

OPEN ACCESS

EDITED BY Raffaele Capasso, University of Naples Federico II, Italy

REVIEWED BY

Miguel Angel Plaza, University of Zaragoza, Spain Ignacio Lizarraga, Ross University School of Veterinary Medicine, Saint Kitts and Nevis

*CORRESPONDENCE

Shen Sun,

sunshen1980@126.com
Shaoqiang Huang,
drhuangsq@163.com

[†]These authors have contributed equally to

this work

RECEIVED 03 July 2025 ACCEPTED 13 October 2025 PUBLISHED 27 October 2025

CITATION

Yang C, Hu J, Cheng Y, Sun S and Huang S (2025) Effects of different opioids on the contractile tension of isolated rat intestinal smooth muscle.

Front. Pharmacol. 16:1617936. doi: 10.3389/fphar.2025.1617936

COPYRIGHT

© 2025 Yang, Hu, Cheng, Sun and Huang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Effects of different opioids on the contractile tension of isolated rat intestinal smooth muscle

Chen Yang^{1†}, Jianying Hu^{1†}, Yan Cheng¹, Shen Sun^{2*} and Shaoqiang Huang^{2*}

¹Department of Anesthesiology, Obstetrics & Gynecology Hospital, Fudan University, Shanghai, China, ²Department of Anesthesiology, Shanghai Fifth People's Hospital, Fudan University, Shanghai, China

Backgroud: To compare how five common opioids affect rat intestinal muscle tension and frequency across concentrations, establishing an animal model to provide translational evidence for selecting opioids with the least gastrointestinal impact.

Methods: Isolated rat small intestine specimens were prepared, and six intestinal segments were selected from the same rat and placed into remifentanil, sufentanil, oxycodone, nalbuphine, butorphanol, and the control group with balanced solution. Experiments were carried out to observe and compare the changes in the contraction tension and contraction frequency of small intestinal smooth muscle during the change in concentration gradients of 10^{-9} , 10^{-8} , 10^{-7} , 10^{-6} , 10^{-5} , and 10^{-4} M. Statistical analysis obtained from 10 rats in the *in vitro* intestinal tube experiment was performed.

Results: The results revealed that when the concentration increased to 10^{-7} M, the intestinal contraction tension in the butorphanol group began to decrease significantly, and the intestinal contraction tension in the nalbuphine and oxycodone groups began to decrease significantly at concentrations of 10^{-6} M. while the sufentanil group and the remifentanil group showed a significant decrease in intestinal contractile tension at 10^{-5} M and 10^{-4} M, respectively. When the concentration of the drug increased to 10^{-4} M, the contraction frequency of the sufentanil group decreased significantly, and there was no statistical difference among concentrations of other drugs.

Conclusion: As the concentration increased, different opioids inhibited the contractile tone of rat intestinal smooth muscle *in vitro*. The intensity of inhibition was butorphanol > nalbuphine \approx oxycodone > sufentanil > remifentanil, and the Contraction frequency was almost unaffected except in the very high concentration sufentanil group.

KEYWORDS

opioids, contractile tension, contractile frequency, intestinal smooth muscle, rat

Introduction

Postoperative gastrointestinal dysfunction (PGID) is a common adverse reaction. Its pathogenesis is complex and affected by multiple factors (Mythen, 2009). Opioids used for anesthesia and analgesia are one of them (Imam et al., 2018; Müller-Lissner et al., 2017). Many animal experiments have confirmed morphine and sufentanil may dose dependently increase the contractile tension and contraction ability of isolated rat small intestine smooth muscle, while dezocine has no significant effect on intestine smooth muscle contraction (Bian et al., 2015). Clinical studies have also found that the incidence of postoperative

