



# Study of the Effects of 3 h of Continuous Cardiopulmonary Resuscitation at 27°C on Global Oxygen Transport and Organ Blood Flow

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Nilsen JH, Valkov S, Mohyuddin R, Schanche T, Kondratiev TV, Næsheim T, Sieck GC and Tveita T (2020) Study of the Effects of 3 h of Continuous Cardiopulmonary Resuscitation at 27°C on Global Oxygen Transport and Organ Blood Flow. Front. Physiol. 11:213. doi: 10.3389/fphys.2020.00213 **Aims:** Complete restitution of neurologic function after 6 h of pre-hospital resuscitation and in-hospital rewarming has been reported in accidental hypothermia patients with cardiac arrest (CA). However, the level of restitution of circulatory function during longlasting hypothermic cardiopulmonary resuscitation (CPR) remains largely unknown. We compared the effects of CPR in replacing spontaneous circulation during 3 h at 27°C vs. 45 min at normothermia by determining hemodynamics, global oxygen transport (DO<sub>2</sub>), oxygen uptake (VO<sub>2</sub>), and organ blood flow.

**Methods:** Anesthetized pigs (n = 7) were immersion cooled to CA at 27°C. Predetermined variables were compared: (1) Before cooling, during cooling to 27°C with spontaneous circulation, after CA and subsequent continuous CPR (n = 7), vs. (2) before CA and during 45 min CPR in normothermic pigs (n = 4).

**Results:** When compared to corresponding values during spontaneous circulation at 38°C: (1) After 15 min of CPR at 27°C, cardiac output (CO) was reduced by 74%, mean arterial pressure (MAP) by 63%, DO<sub>2</sub> by 47%, but organ blood flow was unaltered. Continuous CPR for 3 h maintained these variables largely unaltered except for significant reduction in blood flow to the heart and brain after 3 h, to the kidneys after 1 h, to the liver after 2 h, and to the stomach and small intestine after 3 h. (2) After normothermic CPR for 15 min, CO was reduced by 71%, MAP by 54%, and DO<sub>2</sub> by 63%. After 45 min, hemodynamic function had deteriorated significantly, organ blood flow was undetectable, serum lactate increased by a factor of 12, and mixed venous O<sub>2</sub> content was reduced to 18%.

**Conclusion:** The level to which CPR can replace CO and MAP during spontaneous circulation at normothermia was not affected by reduction in core temperature in our

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setting. Compared to spontaneous circulation at normothermia, 3 h of continuous resuscitation at 27°C provided limited but sufficient O<sub>2</sub> delivery to maintain aerobic metabolism. This fundamental new knowledge is important in that it encourages early and continuous CPR in accidental hypothermia victims during evacuation and transport.

Keywords: accidental hypothermia, cardiac arrest, hemodynamics, organ blood flow, cardiopulmonary resuscitation (CPR)

## INTRODUCTION

Over the past years, patient case reports of accidental hypothermia with cardiac arrest (CA) indicate favorable neurologic outcome after in-hospital rewarming (Walpoth et al., 1990, 1997; Gilbert et al., 2000; Mark et al., 2012; Wanscher et al., 2012; Hilmo et al., 2014). These favorable outcomes appear to be linked to the good quality of pre-hospital emergency medical treatment provided including early start and continued cardiopulmonary resuscitation (CPR) (Monsieurs et al., 2015; Truhlar et al., 2015) in accordance to the latest international guidelines (Truhlar et al., 2015).

Normothermic CPR can provide ~30% of pre-arrest cardiac output (CO) (Wik et al., 1996; Sunde et al., 1998; Pytte et al., 2006) but is usually considered discontinued after 20–30 min if the return of spontaneous circulation is not achieved, as these patients show poor clinical outcome (Torke et al., 2015; Perkins et al., 2015a,b). Preclinical studies (Wik et al., 1996; Gervais et al., 1997; Schwarz et al., 2002; Tomte et al., 2010; Welbourn and Efstathiou, 2018) also support this clinical notion, but the maximum duration of CPR in normothermic patients has so far not been determined (Welbourn and Efstathiou, 2018).

It is well documented that cooling with spontaneous circulation slows metabolic rate and, therefore, reduces CO in parallel with reductions in global O<sub>2</sub> transport (DO<sub>2</sub>) and O<sub>2</sub> consumption (VO<sub>2</sub>) (Black et al., 1976; Kondratiev et al., 2006; Valkov et al., 2019). In addition, low core temperature per se prolongs end-organ survival during hypothermic CPR (Gilbert et al., 2000; Mark et al., 2012; Wanscher et al., 2012). In an intact pig model, we validated these findings and also documented that O2 transport (Filseth et al., 2010) and tissue blood flow (Valkov et al., 2019) are maintained during 1-3 h of stable hypothermia (25°-27°C) as well as during rewarming. However, to date, the effects of prolonged (hours) CPR on global DO<sub>2</sub> and organ blood perfusion have not been assessed in an intact hypothermic animal model. A report from 1986 documented that CPR at 28°C replaced normothermic CO during spontaneous circulation by only 7% after 20 min (Maningas et al., 1986). Subsequent pharmacologic studies on pigs (Krismer et al., 2000; Kornberger et al., 2001) reported the maintenance of coronary perfusion pressure at 28°C during short-lasting CPR, but neither of these studied applied the latest standard CPR algorithm (Truhlar et al., 2015).

New devices make it possible to sustain CPR during evacuation and transport for a longer period of time with a limited number of rescuers (Perkins et al., 2015b). UNN (University Hospital of North Norway) is located in a scarcely populated, subarctic catchment area, making flying time for patients with ambulance airplanes typically around 90 min. This makes evacuation and transportation time 3–4 h for accidental hypothermia patients with CA in need of in-hospital rewarming. Under these circumstances, continuous prehospital CPR is often the only treatment option for these patients.

