

OPEN ACCESS

EDITED BY Wei Peng.

Chengdu University of Traditional Chinese Medicine, China

REVIEWED BY

Qiping Zhan,

Nanjing Agricultural University, China Ziwei Yue,

Beijing Forestry University, China

*CORRESPONDENCE

Zilong Zhao,

bigdragonbrother@163.com

Xinghua Wang,

RECEIVED 04 July 2025 ACCEPTED 03 October 2025 PUBLISHED 13 October 2025

CITATION

Chen J, Zhao Z, Lin L, Wang G, Yang H and Wang X (2025) *Aucklandia lappa* Decne.: a review of its botany, cultivation, ethnopharmacology, phytochemistry, pharmacology, and practical applications. *Front. Pharmacol.* 16:1659831. doi: 10.3389/fphar.2025.1659831

COPYRIGHT

© 2025 Chen, Zhao, Lin, Wang, Yang and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Aucklandia lappa Decne.: a review of its botany, cultivation, ethnopharmacology, phytochemistry, pharmacology, and practical applications

Jing Chen¹, Zilong Zhao²*, Lihua Lin¹, Guangyao Wang³, Haixia Yang⁴ and Xinghua Wang^{3,5}*

¹School of Environmental and Food Engineering, Liuzhou Polytechnic University, Liuzhou, China, ²School of Chemical Engineering, Northwest University, Xi'an, China, ³College of Traditional Chinese Medicine, Nanjing, China, ⁴Brain Hospital Affiliated to Nanjing Medical University, Nanjing, China, ⁵Faculty of Medicine and Health Sciences, Universiti Tunku Abdul Rahman, Selangor, Malaysia

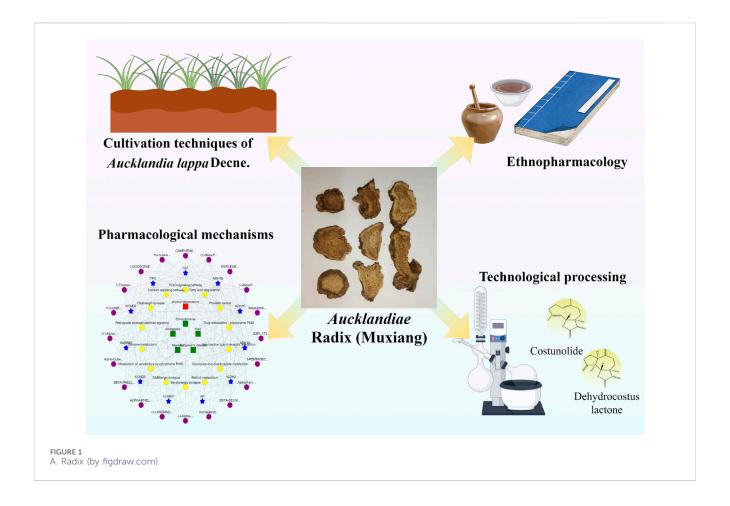
Aucklandia lappa Decne. (ALD), a synonym of Saussurea costus (Falc.) Lipsch., is a traditional Chinese medicinal herb extensively cultivated in China. Aucklandiae Radix (AR, known as "Muxiang" in China), derived from the dried root of ALD, holds a significant position in the clinical application of traditional Chinese medicine, encompassing the enhancement of gastrointestinal motility, antibacterial properties, and antitumor activities. Notably, AR possesses a complex and diverse chemical composition, with costunolide and dehydrocostus lactone being its core active metabolites. This review provides an in-depth exploration of the biological characteristics, cultivation techniques, ethnopharmacology, phytochemistry, pharmacological activities, and processing techniques associated with ALD. To collect relevant research materials, the study systematically retrieved information from authoritative databases such as CNKI, PubMed, Elsevier, Web of Science, and SpringerLink, employing keywords including "cultivation," "phytochemistry," "pharmacology," and the plant names "Aucklandia lappa Decne.," "Saussurea costus (Falc.) Lipsch.," or "Aucklandiae Radix." Despite demonstrating remarkable pharmacological activities and potential for clinical applications, research on ALD still faces several challenges. For instance, its specific mechanisms of action in treating certain diseases remain incompletely understood, and multiple studies have indicated that ALD extracts may cause adverse reactions. Further in-depth research and systematic evaluation can facilitate the optimization of ALD practices to promote further research into its myriad applications.

KEYWORDS

Aucklandia lappa Decne., Aucklandiae Radix, Chinese medicine, applications, Muxiang (Radix Aucklandiae)

1 Introduction

Aucklandia lappa Decne. (ALD), synonym of Saussurea costus (Falc.) Lipsch., is a perennial herb belonging to the Asteraceae family that possesses a rich medicinal history extending over two millennia (Huang et al., 2021; Song et al., 2021). Currently, ALD is predominantly cultivated in the Yunnan, Guizhou, Guangxi and Sichuan provinces of



China (Xue et al., 2020). Aucklandiae Radix (AR), commonly known as Muxiang or Yunmuxiang (Chinese trade name), is derived from the dried root of ALD. It has been utilized extensively as a medicinal material in traditional Chinese medicine (TCM) and is officially recognized in the Chinese Pharmacopoeia (Huang et al., 2021; Shum et al., 2007). Since 2014, when related industrial bases were established in Yunnan and Guizhou Province and Chongqing of China, large-scale planting was carried out based on the growth characteristics of ALD, and its economic benefits have achieved a leapfrog growth. In 2021, the national sales volume of ALD was approximately 6,000 tons, and by 2024, the national sales volume of ALD exceeded 8,300 tons.

The pharmacological effects of AR are diverse and significant (Li S.-Y. et al., 2024). Modern pharmacological researches have demonstrated that AR exhibits several properties, including the promotion of gastrointestinal motility, dilation of bronchial smooth muscles, antibacterial activity, reduction of blood glucose levels, and antitumor effects (Song et al., 2022b; Zhang et al., 2021;

Abbreviations: ALD, Aucklandia lappa Decne.; AR, Aucklandiae Radix; ARR, Aristolochiae Radix; DSS, dextran sulfate sodium; E-nose, electronic nose; GC, gas chromatography; IBD, inflammatory bowel disease; ICPMS, inductively coupled plasma mass spectrometry; IR, Inulae Radix; LC, liquid chromatography; MBC, minimal bactericidal concentration; MIC, minimum inhibitory concentration; MS, mass spectrometry; NMR, nuclear magnetic resonance; TCM, traditional Chinese medicine; TLC, thin-layer chromatography; VR, Vladimiriae Radix.

Zheng et al., 2022; Zhuang et al., 2021). The chemical composition of AR is complex and diverse, primarily encompassing terpenoids, glycosides, and other metabolites (anthraquinones, flavonoids, amino acids, etc.). Costunolide and dehydrocostus lactone are principal active metabolites in AR (Huang Z. et al., 2022). The total content of costunolide (PubChem CID 5281437, $C_{15}H_{20}O_2$) and dehydrocostus lactone (PubChem CID 73174, $C_{15}H_{18}O_2$) in AR must not be less than 1.8%, as stipulated by the Chinese Pharmacopoeia (2020 Edition). Concurrently, the European Union's Traditional Herbal Medicinal Products Directive has officially recognized AR as a certified herbal preparation for the treating functional dyspepsia.

This review provides a comprehensive examination of the biological characteristics, cultivation techniques, chemical composition, traditional Chinese medicinal applications, pharmacological activities, and processing methods associated with ALD (Figure 1). Large-scale cultivation and standardized production, circular economy model innovation, and the development of bio-synthesized active substances along with alternative pathways can facilitate the optimization of ALD practices to promote further research into its diverse applications.

2 Botanical characteristics

ALD is a perennial, tall herb characterized by a thick main root with a distinctive aroma. The basal leaves are triangular-ovate,

featuring pinnately lobed upper long petioles and shallowly lobed margins. The stem leaves are also triangular-ovate or ovate, with the base extending downward and being sessile or having winged petioles. ALD produces capitula with multiple outer bracts, and its flowers are bisexual, dark purple, tubular, with inferior ovaries. The flowering period occurs from July to August, followed by a fruiting period from August to September (Chen, 2025). ALD thrives in cool to cold climates and exhibits significant cold resistance, making it particularly suitable for high-altitude regions with relatively low temperatures and high humidity. This species is a deep-rooted plant, with roots extending 30–50 cm or even deeper into the soil.

3 Cultivation and management

The cultivation of ALD plays a decisive role in determining the quality of its medicinal material. This influence is primarily reflected in four key aspects: geo-authenticity, growing environment, cultivation management, and harvesting and processing practices. Firstly, the specific altitude, climate, and soil conditions in geoauthentic regions promote the accumulation of higher levels of active metabolites (such as costunolide and dehydrocostus lactone) in the plant. In contrast, non-authentic producing areas typically yield inferior medicinal efficacy. Secondly, scientific cultivation management is central to quality assurance. The use of highquality seeds, emphasis on well-rotted organic fertilizers, crop rotation, and integrated green pest management ensure healthy plant growth without pesticide residues. Conversely, excessive use of chemical fertilizers and pesticides compromises quality and introduces safety risks. Finally, timely harvesting standardized post-harvest processing are crucial for preserving the therapeutic potency and aromatic properties. Harvesting too early or too late, or using high-temperature drying methods, can lead to the loss of volatile oils and significantly diminish efficacy. Therefore, standardized management throughout the entire process, is essential for producing ALD that is safe, effective, and of high quality.

3.1 Cultivation techniques

When selecting a cultivation site for ALD, it is imperative to ensure that the slope faces east or north and the slope is maintained at 30°-35°. A shaded slope is preferable. Additionally, the soil should be loose, fertile and possess a deep profile, with loam or sandy loam enriched with humus being optimal, as this composition facilitates effective drainage. Low-lying areas, which are susceptible to flooding and consequently to root rot, should be avoided. For newly developed land, it is essential to thoroughly clear all the weeds and existing vegetation from the wasteland, incorporating these materials into the soil through deep plowing. Deep plowing should be carried out again in the following spring. In addition, cultivators may opt to grow wood-scented plants on land that has previously supported crops such as potatoes and corn (Chen, 2025). Given that ALD can accumulate harmful metals from its native soil, consumption of low-quality ALD may lead to the accumulation of toxic metal elements in the human body (Meng et al., 2021). Inductively coupled plasma mass spectrometry (ICPMS) provides a precise and reliable method for monitoring and controlling contamination levels in the extract, such as Cu, Pb, As, Cd, and Hg in ALD (Chen et al., 2023). The soil used for planting should effectively control the pollution of related metals. The arbuscular mycorrhizal fungi strains (Gigaspora decipiens, Scutellospora calospora, Racocetra coralloidea, Septoglomus deserticola, Entrophospora colombiana, Paraglomus brasilianum), which had a good symbiotic relationship with ALD, are the potential strains to inoculate ALD seedlings under artificial cultivation conditions (Yang et al., 2020). Symbiotic bacteria can significantly increase terpenoids accumulation (costunolide, dehydrocostus lactone, etc.), soluble protein, soluble sugar, antioxidant enzyme activity in the leaves.

3.2 Reproductive methods

Seed propagation serves as the primary method for the cultivation of ALD. In Yunnan Province, the cultivation of ALD predominantly involves seeds sowing, which can be conducted during the winter, autumn, and spring. Typically, spring sowing occurs from mid-March to early April, autumn sowing from late August to mid-September, and winter sowing in early November. Post-harvest, the seeds must be sun-dried and subsequently cleaned of impurities to prepare them for subsequent processes. Prior to sowing, it is essential to prepare the seeds by soaking them in lukewarm water with continuous stirring. As the water cools, impurities and non-viable seeds are removed, leaving only the viable seeds that settle at the bottom. These viable seeds are then soaked for 24 h before being partially dried in preparation for sowing.

If the seeds quantity is insufficient for planting requirements, asexual reproduction can be considered. It is crucial to avoid using fine roots with medicinal value as breeding material. During planting, the layout should be carried out according to the prescribed spacing. And when covering the soil, it is necessary to ensure that the root systems are completely and tightly buried. ALD propagation via cuttings requires a healthy selection of semihardened stems with 2-3 nodes. Stems are trimmed to retain 2-3 leaves for photosynthesis, followed by treatment with rooting hormones or basal soaking in diluted rooting agent for 30 min prior to air-drying. A sterile, well-draining substrate (e.g., a mixture of leaf mould and perlite) is prepared and sterilized to minimize pathogen risk. Processed cuttings are inserted into the substrate at a depth of 1/3-1/2 the stem length. Post-insertion, the medium is moderately watered to maintain moisture without waterlogging. Environmental conditions are maintained at 20 °C-25 °C with indirect light and adequate airflow to avoid direct sunlight exposure. Rooting initiates within 3-4 weeks, accompanied by new shoot development.

Large-scale planting has been carried out based on the growth characteristics of ALD in Yunnan and Guizhou Province and Chongqing of China. This large-scale planting is still mainly based on traditional agricultural cultivation methods. In the future, the yield, quality and production efficiency of ALD can be improved through multi-disciplinary means such as micropropagation and tissue culture technology, molecular biology, intelligent equipment and environmental control.

Micropropagation and tissue culture technology represent a fundamental advancement in contemporary crop cultivation (Alanagh et al., 2014; Lu et al., 2019). By employing asexual reproduction and cellular engineering techniques, these methods effectively address the challenges of low efficiency and genetic instability inherent in traditional seed propagation (Pupilli and Barcaccia, 2012). The root tips of ALD serve as highly differentiated potential explants suitable for plant tissue culture. By precisely regulating the growth system through genetic modification and improving stress resistance and the targeted accumulation of metabolites, the production efficiency and product value of ALD can be significantly enhanced. The precise regulation of nutrient solutions during the cultivation process can now be easily implemented (Lu et al., 2022; Wang et al., 2023). The demand for elements such as nitrogen, phosphorus, potassium, and calcium varies significantly at different stages of plant growth. Through dynamic monitoring and intelligent intervention, the accumulation of secondary metabolites, stress resistance, and growth consistency of medicinal plants like ALD may be significantly improved. Furthermore, based on historical data, a growth prediction model for ALD can be constructed, integrating environmental variables (temperature, precipitation, soil EC value) to output the best irrigation and fertilization decisions. If synthetic biology and intelligent equipment can be effectively combined, the ALD industry is expected to become a benchmark model in the global medicinal plant field.

3.3 Control of pests and diseases

Leaf spot disease and root rot represent significant threats to the growth cycle of ALD, particularly during the rainy season when the prevalence of these diseases increases markedly, with July and August identified as peak periods. To mitigate these challenges, growers should prioritize land with superior drainage and a lower groundwater table for cultivating ALD. During field management, it is crucial to handle the plants carefully to prevent root injury and to rigorously implement quarantine measures to ensure that seeds are free from pathogens. Once infected plants are identified, they should be immediately removed, and the soil should be disinfected with quicklime to curb the further spread of root rot (Tan et al., 2023). In cases where ALD seedlings exhibit symptoms of root rot, growers need to take prompt action by precisely spraying an appropriate amount of thiophanate-methyl or carbendazim on the roots of the affected plants (Gaitnieks et al., 2016). Additionally, during the rainy season, the application of Bordeaux mixture on clear days is advised to effectively prevent leaf spot disease. Upon noticing symptoms of disease on ALD seedlings, an appropriate amount of bactericides (tebuconazole or chlorothalonil) should be evenly sprayed on the leaves, with attention to rotating different pesticides to enhance control effectiveness.

The primary pests impacting ALD include grasshoppers, aphids, cutworms, and grubs. During their nymphal stage, grasshoppers can be effectively managed by spraying with a solution of 90% crystalline trichlorfon diluted to 800 times (Zhang et al., 2019). Aphids can be controlled by spraying with a solution of 40% dimethoate emulsifiable concentrate diluted 800 to 1,500 times (Chandrasena

et al., 2011). Cutworms and grubs, which damage seedlings, roots, and leaves, can be effectively trapped and eradicated using bait made by combining crystalline trichlorfon with wheat bran (Kumar and Pandey, 2022; Smirle et al., 2013).

4 Ethnopharmacology

The earliest documentation of AR appears in the ancient book "Supplements to the *Shennong Bencao* by Medical Masters" (《名医别录》). Subsequently, classical works such as "The Newly Revised Materia Medica" (《新修本草》), "Illustrated Classic of Materia Medica" (《本草图经》), "Essential Documents of the Tang Dynasty" (《唐会要》), "Muslim Medicine" (《回回药方》), and "Compendium of Materia Medica" (《本草纲目》) have systematically recorded its applications, underscoring its integration into both Chinese pharmacology and cross-cultural medical traditions. In the Qing Dynasty, the "Essential Compendium of Materia Medica" (《本草备要》) systematically standardized the processing techniques of ALD, establishing protocols that balanced traditional practices with empirical refinement. At present, it is officially recognized in the Chinese Pharmacopoeia (《中国药典》2020 Edition).

4.1 Identification of AR

AR originates from the dried root of ALD, which is collected during the autumn and winter seasons. Once dried, the coarse outer skin is removed. The final product is typically cylindrical or semicylindrical, measuring between 5 and 10 cm in length and 0.5–5 cm in diameter. Its surface boasts a yellowish-brown to grayish-brown coloration, adorned with prominent wrinkles, longitudinal grooves, and lateral root marks. The texture is notably firm, making it resistant to breaking. Upon sectioning, the interior reveals a grayish-brown to dark-brown coloration, with a grayish-yellow or light brownish-yellow periphery. A distinct brown cambium ring is evident, accompanied by a radial texture. It possesses a unique aroma and a slightly bitter taste (China Pharmacopoeia Committee, 2020a).

4.2 Traditional uses of AR

Four traditional AR-based prescriptions have been recorded in the Chinese Pharmacopoeia (2020 edition): Muxiang Fenqi Wan (China Pharmacopoeia Committee, 2020e), Muxiang Shunqi Wan (Bai, 2023; China Pharmacopoeia Committee, 2020f), Muxiang Binglang Wan (China Pharmacopoeia Committee, 2020d; Gao et al., 2024), and Liuwei Muxiang San (China Pharmacopoeia Committee, 2020c). These traditional medicines primarily treat gastrointestinal stagnation, abdominal distension pain, and bowel obstruction.

Additionally, other traditional prescriptions that were not included in the Chinese Pharmacopoeia have also been studied (Table 1). Simo Decoction has been shown to enhance gastrointestinal motility by influencing the contractions of antral circular smooth muscle strips (Dai et al., 2012). JianPi'I, which

TABLE 1 List of TCM prescriptions related to ALD.

