



Unraveling Brain Microcircuits, Dendritic Spines, and Synaptic Processing Using Multiple Complementary Approaches

Alberto A. Rasia-Filho^{1,2*}

¹ Department of Basic Sciences/Physiology, Graduate Program in Biosciences, Universidade Federal de Ciências da Saúde de Porto Alegre, Porto Alegre, Brazil, ² Graduate Program in Neuroscience, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil

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*Correspondence:

Alberto A. Rasia-Filho
aarf@ufcspa.edu.br;
rasiafilho@yahoo.com

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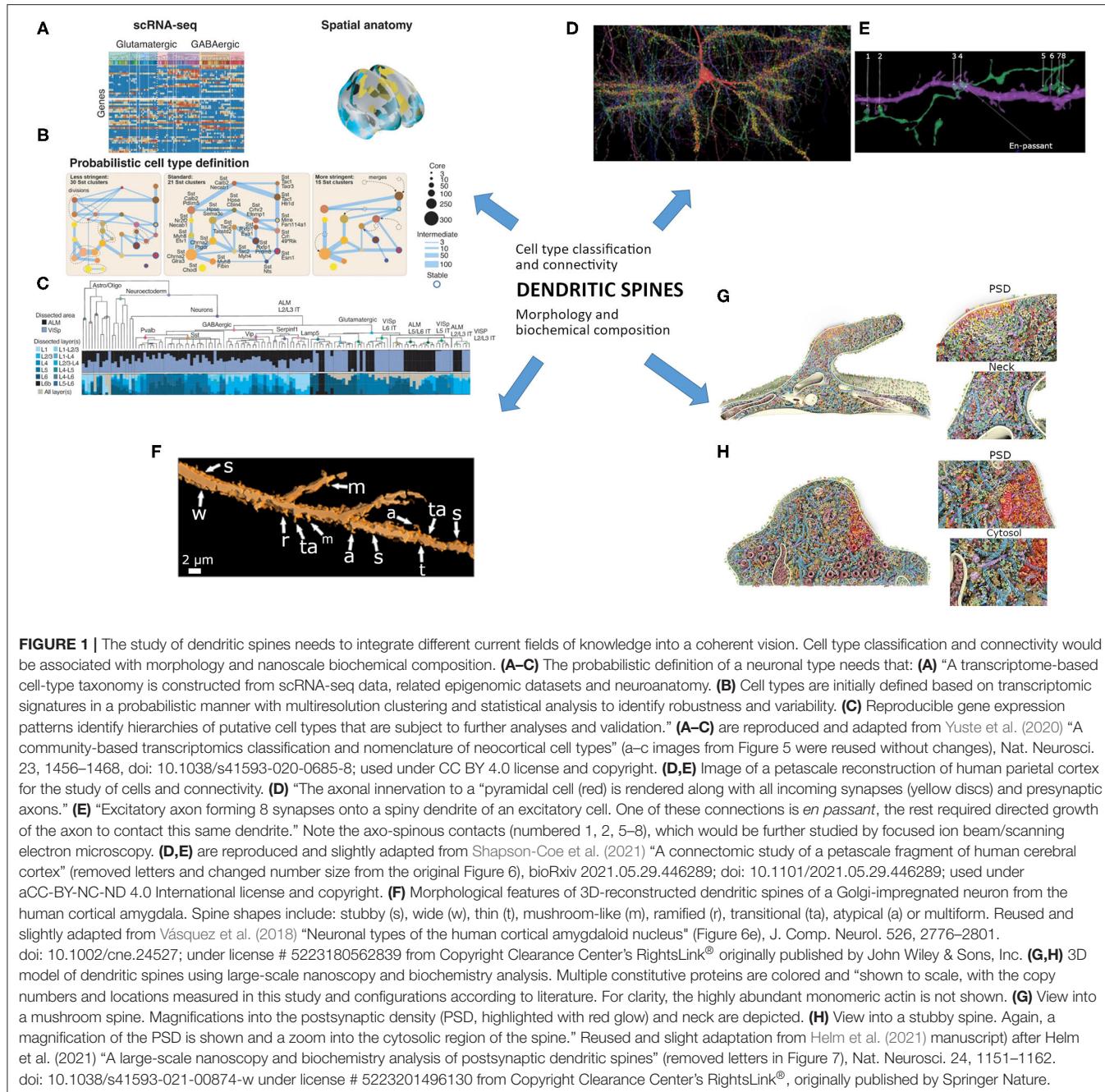
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INTRODUCTION

Innovative experimental approaches and technological advancements have provided an unprecedented level of detail for the nervous system. New findings advanced our knowledge about the complexity of genetic profiles, neuroanatomical and connectional parcellation of cortical areas, and cytoarchitectonic and synaptic organization in humans compared to other species (DeFelipe, 2011; Vogt, 2015; Bruner et al., 2017; Hodge et al., 2019; Assem et al., 2020; Benavides-Piccione et al., 2020; Eze et al., 2021; Girsik et al., 2021; Viscardi et al., 2021). We can now observe neuronal features on a nanoscale level and envisage possible links for cells and circuits when identifying genes, constitutive proteins, subpopulations of neurons, networks with high-speed actions, higher-ordered mental states, and a multitude of disparate behaviors (Fuzik et al., 2016; Turcotte et al., 2019; Hodge et al., 2020; Close et al., 2021; Demas et al., 2021; Helm et al., 2021). Therefore, we need to integrate different fields of knowledge about dendritic spines into a coherent vision of where this field of research is going using new techniques (**Figure 1**).

