



# Paternal Determinants in Preeclampsia

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Preeclampsia is a condition associated with high rates of maternal-fetal morbidity and mortality. It usually occurs in 3-10% of nulliparous women and 18% of previously affected women. Different lines of evidence have demonstrated the role of the father in the onset of preeclampsia. The placenta is the cornerstone of preeclampsia and poses important paternal genetic determinants; in fact, the existence of a "paternal antigen" has been proposed. Nulliparity is a well-known risk factor. Change of partner to a woman without history of preeclampsia increases the risk; however, this change decreases in women with history of the condition. High interval between pregnancies, short sexual intercourse before pregnancy, and conception by intracytoplasmic sperm injection suggest a limited exposure to the so-called paternal antigen. A man who was born from a mother with preeclampsia also increases the risk to his partner. Not only maternal but also paternal obesity is a risk factor for preeclampsia. Fetal HLA-G variants from the father increased the immune incompatibility with the mother and are also significantly associated with preeclampsia in multigravida pregnancies. An analysis of a group of Swedish pregnant women showed that the risk for preeclampsia is attributable to paternal factors in 13% of cases, which could be related to genetic interactions with maternal genetic factors. This review aimed to evaluate the evidences of the father's contribution to the onset of preeclampsia and determine the importance of including them in future studies.

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### BACKGROUND

Preeclampsia is a condition in pregnant women associated with high rates of maternal and fetal morbidity and mortality worldwide. This disease is characterized as a systemic syndrome with *de novo* hypertension occurring after 20 weeks of gestation as well as proteinuria (300 mg in 24 h) (NHBPEP, 2000). In 2013, the Task Force on Hypertension in Pregnancy established new diagnostic criteria for hypertension in pregnancy.

Preeclampsia is related to deficient placental implantation, with multisystem consequences (generalized endotheliosis) in the mother (Roberts and Hubel, 2009). Its incidence in industrialized

countries ranges from 3 to 5% (Stone et al., 1995; Dahlstrom et al., 2006; Wallis et al., 2008), which reaches up to 16% in Nigeria (Osungbade and Ige, 2011). The frequency of maternal and perinatal deaths in Mexico is 34 and 33%, respectively (Peralta Pedrero et al., 2006).

Preeclampsia is classified as mild and severe and can complicate to hemolysis, elevated liver enzymes, and low platelet count (HELLP) syndrome and eclampsia (Geller et al., 2004).

The pathophysiology of preeclampsia is characterized by placental hypoxia and/or ischemia leading to overexpression of hypoxia-inducible factor 1 (HIF1), which in turn increases the expression of the soluble isoform of vascular endothelial growth factor (sFlt) (Nevo et al., 2006). Hypoxic placenta also releases the soluble endoglin, antagonizing the production of endothelial nitric oxide synthase (eNOS) through sequestering of transforming growth factor beta1 (TGFb1) (Sandrim et al., 2008). In addition, the proangiogenic factors such as the placental growth factor (PIGF) and vascular endothelial growth factor (VEGF) (Reuvekamp et al., 1999). Together, they can trigger the antiangiogenic state in the mother, which resulted in generalized endothelial dysfunction.

Risk factors for disease development include nulliparity, pregnancy with multiple products, previous history of preeclampsia, vascular and connective tissue diseases, maternal age of >35 years, Afro-American ethnicity, preexistent renal disease, arterial hypertension, type 2 diabetes, and obesity (Eiland et al., 2012).

Previous studies on preeclampsia have considered the mother's participation almost exclusively. However, the placenta is a transient biparental organ with maternal and paternal contributions. In this way, changes on the expression profile of genes involved in the metabolism and transport in the placenta depend on various maternal and paternal genetic profiles (Zusterzeel et al., 2002; Dekker et al., 2011).

Increasing evidence has demonstrated the father's role in the onset of preeclampsia. In Astin et al. (1981) reported a case of a man who fathered two consecutive women who developed severe preeclampsia and passed away, suggesting the existence of a "fatal father factor."

The risk of preeclampsia is genetically attributable to the mother (35%), fetus (20%), and couple (13%) (Cnattingius et al., 2004). Women and men who were born from a pregnancy with preeclampsia are at higher risk to have a baby, result of a preeclamptic pregnancy (Esplin et al., 2001). The change of partner increases the risk for preeclampsia in 1.6%, which increases up to 2.9% in a woman whose second pregnancy is the result of union with a man who has had a previous partner with preeclampsia (Lie et al., 1998). Change of partner plays a predisposing or protective role, depending of the presence or absence of the disease in the first pregnancy (Wikström et al., 2012). The presence of fetal variants of HLA-G from the father and those outside the mother generate a paternal-fetal susceptibility component for the development of preeclampsia (Tan et al., 2008).

Paternal genetic material plays an important role for the onset of preeclampsia. A triploid (69 XXX) partial mole, with paternal isodisomy in the placenta and a disomic fetus, increases

preeclampsia-like symptoms at 19 weeks of gestation (Yoneda et al., 2013). Although preeclampsia is a human-specific disorder, animal models of the disease have been created to show the role of the paternal genetic factors. A female transgenic mouse for human angiotensinogen gene was mated with a male transgenic mouse for human renin gene, originating a preeclampsia-like syndrome (Takimoto et al., 1996).