Name	Formula (except for AR)	Indications	Ref.
Muxiang Fenqi Wan	Wurfbainia compacta, Syzygium aromaticum L., Santalum album, Cyperus rotundus L., Pogostemon cablin, Citrus reticulata Blanco, Magnolia officinalis, Citrus aurantium L., Curcuma longa L., Crataegus monogyna Jacq., Atractylodes macrocephala Koidz., Nardostachys jatamansi, Areca catechu L., Glycyrrhiza glabra L.	Bloating, abdominal pain	China Pharmacopoeia Committee (2020e)
Muxiang Shunqi Wan	Wurfbainia compacta, Cyperus rotundus L., Areca catechu L., Glycyrrhiza glabra L., Magnolia officinalis, Citrus aurantium L., Atractylodes Lancea, Citrus reticulata Blanco, Zingiber officinale Roscoe	Bloating, abdominal pain	(Bai, 2023; China Pharmacopoeia Committee, 2020f)
Muxiang Binglang Wan	Areca catechu L., Citrus aurantium L., Citrus reticulata Blanco, Cyperus rotundus L., Sparganium emersum Rehmann, Curcuma longa L., Coptis chinensis Franch., Cortex Phellodendri Chinensis, Rheum officinale Baill., Semen Pharbitidis, Natrii Sulfas	Bloating, abdominal pain, uncomfortable bowel movements	(China Pharmacopoeia Committee, 2020d; Gao et al., 2024)
Liuwei Muxiang San (Wan)	Gardenia jasminoides, Punica Granatum, Rhododendron molle, Wurfbainia compacta Rotundus, Piper longum L.	Bloating, abdominal pain, uncomfortable bowel movements	(China Pharmacopoeia Committee, 2020c; Renqing-Dongzhu et al., 2023)
Simo decoction	Citrus aurantium L., Areca catechu L., Lindera aggregata (Sims) Kosterm.	Gastrointestinal dysmotility	(Dai et al., 2012; Yi et al., 2011)
JianPi'I	Codonopsis pilosula (Franch.) Nannf., Atractylodes macrocephala Koidz., Poria cocos, Wurfbainia compacta, Glycyrrhiza glabra L., Citrus reticulata Blanco, Pinellia ternata (Thunb.) Makino	Postprandial distress syndrome	Wang et al. (2016)
Xian-He-Cao-Chang- Yan formula	Agrimonia eupatoria L., Coptis chinensis Franch., Cyperus esculentus L., Acorus verus (L.) Raf., Platycodon grandiflorus Jacq.	Colitis	Li et al. (2021a)
Weichang'an Pill	Lignum Aquilariae Resinatum, Lignum Aantali Albi, Citrus aurantium L., Cortex Magnoliae officinalis, Rheum officinale Baill., Conioselinum anthriscoides, Croton tiglium L., Ziziphus jujuba Mill., Abelmoschus moschatus Medik.	Irritable bowel syndrome and functional dyspepsia	Zhang et al. (2013)
Kangen-karyu (GuanYuan-Ke-Li)	Salvia miltiorrhiza Bunge, Conioselinum anthriscoides, Paeonia lactiflora Pall., Carthamus tinctorius L., Cyperus rotundus L.	Alzheimer's disease	Paudel et al. (2020)
Xianglian pill	Coptis chinensis Franch.	Gastrointestinal disease	Lu et al. (2020)
Wuwei Shexiang pills	Terminalia chebula Retz., Abelmoschus moschatus Medik., Aconitum carmichaelii Debx., Acorus calamus L.	Joint pain	Bai et al. (2023)
Muxiang gel plaster (Muxiang Bing)	Edible gelatin, <i>Rehmannia glutinosa</i> (Gaertn.) Libosch., catalpol, rehmannioside D, tartaric acid, carbomer, sodium polyacrylate, dihydroxyaluminium aminoacetate, glycerol	Mammary hyperplasia	Wu et al. (2024b)
Anshen-Buxin-Liuwei pill	Bos taurus domesticus Gmelin, Choerospondias axillaris, Myristica fragrans Houtt., Eugenia caryophµllata Thunb., Liquidambar formosana	Cardiomyocyte hypoxia/ reoxygenation injury	Huang et al. (2022a)

originates from the classical Chinese medicine formula Xiangshaliujunzi decoction, is known to alleviate symptoms associated with postprandial distress syndrome (Wang et al., 2016). The Xian-He-Cao-Chang-Yan formula has been demonstrated to ameliorate DSS-induced colitis in mice (Li J. et al., 2021). Weichang'an Pill has been widely used for decades in the treatment of irritable bowel syndrome and functional dyspepsia (Zhang et al., 2013). Kangen-karyu (GuanYuan-Ke-Li) is considered a promising therapeutic agent for Alzheimer's disease (Paudel et al., 2020). Xianglian Pill, composed of Rhizoma coptidis and AR, has long been employed in the management of gastrointestinal disorders (Lu et al., 2020). Clinically, Wuwei Shexiang Pill is used in China to reduce joint pain and swelling, as well as to dispel wind and alleviate pain (Bai et al., 2023). Additionally, treatment with Muxiang gel plaster has been found to effectively prevent and mitigate mammary hyperplasia (Wu Y. et al., 2024).

With the development of TCM, the active metabolites in medicinal substances can be concentrated, extracted and combined to enhance their therapeutic efficacy. A metabolite formulation, consisting of three herbal extracts (CO₂ supercritical fluid extract of ginger, ethanol reflux extract of AR, and pogostemonis herba essential oil) has been developed as a promising anti-motion sickness treatment (Zhang et al., 2015).

4.3 Traditional processing

The traditional processing techniques for ALD involves several methods aimed at enhancing its efficacy and adaptability for various Chinese medicinal formulations (Li X. et al., 2021; Song et al., 2022a). These methods include the use of raw AR, stir-fried AR, baked (or grilled) AR, and wine-processed AR. Raw AR denotes the cleaned and dried form of the original medicinal material, which

undergoes impurity removal, washing, slicing, and drying. Stir-fried AR is prepared by lightly frying the slices with bran before they are used in formulations. Baked AR, also known as grilled AR, involves stir-frying the pieces with bran until they turn yellow, after which they are cooled and incorporated into medicinal applications (Wu M.-l. et al., 2024). Wine-processed AR is made by moistening the botanical drug with rice wine, followed by slicing and drying for medicinal use.

These traditional processing techniques for the botanical drug AR significantly influence the quality and therapeutic direction of the final product by carefully controlling the heating intensity and the use of auxiliary materials: raw AR retains abundant volatile oils, offering strong medicinal properties but also greater side effects due to alkaloids and other metabolites; roasting gently heats the botanical drug at low temperatures, promoting the release or transformation of some volatile oils, thereby reducing side effects and mildly enhancing its gastrointestinal therapeutic functions; plain stir-frying and bran-frying reduce toxicity and adverse effects while strengthening its therapeutic efficacy; wineprocessing using rice wine facilitates the extraction of active metabolites, increasing pharmacological activity. Essentially, these methods work by regulating the content and transformation of volatile oils and other metabolites to achieve reduced toxicity, preserved efficacy, enhanced potency, or altered therapeutic targeting (Feng et al., 2023; Li R.-l. et al., 2021).

4.4 Adverse reaction (toxicity)

Although AR has good medicinal value, excessive use may lead to adverse effects. Skin allergic reaction is a common side effect in the use of AR, due to various irritant alkaloids (saussureanine, costunolide, etc.), for people who are sensitive or have existing skin inflammation, contact with these substances is easy to cause allergic reactions. Acute generalized exanthematous pustulosis also could induced by AR (Chu et al., 2019). Additionally, high doses of TCM usually have liver toxicity, and AR is no exception. Oxidative stress should be the primary mechanism for the high-dose ARinduced hepatotoxicity, and Nrf2, HO-1 and NQO1 were the main targets (Song et al., 2024). On the contrary, the appropriate dosage of ethanol extract of AR had a protective effect on liver injury induced by lipopolysaccharide in rats (Song et al., 2023). Reasonable control of the dosage of AR is important for patients to avoid side effects, which can effectively ensure the safety and effectiveness of Chinese herbal medicine.

4.5 Commonly confused botanical drugs

In the field of traditional botanical drugs, species within the AR, Aristolochiae Radix (ARR), Vladimiriae Radix (VR), and Inulae Radix (IR), present persistent identification challenges due to morphological convergence and historical misclassification (Zhuang et al., 2021). These taxonomically related species exhibit overlapping botanical characteristics, particularly in root morphology and histological features, resulting in frequent substitution errors in herbal commerce. Current pharmacognostic authentication relies on UHPLC-QTOF-MS as the gold-standard

methodology for precise differentiation (Guccione et al., 2017; Shum et al., 2007; Wang et al., 2024). This analytical approach enables simultaneous detection of multiple marker metabolites, achieving clear discrimination between structurally analogous aristolochic acid derivatives and sesquiterpene lactone.

VR ("Chuanmuxiang" in China), primarily derived from the dried roots of Vladimiria souliei (Franch.) Ling or V. souliei (Franch.) Lingvar. cinerea Ling, predominantly cultivated in western Sichuan, China, exhibits medicinal properties akin to those of Muxiang, albeit with distinct emphases (China Pharmacopoeia Committee, 2020b). Chemical profiling differentiates AR from VR based on significant differences in their quantitative composition (dihydrodehydrocostus lactone, mokkolactone, α-costol, etc.) (Chen et al., 2020; Liang et al., 2024; Yan et al., 2020). The total lactone extract of VR has the similar protective effects on cholestatic liver injury as AR, even better in terms of anti-inflammatory properties (Chen Z. et al., 2022).

IR ("Tumuxiang" in China) refers to the dried roots of *Inula helenium* L. or *Inula racemosa* Hook.f. (China Pharmacopoeia Committee, 2020g). Historically utilized as a substitute for AR, IR exhibits significant chemical distinctions from AR, with its primary differential metabolites being alantolactone and isoalantolactone (Cai et al., 2021; Rasul et al., 2013). These metabolites have been used to emesis and diarrhoea, and to eliminate parasites (Xu et al., 2015). Alantolactone and isoalantolactone have been confirmed to possess potential hepatotoxicity, nephrotoxicity, and allergenic effects. Long-term or excessive intake may lead to symptoms such as loss of appetite, fatigue, nausea, vomiting, and pain in the liver area. In severe cases, it can cause irreversible liver damage. Due to their toxicity, IR has been rarely used in modern clinical practice of TCM.

ARR ("Qingmuxiang" in China) is the dried roots of Aristolochia debilis Sieb. et Zucc. or Aristolochia contorta Bge. ARR originally appeared as a substitute for AR, but contemporary researches have revealed the presence of aristolochic acid in Aristolochiae species, which can cause nephrotoxic adverse reactions (Cheng et al., 2006; Wang X. et al., 2020; Zhang et al., 2016). Consequently, ARR has been banned from use in TCM formulations (Luo et al., 2024).

In summary, careful identification of AR's species authenticity and quality is essential during procurement and use to prevent confusion with commonly confused botanical drugs. At the same time, it is recommended to use AR and its related botanical drugs under the guidance of a professional physician or pharmacist.

4.6 Botanical drugs with similar therapeutic properties

The commercially available herbal medicines exhibiting comparable qi-regulating, analgesic, spleen-strengthening, and digestion-promoting activities to AR primarily include *Pericarpium Citri Reticulatae*, *Aurantii Fructus*, *Cyperi Rhizoma*, and *Amomi Fructus*. However, their therapeutic emphases and clinical applications exhibit notably divergent profiles. Additionally, AR demonstrates multidimensional industrial advantages over other botanical drugs in its category.

TABLE 2 Main metabolites isolated from AR (Zheng et al., 2022; Zhuang et al., 2021).

Classification 1 Sesquiterpene $C_{15}H_{20}O_2$ $\alpha\text{-}Cyclocostunolide$ lactones eudesmanolides 2 $C_{15}H_{20}O_3$ Santamarine 3 C₁₅H₂₂O₃ 11α,13-Dihydrosantamarine 4 $C_{15}H_{20}O_2$ $\beta\text{-}Cyclocostunolide$ 5 $C_{15}H_{20}O_3$ Reynosin $C_{15}H_{22}O_3$ 11a,13-Dihydroreynosin 6 7 C₁₅H₂₀O₃ Magnolialide 8 C₁₅H₂₂O₃ 11α,13-Dihydromagnolialide 9 $C_{15}H_{22}O_3$ Arbusculin A 10 C₁₅H₂₂O₄ 1β-Hydroxyarbusculin A 11 $C_{15}H_{20}O_2$ Arbusculin B 12 $C_{15}H_{24}O_3$ Colartin $C_{15}H_{20}O_4$ $1\beta\text{-}Hydroxycolartin$ 13 14 C₁₅H₂₀O₂ Alantolactone 15 $C_{15}H_{20}O_2$ Isoalantolactone 16 $C_{15}H_{20}O_3$ Saussureal 17 Dihydro-α- $C_{15}H_{22}O_2$ cyclocostunolide 18 $C_{20}H_{28}NO_5$ Saussureamine D 19 $C_{20}H_{28}NO_5$ Saussureamine E 20 $C_{16} H_{22} O_6 S \\$ 13-Sulfodihydrosantamarine 21 $C_{16}H_{22}O_6S$ 13-Sulfo-dihydroreynosin 22 Sesquiterpene C₁₅H₁₈O₂ Dehydrocostus lactone guaianolides 23 $C_{15}H_{20}O_2$ Dihydrodehydrocostus lactone 24 4β-Methoxy- $C_{15}H_{20}O_3$ dehydrocostuslactone 25 C₁₅H₂₀O₃ 4α-Methoxydehydrocostuslactone 26 $C_{15}H_{20}O_3$ 10α-Methoxydehydrocostuslactone 11,13-Epoxy-27 $C_{15}H_{18}O_3$ dehydrocostuslactone 28 $C_{15}H_{16}O_4$ 11,13-Epoxy-3-ketodehydrocostuslactone 29 C₁₅H₂₀O₂ Mokko lactone 30 $C_{16}H_{22}O_3$ 13-Methoxy-dihydrodehy drocostuslactone 31 $C_{17}H_{22}O_4$ Lappalone

(Continued in next column)

TABLE 2 (Continued) Main metabolites isolated from AR (Zheng et al., 2022; Zhuang et al., 2021).

ZOZZ, Zildang	2022; Zhuang et al., 2021).			
Number	Classification	Molecular formula	Metabolites	
32		$C_{15}H_{18}O_3$	Zaluzanin C	
33		C ₁₇ H ₂₀ O ₄	Zaluzanin D	
34		C ₂₁ H ₃₀ O ₈	11β,13- Dihydroglucozalunin C	
35		C ₁₅ H ₁₈ O ₃	Isozaluzanin C	
36		C ₁₅ H ₂₀ O ₃	11β,13-dihydro-3- epizaluzanin C	
37		C ₁₅ H ₁₈ O ₄	11,13-Epoxyisozaluzanin C	
38		$C_{15}H_{18}O_2$	Isodehydrocostuslactone	
39		C ₁₅ H ₁₆ O ₃	Isodehydrocostuslactone- 15-aldehyde	
40		C ₁₅ H ₁₆ O ₃	Isodehydrocostuslactone- 14β- aldehyde	
41		$\mathrm{C}_{20}\mathrm{H}_{26}\mathrm{NO}_4$	Saussureamine B	
42		C ₁₉ H ₂₄ NO ₅	Saussureamine C	
43		C ₁₅ H ₂₀ O ₅ S	Sulfocostunolide A	
44		C ₁₈ H ₂₀ O ₆	Cynaropicrin	
45		C ₁₅ H ₂₀ O ₅ S	Sulfocostunolide B	
46		C ₂₁ H ₂₈ O ₈	3-O-β-D-Glucopyranoside- 1α ,3 α ,5 α ,7 α H-guaiane- $10(14)$,11 (13)-trien-6 α ,12- olide	
47	Sesquiterpene lactones - germacranolides	$C_{15}H_{20}O_2$	Costunolide	
48		C ₁₅ H ₂₂ O ₂	11α,13-Dihydrocostunolide	
49		C ₁₅ H ₂₂ O ₂	11β,13-Dihydrocostunolide	
50		C ₁₆ H ₂₄ O ₃	13-Methoxydihy drocostunolide	
51		C ₂₀ H ₂₈ NO ₄	Saussureamine A	
52		C ₂₁ H ₃₀ O ₈	Picriside B	
53		C ₁₅ H ₂₂ O ₂	Isodihydrocostunolide	
54		C ₁₅ H ₂₀ O ₃	Soulangianolide A	
55		C ₁₅ H ₂₀ O ₃	Parthenolide	
56		C ₂₀ H ₂₆ O ₆	Eupatoriopicrin	
57		C ₁₅ H ₂₂ O ₂	Saussurea lactone	
58		$C_{20}H_{30}O_{8}$	Saussurea lactone-10-O-β- D-glucoside	
59		C ₁₅ H ₂₀ O ₂	Dehydrosaussurea lactone	
60		C ₃₀ H ₃₈ O ₆	Lappadilactone	
61	Other sesquiterpenoids	C ₁₅ H ₂₄	α-Selinene	
62		C ₁₅ H ₂₄	β-Selinene	
63		C ₁₅ H ₂₄	γ-Selinene	
64		C ₁₅ H ₂₄ O	α-Costol	

TABLE 2 (*Continued*) Main metabolites isolated from AR (Zheng et al., 2022; Zhuang et al., 2021).

65 $C_{15}H_{24}O$ β-Costol 66 $C_{15}H_{24}O$ γ-Costol 67 $C_{15}H_{22}O$ $\alpha\text{-}Costal$ β-Costal 68 $C_{15}H_{22}O$ 69 $C_{15}H_{22}O$ γ-Costal 70 $C_{15}H_{22}O_2$ Costic acid $C_{15}H_{22}O_2$ Isocostic acid 71 72 4β-Hydroxy-11 (13)-C₁₅H₂₆O₂ eudesmane-12-al 73 C₁₅H₂₄O₂ 74 $C_{15}H_{22}O_3$ $5\alpha ext{-Hydroxy-costic}$ acid $\gamma\text{-}Eudesmol$ 75 $C_{15}H_{26}O$ 76 $C_{15}H_{26}O$ α-Eudesmol 77 C₁₅H₂₆O β-Eudesmol 78 $C_{15}H_{24}$ $\gamma\text{-}Muurolene$ 79 Eudesma-3,7 (11)-diene $C_{15}H_{24}$ 1β,6α-Dihydroxycostic acid 80 $C_{17}H_{26}O_4$ ethyl ester 81 $C_{15}H_{24}$ β-Maaliene 82 $C_{15}H_{24}$ (+)-Germacrene 83 $C_{15}H_{24}O$ Germacra-1(10),4,11(13)tiren-12-ol 84 Germacra-1(10),4,11(13)- $C_{15}H_{22}O$ tiren-12-al 85 $C_{15}H_{22}O_2$ Germacra-1(10),4,11(13)tiren-12-oic acid 86 $C_{15}H_{24}$ β-Elemene Elema-1,3,1 (13)-tiren-87 $C_{15}H_{24}O$ 12-ol 88 $C_{15}H_{22}O$ Elemenal 89 $C_{15}H_{28}O\\$ Elemol 90 $C_{15}H_{24}$ α-Humulene 91 β-Humulene $C_{15}H_{24}$ 92 $C_{15}H_{24}$ β -Caryophyllene 93 $C_{15}H_{24}O$ Epoxy-caryophyllene 94 $C_{15}H_{24}$ α-Cedrene 95 $C_{15}H_{24}$ β-Cedrene 96 $C_{15}H_{24}O$ Cedrenol 97 $C_{15}H_{24}$ $\alpha\text{-}Bergamotene$ 2,12-Bergamotadien-14-al 98 $C_{15}H_{22}O$ $C_{14}H_{24}O$ (E)-9-Isopropyl-6-methyl-

(Continued in next column)

TABLE 2 (Continued) Main metabolites isolated from AR (Zheng et al., 2022; Zhuang et al., 2021).