Dendritic spines are specialized postsynaptic compartments (Yuste, 2010; Stewart et al., 2014; Helm et al., 2021) for monosynaptic or, in some cases, multisynaptic inputs (Brusco et al., 2014; Dall’Oglio et al., 2015). Spines are significant in that their morphology appears to change with various inputs and brain disorders (Chidambaram et al., 2019; Baczyńska et al., 2021). Understanding their role in synaptic connectivity is a fruitful approach to elucidating relationships among connectivity of different cellular structures in the brain. However, the study of spines represents a huge challenge when considering neural networks that show region- and cell type-specific characteristics and differences within and among species [Lavenex et al., 2009; Cembrowski and Spruston, 2019; Hodge et al., 2019; Winnubst et al., 2019; Yang et al., 2019; BRAIN Initiative Cell Census Network (BICCN), 2021]. Complementary approaches to the structure-function relationship of the components of an inherently intricate system as the brain are needed. For example, *in vitro* and *in vivo* studies demonstrate the high level of structural and functional complexity of the cerebral cortex (Schepersjans et al., 2008; Rigotti et al., 2013; Vogt, 2015; Glasser et al., 2016; Real et al., 2018; Leopold et al., 2019; Assem et al., 2020; Finkelstein et al., 2021; Foster et al., 2021). In both allocortex and neocortex, morphological heterogeneity within classical



cell types can be the rule as occur for the spiny pyramidal neurons (Morishima and Kawaguchi, 2006; Ramaswamy and Markram, 2015; Cembrowski and Spruston, 2019; Benavides-Piccione et al., 2020; Rasia-Filho et al., 2021). Indeed, discrete and continuous variations may coexist and underlie cell-type diversity, forming a "combination of specification through evolutionarily driven and developmentally regulated genetic mechanisms, and refinement of cellular identities through

intercellular interactions within the network in which the cells are embedded" [BRAIN Initiative Cell Census Network (BICCN), 2021]. Henceforth, the probabilistic definition of each neuronal type will require single-cell transcriptomic data associated to morphology (Figure 1A; Hodge et al., 2019; Yuste et al., 2020). Dendritic spines may be further studied in specific subpopulations of neurons and circuits to address their functional roles in information processing (Figure 1B).

BRAIN NETWORKS, CELLULAR CONNECTIVITY, AND THE RELEVANCE OF DENDRITIC SPINES

Recent approaches advanced the study of brain cells, microcircuits, and connections. The connectomic study of a fragment of the human temporal cortex (1 mm^3 , $>5,000$ slices cut at $\sim 30 \text{ nm}$), imaged using a high-speed multi-beam scanning electron microscopy (EM) and three-dimensional (3D) reconstruction, exhibited 57,216 cells and ~ 133 million synapses in a 1.4-petabyte volume (Shapson-Coe et al., 2021). Dense digital reconstruction of a 0.3 mm^3 cortical circuit containing $\sim 31,000$ neurons, ~ 37 million excitatory and inhibitory synapses, and 55 morphological cell types served to identify hub neurons that could modulate cortical dynamics (Gal et al., 2021). Additional highly multiplexed, high-resolution brain-wide cell-type mapping, and high-throughput spatially resolved transcriptomics approaches can link cell types with connectivity mapping and functional data (Close et al., 2021) for advanced molecular neuroanatomical maps (Ortiz et al., 2021). Some techniques may link functional data with different spatial scales. For example, patch-clamp electrophysiology and single-cell semi-quantitative PCR would identify neuronal subtypes (Fuzik et al., 2016). On the other hand, high-resolution magnetic resonance imaging (MRI) would locate different nuclei in the brain (Saygin et al., 2017 for human amygdala) and help to identify likely borders for each area of interest (e.g., to separate the medial and cortical amygdaloid nuclei, Dall'Oglio et al., 2013; Vásquez et al., 2018). These data are relevant for understanding the complex expression of emotion in different species (Quirk et al., 1995; Zebarjadi et al., 2021; including mice affiliative touch in prosocial interaction, Wu et al., 2021) and what feelings are to humans (Zeki, 2007; Gendron and Barrett, 2009; de Boer et al., 2012; Diano et al., 2017; LeDoux and Brown, 2017; Fogazzi et al., 2020; Šimić et al., 2021).

