cattoglio, C., and Tjian, R. (2014). Looping back to leap forward: transcription enters a new era. *Cell* 157, 13–25. doi:10.1016/j.cell.2014.02.009

Lewis, M. W., King, C. M., Wisniewska, K., Regner, M. J., Coffey, A., Kelly, M. R., et al. (2024). CRISPR screening of transcribed super-enhancers identifies drivers of triplenegative breast cancer progression. *Cancer Res.* 84, 3684–3700. doi:10.1158/0008-5472. CAN-23-3995

Li, J., Wang, J., Wang, Y., Zhao, X., and Su, T. (2023). E2F1 combined with LINC01004 super-enhancer to promote hepatocellular carcinoma cell proliferation and metastasis. *Clin. Epigenetics* 15, 17. doi:10.1186/s13148-023-01428-6

Li, J., Wang, Y., Wang, Z., Wei, Y., Diao, P., Wu, Y., et al. (2024c). Super-enhancer driven LIF/LIFR-STAT3-SOX2 regulatory feedback loop promotes cancer stemness in head and neck squamous cell carcinoma. *Adv. Sci. (Weinheim, Baden-Wurttemberg, Ger.)* 11, e2404476. doi:10.1002/advs.202404476

Li, Q., Han, J., Yang, Y., and Chen, Y. (2022). PD-1/PD-L1 checkpoint inhibitors in advanced hepatocellular carcinoma immunotherapy. *Front. Immunol.* 13, 1070961. doi:10.3389/fimmu.2022.1070961

Li, X., Hu, Z., Shi, Q., Qiu, W., Liu, Y., Liu, Y., et al. (2024b). Elevated choline drives KLF5-dominated transcriptional reprogramming to facilitate liver cancer progression. *Oncogene* 43, 3121–3136. doi:10.1038/s41388-024-03150-w

Li, X., Wu, X. Q., Xu, T., Li, X. F., Yang, Y., Li, W. X., et al. (2016). Role of histone deacetylases(HDACs) in progression and reversal of liver fibrosis. *Toxicol. Appl. Pharmacol.* 306, 58–68. doi:10.1016/j.taap.2016.07.003

Li, X., Zheng, C., Liu, Y., Sun, H., Qian, Y., and Fan, H. (2024a). Co-overexpression of BRD4 and CDK7 promotes cell proliferation and predicts poor prognosis in HCC. *Heliyon* 10, e24389. doi:10.1016/j.heliyon.2024.e24389

Liang, W., Shi, C., Hong, W., Li, P., Zhou, X., Fu, W., et al. (2021). Super-enhancerdriven lncRNA-DAW promotes liver cancer cell proliferation through activation of Wnt/ $\beta$ -catenin pathway. *Mol. Ther. Nucleic Acids* 26, 1351–1363. doi:10.1016/j.omtn. 2021.10.028

Lin, C. H., Kuo, J. C., Li, D., Koenig, A. B., Pan, A., Yan, P., et al. (2022). AZD5153, a bivalent BRD4 inhibitor, suppresses hepatocarcinogenesis by altering BRD4 chromosomal landscape and modulating the transcriptome of HCC cells. *Front. Cell Dev. Biol.* 10, 853652. doi:10.3389/fcell.2022.853652

Lin, J., Rao, D., Zhang, M., and Gao, Q. (2024). Metabolic reprogramming in the tumor microenvironment of liver cancer. *J. Hematol. Oncol.* 17, 6. doi:10.1186/s13045-024-01527-8

Liu, B., Dou, J., and Cao, J. (2024c). Nuclear respiratory factor 1 regulates super enhancer-controlled SPIDR to protect hepatocellular carcinoma cells from oxidative stress. *BMC Gastroenterol.* 24, 97. doi:10.1186/s12876-024-03183-1

Liu, F., Liao, Z., and Zhang, Z. (2023). MYC in liver cancer: mechanisms and targeted therapy opportunities. *Oncogene* 42, 3303–3318. doi:10.1038/s41388-023-02861-w

Liu, K., Du, Y., Li, H., and Lin, X. (2022b). Identification of super-enhancer-associated transcription factors regulating glucose metabolism in poorly differentiated thyroid carcinoma. *Genet. Mol. Biol.* 45, e20210370. doi:10.1590/1678-4685-GMB-2021-0370