Number	Classification	Molecular formula	Metabolites
100		C ₁₅ H ₂₀	α-Calacorene
101		C ₁₅ H ₂₂	α-Curcumene
102		C ₁₅ H ₂₄	γ-Curcumene
103		C ₁₅ H ₂₄	α-Zingiberene
104		C ₁₅ H ₂₄	Valencene
105		C ₁₅ H ₂₄	β-Sesquiphellandrene
106		C ₁₅ H ₂₄ O	Glaucyl alcohol
107		C ₁₅ H ₂₄	β-Bergamotene
108		C ₁₅ H ₂₄	Longifolene
109		C ₁₅ H ₂₄	γ-Gurjunene
110		C ₁₅ H ₂₄	α-Gurjunene
111		C ₁₅ H ₂₄	Bisabolene
112		C ₁₅ H ₂₄	cis-α-Bisabolene
113		C ₁₅ H ₃₀ O	Nerolidol
114		C ₁₅ H ₂₆ O	Viridiflorol
115		C ₁₅ H ₂₆ O	Globulol
116		C ₁₅ H ₂₆ O	Ledol
117		C ₁₅ H ₂₄	α-Longipinene
118		C ₁₅ H ₂₄	β-Guaiene
119		C ₁₅ H ₂₄	α-Guaiene
120		C ₁₅ H ₂₂ O	Santalol
121		C ₁₅ H ₂₄ O	Aromadendrene epoxide
122		C ₁₅ H ₂₄	trans-β-Farnesene
123		C ₁₅ H ₂₄	α-Farnesene
124		C ₁₅ H ₂₄	β-Himachalene
125		C ₁₅ H ₂₆ O	Hedycaryol
126		C ₁₅ H ₂₄	Aromadendrene
127		C ₁₅ H ₂₂ O	Nootkatone
128		C ₁₅ H ₂₄ O	Thujopsanone
129		C ₁₅ H ₂₄ O	trans-α-Bergamotol
130		C ₁₅ H ₂₄ O	Santalcamphor
131		C ₁₅ H ₂₄ O	Longifolenaldehyde
132		C ₁₅ H ₂₄ O	Spathulenol
133		C ₁₅ H ₂₄	α-Copaene
134		C ₁₅ H ₂₄	α-Bulnesene
135		C ₁₅ H ₂₆ O	Valerianol
136		C ₁₅ H ₂₈ O ₂	Cryptomeridiol
137	Monoterpenoids	C ₁₅ H ₂₄ O	γ-Gurjunenepoxide-(2)
138		C ₁₀ H ₁₆	Camphene

TABLE 2 (*Continued*) Main metabolites isolated from AR (Zheng et al., 2022; Zhuang et al., 2021).

Phellandrene 139 $C_{10}H_{16}$ 140 $C_{13}H_{20}O$ α-Ionone 141 $C_{13}H_{20}O$ $\beta\text{-Ionone}$ 142 $C_{13}H_{22}O$ 3,4-Dihydro-α-ionone 143 $C_{13}H_{20}O_2$ (3R,6S)- α -Ionone-3-ol 144 $C_{13}H_{17}O_2$ α-Ionone-4-one 145 $C_{13}H_{20}O_2$ (3R,6R)-α-Ionone-3-ol 146 $C_{10}H_{16}$ Myrcene 147 $C_{10}H_{18}O$ Linalool 148 $C_{10}H_{16}$ $\alpha\text{-}Terpinene$ $C_{10}H_{16}$ $\beta\text{-}Terpinene$ 149 150 $C_{10}H_{18}O$ Isoborneol $C_{10}H_{18}O$ Borneol 152 $C_{10}H_{16}$ γ-Terpinene 153 α-Pinene $C_{10}H_{16}$ β-Pinene 154 $C_{10}H_{16}$ 155 $C_{10}H_{18}O$ 4-Terpineol 156 $C_{10}H_{18}O$ $\alpha\text{-}Terpine ol$ 157 $C_{13}H_{22}O\\$ a-Ionol 158 $C_{10}H_{16}$ Limonene 159 $C_{10}H_{16}$ Terpinolene 160 $C_{13}H_{22}O\\$ Geranylacetone 161 $C_{10}H_{20}O$ Menthol 162 $C_{10}H_{16}O$ Carvotanacetone 163 $C_{12}H_{22}O_2$ Linalyl acetate 164 $C_{10}H_{18}O$ Menthone 165 $C_{10}H_{16}O$ (+)-Camphor 166 Triterpenoids $C_{30}H_{50}O$ α-Amyrin 3β-Acetyl-9 (11)-167 $C_{34}H_{58}O_2$ baccharene Betulinic acid 168 $C_{30}H_{48}O_{3}$ Betulinol 169 $C_{30}H_{50}O_{2}$ 170 $C_{31}H_{50}O_{3}$ Betulinic acid methyl ester 171 $C_{30}H_{50}O$ Taraxsterol 172 $C_{30}H_{50}O$ 3-Filicanone 173 Phenylpropanoids C₁₇H₂₄O₉ Syringin 174 C₁₆H₂₂O₈ 4-Allyl-2,6-. dimethoxy phenol glucoside175 $C_{16}H_{18}O_9$ Chlorogenic acid 176 $C_{10}H_{12}O$ Anethole

(Continued in next column)

TABLE 2 (Continued) Main metabolites isolated from AR (Zheng et al., 2022; Zhuang et al., 2021).

2022; Zhuang	et al., 2021).		
Number	Classification	Molecular formula	Metabolites
177		C ₁₂ H ₁₄ O ₃	Eugenol acetate
178		C ₉ H ₈ O	Cinnamaldehyde
179		C ₁₀ H ₁₀ O ₂	Safrole
180		C ₁₀ H ₁₂ O	Estragole/4-Allylanisole
181		C ₁₂ H ₈ O ₄	Bergapten
182		C ₁₃ H ₁₄ O ₄	6,8-Dimethoxy-3,7- dimethylisocoumarin
183		$C_{26}H_{32}O_{12}$	1-Hydroxyrosinol-1-O-β- D-glucopyranoside
184		$C_{26}H_{34}O_{13}$	(–)-olivil-4"-O-β-D- glucopyranoside
185		$C_{22}H_{26}O_8$	Syringaresinol
186		$C_{20}H_{22}O_8$	Prinsepiol
187		C ₂₉ H ₃₆ O ₁₁	(+) -1-Hydroxypinoresinol- 4"-O-methyl ester-4'-β-D- glucopyranoside
188		$C_{28}H_{34}O_{10}$	(+)-1-Hypinoresinol-4"-Ο- β-D-glucopyranoside
189	Steroids	$C_{30}H_{46}O_{6}$	Lappalanasterol
190		C ₃₁ H ₅₀ O	3-Epi-lappasterol
191		C ₂₁ H ₃₂ O ₂	Pregnenolone
192		C ₃₅ H ₆₀ O ₆	Daucosterol
193		C ₂₉ H ₅₀ O	β-Sitosterol
194		C ₂₉ H ₄₈ O	Stigmasterol
195		C ₂₈ H ₄₈ O	Campesterol
196		C ₂₉ H ₅₀ O	γ-Sitosterol
197	Flavonoids	C ₂₉ H ₄₈ O ₂	Vlasoudiol
198		$C_{46}H_{60}O_{22}$	3'-(3 R-Acetoxy-5,5-dimethylcyclopent-1-ene)-4'-Omethylscutellarein-7-O-(6''')-Oacetyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -[α -L-rhamnopyranosyl- $(1 \rightarrow 2)$]- β -D-glucopyranoside
199		$C_{44}H_{58}O_{21}$	Kaempferol-3-O-β-Dglucopyransoyl- $(1 \rightarrow 4)$ -α-Lrhamnopyranosyl- $(1 \rightarrow 6)$ -β-D-galactopyranoside 7-O- $(6'''$ O-acetyl-β-Dglucopyranosyl- $(1 \rightarrow 3)$ - $[\alpha$ -L-rhamnopyranosyl $(1 \rightarrow 2)]$ -β-D-glucopyranoside
200		C ₄₇ H ₅₄ O ₂₀	Kaempferol-3-O-β-Dglucopyranosyl $(1 \rightarrow 2)$ -β-D-(6a'-Ocaffeoyl)-glucopyranoside 7-O-(6''' '-O-acetyl-β-Dglucopyranosyl- $(1 \rightarrow 3)$ - $[\alpha$ -L-rhamno-pyranosyl- $(1 \rightarrow 2)$]β-D-glucopyranoside
201		C ₄₉ H ₄₈ O ₁₅	

TABLE 2 (Continued) Main metabolites isolated from AR (Zheng et al., 2022; Zhuang et al., 2021).

Number	Classification	Molecular formula	Metabolites
			Kaempferol-3-O- α -L- (2a',3a'-E-di-p- coumaroyl)-rhamnoside 7- O-(α -O-acetyl- β - Dglucopyranosyl-($1 \rightarrow 3$)- [α -Lrhamnopyranosyl-($1 \rightarrow 2$)]- β -D- glucopyranoside
202		C ₁₀ H ₁₀ O ₄	5,7-Dihydroxy-2- methylchromone
203		C ₁₆ H ₁₄ O ₇	1-Hydroxy-2,3,4,7- tetramethoxyxanthone
204		C ₁₅ H ₁₂ O ₆	1,7-Dihydroxy-3,4- dimethoxyxanthon
205	Amino acid	C ₂ H ₅ NO ₂	Aspartic acid
206		C ₄ H ₇ NO ₄	Glycine
207		C ₄ H ₈ N ₂ O ₃	L-Asparagine
208		C ₄ H ₉ NO ₂	4-Aminobutyric acid
209		C ₄ H ₉ NO ₃	L-Threonine
210		C ₃ H ₇ NO ₃	Serine
211		C ₅ H ₉ NO ₄	Glutamic acid
212		C ₅ H ₁₀ N ₂ O ₃	Glutamine
213		C ₃ H ₇ NO ₂	Alanine
214		C ₆ H ₁₃ N ₃ O ₃	Citrulline
215		C ₅ H ₁₁ NO ₂	Valine
216		C ₆ H ₁₂ N ₂ O ₄ S ₂	Cystine
217		C ₆ H ₁₃ NO ₂	Isoleucine
218		C ₆ H ₁₃ NO ₂	Leucine
219		C ₉ H ₁₁ NO ₃	Tyrosine
220		C ₉ H ₁₁ NO ₂	Phenylalanine
221		C ₅ H ₁₂ N ₂ O ₂	Ornithine
222		C ₆ H ₁₄ N ₂ O ₂	Lysine
223		C ₆ H ₉ N ₃ O ₂	Histidine
224		C ₆ H ₁₄ N ₄ O ₂	Arginine
225		C ₂₁ H ₂₀ O ₁₀	Aloe-emodin-8-O-β-D- glucopyranoside
226		C ₂₁ H ₁₈ O ₁₁	Rhein-8-O-β-D- glucopyranoside
227		C ₁₅ H ₁₀ O ₄	Chrysophanic acid
228		C ₁₀ H ₂₀ O ₆	N-Butyl-β-D-fructoside
229		C ₇ H ₁₄ O ₆	Methyl-α-D- frutofuranoside
230		C ₁₂ H ₁₆ O ₆	Phenyl-β-D- glucopyranoside
231		C ₁₃ H ₁₈ O ₆	Benzyl-β-D- glucopyranoside

(Continued in next column)

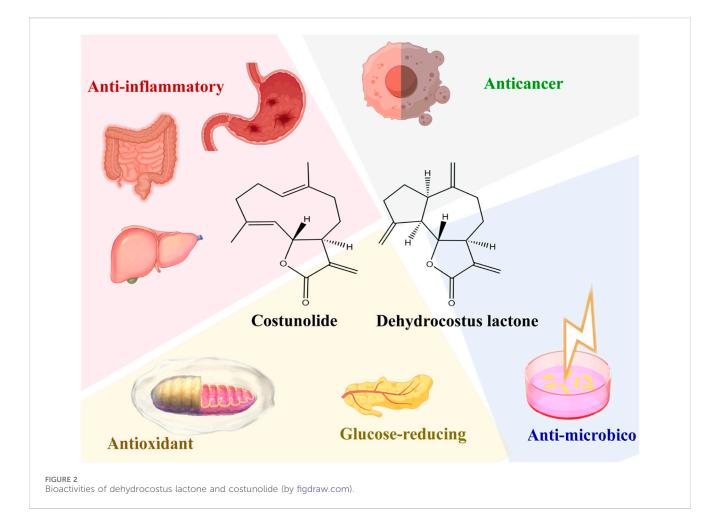
TABLE 2 (Continued) Main metabolites isolated from AR (Zheng et al., 2022; Zhuang et al., 2021).

Number	Classification	Molecular formula	Metabolites
232		C ₁₉ H ₃₂ O ₈	Ascleposide E
233		C ₁₉ H ₁₂ O ₈	β-D-Frutofuranose
234		C ₆ H ₁₂ O ₆	Glucose
235		C ₈ H ₈ O ₃	Vanillin
236		C ₆ H ₆ O ₃	5-Hydroxymethyl- furaldehyde
237		C ₉ H ₁₀ O ₄	3,5-dimethoxy-4-hydroxy- benzaldehyde
238		C ₂₁ H ₃₈ O ₄	Monolinolein
239		C ₄ H ₆ O ₄	Succinic acid
240		C ₁₇ H ₃₂ O	Shikokiol A
241		C ₁₇ H ₃₂ O	Shikokiol B
242		C ₁₇ H ₃₂ O	Shikokiol C
243		C ₇ H ₆ O ₂	p-Hydroxybenzaldehyde
244		C ₉ H ₁₀ O ₄	3,5-Dimethoxy-4- hydroxyacetophenone
245		C ₁₆ H ₃₂ O ₂	Palmitic acid
246		C ₁₀ H ₁₄	p-Cymene
247		C ₁₉ H ₃₄ O ₂	(Z, Z)-9,12- Octadecadienoic acid
248		C ₂₁ H ₃₈ O ₄	(Z, Z)-9,12- Octadecadienoic acid-2- hydroxy-1,3propamedinyl ester

Specifically, the flavonoid metabolites (hesperidin, nobiletin, tangeretin, etc.) in Pericarpium Citri Reticulatae are susceptible to rapid degradation under UV light, resulting in poor stability (Ho and Kuo, 2014; Li Y. et al., 2024). Aurantii Fructus contains alkaloids such as p-tyramine, N-methyltyramine, and tyramine that may cause cardiovascular adverse reactions including hypertension and arrhythmia, thus requiring careful evaluation of the patient's baseline condition in clinical use (Gao et al., 2020). Cyperus volatile oils, rich in α -cyperone, cyperotundone, and limonene, face industrial scalability challenges due to low extraction yields (Bezerra et al., 2025). Amomi Fructus is constrained by its reliance on specific cultivation conditions and high production costs (Suo et al., 2018). In contrast, AR not only has a mild nature and comprehensive effects, but also stands out in terms of the stability of its metabolites, safety, and the maturity of industrialization. Therefore, it is more suitable for large-scale production, pharmaceutical food processing, development of health products.

5 Phytochemistry

Currently, more than 200 metabolites (Table 2) have been isolated and identified from ALD (Huang et al., 2021; Zheng



et al., 2022). These metabolites can be categorized by structural type into sesquiterpene lactones (eudesmanolides, guaianolides, germacranolides, etc.), monoterpenoids, triterpenoids, phenylpropanoids, steroids, flavonoids, amino acids, and other metabolites (Huang et al., 2021; Seo et al., 2015; Wang Y. et al., 2020; Zheng et al., 2022). These metabolites were primarily isolated and identified using techniques such as thin-layer chromatography (TLC), nuclear magnetic resonance (NMR), liquid chromatography (LC), gas chromatography (GC), mass spectrometry (MS), and electronic nose (e-nose) technology.

5.1 Sesquiterpene lactones

The sesquiterpene lactones are the primary and characteristic metabolites of ALD, exhibiting a diverse and abundant range exceeding 130 species (Yuan et al., 2024; Zheng et al., 2022). Sesquiterpene lactones are a class of natural metabolites primarily found in plants of the Asteraceae family, characterized by a 15-carbon sesquiterpenoid backbone coupled with a lactone ring (Amen et al., 2025; Liu et al., 2021). They often serve as defensive metabolites in plants and are largely responsible for their characteristic bitter taste. These metabolites are renowned for their diverse and potent biological activities, including anti-inflammatory, anti-tumor,

antimicrobial, and immunomodulatory effects (Amen et al., 2025). As a result, they represent not only key active metabolites in traditional herbal medicine but also important lead metabolites in modern drug development. However, their bioactivity is dual-edged: some members are strong allergens capable of inducing contact dermatitis and may exhibit cytotoxicity at higher concentrations (da Silva et al., 2021).

Among the sesquiterpene lactones, costunolide and dehydrocostus lactone are the key substances for quality control of AR (Figure 2) (Dong et al., 2018). The total content of costunolide and dehydrocostus lactone in AR must not be less than 1.8%, as stipulated by the Chinese Pharmacopoeia (2020 Edition). They are also the most important active substances in AR and could be quantified synchronously using high-performance liquid chromatography coupled with mass spectrometry (Seo and Shin, 2015; Zhang et al., 2014). Currently, costunolide is already available in substantial quantities through genetic engineering techniques. The biosynthetic pathway for costunolide (Figure 3) has been successfully built in Escherichia coli by the co-expression of three genes (GAS, GAO, COS) involved in costunolide biosynthesis, along with eight genes responsible for converting acetyl-CoA into farnesyl diphosphate via the mevalonate pathway. And costunolide yield was up to 100 mg L⁻¹ in E. coli (Yin et al., 2015). Meanwhile, the co-expression of GAS, GAO, and COS in

yeast and *Nicotiana benthamiana* leaves has also facilitated costunolide production (Blazquez et al., 2011). Revealing the synthetic pathway of costunolide indicates that there are no obstacles at all in constructing transgenic plants of ALD that produce high yields of costunolide. However, despite the structural similarities between dehydrocostus lactone and costunolide, and their concurrent presence in plants, the biosynthetic pathway for dehydrocostus lactone in plants remains unidentified. Consequently, the production of dehydrocostus lactone still relies on the extraction from plant raw materials.

