



# Photodynamic Therapy Using Cerenkov and Radioluminescence Light

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In this short review the potential use of Cerenkov radiation and radioluminescence as internal sources for Photodynamic therapy (PDT) is discussed. PDT has been developed over the course of more than 100 years and is based on the induced photo conversion of a drug called photosensitizer (PS) that triggers the production of cytotoxic reactive oxygen species (ROS) leading to the killing of the cells. In order to overcome the problem of light penetration in the tissues, different solutions were proposed in the past. The use of radioisotopes like: <sup>18</sup>F, <sup>64</sup>Cu, <sup>90</sup>Y, <sup>177</sup>Lu as internal light sources increase the light fluence at the PS compared to an external source, resulting in a larger cytotoxic effect.

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## 1 INTRODUCTION

Photodynamic therapy (PDT) has been developed over the course of more than 100 years as will be described in more details in **section 2** of this review. The basic idea behind PDT is rather simple and is based on the induced photo conversion of a drug called photosensitizer (PS) that triggers the production of cytotoxic reactive oxygen species (ROS), leading to the killing of the cells as will be described in details in **section 3**. The clinical use of PDT is mainly focused on treating different types of cancers; however other disease like herpes acne can be treated [1], showing a wide range of possible applications. The first PS approved for the clinical use to treat cancer was an optimized hematoporphyrin derivative (HPD) known with the commercial name of Photofrin<sup>®</sup>, described in details by [2]. Since the introduction of HPD in the seventies there has been significant research focused on the development of second and third generation PS in order to increase the yield of cytotoxic ROS and introduce targeting strategies such as antibody conjugation or nanoparticles (NPs) [3]. A detailed description of the different PS is well beyond the scope of this review and more details can be found for example in [4, 5]. The main difficulties related to the use of PDT for clinical applications are due to the intrinsic processes (absorption and scattering) of the light in penetrating tissues, the low concentration of the PS at the tumor site and the lack of oxygen in hypoxic tumor regions. In order to overcome the problem of light penetration in the tissues, different solutions have been proposed in the past. In this review we will focus on the recent use of Cerenkov radiation (CR) (**section 4**) and radioluminescence (**section 5**) as internal light sources to excite the PS molecules and, thus, to trigger ROS production inside the tumor region as shown in **Figure 1**. We would like to point out that we did not make any attempt in writing an exhaustive description on PDT and its clinical applications, for this please refer also to these reviews [6, 7].

## 2 HISTORICAL OVERVIEW OF PDT

The use of light to treat diseases can be dated back to antiquity. For example, the ancient Greek physicians suggested the use of sunlight to treat diseases [8]. The use of light was then abandoned for many centuries until the beginning of the 20th century. The modern era of PDT began in the early 1900s with the important contribution of the Danish physician, Niels Finsen. In his work he showed that it was possible to treat pustules or cutaneous tuberculosis using red or UV light. In 1903 Finsen was awarded the Nobel Prize for his discovery [9]. Almost at the same time, the possibility of using light in combination with a drug to kill cells was suggested by Oscar Raab, a medical student at the time. During a thunderstorm, Raab observed that the combination of acridine and light was lethal to *Infusoria*. Starting from this findings his supervisor Herman von Tappeiner showed the important role of oxygen in the reaction, and in 1904, he introduced the term *photodynamic action* [10]. The use of the term *dynamic* did not convince von Tappeiner himself and there has been some attempts (with no success) to modify it into photochemotherapy. A very important advancement in PDT was that hematoporphyrin and its derivative (HPD) was a powerful PS with a strong affinity with neoplastic tissues [11]. Based on this result a clinical trial was performed at Roswell Park Cancer Institute in 1978 [2, 12]. The properties of HPD as PS were improved by purifying the compound to extract the more active monomers, currently known with the commercial name of Photofrin® [2]. Photofrin absorbs light at 400 nm (Soret band) and 630 nm (Q-band) [13]. Contemporary PS are designed with an absorption range of 650–800 nm, longer wavelength range reduces the absorption and scattering properties of the tissues. Wavelengths longer than 800 nm can not be used because in this case the photon energy is too small to induce the generation of singlet oxygen [6]. As will be discussed in **sections 4 and 5** the problem of tissue absorption and scattering can be reduced using CR or radioluminescence as internal light sources.