In this review, we aimed to describe different lines of evidence regarding the paternal contribution on preeclampsia.

# EPIDEMIOLOGICAL AND CLINICAL EVIDENCE

Preeclampsia has been traditionally considered as a disease during first pregnancies; however, an early study by Need (1975) showed that a woman, pregnant by different fathers, had a healthy twin pregnancy with the first men, but severe preeclampsia occurred in the second one. The analysis of 34,201 patients revealed the presence of 47 multigravid patients with severe preeclampsia. A new partner was demonstrated in 13 among them (19.1%), and significant difference was observed than those in the control group (p < 0.01) (Feeney and Scott, 1980). Ikedife (1980) showed that change of partners was observed in 34 (74%) out of 46 multiparous patients with eclampsia. A woman with three different husbands presented preeclampsia only in the first pregnancy of the second and third spouses. Therefore, preeclampsia is not just a primiparous woman disease but also a condition related to the first pregnancy with a particular partner (Chng, 1982). The Guadeloupe study revealed an increased change of partner in multiparous women who were affected with preeclampsia (Robillard et al., 1993). The same group demonstrated that the incidence of preeclampsia was 11.9% among primigravidae, 4% among multigravidae without a change of paternity, and 24% among multigravidae with a new partner; however, these numbers depend on the duration of sexual cohabitation before conception (Robillard et al., 1994).

When evaluating paternal vs. maternal half-sisters with preeclampsia, Lie et al. (1998) found an increased risk for the disease in the former. Likewise, they showed that 13% of primiparous mothers with preeclampsia had recurrence in the second pregnancy, decreasing the influence of partner's change (11.8%), which is opposite in mothers without preeclampsia in their first pregnancy (Lie et al., 1998).

The same results were obtained by Li and Wi (2000). Pipkin (2001) observed that partner's change increases the risk for preeclampsia occurrence in primiparous women.

A woman pregnant by a partner who previously fathered a woman with preeclampsia was highly at risk to develop the disease. In 2004, Cnatinggius et al. evaluated three different scenarios: (1) mothers without preeclampsia and no partner change (control group); (2) mothers with previous preeclampsia, current preeclampsia, and change of spouse who was not fathered by a previous couple with preeclampsia; and (3) mothers with preeclampsia and change of couple who fathered another woman with preeclampsia. The maternal effect was 45%, meanwhile the paternal one was 10% (Cnattingius et al., 2004).

Conversely, Chigbu et al. (2009) evaluated two groups of Nigerian women with and without change of partner in their second pregnancy and found no differences for the development of preeclampsia.

Marti and Herrmann (1977) found that preeclampsia is associated with shorter exposure to spermatozoa in younger women and more frequent use of barrier contraceptive methods. Therefore, the authors coined the term "immunogestosis" to denote the immune nature of preeclampsia. The use of barrier contraceptive methods was significantly higher in women with preeclampsia compared to healthy women [odds ratio (OR) = 2.48], and the number of sexual contacts was inversely related to the risk of preeclampsia (Klonoff-Cohen et al., 1989). Conversely, Mills et al. (1991) and Ness et al. (2004) found no differences in the use of barrier contraceptive methods between women with and without preeclampsia.

Shorter sexual intercourse increases the shorter antigenic seminal exposure (Beer, 1989). The risk of pregnancy-induced hypertension (PIH) was increased when conception is within 12 months of sexual cohabitation: 40% in 0-4 months, 23% in 5-8 months, 15% in 9-12 months, and 5% after 12 months (Robillard et al., 1994). The same author compared patients with simple PIH versus those with preeclampsia and eclampsia and observed that the sexual cohabitation times are shorter (9.5 months) compared to those of unaffected women (26.3 months) (Robillard and Hulsey, 1996). Koelman et al. (2000) found a lower frequency of oral sex in women with preeclampsia compared to healthy pregnant women (p = 0.0003), suggesting a protective role. The authors report lower amounts of soluble HLA A and HLA B in the seminal fluid, in partners of preeclamptic women (Koelman et al., 2000). In Robertson et al. (2003) proposed how repeated semen exposure protects preeclampsia, based on four lines of evidence: (1) the semen contains antigens shared by the conception; (2) after the seminal contact, the maternal mucosa can mount a regulated immune response to semen antigens; (3) semen contains among others, high amounts of TGFb that can inhibit type 1 immunity; and (4) TGFb-dependent changes in T-lymphocytes allow a hyporesponsiveness to paternal antigens. The exposure to seminal fluid through the vagina is inversely correlated with the risk of preeclampsia occurrence, whereas the oral exposure to seminal fluid has no effect on disease development (Saftlas et al., 2014).

A clear increase in the frequency of preeclampsia when the father (OR = 2.1) and mother (OR = 3.3) are products of preeclampsia-complicated pregnancies has been observed; however, this study did not consider the changes of paternity (Conde-Agudelo, 2001). In men who were born after a pregnancy complicated by preeclampsia, the risk in the first pregnancy was moderately increased compared with men who were born after a pregnancy without preeclampsia (OR = 1.5, 1.3–1.7).