5.2 Monoterpenoids and triterpenoids

Monoterpenoids and triterpenoids are two important classes of natural terpenoid metabolites, each with distinct characteristics in structure, distribution, and function (Yang et al., 2025). Monoterpenoids, composed of two isoprene units (C10), are major metabolites of plant essential oils (Bhatti et al., 2014; Jiang and Wang, 2023). In contrast, triterpenoids consist of six isoprene units (C30), featuring higher molecular weight and greater structural complexity (Huang et al., 2024; Zeng et al., 2024). Monoterpenoids and triterpenoids are widely used in the flavor, food, and cosmetics industries. Their biological effects tend to be more profound and systemic, including notable anti-inflammatory, antitumor, immunomodulatory, and cholesterol-lowering activities (Yang et al., 2025). Representative metabolites such as α-amyrin and betulinic acid are core active metabolites in many traditional Chinese medicines. However, some triterpenoids may exhibit hepatotoxicity at high doses.

5.3 Phenylpropanoids

Phenylpropanoids are an important class of plant secondary metabolites characterized by a fundamental C6–C3 skeleton, which consists of a benzene ring (C6) attached to a propene group (C3), also known as the phenylpropane backbone (Vogt, 2010). These

metabolites are primarily synthesized through the shikimate pathway and play crucial roles in plant defense, growth development, and signal transduction (Cao et al., 2025; Zhang et al., 2025). Additionally, they serve as significant sources for numerous pharmaceuticals, flavoring agents, and industrial raw materials.

5.4 Steroids

Steroids represent a significant class of bioactive metabolites in TCM. Phytosterols (β -sitosterol, lappalanasterol, pregnenolone, etc.) are the predominant type in ALD. Structurally, phytosterols resemble cholesterol in animals, sharing the same cyclopentanoperhydrophenanthrene core skeleton, but differ in their side chain configurations. These steroidal metabolites exhibit diverse structures and broad biological functions, serving not only as a fundamental chemical basis for explaining the efficacy and mechanisms of TCM, but also as key resources for modern drug (steroid hormones) development (Ferreira-Guerra et al., 2020).

5.5 Flavonoids

Flavonoids are ubiquitous in traditional Chinese medicine and are one of the key metabolites that enable many medicinal materials to exert their therapeutic effects. Flavonoids are distinguished by their broad spectrum of biological activities, including antioxidant effects, cardiovascular protection, and antiplatelet aggregation, which collectively contribute to reducing the risk of cardiovascular diseases and enhancing blood circulation (Jomova et al., 2025; Xu et al., 2025).

5.6 Others

In addition to the previously mentioned metabolites, ALD has been identified to contain polysaccharides, higher fatty acids, small aliphatic alcohols, aldehydes, acids, amino acids, and cholamine, among other metabolites (Peng et al., 2025; Zhuang et al., 2021). The

TABLE 3 The main pharmacological properties of ALD.

Pharmacological activities	Main findings	References
Antioxidant	The preadministration of ALD extract exerted its protective effect of Th-induced adult male rats mainly through potentiating the antioxidant defense system by decreased lipid peroxidation and NO and increase the glutathione content.	Abdel-Rahman et al. (2020)
	The aqueous solvents of ALD were found superior in their ability to extract the antioxidants and aqueous ethanol was reported more efficient than aqueous methanol.	Ahmed et al. (2016)
	Alcoholic extract of AR (including alkaloids, terpenoid, phenols and others) have comparable antioxidant activity to ascorbic acid (81.96%).	Adel et al. (2025)
	The extract of ALD showed high levels of total phenolic content (188.2 \pm 2.1 mg GAE/g DM) and total flavonoid content (129 \pm 2.6 mg QE/g DM). In antioxidant tests, the extract exhibited strong activity, with the IC50 values of 137.15 µg/mL for ABTS and 175.5 µg/mL for DPPH.	Binobead et al. (2024)
	α,β -Unsaturated carbonyl metabolites (costunolide, dehydrocostus lactone, artemisitene, santamarine, isoalantolactone) were mainly featured as the antioxidant active metabolites of AR.	Wu et al. (2025)
	The 70% ethanol extract of ALD has a higher concentration of total phenolic content, total flavonoids, and antioxidant effect than the 70% methanol and water extracts. Rats pretreated with ALD extracts (70% methanol, 300 mg/kg BW) reduced the harmful effects induced by NaNO ₂ and improved the hematological parameters, liver, and kidney function biomarkers as well as lipid profile as compared to the NaNO ₂ group (75 mg/kg BW, single oral dose for 4 weeks).	Elshaer et al. (2022)
Anti-inflammatory	Dehydrocostuslactone exerts potent anti-hepatocellular carcinoma effects by inducing ER stress-mediated apoptosis via the MAPK pathway, resulting in a 50% reduction in tumor volume <i>in vivo</i> after 45 days of treatment.	Hsu et al. (2009)
	AR alleviates ulcerative colitis by targeting PKM2 to inhibit the NF- κB and NLRP3 pathways, thereby reducing inflammation and modulating immune responses.	Feng et al. (2024)
	Dehydrocostus lactone alleviates irinotecan (CPT-11)-induced intestinal mucositis by inhibiting the TLR4/MD2 complex and suppressing the NF-κB/NLRP3 signaling pathway, without compromising the antitumor efficacy of CPT-11.	Sun et al. (2024)
	The sesquiterpene lactone-rich fraction of ALD alleviates ulcerative colitis by regulating the Nrf2-Hmox-1, NF- κ B, and MAPK pathways.	Chen et al. (2022a)
	ALD demonstrates potential in preventing and treating benign prostatic hyperplasia (BPH) by modulating apoptosis and inflammation, as evidenced by reduced prostate weight, improved apoptotic protein expression, and decreased inflammatory cytokines in a testosterone-induced BPH rat model.	Choi et al. (2021)
	The combined topical application of ALD and <i>Thuja orientalis</i> extracts demonstrates synergistic efficacy in alleviating atopic dermatitis symptoms by reducing pro-inflammatory activity and immune hyperresponsiveness, outperforming either extract alone.	Yang et al. (2017)
	$\alpha_i\beta\text{-Unsaturated carbonyl metabolites}$ are as the key anti-inflammatory and antioxidant metabolites in ALD.	Wu et al. (2025)
	The sesquiterpenoids isolated from ALD, exhibit significant anti-inflammatory activity by inhibiting NO production in LPS-stimulated macrophages at 20 $\mu M.$	Lyu et al. (2023)
	In a DSS-induced murine ulcerative colitis UC model, daily gavage with dehydrocostus lactone at 20, 15, and 10 mg/kg/d from day 4–17 significantly reduced inflammation and enhanced barrier function by suppressing the IL-6/STAT3 pathway.	Zhou et al. (2020)

TABLE 3 (Continued) The main pharmacological properties of ALD.

Pharmacological activities	Main findings	References
	Dehydrocostus lactone alleviates atherosclerosis by promoting cholesterol efflux and inhibiting inflammation via the TLR2/PPAR- γ /NF- κ B signaling pathway in both <i>in vivo</i> and <i>in vitro</i> models.	Hong et al. (2025)
	Alantolactone exerts anti-inflammatory effects in LPS-stimulated macrophages by suppressing NF-κB activation and MAPK phosphorylation via downregulation of the MyD88-dependent signaling pathway.	Chun et al. (2012)
	Owing to the multi-target nature of IBD, the natural formulation KM1608 demonstrates potential therapeutic value by ameliorating colitis symptoms and distributing effectively in the intestinal tract.	Lee et al. (2020)
	Epoxymicheliolide alleviates ulcerative colitis by covalently targeting TAK1 and Keap1 to inhibit NF-κB-mediated inflammation and activate the Nrf2 antioxidant pathway.	He et al. (2022)
	Dehydrocostus lactone significantly ameliorated DSS-induced colitis in mice at doses of 5–15 mg/kg by covalently targeting both IKK α / β and Keap1, thereby suppressing NF- κ B signaling and activating the Nrf2 pathway.	Yuan et al. (2022)
	ALD demonstrates potential as a therapeutic agent for osteoarthritis by exhibiting analgesic and anti-inflammatory effects in both <i>in vivo</i> and <i>in vitro</i> models.	Jo et al. (2021)
	The ethanol extract of ALD demonstrates anti-inflammatory effects by suppressing NF- κ B and MAPK pathways and antioxidant activity through activation of the Nrf2/HO-1 pathway in LPS-stimulated RAW 264.7 cells.	Lim et al. (2020)
Anti-cancer effect	For the ethyl acetate extract of AR (including arbusculin B, α -cyclocostunolide, costunolide, and dehydrocostus lactone), cytotoxic IC $_{\rm SOS}$ of rat skeletal myoblast (L6 cells) from were from 1.6 to 19 μM , and selectivity indices from 0.5 to 6.5.	Julianti et al. (2011)
	The supercritical fluid extraction of oils from ALD obtained at 10 MPa exhibited the strongest antitumor efficacy with IC $_{50}$ values of approximately 0.44, 0.46, and 0.74 µg/mL on HCT, MCF-7, and HepG-2 cells, respectively, whereas those at 20 MPa showed higher IC $_{50}$ values (2.33, 6.59, 19.0 µg/mL), followed by 48 MPa (36.02, 59.5, 96.9 µg/mL).	Ahmed et al. (2022)
	Lyophilized ALD selectively inhibits the growth of breast and cervical cancer cells by inducing alternative apoptotic pathways.	Hasson et al. (2018)
	AR extract demonstrated significant antiproliferative and apoptotic effects, inducing caspase-3/7 and annexin V/PI activity, on MCF-7 breast cancer cells at concentrations of 20–200 $\mu g/mL$ over 24–72 h, supporting its potential as a low-toxicity therapeutic candidate.	Kumar et al. (2024b)
	Stigmasterol, isoboldine, and β-sitosterol could target key prostate cancer-related hub genes (e.g., SRC, FGFR1, HSP90AA1); stigmasterol showed the strongest binding to HSP90AA1, and pathway analysis highlighted involvement of PI3K/AKT signaling.	Kosanam et al. (2024)
	A study on 72 mice with PVP-induced cancer demonstrated that AR extract exhibited anticancer effects by reducing cell proliferation and modulating liver enzyme levels (ALT: 29.01 \pm 1.8, AST: 87.55 \pm 2.9, ALP: 98.12 \pm 8.8 U/L in controls), with dosedependent tissue regeneration observed across treatment groups.	Al-Zayadi et al. (2023)
	AR extract at 500 mg/kg body weight demonstrated the highest anti-neoplastic efficacy in a DMBA-induced rat mammary tumour model, significantly reducing tumour progression, oxidative stress, pro-inflammatory cytokines (TNF- α and NF- κ B), and expression of Ki-67, MMP-9, and VEGF markers.	Kumar et al. (2024a)
	AR extracts and isolated sesquiterpene lactones (particularly isoalantolactone, alantolactone, β -cyclocostunolide, and α -cyclocostunolide) exhibited significant cytotoxicity against A549 and C-6 cancer cells.	Kumar et al. (2014)

TABLE 3 (Continued) The main pharmacological properties of ALD.

Pharmacological activities	Main findings	References
	The hexane and chloroform fractions of ALD exhibited potent anticancer activity against the PC-3 prostate cancer cell line, with IC ₅₀ values of 3.37 \pm 0.14 µg/mL and 7.53 \pm 0.18 µg/mL, respectively.	Bhushan et al. (2023b)
	ALD extracts demonstrated potent anticancer activity by inducing G1 phase arrest and intrinsic apoptosis via the mitochondrial pathway in breast, liver, and colon cancer cells, with IC $_{50}$ values as low as 0.25–2.5 µg/mL for the most active extracts.	Shati et al. (2020)
	Three new eudesmane-type sesquiterpene lactone galactosides, costunosides A–C, were isolated from ALD and identified as the first natural β -galactopyranoside-containing eudesmane glycosides, among which metabolite 3 exhibited cytotoxic activity against several human cancer cell lines with IC50 values ranging from 3.4 to 9.3 μM .	Bhushan et al. (2023a)
	Dehydrocostus lactone demonstrates anti-angiogenic activity by inducing G0/G1 cell cycle arrest via inhibition of the Akt/GSK-3β/cyclin D1 and mTOR pathways, as evidenced in both <i>in vitro</i> and nude mice models.	Ahmad et al. (2012)
	Dehydrocostus lactone from ALD inhibits viability, migration, and proliferation of laryngeal carcinoma cells (Hep-2 and TU212) with low toxicity to normal HBE cells, and suppresses tumor growth <i>in vivo</i> by inducing mitochondrial apoptosis via inhibiting PI3K/Akt/Bad and activating ER stress pathways.	Zhang et al. (2020)
Organ protection	AR extract improved lung injury by reducing iNOS, caspase-3, and microRNA-let-7a, while boosting HO-1.	Attallah et al. (2023)
Anti-diabetic effect	Two proteinaceous amylase inhibitors, ScAI-R (IC50 = 23 $\mu g/mL$, Ki = 0.38 μM) and ScAI-L (IC50 = 28 $\mu g/mL$, Ki = 0.32 μM), which were from ALD, displayed the high affinities towards human salivary and pancreatic α -amylases (up to 90% inhibitory activity).	Ben Abdelmalek et al. (2025)
	The administration of extracts of ALD into Streptozotocin-treated rats separately resulted in a decline in the elevated levels of blood glucose, total cholesterol, triglycerides and improving serum HDL-Cholesterol and body weight.	Gomaa et al. (2021)
	All streptozotocin-treated diabetic rats received the treatments of ALD- extracts (200–400 mg/kg bwt., with isochlorogenic acid A (8393.64 μ g/g) and chlorogenic acid (6,532.65 μ g/g)) orally for 21 days consecutively, the administration significantly mitigated diabetic hyperglycemia.	Abouelwafa et al. (2024)
	The casein encapsulated extract of ALD demonstrated anti-diabetic potential through inhibition of $\alpha\text{-amylase}$ and $\alpha\text{-glucosidase}$ activities and enhanced glucose uptake in HepG2 cells.	Mushtaq et al. (2025)
Antiparasitic effect	The ethyl acetate extract of AR (including arbusculin B, α -cyclocostunolide, costunolide, and dehydrocostus lactone) potently inhibited the growth of <i>Trypanosoma brucei rhodesiense</i> , with ICsos between 0.8 and 22 μ M.	Julianti et al. (2011)
	<i>Trichinella spiralis</i> experimental infection induced DNA damage and oxidative stress in rat skeletal muscles and treatments with AR extract modulates these changes.	Alghabban et al. (2024)
	In vitro, extract of AR showed a significant anthelmintic effect on live adult Ascaridia.galli worms in terms of inhibition of worm motility, with worm motility inhibition of 100% at 24 h post-exposure at the 100 mg/mL.	Mir et al. (2024)
Antimicrobial effect	The highest activity with the lowest MIC value was recorded as 3.12 μ L/mL for the essential oil of ALD (against <i>S. epidermidis</i> and <i>C. albicans</i>), 3.12 mg/mL for the methanolic extract (against <i>S. aureus</i>), and 6.25 mg/mL for both hexane-chloroform and aqueous extracts (against <i>S. aureus</i>).	Ahmed and Coskun (2023)
	The smoke of ALD successfully inhibited <i>P. crustosum</i> growth of Fresh walnuts.	Qiao et al. (2023)

TABLE 3 (Continued) The main pharmacological properties of ALD.

Pharmacological activities	Main findings	References
	Ethanol extracts of AR had inhibitory effects on common plant-pathogenic fungi, with EC50 (concentration for 50% of maximal effect) values ranging from 114.18 mg/L to 414.08 mg/L.	Cai et al. (2022)
	The highest minimum inhibitory concentration was seen in the ethanolic extract of ALD, with an MIC of 50 mg/mL for <i>S. aureus</i> followed by an MIC of 200 mg/mL for <i>K. pneumoniae</i> . It showed a MBC against <i>S. aureus</i> and <i>K. pneumoniae</i> (>50 mg/mL and >200 mg/mL, respectively).	(Mammate et al., 2023; Mishra et al., 2020)
	The acetic acid extract of ALD exhibited significant antimicrobial activity (<i>in vitro</i>) against <i>C. albicans</i> (MIC = 6.25 mg/mL, MFC = 12.5 mg/mL), followed by <i>B. cereus</i> , <i>S. enterica</i> , <i>S. aureus</i> , <i>E. coli</i> , and <i>P. aeruginosa</i> , respectively (MIC = 25 mg/mL, MBC/MFC = 25–50 mg/mL).	Idriss et al. (2023)
	The inhibitory activity of all ALD extracts at three different extraction pressure levels (10, 20, 48 MPa), was higher than gentamicin against all tested bacteria (<i>B. subtilis</i> , and <i>S. aureus</i> , <i>P. aeruginosa</i> , <i>E. coli</i> , <i>K. pneumonia</i> , <i>C. albicans</i> , <i>C. tropicalis</i> , <i>A. flavus</i> , <i>F. oxysporium</i>).	Ahmed et al. (2022)
Antiviral effect	The antiviral activity of ALD acetic acid extract had a significant positive influence against HSV-1 (EC50 = 82.6 g/mL; CC50 = 162.9 g/mL; selectivity index = 1.9). No effect was detected in terms of the inhibition of SARS-CoV2 entry.	Idriss et al. (2023)
	Some sesquiterpenoids isolated from ALD demonstrate anti-HBV activity by inhibiting HBsAg secretion.	Li et al. (2024a)
	The value of the <i>in vitro</i> IC50 of AR extract against low pathogenic human coronavirus (HCoV-229E) and human influenza virus (H1N1) influenza virus were 23.21 \pm 1.1 and 47.6 \pm 2.3 μ g/mL, respectively.	Attallah et al. (2023)
	The ALD phytoconstituents have inhibitory potential against the receptor-binding domain of the spike glycoprotein and the main protease of the SARS-CoV-2 Delta (B.1.617.2) variant of the novel coronavirus via molecular docking, DFT, and ADME/Tox studies.	Houchi and Messasma (2022)
	The bioactive molecules from ALD can be as SARS-CoV-2 main protease inhibitors by computational approach investigation.	Hajji et al. (2022)

presence of these metabolites in AR may provide a pharmacological basis for the utilization of this herbal medicine in the treatment of constipation, intestinal infections, and associated inflammatory diseases. Higher fatty acids, amino acids, and cholamine are essential for maintaining normal physiological functions of the body (Chen and Fang, 2025; Kelling et al., 2024; Wu et al., 2022). Meanwhile, although the specific mechanisms of action of aliphatic alcohols, aldehydes, and acids require further investigation, they may play significant roles in regulating metabolism and participating in immune responses.

