Check for updates

OPEN ACCESS

EDITED BY Anna Bukiya, University of Tennessee Health Science Center (UTHSC), United States

REVIEWED BY Tamer M. Gamal El-Din, University of Washington, United States

*CORRESPONDENCE Stephen G. Waxman, stephen.waxman@yale.edu

SPECIALTY SECTION This article was submitted to Membrane Physiology and Membrane Biophysics, a section of the journal Frontiers in Physiology

RECEIVED 10 October 2022 ACCEPTED 31 October 2022 PUBLISHED 10 November 2022

CITATION

Ghovanloo M-R, Dib-Hajj SD, Goodchild SJ, Ruben PC and Waxman SG (2022), Non-psychotropic phytocannabinoid interactions with voltage-gated sodium channels: An update on cannabidiol and cannabigerol. *Front. Physiol.* 13:1066455. doi: 10.3389/fphys.2022.1066455

COPYRIGHT

© 2022 Ghovanloo, Dib-Hajj, Goodchild, Ruben and Waxman. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY).

The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Non-psychotropic phytocannabinoid interactions with voltage-gated sodium channels: An update on cannabidiol and cannabigerol

Mohammad-Reza Ghovanloo¹, Sulayman D. Dib-Hajj¹, Samuel J. Goodchild², Peter C. Ruben³ and Stephen G. Waxman¹*

¹Department of Neurology, Yale University School of Medicine, New Haven, CT, United States, ²Department of Cellular and Molecular Biology, Xenon Pharmaceuticals Inc., Burnaby, BC, Canada, ³Department of Biomedical Physiology and Kinesiology, Simon Fraser University, Burnaby, BC, Canada

Phytocannabinoids, found in the plant, *Cannabis sativa*, are an important class of natural compounds with physiological effects. These compounds can be generally divided into two classes: psychoactive and non-psychoactive. Those which do not impart psychoactivity are assumed to predominantly function *via* endocannabinoid receptor (CB) -independent pathways and molecular targets, including other receptors and ion channels. Among these targets, the voltage-gated sodium (Nav) channels are particularly interesting due to their well-established role in electrical signalling in the nervous system. The interactions between the main non-psychoactive phytocannabinoid, cannabidiol (CBD), and Nav channels were studied in detail. In addition to CBD, cannabigerol (CBG), is another non-psychoactive molecule implicated as a potential therapeutic for several conditions, including pain *via* interactions with Nav channels. In this mini review, we provide an update on the interactions of Nav channels with CBD and CBG.

KEYWORDS

cannabidiol (CBD), cannabigerol (CBG), voltage-gated sodium (Nav) channels, excitability, pharmacology

Introduction

The cannabis plant contains over 120 active phytocannabinoids (Morales et al., 2017). Among these molecules, there are some that are psychotropic, and others that are not. Cannabidiol (CBD) is the primary non-psychotropic phytocannabinoid (Pertwee, 2008; Ghovanloo and Ruben, 2021). CBD has received extensive attention in recent years due to many anecdotal and some clinically substantiated reports of efficacy against various conditions (Billakota et al., 2019; Ghovanloo and Ruben, 2021). The interest in CBD has increased since the success of Epidiolex (therapeutic CBD) in large-scale clinical trials

against Dravet and Lennox-Gastaut syndromes, which are severe pediatric-onset epileptic encephalopathies (Devinsky et al., 2017; Devinsky et al., 2018). However, despite its clinical efficacy, the exact mechanism of action for CBD remains undetermined.

CBD, the In contrast to main psychotropic phytocannabinoid, Δ9-tetrahydrocannabinol (THC), has a relatively clearcut mode of action. THC is a potent agonist at the human endocannabinoid (CB) receptors (~13-90 nM) (Turner et al., 2017). The physiological function of these receptors is to respond to endogenous lipid agonists, anandamide and 2-arachidonoylglycerol (Devane et al., 1988; Devane et al., 1992; Vogel et al., 1993; Howlett, 2005; Di Marzo, 2008). Similar to CBD, THC has been shown to possess anticonvulsant properties in animal models; however, the noted psychotropic effects of this compound make it a less than ideal therapeutic candidate (Lynch et al., 2004; Pinsger et al., 2006; Russo and Guy, 2006; Mechoulam et al., 2007; Skrabek et al., 2008; Johnson et al., 2010; Ware et al., 2010).

CBD has low affinity for the CB receptors, where it has mild antagonistic effects (Tham et al., 2019). Therefore, CBindependent targets are the most likely molecular mechanisms underlying CBD's efficacy. CBD was shown to interact with GPR55 receptors (Sylantyev et al., 2013; Kaplan et al., 2017), which are proteins that are expressed in excitatory and inhibitory synapses and which modulate synaptic plasticity. CBD is also a modulator of several TRP channels (De Petrocellis et al., 2011; De Petrocellis et al., 2012; Hassan et al., 2014; Pumroy et al., 2019), 5-HT1A receptors (Campos and Guimarães, 2008; Fogaça et al., 2014; Marinho et al., 2015), and an inhibitor of adenosine reuptake by voltagedependent anion channel 1 (Rimmerman et al., 2013). Importantly, CBD is an inhibitor of voltage-dependent sodium (Nav) channels (Ghovanloo et al., 2018; Ghovanloo and Ruben, 2021; Zhang and Bean, 2021), some potassium channels (e.g., Kv2.1) (Ghovanloo et al., 2018; Orvos et al., 2020), calcium channels (Ross et al., 2008) and, in contrast, an activator of Kv7 channels in the nanomolar range (Zhang et al., 2022). Additionally, CBD directly modulates the biophysical properties of the bio-membrane itself (Ghovanloo et al., 2021; Ghovanloo et al., 2022b; Guard et al., 2022), which may facilitate an allosteric modulation of membrane proteins including, but not limited to, those noted above.