This observed risk increases when severe or early preeclampsia is considered (OR = 1.9, 1.4–2.5) (Skjaerven et al., 2005). Lie (2007) found similar numbers: fathers who came from a preeclamptic pregnancy had a 1.5-fold risk [95% CI 1.3–1.7] of fathering a preeclamptic pregnancy.

Since, Need et al. (1983) reported a significantly higher frequency of preeclampsia in women with abortions (15.7%)

than those with normal pregnancy (4.7%). They also found that azoospermia and oligospermia present a lower frequency of preeclampsia of 8.7 and 13.6%, respectively, which could be related to the minimal antigenic sperm exposure. The frequency of preeclampsia was higher in the donor insemination program than those in the father insemination program (OR = 1.20, 95%) (Smith et al., 1997). Hoy et al. (1999) revealed a higher frequency of preeclampsia in donor-inseminated women vs. natural-inseminated women (OR = 1.4, 95% CI). In Salha et al. (1999) compared 72 infertile women subjected to sperm, ovum, or embryo donation, with the same number of pregnant women through insemination with their own ovum or partner's spermatozoa (control group). Fourteen patients developed preeclampsia, 13 of whom belong to the group of donated gametes and 1 to the control group. Kyrou et al. (2010) found a marginally significant (p = 0.05) higher frequency of preeclampsia in women conceiving by a sperm donor compared to partners in spermatozoa insemination. The type of spermatozoa and number of previous insemination cycles were the variables that influenced the risk of preeclampsia (p = 0.012); in fact, the authors observed that the fewer the number of the insemination cycles, the higher the risk of preeclampsia (Kyrou et al., 2010). A recent meta-analysis of seven studies showed the association with preeclampsia in women conceiving with donor sperm (OR = 1.63) (González-Comadran et al., 2014). Another study evaluated the risk of preeclampsia in infertile women subjected only to sperm donation via intrauterine insemination (IUI) or in vitro fertilization (IVF), compared to those with primary sperm donation (IUI or IVF) followed by egg donation. A higher frequency of preeclampsia in the latter was observed; therefore, the authors conclude that double gamete donation is associated with increased risk for preeclampsia (Bartal et al., 2018).

The aforementioned studies support the theory of the immunological basis of preeclampsia.

## **IMMUNOLOGICAL EVIDENCE**

Preeclampsia is a state in which alloantigen (placenta of paternal origin) must be recognized to avoid rejection (Saito et al., 2007). Trophoblast cells must express paternal alloantigens that must be recognized by the mother's immune system. Extravillous trophoblast express different HLA-C, E, F, and G (Hackmon et al., 2017). HLA-C is the ligand of immunoglobulinlike receptors (KIR) that are expressed in decidual natural killer (NK) cells (Sharkey et al., 2008). HLA-C and KIR are polymorphic; therefore, many maternal/paternal different combinations are possible. Two KIR haplotypes exist A and B, with the latter stimulating the expression of chemokines and angiogenic cytokines, promoting trophoblast invasiveness. Therefore, haplotype B could be protective for preeclampsia (Redman and Sargent, 2010). Seminal priming triggers a cascade of events for placental recognition or rejection. Seminal fluid contains high amounts of TGF-b that induces T-regulatory cells (Treg); these cells modulate immune responses in an antigenspecific way. Therefore, the effects of HLA-C/KIR interaction plus the seminal priming activity of TGF-b could in some

extent explain the immune nature of father's involvement in preeclampsia (Redman and Sargent, 2010).

The immune nature of preeclampsia was observed by Need (1975), who demonstrated histoincompatibility in a mother who developed preeclampsia with her second partner through the evaluation of HL-A typing. In Feeney et al. (1977) found a lower incidence of preeclampsia in previously blood-transfused women, compared with the same number of non-transfused primigravidas. A similar effect is observed in patients with kidney transplantation (Feeney et al., 1977). In the same year, Marti and Herrmann, (1977) found a correlation between the number of exposures to semen and lower frequency of preeclampsia; they coined the term immunogestosis to explain both, the immunologic tolerance and immunologic enhancement that abrogates immunoreaction against paternal and fetal histocompatibility antigens.

Birkeland and Kristofferson (1979) evaluated the immune response in mothers with and without preeclampsia and found no leukocyte antigens against the father in women with normal pregnancies, meanwhile these antigens were identified in one woman with severe preeclampsia. The evaluation of HLA A, B, and DR in women with severe and mild preeclampsia as well as their husbands and babies revealed a higher frequency of DR4 in all family members in severe preeclampsia (Kilpatrick et al., 1987). However, HLA A, B, and C are expressed in low amounts in the trophoblast, and HLA-G protein is exclusively expressed in trophoblast cells in high amounts. The evaluation of 1597del/C allele was not associated with preeclampsia (Aldrich et al., 2000). Three polymorphisms in HLA-G were evaluated in 68 primigravida trios, but were not associated with preeclampsia (Bermingham et al., 2000). The evaluation of 15 alleles in 4 exons of HLA-G in 155 family triads showed an overrepresentation of a homozygous HLA-G genotype in 40 pre-eclamptic offspring compared to 70 controls (p = 0.002) among primiparous women; further analyses suggested that the differences between pre-eclamptic cases and controls were primarily accomplished by a different transmission from the father of a 14 bp deletion/insertion polymorphism in the 3'UTR region (14 bp del/in) (p = 0.006) (Hylenius et al., 2004). In Tan et al. (2008) observed a significant association between paternally inherited HLA-G allele G\*0106 in the fetus and an increased risk for preeclampsia, but only in multigravid pregnancies. The 14 bp del/in was evaluated in three different combinations: mother/offspring, father/offspring, and couples; heterozygosity in the mother plus double insertion in babies was significantly higher in severe early-onset preeclampsia (p = 0.023), and the frequency of double deletion in both the father and baby was lower in severe early-onset preeclampsia (p = 0.024). The analysis of couples did not reveal significant differences between cases and controls (Zhang et al., 2012). A recent meta-analysis of 1,625 cases and 2,145 controls in all members of the triads evaluated the influence of 14 bp del/in in the onset of preeclampsia; the results did not reveal the association with the disease in offspring, mothers, or fathers. A stratification showed the association of 14 bp del/in with preeclampsia in European Caucasian offspring, but not found in African descent population (Pabalan et al., 2015).