## 3 PDT MECHANISMS

The starting step of PDT is the absorption of an optical photon by the PS leading to the molecule transition from the singlet state  $S_0$  to the excited singlet state  $S_1$  as shown in **Figure 2**. This excited singlet state has a nanosecond lifetime leading to the emission of the excess energy in the form of fluorescence and heat. It can be also converted to an excited triplet state  $T_1$  by means of intersystem crossing [6]. The triplet state has a longer microsecond lifetime and more importantly is chemically more reactive than the  $S_1$  singlet state. Two different types of reactions can take place. In Type I process an electron of the PS is transferred by reacting with a nearby substrate forming ROS. The Type II reaction involves an energy and spin transfer from the PS triplet state  $T_1$  to the ground-state triplet oxygen to generate ( $^1O_2$ ). Both types of reactions can take place and contribute to the efficiency of PDT. This efficiency is also dependent on the local oxygen concentration, which often decreases during the course of treatment. Unfortunately the oxygen concentration is lower in

hypoxic tumor regions, thus, reducing the cytotoxic effect of PDT. To overcome this problem different compounds like: catalase [14], Platinum nanozymes [15], Gold nanorods [16] were introduced to decompose intracellular hydrogen peroxide into  $H_2$  and  $O_2$  that is then delivered to the tumor.

## 4 PDT USING CERENKOV LIGHT SOURCES

### 4.1 Cerenkov Radiation Production

Before addressing the use of Cerenkov light as a source for PDT it is useful to describe how CR radiation is produced considering its peculiar nature [17–19]. CR is produced when a charged particle travels through a dielectric medium inducing a local polarization, more precisely the atoms behave like elementary dipoles as shown in **Figure 3**. If the particle speed  $v$  exceeds the speed of light  $c$  in a medium with a refracting index  $n$ , or more precisely when  $v \geq c/n$ , the polarization field becomes asymmetric along the particle track, producing a field at larger distances from the particle path. A very peculiar property of CR is the characteristic emission angle  $\theta$  obtained using the following equation:

$$\cos(\theta) = 1/\beta n \quad (1)$$

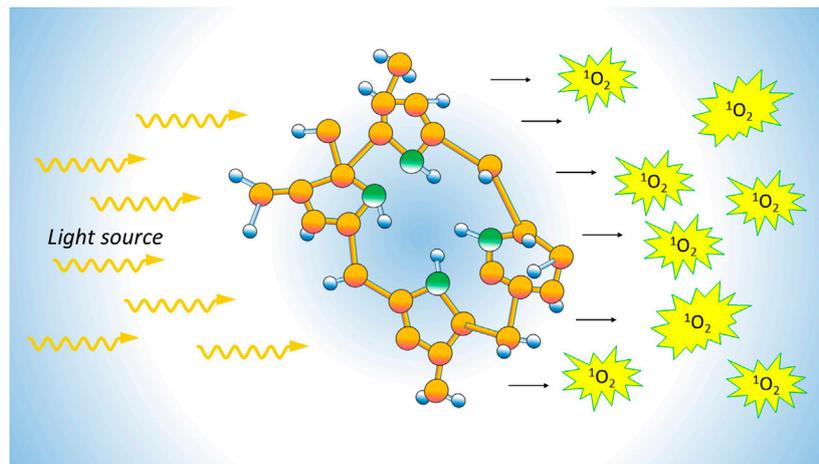
where  $\beta = v/c$ . The number  $N$  of the produced Cerenkov photons per unit of length  $l$  and wavelength  $\lambda$  can be determined from:

$$\frac{dN}{dld\lambda} = 2\pi\alpha \left(1 - \frac{1}{\beta^2 n^2}\right) \frac{1}{\lambda^2} \quad (2)$$

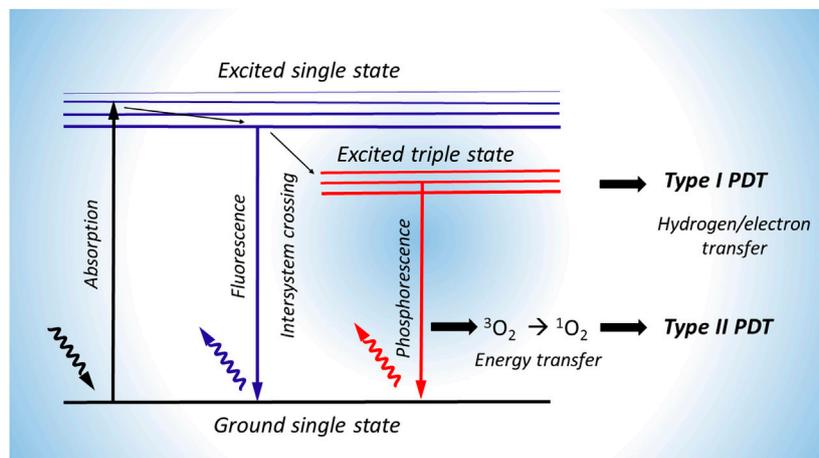
where  $\alpha \approx 1/137$  is the fine structure constant. As can be noticed from **Eq. 2** the spectrum of CR is continuous and more intense at lower wavelengths (e.g., UV and blue region) and for  $\beta \approx 1$  (e.g., relativistic particles). The shape is thus not dependent upon the particle energy, however particles with higher energies produce more Cerenkov photons. In order to find the total number of Cerenkov photons emitted by different radioisotopes [20] performed a series of Monte Carlo simulations and determined that  $^{18}F$  (endpoint energy = 633 keV) and  $^{90}Y$  (endpoint energy = 2.28 MeV), emits in water ( $n = 1.33$ ) per decay respectively a total of 1.4 and 57 photons in the wavelength range 400–800 nm. In order to improve the efficacy of PDT, radioisotopes with a longer half life and higher endpoint energy like:  $^{90}Y$ ,  $^{89}Zr$ ,  $^{68}Ga$ ,  $^{32}P$  should be chosen to increase the local ROS production.