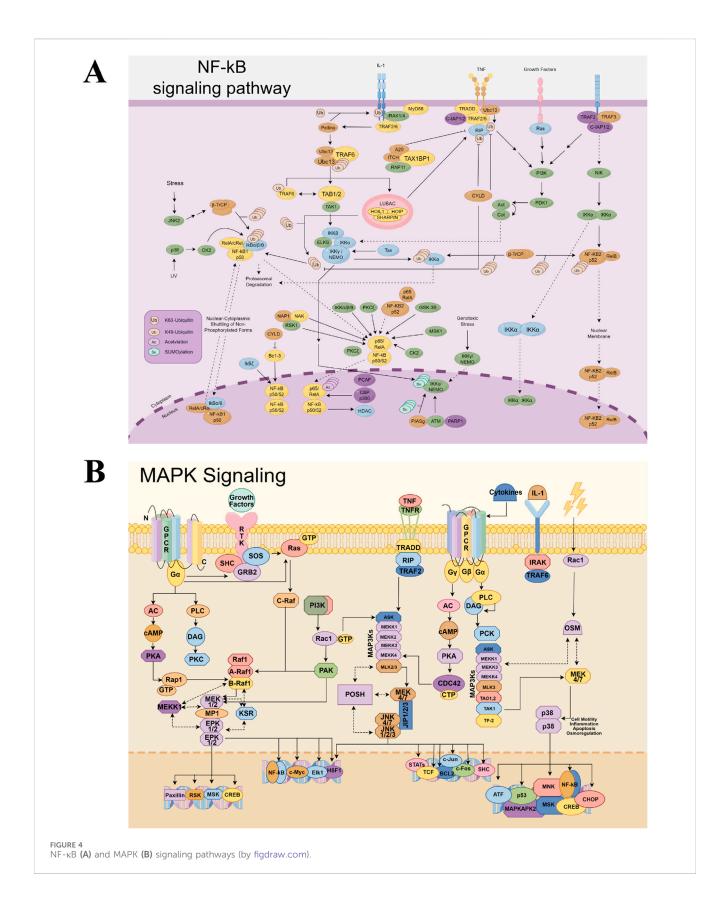
6 Pharmacology

The pharmacological effects of AR are diverse and significant (Table 3).

6.1 Antioxidant

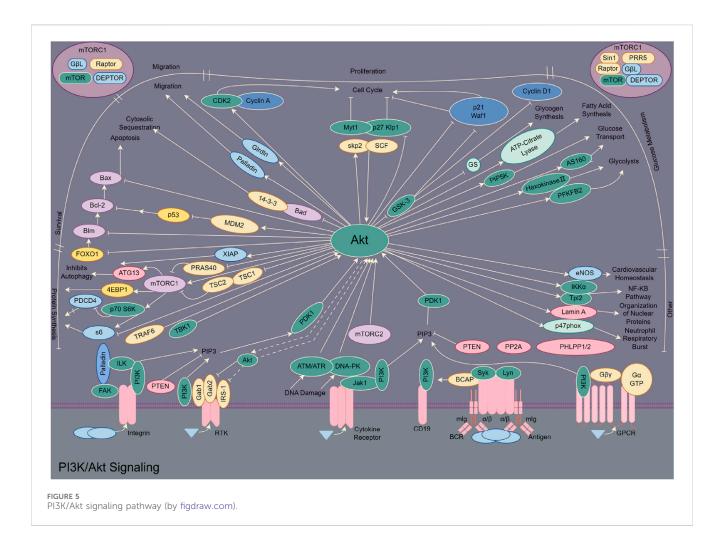
The antioxidant profile of ALD multifaceted, stemming from its rich and diverse phytochemical composition. The high values

for total phenolic (188.2 ± 2.1 mg GAE/g DM) and flavonoid $(129 \pm 2.6 \text{ mg QE/g DM})$ content are paramount (Binobead et al., 2024). Phenolics and flavonoids are renowned antioxidants due to their hydrogen-donating ability, which neutralizes free radicals by stabilizing them. The in vitro assays (DPPH and ABTS) confirm this radical-scavenging capability. The IC₅₀ values (137.15 µg/mL for ABTS, 175.5 µg/mL for DPPH) represent the concentration required to scavenge 50% of the radicals (Binobead et al., 2024). Lower IC₅₀ values indicate higher potency. The extract of AR have comparable antioxidant activity to ascorbic acid (Adel et al., 2025). α,β-Unsaturated carbonyl metabolites (costunolide, dehydrocostus lactone, artemisitene, santamarine, isoalantolactone) were mainly featured as the antioxidant active metabolites of AR (Wu et al., 2025). This molecular structure is a key pharmacophore for antioxidant activity. It can quench free radicals through direct singleelectron transfer. Ultimately, the most significant evidence of its antioxidant power is its in vivo protective effect against NaNO2-induced toxicity. NaNO2 is a potent oxidant that causes methemoglobinemia and oxidative damage to organs. The extract's ability to improve hematological parameters and protect liver and kidney function biomarkers demonstrates that



its antioxidants are bioavailable and active within a living system, effectively mitigating systemic oxidative stress (Elshaer et al., 2022). This bridges the gap from laboratory findings to potential

therapeutic applications, suggesting ALD could be developed into a natural remedy for conditions where oxidative stress is a key pathological factor.



6.2 Anti-inflammatory

The extract of ALD and its isolated bioactive metabolites, particularly sesquiterpene lactones such as dehydrocostus lactone, alantolactone, and epoxymicheliolide, demonstrate significant antiinflammatory activity across multiple experimental models. In LPSstimulated RAW 264.7 macrophages and peritoneal macrophages, ALD and these metabolites consistently suppress the production of key proinflammatory mediators, including NO, PGE2, iNOS, COX-2, and the cytokines IL-6, IL-1β, and TNF-α (He et al., 2022; Lim et al., 2020). This broad inhibition of inflammatory outputs is not limited to macrophage models but extends to in vivo conditions such as osteoarthritis (Jo et al., 2021), where ALD reduces pain and serum IL-1β, and to inflammatory bowel disease (IBD) models (Lee et al., 2020), including dextran sulfate sodium (DSS)-induced colitis (Yuan et al., 2022), where it ameliorates symptoms, protects the colonic barrier, and lowers inflammatory cytokines and enzymes like myeloperoxidase (MPO).

The fundamental mechanism underlying this robust antiinflammatory effect involves the dual suppression of the NF- κ B and MAPK signaling pathways (Figure 4) (Lim et al., 2020). ALD and its metabolites inhibit the LPS-induced phosphorylation and degradation of I κ B α , thereby preventing the nuclear translocation of the NF- κ B subunits p65 and p50 (Chun et al., 2012; Lim et al., 2020). Concurrently, these metabolites inhibit the phosphorylation of MAPKs, including JNK, ERK, and p38 (Chun et al., 2012; Hsu et al., 2009; Lim et al., 2020). Further upstream, alantolactone was shown to suppress the expression of adaptor proteins MyD88 and TIRAP, which are critical for initiating these cascades (Chun et al., Importantly, this anti-inflammatory activity complemented by a potent antioxidative effect mediated through the activation of the Nrf2/HO-1 pathway. Metabolites like dehydrocostus lactone and epoxymicheliolide enhance the nuclear accumulation of Nrf2, leading to the upregulation of antioxidant genes and a reduction in intracellular ROS (Yuan et al., 2022). Mechanistic studies reveal that many of these actions depend on the presence of an α,β -unsaturated carbonyl group, which allows the metabolites to form covalent bonds with key cysteine residues on target proteins like IKKα/β, Keap1, or TAK1 (He et al., 2022; Wu et al., 2025; Yuan et al., 2022).

The therapeutic implications of these mechanisms are vast, as evidenced by efficacy in diverse inflammatory disease models. Beyond colitis and osteoarthritis, ALD and dehydrocostus lactone alleviate atherosclerosis by promoting cholesterol efflux in macrophage-derived foam cells and modulating the TLR2/PPAR- γ /NF- κ B pathway (Hong et al., 2025). They also show protective effects against irinotecan-induced intestinal mucositis by inhibiting the TLR4/NF- κ B/NLRP3 axis and against benign prostatic

hyperplasia by restoring apoptosis balance (Choi et al., 2021; Sun et al., 2024). Furthermore, a sesquiterpene lactone-rich fraction of ALD was significantly more effective than an aqueous extract in treating ulcerative colitis (Chen Y. et al., 2022), underscoring that these specific metabolites are the primary active anti-inflammatory agents. This multifaceted, multitargeted action, targeting both inflammation and oxidative stress, positions ALD and its metabolites as promising candidates for treating complex chronic inflammatory disorders.

6.3 Anti-cancer effect

The supercritical fluid extraction of oils from ALD demonstrates remarkable and concentration-dependent cytotoxic efficacy against various cancer cell lines. Specifically, the extract obtained at 10 MPa exhibited potent antitumor activity with IC50 values of approximately 0.44, 0.46, and 0.74 µg/mL against HCT-116, MCF-7, and HepG-2 cells, respectively, whereas extracts obtained at higher pressures (20 MPa and 48 MPa) showed significantly reduced potency, highlighting the critical influence of extraction parameters on bioactive metabolite efficacy (Ahmed et al., 2022). Furthermore, specific sesquiterpene lactones like costunolide, dehydrocostus lactone, alantolactone, and isoalantolactone, isolated from the roots, have demonstrated significant activity against diverse cancer types including lung (A549), glioma (C-6), and prostate (PC-3) cancer cells, with hexane and chloroform fractions of ALD showing particularly low IC50 values, such as 3.37 ± 0.14 and 7.53 ± 0.18 µg/mL against PC-3 cells, respectively (Bhushan et al., 2023b; Kumar et al., 2014).

The anticancer mechanisms of these extracts and metabolites are primarily mediated through the induction of apoptosis via both intrinsic and extrinsic pathways. Lyophilized ALD significantly suppressed the growth and proliferation of T-47D and HeLa cells by inducing apoptosis, as evidenced by suppressed LDH release, reduced NO production, and activation of death receptors in a dosedependent manner (Hasson et al., 2018). Similarly, ALD extract promoted apoptosis in MCF-7 cells by activating caspase-3/7 and annexin V/PI pathways (Kumar et al., 2024b). In-depth mechanistic studies on dehydrocostus lactone have revealed its ability to inhibit angiogenesis by inducing G0/G1 cell cycle arrest in human umbilical vein endothelial cells (HUVECs) through the abrogation of the Akt/ GSK-3β/cyclin D1 and mTOR signaling pathways (Ahmad et al., 2012). Moreover, in laryngeal carcinoma cells, dehydrocostus lactone induced mitochondrial apoptosis by inhibiting the PI3K/ Akt/Bad pathway and stimulating endoplasmic reticulum stressmediated apoptosis, accompanied by the upregulation of p53 and P21 (Zhang et al., 2020).

In vivo studies substantiate the anticancer potential and reduced systemic toxicity of these natural metabolites. In a 12-dimethylbenz (a) anthracene (DMBA)-induced mammary tumour model in rats, treatment with ALD root extract at 500 mg/kg body weight resulted in significant chemopreventive effects, demonstrated by inhibition of tumour parameters, minimal alterations in liver and kidney enzymes, reduction in oxidative stress, decreased proinflammatory cytokines (TNF- α and NF- κ B), and downregulation of proliferation (Ki-67), metastasis (MMP-9), and angiogenesis (VEGF) markers (Kumar et al., 2024a). Another study in mice

induced with cancer using Polyvinyl pyrrolidone K-30 (PVP) showed that ethanolic extract of ALD reduced cancer cell proliferation and mitigated histopathological damage in the liver and kidneys in a dose-dependent manner, contrasting with the severe damage observed in chemotherapy-treated groups (Al-Zayadi et al., 2023). Furthermore, molecular docking and virtual screening studies identified bioactive metabolites in ALD, such as stigmasterol, isoboldine, and beta-sitosterol, with favourable ADME (Absorption, Distribution, Metabolism, and Excretion) and toxicity profiles, showing strong binding affinities to hub genes like SRC, FGFR1, and HSP90AA1 involved in prostate cancer pathways, particularly PI3K/AKT signaling (Figure 5) (Kosanam et al., 2024). This multi-targeted approach, combined with a favourable safety profile, positions ALD and its bioactive metabolites as promising candidates for further development as anticancer therapeutics.

6.4 Organ protection

AR extract demonstrates significant organ-protective effects, particularly against acute lung injury, through a multi-target and multi-pathway mechanism. The study revealed that AR extract ameliorates cyclophosphamide-induced histological damage in the lung by concurrently modulating inflammatory, oxidative, and apoptotic pathways (Attallah et al., 2023). Specifically, its protective action is mediated through the reduction of iNOS and the caspase-3, which attenuates excessive inflammation and inhibits programmed cell death, respectively. Furthermore, AR extract alleviates oxidative stress by significantly decreasing the level of MDA, a marker of lipid peroxidation, while upregulating the gene expression of the HO-1. The accompanying downregulation of microRNA-let-7a suggests a potential involvement in epigenetic regulation, though its precise role requires further elucidation. This evidence collectively indicates that AR extract's organoprotective efficacy is achieved via a synergistic combination of antiinflammatory, antioxidant, and anti-apoptotic activities.

6.5 Anti-diabetic effect

exhibits multi-faceted anti-diabetic properties, demonstrated through both in vitro and in vivo studies. Two novel non-competitive proteinaceous inhibitors, ScAI-R (IC₅₀ = 23 $\mu g/mL$, $K_i = 0.38 \mu M$) and ScAI-L (IC₅₀ = 28 $\mu g/mL$, $K_i =$ 0.32 µM), purified from the roots and leaves, showed high affinity towards human salivary and pancreatic α-amylases, achieving up to 90% inhibitory activity (Ben Abdelmalek et al., 2025). Further supporting these findings, the casein-encapsulated extract of bioactive metabolites from ALD significantly inhibited α-amylase and α -glucosidase activities and enhanced glucose uptake in HepG2 cells (Mushtaq et al., 2025). In diabetic rat models induced by Streptozotocin, oral administration of ALD extracts—containing high concentrations of phenolic metabolites such as dehydrocostus lactone, azulene, eicosapentaenoic acid, linoelaidic acid, isochlorogenic acid A, and chlorogenic acid resulted markedly reduced blood glucose, total cholesterol, and triglyceride levels, while improving HDL-cholesterol and body

weight (Abouelwafa et al., 2024; Gomaa et al., 2021). These results collectively highlight the potential of ALD a source of effective anti-diabetic agents through enzyme inhibition and metabolic regulation.

promising candidate for addressing the global antibiotic resistance crisis.

6.6 Antiparasitic effect

Based on multiple studies, the extract of ALD and its specific bioactive metabolites, costunolide such as dehydrocostuslactone, demonstrate broad-spectrum antiparasitic properties by exhibiting both direct lethal effects and indirect host-protective mechanisms. The ethyl acetate extract of AR (including arbusculin B, α-cyclocostunolide, costunolide, and dehydrocostuslactone) potently inhibited the growth Trypanosoma brucei rhodesiense, with ICsos between 0.8 and 22 µM (Julianti et al., 2011). In vitro, extract of AR showed a significant anthelmintic effects on live adult Ascaridia.galli worms in terms of inhibition of worm motility, with worm motility inhibition of 100% at 24 h post-exposure at the 100 mg/mL (Mir et al., 2024). Beyond direct parasite killing, research reveals a complementary protective role; in a Trichinella spiralis-infected rat model, the extract significantly modulated infection-induced DNA damage and oxidative stress in host tissues (Alghabban et al., 2024). This collective evidence positions ALD as a highly promising source for novel antiparasitic agents, offering a dual mechanism of action that combines direct potency with alleviation of the pathological damage caused by parasitic infections.

6.7 Antimicrobial effect

The essential oil, hexane-chloroform, methanolic, and aqueous extracts of AR have been proven to have certain antimicrobial effects on Acinetobacter baumannii, Aspergillus flavus, Alternaria alternata, Bacillus cereus, Blumeria graminis, Botrytis cinerea, Candida tropicalis, Colletotrichum gloeosporioides, Candida albicans, Didymella glomerata, Escherichia coli, Enterobacter cloacae, Enterococcus faecalis, Fusarium oxysporum, **Fusarium** graminearum, Fusarium lateritium, Klebsiella pneumonia, Pythium Pseudomonas aeruginosa, aphanidermatum, Phytophthora infestans, Sclerotinia sclerotiorum, Salmonella enterica, Staphylococcus aureus, and Staphylococcus epidermidis (Ahmed and Coskun, 2023; Ahmed et al., 2022; Cai et al., 2022; Idriss et al., 2023). Meanwhile, the smoke of ALD successfully inhibited Penicillium crustosum growth of fresh walnuts (Qiao et al., 2023). The antimicrobial mechanism of the extract of AR is multi-targeted, mainly including: damaging the structure of the cell membrane, inhibiting the formation of biofilms, inhibiting quorum sensing and inducing the production of ROS. The composites of ALD demonstrated significantly stronger antimicrobial activity compared to its individual metabolites, causing clear structural damage to resistant strains, such as chitosan-AR nanoconjugates (Alshubaily, 2019), iron oxide nanoparticles of AR (Al-Shaeri and Al-brahim, 2023), and AR-MgO nanoparticles (Mishra et al., 2020). AR extract is undoubtedly a natural antibacterial agent with great research and development value. Its broad-spectrum antibacterial activity, especially its effective effect on drug-resistant bacteria and biofilms, makes it a

6.8 Antiviral effect

Studies have demonstrated the broad-spectrum antiviral potential of extracts and metabolites from ALD and related botanicals. Some lappanolides from ALD exhibits excellent anti-HBV activity (Li H.-B. et al., 2024). The value of the *in vitro* IC50 of AR extract against low pathogenic human coronavirus (HCoV-229E) and human influenza virus (H1N1) influenza virus were 23.21 \pm 1.1 and 47.6 \pm 2.3 µg/mL, respectively. (Attallah et al., 2023). The antiviral activity of ALD acetic acid extract had a significant positive influence against HSV-1 (EC50 = 82.6 g/mL; CC50 = 162.9 g/mL; selectivity index = 1.9) (Idriss et al., 2023).

The bioactive molecules from ALD can be as SARS-CoV-2 main protease inhibitors by computational approach investigation (Hajji et al., 2022). The ALD phytoconstituents have inhibitory potential against the receptor-binding domain of the spike glycoprotein and the main protease of the SARS-CoV-2 Delta (B.1.617.2) variant of the novel coronavirus using molecular docking, DFT, and ADME/Tox studies (Houchi and Messasma, 2022). However, in the actual experiment, no effect was still detected in terms of the inhibition of SARS-CoV2 entry (Idriss et al., 2023).

Currently, the vast majority of research on the antiviral effects of ALD (especially against SARS-CoV-2) remains at the stage of computer simulations (e.g., molecular docking) and *in vitro* studies. While these findings provide a solid theoretical foundation and valuable guidance for further investigation, there is still a long way to go before clinical application can be realized. Validation through animal studies and human clinical trials is still needed. It is also important to note that ALD a complex multimetabolite system, and its antiviral activity likely results from the synergistic effects of various metabolites rather than a single metabolite. From a modern scientific perspective, its antiviral properties may be associated with holistic regulatory functions, such as anti-inflammatory and immunomodulatory effects.