Among the CBD targets, the family of Nav channels are particularly interesting for three reasons. First, Dravet syndrome, the most notable condition for which CBD is efficacious, is commonly linked to genetic mutations in Nav1.1 (Yu et al., 2006; Kalume et al., 2007, 2015; Dravet, 2011; Devinsky et al., 2014; Peters et al., 2016; Gorman et al., 2021; Fouda et al., 2022). Nav1.1 is a key regulator of excitability in inhibitory circuits within the central nervous system. Second, CBD is reputed to have therapeutic value, substantiated by preclinical and animal studies, for a variety of excitability related disorders including pain, seizures, muscular problems, and arrhythmias, among others (Wade et al., 2004; Iannotti et al., 2019; Ghovanloo and Ruben, 2021). Dysfunction of various Nav channels in different tissues could trigger any of the noted conditions (Ghovanloo et al., 2016; Fouda et al., 2022). Third, amphiphilic compounds (e.g., Triton X-100) (Lundbæk et al., 2010) that modulate membrane elasticity (with properties that are similar to CBD) have been shown to allosterically stabilize Nav channel inactivation (Lundbæk, 2005; Lundbæk et al., 2010; Ghovanloo et al., 2021; Ghovanloo et al., 2022b). These reasons prompted us and others to study effects of CBD on Nav channels in detail over the past several years. CBD is now established as an effective Nav channel inhibitor (Ghovanloo et al., 2018; Fouda et al., 2020; Sait et al., 2020; Ghovanloo et al., 2021; Ghovanloo and Ruben, 2021; Zhang and Bean, 2021). Furthermore, these investigations suggested Nav channels are a promising pathway for cannabinoid-mediated reductions in macro excitability, with a substantial therapeutic potential. This pathway could be explored not just with CBD, but also with other compounds with similar physicochemical properties.

A common precursor for THC and CBD is cannabigerol (CBG) (Nachnani et al., 2021). Like THC (ChEMBL-calculated-LogD = 5.94) and CBD (ChEMBL-calculated-LogD = 6.60), CBG (ChEMBL-calculated-LogD = 7.04) is also a highly hydrophobic compound. Although CBG is less well studied than THC or CBD, the existing literature suggests that CBG's pharmacological profile falls in between these two cannabinoids. Importantly, while CBG's affinity for CB receptors is higher than CBD, CBG is non-psychotropic (Nachnani et al., 2021). This suggests that CBG could work through both CB-dependent and CBindependent (e.g., Nav channels, TRP channels, etc.) pathways without THC's unwanted psychoactive effects (Muller et al., 2019; Ghovanloo et al., 2022a). With this combination of properties, CBG offers the potential to be a superior therapeutic compound than either CBD or THC. A comparison of the key targets between CBD and CBG is provided in Table 1.

Much of the molecular details of CBD's interactions with Nav channels is reviewed in Ghovanloo and Ruben, 2021. In this short report, we provide new important updates on Nav channel mediated-cannabinoid pathway with a focus on CBD and CBG.

Cannabidiol—Mechanism of action on sodium channels

We previously found that CBD is a non-selective Nav channel inhibitor. Using voltage-clamp experiments, we found that CBD inhibits all human Nav1.1-7 from the inactivated states, with potencies ranging from 1.9 to 3.8 μ M, and steep Hill slopes of ~3. We also found that CBD imparts similar effects on Nav gating: inhibiting G_{max} without changing voltage-dependence of activation, but hyperpolarizing steady-state

Target	CBD	CBG	References
CB1	Inverse agonist/ antagonist	Weak agonist	(Cascio et al., 2010; Pollastro et al., 2011; Muller et al., 2019)
CB2	Inverse agonist	Partial agonist	(Cascio et al., 2010; Pollastro et al., 2011; Muller et al., 2019)
GPR55	Antagonist	Unknown	(Kaplan et al., 2017; Harding et al., 2018)
Nav	Inhibitor	Inhibitor	(Ghovanloo et al., 2018; Sait et al., 2020; Ghovanloo et al., 2021; Zhang and Bean, 2021; Ghovanloo et al., 2022a)
TRPA1	Agonist	Agonist	(De Petrocellis et al., 2011; Pollastro et al., 2011; Muller et al., 2019)
TRPV1	Agonist	Agonist	(De Petrocellis et al., 2011; Pollastro et al., 2011; Muller et al., 2019)
TRPV2	Agonist	Agonist	(De Petrocellis et al., 2011; Pollastro et al., 2011; Muller et al., 2019)
TRPV3	Agonist	Agonist	Muller et al. (2019)
TRPV4	Agonist	Agonist	Muller et al. (2019)
TRPM8	Antagonist	Antagonist	(De Petrocellis et al., 2011; Pollastro et al., 2011; Muller et al., 2019)
Kv7	Potentiator	Unknown	Zhang et al. (2022)
Kv2.1	Inhibitor	Unknown	Ghovanloo et al. (2018)
Cav	Inhibitor	Inhibitor	(Ross et al., 2008; Mirlohi et al., 2022)
Biomembrane	Modulator	Unknown	Ghovanloo et al. (2021)

TABLE 1 Comparison of a list of key receptors and ion channel targets between CBD and CBG. See (Almeida and Devi, 2020; Ghovanloo and Ruben, 2021; Nachnani et al., 2021) for more extensive reviews of these targets.

inactivation and slowing recovery from inactivation (Ghovanloo et al., 2018). Furthermore, we found that when Nav channels enter deeper inactivated states, CBD slows the recovery kinetics even further, consistent with state-dependent Nav channel inhibition. CBD has an approximately 10-fold statedependence, which makes it a moderately state-dependent Nav channel inhibitor (Ghovanloo et al., 2018; Ghovanloo and Ruben, 2021). From a molecular perspective, it may be theoretically conceivable to use CBD against Nav channelopathies that greatly impair inactivation (Ghovanloo et al., 2020).

The effects of CBD on Nav channels are the result of interactions at the interface of the channel pore and fenestrations in which CBD directly blocks the pore (in part *via* the local anesthetic phenylalanine), and alterations to the membrane elasticity which indirectly stabilizes Nav channel inactivation (Figure 1A). These results were elucidated using structural- (Sait et al., 2020), functional- (Ghovanloo et al., 2018, 2021), and molecular dynamics simulation-based (Ghovanloo et al., 2021) studies. It is important to note that the drug pathway from the membrane phase and through the Nav channel fenestrations is pharmacologically important, and has been elucidated with various drugs previously (Hille, 1977; Gamal El-Din et al., 2018).