### 4.2 PDT Using Isotopes Emitting Cerenkov Radiation

As mentioned in **section 1**, one of the main problems of PDT is the delivery of light to the PS due to the intrinsic absorption and scattering properties of the tissues. One solution is to use CR created within the tumor as the light source. The main advantage of using Cerenkov sources is the possibility of having in the same location (e.g., tumor site) both the PS and the light source for PDT. The local light emission feature of Cerenkov sources has been exploited also in endoscopic imaging as shown by [21]. Another important feature of CR is its spectrum, which is



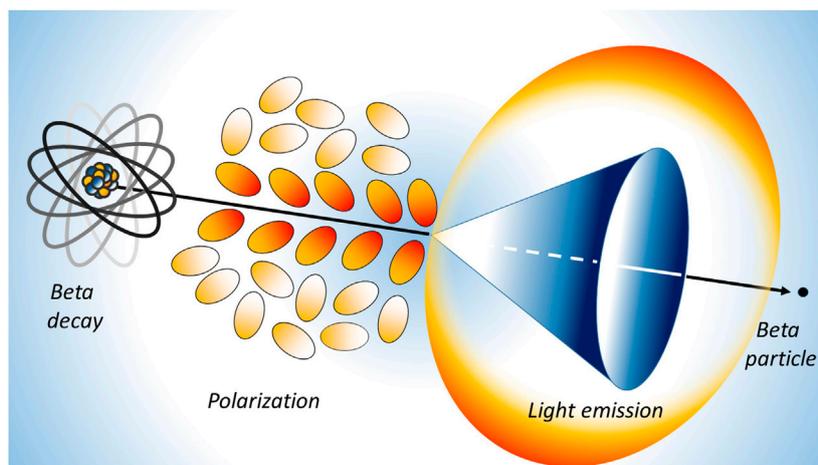
**FIGURE 1** | The figure describes the basic steps of PDT that is based on the induced photo conversion of a PS that trigger the production of cytotoxic reactive oxygen species leading to the killing of tumor cells.



**FIGURE 2** | Scheme of the different photosensitization processes after the absorption of a photon by the PS. Two different types of reactions can take place, in Type I process an electron of the PS is transfer by reacting with a nearby substrate forming ROS. The type II reaction involves an energy transferred from the PS triplet state  $T_1$  to the ground state ground-state triplet oxygen ( $^3O_2$ ) to generate ( $^1O_2$ ).

weighted to the UV and blue regions where some PS absorb strongly. In a preclinical proof-of-principle study with a breast cancer animal model expressing luciferase [22] investigated the possible role of  $^{18}F$ FDG, a widely used positron emission tomography (PET) tracer, as a internal light source to photoactivate caged luciferin. No PDT was performed in this case. In particular they used luciferin 1-(4,5-dimethoxy-2-nitrophenyl) ethyl ester (DMNP-luciferin) to demonstrate *in vitro* and *in vivo* the effect of the CR emitted by  $^{18}F$ FDG as a possible source to activate luciferin. *In vitro* results obtained using breast adenocarcinoma cells (MDA-MB-231-luc-D3H1) showed that the CR emitted by  $^{18}F$ FDG induced similar photoactivation to UV light at 365 nm. As described in [23] transferrin-coated  $TiO_2$  nanoparticles in combination with PET radionuclides like  $^{18}F$  and  $^{64}Cu$  can be used to perform PDT

*in vitro* and *in vivo*. Cell viability was reduced significantly *in vitro* with respect to control groups when combining radionuclides with the PS. Similar results were observed *in vivo* with an aggressive HT1080 subcutaneous mouse tumor model. After 12 days from the start of the treatment, the tumor volume dropped to almost zero while the tumor volumes of the untreated groups were in the 400–600 mm<sup>3</sup> range. Histological analysis confirmed extensive tumor necrotic regions [24]. developed an analytical model with the goal of combining the physics of Cerenkov light production, scattering, and the mechanisms of PDT reaction when using  $TiO_2$  NPs as PS. Their model has been validated using the experimental data found in [23, 25]. The main result of [24] was the establishment of a mechanistic description based on the Frank-Tamm theory of CR production (see Eq. 2) and the