7 Conclusion

In the-present review, we have-presented the biological characteristics, cultivation techniques, chemical composition, pharmacological activities, and processing techniques pertaining to ALD. ALD, as core metabolites in traditional Chinese herbal formulations for treating hepatic and intestinal inflammation, have drawn global attention for their therapeutic potential in biopharmaceutical applications. The future development of ALDderived products must be propelled by technology-driven innovation and supported by institutional synergy. Through standardized systems to elevate industrial upgrading, regulatory science to dismantle market barriers, synthetic biology to redefine resource supply, and clinical research to unlock health potential, ALD-related industries are poised to emerge as global benchmarks in biopharmaceuticals and sustainable development. This transformation will not only alleviate global pressures on natural drug supply chains but also disseminate the wisdom of traditional

Chinese medicine worldwide, advancing the modernization of traditional Chinese medicine into the global health governance framework.

8 Further perspectives

8.1 Limitations

Although ALD demonstrates significant medicinal potential, particularly its notable anticancer activity, there remain major limitations in current research that severely hinder its transition from a traditional remedy to a modern, internationally recognized drug.

First, the standardization of botanical drug sources and quality is the primary obstacle. The origin of ALD is complex, and variations in harvesting seasons and processing methods lead to significant differences in the types and concentrations of its internal chemical metabolites. Existing research has predominantly focused on exploring the activity of specific extracts or metabolites but lacks a comprehensive and systematic quality evaluation system for the botanical drug itself. Most studies rely only on individual active metabolites (such as costunolide and dehydrocostus lactone) as quality control indicators, which fails to comprehensively reflect the holistic nature of the botanical drug and its "multi-metabolite, multi-target" mechanism of action. This lack of standardization makes it difficult to reproduce, compare, and integrate research results from different laboratories, resulting in fragmented data that cannot provide a consistent and reliable material basis for clinical medication.

Secondly, research on the active metabolites and their mechanisms of action remains insufficient. Although numerous bioactive metabolites (costunolide, dehydrocostus lactone, alantolactone, etc.), have been isolated and identified, and their functions in inducing apoptosis and inhibiting angiogenesis have been confirmed, most studies remain at the stage of phenomenological observation and preliminary mechanistic exploration. The majority of mechanistic research relies on in vitro cell models, where the drug concentrations used are often significantly higher than the achievable levels in vivo, raising doubts about the extrapolation of the findings. There is a notable lack of research on how the multiple metabolites in ALD interact synergistically or antagonistically to produce therapeutic effects, which is an aspect central to the philosophy of traditional Chinese medicine. Furthermore, current predominantly on its anticancer properties, while modern scientific explanations of its traditional efficacy are severely lacking. Little effort has been made to integrate newly discovered functions (e.g., anti-inflammatory effects and regulation of gastrointestinal motility) with traditional knowledge.

Thirdly, the pharmacokinetic and safety evaluation systems require significant improvement. The processes of oral absorption, distribution, metabolism, and excretion of the main active metabolites in ALD, such as lactones, remain largely unclear. Key questions persist: What is their bioavailability? In what form do they exert effects *in vivo*, as parent metabolites or metabolites? What kind of pharmacokinetic interactions exist between these metabolites and commonly used chemotherapy drugs? All these

questions remain unanswered. Although some studies suggest that its extracts exhibit low toxicity to normal cells, there is a lack of systematic toxicological studies that meet modern drug approval standards, including investigations into long-term toxicity, reproductive toxicity, and genotoxicity. The potential risk of "nephrotoxicity" has also been frequently mentioned, but solid experimental data to either confirm or refute this claim are still lacking. This uncertainty represents a major concern for its clinical adoption.

Finally, there is a significant lack of clinical research. Almost all encouraging data currently available come from preclinical studies (*in vitro* cell and animal experiments). There is a shortage of rigorously designed and standardized clinical trials to verify the actual efficacy and safety of these treatments in humans. In traditional Chinese clinical practice, ALD is most commonly used in formulations. However, modern research has rarely attempted to simulate such complex environments to explore its role and effects within these formulations. Moreover, the considerable gap between discovering highly active extracts or metabolites in the laboratory and developing them into quality-controlled, standardized preparations suitable for patient use remains largely unaddressed.

8.2 Future research needs

To promote the in-depth development of ALD research and facilitate its clinical translation, future work should focus on the following aspects to meet the needs of a comprehensive research pipeline from basic to applied studies.

Firstly, future research needs to focus on establishing a quality control methodology based on a "holistic perspective". Modern chromatographic, spectroscopic, and coupling techniques (such as HPLC-MS and GC-MS) should be utilized, in combination with chemometric methods, to conduct systematic analysis of ALD from different sources. This will help establish a "fingerprint profiling" standard that covers multiple major active metabolites and characteristic metabolites. At the same time, active research on spectrum-effect relationships should be carried out to correlate chemical fingerprints with pharmacological efficacy data. This will help identify which groups of metabolites are the key material basis for specific pharmacological effects (such as anticancer, anti-inflammatory, and anti-ulcer activities).

Secondly, it is essential to employ multi-omics technologies (genomics, transcriptomics, proteomics, metabolomics) and systemic biological approaches to comprehensively and unbiasedly uncover the target sites and network pathways of ALD and its active metabolites. This represents a core requirement. Comprehensive clinical studies in line with international standards must be initiated. It is imperative to systematically complete pharmacokinetic studies on the main active metabolites and optimized extracts of ALD to clarify their ADME properties. Additionally, toxicological evaluations should be conducted to thoroughly assess the safety of long-term usage and fully elucidate potential risks such as nephrotoxicity.

Thirdly, innovative formulation technologies and translational research should be a top priority for future studies. Given that many lactone metabolites exhibit poor water solubility and low

bioavailability, there is a need to develop novel drug delivery systems such as nanoparticles, liposomes, and phospholipid complexes, to enhance their targeting capability and therapeutic efficacy. Furthermore, these advanced systems can help mitigate systemic toxicity associated with non-selective drug exposure.

8.3 Priorities

Faced with numerous research demands, it is crucial to concentrate resources on addressing key issues. In the short term, priority should be given to foundational work on quality standardization: immediately launching a large-scale chemical composition survey of mainstream commercial ALD, integrating genomics for origin identification, establishing rapid and accurate identification methods based on multi-metabolite quantitative analysis and DNA barcoding technology, and formulating unified and feasible standards for raw medicinal materials. The most promising core active metabolites should be prioritized, with resources focused on completing systematic preclinical pharmacokinetic and pharmacodynamic studies, as well as preliminary acute toxicity and repeated-dose toxicity tests, to quickly obtain an initial assessment of their safety risks. Meanwhile, advanced technologies such as CRISPR screening, molecular docking, and metabolite probes should be employed to precisely identify their direct target sites.

Research on the mechanisms of compatibility in TCM formulations is also one of the priorities. Conducting studies on the synergistic principles and toxicity-reducing effects of pairing ALD with other botanical drugs is essential to provide a scientific basis for the modern application of TCM formulas. For compounds that have been identified as highly active but with low bioavailability, it is necessary to initiate the development of novel drug delivery systems, such as preparing their nanocrystals or phospholipid complexes, and to validate their efficacy enhancement and toxicity reduction in animal models. Additionally, launching small-scale exploratory clinical studies on TCM will help accumulate preliminary data and experience for larger trials.

The establishment of a complete industrial chain can maximize the internationalization of the ALD industry. Facilitating the formation of an integrated industrial chain, which is from large-scale cultivation of ALD bases to standardized extraction and production, and further to the development of final formulations, will help translate research achievements into real industrial applications and build a modern traditional Chinese medicine brand. Meanwhile, establishing an ALD research database to integrate chemical, pharmacological, toxicological, and clinical data will promote global collaboration and knowledge sharing among researchers.

References

Abdel-Rahman, M., Rezk, M. M., Ahmed-Farid, O. A., Essam, S., and Abdel Moneim, A. E. (2020). Saussurea lappa root extract ameliorates the hazards effect of thorium induced oxidative stress and neuroendocrine alterations in adult Male rats. *Environ. Sci. Pollut. Res.* 27 (12), 13237–13246. doi:10.1007/s11356-020-07917-y

Abouelwafa, E., Zaki, A., Sabry, O. M., El-Shiekh, R. A., Caprioli, G., and Abdel-Sattar, E. (2024). Unveiling the chemical profiling and remarkable modulation of carbohydrate metabolism by costus root, *Dolomiaea costus* (Falc.) in streptozotocin (STZ)-induced diabetic rats. *J. Ethnopharmacol.* 326, 117911. doi:10.1016/j.jep.2024.117911

Author contributions

JC: Conceptualization, Funding acquisition, Methodology, Writing – original draft. ZZ: Conceptualization, Writing – original draft, Writing – review and editing. LL: Conceptualization, Software, Writing – review and editing. GW: Supervision, Writing – review and editing. HY: Supervision, Writing – review and editing. XW: Conceptualization, Formal Analysis, Writing – review and editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was financially supported by the Intelligent Detection and Healthcare Research Team of Liuzhou Polytechnic University and the 2024 Major Scientific Research Project of Liuzhou Polytechnic University (2024AK06).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative Al statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Adel, E. M., Al-Tayawi, T. S., Omer, F. H., and Mohammed, M. F. (2025). Phytochemical screening, antioxidant and antibacterial properties of alcoholic extract of *Saussurea costus* roots. *Asian J. Dairy Food Res.* 44 (2), 209–215. doi:10. 18805/ajdfr.DRF-448

Ahmad, A., Wang, C.-Y., Tsai, A.-C., Peng, C.-Y., Chang, Y.-L., Lee, K.-H., et al. (2012). Dehydrocostuslactone suppresses angiogenesis *in vitro* and *in vivo* through inhibition of Akt/GSK-3 β and mTOR signaling pathways. *PLoS One* 7 (2), e31195. doi:10.1371/journal.pone.0031195

- Ahmed, G. S., and Coskun, U. S. Ş. (2023). Investigation of antibacterial and antifungal activity of *Saussurea costus* root extracts. *An. Acad. Bras. Ciênc.* 95 (Suppl. 1), e20230059–17. doi:10.1590/0001-3765202320230059
- Ahmed, A., Ahmad, S., Soni, K., Lapa, B., Afzal, M., Sharma, K., et al. (2016). Suitable solvent and drying condition to enhance phenolics and extractive value of Saussurea costus. *J. Ayurvedic Herb. Med.* 2 (5), 165–170. doi:10.31254/jahm.2016.2504
- Ahmed, H. Y., Kareem, S. M., Atef, A., Safwat, N. A., Shehata, R. M., Yosri, M., et al. (2022). Optimization of supercritical carbon dioxide extraction of Saussurea costus oil and its antimicrobial, antioxidant, and anticancer activities. *Antioxidants* 11 (10), 1960. doi:10.3390/antiox11101960
- Al-Shaeri, M. A. M., and Al-brahim, J. S. (2023). Saussurea costus extract as bio mediator in synthesis iron oxide nanoparticles (IONPs) and their antimicrobial ability. *PLoS One* 18 (3), e0282443. doi:10.1371/journal.pone.0282443
- Al-Zayadi, Z. A., Shanan, H. K., and Akool, A. S. K. (2023). Evaluation of the anticancer effect of *Saussurea costus* root extract against induced hepatic and renal cancer in white mice: a histopathological Study. *Adv. Animal Veterinary Sci.* 11 (7), 1065–1076. doi:10.17582/journal.aavs/2023/11.7.1065.1076
- Alanagh, E. N., Garoosi, G.-a., Haddad, R., Maleki, S., Landín, M., and Gallego, P. P. (2014). Design of tissue culture media for efficient Prunus rootstock micropropagation using artificial intelligence models. *Plant Cell, Tissue Organ Cult.* 117 (3), 349–359. doi:10.1007/s11240-014-0444-1
- Alghabban, A. J. M., Bakr, L., Elbatawy, A. A., El Atrash, A., and Tousson, E. (2024). Impact of Saussurea lappa against foodborne parasite Trichinella spiralis experimental infections induced variation in DNA damage, oxidative stress and PCNA expression in rat skeletal muscles. *Toxicol. Res.* 13 (2), tfae047–6. doi:10.1093/toxres/tfae047
- Alshubaily, F. A. (2019). Enhanced antimycotic activity of nanoconjugates from fungal chitosan and *Saussurea costus* extract against resistant pathogenic Candida strains. *Int. J. Biol. Macromol.* 141, 499–503. doi:10.1016/j.ijbiomac.2019.09.022
- Amen, Y., Abdelwahab, G., Heraiz, A. A., Sallam, M., and Othman, A. (2025). Exploring sesquiterpene lactones: structural diversity and antiviral therapeutic insights. *RSC Adv.* 15 (3), 1970–1988. doi:10.1039/d4ra08125k
- Attallah, N. G. M., Kabbash, A., Negm, W. A., Elekhnawy, E., Binsuwaidan, R., Al-Fakhrany, O. M., et al. (2023). Protective potential of *Saussurea costus* (Falc.) Lipsch. Roots against cyclophosphamide-induced pulmonary injury in rats and its *in vitro* antiviral effect. *Pharmaceuticals* 16 (2), 318. doi:10.3390/ph16020318
- Bai, L. (2023). Effect of muxiang shunqi pills combined with external application of nuangong paste on gastrointestinal adverse reactions and recovery of gastrointestinal function after cesarean section. *Chin. Med. Mod. Distance Educ. China* 21 (20), 102–105. doi:10.3969/j.issn.1672-2779.2023.20.034
- Bai, L., Wu, C., Lei, S., Zou, M., Wang, S., Zhang, Z., et al. (2023). Potential anti-gout properties of wuwei shexiang pills based on network pharmacology and pharmacological verification. *J. Ethnopharmacol.* 305, 116147. doi:10.1016/j.jep.2023. 116147
- Ben Abdelmalek, I., Alhmdi, T. A. A., Ben Bacha, A., and Krayem, N. (2025). Unlocking the therapeutic potential of saussurea costus: purification and functional characterization of α-amylase inhibitors. *Front. Bioeng. Biotechnol.* 13, 1535751. doi:10. 3389/fbioe.2025.1535751
- Bezerra, J. J. L., Pinheiro, A. A. V., and de Oliveira, A. F. M. (2025). Chemical composition and anticancer activity of essential oils from cyperaceae species: a comprehensive review. *Sci. Pharm.* 93, 9. doi:10.3390/scipharm93010009
- Bhatti, H. N., Khan, S. S., Khan, A., Rani, M., Ahmad, V. U., and Choudhary, M. I. (2014). Biotransformation of monoterpenoids and their antimicrobial activities. *Phytomedicine* 21 (12), 1597–1626. doi:10.1016/j.phymed.2014.05.011
- Bhushan, A., Rani, D., Lone, B. A., Tabassum, M., Gupta, A. P., Mondhe, D. M., et al. (2023a). Costunosides A-C: cytotoxic sesquiterpene lactones from the rhizomes of *Aucklandia costus* Falc. *Auckl. costus Falc. Nat. Prod. Res.* 38 (23), 4113–4124. doi:10. 1080/14786419.2023.2275743
- Bhushan, A., Rani, D., Tabassum, M., Kumar, S., Gupta, P. N., Gairola, S., et al. (2023b). HPLC-PDA method for quantification of bioactive compounds in crude extract and fractions of *Aucklandia costus* falc. and cytotoxicity studies against cancer cells. *Molecules* 28 (12), 4815. doi:10.3390/molecules28124815
- Binobead, M. A., Aziz, I. M., Ibrahim, S. M., and Aljowaie, R. M. (2024). Chemical composition and bioactivities of the methanol root extracts of *Saussurea costus. Open Chem.* 22, 20240002. doi:10.1515/chem-2024-0002
- Blazquez, M. A., Liu, Q., Majdi, M., Cankar, K., Goedbloed, M., Charnikhova, T., et al. (2011). Reconstitution of the costunolide biosynthetic pathway in yeast and Nicotiana benthamiana. *PLoS One* 6 (8), e23255. doi:10.1371/journal.pone.0023255
- Cai, Y., Gao, K., Peng, B., Xu, Z., Peng, J., Li, J., et al. (2021). Alantolactone: a natural plant extract as a potential therapeutic agent for cancer. *Front. Pharmacol.* 12, 781033. doi:10.3389/fphar.2021.781033
- Cai, X., Yang, C., Qin, G., Zhang, M., Bi, Y., Qiu, X., et al. (2022). Antimicrobial effects and active compounds of the root of *Aucklandia Lappa* Decne (*Radix Aucklandiae*). *Front. Chem.* 10, 872480. doi:10.3389/fchem.2022.872480
- Cao, S., Zhang, Y., Wang, X., Ba, L., Zhao, Z., Dong, G., et al. (2025). Carvacrol treatment enhances the quality of peach fruit by regulating the metabolism of reactive

oxygen species and the phenyl propanoid pathway. $LWT\,231,\,118329.\,$ doi:10.1016/j.lwt. 2025.118329