One of CBD's main proposed clinical application is in pain treatment (Ward et al., 2014). There are several different Nav channels within the peripheral sensory pathway (Rush et al., 2007; Dib-Hajj et al., 2010), which include both tetrodotoxinsensitive (TTX-S) and resistant (TTX-R) (Nav1.8/9) subtypes (Cummins et al., 2007; Bennett et al., 2019). In contrast to most other Nav channels, the TTX-R channels have a hyperpolarized voltage-dependence of slow inactivation relative to their fast inactivation and a more rapid entry into slow inactivation than other Nav channels (Vilin and Ruben, 2001; Blair and Bean, 2003; Ghovanloo et al., 2016). Therefore, the slow inactivation properties of these channels can limit repetitive firing in their native environment. Because of these properties, drugs that target slow inactivated states could be effective for reducing repetitive firing in sensory neurons.

A recent study determined that CBD at 500 nM has tight binding to the slow inactivated states of Nav1.8 (Zhang and Bean, 2021). These low concentrations of CBD had little effect on the first several action potentials, but as the current injection became larger, CBD reduced firing. Furthermore, CBD reduced the action potential height, widened the action potential, reduced afterhyperpolarization, and increased the propensity of entering depolarization block (Zhang and Bean, 2021).

Another recent study has shown that CBD-dominant nutraceutical products can inhibit Nav channels even more potently than pure CBD (difference is in the order of nanomolar to low micromolar range) (Milligan et al., 2022). This suggests that individual components of these nutraceutical products, such as other phytocannabinoids and terpenes, may synergistically further modulate or inhibit Nav channels (Milligan et al., 2022).

In addition to Nav channels, a new study showed that CBD at sub-micromolar concentrations, hyperpolarizes the voltagedependence of Kv7.2/3. This shift results in an enhancement of the M-current which has a powerful effect on dampening down neuronal excitability and has previously been clinically exploited by effective drugs such as Retigabine (Rundfeldt and Netzer, 2000). This effect may be a key contribution to the anticonvulsive and proposed analgesic activities of CBD, independently of other ion channel modulating effects (Zhang et al., 2022).



FIGURE 1

Cartoon summary of CBD and CBG effects on Nav channels. This figure is a redrawn version of our previously published papers (Ghovanloo and Ruben, 2021; Ghovanloo et al., 2022a). (A) Shows the pathway of CBD from the lipid phase through the mammalian Nav fenestration and into the pore, where it interacts to some extent by the local anesthetic (LA) site with the pore phenylalanine (F) residue. (B) Cartoon representation of the concentration-dependent modality of CBG effects on Nav channels. CBG, like CBD is a state-dependent Nav inhibitor with an increased affinity for the inactivated state. Overall, however, CBG causes a reduction in total conductance/channel opening more potently than inactivation is stabilized in the presence of CBG.

Cannabigerol—A potentially promising avenue for pain treatment via sodium channels

The role of Nav1.7 in the pain pathway is well-established (Dib-Hajj et al., 2013). Many gain- and loss-of-function mutations in Nav1.7 have been identified. Hyperexcitability in this channel has been shown to elicit several pain syndromes (Dib-Hajj et al., 2005, 2013; Fertleman et al., 2006), whilst hypoexcitability in this channel is linked with complete insensitivity to pain, which is not accompanied with any cognitive, cardiac, or motor defects (Cox et al., 2006; Goldberg et al., 2007). These findings highlight the importance of Nav1.7 as an excellent target for pain therapy; however, the efforts that have gone into developing small molecules for Nav1.7 inhibition have thus far been unsuccessful. This lack of success has been attributed to problems in achieving optimum channel occupancy and, thus, problems in effective target engagement (Bankar et al., 2018). To get around this problem, in vivo treatments with many folds above IC₅₀ could be utilized, but at these high dosages, this would cause unwanted side effects.

A potential advantage of highly hydrophobic compounds like cannabinoids is that they might more readily get absorbed into the lipid dense neuronal tissues and nerve membranes. This would require the mode of administration of the compound to reduce exposure in the central nervous system and increase the probability of distribution into the peripheral nerves, to avoid off target effects in the CNS: central nervous system. If the compound also had either structural or functional selectivity for Nav1.7, then efficacy may be achievable. Whilst structural selectivity would refer to a unique amino acid sequence or motif that is present in one channel (Catterall and Swanson, 2015), but not others, functional selectivity refers to the ligand's (in this case CBG) increased affinity for the channel that occupies one particular state within its local environment (Kuo and Bean, 1994). As the resting membrane potential of sensory dorsal root ganglion (DRG) neurons is considerably more depolarized than the availability voltage-dependence of Nav1.7 (relative to other sensory Nav channels), Nav1.7 in these neurons accumulates a lot more inactivation than other Nav channels (Amir et al., 1999; Ghovanloo et al., 2022a). Thus, the state-dependent drug that has a higher apparent potency for the inactivated states of these channels, would be predicted to more effectively inhibit Nav1.7 in DRG neurons than the other Nav channels.

CBD and CBG are both highly hydrophobic compounds with very high distribution coefficients for the hydrophobic phase. In fact, CBG (ChEMBL-calculated-LogD = 7.04) is even more hydrophobic than CBD (ChEMBL-calculated-LogD = 6.60), which suggests that CBG may have a higher propensity to enter and remain within the lipid membranes. However, neither compound has much structural selectivity for Nav channels. As noted above, CBD has been shown to bind at

the Nav channel pore, which is a highly conserved region of the channel (Catterall and Swanson, 2015; Ghovanloo and Ruben, 2021). Thus, like the other classic pore blockers that bind this region (Hille, 1977; Bean et al., 1983; Gamal El-Din et al., 2018), CBD is not structurally selective. Although CBG has not been functionally tested for its binding site on the Nav channel, it likely interacts within the same site at the porefenestration interface.