Semen is not sterile, and its microbes have also been considered as a potential cause of preeclampsia. Repeated exposure to semen seems to create a memory protecting women from preeclampsia and the same could be possible in the case of semen microbes. Therefore, common elements of preeclampsia and infections such as Galectin13, Toll-like receptors, and antiphospholipid syndrome are found (Kenny and Kell, 2018).

# **GENETIC EVIDENCE**

The genetic nature of preeclampsia has also been evaluated through (1) familial cases, (2) twin studies, (3) consanguinity studies, (4) candidate gene evaluation, and (5) linkage analysis. Genes are involved in different pathophysiological mechanisms involved in preeclampsia. Genetic studies that included mother/father/children triads, which allow to determine the sole or joint contribution for preeclampsia, were limited.

evaluation of thrombophilic genes methyle netetrahydrofolate reductase (MTHFR) and FVL in 92 mother/father/child triads revealed an increased risk only in mothers carrying two mutated copies of MTHFR and one mutated allele in FVL. Therefore, the risk of preeclampsia is not increased in the presence of fetal MTHFR or FVL mutations (Vefring et al., 2004). In van Dijk et al. (2010) observed a methylated paternal copy of STOX1 gene and an unmethylated active maternal copy of the gene. Maternal transmission of this gene has been demonstrated in preeclampsia (van Dijk and Oudejans, 2011). The evaluation of GSTP1, eNOS, and LPL genes in 167 preeclamptic and control triads in a Greek cohort, which significantly demonstrated higher frequencies of Val105 allele (GSTP1) and Glu298Asp (eNOS) in control vs. preeclamptic groups in mothers, fathers, and child. The -93 polymorphism (LDL) was higher in preeclampsia only in mothers, but not in fathers or children. The transmission disequilibrium test revealed no differences in the rate of transmission of the studied common vs. mutated alleles (Pappa et al., 2011).

In Zhou et al. (2013) found a significant association of C4599A polymorphism in AGTR2 gene with preeclampsia in mothers with body mass index (BMI) of  $\geq 25 \text{ kg/m}^2$  in their partners and studied children. Galaviz-Hernandez et al. (2016) evaluated the polymorphism rs5370 of EDN1 gene in mothers with preeclampsia and their partners, showing a significant negative association with the disease in case of fathers (OR = 0.42; CI 95%, 0.18-0.94, p = 0.034), which was strengthened after adjusting the paternal protective factors. The evaluation of two polymorphisms in SOD1 (+35A/C) and SOD2 (Ala16Val (C/T) genes in 698 mother/father/infant triads revealed a significant association with preeclampsia in fathers with Ala16Val (TT genotype) [OR = 2.77 (1.32-5.81), p = 0.007]. This study revealed essentially the same risk for preeclampsia in both combined TT genotypes in mothers and fathers [OR = 6.80 (2.32–19.95), p < 0.001] and mother/father/infant triads [OR = 6.46 (2.16-19.31), p < 0.001] (Luo et al., 2018). Polymorphisms in the thrombophilic genes factor V Leiden, prothrombin, and MTHFR were evaluated in women, fetus, and fathers as risk factors for pregnancyassociated complications, including preeclampsia. The authors

found significant differences between cases and controls in maternal Factor V and fetal *MTHFR*, but no differences on any of the polymorphisms analyzed in the father's group (Nevalainen et al., 2018).