Liu, M., Cao, S., He, L., Gao, J., Arab, J. P., Cui, H., et al. (2021). Super enhancer regulation of cytokine-induced chemokine production in alcoholic hepatitis. *Nat. Commun.* 12, 4560. doi:10.1038/s41467-021-24843-w

Liu, S., Wang, Z., Hu, L., Ye, C., Zhang, X., Zhu, Z., et al. (2024d). Pan-cancer analysis of super-enhancer-induced LINC00862 and validation as a SIRT1-promoting factor in cervical cancer and gastric cancer. *Transl. Oncol.* 45, 101982. doi:10.1016/j.tranon.2024. 101982

Liu, W., Tang, J., Gao, W., Sun, J., Liu, G., and Zhou, J. (2024a). PPP2R1B abolishes colorectal cancer liver metastasis and sensitizes Oxaliplatin by inhibiting MAPK/ERK signaling pathway. *Cancer Cell Int.* 24, 90. doi:10.1186/s12935-024-03273-w

Liu, Y., Shi, Q., Su, Y., Chen, Z., and He, X. (2024b). Heat shock transcription factor 1 facilitates liver cancer progression by driving super-enhancer-mediated transcription of MYCN. *Cancer Med.* 13, e70157. doi:10.1002/cam4.70157

Liu, Z., Ye, Y., Liu, Y., Liu, Y., Chen, H., Shen, M., et al. (2022a). RNA helicase DHX37 facilitates liver cancer progression by cooperating with PLRG1 to drive superenhancer-mediated transcription of cyclin D1. *Cancer Res.* 82, 1937–1952. doi:10.1158/0008-5472.CAN-21-3038

Llovet, J. M., Kelley, R. K., Villanueva, A., Singal, A. G., Pikarsky, E., Roayaie, S., et al. (2021). Hepatocellular carcinoma. *Nat. Rev. Dis. Prim.* 7, 6. doi:10.1038/s41572-020-00240-3

Lovén, J., Hoke, H. A., Lin, C. Y., Lau, A., Orlando, D. A., Vakoc, C. R., et al. (2013). Selective inhibition of tumor oncogenes by disruption of super-enhancers. *Cell* 153, 320–334. doi:10.1016/j.cell.2013.03.036

Lu, J., Luo, Y., Rao, D., Wang, T., Lei, Z., Chen, X., et al. (2024a). Myeloid-derived suppressor cells in cancer: therapeutic targets to overcome tumor immune evasion. *Exp. Hematol. Oncol.* 13, 39. doi:10.1186/s40164-024-00505-7

Lu, X., Li, L., Lin, J., Wu, X., Li, W., Tan, C., et al. (2024b). PAARH promotes M2 macrophage polarization and immune evasion of liver cancer cells through VEGF protein. *Int. J. Biol. Macromol.* 281, 136580. doi:10.1016/j.ijbiomac.2024.136580

Luan, Y., Zhang, L., Hu, M., Xu, Y., Hou, Y., Li, X., et al. (2019). Identification and conservation analysis of cis-regulatory elements in pig liver. *Genes* 10, 348. doi:10.3390/genes10050348

Mack, S. C., Pajtler, K. W., Chavez, L., Okonechnikov, K., Bertrand, K. C., Wang, X., et al. (2018). Therapeutic targeting of ependymoma as informed by oncogenic enhancer profiling. *Nature* 553, 101–105. doi:10.1038/nature25169

Maezawa, S., Sakashita, A., Yukawa, M., Chen, X., Takahashi, K., Alavattam, K. G., et al. (2020). Super-enhancer switching drives a burst in gene expression at the mitosisto-meiosis transition. *Nat. Struct. Mol. Biol.* 27, 978–988. doi:10.1038/s41594-020-0488-3

Mahat, D. B., Tippens, N. D., Martin-Rufino, J. D., Waterton, S. K., Fu, J., Blatt, S. E., et al. (2024). Single-cell nascent RNA sequencing unveils coordinated global transcription. *Nature* 631, 216–223. doi:10.1038/s41586-024-07517-7

Manicardi, V., Gugnoni, M., Sauta, E., Donati, B., Vitale, E., Torricelli, F., et al. (2023). *Ex vivo* mapping of enhancer networks that define the transcriptional program driving melanoma metastasis. *Mol. Oncol.* 17, 2728–2742. doi:10.1002/1878-0261.13485