It is well documented that standard CPR can replace CO only to a certain low level. We, here, hypothesize that this level is largely unaffected by core temperature, and thus, cooling with spontaneous circulation will reduce CO to a level, which eventually can be replaced by CPR. Therefore, the aim of this study was to evaluate the value of CPR as part of prehospital interventions for hypothermic patients with CA in more detail using our intact porcine model of CPR instrumented to determine hemodynamics, global DO2, VO2, and organ blood flow during continuous CPR for 3 h at 27°C core temperature. To study the effects of low temperature only, ischemic episodes were avoided before CPR, and 27°C corresponds to the core temperature of surviving patients after hypothermic CA. To more closely mimic a clinical scenario, the porcine model was equipped with a commercially available automated chest compression device.

### MATERIALS AND METHODS

### **Ethical Approval**

The Norwegian Food Safety Authority approved the study (ref. number: 14/56323). Eleven castrated male pigs (wt. 20–29 kg, age 3 months) from NOROC stock were used. On arrival, the animals were acclimated for 2–5 days before the terminal experiment. Animals were fed twice daily, had free access to water at all times, and received humane care in accordance to the Norwegian Animal Welfare Act.

### Anesthesia and Instrumentation

We previously reported the detailed methods for hemodynamic monitoring, immersion cooling, and blood flow measurements using the porcine animal model (Valkov et al., 2019). Briefly, after fasting the animals overnight, premedication was induced by an intramuscular bolus of ketamine hydrochloride (20 mg kg<sup>-1</sup>), midazolam (30 mg), and atropine (1 mg), and anesthesia was induced by a bolus infusion of fentanyl (10  $\mu$ g kg<sup>-1</sup>) and pentobarbital-sodium (10 mg kg<sup>-1</sup>) in an ear vein. After

**Abbreviations:** ACD-CPR, active compression-decompression; CA, cardiac arrest; CPR; ITV-CPR, impedance threshold valve CPR; CPR, cardiopulmonary resuscitation; CVP, central venous pressure; DO<sub>2</sub>, global oxygen transport; ER O<sub>2</sub>, extraction ratio; HR, heart rate; MAP, mean arterial pressure; VO<sub>2</sub>, oxygen uptake; CO, cardiac output; VF, ventricular fibrillation.

tracheotomy and intubation, the animals were connected to a respirator (Siemens Servo 900D, Solna, Sweden), adjusted to maintain  $PaO_2 > 10$  kPa and  $PaCO_2$  at 4.5–6.0 kPa uncorrected for temperature ( $\alpha$ -stat management). After instrumentation, animals were randomly assigned to two groups [hypothermic CA and CPR for 3 h (n = 7), or normothermic CA and CPR for 45 min (n = 4)]. During ventricular fibrillation (VF) and CPR, FiO<sub>2</sub> was set to 1.0. Infusion of fentanyl (20  $\mu$ g kg<sup>-1</sup> h<sup>-1</sup>), midazolam  $(0.3 \ \mu g \ kg^{-1} \ h^{-1})$ , and pentobarbital-sodium (4 mg kg<sup>-1</sup> h<sup>-1</sup>) was continued via a femoral vein catheter. In the hypothermia group, anesthesia was discontinued at 27°C. Microspheres were injected to the left ventricle through a 6F fluid filled pigtail catheter (Cordis Corporation, Miami, FL, United States). Core temperature, CO, and venous and mixed venous blood gases were measured via a 7F Swan-Ganz thermodilution catheter (Edwards Lifesciences LLC, Irvine, CA, United States) positioned in the pulmonary trunk. Thermodilution is described as the gold standard for measuring CO during CPR (Carretero et al., 2010). The tip of another 7F Swan-Ganz thermodilution catheter was positioned in the aortic arch via the left femoral artery to monitor arterial blood gas and mean arterial pressure (MAP) and to collect reference blood samples for the microsphere technique. Urinary output was followed from a 14F urinary bladder catheter introduced via a lower abdominal incision. The animals were given 5,000 IE Heparin and allowed to stabilize for 45 min before the start of the experimental protocol.

### **Regional Blood Flow Measurements**

To determine organ blood flow, stable isotope-labeled microspheres (BioPAL Inc., Worcester, MA, United States) were injected into the left ventricle (Reinhardt et al., 2001). Simultaneously, a reference blood sample was drawn from the tip of a catheter inserted in the left femoral artery and advanced to the aortic arch to calculate regional blood flow. Blood flow was determined in tissue samples from the brain (temporal lobes), kidneys, liver, heart, small intestine, and stomach based on a technique of neutron activation to analyze microsphere content as already described in detail (Valkov et al., 2019).

# **Experimental Protocol**

Animals in the hypothermia group were immersion cooled in ice water to a blood temperature of 27°C, and VF was induced by stimulating the epicardial surface using an alternating current (5-20 mA, 6 Hz, and 30 V) conducted via a 15 cm-long needle electrode (Figure 1). The needle was inserted in the epigastric area and pointed toward the heart apex. Correct needle placement was confirmed when aspirating arterial blood from the left ventricle. CA was defined as asystoly or VF on ECG associated with the absence of fluctuation in arterial pressure. After 90 s of CA, an automated chest compression device (LUCAS chest compression system, Physio-Control Inc., Lund, Sweden) was started. The piston on this compression device was equipped with a suction cup to ensure active decompression with a continuous mode compression/decompression duty cycle of 50  $\pm$  5% at a rate of 100  $\pm$  5 compressions/min, and compression depth was 4-5 cm. In the normothermia group, CPR was continued



for 45 min, whereas in the hypothermia group, CPR was continued for 3 h.

### **Frequency of Measurements**

In the normothermia group: during spontaneous circulation at 38°C before VF, and at 15 and 45 min during CPR. In the hypothermia group: during spontaneous circulation at 38°C, during cooling at 32° and 27°C before VF, and during CPR at 15 min and hourly thereafter. Simultaneously, we recorded electrocardiogram (standard leads), heart rate (HR), MAP, central venous pressure (CVP), and urine output, and microspheres were injected. Blood gases were analyzed by simultaneous sampling from arterial, central venous, and mixed venous blood.

