



## Diabetic Cardiomyopathy: Impact of Biological Sex on Disease Development and Molecular Signatures

#### Ryan Toedebusch<sup>1,2</sup>, Anthony Belenchia<sup>2,3</sup> and Lakshmi Pulakat<sup>1,2,3\*</sup>

<sup>1</sup> Cardiovascular Medicine Division, Department of Medicine, University of Missouri, Columbia, MO, United States, <sup>2</sup> Dalton Cardiovascular Research Center, University of Missouri, Columbia, MO, United States, <sup>3</sup> Department of Nutrition and Exercise Physiology, University of Missouri, Columbia, MO, United States

OPEN ACCESS

#### Edited by:

Miles J. De Blasio, Baker Heart and Diabetes Institute, Australia

#### Reviewed by:

Tamara Paravicini, RMIT University, Australia Silvia Del Ry, Istituto di Fisiologia Clinica (CNR), Italy

> \*Correspondence: Lakshmi Pulakat pulakatl@health.missouri.edu

#### Specialty section:

This article was submitted to Integrative Physiology, a section of the journal Frontiers in Physiology

Received: 01 February 2018 Accepted: 11 April 2018 Published: 03 May 2018

#### Citation:

Toedebusch R, Belenchia A and Pulakat L (2018) Diabetic Cardiomyopathy: Impact of Biological Sex on Disease Development and Molecular Signatures. Front. Physiol. 9:453. doi: 10.3389/fphys.2018.00453 Diabetic cardiomyopathy refers to a unique set of heart-specific pathological variables induced by hyperglycemia and insulin resistance. Given that cardiovascular disease (CVD) is the leading cause of death in the world, and type 2 diabetes incidence continues to rise, understanding the complex interplay between these two morbidities and developing novel therapeutic strategies is vital. Two hallmark characteristics specific to diabetic cardiomyopathy are diastolic dysfunction and cardiac structural maladaptations, arising from cardiac cellular responses to the complex toxicity induced by hyperglycemia with or without hyperinsulinemia. While type 2 diabetes is more prevalent in men compared to women, cardiovascular risk is higher in diabetic women than in diabetic men, suggesting that diabetic women take a steeper path to cardiomyopathy and heart failure. Accumulating evidence from randomized clinical trials indicate that although pre-menopausal women have lower risk of CVDs, compared to age-matched men, this advantage is lost in diabetic pre-menopausal women, which suggests estrogen availability does not protect from increased cardiovascular risk. Notably, few human studies have assessed molecular and cellular mechanisms regarding similarities and differences in the progression of diabetic cardiomyopathy in men versus women. Additionally, most pre-clinical rodent studies fail to include female animals, leaving a void in available data to truly understand the impact of biological sex differences in diabetes-induced dysfunction of cardiovascular cells. Elegant reviews in the past have discussed in detail the roles of estrogen-mediated signaling in cardiovascular protection, sex differences associated with telomerase activity in the heart, and cardiac responses to exercise. In this review, we focus on the emerging cellular and molecular markers that define sex differences in diabetic cardiomyopathy based on the recent clinical and pre-clinical evidence. We also discuss miR-208a, MED13, and AT2R, which may provide new therapeutic targets with hopes to develop novel treatment paradigms to treat diabetic cardiomyopathy uniquely between men and women.

Keywords: cardiac hypertrophy, fibrosis, cardiovascular disease, diabetes, microRNA, estrogen, sex hormones

## INTRODUCTION

Cardiovascular diseases (CVDs) are the number one killer of both men and women worldwide (World Health Organization [WHO], 2017). Type 2 diabetes mellitus (T2DM) is an independent risk factor for CVD (Lotufo et al., 2001; Abdul-Ghani et al., 2017; Archundia Herrera et al., 2017; Davis et al., 2017). Diabetic patients have a two- to fourfold increased risk for CVD development, and undoubtedly, the increased prevalence of T2DM has led to more documented cases of cardiovascular complications (Kannel et al., 1974; Kannel and McGee, 1979; Martin-Timon et al., 2014; Bertoluci and Rocha, 2017). Indeed, while T2DM contributes to overt CVD, it has been documented to be responsible for a unique set of cardiac abnormalities (Miki et al., 2013; Kain and Halade, 2017; Varma et al., 2017), referred to as diabetic cardiomyopathy. Several of the cellular mechanisms documented to underlie the development of contractile dysfunction in diabetic cardiomyopathy include: impaired excitation-contraction coupling, inefficient energy production, reduced coronary flow reserve, and fibrotic remodeling (Miki et al., 2013). Not surprising, many of these mechanisms are also observed in hypertrophic and dilated cardiomyopathies, independent of diabetes, promoting the equivocation and questioning a universal clinical definition for diabetic cardiomyopathy. In further support of the ambiguity, a review by Holscher et al. (2016) reported that "instead of being a cardiomyopathy in the classical sense though, diabetic cardiomyopathy represents a combination of molecular myocardial abnormalities that predispose for the development of myocardial dysfunction." Additional questions regarding diabetic cardiomyopathy and its progression in both insulindependent and insulin-independent diabetic patients remain unanswered. Although a ubiquitous definition has yet been agreed upon, diabetic cardiomyopathy is generally accepted to refer to diabetes-associated changes in the structure and function of the myocardium, independent of other peripheral disease, in otherwise healthy diabetic patients (Dandamudi et al., 2014).

The first documented associations between diabetes mellitus and any cardiovascular complications appeared in the first half of the 20th century (Liebow and Hellerstein, 1949). However, the first reports of what we know today as "diabetic cardiomyopathy" did not appear until the early 1970s when it was documented that diabetic patients with congestive heart failure, who had no evidence of coronary atherosclerosis, had abnormal fibrosis patterns within the myocardium (Rubler et al., 1972). Shortly after, the Framingham Study provided epidemiological data supporting the notion that a specific cardiomyopathy does exist in diabetics, and that cardiovascular mortality was threefold higher in participants with diabetes (Kannel et al., 1974). In the  $\sim$ 40 years since these seminal studies appeared, increased incidence of T2DM, CVD, and the complex interactions between them have underscored the need for a better understanding of the pathology and possible treatments. Beyond epidemiological and associative data describing diabetic cardiomyopathy, the basic science and general understanding of its progression, incidence, and possible treatment(s) remain in their infancy. Nevertheless, mounting evidence suggests that the incidence of

The Framingham Heart Study first revealed that diabetic women have a 5.1-fold increase in heart failure, while diabetic men only have a 2.4-fold increase compared to non-diabetic women and men, respectively (Kannel et al., 1974). Since that time, several studies provided similar evidence that diabetic women have a greater relative risk of CVD compared to diabetic men (Heyden et al., 1980; Barrett-Connor et al., 1991; Simons et al., 1996; Folsom et al., 1997; Natarajan et al., 2003). Laverty et al. (2017) assessed gender differences in hospital admissions for major cardiovascular events in diabetics throughout England over 10 years between 2004 and 2014. They confirmed that diabetic women, compared to men, had increased hospital admission rates for acute myocardial incident (AMI), percutaneous coronary intervention (PCI), and coronary artery bypass grafting (CABG). Specifically, compared to nondiabetic women, diabetic women had a 4.3-fold increase in AMI, 4.4-fold increase in PCI, and 6.2-fold increase in CABG admissions, supporting the notion that diabetic females have an elevated incidence of CVD risk factors, compared to diabetic men. Further, pre-menopausal women usually present with CVD almost a decade later than males; however, this protection is not seen in pre-menopausal women diagnosed with T2DM (Norhammar and Schenck-Gustafsson, 2013).

Considering unique sex-based differences in physiology and pathophysiology, it is worth noting some of the most apparent sex-specific factors that underlie our understanding of the abovementioned observations. First, it has been documented that females have altered systemic glucose regulation, compared to males (Basu et al., 2006; Flanagan et al., 2006). For example, in one clinical study, it was shown that systemic insulin was higher after dextrose infusion in females, suggesting lower insulin-sensitivity at baseline in females compared to males (Flanagan et al., 2006). Second, pre-menopausal women demonstrate different fat distribution compared to men. In contrast to the central (visceral) adiposity observed in males, females tend to have peripheral fat distribution, contributing to higher insulin sensitivity at greater levels of body mass (Wannamethee et al., 2012). Due to this, women typically have increased metabolic disturbances (body mass index and insulin resistance) before overt T2DM diagnosis and therefore incur higher CVD risk (Logue et al., 2011; Gomez-Marcos et al., 2015). Third, and possibly the most important, yet commonly misunderstood, is the role that estrogen-related signaling within the female myocardium plays in disease progression and/or protection. Finally, a strong correlation is reported between the leukocyte telomere length (LTL) shortening and increase in metabolic syndrome components in females (Cheng et al., 2017). LTL shortening and low telomerase activity are shown to be associated with CVD, coronary artery disease, diabetes mellitus, cardiomyopathy, and all-cause mortality (Bar et al., 2014; Yeh and Wang, 2016; Sawhney et al., 2018). It is noteworthy that while telomerase activity in cardiomyocytes decreases in men with aging, it increases in women (Leri et al., 2000; Kajstura et al., 2010). However, how T2DM modulates this effect in the heart tissues of women versus men is currently unclear.

#### DIABETIC CARDIOMYOPATHY AND AN UNRESOLVED DEFINITION

Scientists and clinicians alike face challenges when studying the pathophysiology of diabetic cardiomyopathy. Diabetic cardiomyopathy commonly presents with one or more comorbidities known to exacerbate heart failure. Functionally, diabetic cardiomyopathy is characterized by diastolic dysfunction, defined as a defect in left ventricular relaxation leading to increased pressures and a subsequent impaired filling during diastole (Lorenzo-Almoros et al., 2017). In both type 1 (T1DM) and T2DM, diastolic dysfunction is largely considered a hallmark of diabetic cardiomyopathy, although some reports suggest that upon adjustment for comorbidities, diastolic dysfunction is not statistically significant (Wachter et al., 2007; Fontes-Carvalho et al., 2015). Stahrenberg et al. (2010) demonstrated that along the continuum of diabetic patients, higher HbA1c levels are associated with the severity of diastolic dysfunction, as measured by E/E', a non-invasive estimate of left atrial filling pressure that independently predicts primary cardiac events.

In diabetes, diastolic function as assessed by E/A ratio and hemodynamics has been shown to prematurely deteriorate compared to healthy controls. The E/A ratio represents the ratio of the E wave (peak blood flow velocity in early diastole) to the A wave (peak blood flow velocity in late diastole) caused by atrial contraction. The filling pattern, where there is a reduction in the E/A ratio along with prolongation of the deceleration time of E, indicates impaired relaxation. E/A values measured in young (20-32 years of age) T1DM males match that of healthy men at 50 years of age, suggesting that T1DM negatively effects diastolic function (Berkova et al., 2003). Two more recent reports (Jensen et al., 2014; Suran et al., 2016) using echocardiography evaluation of T1DM patients without known CVD, reported the presence of diastolic dysfunction. Indeed, one study assessed adolescent T1DM patients (mean duration of disease = 6 years) and found, both at rest and during exercise, these patients had reduced diastolic function (Gusso et al., 2012), evidenced by end diastolic volume (Holscher et al., 2016). In contrast, various other reports suggest that in long-term T1DM patients, evidence for diastolic dysfunction is lacking (Zarich et al., 1988; Romanens et al., 1999), indicating that T1DM's ability to cause diastolic dysfunction may be a factor of duration of disease, age of onset, management, and/or environment. The deleterious effects of diabetes on myocardial parameters are not synonymous between patients with T1DM versus T2DM, adding to the vagueness of diabetic cardiomyopathy. For example, T1DM is mostly associated with hyperglycemia, oxidative stress, and resultant myocardial fibrosis and average patient population with T1DM is younger than that with T2DM (Figure 1). In contrast, T2DM is linked to hyperinsulinemia, insulin resistance, obesity, and cardiomyocyte hypertrophy (Lorenzo-Almoros et al., 2017).