nausea and vomiting is lower in patients receiving intravenous oxycodone compared with sufentanil during laparoscopic surgery (Tao et al., 2019). However, the comparison between drugs has not been made in most cases, even if the mechanism of drugs is more clear, it can't play a guiding role in clinical medication selection of opioids. There are many options for opioids during anesthesia and analgesia: MOP receptor agonists (sufentanil, remifentanil), KOP receptor agonists (nalbuphine, butorphanol), MOP and KOP dual receptor agonists (oxycodone), which act on different receptors, and the clinical doses are also different (Kvam et al., 2004; Gepts et al., 1995; Egan, 1998; He et al., 2021; Opioids for pain, 2022; Raff et al., 2019). It is not clear which of these drugs has the least and the most severe inhibition of gastrointestinal function. It is possible to find the drug with the least inhibitory effect on gastrointestinal function through studies comparing several different drugs simultaneously under the same conditions. In this way, on the premise of meeting the requirements of analgesia, drugs with less inhibition on gastrointestinal function can be preferred clinically.

Therefore, we designed this *in vitro* animal experiment to compare the effects of various opioids on the motility of isolated rat intestinal smooth muscle at different concentrations.

Methods

Laboratory animal

All animal experiments were performed in accordance with the basic principles of Fudan University's animal experiments. The Animal Ethics Committee of Shanghai Medical College of Fudan University approved the protocol (201907007Z), and specific-pathogen-free (SPF) 10 Sprague Dawley rats, half male and half female, weighing between 200–220 g, were purchased from Shanghai Jiesi Laboratory Animal Co., Ltd. Before the experiment, they were placed in standard cages and reared for 1 week in a 12 h:12 h light-dark cycle, with food and water *ad libitum*.

Experimental process

Rats were fasted for 10 h before the experiment and had free access to water. On the day of the experiment, all rats were euthanized by inhaling carbon dioxide (replacement rate30%-70%). After the rat was determined to have died, the peritoneal cavity was exposed through a median incision. Identify the ligament of Treitz, isolate the segment from jejunum to ileum, trim away the mesentery along the intestinal wall, and rinse the segment in a Petri dish. Physiological saline solution (PSS) was pre-prepared (8.0 g of NaCl, 0.2 g of KCl, 0.214 g of MgCl₂ 6H₂O, 0.05 g of NaH₂PO₄, 1.0 g of NaHCo₃, 0.2 g of CaCl₂, 1.0 g of glucose, and 1,000 mL of double distilled water). The intestinal contents were rinsed with a 10 mL syringe in PPS, and the intestinal segments were cut into 1 cm small segments (at least six segments per rat).

One end of the intestinal tube was connected with a tension transducer (JZJ01 type, China Chengdu instrument Factory) with cotton thread, and the other end of the intestinal tube was fixed in the smooth muscle groove with an "L"-shaped hook. The intestinal

segment was completely immersed in a smooth muscle tank filled with 30 mL of Tyrode's solution, and the temperature in the bath was kept constant at 37 °C. The smooth muscle tank was inflated with an oxygen pump (95% O2, 5% CO2 ventilation, two air bubbles/s). The connection between the intestinal tube and the tension transducer was adjusted so that it had a certain degree of tension, and the preload was set to 1 g.

The tension transducer is connected to a polyconductive physiological recorder (RM6240BD, China Chengdu Instrument Factory) to record the signal changes of isometric contraction of jejunal smooth muscle. The processed signal of the RM6240 (USB-3.8 version) biological signal acquisition and processing system is displayed on the computer display scree (Figure 1).

n. Six intestinal segments from the same rat were selected, placed in six constant temperature water baths, and randomly divided into six groups for synchronous experiments. Sufentanil citrate, remifentanil hydrochloride, oxycodone hydrochloride, nalbuphine hydrochloride, butorphanol tartrate, and PSS were added respectively in stepwise increasing concentrations.The experimental liquid was calculated and prepared according to the molecular weight of the drug. At 10⁻⁴ M, remifentanil is 41.29 μg/ mL, and sufentanil is 57.87 μg/mL oxycodone 35.18 μg/mL, nalbuphine 42.09 µg/mL, butorphanol 47.76 µg/mL, and configuration concentration gradients of 10⁻⁹, 10⁻⁸, 10⁻⁷, 10⁻⁶, 10^{-5} , and 10^{-4} M were diluted with PSS.