Because CBG was implicated as an analgesic (Evans, 1991; Mammana et al., 2019), we investigated its effects on Nav channels. We determined that CBG is also a moderately state-dependent Nav channel inhibitor, and shares (CBG is slightly less potent than CBD) many of the same features in its modulation of Nav channels with CBD (Ghovanloo et al., 2022a). We found that CBG does not alter the voltagedependence of activation or alter open-state inactivation, it hyperpolarizes inactivation curves, slows recovery from inactivation (with this effect becoming more pronounced as the channels enter deeper inactivated states, e.g., slow inactivation), accelerates onset of closed-state fast inactivation, and reduces spontaneous firing of DRG neurons. Importantly, we found that CBG inhibits total channel conductance more potently than it stabilizes the inactivated state of channels (inactivation shift potency = 13.3 \pm 1.0 μ M, G_{max} inhibition potency=3.4 \pm 1.0 μ M) (Ghovanloo et al., 2022a) (Figure 1B). As Nav1.7 is known as the threshold channel for peripheral firing (Dib-Hajj et al., 2013), these results suggest that CBG may be more effective in preventing pain episodes from initiating. Given that polypharmacological compounds will display more promiscuity in their interactions with various targets at higher concentrations (hence toxicity), the best therapeutic window for CBG's potential therapeutic efficacy will occur by taking advantage of the Nav channel G_{max} block at the low to submicromolar concentrations where CBG's hyperpolarization of inactivation would be physiologically inconsequential (Ghovanloo et al., 2022a). Finally, the noted mechanism could, in principle, work in concert with CBG's CBdependent (without psycho-activity) pathways to target pain.

We suggest that the development of CBD/CBG as Nav channel-targeting drugs may be achievable *via* exploring various modes of administration. For instance, with respect to pain, if the compound could be localized to the nociceptors, then given the physicochemical properties of the compound, along with local resting membrane potential and the availability voltage-dependences of the local Nav channels, the channel that is most inactivated would be most modulated (as noted above, Nav1.7 (Ghovanloo et al., 2018; 2022a)). We also note that the concentrations at which CBD targets slow inactivation of Nav1.8 and Kv7.2/3, on the low end are comparable (Zhang and Bean, 2021; Zhang et al., 2022). Studies suggest that in the case of CBD, mode of administration can substantially alter pharmacokinetics and bioavailability, and hence tissue distribution of the drug (e.g., local administration to work alongside compound hydrophobicity to accumulate in tissues with high lipid content (Ghovanloo et al., 2022a)) (Millar et al., 2018; Lim et al., 2020); however, even then, the drug effect on phenotype would not likely be able to be attributed to activity on a single target (e.g., Na or K currents, TRP channels, etc.), but we propose that Nav channels are likely an important part of the pleiotropic pharmacology of CBD. These considerations would be critical in translation of using these compounds from the bench to the bedside.

Concluding remarks

CBD and CBG, and indeed other cannabinoids and terpenes are intriguing molecules with highly complex pharmacological profiles. Despite enormous progress in recent years, the precise mechanism of clinical efficacy remains unknown. The Nav channel family is vital to nervous system signalling, and it is likely an important receptor for these molecules. Future investigations into the intricate interactions between Nav channels (and other receptors) and cannabinoids will facilitate unravelling how cannabinoids impart their effects on physiology, which could aid the identification of novel therapeutics for various disorders of neuronal excitability.

Author contributions

M-RG wrote the manuscript, and all other co-authors edited, revised, and confirmed the manuscript.

References

Almeida, D. L., and Devi, L. A. (2020). Diversity of molecular targets and signaling pathways for CBD. *Pharmacol. Res. Perspect.* 8. doi:10.1002/prp2.682

Amir, R., Michaelis, M., and Devor, M. (1999). Membrane potential oscillations in dorsal root ganglion neurons: Role in Normal electrogenesis and neuropathic pain. J. Neurosci. 19 (19), 8589–8596. doi:10.1523/JNEUROSCI.19-19-08589.1999

Bankar, G., Goodchild, S. J., Howard, S., Nelkenbrecher, K., Waldbrook, M., Dourado, M., et al. (2018). Selective NaV1.7 antagonists with Long residence Time Show Improved efficacy against Inflammatory and neuropathic pain. *Cell Rep.* 24, 3133–3145. doi:10.1016/j.celrep.2018.08.063

Bean, B. P., Cohen, C. J., and Tsien, R. W. (1983). Lidocaine block of cardiac sodium channels. J. Gen. Physiol. 81, 613-642. doi:10.1085/jgp.81.5.613

Bennett, D. L., Clark, X. A. J., Huang, J., Waxman, S. G., and Dib-Hajj, S. D. (2019). The role of voltage-gated sodium channels in pain signaling. *Physiol. Rev.* 99, 1079–1151. doi:10.1152/physrev.00052.2017

Billakota, S., Devinsky, O., and Marsh, E. (2019). Cannabinoid therapy in epilepsy. Curr. Opin. Neurol. 32, 220–226. doi:10.1097/wco.00000000000660

Blair, N. T., and Bean, B. P. (2003). Role of tetrodotoxin-resistant Na+ current slow inactivation in adaptation of action potential firing in small-diameter dorsal root ganglion neurons. *J. Neurosci.* 23, 10338–10350. doi:10.1523/JNEUROSCI.23-32-10338.2003

Campos, A. C., and Guimarães, F. S. (2008). Involvement of 5HT1A receptors in the anxiolytic-like effects of cannabidiol injected into the dorsolateral periaqueductal gray of rats. *Psychopharmacol. Berl.* 199, 223–230. doi:10.1007/ S00213-008-1168-X

Acknowledgments

The Center for Neuroscience and Regeneration Research is a Collaboration of the Paralyzed Veterans of America with Yale University. M-RG is a Banting Fellow and is supported by the Canadian Institutes of Health Research (CIHR). SG is funded by Xenon Pharmaceuticals Inc. PR is funded by a Discovery grant from Natural Sciences and Engineering Research Council of Canada (NSERC).