A 2003 publication assessed the fact that T1DM patients may present with diabetic autonomic neuropathy (DAN),



**FIGURE 1** Progression of T1DM and T2DM and development of diabetic cardiomyopathy. (A) Illustrates how an immune mediated response in T1DM destroys beta cells within pancreatic islets, leading to hyperglycemia, resulting in cardiac damage which promotes the development of diabetic cardiomyopathy. (B) Shows how environmental influences (diet, inactivity) lead to insulin resistance, hyperinsulinemia, beta cell burnout, and subsequent dysregulation of glucose, leading to hyperglycemia, cardiac damage, and ultimately diabetic cardiomyopathy. Noteworthy is the arrow indicating the progression of diabetic cardiomyopathy and that it occurs and progresses uniquely in certain populations of individuals, namely male and females.

and hypothesized that this may account for diastolic and/or systolic dysfunction (Didangelos et al., 2003). Using radionuclide ventriculography, it was found that T1DM patients had reduced diastolic parameters, including atrial contribution to ventricular filling, peak filling rate, first third filling fraction, and time to peak filling, suggesting diastolic dysfunction (Didangelos et al., 2003). The lack of cardiac dysfunction in long-term T1DM patients may also be related to permanent treatment with exogenous insulin. Others have suggested that myocardial overload and increased peripheral resistance resultant from exogenous insulin may be responsible for the observed diastolic dysfunction, rather than being symptoms of diabetic cardiomyopathy (Holscher et al., 2016). In summary, numerous reports support the fact that diastolic dysfunction is the defining characteristic of diabetic cardiomyopathy in both T1DM and T2DM patients. However, until additional long-term studies are performed, debate will continue until a universal definition is settled upon. Figure 2 summarizes the evidence for and against the (current) understanding and existence of diabetic cardiomyopathy.

Alternatively, when considering rodent models of T1DM, clinical data regarding diastolic dysfunction has been corroborated (Bugger and Abel, 2009). On a cellular and molecular level, rodent models of diabetic cardiomyopathy have helped develop our understanding of myocardial alterations. Due to their resistance to atherosclerosis, they provide a good model to study the effects of diabetes on the myocardium, independent of peripheral complications (Severson, 2004). Through the use of rodent models, mechanisms underlying diabetic cardiomyopathy, including cardiac fibrosis, mitochondrial dysfunction, oxidative stress, lipotoxicity, and metabolic inflexibility have been identified (Schilling and Mann, 2014).

Importantly, there are a few human studies that lend support to rodent findings, including mitochondrial dysfunction, fibrosis, oxidative stress, and metabolic inflexibility (Peterson et al., 2004; Anderson et al., 2009), suggesting that rodent models may be a sufficient means to study diabetic cardiomyopathy progression. For example, consistent with observations from diabetic rat models, intra-myocardial lipid accumulation is seen in the failing heart of diabetic patients. Moreover, intra-myocardial lipid accumulation in T2DM patients correlates with impaired cardiomyocyte contractility and their inability to adapt after myocardial infarction, a phenomenon seen in diabetic rats (Sharma et al., 2004; Borisov et al., 2008). It was also reported that in obese, insulin-resistant men, abnormal left ventricular energy metabolism is observed prior to structural and functional pathological remodeling in the heart (Perseghin et al., 2007).

Together, these data further the potential uniqueness of diabetic cardiomyopathy in different models and buttress the existence of inherent limitations when comparing between clinical (human) and pre-clinical (rodent) experimental data. One feature that has long been known and is under no debate is that cardiomyocytes experience a loss of metabolic flexibility in the face of chronically high glucose in animal models of diabetes (Chatham and Forder, 1993; Stanley et al., 1997). The consequence of this on the progression of diabetic cardiomyopathy is important, yet our understanding of how this contributes to fibrotic remodeling and diastolic dysfunction remains to be fully understood. In this context, Galgani et al. (2008) reported that metabolic inflexibility to glucose in humans with T2DM is mostly related to defective glucose transport. One theme common to rodent and human studies is the disparity for the inclusion of female subjects (Miller and Best, 2011; Mozaffarian et al., 2015), resulting in a lack of data specific to sex differences and treatment options specific to females. The gender-specific differences in diabetic cardiomyopathy and its progression, at the level of the myocardium remain largely unknown. A purpose of this review is to highlight the known data on the human and animal literature interrogating molecular mechanisms and sex differences contributing to diabetic cardiomyopathy.

# CONTRIBUTION OF SEX HORMONES IN DIABETIC CARDIOMYOPATHY

Testosterone is the primary male sex hormone responsible for maturation of sex organs, sperm production, and secondary





sexual characteristics in males (Kloner et al., 2016). It is well established that testosterone levels peak during the third decade of life (Gray et al., 1991), and then decline at a rate of 1-2% annually thereafter (Feldman et al., 2002). Epidemiological studies have shown that low levels of testosterone associate with increased risk of CVD (Rosano et al., 1999; Araujo et al., 2011; Corona et al., 2011; Ruige et al., 2011). Furthermore, one in three type-2 diabetic men are testosterone deficient, suggesting that low testosterone levels are associated with metabolic syndrome and T2DM (Spark, 2007). In a streptozotocin-induced model of T1DM using Sprague-Dawley rats, treatment with testosterone increased cardiac angiogenesis, highlighting the cardioprotective role of testosterone (Chodari et al., 2016). Given the associations between testosterone, aging, CVD, and metabolic diseases such as T2DM, further explorations into the possible role that testosterone levels play in the progression of diabetic cardiomyopathy are warranted. There is a paucity of data describing the specific effects of low testosterone levels on the development/progression of diabetic cardiomyopathy in males, or the underlying molecular and cellular mechanisms.

Estrogen, along with progesterone, are two female sexualreproductive hormones that are precisely regulated throughout the life course. Estrogens (estrone, estriol, and the biologically active 17β-estradiol) are derived from cholesterol and produced via aromatization of androgens. Females, by virtue of higher estrogen levels, compared to males, have higher protection during pre-menopausal years on various organ systems, including cardiovascular and others (Karas et al., 2001; Mendelsohn, 2002). However, these protective effects are attenuated, and often times lost, when estrogen levels drop off significantly at the onset of menopause. In pre-menopausal women, serum estrogen concentrations range from 367 pmol per liter to 2200 pmol per liter. Upon menopause, these levels decrease significantly to between 18 and 74 pmol per liter, analogous to levels in similarly aged men (Yen and Jade, 1991). Free estrogen in the circulation binds to one of two receptors, estrogen receptor alpha (ER $_{\alpha}$ ) and estrogen receptor beta  $(ER_{\beta})$ , both of which vary in expression depending on tissue type and age (Nilsson and Gustafsson, 2011; Vrtacnik et al., 2014).

In addition to binding to its own receptor and subsequent intracellular signaling, estrogen can affect various physiological functions, not limited to serum lipid concentrations, antioxidant systems, nitric oxide (NO) production, and coagulation pathways (Mendelsohn and Karas, 1999). Estrogen has rapid effects (non-genomic), such as increased NO production and vasodilation, as well as long-term effects (genomic), including decreased atherosclerosis, vascular injury, smooth muscle growth, and increased endothelial cell growth and smooth muscle differentiation (Mendelsohn and Karas, 1999; Wu et al., 2015). Endothelial and vascular smooth muscle cells both express estrogen receptors. Endothelial cells are the first line of defense against any foreign substance that remains in the blood stream and may contribute to vascular disease (Karas et al., 1994; Venkov et al., 1996; Gargett et al., 2002). In vitro, estrogen is capable of increasing NO

release from endothelial cells rapidly, prior to any changes in gene expression (Caulin-Glaser et al., 1997; Lantin-Hermoso et al., 1997). Released NO signals to the vascular smooth muscle cells (VSMCs) to relax, contributing to vasodilation and inhibiting platelet activation (Benjamin et al., 1991; Sandoo et al., 2010). Endothelial nitric oxide synthase (eNOS) and neuronal nitric oxide synthase (nNOS), responsible for synthesizing NO, have also been shown to be regulated by endogenous estrogen levels. This suggests that the primary mediators of vascular tone, NOS and NO, are involved in the dysregulation and/or increased protective effects in both males and females, during the development of diabetic cardiomyopathy (Weiner et al., 1994), but are likely to play a more significant role in females. A New England Journal of Medicine article concludes "Direct myocardial effects of estrogen on cardiac structure and function are likely to be important as well and deserve greater attention" (Mendelsohn and Karas, 1999). In female rats, the absence of estrogen is associated with left ventricular hypertrophy, collagen deposition, and increased sensitivity to angiotensin II (Xu et al., 2003; Dean et al., 2005). Estrogen offers protection to cardiomyocytes via prevention of apoptosis and reduced infarct size (Olivetti et al., 1995; Patten et al., 2004; Kim et al., 2006).

Considering that one hallmark characteristic of diabetic cardiomyopathy is damage to the myocardium in the absence of atherosclerosis or peripheral artery disease, it is important to realize the effects of estrogen on the myocardium and cardiomyocytes. With the onset of menopause, and subsequent decrease in estrogen levels, many women begin hormone replacement therapy (HRT) in order to maintain higher physiological levels of estrogens. While this is not without side effects [see reviews for HRT negative consequences (Yang and Reckelhoff, 2011)], evidence from both prospective and retrospective observational studies show that HRT offers protection from CVD in previously healthy women by 35-50% (Grady et al., 1992; Mendelsohn and Karas, 1994; Grodstein et al., 1996, 1997; Barrett-Connor, 1997; Iorga et al., 2017). What is less clear are the many interactions between low levels of estrogen, CVD, and existing T2DM, and if HRT provides protection from diabetic cardiomyopathy in longstanding diabetics. Attempts to tease out the specific underlying mechanisms, whereby chronically high glucose damages the myocardium and the role estrogen may play in preventing this in females remain in their infancy and require further attention.