### **CONCLUSION AND PERSPECTIVES**

Epidemiological, clinical, immunological, and genetic evidences supported the contribution of fathers in the onset of preeclampsia. Despite this, few studies were intended to evaluate the spouses' role. In this way, the evaluation of paternal-derived immune and genetic materials was performed in order to identify the risk and prognostic markers of preeclampsia. Recent advances in placental epigenetics, along with the use of OMICS tools, ensure the identification of molecular markers

### REFERENCES

- Aldrich, C., Verp, M. S., Walker, M. A., and Ober, C. (2000). A null mutation in HLA-G is not associated with preeclampsia or intrauterine growth retardation. *J. Reprod. Immunol.* 47, 41–48. doi: 10.1016/S0165-0378(00)00052-8
- Astin, M., Scott, J. R., and Worley, R. J. (1981). Pre-eclampsia/eclampsia: a fatal father factor. *Lancet* 2:533. doi: 10.1016/S0140-6736(81)90925-9
- Bartal, M. F., Sibai, B. M., Bart, Y., Shina, A., Mazaki-Tovi, S., Eisen, I. S., et al. (2018). The impact of sperm and egg donation on the risk of pregnancy complications. Am. J. Perinatol. doi: 10.1055/s-0038-1667029 [Epub ahead of print].
- Beer, A. E. (1989). Immunology, contraception, and preeclampsia. *JAMA* 262:3184. doi: 10.1001/jama.1989.03430220107040
- Bermingham, J., Jenkins, D., McCarthy, T., and O'Brien, M. (2000). Genetic analysis of insulin-like growth factor II and HLA-G in pre-eclampsia. *Biochem. Soc. Trans.* 28, 215–219. doi: 10.1042/bst0280215
- Birkeland, S. A., and Kristofferson, K. (1979). Pre-eclampsia-a state of mother-fetus immune imbalance. *Lancet* 2, 720-723. doi: 10.1016/S0140-6736(79) 90645-7
- Chigbu, C. O., Okezie, O. A., and Odugu, B. U. (2009). Women in southern Nigeria with change in paternity do not have increased incidence of pre-eclampsia. J. Obstet. Gynaecol. 29, 94–97. doi: 10.1080/01443610802660927
- Chng, P. K. (1982). Occurrence of pre-eclampsia in pregnancies to three husbands. Case report. *Br. J. Obstet. Gynaecol.* 89, 862–863. doi: 10.1111/j.1471-0528.1982. tb05042.x
- Cnattingius, S., Reilly, M., Pawitan, Y., and Lichtenstein, P. (2004). Maternal and fetal genetic factors account for most of familial aggregation of preeclampsia: a population-based Swedish cohort study. *Am. J. Med. Genet.* 130A, 365–371. doi: 10.1002/ajmg.a.30257
- Conde-Agudelo, A. (2001). Paternal and maternal components of the predisposition to preeclampsia. N. Engl. J. Med. 345:149. doi: 10.1056/ NEJM200107123450217
- Dahlstrom, B. L., Engh, M. E., Bukholm, G., and Oian, P. (2006). Changes in the prevalence of pre-eclampsia in Akershus County and the rest of Norway during the past 35 years. Acta Obstet. Gynecol. Scand. 85, 916–921. doi: 10.1080/ 00016340500442449
- Dekker, G., Robillard, P. Y., and Roberts, C. (2011). The etiology of preeclampsia: the role of the father. *J. Reprod. Immunol.* 89, 126–132. doi: 10.1016/j.jri.2010. 12.010
- Eiland, E., Nzerue, C., and Faulkner, M. (2012). Preeclampsia 2012. *J. Pregnancy* 2012:586578. doi: 10.1155/2012/586578
- Esplin, M. S., Fausett, M. B., Fraser, A., Kerber, R., Mineau, G., Carrillo, J., et al. (2001). Paternal and maternal components of the predisposition to preeclampsia. N. Engl. J. Med. 344, 867–872. doi: 10.1056/NEJM20010 3223441201
- Feeney, J. G., and Scott, J. S. (1980). Pre-eclampsia and changed paternity. Eur. J. Obstet. Gynecol. Reprod. Biol. 11, 35–38. doi: 10.1016/0028-2243(80)90051-9

associated with the role of fathers in the development of preeclampsia.

### **AUTHOR CONTRIBUTIONS**

CG-H gathered and analyzed the data, and wrote the manuscript. MS-M gathered and organized the data and references. ET organized the data. MS-M, JG-O, and ET critically lectured the manuscript. BL-R gathered the data.

