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Editorial: Mobilization of hematopoietic cells from the bone marrow to the peripheral blood: Challenges and new therapeutic targets

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Editorial on the Research Topic

Mobilization of hematopoietic cells from the bone marrow to the peripheral blood: challenges and new therapeutic targets

Every year, hematopoietic cell transplantation (HCT) saves the lives of tens of thousands of patients world-wide. HCT is primarily used to treat hematologic malignancies and non-malignant bone marrow disorders, as well as blood- and immune-related genetic conditions such as hemoglobinopathies and severe combined immunodeficiency (Atsuta et al., 2024). It is also occasionally used in the treatment of certain solid tumors, such as neuroblastoma (Storb et al., 2003). Advances in gene therapy and gene-editing technologies are further expanding the potential applications of HCT (Haltalli et al., 2022). However, there remains a pressing need to refine transplant procedures and improve their safety for recipients.

In the early days of HCT, bone marrow aspirates were the sole source of hematopoietic stem cells (HSCs) for transplantation. While bone marrow harvests remain the preferred graft source for certain uses, such as pediatric transplants and non-malignant disorders, mobilized peripheral blood has become the primary graft source for approximately 90% of transplants (Atsuta et al., 2024; Arai and Klingemann, 2003; Passweg et al., 2021). In this approach, donors receive mobilizing factors that stimulate the release of HSCs from the bone marrow into the bloodstream, making collection safer and more convenient. Umbilical cord blood (UCB)-derived hematopoietic stem and progenitor cells (HSPCs) are also used as an alternative graft source for HCT (Atsuta et al., 2024; Passweg et al., 2021). However, a key challenge with UCB—and sometimes bone marrow and mobilized blood—is obtaining a sufficient number of HSPCs (To et al., 2011; Kuang et al., 2022; Ahn et al., 2024). Research is ongoing to develop

methods to increase the number of mobilized cells in donors (Luo et al., 2021; Hoggatt et al., 2018; Szade et al., 2019), reliably expand HSPCs *ex vivo*, and improve HSPC survival during and after transplantation.

Two studies in this collection address these challenges. Li et al. describe the *ex vivo* expansion of UCB-derived HSCs using the small molecule chrysin, which delays HSC differentiation and inhibits apoptosis induced by reactive oxygen species (ROS). Prchal-Murphy et al. report a strategy that combines existing drugs to enhance the survival of umbilical cord-derived HSPCs. Their work demonstrates that sequential priming of murine or human HSPCs with treprostinil and forskolin, followed by cinacalcet treatment in recipients, accelerates hematopoietic recovery and improves survival in transplanted mice.

From the recipient's perspective, effective conditioning is crucial for replacement of the hematolymphoid system and engraftment of transplanted cells. Conditioning removes the patient's endogenous HSCs from their bone marrow niches, eliminating the recipient's diseased cells and creating space for the transplanted graft (Szade et al., 2018). In cancer patients undergoing HCT, conditioning is also needed to deplete residual cancer cells and minimize the risk of recurrence before transplanting isolated cells. This is typically achieved using chemotherapy, total body irradiation (TBI), or a combination of both. The choice of regimen depends on factors such as graft used, the type of disease, remission status, and patientspecific factors, such as age, comorbidities, organ function, prior treatments, and the risk of transplant-related complications (Gyurkocza and Sandmaier, 2014). Busulfan combined with cyclophosphamide is a commonly used myeloablative conditioning regimen in allogeneic HCT. (Gyurkocza and Sandmaier, 2014; Sugita and Yanada, 2024). However, Shi et al. report on a Phase I study evaluating the safety and efficacy of various myeloablative conditioning regimens, including busulfan, cyclophosphamide, cytarabine, and a purine nucleoside analog, cladribine, for autologous HCT in acute myeloid leukemia.