Conflict of interest

SG is an employee of Xenon Pharmaceuticals Inc. No funders were involved in the study design, collection, analysis, interpretation of data, the writing of this article or the decision to submit it for publication.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Cascio, M. G., Gauson, L. A., Stevenson, L. A., Ross, R. A., and Pertwee, R. G. (2010). Evidence that the plant cannabinoid cannabigerol is a highly potent alpha2adrenoceptor agonist and moderately potent 5HT1A receptor antagonist. *Br. J. Pharmacol.* 159, 129–141. doi:10.1111/j.1476-5381.2009.00515.x

Catterall, W. A., and Swanson, T. M. (2015). Structural basis for pharmacology of voltage-gated sodium and calcium channels. *Mol. Pharmacol.* 88, 141–150. doi:10. 1124/mol.114.097659

Cox, J. J., Reimann, F., Nicholas, A. K., Thornton, G., Roberts, E., Springell, K., et al. (2006). An SCN9A channelopathy causes congenital inability to experience pain. *Nature* 444, 894–898. doi:10.1038/nature05413

Cummins, T. R., Sheets, P. L., and Waxman, S. G. (2007). The roles of sodium channels in nociception: Implications for mechanisms of pain. *Pain* 131, 243–257. doi:10.1016/J.PAIN.2007.07.026

De Petrocellis, L., Ligresti, A., Moriello, A. S., Allarà, M., Bisogno, T., Petrosino, S., et al. (2011). Effects of cannabinoids and cannabinoid-enriched Cannabis extracts on TRP channels and endocannabinoid metabolic enzymes. *Br. J. Pharmacol.* 163, 1479–1494. doi:10.1111/j.1476-5381.2010.01166.x

De Petrocellis, L., Orlando, P., Moriello, A. S., Aviello, G., Stott, C., Izzo, A. A., et al. (2012). Cannabinoid actions at TRPV channels: Effects on TRPV3 and TRPV4 and their potential relevance to gastrointestinal inflammation. *Acta Physiol.* 204, 255–266. doi:10.1111/j.1748-1716.2011.02338.x

Devane, W. A., Dysarz, F. A., Johnson, M. R., Melvin, L. S., and Howlett, A. C. (1988). Determination and characterization of a cannabinoid receptor in rat brain. *Mol. Pharmacol.* 34, 605–613. (Accessed February 11, 2019).

Devane, W. A., Hanuš, L., Breuer, A., Pertwee, R. G., Stevenson, L. A., Griffin, G., et al. (1992). Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science* 258, 1946–1949. doi:10.1126/SCIENCE.1470919

Devinsky, O., Cilio, M. R., Cross, H., Fernandez-Ruiz, J., French, J., Hill, C., et al. (2014). Cannabidiol: Pharmacology and potential therapeutic role in epilepsy and other neuropsychiatric disorders. *Epilepsia* 55, 791–802. doi:10.1111/epi.12631

Devinsky, O., Cross, J. H., Laux, L., Marsh, E., Miller, I., Nabbout, R., et al. (2017). Trial of cannabidiol for drug-resistant seizures in the Dravet syndrome. *N. Engl. J. Med.* 376, 2011–2020. doi:10.1056/NEJMoa1611618

Devinsky, O., Patel, A. D., Cross, J. H., Villanueva, V., Wirrell, E. C., Privitera, M., et al. (2018). Effect of cannabidiol on Drop seizures in the Lennox–Gastaut syndrome. *N. Engl. J. Med.* 378, 1888–1897. doi:10.1056/nejmoa1714631

Di Marzo, V. (2008). CB(1) receptor antagonism: Biological basis for metabolic effects. *Drug Discov. Today* 13, 1026–1041. doi:10.1016/J.DRUDIS.2008.09.001

Dib-Hajj, S. D., Cummins, T. R., Black, J. A., and Waxman, S. G. (2010). Sodium channels in Normal and Pathological pain. *Annu. Rev. Neurosci.* 33, 325–347. doi:10.1146/annurev-neuro-060909-153234

Dib-Hajj, S. D., Rush, A. M., Cummins, T. R., Hisama, F. M., Novella, S., Tyrrell, L., et al. (2005). Gain-of-function mutation in Nav1.7 in familial erythromelalgia induces bursting of sensory neurons. *Brain* 128, 1847–1854. doi:10.1093/brain/awh514

Dib-Hajj, S. D., Yang, Y., Black, J. A., and Waxman, S. G. (2013). The Na v 1.7 sodium channel: From molecule to man. *Nat. Rev. Neurosci.* 14, 49–62. doi:10. 1038/nrn3404

Dravet, C. (2011). The core Dravet syndrome phenotype. *Epilepsia* 52, 3–9. doi:10.1111/j.1528-1167.2011.02994.x

Evans, F. J. (1991). Cannabinoids: The separation of central from peripheral effects on a structural basis. *Planta Med.* 57, S60–S67. doi:10.1055/s-2006-960231

Fertleman, C. R., Baker, M. D., Parker, K. A., Moffatt, S., Elmslie, F. V., Abrahamsen, B., et al. (2006). SCN9A mutations in Paroxysmal Extreme pain disorder: Allelic Variants underlie Distinct channel defects and phenotypes. *Neuron* 52, 767–774. doi:10.1016/j.neuron.2006.10.006

Fogaça, M. V., Reis, F. M. C. V., Campos, A. C., and Guimarães, F. S. (2014). Effects of intra-prelimbic prefrontal cortex injection of cannabidiol on anxiety-like behavior: Involvement of 5HT1A receptors and previous stressful experience. *Eur. Neuropsychopharmacol.* 24, 410–419. doi:10.1016/J.EURONEURO.2013.10.012

Fouda, M. A., Ghovanloo, M.-R., and Ruben, P. C. (2020). Cannabidiol protects against high glucose-induced oxidative stress and cytotoxicity in cardiac voltage-gated sodium channels. Br. J. Pharmacol. 177, 2932–2946. doi:10.1111/bph.15020