Mansour, M. R., Abraham, B. J., Anders, L., Berezovskaya, A., Gutierrez, A., Durbin, A. D., et al. (2014). Oncogene regulation. An oncogenic super-enhancer formed through somatic mutation of a noncoding intergenic element. *Science (New York, N.Y.)* 346, 1373–1377. doi:10.1126/science.1259037

McArthur, E., and Capra, J. A. (2021). Topologically associating domain boundaries that are stable across diverse cell types are evolutionarily constrained and enriched for heritability. *Am. J. Hum. Genet.* 108, 269–283. doi:10.1016/j.ajhg.2021.01.001

Meng, J., Han, J., Wang, X., Wu, T., Zhang, H., An, H., et al. (2023). Twist1-YY1p300 complex promotes the malignant progression of HCC through activation of miR-9 by forming phase-separated condensates at super-enhancers and relieved by metformin. *Pharmacol. Res.* 188, 106661. doi:10.1016/j.phrs.2023.106661

Naghavi, M., Ong, K. L., Aali, A., Ababneh, H. S., Abate, Y. H., Abbafati, C., et al. (2024). Global burden of 288 causes of death and life expectancy decomposition in 204 countries and territories and 811 subnational locations, 1990–2021: a systematic analysis for the Global Burden of Disease Study 2021. *Lancet (London, Engl.)* 403, 2100–2132. doi:10.1016/s0140-6736(24)00367-2

Nakamura, K., Hernández, G., Sharma, G. G., Wada, Y., Banwait, J. K., González, N., et al. (2022). A liquid biopsy signature for the detection of patients with early-onset colorectal cancer. *Gastroenterology* 163, 1242–1251.e2. doi:10.1053/j.gastro.2022.06.089

Nguyen, T. T. T., Zhang, Y., Shang, E., Shu, C., Torrini, C., Zhao, J., et al. (2020). HDAC inhibitors elicit metabolic reprogramming by targeting super-enhancers in glioblastoma models. J. Clin. Investigat. 130, 3699–3716. doi:10.1172/JCI129049

Niu, Y., Lin, Z., Wan, A., Sun, L., Yan, S., Liang, H., et al. (2021). Loss-of-Function genetic screening identifies aldolase A as an essential driver for liver cancer cell growth under hypoxia. *Hepatology* 74, 1461–1479. doi:10.1002/hep.31846

Ottema, S., Mulet-Lazaro, R., Erpelinck-Verschueren, C., van Herk, S., Havermans, M., Arricibita Varea, A., et al. (2021). The leukemic oncogene EVI1 hijacks a MYC super-enhancer by CTCF-facilitated loops. *Nat. Commun.* 12, 5679. doi:10.1038/s41467-021-25862-3

Parker, S. C., Stitzel, M. L., Taylor, D. L., Orozco, J. M., Erdos, M. R., Akiyama, J. A., et al. (2013). Chromatin stretch enhancer states drive cell-specific gene regulation and harbor human disease risk variants. *Proc. Natl. Acad. Sci. U. S. A.* 110, 17921–17926. doi:10.1073/pnas.1317023110

Pasquali, L., Gaulton, K. J., Rodríguez-Seguí, S. A., Mularoni, L., Miguel-Escalada, I., Akerman, İ., et al. (2014). Pancreatic islet enhancer clusters enriched in type 2 diabetes risk-associated variants. *Nat. Genet.* 46, 136–143. doi:10.1038/ng.2870

Pei, R., Zhao, L., Ding, Y., Su, Z., Li, D., Zhu, S., et al. (2023). JMJD6-BRD4 complex stimulates lncRNA HOTAIR transcription by binding to the promoter region of HOTAIR and induces radioresistance in liver cancer stem cells. *J. Transl. Med.* 21, 752. doi:10.1186/s12967-023-04394-y

Peng, J., Li, J., Huang, J., Xu, P., Huang, H., Liu, Y., et al. (2019b). p300/CBP inhibitor A-485 alleviates acute liver injury by regulating macrophage activation and polarization. *Theranostics* 9, 8344–8361. doi:10.7150/thno.30707

Peng, J. Y., Cai, D. K., Zeng, R. L., Zhang, C. Y., Li, G. C., Chen, S. F., et al. (2022). Upregulation of superenhancer-driven LncRNA FASRL by USF1 promotes *de novo* fatty acid biosynthesis to exacerbate hepatocellular carcinoma. *Adv. Sci. (Weinheim, Baden-Wurttemberg, Ger.)* 10, e2204711. doi:10.1002/advs.202204711