- Chandrasena, D., DiFonzo, C., and Byrne, A. (2011). An aphid-dip bioassay to evaluate susceptibility of soybean aphid (Hemiptera: aphididae) to pyrethroid, organophosphate, and neonicotinoid insecticides. *J. Econ. Entomol.* 104 (4), 1357–1363. doi:10.1603/ec10414
- Chen, Z. (2025). Research on the cultivation and management technology of Chinese herbal medicine *Aucklandia lappa* Decne. *Seed Sci. & Technol.* 43 (1), 77–79. doi:10. 19904/j.cnki.cn14-1160/s.2025.01.025
- Chen, Y., and Fang, J.-Y. (2025). The role of colonic microbiota amino acid metabolism in gut health regulation. *Cell Insight* 4 (2), 100227. doi:10.1016/j.cellin. 2025.100227
- Chen, Z., Li, Q., Yu, Z., Yan, X., Wang, W., Xie, Y., et al. (2020). Analysis of the similarities and differences between *Auclandia* and *Vladimirae* rhizomes by chemical profiling and chemometric analysis. *J. Ethnopharmacol.* 255, 112719. doi:10.1016/j.jep. 2020.112719
- Chen, Y., Miao, Z., Sheng, X., Li, X., Ma, J., Xu, X., et al. (2022a). Sesquiterpene lactones-rich fraction from *Aucklandia lappa* Decne. alleviates dextran sulfate sodium induced ulcerative colitis through co-regulating MAPK and Nrf2/Hmox-1 signaling pathway. *J. Ethnopharmacol.* 295, 115401. doi:10.1016/j.jep.2022.115401
- Chen, Z., Wei, C., Yu, Z., Yang, K., Huang, Z., Hu, H., et al. (2022b). An effective method for preventing cholestatic liver injury of *Aucklandiae* Radix and *Vladimiriae* Radix: inflammation suppression and regulate the expression of bile acid receptors. *J. Ethnopharmacol.* 294, 115330. doi:10.1016/j.jep.2022.115330
- Chen, X., Li, F., Xie, X., Zhao, L., and Zan, K. (2023). Comparison the content changes of heavy metals and harmful elements in *Aucklandiae Radix* before and after processing by ICP-MS. *Special Wild Econ. Animal Plant Res.* 45 (4), 135–138. doi:10.16720/j.cnki. tcyj.2023.127
- Cheng, C.-L., Chen, K.-J., Shih, P.-H., Lu, L.-Y., Hung, C.-F., Lin, W.-C., et al. (2006). Chronic renal failure rats are highly sensitive to aristolochic acids, which are nephrotoxic and carcinogenic agents. *Cancer Lett.* 232 (2), 236–242. doi:10.1016/j. canlet.2005.02.021
- China Pharmacopoeia Committee (2020a). Pharmacopoeia of the People's Republic of China muxiang. Beijing: China Medical Science and Technology Press.
- China Pharmacopoeia Committee (2020b). Pharmacopoeia of the People'S Republic of China chuanmuxiang. Beijing: China Medical Science and Technology Press.
- China Pharmacopoeia Committee (2020c). Pharmacopoeia of the People's Republic of China liuwei Muxiang San. Beijing: China Medical Science and Technology Press.
- China Pharmacopoeia Committee (2020d). Pharmacopoeia of the People's Republic of China muxiang binglang wan. Beijing: China Medical Science and Technology Press.
- China Pharmacopoeia Committee (2020e). Pharmacopoeia of the People's Republic of China muxiang fenqi wan. Beijing: China Medical Science and Technology Press.
- China Pharmacopoeia Committee (2020f). Pharmacopoeia of the People's Republic of China muxiang shunqi wan. Beijing: China Medical Science and Technology Press.
- China Pharmacopoeia Committee (2020g). Pharmacopoeia of the People's Republic of China tumuxiang. Beijing: China Medical Science and Technology Press.
- Choi, D. H., Kim, J. Y., An, J. H., Sung, S. H., and Kong, H. S. (2021). Effects of saussurea costus on apoptosis imbalance and inflammation in benign prostatic hyperplasia. J. Ethnopharmacol. 279, 114349. doi:10.1016/j.jep.2021.114349
- Chu, C.-Y., Ho, P.-H., and Cho, Y.-T. (2019). Radix Aucklandiae (dried root of *Saussurea costus*)-induced acute generalized exanthematous pustulosis confirmed by patch testing. *Dermatol. Sin.* 37 (2), 98–102. doi:10.4103/ds.ds_16_18
- Chun, J., Choi, R. J., Khan, S., Lee, D.-S., Kim, Y.-C., Nam, Y.-J., et al. (2012). Alantolactone suppresses inducible nitric oxide synthase and cyclooxygenase-2 expression by down-regulating NF-κB, MAPK and AP-1 *via* the MyD88 signaling pathway in LPS-activated RAW 264.7 cells. *Int. Immunopharmacol.* 14 (4), 375–383. doi:10.1016/j.intimp.2012.08.011
- da Silva, L. P., Borges, B. A., Veloso, M. P., Chagas-Paula, D. A., Gonçalves, R. V., and Novaes, R. D. (2021). Impact of sesquiterpene lactones on the skin and skin-related cells? A systematic review of *in vitro* and *in vivo* evidence. *Life Sci.* 265, 118815. doi:10. 1016/i.lfs.2020.118815
- Dai, C., Liu, N., Chen, W., Qian, W., and Hou, X. (2012). Simo decoction promotes contraction of antral circular smooth muscle mainly *via* muscarinic M3 receptor. *J. Ethnopharmacol.* 144 (2), 270–276. doi:10.1016/j.jep.2012.09.008
- Dong, S., Ma, L.-Y., Liu, Y.-T., Yu, M., Jia, H.-M., Zhang, H.-W., et al. (2018). Pharmacokinetics of costunolide and dehydrocostuslactone after oral administration of *Radix aucklandiae* extract in normal and gastric ulcer rats. *J. Asian Nat. Prod. Res.* 20 (11), 1055–1063. doi:10.1080/10286020.2018.1489379
- Elshaer, S. E., Hamad, G. M., Hafez, E. E., Baghdadi, H. H., El-Demerdash, F. M., and Simal-Gandara, J. (2022). Root extracts of *Saussurea costus* as prospective detoxifying food additive against sodium nitrite toxicity in male rats. *Food Chem. Toxicol.* 166, 113225. doi:10.1016/j.fct.2022.113225
- Feng, Z.-G., Cai-Rang, X.-D., Tan, X.-Y., Li, C.-Y., Zeng, S.-Y., Liu, Y., et al. (2023). Processing methods and the underlying detoxification mechanisms for toxic medicinal

materials used by ethnic minorities in China: a review. *J. Ethnopharmacol.* 305, 116126. doi:10.1016/j.jep.2022.116126

- Feng, Y.-L., Xu, X.-R., Zhu, Q.-M., Chang, J., Zhang, H.-L., Wang, N., et al. (2024). *Aucklandiae* radix targeted PKM2 to alleviate ulcerative colitis: insights from the photocrosslinking target fishing technique. *Phytomedicine* 134, 155973. doi:10.1016/j.phymed.2024.155973
- Ferreira-Guerra, M., Marquès-Bueno, M., Mora-García, S., and Caño-Delgado, A. I. (2020). Delving into the evolutionary origin of steroid sensing in plants. *Curr. Opin. Plant Biol.* 57, 87–95. doi:10.1016/j.pbi.2020.06.005
- Gaitnieks, T., Klavina, D., Muiznieks, I., Pennanen, T., Velmala, S., Vasaitis, R., et al. (2016). Impact of Heterobasidion root-rot on fine root morphology and associated fungi in picea abies stands on peat soils. *Mycorrhiza* 26 (5), 465–473. doi:10.1007/s00572-016-0685-4
- Gao, T., Jiang, M., Deng, B., Zhang, Z., Fu, Q., and Fu, C. (2020). *Aurantii fructus*: a systematic review of ethnopharmacology, phytochemistry and pharmacology. *Phytochem. Rev.* 20 (5), 909–944. doi:10.1007/s11101-020-09725-1
- Gao, Y., Zhuang, T., Gao, L., Guo, Z., Yu, J., and Tang, Y. (2024). Effects of muxiang binglang Wan on tumor necrosis factor a content and pepsinogen in mice. *J. Baotou Med. Coll.* 40 (4), 15–20. doi:10.16833/j.cnki.jbmc.2024.04.003
- Gomaa, H. F., Abdelmalek, I. B., and Abdel-Wahhab, K. G. (2021). The anti-diabetic effect of some plant extracts against Streptozotocin induced diabetes type 2 in Male albino rats. *Endocr. Metab. Immune* 21 (8), 1431–1440. doi:10.2174/1871530320666201016145502
- Guccione, C., Ros, G., Gallori, S., Bergonzi, M. C., and Bilia, A. R. (2017). Rapid and efficient extraction and HPLC analysis of sesquiterpene lactones from *Aucklandia lappa* root. *Nat. Prod. Commun.* 12 (2), 1934578X1701200218–216. doi:10.1177/1934578X1701200218
- Hajji, H., Alaqarbeh, M., Lakhlifi, T., Ajana, M. A., Alsakhen, N., and Bouachrine, M. (2022). Computational approach investigation bioactive molecules from *saussurea costus* plant as SARS-CoV-2 main protease inhibitors using reverse docking, molecular dynamics simulation, and pharmacokinetic ADMET parameters. *Comput. Biol. Med.* 150, 106209. doi:10.1016/j.compbiomed.2022.106209
- Hasson, S. S., H Al-Shubi, A. S., Al-Busaidi, J. Z., Al-Balushi, M. S., Hakkim, F. L., Rashan, L., et al. (2018). Potential of *Aucklandia lappa* Decne Ethanolic extract to trigger apoptosis of human T47D and Hela cells. *Asian pac. J. Cancer* 19 (7), 1917–1925. doi:10. 22034/APJCP.2018.19.7.1917
- He, J., Liu, L., Liu, X., Chen, H., Liu, K., Huang, N., et al. (2022). Epoxymicheliolide prevents dextran sulfate sodium-induced colitis in mice by inhibiting TAK1-NF-κB pathway and activating Keap1-NRF2 signaling in macrophages. *Int. Immunopharmacol.* 113, 109404. doi:10.1016/j.intimp.2022.109404
- Ho, S.-C., and Kuo, C.-T. (2014). Hesperidin, nobiletin, and tangeretin are collectively responsible for the anti-neuroinflammatory capacity of tangerine peel (*Citri reticulatae* pericarpium). *Food Chem. Toxicol.* 71, 176–182. doi:10.1016/j.fct.2014.06.014
- Hong, W., Chen, X., Xiao, J., Chen, G., Yang, J., Zhang, P., et al. (2025). Dehydrocostus lactone attenuates atherogenesis by promoting cholesterol efflux and inhibiting inflammation via TLR2/PPAR- γ /NF- κ B signaling pathway. *Mol. Med.* 31 (1), 243. doi:10.1186/s10020-025-01265-8
- Houchi, S., and Messasma, Z. (2022). Exploring the inhibitory potential of Saussurea costus and Saussurea involucrata phytoconstituents against the Spike glycoprotein receptor binding domain of SARS-CoV-2 Delta (B.1.617.2) variant and the main protease (Mpro) as therapeutic candidates, using molecular docking, DFT, and ADME/Tox studies. *J. Mol. Struct.* 1263, 133032. doi:10.1016/j.molstruc.2022.133032
- Hsu, Y.-L., Wu, L.-Y., and Kuo, P.-L. (2009). Dehydrocostuslactone, a medicinal plant-derived sesquiterpene lactone, induces apoptosis coupled to endoplasmic reticulum stress in liver cancer cells. *J. Pharmacol. Exp. Ther.* 329 (2), 808–819. doi:10.1124/jpet.108.148395
- Huang, Z., Wei, C., Yang, K., Yu, Z., Wang, Z., and Hu, H. (2021). *Aucklandiae* radix and *vladimiriae* radix: a systematic review in ethnopharmacology, phytochemistry and pharmacology. *J. Ethnopharmacol.* 280, 114372. doi:10.1016/j.jep.2021.114372
- Huang, Y. J., Tong, H. Y., Huang, X. J., Xiao, X. C., Dong, Y., and Iqbal, M. S. (2022a). Anshen-buxin-liuwei pill, a Mongolian medicinal formula could alleviate cardiomyocyte hypoxia/reoxygenation injury *via* mitochondrion pathway. *Mol. Biol. Rep.* 49 (2), 885–894. doi:10.1007/s11033-021-06867-z
- Huang, Z., Xu, C., Zhao, L., Wei, C., Wu, Y., Qiu, J., et al. (2022b). Preparation, optimization and *in vivo* study of gastric floating tablets of constunolide and dehydrocostus lactone with ideal therapeutic effect on gastric diseases. *J. Drug Deliv. Sci. Technol.* 78, 103942. doi:10.1016/j.jddst.2022.103942
- Huang, X.-F., Xue, Y., Yong, L., Wang, T.-T., Luo, P., and Qing, L.-S. (2024). Chemical derivatization strategies for enhancing the HPLC analytical performance of natural active triterpenoids. *J. Pharm. Anal.* 14 (3), 295–307. doi:10.1016/j.jpha.2023.07.004
- Idriss, H., Siddig, B., González-Maldonado, P., Elkhair, H. M., Alakhras, A. I., Abdallah, E. M., et al. (2023). Inhibitory activity of saussurea Costus extract against bacteria, Candida, herpes, and SARS-CoV-2. Plants 12 (3), 460. doi:10.3390/plants12030460
- Jiang, H., and Wang, X. (2023). Biosynthesis of monoterpenoid and sesquiterpenoid as natural flavors and fragrances. *Biotechnol. Adv.* 65, 108151. doi:10.1016/j.biotechadv. 2023.108151

- Jo, H.-G., Lee, G.-Y., Baek, C. Y., Song, H. S., and Lee, D. (2021). Analgesic and antiinflammatory effects of Aucklandia lappa root extracts on acetic acid-induced writhing in mice and monosodium iodoacetate-induced osteoarthritis in rats. *Plants* 10, 42. doi:10.3390/plants10010042
- Jomova, K., Alomar, S. Y., Valko, R., Liska, J., Nepovimova, E., Kuca, K., et al. (2025). Flavonoids and their role in oxidative stress, inflammation, and human diseases. *Chem. Biol. Interact.* 413, 111489. doi:10.1016/j.cbi.2025.111489
- Julianti, T., Hata, Y., Zimmermann, S., Kaiser, M., Hamburger, M., and Adams, M. (2011). Antitrypanosomal sesquiterpene lactones from *Saussurea costus*. *Fitoterapia* 82 (7), 955–959. doi:10.1016/j.fitote.2011.05.010
- Kelling, M., Dimza, M., Bartlett, A., Traktuev, D. O., Duarte, J. D., and Keeley, E. C. (2024). Omega-3 fatty acids in the treatment of heart failure. *Curr. Prob. Cardiol.* 49 (9), 102730. doi:10.1016/j.cpcardiol.2024.102730
- Kosanam, S., Pasupula, R., and James, D. (2024). Exploring the anticancer potential of Saussurea costus phytoconstituents against prostate cancer using virtual screening and molecular docking. *J. Nat. Remedies* 24 (11), 2459–2467. doi:10.18311/jnr/2024/41950
- Kumar, S., and Pandey, A. K. (2022). Bio-efficacy of various insecticides against white grubs (Coleoptera: scarabaeidae) infesting sugarcane. *Int. J. Trop. Insect Sci.* 42 (5), 3319–3325. doi:10.1007/s42690-022-00820-8
- Kumar, A., Kumar, S., Kumar, D., and Agnihotri, V. K. (2014). UPLC/MS/MS method for quantification and cytotoxic activity of sesquiterpene lactones isolated from Saussurea lappa. J. Ethnopharmacol. 155 (2), 1393–1397. doi:10.1016/j.jep.2014.07.037
- Kumar, R., Bhardwaj, P., Soni, M., Singh, R., Choudhary, S., Virmani, N., et al. (2024a). Modulation of mammary tumour progression using murine model by ethanol root extract of *Saussurea costus* (falc.) lipsch. *J. Ethnopharmacol.* 319, 117302. doi:10.1016/j.jep.2023.117302
- Kumar, R., Choudhary, S., Soni, M., Singh, R., Kumar, S., and Asrani, R. K. (2024b). Augmentation of cytotoxicity and apoptotic activity in human breast cancer MCF-7 cells by Saussurea costus (Falc.) Lipsch roots. *Ann. Phytomedicine* 13 (2), 433–438. doi:10.54085/ap.2024.13.2.42
- Lee, S., Kim, S.-B., Lee, J., Park, J., Choi, S., Hwang, G. S., et al. (2020). Evaluation of anti-colitis effect of KM1608 and biodistribution of dehydrocostus lactone in mice using bioimaging analysis. *Plants* 9 (9), 1175. doi:10.3390/plants9091175
- Li, J., Li, M., Ye, K., Jiang, Q., Wang, M., Wen, X., et al. (2021a). Chemical profile of xian-he-cao-chang-yan formula and its effects on ulcerative colitis. *J. Ethnopharmacol.* 267, 113517. doi:10.1016/j.jep.2020.113517
- Li, R.-l., Zhang, Q., Liu, J., He, L.-y., Huang, Q.-w., Peng, W., et al. (2021b). Processing methods and mechanisms for alkaloid-rich Chinese herbal medicines: a review. *J. Integr. Med.* 19 (2), 89–103. doi:10.1016/j.joim.2020.12.003
- Li, X., Wang, L., Ma, Y., and Huang, P. (2021c). Study on the processing technology of Aucklandia lappa Decne. *Strait Pharm. J.* 33 (10), 26–28.
- Li, H.-B., Bai, S.-Q., Shu, T.-Y., Wang, Q., Chen, H., Su, L.-H., et al. (2024a). Lappanolides A–N, fourteen undescribed sesquiterpenoids from *Saussurea costus* (Syn. Saussurea lappa) and their anti HBV activity. *Phytochemistry* 226, 114207. doi:10.1016/j.phytochem.2024.114207
- Li, S.-Y., Xu, D.-Q., Chen, Y.-Y., Fu, R.-J., and Tang, Y.-P. (2024b). Several major herb pairs containing Coptidis rhizoma: a review of key traditional uses, constituents and compatibility effects. *Front. Pharmacol.* 15, 1399460. doi:10.3389/fphar.2024.1399460
- Li, Y., Chen, Y., Zhou, Y., He, J., Zhou, Q., and Wang, M. (2024c). Unveiling the potentials and action mechanisms of *Citri reticulatae* Pericarpium as an anti-inflammatory food. *Food Front.* 6, 163–184. doi:10.1002/fft2.506
- Liang, S., Chen, R., Ciren, Z., Jiang, G., and Du, L. (2024). Chemical composition of volatile oils from different processed products of *Aucklandia Lappa* decne. and *Vladimiria Souliei* (Franch.) ling based on Gaschromatography-Mass spectrometry. *J. Chengdu Univ. TCM* 47 (2), 1–10. doi:10.13593/j.cnki.51-1501/r.2024.0.001
- Lim, J. S., Lee, S. H., Lee, S. R., Lim, H. J., Roh, Y. S., Won, E. J., et al. (2020). Inhibitory effects of *Aucklandia lappa* decne. Extract on inflammatory and oxidative responses in LPS-Treated macrophages. *Molecules* 25 (6), 1336. doi:10.3390/molecules25061336
- Liu, X.-n., Li, H.-m., Wang, S.-p., Zhang, J.-z., and Liu, D.-l. (2021). Sesquiterpene lactones of Aucklandia lappa: pharmacology, pharmacokinetics, toxicity, and structure–activity relationship. *Chin. Herb. Med.* 13 (2), 167–176. doi:10.1016/j. chmed.2020.11.005
- Lu, J.-j., Ali, A., He, E.-q., Yan, G.-q., Arak, T.-u., and Gao, S.-J. (2019). Establishment of an open, sugar-free tissue culture system for sugarcane micropropagation. *Sugar Tech.* 22 (1), 8–14. doi:10.1007/s12355-019-00758-1
- Lu, M., Chu, Z., Wang, L., Liang, C., Sun, P., Xiong, S., et al. (2020). Pharmacokinetics and tissue distribution of four major bioactive components in rats after oral administration of Xianglian pill. *Biomed. Chromatogr.* 34 (3), e4770. doi:10.1002/bmc.4770
- Lu, T., Yu, H. J., Wang, T. Y., Zhang, T. Y., Shi, C. H., and Jiang, W. J. (2022). Influence of the electrical conductivity of the nutrient solution in different phenological stages on the growth and yield of cherry tomato. *Horticulturae* 8 (5), 378. doi:10.3390/horticulturae8050378
- Luo, J.-n., Li, J.-y., Wang, H., Wang, Z.-x., Ding, Y., Sun, H., et al. (2024). Content determination of aristolochic acid | and AristolochicaAcid | in Arastolochia

moupinensis Franch. *UPLC-MS/MS*. Mod. Chin. Med. 26 (8), 1326–1331. doi:10.13313/j.issn.1673-4890.20230615001