### **Statistics**

Statistical analyses were performed using SigmaPlot statistical software version 14 (Systat Software Inc., Richmond, CA, United States). Normal distribution was checked using the Shapiro-Wilk test. Comparisons between normothermia and hypothermia groups as well as intragroup comparisons at 38°C baseline, 15 min, and 45/60 min of CPR were performed by twoway repeated measures ANOVA. Where significant differences were found, all pairwise multiple comparison procedures were done using the Holm-Sidak test. To compare values within the hypothermia group, we used one-way repeated measures ANOVA for normal distributed variables, and Friedman repeated measures ANOVA on ranks for non-normal distributed variables. Where significant differences were found, Dunnett's test was used to compare all values within the hypothermia group vs. 38°C baseline, as well as values obtained during CPR at 60, 120, and 180 min vs. CPR at 15 min. The level of significance was set at p < 0.05. Data are presented as means and SD.

# RESULTS

No statistically significant differences were found between groups in any of the variables recorded at the start of the experiments.



**FIGURE 2** Hemodynamic variables, and global O<sub>2</sub> transport and O<sub>2</sub> uptake during CPR at normothermia and hypothermia. (A) Cardiac output (CO), (B) mean arterial pressure (MAP), (C) global oxygen delivery (DO<sub>2</sub>), and (D) global oxygen consumption (VO<sub>2</sub>). CPR at normothermia (NT 38°C, n = 4), CPR at hypothermia (HT 27°C, n = 7). 38°C baseline (BL) (38°C); normal sinus rhythm (NSR); cardiopulmonary resuscitation (CPR); ventricular fibrillation (VF). Comparisons between groups and intragroup comparisons at 38°C baseline, 15 and 45/60 min CPR were done using two-way repeated measures ANOVA and pairwise multiple comparison using Holm–Sidak test. Values are mean ± SD. \*p < 0.05 vs. intragroup 38°C baseline; #p < 0.05 vs. intragroup 15 min of CPR; § p < 0.05 vs. corresponding value between groups.

Animals in both groups had spontaneous circulation before induction of VF. Stability of the actual pig model related to hemodynamic function and organ blood flow at normothermia has previously been documented in experiments lasting up to 7 h (Filseth et al., 2010; Valkov et al., 2019). Multiple costal fractures occurred in all animals after CPR, and after 3 h of CPR also sternal fractures were observed in six out of seven animals.

# Comparing Effects of CPR for VF After 15 and 45/60 min, at the Two Different Core Temperatures, 38° and 27°C Hemodynamic Variables

### 38° C.

Compared to spontaneous circulation at 38°C, after CPR for 15 min at this temperature, CO was significantly reduced from 3.0  $\pm$  0.6 to 0.9  $\pm$  0.2 L min<sup>-1</sup> (-71%) and MAP was significantly reduced from 85  $\pm$  5 to 38  $\pm$  12 mmHg (-55%)

(Figures 2A,B). After 45 min of CPR at 38°C, CO and MAP ended up being reduced by 80 and 78% (0.6  $\pm$  0 L min<sup>-1</sup> and 18  $\pm$  6 mmHg), respectively, but the reduction in MAP was statistically significant.

### 27°C.

Compared to spontaneous circulation at 38°C, after CPR for 15 min at 27°C, CO was significantly reduced from  $3.2 \pm 0.6$  to  $0.8 \pm 0.1$  L min<sup>-1</sup> (-74%) and MAP from 89 ± 14 to  $33 \pm 9$  mmHg (-63%). After 60 min of CPR at 27°C, CO and MAP were reduced by 74 and 65% ( $0.8 \pm 0.2$  L min<sup>-1</sup> and  $31 \pm 8$  mmHg), respectively.

### Comparisons.

No significant differences between groups in CO or MAP were found after 15 min of CPR. In the 38°C group, MAP after 45 min CPR was significantly lower than MAP after 60 min of CPR in the



**FIGURE 3** Oxygen extraction ratio, and regional blood flow during CPR at normothermia and hypothermia. (A) Oxygen extraction ratio ( $O_2$  ER), (**B–D**) regional blood flow in the heart, brain, and kidneys. CPR at normothermia (NT 38°C, n = 4), CPR at hypothermia (HT 27°C, n = 7). 38°C baseline (BL) (38°C); normal sinus rhythm (NSR); cardiopulmonary resuscitation (CPR); ventricular fibrillation (VF). Comparisons between groups and intragroup comparisons at 38°C baseline, 15 and 45/60 min of CPR were done using two-way repeated measures ANOVA and pairwise multiple comparison using the Holm–Sidak test. Values are mean  $\pm$  SD. \*p < 0.05 vs. intragroup 38°C baseline; #p < 0.05 vs. intragroup 15 min of CPR;  $^{\circ}p < 0.05$  vs. corresponding value between groups.

27°C group. This is in essential contrast to the 27°C group where both MAP and CO remained unchanged during 60 min of CPR.

# Oxygen Transport and Extraction

38°*C*.

Compared to during spontaneous circulation at 38°C, after 15 min of CPR, DO<sub>2</sub> decreased significantly from 13.6  $\pm$  2.2 to 4.8  $\pm$  1.9 ml min<sup>-1</sup> kg<sup>-1</sup> (-64%), and VO<sub>2</sub> from 5.8  $\pm$  0.5 to 4.1  $\pm$  1.8 ml min<sup>-1</sup> kg<sup>-1</sup> (-30%) (**Figures 2C,D**). Accordingly, the O<sub>2</sub> extraction ratio (ER; DO<sub>2</sub>/VO<sub>2</sub>) (**Figure 3A**) was significantly increased after 15 min of CPR (from 0.43  $\pm$  0.03 to 0.84  $\pm$  0.04), exceeding the reported critical extraction ratio (ER<sub>crit</sub>) value (0.6–0.7) to provide aerobic metabolism (Leach and Treacher, 1992). After 45 min, ER remained significantly elevated (0.84  $\pm$  0.04). Simultaneously, a reduction in mixed venous O<sub>2</sub> saturation (SvO<sub>2</sub>) (**Figure 4B**) from 56  $\pm$  3% during spontaneous

circulation to 15 ± 3 and 18 ± 6% after 15 and 45 min of CPR, respectively, took place in parallel with a significant increase in the serum lactate level (**Figure 4A**), from 0.63 ± 0.15 during spontaneous circulation to 3.78 ± 0.87 after 15 min of CPR increasing to 7.38 ± 1.51 mmol L<sup>-1</sup> after 45 min of CPR, underlining the existence of inadequate O<sub>2</sub> transport throughout the last 30 min of CPR at 38°C.