The small intestine segments mounted longitudinally equilibrated for 30 min, and after the spontaneous contraction activity was stable, the contraction tension and contraction frequency were observed, and recorded as baseline values. Then, the experiment proceeded with stepwise increases in concentration from low to high, without washout between concentrations. Each concentration was studied for 10 min, the intestinal segment was balanced at this concentration for the first 5min, and the intestinal muscle contraction frequency and contraction tension were recorded for the next 5 min. During the whole experiment, the control group synchronously replaced the same PSS and the contraction tension, contraction frequency was recorded, the total contraction time remained consistent. In order to exclude individual differences of intestinal muscle contractility in different rats, we used the baseline values of contractile tension and contractile frequency of the same rat to calculate the ratio of intestinal muscle contractile tension and contractile frequency at different concentrations of each drug, and then compared them.

Statistical analysis

Data analysis was performed using SPSS 22.0 (SPSS, Inc., Chicago, IL, USA). The Kolmogorov–Smirnov test was used to test the distribution of univariate data. The dose–response curve was fitted with the logarithm of the drug concentration as the abscissa and the ratio of contraction tension and contraction frequency as the ordinate. Variables were expressed as medians (interquartile range), the Kruskal–Wallis rank-sum test was used for comparisons between different concentrations of the same drug, and multiple comparisons were performed using the Dunnet statistical method. P < 0.05 was considered statistically significant.

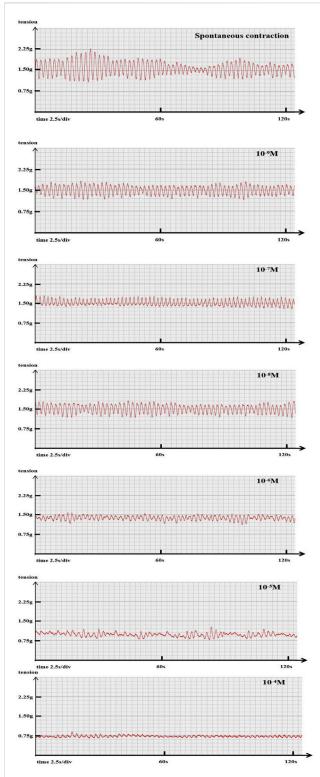


FIGURE 1 Representative original trace A representative example showing the changes in ileal segment contraction tension and frequency with increasing sufentanil concentration. Baseline: tension 1.15 g, frequency 27 cycles/min. Sufentanil 10^{-9} M: tension 0.89 g, frequency 24 cycles/min; 10^{-8} M: 0.77 g, 17 cycles/min; 10^{-7} M: 0.71 g, 17 cycles/min; 10^{-8} M: 0.67 g, 14 cycles/min; 10^{-5} M: 0.52 g, 14 cycles/min; 10^{-6} M: 0.34 g, 10 cycles/min.

Results

The data obtained from the *in vitro* intestinal tube experiment of 10 rats showed in Table 1. Statistical analysis found that butorphanol (10⁻⁹-10⁻⁴ M) decreased the tension ratio in a concentrationdependent manner. The first significant reduction occurred at 10^{-7} M (0.85 \pm 0.26; p = 0.013 versus baseline). Significant inhibition persisted at 10^{-6} , 10^{-5} and 10^{-4} M (p < 0.05). Nalbuphine produced its first significant effect at 10^{-6} M (0.80 \pm 0.24; p = 0.027) and remained lower at 10^{-5} and 10^{-4} M (p < 0.05). Oxycodone significantly reduced tension at 10^{-6} M (0.85 \pm 0.20; p = 0.024) and maintained inhibition up to 10^{-4} M (p < 0.05). Sufentanil and remifentanil decreased tension at 10^{-5} M (p = 0.010) and 10^{-4} M (p = 0.007), respectively. The control group showed no significant change, only a slight drift (Figure 2). Meanwhile, the tension ratio were compared with time-match control group, all differences remained significant except at 10⁻⁶ M butorphanol and nalbuphine produced significant changes at 10⁻⁷ M.