Peng, L., Jiang, B., Yuan, X., Qiu, Y., Peng, J., Huang, Y., et al. (2019a). Superenhancer-associated long noncoding RNA HCCL5 is activated by ZEB1 and promotes the malignancy of hepatocellular carcinoma. *Cancer Res.* 79, 572–584. doi:10.1158/ 0008-5472.CAN-18-0367

Peng, L., Sferruzza, G., Yang, L., Zhou, L., and Chen, S. (2024). CAR-T and CAR-NK as cellular cancer immunotherapy for solid tumors. *Cell. Mol. Immunol.* 21, 1089–1108. doi:10.1038/s41423-024-01207-0

Pope, E. D., 3rd, Kimbrough, E. O., Vemireddy, L. P., Surapaneni, P. K., Copland, J. A., 3rd, and Mody, K. (2019). Aberrant lipid metabolism as a therapeutic target in liver cancer. *Expert Opin. Ther. Targets* 23, 473–483. doi:10.1080/14728222.2019.1615883

Puustinen, M. C., and Sistonen, L. (2020). Molecular mechanisms of heat shock factors in cancer. *Cells* 9, 1202. doi:10.3390/cells9051202

Ramachandran, S., and Henikoff, S. (2016). Transcriptional regulators compete with nucleosomes post-replication. *Cell* 165, 580–592. doi:10.1016/j.cell.2016.02.062

Sabari, B. R., Dall'Agnese, A., Boija, A., Klein, I. A., Coffey, E. L., Shrinivas, K., et al. (2018). Coactivator condensation at super-enhancers links phase separation and gene control. *Science (New York, N.Y.)* 361, eaar3958. doi:10.1126/science.aar3958

Sang, X., Zhang, Y., Fang, F., Gao, L., Tao, Y., Li, X., et al. (2022). BRD4 inhibitor GNE-987 exerts anticancer effects by targeting super-enhancer-related gene LYL1 in acute myeloid leukemia. *J. Immunol. Res.* 2022, 7912484. doi:10.1155/2022/7912484

Schulze, K., Nault, J. C., and Villanueva, A. (2016). Genetic profiling of hepatocellular carcinoma using next-generation sequencing. *J. Hepatol.* 65, 1031–1042. doi:10.1016/j. jhep.2016.05.035

Seehawer, M., D'Artista, L., and Zender, L. (2019). The worst from both worlds: cHCC-ICC. Cancer Cell 35, 823–824. doi:10.1016/j.ccell.2019.05.008

Shi, C., Chen, L., Pi, H., Cui, H., Fan, C., Tan, F., et al. (2023). Identifying a locus in super-enhancer and its resident NFE2L1/MAFG as transcriptional factors that drive PD-L1 expression and immune evasion. *Oncogenesis* 12, 56. doi:10.1038/s41389-023-00500-3

Shin, J., He, M., Liu, Y., Paredes, S., Villanova, L., Brown, K., et al. (2013). SIRT7 represses Myc activity to suppress ER stress and prevent fatty liver disease. *Cell Rep.* 5, 654–665. doi:10.1016/j.celrep.2013.10.007

Sia, D., Villanueva, A., Friedman, S. L., and Llovet, J. M. (2017). Liver cancer cell of origin, molecular class, and effects on patient prognosis. *Gastroenterology* 152, 745–761. doi:10.1053/j.gastro.2016.11.048

Song, P., Han, R., and Yang, F. (2024). Super enhancer lncRNAs: a novel hallmark in cancer. *Cell Commun. Signal.* 22, 207. doi:10.1186/s12964-024-01599-6

Su, T., Zhang, N., Wang, T., Zeng, J., Li, W., Han, L., et al. (2023). Super enhancerregulated LncRNA LINC01089 induces alternative splicing of DIAPH3 to drive hepatocellular carcinoma metastasis. *Cancer Res.* 83, 4080–4094. doi:10.1158/0008-5472.CAN-23-0544

Sun, Y., Zhang, H., Meng, J., Guo, F., Ren, D., Wu, H., et al. (2022). S-palmitoylation of PCSK9 induces sorafenib resistance in liver cancer by activating the PI3K/AKT pathway. *Cell Rep.* 40, 111194. doi:10.1016/j.celrep.2022.111194

Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., et al. (2021). Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 71, 209–249. doi:10.3322/caac.21660

Tang, L. (2019). Liquid phase separation. Nat. methods 16, 18. doi:10.1038/s41592-018-0269-7

Tasdemir, N., Banito, A., Roe, J. S., Alonso-Curbelo, D., Camiolo, M., Tschaharganeh, D. F., et al. (2016). BRD4 connects enhancer remodeling to senescence immune surveillance. *Cancer Discov.* 6, 612–629. doi:10.1158/2159-8290.CD-16-0217

Teng, S., Li, Y. E., Yang, M., Qi, R., Huang, Y., Wang, Q., et al. (2020). Tissue-specific transcription reprogramming promotes liver metastasis of colorectal cancer. *Cell Res.* 30, 34–49. doi:10.1038/s41422-019-0259-z

Tsang, F. H., Law, C. T., Tang, T. C., Cheng, C. L., Chin, D. W., Tam, W. V., et al. (2019). Aberrant super-enhancer landscape in human hepatocellular carcinoma. *Hepatology* 69, 2502–2517. doi:10.1002/hep.30544

Tu, Y., Wu, H., Zhong, C., Liu, Y., Xiong, Z., Chen, S., et al. (2024). Pharmacological activation of STAT1-GSDME pyroptotic circuitry reinforces epigenetic immunotherapy for hepatocellular carcinoma. *Gut* 74, 613–627. doi:10.1136/gutjnl-2024-332281

Tzelepis, K., De Braekeleer, E., Aspris, D., Barbieri, I., Vijayabaskar, M. S., Liu, W. H., et al. (2018). SRPK1 maintains acute myeloid leukemia through effects on isoform usage of epigenetic regulators including BRD4. *Nat. Commun.* 9, 5378. doi:10.1038/s41467-018-07620-0

Wang, A., Yan, S., Liu, J., Chen, X., Hu, M., Du, X., et al. (2025). Endoplasmic reticulum stress-related super enhancer promotes epithelial-mesenchymal transformation in hepatocellular carcinoma through CREB5 mediated activation of TNC. *Cell Death Dis.* 16, 73. doi:10.1038/s41419-025-07356-y

Wang, M., Chen, Q., Wang, S., Xie, H., Liu, J., Huang, R., et al. (2023). Superenhancers complexes zoom in transcription in cancer. *J. Exp. Clin. Cancer Res.* 42, 183. doi:10.1186/s13046-023-02763-5

Wang, S. W., Gao, C., Zheng, Y. M., Yi, L., Lu, J. C., Huang, X. Y., et al. (2022). Current applications and future perspective of CRISPR/Cas9 gene editing in cancer. *Mol. Cancer* 21, 57. doi:10.1186/s12943-022-01518-8

Wang, W., Tang, Y. A., Xiao, Q., Lee, W. C., Cheng, B., Niu, Z., et al. (2021). Stromal induction of BRD4 phosphorylation results in chromatin remodeling and BET inhibitor resistance in colorectal cancer. *Nat. Commun.* 12, 4441. doi:10.1038/s41467-021-24687-4

Wang, W., Wang, C., Xu, H., and Gao, Y. (2020). Aldehyde dehydrogenase, liver disease and cancer. Int. J. Biol. Sci. 16, 921-934. doi:10.7150/ijbs.42300

Wang, Y., Chen, S., Xue, M., Ma, J., Yi, X., Li, X., et al. (2024). Epigenetic regulation of key gene of PCK1 by enhancer and super-enhancer in the pathogenesis of fatty liver hemorrhagic syndrome. *Anim. Biosci.* 37, 1317–1332. doi:10.5713/ab.23.0423

Watanabe, K., Yamamoto, M., Xin, B., Ooshio, T., Goto, M., Fujii, K., et al. (2019). Emergence of the dedifferentiated phenotype in hepatocyte-derived tumors in mice: roles of oncogene-induced epigenetic alterations. *Hepatol. Commun.* 3, 697–715. doi:10. 1002/hep4.1327

Wei, D., Yuan, L., Xu, X., Wu, C., Huang, Y., Zhang, L., et al. (2024). Exploring epigenetic dynamics unveils a super-enhancer-mediated NDRG1- $\beta$ -catenin axis in modulating gemcitabine resistance in pancreatic cancer. *Cancer Lett.* 605, 217284. doi:10.1016/j.canlet.2024.217284