Fouda, M. A., Ghovanloo, M.-R., and Ruben, P. C. (2022). Late sodium current: Incomplete inactivation triggers seizures, myotonias, arrhythmias, and pain syndromes. J. Physiol. 600, 2835–2851. doi:10.1113/JP282768

Gamal El-Din, T. M., Lenaeus, M. J., Zheng, N., and Catterall, W. A. (2018). Fenestrations control resting-state block of a voltage-gated sodium channel. *Proc. Natl. Acad. Sci. U. S. A.* 115, 13111–13116. doi:10.1073/pnas.1814928115

Ghovanloo, M.-R., Aimar, K., Ghadiry-Tavi, R., Yu, A., and Ruben, P. C. (2016). Physiology and Pathophysiology of sodium channel inactivation. *Curr. Top. Membr.* 78, 479–509. doi:10.1016/bs.ctm.2016.04.001

Ghovanloo, M.-R., Atallah, J., Escudero, C. A., and Ruben, P. C. (2020). Biophysical characterization of a novel SCN5A mutation associated with an Atypical phenotype of Atrial and Ventricular arrhythmias and Sudden death. *Front. Physiol.* 11, 610436. doi:10.3389/fphys.2020.610436

Ghovanloo, M.-R., Choudhury, K., Bandaru, T. S., Fouda, M. A., Rayani, K., Rusinova, R., et al. (2021). Cannabidiol inhibits the skeletal muscle nav1.4 by blocking its pore and by altering membrane elasticity. *J. Gen. Physiol.* 153, e202012701. doi:10.1085/jgp.202012701

Ghovanloo, M.-R., Estacion, M., Higerd-Rusli, G. P., Zhao, P., Dib-Hajj, S., and Waxman, S. G. (2022a). Inhibition of sodium conductance by cannabigerol contributes to a reduction of dorsal root ganglion neuron excitability. *Br. J. Pharmacol.* 179, 4010–4030. doi:10.1111/bph.15833

Ghovanloo, M.-R., Goodchild, S. J., and Ruben, P. C. (2022b). Cannabidiol increases gramicidin current in human embryonic kidney cells: An observational study. *PLoS One* 17, e0271801. doi:10.1371/journal.pone.0271801

Ghovanloo, M.-R., and Ruben, P. C. (2021). Cannabidiol and sodium channel pharmacology: General overview, mechanism, and clinical Implications. *Neuroscientist.* 28, 318–334. doi:10.1177/10738584211017009

Ghovanloo, M.-R., Shuart, N. G., Mezeyova, J., Dean, R. A., Ruben, P. C., and Goodchild, S. J. (2018). Inhibitory effects of cannabidiol on voltage-dependent sodium currents. *J. Biol. Chem.* 293, 16546–16558. doi:10.1074/jbc.RA118.004929

Goldberg, Y. P., Macfarlane, J., Macdonald, M. L., Thompson, J., Dube, M. P., Mattice, M., et al. (2007). Loss-of-function mutations in the Nav1.7 gene underlie

congenital indifference to pain in multiple human populations. Clin. Genet. 71, 311-319. doi:10.1111/j.1399-0004.2007.00790.x

Gorman, K. M., Peters, C. H., Lynch, B., Jones, L., Bassett, D. S., King, M. D., et al. (2021). Persistent sodium currents in SCN1A developmental and degenerative epileptic dyskinetic encephalopathy. *Brain Commun.* 3, fcab235. doi:10.1093/ braincomms/fcab235

Guard, S. E., Chapnick, D. A., Poss, Z. C., Ebmeier, C. C., Jacobsen, J., Nemkov, T., et al. (2022). Multi-omic analysis Reveals Disruption of Cholesterol Homeostasis by cannabidiol in human cell Lines. *Mol. Cell. Proteomics* 21 (10), 100262. doi:10.1016/j.mcpro.2022.100262

Harding, S. D., Sharman, J. L., Faccenda, E., Southan, C., Pawson, A. J., Ireland, S., et al. (2018). The IUPHAR/BPS Guide to PHARMACOLOGY in 2018: Updates and expansion to encompass the new guide to IMMUNOPHARMACOLOGY. *Nucleic Acids Res.* 46, D1091–D1106. doi:10.1093/nar/gkx1121

Hassan, S., Eldeeb, K., Millns, P. J., Bennett, A. J., Alexander, S. P. H., and Kendall, D. A. (2014). Cannabidiol enhances microglial phagocytosis via transient receptor potential (TRP) channel activation. *Br. J. Pharmacol.* 171, 2426–2439. doi:10.1111/ BPH.12615

Hille, B. (1977). Local anesthetics: Hydrophilic and hydrophobic pathways for the drug-receptor reaction. J. Gen. Physiol. 69, 497-515. doi:10.1085/jgp.69.4.497

Howlett, A. C. (2005). Cannabinoid receptor signaling. *Handb. Exp. Pharmacol.* 168, 53–79. doi:10.1007/3-540-26573-2_2

Iannotti, F. A., Pagano, E., Moriello, A. S., Alvino, F. G., Sorrentino, N. C., D'Orsi, L., et al. (2019). Effects of non-euphoric plant cannabinoids on muscle quality and performance of dystrophic mdx mice. *Br. J. Pharmacol.* 176, 1568–1584. doi:10. 1111/bph.14460

Johnson, J. R., Burnell-Nugent, M., Lossignol, D., Ganae-Motan, E. D., Potts, R., and Fallon, M. T. (2010). Multicenter, double-blind, randomized, placebocontrolled, Parallel-Group study of the efficacy, Safety, and Tolerability of THC: CBD Extract and THC Extract in patients with Intractable Cancer-related pain. *J. Pain Symptom Manage.* 39, 167–179. doi:10.1016/j.jpainsymman.2009.06.008

Kalume, F., Oakley, J. C., Westenbroek, R. E., Gile, J., de la Iglesia, H. O., Scheuer, T., et al. (2015). Sleep impairment and reduced interneuron excitability in a mouse model of Dravet Syndrome. *Neurobiol. Dis.* 77, 141–154. doi:10.1016/J.NBD.2015. 02.016