Regarding the effect on the contraction frequency of intestinal smooth muscle (Table 2), the ratio of contraction frequency decreased significantly in the sufentanil group when the drug concentration increased to 10^{-4} M compared with the baseline (0.7 [0.23], P < 0.001). There was no statistical difference among different concentrations of other drugs. In the control group, the contraction frequency remained virtually unchanged over time (Figure 3). After comparing the contraction frequency ratio with the time-matched control group, the statistical results were consistent.

Discussion

In this study, by comparing the effects of commonly used opioid analgesics on the contraction tension and contraction frequency of the isolated intestinal canal at different concentrations, it was found that the inhibitory effect of opioids on isolated intestinal contraction was concentration dependent. The concentrations at which the intestinal contraction tension began to decrease significantly were butorphanol 10^{-7} M, nalbuphine and oxycodone 10^{-6} M, sufentanil 10^{-5} M, and remifentanil 10^{-4} M. According to the above results, the order of drugs for inhibiting intestinal contraction tension was as follows: butorphanol > nalbuphine \approx oxycodone > sufentanil > remifentanil.

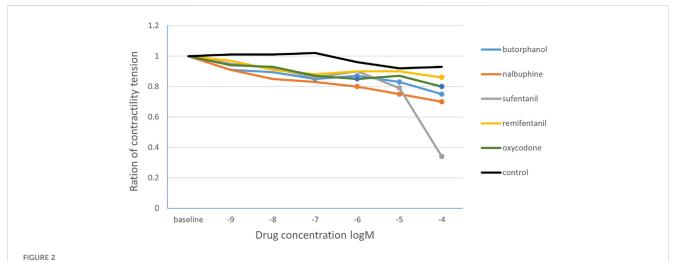
Opioids lead to postoperative gastrointestinal dysfunction by activating opioid receptors. The classic opioid receptors have three main receptor subtypes, MOP, KOP and DOP. Among the MOP receptor, MOP1 receptor are involved in analgesia and sedation; MOP2 receptor are involved in respiratory depression, pruritus, and gastrointestinal function inhibition; DOP receptors are involved in spinal cord analgesia, mediate smooth muscle effects and regulate MOP receptor activity (Hill and Canals, 2022; Galligan and Sternini, 2017; DiCello et al., 2020); and while the KOP receptor is involved in intestinal inhibition. Different research results have been reported. The KOP receptor agonist U50488H (1–100 mg/kg) was found to inhibit gastric emptying and small intestinal motility in male Wistar rats in a study of intraperitoneal injection (Asai et al., 1998), but the

TABLE 1 The ratio of intestinal smooth muscle contraction tension with different concentrations of opioids.

Drug concentration	Butorphanol	Nalbuphine	Oxycodone	Sufentanil	Remifentanil	Control
10 ⁻⁹ M	0.91 (0.20)	0.91 (0.12)	0.94 (0.09)	0.95 (0.06)	0.97 (0.20)	1.01 (0.08)
10^{-8} M	0.89 (0.21)	0.85 (0.14)	0.93 (0.14)	0.92 (0.14)	0.91 (0.29)	1.01 (0.13)
10 ⁻⁷ M	0.85 (0.26)*#	0.83 (0.25)#	0.87 (0.19)	0.86 (0.18)	0.89 (0.27)	1.02 (0.13)
10 ⁻⁶ M	0.87 (0.20)*	0.80 (0.24)*#	0.85 (0.20)*#	0.90 (0.28)	0.90 (0.37)	0.96 (0.02)
10 ⁻⁵ M	0.83 (0.29)*#	0.75 (0.25)*#	0.87 (0.24)	0.79 (0.25)*#	0.90 (0.26)	0.92 (0.14)
10^{-4} M	0.75 (0.25)*#	0.70 (0.38)*#	0.80 (0.24)*#	0.34 (0.27)*#	0.86 (0.65)*	0.93 (0.16)

^{*}Statistically significant difference from baseline P < 0.05.

Data are presented as the median (interquartile range).