Non-genotoxic therapies are being explored for transplant conditioning, such as antibodies that block interactions between HSCs and their niches (Czechowicz et al., 2007; Griffin et al., 2022). In cancer patients, however, conditioning also serves to eliminate tumor cells though studies have shown that the donor immune response against residual tumor cells-known also as graft-versustumor (GVT) or graft-versus-leukemia (GVL)-is critical in reducing recurrence risk (Gyurkocza and Sandmaier, 2014; Sugita and Yanada, 2024). Unfortunately, the immune reaction of donor cells against the recipient's tissues, termed graft-versus-host disease (GVHD), remains a serious complication of allogeneic HCT and occurs more frequently with mobilized peripheral blood grafts (Atsuta et al., 2024; Nassereddine et al., 2017). Clinical data show that the risk of chronic GVHD is higher when transplanting mobilized blood compared to bone marrow (Flowers and Martin, 2015; Zhang et al., 2023; Lacan et al., 2024). To prevent and treat GVHD. various immunosuppressive drugs, primarily corticosteroids, are used. However, many patients develop steroid-refractory GVHD, which is challenging to treat (Flinn and Gennery, 2023). New drugs are being investigated and introduced for this condition (Martini et al., 2022). Among these, ruxolitinib-a JAK inhibitor originally approved for treatment of myelofibrosis-has shown promise. In this Research Topic, Zhang et al. explore its effectiveness in treating bronchiolitis obliterans syndrome (BOS), a pulmonary manifestation of GVHD. Another study by Liu et al. focuses on optimizing dosing and pharmacokinetics of tacrolimus, a calcineurin inhibitor, in pediatric HCT recipients.

Hematopoietic cell transplantation is a curative treatment for many blood and immune diseases, though treatment approaches are constantly evolving. Especially as mobilized peripheral blood becomes the primary source of transplanted HSCs, it is crucial to enhance our understanding of GVHD mechanisms, the interplay between GVHD and GVT, and effective strategies for obtaining high-quality, transplantable HSCs. While many questions remain unanswered, we hope this Research Topic will offer valuable new insights.

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Conflict of interest

AC is a co-inventor on various cell and gene therapy patents, including those related to antibody-based conditioning. Additionally she is a past or current consultant, advisor, research partner, member of founding team, or equity holder including through intellectual property licensing to the following gene and/or cell therapy companies: Beam Therapeutics, Bluebird Bio, CSL, Decibel Therapeutics, Editas Medicine, Forty Seven Inc, Global Blood Therapeutics, Inograft Biotherapeutics, Jasper Therapeutics, Kyowa Kirin, Magenta Therapeutics, Prime Medicine, Rocket Pharmaceuticals, Spotlight Therapeutics, STRM.Bio and Teiko Bio. AS is the co-inventor of patents related to the use of cobalt porphyrins for the treatment of blood-related disorders. GG is a co-founder of CytoTRACE Biosciences, holds equity in the company, and is listed as an inventor on patents related to stem cell discovery using single-cell RNA sequencing (e.g., US20200370112A1).

Correction note

This article has been corrected with minor changes. These changes do not impact the scientific content of the article.

Generative AI statement

The author(s) declare that Generative AI was used in the creation of this manuscript. AI (ChatGPT-4) was used solely for assessing and correcting the initial draft for clarity and grammar.

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References

Ahn, W. K., Nam, H.-J., Lee, H. W., Hahn, S., Han, J. W., Lyu, C. J., et al. (2024). Poor mobilization-associated factors in autologous hematopoietic stem cell harvest. *Cancers* 16, 1821. doi:10.3390/cancers16101821

Arai, S., and Klingemann, H.-G. (2003). Hematopoietic stem cell transplantation: bone marrow vs. mobilized peripheral blood. *Arch. Med. Res.* 34, 545–553. doi:10.1016/j.arcmed.2003.07.002

Atsuta, Y., Baldomero, H., Neumann, D., Sureda, A., DeVos, J. D., Iida, M., et al. (2024). Continuous and differential improvement in worldwide access to hematopoietic cell transplantation: activity has doubled in a decade with a notable increase in unrelated and non-identical related donors. *Haematologica* 109, 3282–3294. doi:10.3324/ haematol.2024.285002

Czechowicz, A., Kraft, D., Weissman, I. L., and Bhattacharya, D. (2007). Efficient transplantation via antibody-based clearance of hematopoietic stem cell niches. *Science* 318, 1296–1299. doi:10.1126/science.1149726

Flinn, A. M., and Gennery, A. R. (2023). Recent advances in graft-versus-host disease. *Fac. Rev.* 12, 4. doi:10.12703/r/12-4

Flowers, M. E. D., and Martin, P. J. (2015). How we treat chronic graft-versus-host disease. *Blood* 125, 606–615. doi:10.1182/blood-2014-08-551994

Griffin, J. M., Healy, F. M., Dahal, L. N., Floisand, Y., and Woolley, J. F. (2022). Worked to the bone: antibody-based conditioning as the future of transplant biology. *J. Hematol. Oncol* 15, 65. doi:10.1186/s13045-022-01284-6