**FIGURE 3** | Schematic representation of the Cerenkov light emission process. A beta particle, emitted by a radionuclide, travels in a medium faster than the light speed in the medium itself. The fast de-polarization of the molecules produces a cone of light known as Cerenkov radiation.

photocatalytic generation of free radicals. It has been found that the ROS production depends on the isotope activity (MBq), the NPs concentration (mol/L) and diameter (in this case 25 nm). The model agrees well with the experimental data found in the literature and possibly can be applied to other types of radionuclides, PS, or can be used to optimize the NPs size. Recently [26] proposed the combination of Doxorubicin (DOX), a chemotherapeutic agent that has been used to treat different types of haematological and solid cancers, with the CR emitted by  $^{18}\text{F}$ . Unfortunately DOX has severe side effects like liver damage or cardiotoxicity, however its alternative use as PS was proposed by [27] after a strong reduction of the injected dose. The interesting feature of DOX is its excitation band centered at 480 nm that matches well the spectrum of the CR as described in **section 4.1**. The cytotoxic effect of such combination was evaluated *in vitro* using a breast cancer cell line (T47D). It was shown that cell viability was reduced up to 85% by increasing  $^{18}\text{F}$  activity ( $=140\ \mu\text{Ci}$ ) in combination with DOX. In order to increase the Cerenkov light yield [13] developed mesoporous silica nanoparticles (HMSNs) able to encapsulate a second-generation PS like chlorin e6 (Ce6) [28] and  $^{89}\text{Zr}$ . The Chlorin e6 (Ce6) shows interesting PS properties as high efficiency and low dark toxicity, and, at the same time, the use of  $^{89}\text{Zr}$  allows a greater Cerenkov light yield production having an higher beta endpoint energy (909 keV) and also a longer half life (78.4 h) compared to  $^{18}\text{F}$  and  $^{64}\text{Cu}$ . In their work [13] found with *in vitro* experiments a strong dose-dependent cell viability reduction depending on the concentration of Ce6 and  $^{89}\text{Zr}$ . In particular cell viability was reduced to 10% when using an  $^{89}\text{Zr}$  activity of 1 MBq and a concentration of HMSN-Ce6 equal to  $40\ \mu\text{M}$ . An *in vivo* study was performed using 4T1 tumors in Balb/c mice with a size of  $\approx 200\ \text{mm}^3$  and showed a strong tumor control for 14 days after a single dose of [ $^{89}\text{Zr}$ ]HMSN-Ce6 (15 MBq,  $50\ \mu\text{l}$ ). The damage to the tumor tissue was also confirmed by H&E stained tumor slices taken 7 days after treatment. In [29] a novel hybrid tetramodal imaging

nanotheranostics tool based on  $^{89}\text{Zr}$  for photothermal and PDT was described. More precisely they developed a nanoconstruct by assembling copper sulfide (CuS) nanoparticles on the surface of  $^{89}\text{Zr}$ -labeled hollow mesoporous silica nanoshells filled with porphyrin molecules. In order to investigate *in vivo* the tetramodal properties of [ $^{89}\text{Zr}$ ]CSNC-PEG $_{10k}$  its biodistribution was imaged using: PET, fluorescence, Cerenkov luminescence and CR energy transfer imaging. It has been found for a subcutaneous 4T1 murine breast tumor model using these imaging techniques, that [ $^{89}\text{Zr}$ ]CSNC-PEG $_{10k}$  showed a specific uptake even 15 days after intravenous injection. Recently [30] investigated the use of  $^{131}\text{I}$ -bovine serum albumin loaded into chlorin-lipid nanovesicle cavity. The main goal was to combine the anti-tumor effect of  $^{131}\text{I}$  radiotherapy with PDT induced by Cerenkov light emitted by  $^{131}\text{I}$ . Their results showed a reduced lung tumor growth in mice. In particular the survival curve was significantly prolonged and, at the same time, histopathology on different organs found no strong side effects. The use of  $^{131}\text{I}$  as both radiotherapy and PDT Cerenkov light source was also investigated by [31] using a Zn based nanoplatfrom. Both *in vitro* and *in vivo* experiments were carried out using 4T1 cell line. Cellular viability was measured and it has been found equal to 29.2% using an activity of  $40\ \mu\text{Ci}$ . The same cell line has been used to establish a tumor model with a subcutaneous injection of  $5 \times 10^6$  cells. After 14 days from the treatment the average tumor volume of the treated mice was about  $180\ \text{mm}^3$  while the volume of the untreated group was  $850\ \text{mm}^3$ . *Ex vivo* analysis using hematoxylin and eosin staining was performed and showed more tumor cells with shranked nuclei and, at the same time no particular damage to other organs like: heart, liver, spleen and kidney. Using the same 4T1 cell line [25] investigated the use of  $^{68}\text{Ga}$  (endpoint energy 1.899 MeV) as a more intense CR source to activate a dextran-modified  $\text{TiO}_2$  PS. A comparison between  $^{18}\text{F}$ -FDG, and  $^{68}\text{Ga}$ -labeled bovine serum albumin was performed, and it has been found a stronger inhibitor of the