The change curve of the ratio of intestinal smooth muscle contraction tension under different concentrations of opioids. The solid circles are the drug concentrations corresponding to the statistically significant changes in the ratio of the contraction tension. Butorphanol 10^{-7} M, 10^{-6} M, 10^{-5} M, 10^{-4} M concentrations; nalbuphine 10^{-6} M, 10^{-6} M, 10^{-4} M concentration; sufentanil 10^{-5} M concentration; remifentanil 10^{-4} M concentration.

TABLE 2 The ratio of intestinal smooth muscle contraction frequency of different concentrations of opioids.

Drug concentration	Butorphanol	Nalbuphine	Oxycodone	Sufentanil	Remifentanil	Control
10 ⁻⁹ M	1.00 (0.28)	1.00 (0.16)	0.98 (0.10)	0.96 (0.37)	1.00 (0.24)	1.00 (0.10)
10 ⁻⁸ M	0.91 (0.14)	0.94 (0.34)	0.97 (0.13)	0.97 (0.41)	0.95 (0.32)	1.00 (0.09)
10 ⁻⁷ M	0.86 (0.40)	0.96 (0.14)	1.00 (0.19)	0.96 (0.39)	0.94 (0.30)	0.93 (0.19)
10^{-6} M	0.89 (0.37)	0.93 (0.17)	0.98 (0.07)	0.9 (0.56)	0.89 (0.43)	0.97 (0.27)
10 ⁻⁵ M	0.89 (0.14)	0.97 (0.30)	0.95 (0.22)	0.9 (0.15)	0.83 (0.43)	0.97 (0.20)
10 ⁻⁴ M	0.98 (0.71)	0.89 (0.33)	0.9 (0.26)	0.7 (0.23)*	0.88 (0.48)	0.98 (0.19)

^{*}Statistically significant change from baseline P < 0.05.

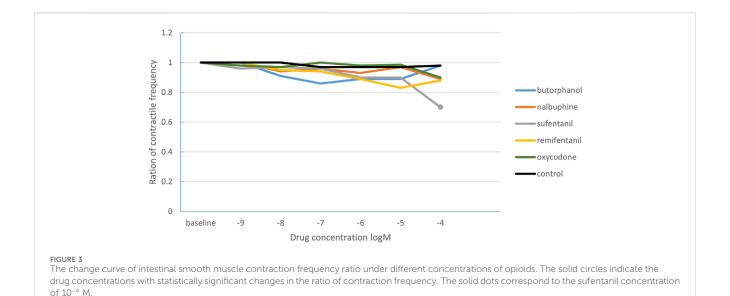
Data are presented as the median (interquartile range).

study on voluntary intestinal smooth muscle found that the KOP-receptor agonist U50488H had no effect on spontaneous contractile activity of muscle strips, which may partly account for the divergent results (Chamouard et al., 1993).

There are many types of opioids that we can choose clinically. The five drugs in the study are commonly used intraoperative and postoperative analgesics, and sufentanil and remifentanil are MOP receptor agonists (Gepts et al., 1995; Egan, 1998). Nalbuphine is a complete agonist of KOP receptors and a partial antagonist of MOP receptors (He et al., 2021); butorphanol mainly stimulates KOP receptors and has dual effects of agonism and antagonism on MOP receptors. The intensity of butorphanol on KOP:MOP:DOP

[#] Statistically significant difference compared with time-match control group P < 0.05.

[#] Statistically significant difference compared with time-match control group P < 0.05.



receptors is $25:4:1,^{10}$ and oxycodone is a synthetic pure opioid receptor μ -k dual agonist (Raff et al., 2019). At present, no study has compared their effects on gastrointestinal function at the same time. Because of individual differences and drug limitations, it is difficult to study five drugs at the same time with postoperative gastrointestinal dysfunction as the main result. Therefore, we chose the rat intestinal tube for *in vitro* experiments for comparison. To minimize temporal bias, we used the same intestinal segment from each rat and included a time-matched control group, compared with time-match control group, the results remained virtually identical, indicating a weak time-dependent effect.