- Lyu, H.-Y., Bao, M.-Y., Io, C.-C., Xiong, H.-M., Chen, F.-L., Bai, L.-P., et al. (2023). Sesquiterpenoids from the roots of *Aucklandia costus* and their anti-inflammatory activities. *Fitoterapia* 169, 105604. doi:10.1016/j.fitote.2023.105604
- Mammate, N., El oumari, F. E., Imtara, H., Belchkar, S., Benjelloun Touimi, G., Al-Zharani, M., et al. (2023). Anti-struvite, antimicrobial, and anti-inflammatory activities of aqueous and ethanolic extracts of *Saussurea costus* (Falc) lipsch asteraceae. *Molecules* 28 (2), 667. doi:10.3390/molecules28020667
- Meng, C., Wang, P., Hao, Z., Gao, Z., Li, Q., Gao, H., et al. (2021). Ecological and health risk assessment of heavy metals in soil and Chinese herbal medicines. *Environ. Geochem. Health* 44 (3), 817–828. doi:10.1007/s10653-021-00978-z
- Mir, F. H., Tanveer, S., Bharti, P., and Para, B. A. (2024). Anthelmintic activity of Saussurea costus (Falc.) Lipsch. Against Ascaridia galli, a pathogenic nematode in poultry: *in vitro* and *in vivo* studies. *Acta Parasitol*. 69 (2), 1192–1200. doi:10.1007/s11686-024-00837-8
- Mishra, Y. K., Amina, M., Al Musayeib, N. M., Alarfaj, N. A., El-Tohamy, M. F., Oraby, H. F., et al. (2020). Biogenic green synthesis of MgO nanoparticles using Saussurea costus biomasses for a comprehensive detection of their antimicrobial, cytotoxicity against MCF-7 breast cancer cells and photocatalysis potentials. PLoS One 15 (8), e0237567. doi:10.1371/journal.pone.0237567
- Mushtaq, M., Amin, Q. A., Wani, T. A., Bhat, T. A., Parveen, S., and Beigh, M. A. (2025). Supercritical fluid extraction and micro-encapsulation of *Saussurea costus* roots for next gen functional foods: antioxidant potential, surface morphology and anti-diabetic potential. *Food Chem.* 495, 146290. doi:10.1016/j.foodchem.2025.146290
- Paudel, P., Park, C. H., Jung, H. A., Yokozawa, T., and Choi, J. S. (2020). A systematic review on anti-alzheimer's disease activity of prescription Kangen-karyu. *Drug Discov. & Ther.* 14 (2), 61–66. doi:10.5582/ddt.2020.03013
- Peng, M., Zhao, H., Fu, J., Zhang, Z., Yuan, Z., and Zhang, B. (2025). Structural characterization of two low-molecular-weight polysaccharides from the roots of *Aucklandia lappa* Decne. and their bioactivity on the gut microbiota of immunocompromised mice. *J. Food Sci.* 90 (6), e70013–e70016. doi:10.1111/1750-3841.70013
- Pupilli, F., and Barcaccia, G. (2012). Cloning plants by seeds: inheritance models and candidate genes to increase fundamental knowledge for engineering apomixis in sexual crops. *J. Biotechnol.* 159 (4), 291–311. doi:10.1016/j.jbiotec.2011.08.028
- Qiao, L., Jiao, Y., Li, X., Zhang, Y., Lu, L., Zhang, X., et al. (2023). Herbal smoke fumigation for controlling Penicillium crustosum in fresh walnuts. *Food Res. Int.* 167, 112709. doi:10.1016/j.foodres.2023.112709
- Rasul, A., Khan, M., Ali, M., Li, J., Li, X., Jann, M. W., et al. (2013). Targeting apoptosis pathways in cancer with alantolactone and isoalantolactone. *Sci. World J.* 2013 (1), 248532. doi:10.1155/2013/248532
- Renqing, D., Feng, X., Luosang, D., Hua, Q., and San, Z. (2023). Molecular mechanism of Tibetan medicine Liuwei Muxiang pills for treating chronic Non atrophic Gastritis based on serum active chemical components. *Plateau Sci. Res.* 7 (3), 58–71. doi:10. 16249/j.cnki.2096-4617.2023.03.007
- Seo, C. S., and Shin, H. K. (2015). Simultaneous determination of three sesquiterpene lactones in *Aucklandia lappa* decne by high-performance liquid chromatography. *Pharmacogn. Mag.* 11 (43), 562–566. doi:10.4103/0973-1296.160471
- Seo, C. S., Lim, H. S., Jeong, S. J., and Shin, H. K. (2015). Anti-allergic effects of sesquiterpene lactones from the root of Aucklandia lappa decne. *Mol. Med. Rep.* 12 (5), 7789–7795. doi:10.3892/mmr.2015.4342
- Shati, A. A., Alkahtani, M. A., Alfaifi, M. Y., Elbehairi, S. E. I., Elsaid, F. G., Prasanna, R., et al. (2020). Secondary metabolites of saussurea costus leaf extract induce apoptosis in breast, liver, and Colon cancer cells by Caspase-3-Dependent intrinsic pathway. *Biomed. Res. Int.* 2020 (1), 1608942. doi:10.1155/2020/1608942
- Shum, K. C., Chen, F., Li, S. L., Wang, J., But, P. P. H., and Shaw, P. C. (2007). Authentication of Radix *aucklandiae* and its substitutes by GC-MS and hierarchical clustering analysis. *J. Sep. Sci.* 30 (18), 3233–3239. doi:10.1002/jssc.200700232
- Smirle, M. J., Zurowski, C. L., Lowery, D. T., and Mostafa, A. M. (2013). Insecticide susceptibility of three species of cutworm (lepidoptera: noctuidae) pests of grapes. *J. Econ. Entomol.* 106 (5), 2135–2140. doi:10.1603/ec13110
- Song, Y. Y., Zhang, T. T., Tang, H., Xu, L., Xing, Y. P., Zhao, R., et al. (2021). The complete mitochondrial genome of *Aucklandia lappa* decne. (asteraceae, aucklandia falc.). Mitochondrial DNA B 6 (6), 1691–1693. doi:10.1080/23802359.2021.1914524
- Song, S., Qiu, R., Jin, X., Zhou, Z., Yan, J., Ou, Q., et al. (2022a). Mechanism exploration of ancient pharmaceutic processing (Paozhi) improving the gastroprotective efficacy of *Aucklandiae Radix*. *J. Ethnopharmacol.* 287, 114911. doi:10.1016/j.jep.2021.114911
- Song, S., Zhou, J., Li, Y., Liu, J., Li, J., and Shu, P. (2022b). Network pharmacology and experimental verification based research into the effect and mechanism of Aucklandiae Radix–Amomi fructus against gastric cancer. *Sci. Rep.* 12 (1), 9401. doi:10.1038/s41598-022-13223-z
- Song, S., Wei, D., Qiu, R., Huang, Y., He, J., Du, X., et al. (2023). Study on the protective effect of ethanol extract from Aucklandiae Radix on lipopolysaccharide-induced liver injury. *China J. Traditional Chin. Med. Pharm.* 38 (12), 6020–6023.

- Song, S., Qiu, R., Huang, Y., Zhou, Z., Yan, J., Ou, Q., et al. (2024). Study on the mechanism of hepatotoxicity of Aucklandiae radix through liver metabolomics and network pharmacology. *Toxicol. Res.* 13 (4), tfae123–13. doi:10.1093/toxres/tfae123
- Sun, M., Zhan, H., Long, X., Alsayed, A. M., Wang, Z., Meng, F., et al. (2024). Dehydrocostus lactone alleviates irinotecan-induced intestinal mucositis by blocking TLR4/MD2 complex formation. *Phytomedicine* 128, 155371. doi:10.1016/j.phymed. 2024.155371
- Suo, S., Lai, Y., Li, M., Song, Q., Cai, J., Zhao, J., et al. (2018). Phytochemicals, pharmacology, clinical application, patents, and products of Amomi fructus. *Food Chem. Toxicol.* 119, 31–36. doi:10.1016/j.fct.2018.05.051
- Tan, W., Li, K., Liu, D., and Xing, W. (2023). Cercospora leaf spot disease of sugar beet. *Plant Signal. & Behav.* 18 (1), 2214765. doi:10.1080/15592324.2023.2214765
- Vogt, T. (2010). Phenylpropanoid biosynthesis. *Mol. Plant* 3 (1), 2–20. doi:10.1093/mp/ssp106
- Wang, F.-Y., Zhong, L. L. D., Kang, N., Dai, L., Lv, L., Bian, L.-Q., et al. (2016). Chinese herbal formula for postprandial distress syndrome: study protocol of a double-blinded, randomized, placebo-controlled trial. *Eur. J. Integr. Med.* 8 (5), 688–694. doi:10.1016/j.eujim.2016.03.010
- Wang, X., Giusti, A., Ny, A., and de Witte, P. A. (2020a). Nephrotoxic effects in zebrafish after prolonged exposure to aristolochic acid. *Toxins* 12 (4), 217. doi:10.3390/toxins12040217
- Wang, Y., Fan, X.-x., Yang, J., Wang, Z.-q., Wang, N., Chen, J.-q., et al. (2020b). Research progress on terpenes and pharmacological effects of Saussurea lappa. *China J. Chin. Materia Medica* 45 (24), 5917–5928. doi:10.19540/j.cnki.cjcmm. 20200903.601
- Wang, Y. G., Zhang, N., Chen, C. L., Jiang, Y. C., and Liu, T. (2023). Nonlinear adaptive generalized predictive control for PH model of nutrient solution in plant factory based on ANFIS. *Processes* 11 (8), 2317. doi:10.3390/pr11082317
- Wang, X. r., Zhang, J. t., Guo, X. h., Li, M. h., Jing, W. g., Cheng, X. l., et al. (2024). Digital identification of Aucklandiae radix, Vladimiriae radix, and Inulae radix based on multivariate algorithms and UHPLC-QTOF-MS analysis. *Phytochem. Anal.* 36, 92–100. doi:10.1002/pca.3421
- Wu, G., Li, Z., Zheng, Y., Zhang, Y., Liu, L., Gong, D., et al. (2022). Supplementing cholamine to diet lowers laying rate by promoting liver fat deposition and altering intestinal microflora in laying hens. *Poult. Sci.* 101 (10), 102084. doi:10.1016/j.psj.2022. 102084
- Wu, M.-l., Bao, S.-y., Huang, H.-x., and Sun, L.-e. (2024a). Quality standard of bran Radix Aucklandiae formula granules based on standard decoction. *Cent. South Pharm.* 22 (11), 3014–3020. doi:10.7539/j.issn.1672-2981.2024.11.029
- Wu, Y., Wu, J., Li, L., OuYang, H., Wu, L., Yang, C., et al. (2024b). A gel plaster in the form of nipple cover: a comfortable and safe transdermal delivery method for mammary hyperplasia. *Int. J. Pharm.* 662, 124500. doi:10.1016/j.ijpharm.2024.124500
- Wu, H., Luo, J., Chen, Y., Zhang, J., Han, T., Tan, B., et al. (2025). Integrated chemometrics and biological validation assay to identify α,β -unsaturated carbonyl compounds as key anti-inflammatory and antioxidant agents in *Aucklandiae Radix. Fitoterapia* 186, 106797. doi:10.1016/j.fitote.2025.106797
- Xu, R., Zhou, G., Peng, Y., Wang, M., and Li, X. (2015). Pharmacokinetics, tissue distribution and excretion of Isoalantolactone and alantolactone in rats after oral administration of Radix inulae extract. *Molecules* 20 (5), 7719–7736. doi:10.3390/molecules20057719
- Xu, Y., Ge, X., Lv, Y., Yang, Z., Li, F., and Yang, Z. (2025). Engineering plant hosts for high-efficiency accumulation of flavonoids: advances, challenges and perspectives. *Biotechnol. Adv.* 84, 108692. doi:10.1016/j.biotechadv.2025.108692
- Xue, R., Deng, C., Cao, H., Zhang, K., Lu, T., and Mao, C. (2020). Quality assessment of raw and baked *Aucklandia lappa* Decne. by color measurement and fingerprint analysis. *J. Sep. Sci.* 43 (15), 3017–3026. doi:10.1002/jssc.202000308
- Yan, X., Wang, W., Chen, Z., Xie, Y., Li, Q., Yu, Z., et al. (2020). Quality assessment and differentiation of *Aucklandiae* radix and *Vladimiriae* Radix based on GC-MS fingerprint and chemometrics analysis: basis for clinical application. *Anal. Bioanal. Chem.* 412 (7), 1535–1549. doi:10.1007/s00216-019-02380-2
- Yang, H. J., Kim, M. J., Kang, S., Moon, N. R., Kim, D. S., Lee, N. R., et al. (2017). Topical treatments of *Saussurea costus* root and *Thuja orientalis* L. synergistically alleviate atopic dermatitis-like skin lesions by inhibiting protease-activated receptor-2 and NF-κB signaling in HaCaT cells and Nc/Nga mice. *J. Ethnopharmacol.* 199, 97–105. doi:10.1016/j.jep.2017.01.055
- Yang, H.-S., Mauki, D. H., Zheng, Y.-X., Wang, T.-H., and He, X.-Y. (2025). Terpenoids: a promising traditional chinese medicine for neuropathic pain relief. *Pharmacol. Res.* 216, 107789. doi:10.1016/j.phrs.2025.107789
- Yang, W., Guo, D., Cao, W., Pan, X., Xue, Y., Zhang, J., et al. (2020). Effects of 27 strains of Arbuscular Mycorrhizal Fungi inoculation on physiology and biochemistry and major components of terpenoids in potted saussurea costus. *Chin J. Trop. Crop.* 41, 1822–1830. doi:10.3969/j.issn.1000-2561.2020.09.015
- Yi, Y.-n., Cheng, X.-m., Liu, L.-a., Hu, G.-y., Cai, G.-x., Deng, Y.-d., et al. (2011). HPLC fingerprint with multi-components analysis for quality consistency evaluation of traditional Chinese medicine si-mo-tang oral liquid preparation. *Chem. Res. Chin. Univ.* 27 (5), 756–763.

Yin, H., Zhuang, Y.-b., Li, E. e., Bi, H.-p., Zhou, W., and Liu, T. (2015). Heterologous biosynthesis of costunolide in *Escherichia coli* and yield improvement. *Biotechnol. Lett.* 37 (6), 1249–1255. doi:10.1007/s10529-015-1784-6

- Yuan, Y., Hu, Q., Liu, L., Xie, F., Yang, L., Li, Y., et al. (2022). Dehydrocostus lactone suppresses Dextran Sulfate sodium-induced colitis by targeting the IKK α / β -NF- κ B and Keap1-Nrf2 signalling pathways. *Front. Pharmacol.* 13, 817596. doi:10.3389/fphar.2022. 817596
- Yuan, W., Yu, X., Wu, H., Xu, M., Dai, Y., Xinru, L., et al. (2024). Research progress on sesquiterpenoid components and pharmacological effects of *Aucklandiae Radix*. *Yunnan Chem. Technol.* 51 (8), 1–5. doi:10.3969/j.issn.1004-275X.2024.08.01
- Zeng, L. R., Pan, B. W., Cai, J., Liu, L. J., Dong, Z. C., Zhou, Y., et al. (2024). Construction, structural modification, and bioactivity evaluation of pentacyclic triterpenoid privileged scaffolds in active natural products. *RSC Adv.* 14 (53), 39436–39461. doi:10.1039/d4ra07602h
- Zhang, J., Gao, W., Liu, Z., and Zhang, Z. (2013). Identification and simultaneous determination of twelve active components in the methanol extract of traditional medicine weichang'an pill by HPLC-DAD-ESI-MS/MS. *Iran. J. Pharm. Res.* 12 (1), 15–24.
- Zhang, J., Hu, X., Gao, W., Qu, Z., Guo, H., Liu, Z., et al. (2014). Pharmacokinetic study on costunolide and dehydrocostuslactone after oral administration of traditional medicine *Aucklandia lappa* Decne. by LC/MS/MS. *J. Ethnopharmacol.* 151 (1), 191–197. doi:10.1016/j.jep.2013.10.024
- Zhang, M., Zhang, C., Chen, Y., and Fu, X. (2015). Three statistical experimental designs for enhancing yield of active compounds from herbal medicines and anti motion sickness bioactivity. *Pharmacogn. Mag.* 11 (43), 435–443. doi:10.4103/0973-1296.160444
- Zhang, J., Wei, Z., Zhang, A., Zhao, Y., Zhao, G., Cai, X., et al. (2016). Detection aristolochic acids 1 and 2 in costustoot via electrochemical method and liquid

- chromatography. Int. J. Electrochem. Sci. 11 (8), 6830-6837. doi:10.20964/2016.
- Zhang, L., Lecoq, M., Latchininsky, A., and Hunter, D. (2019). Locust and grasshopper management. *Annu. Rev. Entomol.* 64 (1), 15–34. doi:10.1146/annurevento-011118-112500
- Zhang, R., Hao, J., Wu, Q., Guo, K., Wang, C., Zhang, W. K., et al. (2020). Dehydrocostus lactone inhibits cell proliferation and induces apoptosis by PI3K/Akt/Bad and ERS signalling pathway in human laryngeal carcinoma. *J. Cell. Mol. Med.* 24 (11), 6028–6042. doi:10.1111/jcmm.15131
- Zhang, L., Xiao, Y., Yang, R., Wang, S., Ma, S., Liu, J., et al. (2021). Systems pharmacology to reveal multi-scale mechanisms of traditional Chinese medicine for gastric cancer. *Sci. Rep.* 11 (1), 22149. doi:10.1038/s41598-021-01535-5
- Zhang, L., Fan, H., Pan, X., Han, Z., Tian, C., and Schneiter, R. (2025). Comparative multi-omics reveals phenylpropanoid pathway activation in resistant poplar species challenged by Cytospora chrysosperma. *Ind. Crops Prod.* 234, 121620. doi:10.1016/j. indcrop.2025.121620
- Zheng, J.-m., Shang, M.-y., Wang, J.-l., Dai, G.-n., Song, J.-y., and Duan, B.-z. (2022). Research progress on chemical constituents, pharmacological effects and clinical applications of *Aucklandiae* Radix and prediction analysis on Q-Marker. *Chin. Traditional Herb. Drugs* 53 (13), 4198–4213. doi:10.7501/j.issn.0253-2670. 2022.13.033
- Zhou, Q., Zhang, W.-x., He, Z.-q., Wu, B.-s., Shen, Z.-f., Shang, H.-t., et al. (2020). The possible anti-inflammatory effect of Dehydrocostus lactone on DSS-Induced colitis in mice. *Evid.-Based Compl. Alt.* 2020 (1), 5659738. doi:10.1155/2020/5659738
- Zhuang, K., Xia, Q., Zhang, S., Maharajan, K., Liu, K., and Zhang, Y. (2021). A comprehensive chemical and pharmacological review of three confusable Chinese herbal medicine—*Aucklandiae* radix, *Vladimiriae* radix, and *Inulae* radix. *Phytother. Res.* 35 (12), 6655–6689. doi:10.1002/ptr.7250