#### OXIDATIVE STRESS AND DIABETIC CARDIOMYOPATHY

Cardiac oxidative stress is thought to be one of the primary insults leading to subsequent fibrosis, apoptosis, cellular damage, and hypertrophy (Huynh et al., 2014). One study examined 12-week-old female Sprague–Dawley rats after induction of T1DM using Alloxan and reported that markers of oxidative stress were dysregulated in diabetic animals compared to age-matched control rats. The authors suggest that in their model, Alloxan-induced diabetes caused cardiomyopathic changes, and these changes are likely mediated by oxidative stress (Aksakal et al., 2011). Additional studies have documented the association between oxidative stress and T1DM-induced cardiac damage (Zhang et al., 2017). Similarly, pre-clinical models of T2DM have been used to assess the role of oxidative stress in the progression of diabetic cardiomyopathy, with several transgenic, and diet-induced models suggesting a strong connection between diabetic cardiomyopathy and oxidative stress (Zhang et al., 2010; Calligaris et al., 2013; Fuentes-Antras et al., 2015; Chong et al., 2017). Evidence suggests that oxidative stress exhibits sex differences. For example, superoxide dismutase activity was found to be higher in female rat heart compared to male rat heart indicating that oxidative stress is lower in female rat heart compared to male rat heart (Barp et al., 2002). In humans, in vivo biomarkers of oxidative stress are reported to be higher in young men than in women of the same age (Ide et al., 2002). Furthermore, reactive oxygen species (ROS) production in the vascular cells from males are higher than cells from females (Matarrese et al., 2011). Sex differences in oxidative stress are also seen in healthy term neonates and their mothers (Diaz-Castro et al., 2016). Femalespecific reduced systemic oxidative stress may play a role in improved cardio-protection in healthy females compared to males.

Studies in db/db mice show that while both males and females exhibited age-associated increases in the left ventricular atherosclerosis biomarker, plasminogen activator inhibitor 1 (PAI-1), females had a marked increase as diabetes progressed, compared to males, indicating a possible role for PAI-1 in diabetic cardiomyopathy (Zhao R. et al., 2013; Bowden et al., 2015). Bowden et al. (2015) demonstrated that many of the deleterious diabetic complications were exacerbated in female db/db mice compared to their male counterparts, lending support to the hypothesis that diabetic women are more susceptible to diabetic cardiomyopathy (Peters et al., 2014). Surprisingly, LV fibrosis and SERCA2a expression were higher in diabetic mice, compared to controls, but did not show a sex-specific effect. However, in these same animals, oxidative stress was greater in db/db females compared to males (Bowden et al., 2015).

Cardiac fibrosis, which occurs as a by-product of oxidative stress, is a hallmark feature of CVD. In the diabetic heart, fibrosis has been documented to progress in the left ventricle as a result of increased expression and presence of collagens and other extracellular matrix proteins, which stiffens the ventricular walls (Wu et al., 2000; Asbun and Villarreal, 2006; Huynh et al., 2012). As a result, it is common for compensatory left ventricular hypertrophy to occur due to the heart attempting to compensate for increased stiffness. Clinical studies using cardiovascular magnetic resonance have shown significant sex differences in left ventricular remodeling and myocardial fibrosis among patients with heart disease and women show higher New York Heart Association (NYHA) class and greater left ventricular remodeling index (Chen et al., 2015; Li et al., 2017). The population of patients in this study included similar numbers of male and female patients with diabetes. Another study showed that women have higher levels of fibrosis than men among patients with long-standing persistent atrial fibrillation (Li et al., 2017).

#### HUMAN DATA SUPPORTING SEX DIFFERENCES IN DIABETIC CARDIOMYOPATHY

The rates of T2DM incidence are roughly 6.6 and 5.9% of the population for men and women, respectively (Centers for Disease Control and Prevention, 2017). Interestingly, diabetic cardiomyopathy incidence remains higher in diabetic females compared to diabetic males, and this is further complicated by the pre- and post-menopausal stages of women and the vital role that estrogen may be playing in disease protection and progression. A 2014 meta-analysis that analyzed 64 cohorts, including 858, 506 individuals, and 28,203 cardiac events, concluded that diabetic women have a 40% greater risk of incident coronary heart disease compared with diabetic men (Peters et al., 2014). Note that most of the studies to date focus on coronary heart disease rather than diabetic cardiomyopathy implying that the participants were not free from peripheral disease, including hypertension, coronary artery disease and/or atherosclerosis. This is due to the absence of large cohort studies specifically looking at diabetic cardiomyopathy and sex differences. Additionally, virtually no large-scale human studies have inquired specifically into any mechanistic understanding of diabetic cardiomyopathy and the role that sex and sex hormones play.

A 2015 population-based survey examined 2,042 randomly selected residents in Minnesota, aged 45 years or older (Dandamudi et al., 2014). Participants underwent initial echocardiographic assessment of systolic and diastolic function and were followed for 3 years. It was reported that the incidence of diabetic cardiomyopathy in diabetic patients in this cohort was 16.9% and the prevalence of diastolic dysfunction was 54%. Indeed, the authors demonstrated that diabetic cardiomyopathy is associated with a high cumulative probability of the development of heart failure and death. The authors concluded that diabetic cardiomyopathy is relatively common in the studied community and that morbidity and mortality in patients with diabetic cardiomyopathy is high, reaching 31% in the studied population. Notably, this survey did not look at the differences between males and females, and more specifically, the role that sex hormones play in the incidence and progression of diabetic cardiomyopathy.

The Rancho Bernardo Study is a prospective population-based study of older adult inhabitants of Southern California. Subjects were recruited and monitored yearly with questionnaires and every 4 years by clinical evaluation. The study evaluated both male and female T2DM patients for 14 years and, among other outcome measures, concluded that "diabetes in women overrides their natural advantages and that is not entirely...mediated by many other conventional heart disease risk factors." One aspect of this natural advantage that they discuss is the levels of high-density lipoprotein cholesterol (HDLC) being lower in diabetic women compared with both healthy women and diabetic men (Barrett-Connor et al., 1991). Furthermore, data suggests that HDLC at normal or elevated levels confers more protection in women than men (Crouse, 1989; Gordon et al., 1989).

#### RODENT DATA SUPPORTING SEX DIFFERENCES IN DIABETIC CARDIOMYOPATHY

Our understanding of diabetic cardiomyopathy incidence in humans has largely been a result of the previously mentioned epidemiological and cross-sectional studies, and meta-analysis. However, the current understanding of diabetic cardiomyopathy progression and the mechanisms involved are largely a result of pre-clinical rodent studies. Several experimental rodent models have been developed to study various contributing factors to diabetic cardiomyopathy; including several genetically modified models; db/db mice, ob/ob mice, Otsuka Long-Evans Tokushima (OLETF) rats, CIRKO (cardiomyocyte deletion of insulin receptor) mice, cardiac lipotoxic mice [cardiomyocyte-specific long-chain acyl-CoA synthetase (ACS) and fatty acid transport protein (FATP1) overexpressing], Zucker diabetic fatty (ZDF), Zucker obese (ZO), lean (ZL) rats, and various other diet-induced obesity (DIO) strains. Monogenic, polygenic, and pharmacologically altered rodent models all provide insight into disease progression but are not without challenges when comparing back to human disease. Indeed, human diabetic cardiomyopathy is the result of a sequence of deleterious events that occur in succession, leading to eventual cardiac dysfunction in diabetic patients. The mechanisms elucidated from rodents and currently thought to be responsible for diabetic cardiomyopathy progression include: increased myocardial lipotoxicity, hypertrophy, decreased cardiac function, altered cytokine profile, increased oxidative stress, interstitial fibrosis, contractile and mitochondrial dysfunction, and altered myocardial metabolism.

The db/db and ob/ob genetic mouse models, both of which have altered leptin signaling and become obese, insulin-resistant and mild to severely hyperglycemic, provide a useful model to look at the damaging effects of diabetes on the myocardium. Noteworthy, few studies have included both males and females in an attempt to understand sex differences in these models. The db/db mice myocardium is characterized by increased oxidative stress, [(Boudina et al., 2007) male mice], cardiomyocyte apoptosis (Barouch et al., 2006), diastolic dysfunction [(Semeniuk et al., 2002) – male mice]. Isolated db/db hearts exhibit decreased left ventricular function compared to ob/ob hearts (Mazumder et al., 2004; Hafstad et al., 2006). Indeed, ob/ob mice have a reduction in 'cardiac metabolic flexibility,' meaning that in times of stress, i.e., hypoxia or ischemia, when a healthy heart would switch from fatty acids to glucose for fuel, obese ob/ob hearts are unable to switch substrate utilization (Boudina et al., 2005). Cardiac metabolism in db/db and ob/ob mouse models demonstrate decreased glucose oxidation and increased fatty acid oxidation. Indeed, this attenuation of metabolic flexibility increases the risk of ischemic damage and associates with insulin resistance and reductions in mitochondrial oxidative capacity.

The male ZDF rat has been extensively used as a rodent model of hyperglycemia and T2DM. Young ZDF-male rats exhibit severe hyperglycemia, diastolic dysfunction with preserved ejection fraction, reduced cardiac capillary density and cardiac structural damage caused by fibrosis and mitochondrial disorganization (Baynes and Murray, 2009; Pulakat et al., 2011). Diabetic cardiomyopathy in young hyperglycemic ZDF-female rats shares the features of reduced capillary density and mitochondrial damage with ZDF-male rats (Lum-Naihe et al., 2017). However, ZDF-female rats did not exhibit cardiac fibrosis, but demonstrated increase in heart weight and cardiomyocyte hypertrophy and higher levels of phosphorylation of cardiac mTOR (mechanistic target of rapamycin).

Myocardial contractility and structural parameters of the heart are different between male and female rats. Specifically, cardiomyocytes from male Wistar rats exhibit hypertrophy and undergo a greater degree of postnatal growth compared to female rats (Capasso et al., 1983). Similarly, 5-month-old ZL-male rats have larger hearts compared to ZL-female rats. Presence of T2DM is associated with increased risk of left ventricular hypertrophy, however, prolonged T2DM and severe hyperglycemia causes muscle loss. The 5-month-old ZDF-female rats that exhibited both hyperinsulinemia and hyperglycemia showed a greater degree of hypertrophy, both in terms of cardiomyocyte size and total heart size (adjusted to tibia length), compared to their lean counterparts, whereas agematched ZDF male rats that had prolonged hyperglycemia did not show cardiac hypertrophy (Lum-Naihe et al., 2017). Blenck et al. (2016) have compared the sexual dimorphism in human and rodent cardiovascular physiology in a recent review.

Previous studies have shown that there is a pan suppression of intracardiac cytokines in another T2DM rat model, the ZOmale rat (Luck et al., 2017). The ZO-male rat also has a mutation in the leptin receptor and is characterized by obesity, hyperinsulinemia, mild hyperglycemia, diastolic dysfunction with preserved ejection fraction, cardiac hypertrophy, and fibrosis (Gul et al., 2015; Luck et al., 2017). Compared to ZL-males, both ZO-male and ZDF-male rats exhibited a suppression of GM-CSF, IL-10, and IFN-y (Luck et al., 2017; Lum-Naihe et al., 2017). GM-CSF, IL-10 (Wilson et al., 2007; Shi et al., 2014), and IFN-γ (Oldroyd et al., 1999; Emmez et al., 2008) are anti-fibrotic cytokines and their suppression could have contributed to cardiac fibrosis in these male T2DM models. Conversely, expression of these anti-fibrotic intracardiac cytokines was comparable between ZDF-female rats and ZL-female rats. This sex difference in the expression of anti-fibrotic intracardiac cytokines may be one of the reasons underlying the sex differences in diabetic cardiomyopathy (**Figure 3**). Interestingly, suppression of neuropilin-1, associated with increased cardiomyopathy (Wang et al., 2015), was observed in healthy females compared to male rats and also in diabetic rats of both sexes, compared to their healthy counterparts. This observation suggests that reduction in neuropilin-1 is an important contributor to diabetic cardiomyopathy.