Of course, the effect of opioids on gastrointestinal function is a multichannel and complex process. In addition to directly acting on opioid receptors, it also inhibits the excitability of neurons in the enteric nervous system, resulting in the inhibition of the fluidity of intestinal contents. Drugs act on secretion-promoting neurons in the submucosal nerve plexus, inhibiting the release of neurotransmitters that induce secretory activity in the intestinal recess, reducing intestinal secretion, leading to decreased gastrointestinal function and the formation of constipation (Hughes et al., 2016; Kokki et al., 2012). While clinical research on opioids mainly focus on analgesic efficacy, postoperative gastrointestinal dysfunction often as incidental results (Sun et al., 2020; Han et al., 2018; Guo et al., 2022), which may be the reason why the results of clinical studies on the effects of opioids on gastrointestinal function differ from *in vitro* studies.

For this *in vitro* experiment, different molar concentrations of opioids calculated by molecular weight were selected for longitudinal comparison. Considering the clinical dose of each drug, the application of nalbuphine, butorphanol, and oxycodone is in mg for intraoperative and postoperative analgesia, while sufentanil and remifentanil are in µg. The blood concentrations of sufentanil and remifentanil at commonly used clinical doses are approximately 100–1,000 times lower than those of butorphanol and the other three drugs. Considering the equivalent analgesic dose between different drugs, we believe that the ranking of inhibition of these drugs on the intestinal smooth muscle will not change, and the difference will be more obvious. A previous *in vitro*

study observed the effects of low, medium and high concentrations of morphine, sufentanil, and dezocine on ileal intestinal muscle contraction and further measured the changes of intestinal propulsion motion after intraperitoneal injection of three opioids in male Sprague-Dawley rats (Bian et al., 2015). While keeping the 20 mL bathing volume constant, the study showed that sufentanil at high doses (40-120 µg/L) concentration-dependently increased the tension of isolated rat small-intestinal smooth muscle, a finding that aligns with our results; dezocine, in contrast, had no significant effect on smooth-muscle contraction. Both opioids, however, may impair intestinal propulsive function. The results of Guo's (Yan et al., 2021) research recently also supports our conjecture. The study investigated the effect of equivalent doses of fentanyl, oxycodone and butorphanol on small intestine segments function in patients undergoing laparoscopic hysterectomy and found that butorphanol significantly prolonged postoperative intestinal function recovery, followed by oxycodone. The first postoperative exhaust time of butorphanol, oxycodone and fentanyl was 45.2 \pm 11.6, 36.2 \pm 10.9 and 33.1 \pm 11.2 h, respectively.

This study has several limitations. First, as mentioned above, our study chose molecular weight to calculate the unified experimental drug concentration, longitudinally compared the concentration gradient of opioids affecting gastrointestinal function, and made an indirect horizontal comparison based on the obtained results. Second, intestinal smooth muscle undergoes rhythmic contractions and it also performs circular contractions to complete the compound movement of the intestinal migration (Waterman et al., 1992), which may differ among segments, however we did not differentiate between intestinal segments. The effects of opioids on intestinal smooth muscle are multifaceted; in the present study we examined only changes in basal tension and did not explore potential non-opioid mechanisms or the influence of drugs on electrically or chemically evoked contractions. Finally, in vitro findings cannot be directly extrapolated to the in vivo situation; systemic neural and humoral inputs interact with the drug itself (Chen et al., 2012; Díaz-Ruano et al., 2019), and both opioid-receptor expression and intestinal physiology differ across species. The animal model offers a critical reference point for

subsequent clinical work, but the study's conclusions must await translational validation before they can be applied in practice.

Analysis, Writing – original draft. SS: Writing – review and editing. SH: Writing – review and editing.