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- Feeney, J. G., Tovey, L. A., and Scott, J. S. (1977). Influence of previous blood-transfusion on incidence of pre-eclampsia. *Lancet* 1, 874–875. doi: 10.1016/S0140-6736(77)91199-0
- Galaviz-Hernandez, C., Arámbula-Meraz, E., Medina-Bastidas, D., Sosa-Macías, M., Lazalde-Ramos, B. P., Ortega-Chávez, M., et al. (2016). The paternal polymorphism rs5370 in the EDN1 gene decreases the risk of preeclampsia. *Pregnancy Hypertens*. 6, 327–332. doi: 10.1016/j.preghy.2016. 07.002
- Geller, S. E., Ahmed, S., Brown, M. L., Cox, S. M., Rosenberg, D., and Kilpatrick, S. J. (2004). International classification of diseases-9th revision coding for preeclampsia: how accurate is it? Am. J. Obstet. Gynecol. 190, 1629–1633. doi:10.1016/j.ajog.2004.03.061
- González-Comadran, M., Urresta Avila, J., Saavedra Tascón, A., Jimenéz, R., Solà, I., Brassesco, M., et al. (2014). The impact of donor insemination on the risk of preeclampsia: a systematic review and meta-analysis. Eur. J. Obstet. Gynecol. Reprod. Biol. 182, 160–166. doi: 10.1016/j.ejogrb.2014.09.022
- Hackmon, R., Pinnaduwage, L., Zhang, J., Lye, S. J., Geraghty, D. E., and Dunk, C. E. (2017). Definitive class I human leukocyte antigen expression in gestational placentation: HLA-F, HLA-E, HLA-C, and HLA-G in extravillous trophoblast invasion on placentation, pregnancy, and parturition. Am. J. Reprod. Immunol. 77:e12643. doi: 10.1111/aji.12643
- Hoy, J., Venn, A., Halliday, J., Kovacs, G., and Waalwyk, K. (1999). Perinatal and obstetric outcomes of donor insemination using cryopreserved semen in Victoria. Aust. Hum. Reprod. 14, 1760–1764. doi: 10.1093/humrep/14.7. 1760
- Hylenius, S., Andersen, A. M., Melbye, M., and Hviid, T. V. (2004). Association between HLA-G genotype and risk of pre-eclampsia: a case-control study using family triads. *Mol. Hum. Reprod.* 10, 237–246. doi: 10.1093/molehr/gah035
- Ikedife, D. (1980). Eclampsia in multipara. Br. Med. J. 280, 985–986. doi: 10.1136/bmj.280.6219.985-a
- Kenny, L. C., and Kell, D. B. (2018). Immunological tolerance, pregnancy, and preeclampsia: the roles of semen microbes and the father. *Front. Med.* 4:239. doi: 10.3389/fmed.2017.00239
- Kilpatrick, D. C., Liston, W. A., Jazwinska, E. C., and Smart, G. E. (1987). Histocompatibility studies in pre-eclampsia. *Tissue Antigens* 29, 232–236. doi: 10.1111/j.1399-0039.1987.tb01582.x
- Klonoff-Cohen, H. S., Savitz, D. A., Cefalo, R. C., and McCann, M. F. (1989). An epidemiologic study of contraception and preeclampsia. *JAMA* 262, 3143–3147. doi: 10.1001/jama.1989.03430220066032
- Koelman, C. A., Coumans, A. B., Nijman, H. W., Doxiadis, I. I., Dekker, G. A., and Claas, F. H. (2000). Correlation between oral sex and a low incidence of preeclampsia: a role for soluble HLA in seminal fluid? *J. Reprod. Immunol.* 46, 155–166. doi: 10.1016/S0165-0378(99)00062-5
- Kyrou, D., Kolibianakis, E. M., Devroey, P., and Fatemi, H. M. (2010). Is the use of donor sperm associated with a higher incidence of preeclampsia in women who achieve pregnancy after intrauterine insemination? *Fertil. Steril.* 93, 1124–1127. doi: 10.1016/j.fertnstert.2008.12.021

Li, D. K., and Wi, S. (2000). Changing paternity and the risk of preeclampsia/eclampsia in the subsequent pregnancy. Am. J. Epidemiol. 151, 57–62. doi: 10.1093/oxfordjournals.aje.a010122