growth of 4T1 cells and stronger DNA damage to tumor cells when using  $^{68}\text{Ga}$  to excite the PS. Another interesting isotope for PDT is  $^{90}\text{Y}$  considering its half-life of 64.1 h and the endpoint energy of 2.278 MeV leading to an higher number of emitted Cerenkov photons as described in **section 4.1**. The use of  $^{90}\text{Y}$  as PDT source was investigated *in vitro* by [32] with the C6 glioma cell line in combination with the widely used aminolevulinic acid (ALA) or the porphyrin-based PS TTPS<sub>2a</sub>. It has been found that the cell viability was reduced respectively down to 60 and 45% for the combination of  $^{90}\text{Y}$  with ALA or TTPS<sub>2a</sub> using an activity equal to 60  $\mu\text{Ci}$  in the well.

## 5 PDT USING RADIOLUMINESCENCE LIGHT

In **section 4** we focused our attention on the use of Cerenkov light emitted by beta plus or beta minus radioisotopes as a possible internal source for PDT. Another possible source that has been investigated is radioluminescence light that is normally produced by the interaction of ionizing radiations with a scintillator material. However it has been recently shown that radioluminescence light can also be produced in non scintillating material like: Plexiglas, glass [33] small animals [34] or *ex vivo tissue* [35]. An alternative solution to solve the problem of light tissue penetration is the use of an external X-rays source in combination with scintillating nanoparticles (ScNPs). This type of NPs behave as a local X-ray converter, more precisely the ScNPs are able to absorb X-ray and then produce scintillation light that can excite the PS and induce ROS production. An example of the use of Fluoride-based material for X-ray excited optical imaging can be found in [36] where NaGdF<sub>4</sub>:Eu<sup>3+</sup> NPs were used to convert X-ray into optical photons within the 600–700 nm range. By using this NPs it was possible to perform X-ray luminescence optical tomography. Another interesting approach to perform X-ray excited optical imaging is the use of quantum dots (QDs) to convert X-ray into visible photons. A recent example of this method was described in [37] where core-type CdTe has been used to convert X-rays with energies between 20 and 120 keV. Phantom experiments were carried out by filling a syringe with 0.1 ml aqueous suspensions of 715 nm-emitting CdTe QDs, with 0, 0.005, 0.016, 0.13, and 0.46 mg of CdTe. The syringe was then irradiated at: 55, 70, and 100 kVp resulting in intense photons emission that could be imaged in 1 s using a sensitive EM-CCD detector. Mouse imaging was performed by irradiating at 55 kV *p* and 500 mA a mouse subcutaneously injected with 0.46 mg of CdTe in back of the neck. The corresponding optical emission image shows a good match with the corresponding anatomical region. One of the first attempts to use scintillating light for PDT *in vivo* was described in [38]. In this paper the authors combined porphyrins with scintillating rare Earth NPs excited by X-rays in order to obtain X-rays excited optical luminescence (XEOL). The interesting feature of rare Earth NPs is the emission peak at around 400 nm that matches well with the strong absorption peak

(Soret band) of porphyrins. In [39] it has been reported the synthesis of LaF<sub>3</sub>:Tb<sup>3+</sup>-meso-tetra(4-carboxyphenyl)porphine (MTCP) NPs conjugates with the addition of folic acid to improve the targeting to folate receptors. It has been found that this type of NPs can potentially be used to trigger PDT to treat deep cancer. In [40] a micellar system based on amphiphilic lanthanide chelates that includes the Hyp PS inside the core has been developed. This micellar system, once excited by X-ray, was able to produce singlet oxygen production through PDT. It is also important to note that the inclusion of lanthanides in the structure allow the detection of the micellar systems using optical or magnetic resonance imaging. More recently [41] investigated the use of titania NP in particular dumbbell-like Au-TiO<sub>2</sub> NPs (DATs). This type of NP can be activated by X-rays resulting in the production of secondary photons or electrons, leading to ROS production.