Conclusion

The study was found that different opioids affected the contractile tension of isolated rat intestinal smooth muscle with increasing drug concentration: butorphanol > nalbuphine \approx oxycodone > sufentanil > remifentanil, and the contraction frequency was almost unaffected except in the very high-concentration sufentanil group. The consistency of this conclusion provides preliminary animal-model data on opioids, serving as a key reference for future in-vivo or clinical investigations aimed at guiding perioperative opioid selection with minimal gastrointestinal impact.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The animal study was approved by The Animal Ethics Committee of Shanghai Medical College of Fudan University approved the protocol (201907007Z). The study was conducted in accordance with the local legislation and institutional requirements.

Author contributions

CY: Formal Analysis, Methodology, Software, Writing – original draft. JH: Writing – original draft. YC: Data curation, Formal

References

Asai, T., Mapleson, W. W., and Power, I. (1998). Effects of nalbuphine, pentazocine and U50488H on gastric emptying and gastrointestinal transit in the rat. *Br. J. Anaesth.* 80 (6), 814–819. doi:10.1093/bja/80.6.814

Bian, X., Zhou, R., Yang, Y., Li, P., Hang, Y., Hu, Y., et al. (2015). Divergent effect of dezocine, morphine and sufentanil on intestinal motor function in rats. *Int. J. Med. Sci.* 12 (11), 848–852. doi:10.7150/ijms.12616

Chamouard, P., Klein, A., Martin, E., Adloff, M., and Angel, F. (1993). Regulatory role of enteric kappa opioid receptors in human colonic motility. *Life Sci.* 53 (14), 1149–1156. doi:10.1016/0024-3205(93)90551-d

Chen, W., Chung, H. H., and Cheng, J. T. (2012). Opiate-induced constipation related to activation of small intestine opioid μ 2-receptors. World J. Gastroenterol. 18 (12), 1391–1396. doi:10.3748/wjg.v18.i12.1391

Díaz-Ruano, S., López-Pérez, A. E., Girón, R., Pérez-García, I., Martín-Fontelles, M. I., and Abalo, R. (2019). Fluoroscopic characterization of colonic dysmotility associated to opioid and cannabinoid agonists in conscious rats. *J. Neurogastroenterol. Motil.* 25 (2), 300–315. doi:10.5056/jnm18202

DiCello, J. J., Carbone, S. E., Saito, A., Rajasekhar, P., Ceredig, R. A., Pham, V., et al. (2020). Mu and Delta opioid receptors are coexpressed and functionally interact in the enteric nervous system of the mouse Colon. *Cell Mol. Gastroenterol. Hepatol.* 9 (3), 465–483. doi:10.1016/j.jcmgh.2019.11.006

Egan, T. D. (1998). The clinical pharmacology of remifentanil: a brief review. J. Anesth. 12 (4), 195–204. doi:10.1007/BF02481730

Galligan, J. J., and Sternini, C. (2017). Insights into the role of opioid receptors in the GI tract: experimental evidence and therapeutic relevance. *Handb. Exp. Pharmacol.* 239, 363–378. doi:10.1007/164_2016_116

Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Gepts, E., Shafer, S. L., Camu, F., Stanski, D. R., Woestenborghs, R., Van Peer, A., et al. (1995). Linearity of pharmacokinetics and model estimation of sufentanil. *Anesthesiology* 83 (6), 1194–1204. doi:10.1097/00000542-199512000-00010

Guo, M., Liu, S., Gao, J., Han, C., Yang, C., and Liu, C. (2022). The effects of fentanyl, oxycodone, and butorphanol on gastrointestinal function in patients undergoing laparoscopic hysterectomy: a prospective, double-blind, randomized controlled trial. *BMC Anesthesiol.* 22 (1), 53. doi:10.1186/s12871-022-01594-9

Han, L., Su, Y., Xiong, H., Niu, X., Dang, S., Du, K., et al. (2018). Oxycodone *versus* sufentanil in adult patient-controlled intravenous analgesia after abdominal surgery: a prospective randomized double-blinded multiple-center clinical trial. *Med. Baltim.* 97 (31), e11552. doi:10.1097/MD.0000000000011552