- Lie, R. T. (2007). Intergenerational exchange and perinatal risks: a note on interpretation of generational recurrence risks. *Paediatr. Perinat. Epidemiol.* 21, 13–18. doi: 10.1111/j.1365-3016.2007.00832.x
- Lie, R. T., Rasmussen, S., Brunborg, H., Gjessing, H. K., Lie-Nielsen, E., and Irgens, L. M. (1998). Fetal and maternal contributions to risk of pre-eclampsia: population based study. BMJ 316, 1343–1347. doi: 10.1136/bmj.316.7141. 1343
- Luo, Z. C., Julien, P., Wei, S. Q., Audibert, F., Fraser, W. D., and Maternal and Infant Research on Oxidative Stress (MIROS) study group (2018). Association of. (pre)-eclampsia with SOD2 Ala16Val polymorphism among mother-fatherinfant triads. *Int. J. Gynaecol. Obstet.* 142, 221–227. doi: 10.1002/ijgo.12528
- Marti, J. J., and Herrmann, U. (1977). Immunogestosis: a new etiologic concept of "essential" EPH gestosis, with special consideration of the primigravid patient; preliminary report of a clinical study. Am. J. Obstet. Gynecol. 128, 489–493. doi: 10.1016/0002-9378(77)90030-8
- Mills, J. L., Klebanoff, M. A., Graubard, B. I., Carey, J. C., and Berendes, H. W. (1991). Barrier contraceptive methods and preeclampsia. *JAMA* 265, 70–73. doi: 10.1001/jama.1991.03460010070033
- Need, J. A. (1975). Pre-eclampsia in pregnancies by different fathers: immunological studies. Br. Med. J. 1, 548–549. doi: 10.1136/bmj.1.5957.548
- Need, J. A., Bell, B., Meffin, E., and Jones, W. R. (1983). Pre-eclampsia in pregnancies from donor inseminations. J. Reprod. Immunol. 5, 329–338. doi: 10.1016/0165-0378(83)90242-5
- Ness, R. B., Markovic, N., Harger, G., and Day, R. (2004). Barrier methods, length of preconception intercourse, and preeclampsia. *Hypertens. Pregnancy* 23, 227–235. doi: 10.1081/PRG-200030293
- Nevalainen, J., Ignatius, J., Savolainen, E. R., Ryynanen, M., and Jarvenpaa, J. (2018). Placenta-mediated pregnancy complications are not associated with fetal or paternal factor V Leiden mutation. Eur. J. Obstet. Gynecol. Reprod. Biol. 230, 32–35. doi: 10.1016/j.ejogrb.2018.09.016
- Nevo, O., Soleymanlou, N., Wu, Y., Xu, J., Kingdom, J., Many, A., et al. (2006). Increased expression of sFlt-1 in in vivo and in vitro models of human placental hypoxia is mediated by HIF-1. Am. J. Physiol. Regul. Integr. Comp. Physiol. 291, R1085–R1093. doi: 10.1152/ajpregu.00794.2005
- NHBPEP (2000). Report of the national high blood pressure education program working group on high blood pressure in pregnancy. *Am. J. Obstet. Gynecol.* 183, S1–S22. doi: 10.1067/mob.2000.107928
- Osungbade, K. O., and Ige, O. K. (2011). Public health perspectives of preeclampsia in developing countries: implication for health system strengthening. *J. Pregnancy* 2011:481095. doi: 10.1155/2011/481095
- Pabalan, N., Jarjanazi, H., Sun, C., and Iversen, A. C. (2015). Meta-analysis of the human leukocyte antigen-G (HLA-G) 14 bp insertion/deletion polymorphism as a risk factor for preeclampsia. *Tissue Antigens* 86, 186–194. doi: 10.1111/tan. 12627
- Pappa, K. I., Roubelakis, M., Vlachos, G., Marinopoulos, S., Zissou, A., Anagnou, N. P., et al. (2011). Variable effects of maternal and paternal-fetal contribution to the risk for preeclampsia combining GSTP1, eNOS, and LPL gene polymorphisms. J. Matern. Fetal Neonatal Med. 24, 628–635. doi: 10.3109/14767058.2010.511351
- Peralta Pedrero, M. L., Guzmán Ibarra, M. L., Basavilvazo Rodríguez, M. A., Sánchez Ambríz, S., Cruz Avelar, A., Lemus Rocha, R., et al. (2006). Elaboración y validación de un índice para el diagnóstico de preeclampsia. Ginecol. Obstet. Mex. 74, 205–214.
- Pipkin, F. B. (2001). Risk factors for preeclampsia. N. Engl. J. Med. 344, 925–926. doi: 10.1056/NEJM200103223441209
- Redman, C. W., and Sargent, I. L. (2010). Immunology of pre-eclampsia. *Am. J. Reprod. Immunol.* 63, 534–543. doi: 10.1111/j.1600-0897.2010.00831.x
- Reuvekamp, A., Velsing-Aarts, F. V., Poulina, I. E., Capello, J. J., and Duits, A. J. (1999). Selective deficit of angiogenic growth factors characterises pregnancies complicated by pre-eclampsia. *Br. J. Obstet. Gynaecol.* 106, 1019–1022. doi: 10.1111/j.1471-0528.1999.tb08107.x
- Roberts, J. M., and Hubel, C. A. (2009). The two stage model of pre-eclampsia: variations on the theme. *Placenta* 30(Suppl. A), S32–S37. doi: 10.1016/j. placenta.2008.11.009

Robertson, S. A., Bromfield, J. J., and Tremellen, K. P. (2003). Seminal 'priming' for protection from pre-eclampsia-a unifying hypothesis. *J. Reprod. Immunol.* 59, 253–265. doi: 10.1016/S0165-0378(03)00052-4