Wei, X., Zhou, Z., Long, M., Lin, Q., Qiu, M., Chen, P., et al. (2023). A novel signature constructed by super-enhancer-related genes for the prediction of prognosis in hepatocellular carcinoma and associated with immune infiltration. *Front. Oncol.* 13, 1043203. doi:10.3389/fonc.2023.1043203

Whyte, W. A., Orlando, D. A., Hnisz, D., Abraham, B. J., Lin, C. Y., Kagey, M. H., et al. (2013). Master transcription factors and mediator establish super-enhancers at key cell identity genes. *Cell* 153, 307–319. doi:10.1016/j.cell.2013.03.035

Wong, M., Sun, Y., Xi, Z., Milazzo, G., Poulos, R. C., Bartenhagen, C., et al. (2019). JMJD6 is a tumorigenic factor and therapeutic target in neuroblastoma. *Nat. Commun.* 10, 3319. doi:10.1038/s41467-019-11132-w

Wu, F., Xu, L., Tu, Y., Cheung, O. K., Szeto, L. L., Mok, M. T., et al. (2022). Sirtuin 7 super-enhancer drives epigenomic reprogramming in hepatocarcinogenesis. *Cancer Lett.* 525, 115–130. doi:10.1016/j.canlet.2021.10.039

Wu, J., Chai, H., Li, F., Ren, Q., and Gu, Y. (2020). SETD1A augments sorafenib primary resistance via activating YAP in hepatocellular carcinoma. *Life Sci.* 260, 118406. doi:10.1016/j.lfs.2020.118406

Wu, Q., Li, P., Tao, X., Lin, N., Mao, B., and Xie, X. (2024). A novel super-enhancerrelated risk model for predicting prognosis and guiding personalized treatment in hepatocellular carcinoma. *BMC Cancer* 24, 1087. doi:10.1186/s12885-024-12874-7

Wu, Q., Tao, X., Luo, Y., Zheng, S., Lin, N., and Xie, X. (2023b). A novel superenhancer-related gene signature predicts prognosis and immune microenvironment for breast cancer. *BMC Cancer* 23, 776. doi:10.1186/s12885-023-11241-2

Wu, Y., Chen, S., Shao, Y., Su, Y., Li, Q., Wu, J., et al. (2023a). KLF5 promotes tumor progression and parp inhibitor resistance in ovarian cancer. *Adv. Sci. (Weinheim, Baden-Wurttemberg, Ger.)* 10, e2304638. doi:10.1002/advs.202304638

Xi, Y., Wang, R., Qu, M., Pan, Q., Wang, M., Ai, X., et al. (2025). Super-enhancerhijacking RBBP7 potentiates metastasis and stemness of breast cancer via recruiting NuRD complex subunit LSD1. *J. Transl. Med.* 23, 266. doi:10.1186/s12967-025-06270-3

Xu, C., Xu, Z., Zhang, Y., Evert, M., Calvisi, D. F., and Chen, X. (2022). β-Catenin signaling in hepatocellular carcinoma. *J. Clin. Investigat.* 132. doi:10.1172/jci154515

Xue, J., Ge, P., and Wu, Y. (2023). The prognosis and clinicopathological significance of histone deacetylase in hepatocellular carcinoma: a meta-analysis. *Clin. Exp. Med.* 23, 1515–1536. doi:10.1007/s10238-022-00934-w

Xue, R., Zhang, Q., Cao, Q., Kong, R., Xiang, X., Liu, H., et al. (2022). Liver tumour immune microenvironment subtypes and neutrophil heterogeneity. *Nature* 612, 141–147. doi:10.1038/s41586-022-05400-x

Yang, J. D., Altekruse, S. F., Nguyen, M. H., Gores, G. J., and Roberts, L. R. (2017). Impact of country of birth on age at the time of diagnosis of hepatocellular carcinoma in the United States. *Cancer* 123, 81–89. doi:10.1002/cncr.30246

Yang, R., Gao, W., Wang, Z., Jian, H., Peng, L., Yu, X., et al. (2024a). Polyphyllin I induced ferroptosis to suppress the progression of hepatocellular carcinoma through activation of the mitochondrial dysfunction via Nrf2/HO-1/GPX4 axis. *Phytomedicine Int. J. Phytotherapy Phytopharm.* 122, 155135. doi:10.1016/j.phymed.2023.155135