He, K., Ji, W., Zhao, H., Wei, Y., Yang, S., and Wen, Q. (2021). Pharmacokinetic comparison of nalbuphine with single injection and patient-controlled analgesia mimic method in healthy Chinese volunteers. *J. Clin. Pharm. Ther.* 46 (4), 1166–1172. doi:10. 1111/jcpt.13421

Hill, R., and Canals, M. (2022). Experimental considerations for the assessment of *in vivo* and *in vitro* opioid pharmacology. *Pharmacol. Ther.* 230, 107961. doi:10.1016/j. pharmthera.2021.107961

Hughes, P. A., Costello, S. P., Bryant, R. V., and Andrews, J. M. (2016). Opioidergic effects on enteric and sensory nerves in the lower GI tract: basic mechanisms and clinical implications. *Am. J. Physiol. Gastrointest. Liver Physiol.* 311 (3), G501–G513. doi:10.1152/ajpgi.00442.2015

Imam, M. Z., Kuo, A., Ghassabian, S., and Smith, M. T. (2018). Progress in understanding mechanisms of opioid-induced gastrointestinal adverse effects and

respiratory depression. Neuropharmacology 131, 238-255. doi:10.1016/j.neuropharm. 2017.12.032

Kokki, M., Franco, M. G., Raatikainen, K., Välitalo, P., Sankilampi, U., Heinonen, S., et al. (2012). Intravenous oxycodone for pain relief in the first stage of labour--maternal pharmacokinetics and neonatal exposure. *Basic Clin. Pharmacol. Toxicol.* 111 (3), 182–188. doi:10.1111/j.1742-7843.2012.00884.x

Kvam, T. M., Baar, C., Rakvåg, T. T., Kaasa, S., Krokan, H. E., and Skorpen, F. (2004). Genetic analysis of the murine mu opioid re-ceptor: increased complexity of oprm gene splicing. *J. Mol. Med. Berl.* 82 (4), 250–255. doi:10.1007/s00109-003-0514-z

Müller-Lissner, S., Bassotti, G., Coffin, B., Drewes, A. M., Breivik, H., Eisenberg, E., et al. (2017). Opioid-induced constipation and bowel dysfunction: a clinical Guideline. *Pain Med.* 18 (10), 1837–1863. doi:10.1093/pm/pnw255

Mythen, M. G. (2009). Postoperative gastrointestinal tract dysfunction: an overview of causes and management strategies. *Cleve Clin. J. Med.* 76 (Suppl. 4), 66–71. doi:10.3949/ccjm.76.s4.11

Opioids for pain (2022). Opioids for pain. Med. Lett. Drugs Ther. 64(1665): 193-200.

Raff, M., Belbachir, A., El-Tallawy, S., Ho, K. Y., Nagtalon, E., Salti, A., et al. (2019). Intravenous oxycodone *versus* other intravenous strong opioids for acute postoperative pain control: a systematic review of randomized controlled trials. *Pain Ther.* 8 (1), 19–39. doi:10.1007/s40122-019-0122-4

Sun, S., Guo, Y., Wang, T., and Huang, S. (2020). Analgesic effect comparison between Nalbuphine and sufentanil for patient-controlled intravenous Analgesia after cesarean section. *Front. Pharmacol.* 11, 574493. doi:10.3389/fphar.2020.574493

Tao, B., Liu, K., Wang, D., Ding, M., and Zhao, P. (2019). Effect of intravenous oxycodone *versus* sufentanil on the incidence of postoperative nausea and vomiting in patients undergoing gynecological laparoscopic surgery. *J. Clin. Pharmacol.* 59 (8), 1144–1150. doi:10.1002/jcph.1408

Waterman, S. A., Costa, M., and Tonini, M. (1992). Modulation of peristalsis in the guinea-pig isolated small intestine by exogenous and endogenous opioids. Br. J. Pharmacol. 106 (4), 1004–1010. doi:10.1111/j.1476-5381.1992.tb14448.x

Yan, Y., Chen, Y., and Zhang, X. (2021). The effect of opioids on gastrointestinal function in the ICU. $Crit.\ Care\ 25\ (1),\ 370.\ doi:10.1186/s13054-021-03793-1$