- Robillard, P. Y., and Hulsey, T. C. (1996). Association of pregnancyinduced-hypertension, pre-eclampsia, and eclampsia with duration of sexual cohabitation before conception. *Lancet* 347:619. doi: 10.1016/S0140-6736(96) 91315-X
- Robillard, P. Y., Hulsey, T. C., Alexander, G. R., Keenan, A., de Caunes, F., and Papiernik, E. (1993). Paternity patterns and risk of preeclampsia in the last pregnancy in multiparae. *J. Reprod. Immunol.* 24, 1–12. doi: 10.1016/0165-0378(93)90032-D
- Robillard, P. Y., Hulsey, T. C., Périanin, J., Janky, E., Miri, E. H., and Papiernik, E. (1994). Association of pregnancy-induced hypertension with duration of sexual cohabitation before conception. *Lancet* 344, 973–975. doi: 10.1016/S0140-6736(94)91638-1
- Saftlas, A. F., Rubenstein, L., Prater, K., Harland, K. K., Field, E., and Triche, E. W. (2014). Cumulative exposure to paternal seminal fluid prior to conception and subsequent risk of preeclampsia. *J. Reprod. Immunol.* 10, 104–110. doi: 10.1016/j.jri.2013.07.006
- Saito, S., Sakai, M., Sasaki, Y., Nakashima, A., and Shiozaki, A. (2007). Inadequate tolerance induction may induce preeclampsia. *J. Reprod. Immunol.* 76, 30–39. doi: 10.1016/j.jri.2007.08.002
- Salha, O., Sharma, V., Dada, T., Nugent, D., Rutherford, A. J., Tomlinson, A. J., et al. (1999). The influence of donated gametes on the incidence of hypertensive disorders of pregnancy. *Hum. Reprod.* 14, 2268–2273. doi: 10.1093/humrep/14. 9.2268
- Sandrim, V. C., Palei, A. C., Metzger, I. F., Gomes, V. A., Cavalli, R. C., and Tanus-Santos, J. E. (2008). Nitric oxide formation is inversely related to serum levels of antiangiogenic factors soluble fms-like tyrosine kinase-1 and soluble endogline in preeclampsia. *Hypertension* 52, 402–407. doi: 10.1161/ HYPERTENSIONAHA.108.115006
- Sharkey, A. M., Gardner, L., Hiby, S., Farrell, L., Apps, R., Masters, L., et al. (2008). Killer Ig-like receptor expression in uterine NK cells is biased toward recognition of HLA-C and alters with gestational age. *J. Immunol.* 181, 39–46. doi: 10.4049/jimmunol.181.1.39
- Skjaerven, R., Vatten, L. J., Wilcox, A. J., Rønning, T., Irgens, L. M., and Lie, R. T. (2005). Recurrence of pre-eclampsia across generations: exploring fetal and maternal genetic components in a population-based cohort. BMJ 331:877. doi: 10.1136/bmj.38555.462685.8F
- Smith, G. N., Walker, M., Tessier, J. L., and Millar, K. G. (1997). Increased incidence of preeclampsia in women conceiving by intrauterine insemination with donor versus partner sperm for treatment of primary infertility. *Am. J. Obstet. Gynecol.* 177, 455–458. doi: 10.1016/S0002-9378(97)70215-1
- Stone, P., Cook, D., Hutton, J., Purdie, G., Murray, H., and Harcourt, L. (1995). Measurements of blood pressure, oedema and proteinuria in a pregnant population of New Zealand. Aust. N. Z. J. Obstet. Gynaecol. 35, 32–37. doi: 10.1111/j.1479-828X.1995.tb01826.x
- Takimoto, E., Ishida, J., Sugiyama, F., Horiguchi, H., Murakami, K., and Fukamizu, A. (1996). Hypertension induced in pregnant mice by placental renin and maternal angiotensinogen. *Science* 274, 995–998. doi: 10.1126/ science.274.5289.995
- Tan, C. Y., Ho, J. F., Chong, Y. S., Loganath, A., Chan, Y. H., and Ravichandran, J. (2008). Paternal contribution of HLA-G\*0106 significantly increases risk for pre-eclampsia in multigravid pregnancies. *Mol. Hum. Reprod.* 14, 317–324. doi: 10.1093/molehr/gan013
- van Dijk, M., Drewlo, S., and Oudejans, C. B. (2010). Differential methylation of STOX1 in human placenta. *Epigenetics* 5, 736–742. doi: 10.4161/epi.5.8.13084
- van Dijk, M., and Oudejans, C. B. (2011). STOX1: key player in trophoblast dysfunction underlying early onset preeclampsia with growth retardation. *J. Pregnancy* 2011:521826. doi: 10.1155/2011/521826
- Vefring, H., Lie, R. T., ØDegård, R., Mansoor, M. A., and Nilsen, S. T. (2004). Maternal and fetal variants of genetic thrombophilias and the risk of preeclampsia. *Epidemiology* 15, 317–322. doi: 10.1097/01.ede.0000112217. 33111.23
- Wallis, A. B., Saftlas, A. F., Hsia, J., and Atrash, H. K. (2008). Secular trends in the rates of pre-eclampsia, eclampsia, and gestational hypertension, United States, 1987–2004. Am. J. Hypertens. 21, 521–526. doi: 10.1038/ajh.2008.20

Wikström, A. K., Gunnarsdóttir, J., and Cnattingius, S. (2012). The paternal role in pre-eclampsia and giving birth to a small for gestational age infant; a population-based cohort study. BMJ Open 2:e001178. doi: 10.1136/bmjopen-2012-001178

- Yoneda, N., Shiozaki, A., Miura, K., Yonezawa, R., Takemura, K., Yoneda, S., et al. (2013). A triploid partial mole placenta from paternal isodisomy with a diploid fetus derived from one sperm and one oocyte may have caused angiogenic imbalance leading to preeclampsia-like symptoms at 19 weeks of gestation. *Placenta* 34, 631–634. doi: 10.1016/j.placenta.2013. 03.015
- Zhang, Z., Li, Y., Zhang, L. L., Jia, L. T., and Yang, X. Q. (2012). Association of 14 bp insertion/deletion polymorphism of the HLA-G gene in father with severe preeclampsia in Chinese. *Tissue Antigens* 80, 158–164. doi: 10.1111/j.1399-0039. 2012.01907.x
- Zhou, A., Dekker, G. A., Lumbers, E. R., Lee, S. Y., Thompson, S. D., McCowan, L. M., et al. (2013). The association of AGTR2 polymorphisms with preeclampsia and uterine artery bilateral notching is modulated

- by maternal BMI. *Placenta* 34, 75–81. doi: 10.1016/j.placenta.2012.
- Zusterzeel, P. L., Morsche, R., Raijmakers, M. T., Roes, E. M., Peters, W. H., and Steegers, E. A. (2002). Paternal contribution to the risk for pre-eclampsia. J. Med. Genet. 39, 44–45. doi: 10.1136/jmg.39.1.44
- **Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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