Noteworthy is the sex difference in young ZDF-males and females in regards to DIO and T2DM (Mulder et al., 2010). ZDFmales develop hyperinsulinemia, hyperglycemia, and T2DM on standard chow. ZDF-females require a high fat diet with 48 kcal% Lard (Diet #12468 formulated by Edward A. Ulman, Ph.D. Research Diets, Inc.), in order to induce metabolic disease and T2DM. Thus, ZDF females need an additional insult, a high fat diet to induce T2DM. Importantly, this leads to the speculation that, in this model, females possess some level of "protection" and with it, an ability to delay hyperglycemia, hyperinsulinemia, and T2DM. Interestingly, the effect of DIO on the onset of T2DM differs also in mice and this effect depends on the age at which the high fat diet is started. In juvenile mice, DIO induces T2DM in males, but not in females (Salinero et al., 2018). However, if the DIO is initiated in adult mice, this sex difference in the induction of T2DM disappears.

Unlike the above rodent models that are characterized by systemic metabolic disruption, other models have been created that are cardiac specific and allow the deleterious effects of prolonged diabetes to be produced quickly in cardiac tissue. These models include CIRKO mice and cardiac lipotoxic mice (ACS and FATP1 overexpressing). The CIRKO mouse was first characterized in 2002 and provided a model that separated the intrinsic defects in cardiomyocytes from the potential confounding effects of altered systemic hyperglycemia (Belke et al., 2002). Similar to the CIRKO mouse, models of cardiac myocyte-specific liptoxicity provide an alternative lens to look at how fatty acid accumulation damages the heart muscle independent of peripheral and systemic disturbances. There have been two models developed that rely on excess fat entry into cardiomyocytes, mimicking severe diabetic cardiomyopathies. These models include cardiomyocyte-specific ACS and FATP1 overexpressing mice (Chiu et al., 2001, 2005). The resultant pathology exhibited large increases in fatty acids within the myocardium, and therefore, many of the deleterious effects on cardiac function are seen as in diabetic cardiomyopathy. These perturbations in lipid homeostasis within cardiomyocytes offers an experimental model to recapitulate the negative cellular effects commonly observed in clinical diabetic cardiomyopathy. However, these studies did not explore how biological sex differences modulate cardiac pathology of these models.

#### EMERGING MOLECULAR MARKERS OF SEX DIFFERENCES IN DIABETIC CARDIOMYOPATHY

Given the uniqueness of the molecular signature of diabetic cardiomyopathy, development and understanding of the targets and specific sex differences remains a vitally important area of



diabetic cardiomyopathy. In male, mTORC1 activation and hypertrophy are observed in some models but not others (indicated by dashed outer cardiomyocyte outline), while AT2R expression is not changed. While fatty acid (FA) uptake, fibrosis, collagen 1a, and miR-29a/b/c/ are increased with disease progression; the following are all reduced with disease progression; FA utilization, capillary density, mitochondrial function, and *Med13*-signaling, and several cytokines including IL-2, IL-10, IFN-y, and GM-CSF. **(B)** Diagram of a female cardiomyocyte as it progresses from healthy to diabetic cardiomyopathy. In the female, mTORC1 activation and hypertrophy are observed. Additionally, miR-208a and miR-29a/b/c are increased, while FA utilization, mitochondrial function, fibrosis, *Agtr2-Med13* signaling, capillary density, collagen 1a, and NRP-1 are decreased (Widdop et al., 2003; Miki et al., 2013; Schilling and Mann, 2014; Lum-Naihe et al., 2017).

inquiry. One of the molecules that regulates cardiac conduction and hypertrophy is the microRNA (miRNA) miR-208a (Callis et al., 2009). Interestingly, miR-208a exhibits sex differences, with expression being higher in the heart tissues of ZL-female rats compared to ZL-male rats (Lum-Naihe et al., 2017). T2DM increased miR-208a expression in both ZDF-male and ZDFfemale hearts, resulting in ZDF-females having the highest expression. Thus miR-208a is a miRNA that links T2DM and cardiac dysfunction and hypertrophy. Notably, ex vivo studies on mouse aorta showed that miR-208a was suppressed by estrogen (Zhao J. et al., 2013). However, in healthy female rats with intact ovary, cardiac miR-208a expression was higher than that seen in males, indicating that estrogen-mediated regulation of miR-208a in vivo is different. A recent review by Florijn et al. (2017) discusses sex bias in miRNA expression and its correlation with the higher prevalence of heart failure with preserved ejection fraction (HFpEF) in women. HFpEF is a hall mark of diabetic cardiomyopathy. Pre-clinical studies in healthy mice and in vitro studies using estrogen have identified miRNAs either regulated by estrogen (miR-203, miR-126, miR-23a, miR-21, miR-24, miR-27a and b, and miR-106a and b) or encoded by X-chromosome (miR-98, miR-652, miR-221, miR-222, miR-223, miR-361, miR-421, miR-325, miR-188, miR-92a, miR-424, miR-503, miR-505). Authors suggest that estrogen-mediated regulation and/or link to X-chromosome play a role in sex bias of these miRNAs that contribute to CVD (Florijn et al., 2017).

Cardiac Mediator Complex 13 (MED13), encoded by the Med13 gene, plays a critical role in systemic energy homeostasis and confers resistance to weight gain (Grueter et al., 2012). MED13 also improves systemic insulin sensitivity and glucose tolerance and is increased by treatment with mTOR complex 1 inhibitors rapamycin and nebivolol (Gul et al., 2015). Heart tissues of ZL-females exhibit higher levels of Med13 mRNA compared to ZL-male rats, indicating that MED13 is a molecule that may provide increased cardioprotection in healthy female rat hearts. Med13 mRNA is an established target of miR-208a (Grueter et al., 2012; Gul et al., 2015). T2DM suppressed cardiac Med13 mRNA levels in both sexes, which is consistent with the T2DM-induced increase in miR-208a. Notably, both miR-208a and Med13 expression levels were higher in ZL-female rats, indicating that in healthy female rats, other mechanisms contributed to increased expression of Med13; however, this advantage was lost in diabetic ZDF-female rats.

Angiotensin II type 2 receptor (AT2R), encoded by the X-linked *Agtr2* gene, has been documented to confer cardiovascular protective and reparative effects. Activating AT2R signaling by agonists and increasing *Agtr2* gene copy number by genetic manipulation in murine models improves cardiac repair and enhances cardiac function (Altarche-Xifro et al., 2009; Qi et al., 2012; Skorska et al., 2015). AT2R exhibits sex differences in cardiovascular expression (Okumura et al., 2005; Sampson et al., 2008; Hilliard et al., 2012; Lum-Naihe et al., 2017). AT2R expression is higher in the vasculature of healthy murine female models. This female-specific increased AT2R expression contributes to their increased resistance to hypertension and protection from cardiovascular and renal injury compared to males.

### CONCLUSION AND FUTURE DIRECTIONS

The NIH (Clayton and Collins, 2014) has long recognized sex disparities in human clinical trials as a problem, and in 1993, passed the NIH Revitalization Act. In the 2 decades since, over half of NIH-funded clinical research participants are now women. The area that is still experiencing sex bias are pre-clinical studies, including in vitro and in vivo studies. In fact, a 2010 Nature opinion article shows that males still dominate animal studies (Zucker and Beery, 2010). Based on current trends, diabetes incidence will continue to increase and with it the co-morbidities, such as obesity, hypertension, and diabetic cardiomyopathy. Given the unique sex differences observed between males and females, future work needs to be aimed at understanding the role of sexspecificity on the progression of diabetic cardiomyopathy. There is desperately the need for a push for possible unique therapeutic treatment options that will address sex differences. Pre-clinical models can continue to help identify, test, and optimize possible therapies, with the hopes to make the leap into human trials. Paramount to this is the need for a continued scientific and medical discussion and elucidation of an agreed upon clinical definition of diabetic cardiomyopathy. Although the role of estrogen-mediated signaling in better cardiovascular protection of healthy females compared to males is well established, this female advantage is lost in T2DM and diabetic cardiomyopathy. Therefore, identifying additional therapeutic targets to improve cardiovascular outcomes of diabetic females is a critical need. AT2R and MED13 are cardioprotective molecules that exhibit female-specific increased expression in rodent studies. Moreover, currently there are no AT2R agonists in clinic to treat heart disease. Developing drugs that can increase expression of MED13 and AT2R in diabetic heart can lead to better treatment paradigms for diabetic cardiomyopathy and to mitigate sex differences in this pathology.

## **AUTHOR CONTRIBUTIONS**

RT wrote the manuscript. RT, AB, and LP discussed the literature and figures, contributed to the intellectual input, and edited the manuscript. RT, AB, and LP approved the final version.

## FUNDING

This work was supported, in part, by the Life Science Mission Enhancement Fund from UM-Columbia (LP) and NIH NHLBI 1R01HL118376 (LP) and R01HL138988-01A1.

## ACKNOWLEDGMENTS

We greatly acknowledge the facilities at Dalton Cardiovascular Research Center.