Kalume, F., Yu, F. H., Westenbroek, R. E., Scheuer, T., and Catterall, W. A. (2007). Reduced sodium current in Purkinje neurons from NaV1.1 mutant mice: Implications for Ataxia in severe myoclonic epilepsy in infancy. *J. Neurosci.* 27, 11065–11074. doi:10.1523/JNEUROSCI.2162-07.2007

Kaplan, J. S., Stella, N., Catterall, W. A., and Westenbroek, R. E. (2017). Cannabidiol attenuates seizures and social deficits in a mouse model of Dravet syndrome. *Proc. Natl. Acad. Sci. U. S. A.* 114, 11229–11234. doi:10.1073/pnas. 1711351114

Kuo, C. C., and Bean, B. P. (1994). Slow binding of phenytoin to inactivated sodium channels in rat hippocampal neurons. *Mol. Pharmacol.* 46, 716–725.

Lim, S. Y., Sharan, S., and Woo, S. (2020). Model-based analysis of cannabidiol Dose-exposure relationship and bioavailability. *Pharmacotherapy* 40, 291–300. doi:10.1002/phar.2377

Lundbæk, J. A., Collingwood, S. A., Ingólfsson, H. I., Kapoor, R., and Andersen, O. S. (2010). Lipid bilayer regulation of membrane protein function: Gramicidin channels as molecular force probes. *J. R. Soc. Interface* 7, 373–395. doi:10.1098/rsif. 2009.0443

Lundbæk, J., Birn, P., Tape, S. E., Toombes, G. E. S., SogaaRd, R., Koeppe, R. E., et al. (2005). Capsaicin Regulates voltage-dependent sodium Channelsby altering lipid bilayer elasticity. *Mol. Pharmacol.* 68, 680–689. doi:10.1124/mol.105.013573

Lynch, J. J., Wade, C. L., Zhong, C. M., Mikusa, J. P., and Honore, P. (2004). Attenuation of mechanical allodynia by clinically utilized drugs in a rat chemotherapy-induced neuropathic pain model. *Pain* 110, 56–63. doi:10.1016/j. pain.2004.03.010

Mammana, S., Cavalli, E., Gugliandolo, A., Silvestro, S., Pollastro, F., Bramanti, P., et al. (2019). Could the combination of two non-psychotropic cannabinoids counteract neuroinflammation? Effectiveness of cannabidiol associated with cannabigerol. *Medicina* 55, E747. doi:10.3390/medicina55110747

Marinho, A. L. Z., Vila-Verde, C., Fogaça, M. V., and Guimarães, F. S. (2015). Effects of intra-infralimbic prefrontal cortex injections of cannabidiol in the modulation of emotional behaviors in rats: Contribution of 5HT1A receptors and stressful experiences. *Behav. Brain Res.* 286, 49–56. doi:10.1016/J.BBR.2015.02.023

Mechoulam, R., Peters, M., Murillo-Rodriguez, E., and Hanuš, L. O. (2007). Cannabidiol - recent advances. *Chem. Biodivers.* 4, 1678–1692. doi:10.1002/cbdv.200790147

Millar, S. A., Stone, N. L., Yates, A. S., and O'Sullivan, S. E. (2018). A Systematic review on the pharmacokinetics of cannabidiol in humans. *Front. Pharmacol.* 9, 1365. doi:10.3389/fphar.2018.01365 Milligan, C. J., Anderson, L. L., Bowen, M. T., Banister, S. D., McGregor, I. S., Arnold, J. C., et al. (2022). A nutraceutical product, extracted from Cannabis sativa, modulates voltage-gated sodium channel function. *J. cannabis Res.* 4, 30. doi:10. 1186/S42238-022-00136-X

Mirlohi, S., Bladen, C., Santiago, M. J., Arnold, J. C., McGregor, I., and Connor, M. (2022). Inhibition of human recombinant T-type calcium channels by phytocannabinoids *in vitro*. Br. J. Pharmacol. 179, 4031–4043. doi:10.1111/BPH. 15842

Morales, P., Hurst, D. P., and Reggio, P. H. (2017). Molecular targets of the phytocannabinoids: A complex Picture. *Prog. Chem. Org. Nat. Prod.* 103, 103–131. doi:10.1007/978-3-319-45541-9_4

Muller, C., Morales, P., and Reggio, P. H. (2019). Cannabinoid ligands targeting TRP channels. *Front. Mol. Neurosci.* 11, 487. doi:10.3389/fnmol.2018.00487

Nachnani, R., Raup-Konsavage, W. M., and Vrana, K. E. (2021). The pharmacological case for cannabigerol. J. Pharmacol. Exp. Ther. 376, 204–212. doi:10.1124/jpet.120.000340

Orvos, P., Pászti, B., Topal, L., Gazdag, P., Prorok, J., Polyák, A., et al. (2020). The electrophysiological effect of cannabidiol on hERG current and in Guineapig and rabbit cardiac preparations. *Sci. Rep.* 10, 16079. doi:10.1038/s41598-020-73165-2

Pertwee, R. G. (2008). The diverse CB1 and CB2 receptor pharmacology of three plant cannabinoids: delta9-tetrahydrocannabinol, cannabidiol and delta9-tetrahydrocannabivarin. *Br. J. Pharmacol.* 153, 199–215. doi:10.1038/ sj.bjp.0707442

Peters, C., Rosch, R. E., Hughes, E., and Ruben, P. C. (2016). Temperaturedependent changes in neuronal dynamics in a patient with an SCN1A mutation and hyperthermia induced seizures. *Sci. Rep.* 6, 31879. doi:10.1038/srep31879

Pinsger, M., Schimetta, W., Volc, D., Hiermann, E., Riederer, F., and Pölz, W. (2006). Benefits of an add-on treatment with the synthetic cannabinomimetic nabilone on patients with chronic pain--a randomized controlled trial. *Wien. Klin. Wochenschr.* 118, 327–335. doi:10.1007/S00508-006-0611-4