### 27°C.

Compared to during spontaneous circulation at 38°C, after 15 min of CPR at 27°C, DO<sub>2</sub> decreased significantly from 13.7  $\pm$  1.7 to 3.6  $\pm$  0.8 ml min<sup>-1</sup> kg<sup>-1</sup> (-73%), and VO<sub>2</sub> from 5.7  $\pm$  0.7 to 2.0  $\pm$  0.7 ml min<sup>-1</sup> kg<sup>-1</sup> (-64%) (**Figures 2A,B**). This caused a significant increase in ER from 0.42  $\pm$  0.07 to 0.55  $\pm$  0.12%, but this value is below ER<sub>crit</sub> (0.6–0.7) (**Figure 3A**). Simultaneously, SvO<sub>2</sub> (**Figure 4B**) fell from 58  $\pm$  6 at 38°C to



**FIGURE 4** | Lactate and mixed venous oxygen saturation during CPR at normothermia and hypothermia. (A) Lactate and (B) mixed venous oxygen saturation (SvO<sub>2</sub>). CPR at normothermia (NT 38°C, n = 4), CPR at hypothermia (HT 27°C, n = 7). 38°C baseline (BL) (38°C); normal sinus rhythm (NSR); cardiopulmonary resuscitation (CPR); ventricular fibrillation (VF). Comparisons between groups and intragroup comparisons at 38°C baseline, 15 and 45/60 min CPR were done using two way repeated measures ANOVA and pairwise multiple comparison using the Holm–Sidak test. Values are mean  $\pm$  SD. \*p < 0.05 vs. intragroup 38°C baseline; #p < 0.05 vs. intragroup 15 min of CPR; §p < 0.05 vs. corresponding value between groups.

46 ± 14 after 15 min of CPR, and after 60 min of CPR, SvO<sub>2</sub> was reduced to 27 ± 11%, and lactate (**Figure 4A**) increased after 15 min and 60 min of CPR from  $1.17 \pm 0.65$  to  $2.81 \pm 0.98$  mmol  $L^{-1}$ , respectively.

### Differences

Owing to the low core temperature, VO<sub>2</sub> in the 27°C group was significantly lower than VO<sub>2</sub> in the 38°C group, whereas no differences between groups in DO<sub>2</sub> were found. The reduction in SvO<sub>2</sub> was significantly lower in the 27°C group than in the 38°C group. Also, the increase in lactate in the 27°C group was significantly lower than in the 38°C. Taken together, the higher the SvO<sub>2</sub>, the modest increase in lactate, and the less increase in ER during 60 min of CPR in the 27°C group.

### **Organ Blood Flow**

Already after 15 min of CPR in the 38°C group, there was a statistically significant reduction in blood flow to the brain, heart, kidneys, stomach, liver, and small intestine (**Figures 3B,D**). After 45 min of CPR, practically no organ blood flow was detectable. A similar reduction in organ blood flow was measured after 15 min of CPR in the 27°C group, but in essential contrast to the 38°C group, organ blood flow was maintained at this reduced level throughout 60 min of CPR.

# Cooling to 27°C With Spontaneous Circulation

### Hemodynamic Variables

Compared to corresponding values at 38°C, cooling with maintained spontaneous circulation gave an almost linear reduction in CO and MAP (**Figures 5A,B**). At 27°C, CO was significantly reduced from  $3.2 \pm 0.6$  to  $1.6 \pm 0.4$  L min<sup>-1</sup>

(-50%), and MAP significantly reduced from 89  $\pm$  14 to 60  $\pm$  14 mmHg (-33%).

### **Oxygen Transport and Extraction**

Global DO<sub>2</sub> and VO<sub>2</sub> (**Figure 5C**) were significantly reduced from 13.7  $\pm$  1.7 to 7.1  $\pm$  1.8 ml min<sup>-1</sup> kg<sup>-1</sup> (-48%), and from 5.7  $\pm$  0.7 to 1.8  $\pm$  0.2 ml min<sup>-1</sup> kg<sup>-1</sup> (-68%), respectively, giving an ER (**Figure 5D**) of 0.26, which is far below the ER<sub>crit</sub> of 0.6–0.7 (Leach and Treacher, 1992).

### **Organ Blood Flow**

During cooling to  $27^{\circ}$ C, an apparent reduction in blood flow to all organs measured took place, but these changes did not reach statistical significance (**Figure 6**).

# Effects of Continuous CPR for VF During 3 h at 27°C

# Hemodynamic Variables

CPR for 15 min at 27°C gave a significant reduction in CO from  $3.2 \pm 0.6$  to  $0.8 \pm 0.1$  L min<sup>-1</sup> (-74%), and a significant reduction in MAP from  $89 \pm 14$  to  $33 \pm 9$  mmHg (-63%) when compared to corresponding values during spontaneous circulation at 38°C (**Figures 5A,B**). Both CO and MAP remained unchanged at these reduced levels throughout the 3 h of CPR at 27°C.