#### REFERENCES

- Abdul-Ghani, M. A., Jayyousi, A., DeFronzo, R. A., Asaad, N., and Al-Suwaidi, J. (2017). Insulin resistance the link between T2DM and CVD: basic mechanisms and clinical implications. *Curr. Vasc. Pharmacol.* doi: 10.2174/ 1570161115666171010115119 [Epub ahead of print].
- Aksakal, E., Akaras, N., Kurt, M., Tanboga, I. H., Halici, Z., Odabasoglu, F., et al. (2011). The role of oxidative stress in diabetic cardiomyopathy: an experimental study. *Eur. Rev. Med. Pharmacol. Sci.* 15, 1241–1246.
- Altarche-Xifro, W., Curato, C., Kaschina, E., Grzesiak, A., Slavic, S., Dong, J., et al. (2009). Cardiac c-kit+AT2+ cell population is increased in response to ischemic injury and supports cardiomyocyte performance. *Stem Cells* 27, 2488–2497. doi: 10.1002/stem.171
- Anderson, E. J., Kypson, A. P., Rodriguez, E., Anderson, C. A., Lehr, E. J., and Neufer, P. D. (2009). Substrate-specific derangements in mitochondrial metabolism and redox balance in the atrium of the type 2 diabetic human heart. *J. Am. Coll. Cardiol.* 54, 1891–1898. doi: 10.1016/j.jacc.2009.07.031
- Araujo, A. B., Dixon, J. M., Suarez, E. A., Murad, M. H., Guey, L. T., and Wittert, G. A. (2011). Clinical review: endogenous testosterone and mortality in men: a systematic review and meta-analysis. *J. Clin. Endocrinol. Metab.* 96, 3007–3019. doi: 10.1210/jc.2011-1137
- Archundia Herrera, M. C., Subhan, F. B., and Chan, C. B. (2017). Dietary patterns and cardiovascular disease risk in people with type 2 diabetes. *Curr. Obes. Rep.* 6, 405–413. doi: 10.1007/s13679-017-0284-5
- Asbun, J., and Villarreal, F. J. (2006). The pathogenesis of myocardial fibrosis in the setting of diabetic cardiomyopathy. J. Am. Coll. Cardiol. 47, 693–700. doi: 10.1016/j.jacc.2005.09.050
- Bar, C., Bernardes de Jesus, B., Serrano, R., Tejera, A., Ayuso, E., Jimenez, V., et al. (2014). Telomerase expression confers cardioprotection in the adult mouse heart after acute myocardial infarction. *Nat. Commun.* 5:5863. doi: 10.1038/ ncomms6863
- Barouch, L. A., Gao, D., Chen, L., Miller, K. L., Xu, W., Phan, A. C., et al. (2006). Cardiac myocyte apoptosis is associated with increased DNA damage and decreased survival in murine models of obesity. *Circ. Res.* 98, 119–124. doi: 10.1161/01.RES.0000199348.10580.1d
- Barp, J., Araujo, A. S., Fernandes, T. R., Rigatto, K. V., Llesuy, S., Bello-Klein, A., et al. (2002). Myocardial antioxidant and oxidative stress changes due to sex hormones. *Braz. J. Med. Biol. Res.* 35, 1075–1081. doi: 10.1590/S0100-879X2002000900008
- Barrett-Connor, E. (1997). Sex differences in coronary heart disease. Why are women so superior? The 1995 Ancel Keys Lecture. *Circulation* 95, 252–264. doi: 10.1161/01.CIR.95.1.252
- Barrett-Connor, E. L., Cohn, B. A., Wingard, D. L., and Edelstein, S. L. (1991). Why is diabetes mellitus a stronger risk factor for fatal ischemic heart disease in women than in men? The Rancho Bernardo Study. JAMA 265, 627–631. doi: 10.1001/jama.1991.03460050081025
- Basu, R., Dalla Man, C., Campioni, M., Basu, A., Klee, G., Toffolo, G., et al. (2006). Effects of age and sex on postprandial glucose metabolism: differences in glucose turnover, insulin secretion, insulin action, and hepatic insulin extraction. *Diabetes Metab. Res. Rev.* 55, 2001–2014. doi: 10.2337/db05-1692
- Baynes, J., and Murray, D. B. (2009). Cardiac and renal function are progressively impaired with aging in Zucker diabetic fatty type II diabetic rats. Oxid. Med. Cell. Longev. 2, 328–334. doi: 10.4161/oxim.2.5.9831
- Belke, D. D., Betuing, S., Tuttle, M. J., Graveleau, C., Young, M. E., Pham, M., et al. (2002). Insulin signaling coordinately regulates cardiac size, metabolism, and contractile protein isoform expression. *J. Clin. Invest.* 109, 629–639. doi: 10.1172/JCI13946
- Benjamin, N., Dutton, J. A., and Ritter, J. M. (1991). Human vascular smooth muscle cells inhibit platelet aggregation when incubated with glyceryl trinitrate: evidence for generation of nitric oxide. Br. J. Pharmacol. 102, 847–850. doi: 10.1111/j.1476-5381.1991.tb12264.x
- Berkova, M., Opavsky, J., Berka, Z., Skranka, V., and Salinger, J. (2003). Left ventricular diastolic filling in young persons with type 1 diabetes mellitus. *Biomed. Pap. Med. Fac. Univ. Palacky.Olomouc Czech. Repub.* 147, 57–61. doi: 10.5507/bp.2003.008
- Bertoluci, M. C., and Rocha, V. Z. (2017). Cardiovascular risk assessment in patients with diabetes. *Diabetol. Metab. Syndr.* 9:25. doi: 10.1186/s13098-017-0225-1

- Blenck, C. L., Harvey, P. A., Reckelhoff, J. F., and Leinwand, L. A. (2016). The importance of biological sex and estrogen in rodent models of cardiovascular health and disease. *Circ. Res.* 118, 1294–1312. doi: 10.1161/CIRCRESAHA.116. 307509
- Borisov, A. B., Ushakov, A. V., Zagorulko, A. K., Novikov, N. Y., Selivanova, K. F., Edwards, C. A., et al. (2008). Intracardiac lipid accumulation, lipoatrophy of muscle cells and expansion of myocardial infarction in type 2 diabetic patients. *Micron* 39, 944–951. doi: 10.1016/j.micron.2007.11.002
- Boudina, S., Sena, S., O'Neill, B. T., Tathireddy, P., Young, M. E., and Abel, E. D. (2005). Reduced mitochondrial oxidative capacity and increased mitochondrial uncoupling impair myocardial energetics in obesity. *Circulation* 112, 2686–2695. doi: 10.1161/CIRCULATIONAHA.105.554360
- Boudina, S., Sena, S., Theobald, H., Sheng, X., Wright, J. J., Hu, X. X., et al. (2007). Mitochondrial energetics in the heart in obesity-related diabetes: direct evidence for increased uncoupled respiration and activation of uncoupling proteins. *Diabetes Metab. Res. Rev.* 56, 2457–2466. doi: 10.2337/db07-0481
- Bowden, M. A., Tesch, G. H., Julius, T. L., Rosli, S., Love, J. E., and Ritchie, R. H. (2015). Earlier onset of diabesity-Induced adverse cardiac remodeling in female compared to male mice. *Obesity* 23, 1166–1177. doi: 10.1002/oby. 21072
- Bugger, H., and Abel, E. D. (2009). Rodent models of diabetic cardiomyopathy. Dis. Model Mech. 2, 454–466. doi: 10.1242/dmm.001941
- Calligaris, S. D., Lecanda, M., Solis, F., Ezquer, M., Gutierrez, J., Brandan, E., et al. (2013). Mice long-term high-fat diet feeding recapitulates human cardiovascular alterations: an animal model to study the early phases of diabetic cardiomyopathy. *PLoS One* 8:e60931. doi: 10.1371/journal.pone. 0060931
- Callis, T. E., Pandya, K., Seok, H. Y., Tang, R. H., Tatsuguchi, M., Huang, Z. P., et al. (2009). MicroRNA-208a is a regulator of cardiac hypertrophy and conduction in mice. J. Clin. Invest. 119, 2772–2786. doi: 10.1172/JCI36154
- Capasso, J. M., Remily, R. M., Smith, R. H., and Sonnenblick, E. H. (1983). Sex differences in myocardial contractility in the rat. *Basic Res. Cardiol.* 78, 156–171. doi: 10.1007/BF01906669
- Caulin-Glaser, T., Garcia-Cardena, G., Sarrel, P., Sessa, W. C., and Bender, J. R. (1997). 17 beta-estradiol regulation of human endothelial cell basal nitric oxide release, independent of cytosolic Ca2+ mobilization. *Circ. Res.* 81, 885–892. doi: 10.1161/01.RES.81.5.885
- Centers for Disease Control and Prevention (2017). National Diabetes Statistics Report 2017. Atlanta, GA: Centers for Disease Control and Prevention.
- Chatham, J. C., and Forder, J. R. (1993). A 13C-NMR study of glucose oxidation in the intact functioning rat heart following diabetes-induced cardiomyopathy. *J. Mol. Cell Cardiol.* 25, 1203–1213. doi: 10.1006/jmcc.1993.1133
- Chen, Y. Z., Qiao, S. B., Hu, F. H., Yuan, J. S., Yang, W. X., Cui, J. G., et al. (2015). Left ventricular remodeling and fibrosis: sex differences and relationship with diastolic function in hypertrophic cardiomyopathy. *Eur. J. Radiol.* 84, 1487–1492. doi: 10.1016/j.ejrad.2015.04.026
- Cheng, Y. Y., Kao, T. W., Chang, Y. W., Wu, C. J., Peng, T. C., Wu, L. W., et al. (2017). Examining the gender difference in the association between metabolic syndrome and the mean leukocyte telomere length. *PLoS One* 12:e0180687. doi: 10.1371/journal.pone.0180687
- Chiu, H. C., Kovacs, A., Blanton, R. M., Han, X., Courtois, M., Weinheimer, C. J., et al. (2005). Transgenic expression of fatty acid transport protein 1 in the heart causes lipotoxic cardiomyopathy. *Circ. Res.* 96, 225–233. doi: 10.1161/01.RES. 0000154079.20681.B9
- Chiu, H. C., Kovacs, A., Ford, D. A., Hsu, F. F., Garcia, R., Herrero, P., et al. (2001). A novel mouse model of lipotoxic cardiomyopathy. *J. Clin. Invest.* 107, 813–822. doi: 10.1172/JCI10947
- Chodari, L., Mohammadi, M., Ghorbanzadeh, V., Dariushnejad, H., and Mohaddes, G. (2016). Testosterone and voluntary exercise promote angiogenesis in hearts of rats with diabetes by enhancing expression of VEGF-A and SDF-1a. *Can. J. Diabetes* 40, 436–441. doi: 10.1016/j.jcjd.2016. 03.004
- Chong, C. R., Clarke, K., and Levelt, E. (2017). Metabolic remodeling in diabetic cardiomyopathy. *Cardiovasc. Res.* doi: 10.1093/cvr/cvx018 [Epub ahead of print].
- Clayton, J. A., and Collins, F. S. (2014). Policy: NIH to balance sex in cell and animal studies. *Nature* 509, 282–283. doi: 10.1038/509282a