Gyurkocza, B., and Sandmaier, B. M. (2014). Conditioning regimens for hematopoietic cell transplantation: one size does not fit all. *Blood* 124, 344–353. doi:10.1182/blood-2014-02-514778

Haltalli, M. L. R., Wilkinson, A. C., Rodriguez-Fraticelli, A., and Porteus, M. (2022). Hematopoietic stem cell gene editing and expansion: state-of-the-art technologies and recent applications. *Exp. Hematol.* 107, 9–13. doi:10.1016/j.exphem.2021.12.399

Hoggatt, J., Singh, P., Tate, T. A., Chou, B.-K., Datari, S. R., Fukuda, S., et al. (2018). Rapid mobilization reveals a highly engraftable hematopoietic stem cell. *Cell* 172, 191–204. doi:10.1016/j.cell.2017.11.003

Kuang, P., Lin, T., Chen, X., Yang, Y., Ji, J., Dong, T., et al. (2022). Plerixafor as a preemptive or salvage therapy for healthy donors with poor mobilization of hematopoietic stem cells. *Bone Marrow Transpl.* 57, 1737–1739. doi:10.1038/s41409-022-01789-1

Lacan, C., Lambert, J., Forcade, E., Robin, M., Chevallier, P., Loron, S., et al. (2024). Bone marrow graft versus peripheral blood graft in haploidentical hematopoietic stem organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

cells transplantation: a retrospective analysis in1344 patients of SFGM-TC registry. J. Hematol. Oncol 17, 2. doi:10.1186/s13045-023-01515-4

Luo, C., Wang, L., Wu, G., Huang, X., Zhang, Y., Ma, Y., et al. (2021). Comparison of the efficacy of hematopoietic stem cell mobilization regimens: a systematic review and network meta-analysis of preclinical studies. *Stem Cell Res. Ther.* 12, 310. doi:10.1186/s13287-021-02379-6

Martini, D. J., Chen, Y.-B., and DeFilipp, Z. (2022). Recent FDA approvals in the treatment of graft-versus-host disease. *Oncologist* 27, 685–693. doi:10.1093/oncolo/ oyac076

Nassereddine, S., Rafei, H., Elbahesh, E., and Tabbara, I. (2017). Acute graft versus host disease: a comprehensive review. *Anticancer Res.* 37, 1547–1555. doi:10.21873/anticanres.11483

Passweg, J. R., Baldomero, H., Chabannon, C., Basak, G. W., de la Cámara, R., Corbacioglu, S., et al. (2021). Hematopoietic cell transplantation and cellular therapy survey of the EBMT: monitoring of activities and trends over 30 years. *Bone Marrow Transpl.* 56, 1651–1664. doi:10.1038/s41409-021-01227-8

Storb, R. F., Lucarelli, G., McSweeney, P. A., and Childs, R. W. (2003). Hematopoietic cell transplantation for benign hematological disorders and solid tumors. *Hematology* 2003, 372–397. doi:10.1182/asheducation-2003.1.372

Sugita, J., and Yanada, M. (2024). Current status of conditioning regimens in haploidentical hematopoietic cell transplantation. *Hematology* 29, 2332866. doi:10. 1080/16078454.2024.2332866

Szade, A., Szade, K., Nowak, W. N., Bukowska-Strakova, K., Muchova, L., Gońka, M., et al. (2019). Cobalt protoporphyrin IX increases endogenous G-CSF and mobilizes HSC and granulocytes to the blood. *EMBO Mol. Med.* 11, e09571. doi:10.15252/emmm. 201809571

Szade, K., Gulati, G. S., Chan, C. K. F., Kao, K. S., Miyanishi, M., Marjon, K. D., et al. (2018). Where hematopoietic stem cells live: the bone marrow niche. *Antioxid. Redox Signal* 29, 191–204. doi:10.1089/ars.2017.7419

To, L. B., Levesque, J.-P., and Herbert, K. E. (2011). How I treat patients who mobilize hematopoietic stem cells poorly. *Blood* 118, 4530–4540. doi:10.1182/blood-2011-06-318220

Zhang, Z., Zhou, X., Cheng, Z., and Hu, Y. (2023). Peripheral blood stem cell transplantation vs. bone marrow transplantation for aplastic anemia: a systematic review and meta-analysis. *Front. Med.* 10, 1289180. doi:10.3389/fmed. 2023.1289180