## 6 DISCUSSION AND CONCLUSION

In this short review we discussed the potential use of Cerenkov and radioluminescence light as internal sources for PDT. The main advantages of using these excitation sources are reduced light scattering and absorption by the tissue and, at the same time, a lower cytotoxic effect on normal tissue since most of the light is emitted inside the target (e.g., tumor) region. The use of Cerenkov and radioluminescence light for PDT significantly expand the more common applications as: preclinical [39, 42, 43], clinical [44] and *ex vivo* [45] diagnostic tool. An important aspect that should be further exploited regarding the use of Cerenkov and radioluminescence light sources, is the possibility of enhancing the cytotoxic effect of PDT. This can be achieved by inducing further damage using the so called radiotherapy cross fire effect obtained with beta emitters like:  $^{90}\text{Y}$ ,  $^{177}\text{Lu}$  or  $^{32}\text{P}$  [46]. Another interesting research line should focus on a better integration between the PS with different radioisotopes in order to optimize photon absorption, at the same time reducing the distances and, thus, possible photons loss. PDT based on radioluminescence light should be further exploited by developing more efficient ScNPs, for example. In conclusion the use of Cerenkov and radioluminescence light sources is a promising clinical approach for PDT considering also the possibility of using several radiopharmaceuticals already approved for clinical use. Secondly, and more importantly, the combination of radiotherapy and PDT can have an important local synergistic effect for cancer treatment, while possibly reducing side effects to normal tissues.

## AUTHOR CONTRIBUTIONS

AS performed the literature review and wrote the manuscript. FB contributed to manuscript revision and figures. All authors have read and agreed to the published version of the article.