- Corona, G., Rastrelli, G., Monami, M., Guay, A., Buvat, J., Sforza, A., et al. (2011). Hypogonadism as a risk factor for cardiovascular mortality in men: a meta-analytic study. *Eur. J. Endocrinol.* 165, 687–701. doi: 10.1530/EJE-11-0447
- Crouse, J. R. III (1989). Gender, lipoproteins, diet, and cardiovascular risk. Sauce for the goose may not be sauce for the gander. *Lancet* 1, 318–320. doi: 10.1016/ S0140-6736(89)91320-2
- Dandamudi, S., Slusser, J., Mahoney, D. W., Redfield, M. M., Rodeheffer, R. J., and Chen, H. H. (2014). The prevalence of diabetic cardiomyopathy: a populationbased study in Olmsted County, Minnesota. J. Card. Fail. 20, 304–309. doi: 10.1016/j.cardfail.2014.02.007
- Davis, I. C., Ahmadizadeh, I., Randell, J., Younk, L., and Davis, S. N. (2017). Understanding the impact of hypoglycemia on the cardiovascular system. *Expert Rev. Endocrinol. Metab.* 12, 21–33. doi: 10.1080/17446651.2017.12 75960
- Dean, S. A., Tan, J., O'Brien, E. R., and Leenen, F. H. (2005). 17beta-estradiol downregulates tissue angiotensin-converting enzyme and ANG II type 1 receptor in female rats. Am. J. Physiol. Regul. Integr. Comp. Physiol. 288, R759–R766. doi: 10.1152/ajpregu.00595.2004
- Diaz-Castro, J., Pulido-Moran, M., Moreno-Fernandez, J., Kajarabille, N., de Paco, C., Garrido-Sanchez, M., et al. (2016). Gender specific differences in oxidative stress and inflammatory signaling in healthy term neonates and their mothers. *Pediatr. Res.* 80, 595–601. doi: 10.1038/pr.2016.112
- Didangelos, T. P., Arsos, G. A., Karamitsos, D. T., Athyros, V. G., and Karatzas, N. D. (2003). Left ventricular systolic and diastolic function in normotensive type 1 diabetic patients with or without autonomic neuropathy: a radionuclide ventriculography study. *Diabetes Care* 26, 1955–1960. doi: 10.2337/diacare.26. 7.1955
- Emmez, H., Kardes, O., Dogulu, F., Kurt, G., Memis, L., and Baykaner, M. K. (2008). Role of antifibrotic cytokine interferon-gamma in the prevention of postlaminectomy peridural fibrosis in rats. *Neurosurgery* 62, 1351–1357; discussion 1357–1358. doi: 10.1227/01.neu.0000333307.02802.04
- Feldman, H. A., Longcope, C., Derby, C. A., Johannes, C. B., Araujo, A. B., Coviello, A. D., et al. (2002). Age trends in the level of serum testosterone and other hormones in middle-aged men: longitudinal results from the Massachusetts male aging study. J. Clin. Endocrinol. Metab. 87, 589–598. doi: 10.1210/jcem. 87.2.8201
- Flanagan, D. E., Holt, R. I., Owens, P. C., Cockington, R. J., Moore, V. M., Robinson, J. S., et al. (2006). Gender differences in the insulin-like growth factor axis response to a glucose load. *Acta Physiol.* 187, 371–378. doi: 10.1111/j.1748-1716.2006.01581.x
- Florijn, B. W., Bijkerk, R., van der Veer, E., and van Zonneveld, A. J. (2017). Gender and cardiovascular disease: are sex-biased miRNA networks a driving force behind heart failure with preserved ejection fraction in women? *Cardiovasc. Res.* 114, 210–225. doi: 10.1093/cvr/cvx223
- Folsom, A. R., Szklo, M., Stevens, J., Liao, F., Smith, R., and Eckfeldt, J. H. (1997). A prospective study of coronary heart disease in relation to fasting insulin, glucose, and diabetes. The Atherosclerosis Risk in Communities (ARIC) Study. *Diabetes Care* 20, 935–942. doi: 10.2337/diacare.20.6.935
- Fontes-Carvalho, R., Ladeiras-Lopes, R., Bettencourt, P., Leite-Moreira, A., and Azevedo, A. (2015). Diastolic dysfunction in the diabetic continuum: association with insulin resistance, metabolic syndrome and type 2 diabetes. *Cardiovasc. Diabetol.* 14:4. doi: 10.1186/s12933-014-0168-x
- Fuentes-Antras, J., Picatoste, B., Gomez-Hernandez, A., Egido, J., Tunon, J., and Lorenzo, O. (2015). Updating experimental models of diabetic cardiomyopathy. *J. Diabetes Res.* 2015, 656795. doi: 10.1155/2015/656795
- Galgani, J. E., Heilbronn, L. K., Azuma, K., Kelley, D. E., Albu, J. B., Pi-Sunyer, X., et al. (2008). Metabolic flexibility in response to glucose is not impaired in people with type 2 diabetes after controlling for glucose disposal rate. *Diabetes Metab. Res. Rev.* 57, 841–845. doi: 10.2337/db08-0043
- Gargett, C. E., Bucak, K., Zaitseva, M., Chu, S., Taylor, N., Fuller, P. J., et al. (2002). Estrogen receptor-alpha and -beta expression in microvascular endothelial cells and smooth muscle cells of myometrium and leiomyoma. *Mol. Hum. Reprod.* 8, 770–775. doi: 10.1093/molehr/8.8.770
- Gomez-Marcos, M. A., Recio-Rodriguez, J. I., Gomez-Sanchez, L., Agudo-Conde, C., Rodriguez-Sanchez, E., Maderuelo-Fernandez, J., et al. (2015). Gender differences in the progression of target organ damage in patients with increased insulin resistance: the LOD-DIABETES study. *Cardiovasc. Diabetol.* 14:132. doi: 10.1186/s12933-015-0293-1

- Gordon, D. J., Probstfield, J. L., Garrison, R. J., Neaton, J. D., Castelli, W. P., Knoke, J. D., et al. (1989). High-density lipoprotein cholesterol and cardiovascular disease. Four prospective American studies. *Circulation* 79, 8–15. doi: 10.1161/ 01.CIR.79.1.8
- Grady, D., Rubin, S. M., Petitti, D. B., Fox, C. S., Black, D., Ettinger, B., et al. (1992). Hormone therapy to prevent disease and prolong life in postmenopausal women. *Ann. Intern. Med.* 117, 1016–1037. doi: 10.7326/0003-4819-117-12-1016
- Gray, A., Feldman, H. A., McKinlay, J. B., and Longcope, C. (1991). Age, disease, and changing sex hormone levels in middle-aged men: results of the Massachusetts Male Aging Study. J. Clin. Endocrinol. Metab. 73, 1016–1025. doi: 10.1210/jcem-73-5-1016
- Grodstein, F., Stampfer, M. J., Colditz, G. A., Willett, W. C., Manson, J. E., Joffe, M., et al. (1997). Postmenopausal hormone therapy and mortality. *N. Engl. J. Med.* 336, 1769–1775. doi: 10.1056/NEJM199706193362501
- Grodstein, F., Stampfer, M. J., Manson, J. E., Colditz, G. A., Willett, W. C., Rosner, B., et al. (1996). Postmenopausal estrogen and progestin use and the risk of cardiovascular disease. *N. Engl. J. Med.* 335, 453–461. doi: 10.1056/ NEJM199608153350701
- Grueter, C. E., van Rooij, E., Johnson, B. A., DeLeon, S. M., Sutherland, L. B., Qi, X., et al. (2012). A cardiac microRNA governs systemic energy homeostasis by regulation of MED13. *Cell* 149, 671–683. doi: 10.1016/j.cell.2012. 03.029
- Gul, R., Mahmood, A., Luck, C., Lum-Naihe, K., Alfadda, A. A., Speth, R. C., et al. (2015). Regulation of cardiac miR-208a, an inducer of obesity, by rapamycin and nebivolol. *Obesity* 23, 2251–2259. doi: 10.1002/oby.21227
- Gusso, S., Pinto, T. E., Baldi, J. C., Robinson, E., Cutfield, W. S., and Hofman, P. L. (2012). Diastolic function is reduced in adolescents with type 1 diabetes in response to exercise. *Diabetes Care* 35, 2089–2094. doi: 10.2337/dc11-2331
- Hafstad, A. D., Solevag, G. H., Severson, D. L., Larsen, T. S., and Aasum, E. (2006). Perfused hearts from Type 2 diabetic (db/db) mice show metabolic responsiveness to insulin. Am. J. Physiol. Heart Circ. Physiol. 290, H1763–H1769. doi: 10.1152/ajpheart.01063.2005
- Heyden, S., Heiss, G., Bartel, A. G., and Hames, C. G. (1980). Sex differences in coronary mortality among diabetics in Evans County, Georgia. J. Chronic Dis. 33, 265–273. doi: 10.1016/0021-9681(80)90021-1
- Hilliard, L. M., Jones, E. S., Steckelings, U. M., Unger, T., Widdop, R. E., and Denton, K. M. (2012). Sex-specific influence of angiotensin type 2 receptor stimulation on renal function: a novel therapeutic target for hypertension. *Hypertension* 59, 409–414. doi: 10.1161/HYPERTENSIONAHA.111. 184986
- Holscher, M. E., Bode, C., and Bugger, H. (2016). Diabetic cardiomyopathy: does the type of diabetes matter? *Int. J. Mol. Sci.* 17:2136. doi: 10.3390/ijms17122136
- Huynh, K., Bernardo, B. C., McMullen, J. R., and Ritchie, R. H. (2014). Diabetic cardiomyopathy: mechanisms and new treatment strategies targeting antioxidant signaling pathways. *Pharmacol. Ther.* 142, 375–415. doi: 10.1016/j. pharmthera.2014.01.003
- Huynh, K., Kiriazis, H., Du, X. J., Love, J. E., Jandeleit-Dahm, K. A., Forbes, J. M., et al. (2012). Coenzyme Q10 attenuates diastolic dysfunction, cardiomyocyte hypertrophy and cardiac fibrosis in the db/db mouse model of type 2 diabetes. *Diabetologia* 55, 1544–1553. doi: 10.1007/s00125-012-2495-3
- Ide, T., Tsutsui, H., Ohashi, N., Hayashidani, S., Suematsu, N., Tsuchihashi, M., et al. (2002). Greater oxidative stress in healthy young men compared with premenopausal women. *Arterioscler. Thromb. Vasc. Biol.* 22, 438–442. doi: 10. 1161/hq0302.104515
- Iorga, A., Cunningham, C. M., Moazeni, S., Ruffenach, G., Umar, S., and Eghbali, M. (2017). The protective role of estrogen and estrogen receptors in cardiovascular disease and the controversial use of estrogen therapy. *Biol. Sex Differ.* 8:33. doi: 10.1186/s13293-017-0152-8
- Jensen, M. T., Sogaard, P., Andersen, H. U., Bech, J., Hansen, T. F., Galatius, S., et al. (2014). Prevalence of systolic and diastolic dysfunction in patients with type 1 diabetes without known heart disease: the Thousand & 1 Study. *Diabetologia* 57, 672–680. doi: 10.1007/s00125-014-3164-5
- Kain, V., and Halade, G. V. (2017). Metabolic and biochemical stressors in diabetic cardiomyopathy. Front. Cardiovasc. Med. 4:31. doi: 10.3389/fcvm.2017.00031
- Kajstura, J., Gurusamy, N., Ogorek, B., Goichberg, P., Clavo-Rondon, C., Hosoda, T., et al. (2010). Myocyte turnover in the aging human heart. *Circ. Res.* 107, 1374–1386. doi: 10.1161/CIRCRESAHA.110.231498