### **Oxygen Transport and Extraction**

During 3 h of CPR, the differences between  $DO_2$  and  $VO_2$  diminished (**Figure 5C**), but when compared to their corresponding values after 15 min of CPR at  $27^{\circ}C$  (7.1  $\pm$  1.8 and  $1.8 \pm 0.2$  ml min<sup>-1</sup> kg<sup>-1</sup>, respectively),  $DO_2$  and  $VO_2$  were further significantly reduced first after 2 h of CPR ( $2.8 \pm 0.7$  and  $1.9 \pm 0.5$  ml min<sup>-1</sup> kg<sup>-1</sup>, respectively). By 1 h of CPR,  $ER_{crit}$  (**Figure 5D**) was approached (0.71  $\pm$  0.08) and remained at or



Normal sinus rhythm (NSR); cardiopulmonary resuscitation (CPR); ventricular fibrillation (VF). To compare values within groups, we used one-way repeated measures ANOVA for normal distributed variables. Dunnett's test was used to compare all values obtained vs. 38°C baseline and also for values obtained during CPR vs. corresponding values after CPR for 15 min. Values are mean  $\pm$  SD. \*p < 0.05 vs. intragroup 38°C baseline; †p < 0.05 vs. intragroup 15 min of CPR at 27°C.

near this critical value over the next 2 h of CPR (0.68  $\pm$  0.05). After 15 min of CPR, SvO<sub>2</sub> (**Figure 7**) was reduced from 75  $\pm$  7 to 46  $\pm$  14% followed by a further gradual reduction in SvO<sub>2</sub> over the next 1 h to 27  $\pm$  11%, but with no change during the next 2 h. After 1 h of CPR at 27°C, a modest but significant elevation of serum lactate from 0.8  $\pm$  0.65 to 2.81  $\pm$  0.98 mmol L<sup>-1</sup> took place. Serum lactate level increased linearly during the rest of the CPR period reaching 5.56  $\pm$  1.84 mmol L<sup>-1</sup>after 3 h.

### **Organ Blood Flow**

Compared to after 15 min of CPR at  $27^{\circ}$ C, blood flow to the heart remained unaltered during the first 2 h of CPR (**Figure 6A**). A significant reduction in blood flow to the heart from  $0.3 \pm 0.2$  to  $0.1 \pm 0.2$  ml min<sup>-1</sup> g<sup>-1</sup> was found first after 3 h of CPR. Blood flow to both brain hemispheres (**Figure 6B**) remained unaltered after 1 h of CPR when compared to flow after 15 min of CPR, but

flow to both left hemispheres was significantly reduced after 2 h of CPR. First, after 3 h of CPR blood flow to both hemispheres was significantly reduced compared to 15 min of CPR: left hemisphere from  $0.16 \pm 0.16$  to  $0.02 \pm 0.02$  and right hemisphere from  $0.17 \pm 0.15$  to  $0.03 \pm 0.03$ , all values in ml min<sup>-1</sup>g<sup>-1</sup>. Blood flow to kidneys (**Figure 6C**) was significantly reduced already after 1 h of CPR, liver blood flow was significantly reduced after 3 h, whereas flow to the small intestine and stomach was reduced after CPR for 2 and 3 h (**Figure 6D**).

# DISCUSSION

This experiment, comparing hemodynamic function,  $O_2$  transport, and organ blood flow during spontaneous circulation at 38°C, and during subsequent CPR for VF at 38°C or 27°C, demonstrated that after 15 min of CPR, these variables were



**FIGURE 6** [Regional blood flow during cooling and 3 h CPR at 27°C (n = 7). (A–D) Regional blood flow. Normal sinus rhythm (NSR); cardiopulmonary resuscitation (CPR); ventricular fibrillation (VF). To compare values within groups, we used one-way repeated measures ANOVA for normal distributed variables, and Friedman repeated measures ANOVA on ranks for non-normal distributed variables. Dunnett's test was used to compare all values obtained vs. 38°C baseline and also for values obtained during CPR vs. corresponding values after CPR for 15 min. Values are mean  $\pm$  SD. \*p < 0.05 vs. intragroup 38°C baseline; #, †p < 0.05 vs. intragroup 15 min of CPR at 27°C.

generated at the same reduced levels irrespective of the core temperature. This corresponds with our working hypothesis. Continued CPR for a total of 3 h at  $27^{\circ}$ C maintained MAP and CO, blood flow to the brain and heart at the same reduced level, and provided adequate DO<sub>2</sub> to support aerobic metabolism. The results also indicate the presence of a compensatory autoregulation of organ blood flow during hypothermic CPR with critically reduced circulation. By contrast, after 45 min of CPR at 38°C, MAP was significantly reduced, and blood flow to vital organs became undetectable in parallel with derangements in organ metabolism.

# **Cardiopulmonary Resuscitation**

The European Resuscitation Council guidelines (2015) (Truhlar et al., 2015) recommend that in accidental hypothermia patients with CA, CPR should be started immediately and continued during evacuation and transport. These guidelines also advocate using mechanical chest compression devices in special scenarios, e.g., during rescue of hypothermic patients in a moving ambulance, even though it is still debated if compression devices are superior to manual compression for CA (Rubertsson and Karlsten, 2005; Aufderheide et al., 2011; Putzer et al., 2013).

Several studies using the pig as experimental model have explored the effects of different techniques for CPR on pressure generation and organ blood flow during normothermia, with and without the aid of compression devices (Wik et al., 1996; Sunde et al., 1997, 1998; Langhelle et al., 2002; Schwarz et al., 2002; Pytte et al., 2006; Tomte et al., 2010; Karlis et al., 2014). Alternative techniques most often tested are active compression– decompression CPR (ACD-CPR) with and without use of an impedance threshold valve CPR (ITV-CPR) in comparison to standard CPR (Langhelle et al., 2002; Schwarz et al., 2002). These studies consistently reported better pressure end flow generation during ACD-CPR, attributed the elevated negative intrathoracic



pressure during active decompression facilitating better filling of the heart between compressions. Mechanical chest compression devices most often offer ACD-CPR, as used in the present experiment, but guidelines do not consider ACD-CPR superior to good standard CPR (Wenzel et al., 2010).