## REFERENCES

- Oniszczuk A, Wojtunik-Kulesza KA, Oniszczuk T, Kasprzak K. The potential of photodynamic therapy (PDT)-Experimental investigations and clinical use. *Biomed Pharmacother* (2016) 83:912–29. doi:10.1016/j.biopha.2016.07.058
- Dougherty TJ. A brief history of clinical photodynamic therapy development at roswell park cancer institute. *J Clin Laser Med Surg* (1996) 14:219–21. doi:10.1089/clm.1996.14.219
- Baskaran R, Lee J, Yang SG. Clinical development of photodynamic agents and therapeutic applications. *Biomater Res* (2018) 22:25. doi:10.1186/s40824-018-0140-z
- Chen C, Wang J, Li X, Liu X, Han X. Recent advances in developing photosensitizers for photodynamic cancer therapy. *Comb Chem High Throughput Screen* (2017) 20: 414–22. doi:10.2174/1386207320666170113123132
- Zhang J, Jiang C, Figueiró Longo JP, Azevedo RB, Zhang H, Muehlmann LA. An updated overview on the development of new photosensitizers for anticancer photodynamic therapy. *Acta Pharmaceutica Sinica B* (2018a) 8:137–46. doi:10.1016/j.apsb.2017.09.003
- Agostinis P, Berg K, Cengel KA, Foster TH, Girotti AW, Gollnick SO, et al. Photodynamic therapy of cancer: an update. *CA: A Cancer J Clinicians* 61 (2011) 250–81. doi:10.3322/caac.20114
- Hahn K, Zhang Y, Mu C, Xu Q, Jing X, Wang D, et al. Versatile nanoplatforms with enhanced photodynamic therapy: designs and applications. *Theranostics* (2020) 10:7287–318. doi:10.7150/thno.46288
- Dang MD, Hill JS. A history of photodynamic therapy. *ANZ J Surg* (1991) 61: 340–8. doi:10.1111/j.1445-2197.1991.tb00230.x
- Ackroyd R, Kely C, Brown N, Reed M. The history of photodetection and photodynamic therapy. *Photochem Photobiol* (2001) 74:656–69. doi:10.1562/0031-8655(2001)074<0656:thopap>2.0.co;2
- Moan J, Peng Q. An outline of the hundred-year history of PDT. *Anticancer Res* (2003) 23:3591–600.
- Lipson RL, BALDES EJ, OLSEN AM. The use of a derivative of hematoporphyrin in tumor detection. *J Natl Cancer Inst* (1961) 26:1–11.
- Dougherty TJ, Kaufman JE, Goldfarb A, Weishaupt KR, Boyle D, Mittleman A. Photoradiation therapy for the treatment of malignant tumors. *Cancer Res* (1978) 38:2628–35.
- Kamkaew A, Cheng L, Goel S, Valdovinos HF, Barnhart TE, Liu Z, et al. Cerenkov radiation induced photodynamic therapy using chlorin e6-loaded hollow mesoporous silica nanoparticles. *ACS Appl Mater Inter* (2016) 8: 26630–7. doi:10.1021/acsami.6b10255
- Cai SZF, Yang G, Lim WQ, Verma A, Chen H, Thanabalu T, et al. Catalase-integrated hyaluronic acid as nanocarriers for enhanced photodynamic therapy in solid tumor. *ACS Nano* (2019) 13:4742–51. doi:10.1021/acsnano.9b01087
- Zhao Y, Wang F, Liu C, Wang Z, Kang L, Huang Y, et al. Nanozyme decorated metal-organic frameworks for enhanced photodynamic therapy. *ACS Nano* (2018b) 12:651–61. doi:10.1021/acsnano.7b07746
- Dong T, Shen X, Li L, Guan Z, Gao N, Yuan P, et al. Gold nanorods as dual photo-sensitizing and imaging agents for two-photon photodynamic therapy. *Nanoscale* (2012) 4:7712–9. doi:10.1039/C2NR32196C
- Yao JV. Cerenkov radiation and its applications. *Br J Appl Phys* (1955) 6: 227–32. doi:10.1088/0508-3443/6/7/301
- Spinelli AE, Boschi F. Novel biomedical applications of Cerenkov radiation and radioluminescence imaging. *Physica Med* (2015) 31:120–9. doi:10.1016/j.ejmp.2014.12.003
- Boschi F, Spinelli AE. Nanoparticles for cerenkov and radioluminescent light enhancement for imaging and radiotherapy. *Nanomaterials (Basel)* (2020) 10. doi:10.3390/nano10091771
- Mitchell GS, Gill RK, Boucher DL, Li C, Cherry SR. *In vivo* Cerenkov luminescence imaging: a new tool for molecular imaging. *Phil Trans R Soc A* (2011) 369:4605–19. doi:10.1098/rsta.2011.0271
- Zhang Z, Cai M, Bao C, Hu Z, Tian J. Endoscopic Cerenkov luminescence imaging and image-guided tumor resection on hepatocellular carcinoma-bearing mouse models. *Nanomedicine: Nanotechnology, Biol Med* (2019) 17:62–70. doi:10.1016/j.nano.2018.12.017
- Ran C, Zhang Z, Hooker J, Moore A. *In vivo* photoactivation without “light”: use of cerenkov radiation to overcome the penetration limit of light. *Mol Imaging Biol* (2012) 14:156–62. doi:10.1007/s11307-011-0489-z
- Kotagiri N, Sudlow GP, Akers WJ, Achilefu S. Breaking the depth dependency of phototherapy with Cerenkov radiation and low-radiance-responsive nanophotosensitizers. *Nat Nanotech* (2015) 10:370–9. doi:10.1038/nnano.2015.17
- Kavadiya S, Biswas P. Design of cerenkov radiation-assisted photoactivation of TiO<sub>2</sub> nanoparticles and reactive oxygen species generation for cancer treatment. *J Nucl Med* (2019) 60:702–9. doi:10.2967/jnumed.118.215608
- Duan D, Liu H, Xu Y, Han Y, Xu M, Zhang Z, et al. Activating tio<sub>2</sub> nanoparticles: gallium-68 serves as a high-yield photon emitter for cerenkov-induced photodynamic therapy. *ACS Appl Mater Inter* (2018) 10: 5278–86. doi:10.1021/acsami.7b17902
- Liu HA, Aranda-Lara L, Morales-Ávila E, Torres-García E, Camacho-López MÁ, Sánchez-Holguín M, et al. *In vitro* irradiation of doxorubicin with 18F-fdg cerenkov radiation and its potential application as a theragnostic system. *J Photochem Photobiol B: Biol* (2020) 210:111961. doi:10.1016/j.jphotobiol.2020.111961
- Luna-Gutiérrez KW, Gao J-P, Sharma T. Photodynamic enhancement of doxorubicin cytotoxicity. *Cancer Chemother Pharmacol* (1994) 35:17–20. doi:10.1007/bf00686279
- Martynenko IV, Kuznetsova VA, Orlova O, Kanaev PA, Maslov VG, Loudon A, et al. Chlorin e6-ZnSe/ZnS quantum dots based system as reagent for photodynamic therapy. *Nanotechnology* (2015) 26:055102. doi:10.1088/0957-4484/26/5/055102
- Goel S, Ferreira CA, Chen F, Ellison PA, Siamof CM, Barnhart TE, et al. Activatable hybrid nanotheranostics for tetramodal imaging and synergistic photothermal/photodynamic therapy. *Adv Mater* (2018) 30:1704367. doi:10.1002/adma.201704367
- Cai P, Yang W, He Z, Jia H, Wang H, Zhao W, et al. A chlorin-lipid nanovesicle nucleus drug for amplified therapeutic effects of lung cancer by internal radiotherapy combined with the cerenkov radiation-induced photodynamic therapy. *Biomater Sci* (2020) 8:4841–51. doi:10.1039/d0bm00778a
- Gao Q, Liu N, Hou Z, Shi J, Su X, Sun X. Radiiodinated persistent luminescence nanoplatform for radiation-induced photodynamic therapy and radiotherapy. *Adv Healthc Mater.* (2020) 10:2000802. doi:10.1002/adhm.202000802
- Hartl BA, Hirschberg H, Marcu L, Cherry SR. Activating photodynamic therapy *in vitro* with cerenkov radiation generated from yttrium-90. *J Environ Pathol Toxicol Oncol* (2016) 35:185–92. doi:10.1615/jenviroxpathtoxicoloncol.2016016903
- Pagliazzi M, Boschi F, Spinelli AE. Imaging of luminescence induced by beta and gamma emitters in conventional non-scintillating materials. *RSC Adv* (2014) 4:13687–92. doi:10.1039/C3RA47102K
- Spinelli AE, Lo Meo S, Calandrino R, Sbarbati A, Boschi F. Optical imaging of Tc-99m-based tracers: *in vitro* and *in vivo* results. *J Biomed Opt* (2011) 16: 116023. doi:10.1117/1.3653963
- Ackerman NL, Boschi F, Spinelli AE. Monte Carlo simulations support non-Cerenkov radioluminescence production in tissue. *J Biomed Opt* (2017) 22: 1–11. doi:10.1117/1.jbo.22.8.086002
- Sudheendra L, Das GK, Li C, Stark D, Cena J, Cherry S, et al. NaGdF<sub>4</sub>:Eu<sup>3+</sup> nanoparticles for enhanced X-ray excited optical imaging. *Chem Mater* (2014) 26:1881–8. doi:10.1021/cm404044n
- Kennedy SG, Butler MN, Adeyemi SS, Kalber T, Patrick PS, Zaw Thin M, et al. Imaging of X-ray-excited emissions from quantum dots and biological tissue in whole mouse. *Sci Rep* (2019) 9:19223. doi:10.1038/s41598-019-55769-5
- Chen W, Zhang J. Using nanoparticles to enable simultaneous radiation and photodynamic therapies for cancer treatment. *J Nanosci Nanotech* (2006) 6: 1159–66. doi:10.1166/jnn.2006.327
- Liu Y, Chen W, Wang S, Joly AG. Investigation of water-soluble x-ray luminescence nanoparticles for photodynamic activation. *Appl Phys Lett* (2008) 92:043901. doi:10.1063/1.2835701
- Kascakova S, Giuliani A, Lacerda S, Pallier A, Mercere P, Toth E, et al. X-ray-induced radiophotodynamic therapy (RPDT) using lanthanide micelles: beyond depth limitations. *Nano Research* (2015) 8:2373–9. doi:10.1007/s12274-015-0747-5
- Cheng K, Sano M, Jenkins CH, Zhang G, Vernekohl D, Zhao W, et al. Synergistically enhancing the therapeutic effect of radiation therapy with radiation activatable and reactive oxygen species-releasing nanostructures. *ACS Nano* (2018) 12:4946–58. doi:10.1021/acsnano.8b02038