- Kannel, W. B., Hjortland, M., and Castelli, W. P. (1974). Role of diabetes in congestive heart failure: the Framingham study. Am. J. Cardiol. 34, 29–34. doi: 10.1016/0002-9149(74)90089-7
- Kannel, W. B., and McGee, D. L. (1979). Diabetes and cardiovascular disease. The Framingham study. JAMA 241, 2035–2038. doi: 10.1001/jama.1979. 03290450033020
- Karas, R. H., Patterson, B. L., and Mendelsohn, M. E. (1994). Human vascular smooth muscle cells contain functional estrogen receptor. *Circulation* 89, 1943–1950. doi: 10.1161/01.CIR.89.5.1943
- Karas, R. H., Schulten, H., Pare, G., Aronovitz, M. J., Ohlsson, C., Gustafsson, J. A., et al. (2001). Effects of estrogen on the vascular injury response in estrogen receptor alpha, beta (double) knockout mice. *Circ. Res.* 89, 534–539. doi: 10.1161/hh1801.097239
- Kautzky-Willer, A., Harreiter, J., and Pacini, G. (2016). Sex and gender differences in risk, pathophysiology and complications of Type 2 diabetes mellitus. *Endocr. Rev.* 37, 278–316. doi: 10.1210/er.2015-1137
- Kim, J. K., Pedram, A., Razandi, M., and Levin, E. R. (2006). Estrogen prevents cardiomyocyte apoptosis through inhibition of reactive oxygen species and differential regulation of p38 kinase isoforms. J. Biol. Chem. 281, 6760–6767. doi: 10.1074/jbc.M511024200
- Kloner, R. A., Carson, C. III, Dobs, A., Kopecky, S., and Mohler, E. R. III (2016). Testosterone and cardiovascular disease. J. Am. Coll. Cardiol. 67, 545–557. doi: 10.1016/j.jacc.2015.12.005
- Lantin-Hermoso, R. L., Rosenfeld, C. R., Yuhanna, I. S., German, Z., Chen, Z., and Shaul, P. W. (1997). Estrogen acutely stimulates nitric oxide synthase activity in fetal pulmonary artery endothelium. *Am. J. Physiol.* 273(1 Pt 1), L119–L126. doi: 10.1152/ajplung.1997.273.1.L119
- Laverty, A. A., Bottle, A., Kim, S. H., Visani, B., Majeed, A., Millett, C., et al. (2017). Gender differences in hospital admissions for major cardiovascular events and procedures in people with and without diabetes in England: a nationwide study 2004-2014. *Cardiovasc. Diabetol.* 16, 1–13. doi: 10.1186/s12933-017-0580-0
- Leri, A., Malhotra, A., Liew, C. C., Kajstura, J., and Anversa, P. (2000). Telomerase activity in rat cardiac myocytes is age and gender dependent. J. Mol. Cell. Cardiol. 32, 385–390. doi: 10.1006/jmcc.1999.1084
- Li, Z., Wang, Z., Yin, Z., Zhang, Y., Xue, X., Han, J., et al. (2017). Gender differences in fibrosis remodeling in patients with long-standing persistent atrial fibrillation. *Oncotarget* 8, 53714–53729. doi: 10.18632/oncotarget. 16342
- Liebow, I. M., and Hellerstein, H. K. (1949). Cardiac complications of diabetes mellitus. Am. J. Med. 7, 660–670. doi: 10.1016/0002-9343(49)90388-5
- Logue, J., Walker, J. J., Colhoun, H. M., Leese, G. P., Lindsay, R. S., McKnight, J. A., et al. (2011). Do men develop type 2 diabetes at lower body mass indices than women? *Diabetologia* 54, 3003–3006. doi: 10.1007/s00125-011-2313-3
- Lorenzo-Almoros, A., Tunon, J., Orejas, M., Cortes, M., Egido, J., and Lorenzo, O. (2017). Diagnostic approaches for diabetic cardiomyopathy. *Cardiovasc. Diabetol.* 16:28. doi: 10.1186/s12933-017-0506-x
- Lotufo, P. A., Gaziano, J. M., Chae, C. U., Ajani, U. A., Moreno-John, G., Buring, J. E., et al. (2001). Diabetes and all-cause and coronary heart disease mortality among US male physicians. *Arch. Intern. Med.* 161, 242–247. doi: 10.1001/ archinte.161.2.242
- Luck, C., DeMarco, V. G., Mahmood, A., Gavini, M. P., and Pulakat, L. (2017). Differential regulation of cardiac function and intracardiac cytokines by rapamycin in healthy and diabetic rats. Oxid. Med. Cell Longev. 2017:5724046. doi: 10.1155/2017/5724046
- Lum-Naihe, K. T., Toedebusch, R., Mahmood, A., Bajwa, J., Carmack, T., Kumar, S., et al. (2017). Cardiovascular disease progression in female Zucker Diabetic Fatty rats occurs via unique mechanisms compared to males. *Sci. Rep.* 7:17823. doi: 10.1038/s41598-017-18003-8
- Martin-Timon, I., Sevillano-Collantes, C., Segura-Galindo, A., and Del Canizo-Gomez, F. J. (2014). Type 2 diabetes and cardiovascular disease: have all risk factors the same strength? *World J. Diabetes* 5, 444–470. doi: 10.4239/wjd.v5. i4.444
- Matarrese, P., Colasanti, T., Ascione, B., Margutti, P., Franconi, F., Alessandri, C., et al. (2011). Gender disparity in susceptibility to oxidative stress and apoptosis induced by autoantibodies specific to RLIP76 in vascular cells. *Antioxid. Redox Signal.* 15, 2825–2836. doi: 10.1089/ars.2011.3942

- Mazumder, P. K., O'Neill, B. T., Roberts, M. W., Buchanan, J., Yun, U. J., Cooksey, R. C., et al. (2004). Impaired cardiac efficiency and increased fatty acid oxidation in insulin-resistant ob/ob mouse hearts. *Diabetes Metab. Res. Rev.* 53, 2366–2374. doi: 10.2337/diabetes.53.9.2366
- Mendelsohn, M. E. (2002). Protective effects of estrogen on the cardiovascular system. Am. J. Cardiol. 89, 12E–17E; discussion 17E–18E. doi: 10.1016/S0002-9149(02)02405-0
- Mendelsohn, M. E., and Karas, R. H. (1994). Estrogen and the blood vessel wall. *Curr. Opin. Cardiol.* 9, 619–626. doi: 10.1097/00001573-199409000-00018
- Mendelsohn, M. E., and Karas, R. H. (1999). The protective effects of estrogen on the cardiovascular system. N. Engl. J. Med. 340, 1801–1811. doi: 10.1056/ NEJM199906103402306
- Miki, T., Yuda, S., Kouzu, H., and Miura, T. (2013). Diabetic cardiomyopathy: pathophysiology and clinical features. *Heart Fail. Rev.* 18, 149–166. doi: 10. 1007/s10741-012-9313-3
- Miller, V. M., and Best, P. J. (2011). Implications for reproductive medicine: sex differences in cardiovascular disease. Sex. Reprod. Menopause 9, 21–28.
- Mozaffarian, D., Benjamin, E. J., Go, A. S., Arnett, D. K., Blaha, M. J., Cushman, M., et al. (2015). Heart disease and stroke statistics–2015 update: a report from the American Heart Association. *Circulation* 131, e29–322. doi: 10.1161/CIR. 00000000000152
- Mulder, G. B., Luo, S., and Gramlich, P. (2010). The Zucker Diabetic Fatty (ZDF) Rat Diet Evaluation for the Induction of Type 2 Diabetes in Obese Female ZDF Rats. Available at: http://www.criver.com/files/pdfs/rms/zdf/rm\_rm\_r\_ zdf\_diet\_eval\_tech\_sheet.aspx
- Natarajan, S., Liao, Y., Cao, G., Lipsitz, S. R., and McGee, D. L. (2003). Sex differences in risk for coronary heart disease mortality associated with diabetes and established coronary heart disease. *Arch. Intern. Med.* 163, 1735–1740. doi: 10.1001/archinte.163.14.1735
- Nilsson, S., and Gustafsson, J. A. (2011). Estrogen receptors: therapies targeted to receptor subtypes. Clin. Pharmacol. Ther. 89, 44–55. doi: 10.1038/clpt.2010.226
- Norhammar, A., and Schenck-Gustafsson, K. (2013). Type 2 diabetes and cardiovascular disease in women. *Diabetologia* 56, 1–9. doi: 10.1007/s00125-012-2694-y
- Okumura, M., Iwai, M., Ide, A., Mogi, M., Ito, M., and Horiuchi, M. (2005). Sex difference in vascular injury and the vasoprotective effect of valsartan are related to differential AT2 receptor expression. *Hypertension* 46, 577–583. doi: 10.1161/01.HYP.0000178564.14464.80
- Oldroyd, S. D., Thomas, G. L., Gabbiani, G., and El Nahas, A. M. (1999). Interferongamma inhibits experimental renal fibrosis. *Kidney Int.* 56, 2116–2127. doi: 10.1046/j.1523-1755.1999.00775.x
- Olivetti, G., Giordano, G., Corradi, D., Melissari, M., Lagrasta, C., Gambert, S. R., et al. (1995). Gender differences and aging: effects on the human heart. J. Am. Coll. Cardiol. 26, 1068–1079. doi: 10.1016/0735-1097(95) 00282-8
- Patten, R. D., Pourati, I., Aronovitz, M. J., Baur, J., Celestin, F., Chen, X., et al. (2004). 17beta-estradiol reduces cardiomyocyte apoptosis in vivo and in vitro via activation of phospho-inositide-3 kinase/Akt signaling. *Circ. Res.* 95, 692–699. doi: 10.1161/01.RES.0000144126.57786.89
- Perseghin, G., Ntali, G., De Cobelli, F., Lattuada, G., Esposito, A., Belloni, E., et al. (2007). Abnormal left ventricular energy metabolism in obese men with preserved systolic and diastolic functions is associated with insulin resistance. *Diabetes Care* 30, 1520–1526. doi: 10.2337/dc06-2429
- Peters, S. A., Huxley, R. R., and Woodward, M. (2014). Diabetes as risk factor for incident coronary heart disease in women compared with men: a systematic review and meta-analysis of 64 cohorts including 858,507 individuals and 28,203 coronary events. *Diabetologia* 57, 1542–1551. doi: 10.1007/s00125-014-3260-6
- Peterson, L. R., Herrero, P., Schechtman, K. B., Racette, S. B., Waggoner, A. D., Kisrieva-Ware, Z., et al. (2004). Effect of obesity and insulin resistance on myocardial substrate metabolism and efficiency in young women. *Circulation* 109, 2191–2196. doi: 10.1161/01.CIR.0000127959.28627.F8
- Pulakat, L., DeMarco, V. G., Ardhanari, S., Chockalingam, A., Gul, R., Whaley-Connell, A., et al. (2011). Adaptive mechanisms to compensate for overnutrition-induced cardiovascular abnormalities. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 301, R885–R895. doi: 10.1152/ajpregu.00316.2011
- Qi, Y., Li, H., Shenoy, V., Li, Q., Wong, F., Zhang, L., et al. (2012). Moderate cardiac-selective overexpression of angiotensin II type 2 receptor protects

cardiac functions from ischaemic injury. *Exp. Physiol.* 97, 89–101. doi: 10.1113/ expphysiol.2011.060673