Yang, S., Zhang, D., Sun, Q., Nie, H., Zhang, Y., Wang, X., et al. (2024b). Single-cell and spatial transcriptome profiling identifies the transcription factor BHLHE40 as a driver of EMT in metastatic colorectal cancer. *Cancer Res.* 84, 2202–2217. doi:10.1158/ 0008-5472.CAN-23-3264 Yao, N., Li, W., Xu, G., Duan, N., Yu, G., and Qu, J. (2023). Choline metabolism and its implications in cancer. *Front. Oncol.* 13, 1234887. doi:10.3389/fonc.2023.1234887

Yao, Z., Song, P., and Jiao, W. (2024). Pathogenic role of super-enhancers as potential therapeutic targets in lung cancer. *Front. Pharmacol.* 15, 1383580. doi:10.3389/fphar. 2024.1383580

Yi, M., Tan, Y., Wang, L., Cai, J., Li, X., Zeng, Z., et al. (2020). TP63 links chromatin remodeling and enhancer reprogramming to epidermal differentiation and squamous cell carcinoma development. *Cell. Mol. Life Sci.* 77, 4325–4346. doi:10.1007/s00018-020-03539-2

Yoo, K. H., Hennighausen, L., and Shin, H. Y. (2019). Dissecting tissue-specific superenhancers by integrating genome-wide analyses and CRISPR/Cas9 genome editing. J. Mammary Gl. Biol. Neoplasia 24, 47–59. doi:10.1007/s10911-018-9417-z

Yoshizawa, T., Karim, M. F., Sato, Y., Senokuchi, T., Miyata, K., Fukuda, T., et al. (2014). SIRT7 controls hepatic lipid metabolism by regulating the ubiquitin-proteasome pathway. *Cell Metab.* 19, 712–721. doi:10.1016/j.cmet.2014.03.006

Yu, D., Yang, X., Lin, J., Cao, Z., Lu, C., Yang, Z., et al. (2021). Super-enhancer induced IL-20ra promotes proliferation/metastasis and immune evasion in colorectal cancer. *Front. Oncol.* 11, 724655. doi:10.3389/fonc.2021.724655

Yuan, X., Seneviratne, J. A., Du, S., Xu, Y., Chen, Y., Jin, Q., et al. (2022). Single-cell profiling of peripheral neuroblastic tumors identifies an aggressive transitional state that bridges an adrenergic-mesenchymal trajectory. *Cell Rep.* 41, 111455. doi:10.1016/j.celrep.2022.111455

Yuan, X. Q., Zhou, N., Wang, J. P., Yang, X. Z., Wang, S., Zhang, C. Y., et al. (2023). Anchoring super-enhancer-driven oncogenic lncRNAs for anti-tumor therapy in hepatocellular carcinoma. *Mol. Ther. J. Am. Soc. Gene Ther.* 31, 1756–1774. doi:10. 1016/j.ymthe.2022.11.013

Zhang, C., Wei, S., Sun, W. P., Teng, K., Dai, M. M., Wang, F. W., et al. (2020). Superenhancer-driven AJUBA is activated by TCF4 and involved in epithelial-mesenchymal transition in the progression of hepatocellular carcinoma. *Theranostics* 10, 9066–9082. doi:10.7150/thno.45349 Zhang, H., Xu, R., Li, B., Xin, Z., Ling, Z., Zhu, W., et al. (2022a). LncRNA NEAT1 controls the lineage fates of BMSCs during skeletal aging by impairing mitochondrial function and pluripotency maintenance. *Cell Death Differ.* 29, 351–365. doi:10.1038/s41418-021-00858-0

Zhang, J., Liu, S., Ye, Q., and Pan, J. (2019). Transcriptional inhibition by CDK7/ 9 inhibitor SNS-032 abrogates oncogene addiction and reduces liver metastasis in uveal melanoma. *Mol. Cancer* 18, 140. doi:10.1186/s12943-019-1070-7

Zhang, M., Hoyle, R. G., Ma, Z., Sun, B., Cai, W., Cai, H., et al. (2021). FOSL1 promotes metastasis of head and neck squamous cell carcinoma through super-enhancer-driven transcription program. *Mol. Ther. J. Am. Soc. Gene Ther.* 29, 2583–2600. doi:10.1016/j.ymthe.2021.03.024