From clinical practice and preclinical experiments, we know that the effectiveness of normothermic CPR to maintain blood flow rapidly deteriorates. In the present experiment, hemodynamics and organ blood flow were largely maintained during the first 2 h of hypothermic CPR. After that, a modest reduction in these variables was accompanied with a small increase in lactate production, but far from ending in a circulatory collapse as demonstrated in normothermic animals after 45 min of CPR.

# **Hemodynamics and Blood Flow**

In pig models of normothermic CPR, hemodynamic and organ blood flow data are well documented after 5–10 min of CPR (Sunde et al., 1997, 1998; Langhelle et al., 2002; Steen et al., 2002). The present study evaluated the effects of CPR after 15 min, and the actual hemodynamic results are comparable to those reported by Carretero et al. (2010) after 15 min and by Steen et al. (2002) after only 5 min of CPR. Similar to our results, Gervais et al. (1997) reported almost complete cessation of myocardial and cerebral blood flow after 45 min of normothermic CPR.

# **Oxygen Transport**

With compromised circulation,  $VO_2$  will eventually become dependent on the  $DO_2$ . The  $O_2$  extraction ratio (ER), the ratio between  $VO_2$  and  $DO_2$ , can be used to demonstrate this physiologic mechanism. With limited O<sub>2</sub> supply, ER approaches a critical value, ER<sub>crit</sub>, where tissue O<sub>2</sub> consumption is ruled by O2 supply (Schumacker et al., 1987), reported to be 0.6-0.7 (Leach and Treacher, 1992) at normothermia. During hypothermia, the ERcrit is unchanged (~0.65) (Schumacker et al., 1987), even with an increase in vascular resistance, which may override the physiologic local metabolic feedback to arterioles to regulate blood flow (Schumacker et al., 1987). During spontaneous circulation at 27°C, the reduced metabolism caused a significantly reduced  $VO_2$  in parallel with the reduced  $DO_2$ . Importantly, we found that during the first 2 h of CPR at 27°C, despite the 74% reduction in CO, DO2 was maintained at the same low level as during spontaneous circulation. Beyond 2 h of CPR, a significant fall in DO<sub>2</sub> took place, followed by a moderate, yet significant, increase in lactate production and a fall in SvO<sub>2</sub>. For comparison, also in our previous pig experiment mentioned with 3 h of spontaneous circulation at 27°C, we found an elevation of ER to a level approaching ERcrit indicating a marginal O<sub>2</sub> supply during hypothermia, but this marginal supply was followed by a successful rewarming (Valkov et al., 2019).

Taken together, the actual values for lactate production,  $\text{SvO}_2$ , and ER demonstrate that 3 h of CPR at 27°C provides marginal, but sufficient, O<sub>2</sub> transport for aerobic metabolism in vital organs. Importantly, we also found that VO<sub>2</sub> was maintained during 3 h of CPR, indicating a use-dependent O<sub>2</sub> consumption. This is in essential contrast to normothermic CPR where DO<sub>2</sub> after 15 min was unable to provide aerobic metabolism, reflected by the significant elevation of ER exceeding the ER<sub>crit</sub> value, in parallel with a substantial elevation in serum lactate and lowering of SvO<sub>2</sub>.

# LIMITATIONS

The porcine model was chosen in our experiment because of its previous use in exploring the effects of CPR, especially when using a compression device (Steen et al., 2002; Rubertsson and Karlsten, 2005). However, the incidence of fractures of the sternum and multiple costa after 3 h of CPR in all our animals clearly demonstrates a major limitation when applying equipment designed for humans. A multicenter survey has documented the absence of fatal injuries in patients after conventional CPR using a compression device (Smekal et al., 2014), but little is known about damage to the thoracic skeleton or to the heart of patients after prolonged mechanical chest compressions.

# **Potential Translational Value**

From a clinical perspective, to date, the only possible way to treat patients with continuous CPR during evacuation and transport in an ambulance car is by using an automated chest compression device. Therefore, to treat accidental hypothermia patients with cardiac arrest in our catchment area, we aim to equip all car ambulances with automated compression devices. Further, the beneficial physiologic effects of prolonged CPR for cardiac arrest at reduced core temperatures, emerges to build a new basis with unknown potential for patient survival after the ensuing in-hospital rewarming. From clinical reports already listed, extracorporal circulation has evolved to become the method of choice for rewarming these patients. Preferentially, extra corporal membrane oxygenation (ECMO) is increasingly being used. By applying ECMO for rewarming, cardio/respiratory support can be continued during an ensuing ICU stay for days, if needed. This is to treat rewarming injury, which often manifests itself as cardiac dysfunction caused by hypothermia/rewarming, in addition to the unknown consequences of prolonged CPR to cause physical damage to the heart, lungs, and the thoracic skeleton. However, although effects of extracorporal rewarming to increase survival seems promising, more experimental and clinical work is needed to customize this method for rewarming patients from accidental hypothermia.

# REFERENCES

- Aufderheide, T. P., Frascone, R. J., Wayne, M. A., Mahoney, B. D., Swor, R. A., Domeier, R. M., et al. (2011). Standard cardiopulmonary resuscitation versus active compression-decompression cardiopulmonary resuscitation with augmentation of negative intrathoracic pressure for out-of-hospital cardiac arrest: a randomised trial. *Lancet* 377, 301–311. doi: 10.1016/S0140-6736(10) 62103-4
- Black, P. R., van, D. S., and Cohn, L. H. (1976). Effects of hypothermia on systemic and organ system metabolism and function. J. Surg. Res. 20, 49–63.
- Carretero, M. J., Fontanals, J., Agusti, M., Arguis, M. J., Martinez-Ocon, J., Ruiz, A., et al. (2010). Monitoring in resuscitation: comparison of cardiac output measurement between pulmonary artery catheter and NICO. *Resuscitation* 81, 404–409. doi: 10.1016/j.resuscitation.2009. 12.021

# CONCLUSION

The results of this study support our working hypothesis and show the favorable effects of CPR when given at  $27^{\circ}$ C in comparison to normothermic CPR. The beneficial effects of CPR in providing a limited, but adequate, DO<sub>2</sub> during a 3 h period at  $27^{\circ}$ C, combined with an apparent patent peripheral circulatory function, make continuous CPR the most valuable prehospital intervention as a bridge for hypothermic patients to receive in-hospital rewarming.

# DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation, to any qualified researcher.

# **AUTHOR CONTRIBUTIONS**

JN, TT, GS, and TK contributed to the conception and design. JN, SV, RM, TS, TN, and TK contributed to the completion of experiments and collection of data. TT, GS, TK, JN, SV, RM, and TS contributed to the data analysis and interpretation. JN, TT, and GS contributed to the drafting the manuscript for intellectual content. JN, SV, RM, TS, TK, TN, GS, and TT contributed to the revision of the manuscript.

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- Filseth, O. M., How, O. J., Kondratiev, T., Gamst, T. M., and Tveita, T. (2010). Posthypothermic cardiac left ventricular systolic dysfunction after rewarming in an intact pig model. *Crit. Care* 14, R211. doi: 10.1186/cc9334
- Gervais, H. W., Eberle, B., Hennes, H. J., Grimm, W., Kilian, A., Konietzke, D., et al. (1997). High dose naloxone does not improve cerebral or myocardial blood flow during cardiopulmonary resuscitation in pigs. *Resuscitation* 34, 255–261.
- Gilbert, M., Busund, R., Skagseth, A., Nilsen, P., and Solbø, J. (2000). Resuscitation from accidental hypothermia of 13.7 degrees C with circulatory arrest. *Lancet* 355, 375–376.
- Hilmo, J., Naesheim, T., and Gilbert, M. (2014). "Nobody is dead until warm and dead": prolonged resuscitation is warranted in arrested hypothermic victims also in remote areas–a retrospective study from northern Norway. *Resuscitation* 85, 1204–1211. doi: 10.1016/j.resuscitation.2014.04.029
- Karlis, G., Iacovidou, N., Lelovas, P., Niforopoulou, P., Zacharioudaki, A., Papalois, A., et al. (2014). Effects of early amiodarone administration during and

immediately after cardiopulmonary resuscitation in a swine model. Acta Anaesthesiol. Scand. 58, 114–122. doi: 10.1111/aas.12226

- Kondratiev, T. V., Flemming, K., Myhre, E. S., Sovershaev, M. A., and Tveita, T. (2006). Is oxygen supply a limiting factor for survival during rewarming from profound hypothermia? *Am. J. Physiol. Heart Circ. Physiol.* 291, H441–H450. doi: 10.1152/ajpheart.01229.2005
- Kornberger, E., Lindner, K. H., Mayr, V. D., Schwarz, B., Rackwitz, K. S., Wenzel, V., et al. (2001). Effects of epinephrine in a pig model of hypothermic cardiac arrest and closed-chest cardiopulmonary resuscitation combined with active rewarming. *Resuscitation* 50, 301–308.
- Krismer, A. C., Lindner, K. H., Kornberger, R., Wenzel, V., Mueller, G., Hund, W., et al. (2000). Cardiopulmonary resuscitation during severe hypothermia in pigs: does epinephrine or vasopressin increase coronary perfusion pressure? *Anesth. Analg.* 90, 69–73.
- Langhelle, A., Stromme, T., Sunde, K., Wik, L., Nicolaysen, G., and Steen, P. A. (2002). Inspiratory impedance threshold valve during CPR. *Resuscitation* 52, 39–48.
- Leach, R. M., and Treacher, D. F. (1992). The pulmonary physician and critical care.
  Oxygen transport: the relation between oxygen delivery and consumption. *Thorax* 47, 971–978.
- Maningas, P. A., DeGuzman, L. R., Hollenbach, S. J., Volk, K. A., and Bellamy, R. F. (1986). Regional blood flow during hypothermic arrest. *Ann. Emerg. Med.* 15, 390–396.
- Mark, E., Jacobsen, O., Kjerstad, A., Naesheim, T., Busund, R., Bahar, R., et al. (2012). Hypothermic cardiac arrest far away from the center providing rewarming with extracorporeal circulation. *Int. J. Emerg. Med.* 5:7. doi: 10.1186/ 1865-1380-5-7
- Monsieurs, K. G., Nolan, J. P., Bossaert, L. L., Greif, R., Maconochie, I. K., Nikolaou, N. I., et al. (2015). European resuscitation council guidelines for resuscitation 2015: section 1. executive summary. *Resuscitation* 95, 1–80. doi: 10.1016/j. resuscitation.2015.07.038
- Perkins, G. D., Handley, A. J., Koster, R. W., Castren, M., Smyth, M. A., Olasveengen, T., et al. (2015a). European resuscitation council guidelines for resuscitation 2015: section 2. Adult basic life support and automated external defibrillation. *Resuscitation* 95, 81–99. doi: 10.1016/j.resuscitation.2015. 07.015
- Perkins, G. D., Lall, R., Quinn, T., Deakin, C. D., Cooke, M. W., Horton, J., et al. (2015b). Mechanical versus manual chest compression for out-of-hospital cardiac arrest (PARAMEDIC): a pragmatic, cluster randomised controlled trial. *Lancet* 385, 947–955. doi: 10.1016/S0140-6736(14)61886-9
- Putzer, G., Braun, P., Zimmermann, A., Pedross, F., Strapazzon, G., Brugger, H., et al. (2013). LUCAS compared to manual cardiopulmonary resuscitation is more effective during helicopter rescue-a prospective, randomized, cross-over manikin study. Am. J. Emerg. Med. 31, 384–389. doi: 10.1016/j.ajem.2012.07. 018
- Pytte, M., Kramer-Johansen, J., Eilevstjonn, J., Eriksen, M., Stromme, T. A., Godang, K., et al. (2006). Haemodynamic effects of adrenaline (epinephrine) depend on chest compression quality during cardiopulmonary resuscitation in pigs. *Resuscitation* 71, 369–378. doi: 10.1016/j.resuscitation.2006.05.003
- Reinhardt, C. P., Dalhberg, S., Tries, M. A., Marcel, R., and Leppo, J. A. (2001). Stable labeled microspheres to measure perfusion: validation of a neutron activation assay technique. *Am. J. Physiol. Heart Circ. Physiol.* 280, H108–H116. doi: 10.1152/ajpheart.2001.280.1.H108
- Rubertsson, S., and Karlsten, R. (2005). Increased cortical cerebral blood flow with LUCAS; a new device for mechanical chest compressions compared to standard external compressions during experimental cardiopulmonary resuscitation. *Resuscitation* 65, 357–363. doi: 10.1016/j.resuscitation.2004. 12.006
- Schumacker, P. T., Rowland, J., Saltz, S., Nelson, D. P., and Wood, L. D. (1987). Effects of hyperthermia and hypothermia on oxygen extraction by tissues during hypovolemia. J. Appl. Physiol. 63, 1246–1252. doi: 10.1152/jappl.1987. 63.3.1246
- Schwarz, B., Mair, P., Raedler, C., Deckert, D., Wenzel, V., and Lindner, K. H. (2002). Vasopressin improves survival in a pig model of hypothermic cardiopulmonary resuscitation. *Crit. Care Med.* 30, 1311–1314.