- Romanens, M., Fankhauser, S., Saner, B., Michaud, L., and Saner, H. (1999). No evidence for systolic or diastolic left ventricular dysfunction at rest in selected patients with long-term type I diabetes mellitus. *Eur. J. Heart Fail.* 1, 169–175. doi: 10.1016/S1388-9842(99)00012-4
- Rosano, G. M., Leonardo, F., Pagnotta, P., Pelliccia, F., Panina, G., Cerquetani, E., et al. (1999). Acute anti-ischemic effect of testosterone in men with coronary artery disease. *Circulation* 99, 1666–1670. doi: 10.1161/01.CIR.99.13.1666
- Rubler, S., Dlugash, J., Yuceoglu, Y. Z., Kumral, T., Branwood, A. W., and Grishman, A. (1972). New type of cardiomyopathy associated with diabetic glomerulosclerosis. Am. J. Cardiol. 30, 595–602. doi: 10.1016/0002-9149(72) 90595-4
- Ruige, J. B., Mahmoud, A. M., De Bacquer, D., and Kaufman, J. M. (2011). Endogenous testosterone and cardiovascular disease in healthy men: a metaanalysis. *Heart* 97, 870–875. doi: 10.1136/hrt.2010.210757
- Salinero, A. E., Anderson, B. M., and Zuloaga, K. L. (2018). Sex differences in the metabolic effects of diet-induced obesity vary by age of onset. *Int. J. Obes.* doi: 10.1038/s41366-018-0023-3 [Epub ahead of print].
- Sampson, A. K., Moritz, K. M., Jones, E. S., Flower, R. L., Widdop, R. E., and Denton, K. M. (2008). Enhanced angiotensin II type 2 receptor mechanisms mediate decreases in arterial pressure attributable to chronic lowdose angiotensin II in female rats. *Hypertension* 52, 666–671. doi: 10.1161/ HYPERTENSIONAHA.108.114058
- Sandoo, A., van Zanten, J. J., Metsios, G. S., Carroll, D., and Kitas, G. D. (2010). The endothelium and its role in regulating vascular tone. *Open Cardiovasc. Med. J.* 4, 302–312. doi: 10.2174/1874192401004010302
- Sawhney, V., Brouilette, S., Campbell, N., Coppen, S., Baker, V., Hunter, R., et al. (2018). Association of genetic variation in telomere-related SNP and telomerase with ventricular arrhythmias in ischemic cardiomyopathy. *Pacing Clin. Electrophysiol.* doi: 10.1111/pace.13284 [Epub ahead of print].
- Schilling, J. D., and Mann, L. D. (2014). Diabetic cardiomyopathy: distinct and preventable entity or inevitable consequence? *Curr. Cariovasc. Risk Rep.* 8:417. doi: 10.1007/s12170-014-0417-2
- Semeniuk, L. M., Kryski, A. J., and Severson, D. L. (2002). Echocardiographic assessment of cardiac function in diabetic db/db and transgenic db/db-hGLUT4 mice. Am. J. Physiol. Heart Circ. Physiol. 283, H976–H982. doi: 10.1152/ ajpheart.00088.2002
- Severson, D. L. (2004). Diabetic cardiomyopathy: recent evidence from mouse models of type 1 and type 2 diabetes. *Can. J. Physiol. Pharmacol.* 82, 813–823. doi: 10.1139/y04-065
- Sharma, S., Adrogue, J. V., Golfman, L., Uray, I., Lemm, J., Youker, K., et al. (2004). Intramyocardial lipid accumulation in the failing human heart resembles the lipotoxic rat heart. *FASEB J.* 18, 1692–1700. doi: 10.1096/fj.04-2263com
- Shi, J., Li, J., Guan, H., Cai, W., Bai, X., Fang, X., et al. (2014). Anti-fibrotic actions of interleukin-10 against hypertrophic scarring by activation of PI3K/AKT and STAT3 signaling pathways in scar-forming fibroblasts. *PLoS One* 9:e98228. doi: 10.1371/journal.pone.0098228
- Simons, L. A., McCallum, J., Friedlander, Y., and Simons, J. (1996). Diabetes, mortality and coronary heart disease in the prospective Dubbo study of Australian elderly. *Aust. N. Z. J. Med.* 26, 66–74. doi: 10.1111/j.1445-5994.1996. tb02909.x
- Skorska, A., von Haehling, S., Ludwig, M., Lux, C. A., Gaebel, R., Kleiner, G., et al. (2015). The CD4(+) AT2R(+) T cell subpopulation improves post-infarction remodelling and restores cardiac function. J. Cell. Mol. Med. 19, 1975–1985. doi: 10.1111/jcmm.12574
- Spark, R. F. (2007). Testosterone, diabetes mellitus, and the metabolic syndrome. Curr. Urol. Rep. 8, 467–471. doi: 10.1007/s11934-007-0050-4
- Stahrenberg, R., Edelmann, F., Mende, M., Kockskamper, A., Dungen, H. D., Scherer, M., et al. (2010). Association of glucose metabolism with diastolic function along the diabetic continuum. *Diabetologia* 53, 1331–1340. doi: 10. 1007/s00125-010-1718-8
- Stanley, W. C., Lopaschuk, G. D., and McCormack, J. G. (1997). Regulation of energy substrate metabolism in the diabetic heart. *Cardiovasc. Res.* 34, 25–33. doi: 10.1016/S0008-6363(97)00047-3
- Suran, D., Sinkovic, A., and Naji, F. (2016). Tissue Doppler imaging is a sensitive echocardiographic technique to detect subclinical systolic and diastolic

dysfunction of both ventricles in type 1 diabetes mellitus. *BMC Cardiovasc. Disord.* 16:72. doi: 10.1186/s12872-016-0242-2

- Varma, U., Koutsifeli, P., Benson, V. L., Mellor, K. M., and Delbridge, L. M. D. (2017). Molecular mechanisms of cardiac pathology in diabetes - Experimental insights. *Biochim. Biophys. Acta* 1864, 1949–1959. doi: 10.1016/j.bbadis.2017. 10.035
- Venkov, C. D., Rankin, A. B., and Vaughan, D. E. (1996). Identification of authentic estrogen receptor in cultured endothelial cells. A potential mechanism for steroid hormone regulation of endothelial function. *Circulation* 94, 727–733. doi: 10.1161/01.CIR.94.4.727
- Vrtacnik, P., Ostanek, B., Mencej-Bedrac, S., and Marc, J. (2014). The many faces of estrogen signaling. *Biochem. Med.* 24, 329–342. doi: 10.11613/BM.2014.035
- Wachter, R., Luers, C., Kleta, S., Griebel, K., Herrmann-Lingen, C., Binder, L., et al. (2007). Impact of diabetes on left ventricular diastolic function in patients with arterial hypertension. *Eur. J. Heart Fail.* 9, 469–476. doi: 10.1016/j.ejheart.2007. 01.001
- Wang, Y., Cao, Y., Yamada, S., Thirunavukkarasu, M., Nin, V., Joshi, M., et al. (2015). Cardiomyopathy and worsened ischemic heart failure in SM22alpha Cre-mediated neuropilin-1 null mice: dysregulation of PGC1alpha and mitochondrial homeostasis. *Arterioscler. Thromb. Vasc. Biol.* 35, 1401–1412. doi: 10.1161/ATVBAHA.115.305566
- Wannamethee, S. G., Papacosta, O., Lawlor, D. A., Whincup, P. H., Lowe, G. D., Ebrahim, S., et al. (2012). Do women exhibit greater differences in established and novel risk factors between diabetes and non-diabetes than men? The British Regional Heart Study and British Women's Heart Health Study. *Diabetologia* 55, 80–87. doi: 10.1007/s00125-011-2284-4
- Weiner, C. P., Lizasoain, I., Baylis, S. A., Knowles, R. G., Charles, I. G., and Moncada, S. (1994). Induction of calcium-dependent nitric oxide synthases by sex hormones. *Proc. Natl. Acad. Sci. U.S.A.* 91, 5212–5216. doi: 10.1073/pnas. 91.11.5212
- Widdop, R. E., Jones, E. S., Hannan, R. E., and Gaspari, T. A. (2003). Angiotensin AT2 receptors: cardiovascular hope or hype? *Br. J. Pharmacol.* 140, 809–824. doi: 10.1038/sj.bjp.0705448
- Wilson, M. S., Elnekave, E., Mentink-Kane, M. M., Hodges, M. G., Pesce, J. T., Ramalingam, T. R., et al. (2007). IL-13Ralpha2 and IL-10 coordinately suppress airway inflammation, airway-hyperreactivity, and fibrosis in mice. J. Clin. Invest. 117, 2941–2951. doi: 10.1172/JCI31546
- World Health Organization [WHO] (2017). Cardiovascular Diseases (CVD) [Online]. Geneva: World Health Organization.
- Wu, Y., Cazorla, O., Labeit, D., Labeit, S., and Granzier, H. (2000). Changes in titin and collagen underlie diastolic stiffness diversity of cardiac muscle. J. Mol. Cell Cardiol. 32, 2151–2162. doi: 10.1006/jmcc.2000.1281
- Wu, Y., Shen, Y., Kang, K., Zhang, Y., Ao, F., Wan, Y., et al. (2015). Effects of estrogen on growth and smooth muscle differentiation of vascular wall-resident CD34(+) stem/progenitor cells. *Atherosclerosis* 240, 453–461. doi: 10.1016/j. atherosclerosis.2015.04.008
- Xu, Y., Arenas, I. A., Armstrong, S. J., and Davidge, S. T. (2003). Estrogen modulation of left ventricular remodeling in the aged heart. *Cardiovasc. Res.* 57, 388–394. doi: 10.1016/S0008-6363(02)00705-8
- Yang, X. P., and Reckelhoff, J. F. (2011). Estrogen, hormonal replacement therapy and cardiovascular disease. *Curr. Opin. Nephrol. Hypertens.* 20, 133–138. doi: 10.1097/MNH.0b013e3283431921
- Yeh, J. K., and Wang, C. Y. (2016). Telomeres and telomerase in cardiovascular diseases. *Genes* 7:E58. doi: 10.3390/genes7090058
- Yen, S. S. C., and Jade, R. B. (eds) (1991). Reproductive Endocrinology: Physiology, Pathophysiology and Clinical Management. Philadelphia, PA: W.B. Saunders.
- Zarich, S. W., Arbuckle, B. E., Cohen, L. R., Roberts, M., and Nesto, R. W. (1988). Diastolic abnormalities in young asymptomatic diabetic patients assessed by pulsed Doppler echocardiography. J. Am. Coll. Cardiol. 12, 114–120. doi: 10. 1016/0735-1097(88)90364-6
- Zhang, B., Shen, Q., Chen, Y., Pan, R., Kuang, S., Liu, G., et al. (2017). Myricitrin alleviates oxidative stress-induced inflammation and apoptosis and protects mice against diabetic cardiomyopathy. *Sci. Rep.* 7:44239. doi: 10.1038/srep44239
- Zhang, H., Morgan, B., Potter, B. J., Ma, L., Dellsperger, K. C., Ungvari, Z., et al. (2010). Resveratrol improves left ventricular diastolic relaxation in type 2 diabetes by inhibiting oxidative/nitrative stress: in vivo demonstration with magnetic resonance imaging. Am. J. Physiol. Heart Circ. Physiol. 299, H985– H994. doi: 10.1152/ajpheart.00489.2010

- Zhao, J., Imbrie, G. A., Baur, W. E., Iyer, L. K., Aronovitz, M. J., Kershaw, T. B., et al. (2013). Estrogen receptor-mediated regulation of microRNA inhibits proliferation of vascular smooth muscle cells. *Arterioscler. Thromb. Vasc. Biol.* 33, 257–265. doi: 10.1161/ATVBAHA.112.300200
- Zhao, R., Le, K., Moghadasian, M. H., and Shen, G. X. (2013). Regulatory role of NADPH oxidase in glycated LDL-induced upregulation of plasminogen activator inhibitor-1 and heat shock factor-1 in mouse embryo fibroblasts and diabetic mice. *Free Radic. Biol. Med.* 61, 18–25. doi: 10.1016/j.freeradbiomed. 2013.03.009
- Zucker, I., and Beery, A. K. (2010). Males still dominate animal studies. *Nature* 465:690. doi: 10.1038/465690a

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Toedebusch, Belenchia and Pulakat. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.