Zhang, T., Xia, W., Song, X., Mao, Q., Huang, X., Chen, B., et al. (2022b). Superenhancer hijacking LINC01977 promotes malignancy of early-stage lung adenocarcinoma addicted to the canonical TGF- $\beta$ /SMAD3 pathway. *J. Hematol. Oncol.* 15, 114. doi:10.1186/s13045-022-01331-2

Zhang, Y., Zhang, J., Wang, C., Dai, H., Du, X., Li, Q., et al. (2023). The superenhancer repertoire in porcine liver. *J. Animal Sci.* 101, skad056. doi:10.1093/jas/ skad056

Zheng, C., Liu, M., Ge, Y., Qian, Y., and Fan, H. (2022). HBx increases chromatin accessibility and ETV4 expression to regulate dishevelled-2 and promote HCC progression. *Cell Death Dis.* 13, 116. doi:10.1038/s41419-022-04563-9

Zhu, S., Zhu, W., Zhao, K., Yu, J., Lu, W., Zhou, R., et al. (2024b). Discovery of a novel hybrid coumarin-hydroxamate conjugate targeting the HDAC1-Sp1-FOSL2 signaling axis for breast cancer therapy. *Cell Commun. Signal.* 22, 361. doi:10.1186/s12964-024-01733-4

Zhu, Y., Li, J., Li, S., Yang, Z., Qiao, Z., Gu, X., et al. (2024a). ZMAT2 condensates regulate the alternative splicing of TRIM28 to reduce cellular ROS accumulation, thereby promoting the proliferation of HCC cells. *Cell Commun. Signal.* 22, 407. doi:10. 1186/s12964-024-01790-9

# Glossary

ACACA	acetyl-CoA carboxylase 1	MED	mediator complex
ADC	antibody-drug conjugates	PDX	patient-derived xenograft
AH	Alcohol-related hepatitis	PLC	primary liver cancer
AJUBA	ajuba LIM protein	PLRG1	pleiotropic regulator 1
ALD	alcohol-associated liver disease	ROS	oxidative stress response
BRD4	bromodomain-containing protein 4	pri-miRNAs	primary microRNAs
CCND1	cyclin D1	SAM	S-adenosylmethionine
CCT	phosphocholine cytidylyltransferase	SEs	super-enhancers
CDKs	cyclin-dependent kinases	SEdb	super-enhancer database
CHC	combined hepatocellular-cholangiocarcinoma	SEMet	super-enhancer-related metastatic
СНКА	choline kinase-a	SERGs	super-enhancer-related genes
CNNs	convolutional neural networks	SETD1A	SET domain containing 1A
CRC	colorectal cancer	SIRT7	Sirtuin 7
CREB5	CAMP responsive element binding protein 5	SOD	superoxide dismutase
EMT	epithelial-mesenchymal transition	TACE	trans-arterial chemoembolization
ERS	endoplasmic reticulum stress	TAD	topologically associating domain
FA	fatty acid	TARE	trans-arterial radioembolization
FASRL	synthesis-related lncRNA	TCF4	transcription factor 4
FLHS	fatty liver haemorrhage syndrome	TCGA	Tumor Cancer Genome Atlas
GO	Gene Ontology	TEs	typical enhancers
H3K4me1	histone H3 lysine 4 mono methylation	TFs	transcription factors
H3K27ac	acetylation of histone H3 at lysine 27	TNC	tenascin-C
HATs	histone acetyltransferases	USF1	upstream stimulatory factor 1
HBV	hepatitis B virus		
HBx	Hepatitis B virus protein		
HCC	hepatocellular carcinoma		
HCV	chronic hepatitis C virus		
HDACs	histone deacetylases		
HFD	high-fat diet		
HLP	hyperlipidemia		
HM	hepatic metastatic		
HNF1A	hepatocyte nuclear factor 1-alpha		
HSF1	heat shock transcription factor 1		
ICC	intra-hepatic cholangiocarcinoma		
ICGC	International Cancer Genome Consortium		
ICIs	immune chemotherapy inhibitors		
KEGG	Kyoto Encyclopedia of Genes and Genomes		
LETFs	liver-enriched transcription factors		
LIVER-ID	liver-identity		

liquid-liquid phase separation

metabolic dysfunction-associated steatotic liver disease

Long non-coding RNAs

LLPS

LncRNAs

MASLD