Pollastro, F., Taglialatela-Scafati, O., Allarà, M., Muñoz, E., Di Marzo, V., De Petrocellis, L., et al. (2011). Bioactive prenylogous cannabinoid from fiber hemp (Cannabis sativa). J. Nat. Prod. 74, 2019–2022. doi:10.1021/np200500p

Pumroy, R. A., Samanta, A., Liu, Y., Hughes, T. E., Zhao, S., Yudin, Y., et al. (2019). Molecular mechanism of TRPV2 channel modulation by cannabidiol. *Elife* 8, e48792. doi:10.7554/elife.48792

Rimmerman, N., Ben-Hail, D., Porat, Z., Juknat, A., Kozela, E., Daniels, M. P., et al. (2013). Direct modulation of the outer mitochondrial membrane channel, voltage-dependent anion channel 1 (VDAC1) by cannabidiol: A novel mechanism for cannabinoid-induced cell death. *Cell Death Dis.* 4, e949. doi:10.1038/CDDIS. 2013.471

Ross, H. R., Napier, I., and Connor, M. (2008). Inhibition of recombinant human T-type calcium channels by Delta9-tetrahydrocannabinol and cannabidiol. *J. Biol. Chem.* 283, 16124–16134. doi:10.1074/jbc.M707104200

Rundfeldt, C., and Netzer, R. (2000). The novel anticonvulsant retigabine activates M-currents in Chinese hamster ovary-cells tranfected with human KCNQ2/3 subunits. *Neurosci. Lett.* 282, 73–76. doi:10.1016/S0304-3940(00) 00866-1

Rush, A. M., Cummins, T. R., and Waxman, S. G. (2007). Multiple sodium channels and their roles in electrogenesis within dorsal root ganglion neurons. *J. Physiol.* 579, 1–14. doi:10.1113/jphysiol.2006.121483

Russo, E., and Guy, G. W. (2006). A tale of two cannabinoids: The therapeutic rationale for combining tetrahydrocannabinol and cannabidiol. *Med. Hypotheses* 66, 234–246. doi:10.1016/j.mehy.2005.08.026

Sait, L. G., Sula, A., Ghovanloo, M.-R., Hollingworth, D., Ruben, P. C., and Wallace, B. A. (2020). Cannabidiol interactions with voltage-gated sodium channels. *Elife* 9, 585933–e58617. doi:10.7554/eLife.58593

Skrabek, R. Q., Galimova, L., Ethans, K., and Perry, D. (2008). Nabilone for the treatment of pain in Fibromyalgia. J. Pain 9, 164–173. doi:10.1016/j.jpain.2007.09.002

Sylantyev, S., Jensen, T. P., Ross, R. A., and Rusakov, D. A. (2013). Cannabinoidand lysophosphatidylinositol-sensitive receptor GPR55 boosts neurotransmitter release at central synapses. *Proc. Natl. Acad. Sci. U. S. A.* 110, 5193–5198. doi:10. 1073/pnas.1211204110

Tham, M., Yilmaz, O., Alaverdashvili, M., Kelly, M. E. M., Denovan-Wright, E. M., and Laprairie, R. B. (2019). Allosteric and orthosteric pharmacology of cannabidiol and cannabidiol-dimethylheptyl at the type 1 and type 2 cannabinoid receptors. *Br. J. Pharmacol.* 176, 1455–1469. doi:10.1111/bph.14440

Turner, S. E., Williams, C. M., Iversen, L., and Whalley, B. J. (2017). Molecular pharmacology of phytocannabinoids. *Prog. Chem. Org. Nat. Prod.* 103, 61–101. doi:10.1007/978-3-319-45541-9_3

Vilin, Y. Y., and Ruben, P. C. (2001). Slow inactivation in voltage-gated sodium channels: Molecular substrates and contributions to channelopathies. *Cell biochem. Biophys.* 35, 171–190. doi:10.1385/CBB:35:2:171

Vogel, Z., Barg, J., Levy, R., Saya, D., Heldman, E., and Mechoulam, R. (1993). Anandamide, a brain endogenous compound, interacts specifically with cannabinoid receptors and inhibits adenylate cyclase. *J. Neurochem.* 61, 352–355. doi:10.1111/J.1471-4159.1993.TB03576.X

Wade, D. T., Makela, P., Robson, P., House, H., and Bateman, C. (2004). Do cannabis-based medicinal extracts have general or specific effects on symptoms in multiple sclerosis? A double-blind, randomized, placebo-controlled study on 160 patients. *Mult. Scler.* 10, 434–441. doi:10.1191/1352458504ms10820a

Ward, S. J., McAllister, S. D., Kawamura, R., Murase, R., Neelakantan, H., and Walker, E. A. (2014). Cannabidiol inhibits paclitaxel-induced neuropathic pain through 5-HT 1A receptors without diminishing nervous system function or chemotherapy efficacy. *Br. J. Pharmacol.* 171, 636–645. doi:10.1111/bph.12439

Ware, M. A., Wang, T., Shapiro, S., Robinson, A., Ducruet, T., Huynh, T., et al. (2010). Smoked cannabis for chronic neuropathic pain: A randomized controlled trial. *C. Can. Med. Assoc. J.* 182, E694–E701. doi:10.1503/CMAJ.091414

Yu, F. H., Mantegazza, M., Westenbroek, R. E., Robbins, C. A., Kalume, F., Burton, K. A., et al. (2006). Reduced sodium current in GABAergic interneurons in a mouse model of severe myoclonic epilepsy in infancy. *Nat. Neurosci.* 9, 1142–1149. doi:10.1038/nn1754

Zhang, H. X. B., and Bean, B. P. (2021). Cannabidiol inhibition of murine primary nociceptors: Tight binding to slow inactivated states of Nav1.8 channels. J. Neurosci. 41, 6371–6387. doi:10.1523/JNEUROSCI.3216-20.2021

Zhang, H. X. B., Heckman, L., Niday, Z., Jo, S., Fujita, A., Shim, J., et al. (2022). Cannabidiol activates neuronal Kv7 channels. *Elife* 11, e73246. doi:10.7554/ELIFE.73246