- Smekal, D., Lindgren, E., Sandler, H., Johansson, J., and Rubertsson, S. (2014). CPR-related injuries after manual or mechanical chest compressions with the LUCAS device: a multicentre study of victims after unsuccessful resuscitation. *Resuscitation* 85, 1708–1712. doi: 10.1016/j.resuscitation.2014. 09.017
- Steen, S., Liao, Q., Pierre, L., Paskevicius, A., and Sjoberg, T. (2002). Evaluation of LUCAS, a new device for automatic mechanical compression and active decompression resuscitation. *Resuscitation* 55, 285–299.
- Sunde, K., Wik, L., Naess, P. A., Grund, F., Nicolaysen, G., and Steen, P. A. (1998). Improved haemodynamics with increased compression-decompression rates during ACD-CPR in pigs. *Resuscitation* 39, 197–205.
- Sunde, K., Wik, L., and Steen, P. A. (1997). Quality of mechanical, manual standard and active compression-decompression CPR on the arrest site and during transport in a manikin model. *Resuscitation* 34, 235–242.
- Tomte, O., Sjaastad, I., Wik, L., Kuzovlev, A., Eriksen, M., Norseng, P. A., et al. (2010). Discriminating the effect of accelerated compression from accelerated decompression during high-impulse CPR in a porcine model of cardiac arrest. *Resuscitation* 81, 488–492. doi: 10.1016/j.resuscitation.2009.12.028
- Torke, A. M., Bledsoe, P., Wocial, L. D., Bosslet, G. T., and Helft, P. R. (2015). CEASE: a guide for clinicians on how to stop resuscitation efforts. Ann. Am. Thorac. Soc. 12, 440–445. doi: 10.1513/AnnalsATS.201412-552PS
- Truhlar, A., Deakin, C. D., Soar, J., Khalifa, G. E., Alfonzo, A., Bierens, J. J., et al. (2015). European resuscitation council guidelines for resuscitation 2015: section 4. Cardiac arrest in special circumstances. *Resuscitation* 95, 148–201. doi: 10.1016/j.resuscitation.2015.07.017
- Valkov, S., Mohyuddin, R., Nilsen, J. H., Schanche, T., Kondratiev, T. V., Sieck, G. C., et al. (2019). Organ blood flow and O2 transport during hypothermia (27 degrees C) and rewarming in a pig model. *Exp. Physiol.* 104, 50–60. doi: 10.1113/EP087205
- Walpoth, B. H., Locher, T., Leupi, F., Schupbach, P., Muhlemann, W., and Althaus, U. (1990). Accidental deep hypothermia with cardiopulmonary arrest: extracorporeal blood rewarming in 11 patients. *Eur. J. Cardiothorac. Surg.* 4, 390–393.
- Walpoth, B. H., Walpoth-Aslan, B. N., Mattle, H. P., Radanov, B. P., Schroth, G., Schaeffler, L., et al. (1997). Outcome of survivors of accidental deep hypothermia and circulatory arrest treated with extracorporeal blood warming [see comments]. N. Engl. J. Med. 337, 1500–1505.
- Wanscher, M., Agersnap, L., Ravn, J., Yndgaard, S., Nielsen, J. F., Danielsen, E. R., et al. (2012). Outcome of accidental hypothermia with or without circulatory arrest: experience from the Danish Praesto Fjord boating accident. *Resuscitation* 83, 1078–1084. doi: 10.1016/j.resuscitation.2012.05.009
- Welbourn, C., and Efstathiou, N. (2018). How does the length of cardiopulmonary resuscitation affect brain damage in patients surviving cardiac arrest? A systematic review. Scand. J. Trauma Resusc. Emerg. Med. 26:77. doi: 10.1186/ s13049-018-0476-3
- Wenzel, V., Russo, S. G., Arntz, H. R., Bahr, J., Baubin, M. A., Bottiger, B. W., et al. (2010). Comments on the 2010 guidelines on cardiopulmonary resuscitation of the European Resuscitation Council. *Anaesthesist* 59, 1105–1123. doi: 10.1007/ s00101-010-1820-9
- Wik, L., Naess, P. A., Ilebekk, A., Nicolaysen, G., and Steen, P. A. (1996). Effects of various degrees of compression and active decompression on haemodynamics, end-tidal CO2, and ventilation during cardiopulmonary resuscitation of pigs. *Resuscitation* 31, 45–57.

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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