42. Wei F, Calderan L, D'Ambrosio D, Marengo M, Fenzi A, Calandrino R, et al. *In vivo* 18F-FDG tumour uptake measurements in small animals using Cerenkov radiation. *Eur J Nucl Med Mol Imaging* (2011) 38:120–7. doi:10.1007/s00259-010-1630-y
43. Sbarbati W, Qin W, Hu Z, Suo Y, Zhao R, Ma X, et al. Comparison of Cerenkov luminescence imaging (CLI) and gamma camera imaging for visualization of let-7 expression in lung adenocarcinoma A549 Cells. *Nucl Med Biol* (2012) 39: 948–53. doi:10.1016/j.nucmedbio.2012.05.004
44. Ma AE, Ferdeghini M, Cavedon C, Zivelonghi E, Calandrino R, Fenzi A, et al. First human cerenkography. *J Biomed Opt* (2013) 18 020502. doi:10.1117/1.jbo.18.2.020502
45. Sbarbati AE, Schiariti MP, Grana CM, Ferrari M, Cremonesi M, Boschi F. Cerenkov and radioluminescence imaging of brain tumor specimens during neurosurgery. *J Biomed Opt* (2016) 21:050502. doi:10.1117/1.jbo.21.5.050502
46. Galiè M, Boschi F, Scambi I, Merigo F, Marzola P, Altabella L, et al. Theranostic role of <sup>32</sup>P-ATP as radiopharmaceutical for the induction of massive cell death within avascular tumor core. *Theranostics* (2017) 7: 4399–409. doi:10.7150/thno.21403

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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