To process manifold stimuli from external and internal milieux engenders specialization and functional integration of neural areas, cells, and networks (e.g., Rasia-Filho, 2006; Rasia-Filho et al., 2018; Freiwald, 2020; Barnett et al., 2021). Dendritic spine function comprises an important part of this complex scenario (Ramón y Cajal, 1909–1911; Bourne and Harris, 2009; von Bohlen und Halbach, 2009; Yuste, 2010; Spruston et al., 2013; Dall'Oglio et al., 2015; Helm et al., 2021). That is, spines increase the connectivity between neurons and the packing density of synapses without increasing the brain's overall volume (Bourne and Harris, 2009). This feature adds and maximizes the connectivity repertoire governing the shape of dendritic arbors (Wen et al., 2009). Dendritic spines modulate the excitatory synaptic transmission in the brain. The majority of input contacts on dendritic spines are from glutamatergic axon terminals (Yuste, 2013; but see also GABAergic and dopaminergic innervation in Brusco et al., 2014; Kubota et al., 2016; Iino et al., 2020; Kasai et al., 2021). Spines are morphologically diverse, ranging in a *continuum* of number, shape, and size classified according to their head and neck features (Figure 1C). These include: stubby/wide, thin, mushroom, ramified, “atypical”

or multiform spines, including “intermediate” shapes, “double” spines, and thorny excrescences, among others (Fiala and Harris, 1999; Arellano et al., 2007; Bourne and Harris, 2007, 2009; Stewart et al., 2014; Fuentealba-Villarroel et al., 2021; see also Ruszczycki et al., 2012; Pchitskaya and Bezprozvanny, 2020). Spine shape involves local actin organization, second messengers, and organelles (e.g., endoplasmic reticulum and ribosomes, Yuste, 2010; Sala and Segal, 2014; Miermans et al., 2017; Okabe, 2020; for mitochondria see Li et al., 2004). This can lead to biochemical compartmentalization and affect the electrical signaling of synapses (Chen and Sabatini, 2012; Tønnesen and Nägerl, 2016; Obashi et al., 2021). The balance between spine number, structure, and function may represent synaptic processing for learning and memory (Bourne and Harris, 2007, 2009) with stimulus-specific features (Knafo et al., 2005) in selective synaptic ensembles (Hayashi-Takagi et al., 2015). Optogenetic manipulation allowed the identification and erasure of specific synaptic memory traces in potentiated spines of the mouse motor cortex (Hayashi-Takagi et al., 2015). This was a remarkable achievement since the functional mapping of single-spine synaptic inputs to the same dendrite can be highly heterogeneous, as revealed by high-resolution two-photon imaging of auditory-evoked NMDA-dependent calcium transients in mouse cortical neurons *in vivo* (Chen et al., 2011).

Dendritic spine dynamics in different neural circuits result from various phenomena. These include phylogenetic, ontogenetic, and epigenetic events (García-López et al., 2010; DeFelipe, 2011; Reza-Zaldivar et al., 2020). Activity-dependent and activity-independent actions promote stabilization, differentiation, and remodeling with enlargement or shrinkage and pruning of spines (Oray et al., 2006; Zancan et al., 2018; Runge et al., 2020; Kasai et al., 2021). Spines can be found relatively isolated or in clusters in the same dendritic segments, as evidenced after 3D image reconstruction of Golgi-impregnated neurons in humans from our laboratory (Reberger et al., 2018; Rasia-Filho et al., 2021) and other approaches using transmission EM (Arellano et al., 2007; Bourne and Harris, 2009; Brusco et al., 2014; Stewart et al., 2014), rapid structured illumination microscopy and enhanced resolution confocal microscopy (for spinules, Zaccard et al., 2020), high-resolution transmission, focused ion beam (FIB) scanning and EM tomography (Rollenhagen et al., 2020), and/or FIB/scanning EM in humans and other animals (Cano-Astorga et al., 2021). Clustered spines can show spike-timing-dependent cooperativity and plasticity (Tazerart et al., 2020). Therefore, synaptic integration made by each spine type can impact cellular activity differently depending upon its location and spatiotemporal processing along proximal to distal dendritic domains (Spruston et al., 2013). In addition, its passive and/or active biophysical properties associated with those of parent dendrites may play a role (Sala and Segal, 2014; Gidon et al., 2020; Obashi et al., 2021). Dendritic spines modulate both stable and/or transitory connections (Oray et al., 2006) and synaptic plasticity using various molecules in variable biochemical pathways for short-term to long-term cellular effects (Sala and Segal, 2014; Chidambaram et al., 2019).

There are many frontiers to explore the structure and integrated function of dendritic spines for synaptic plasticity. The impact of heterogeneous glial cells and the role of the extracellular matrix in tetrapartite synapses need to be addressed (Chelini et al., 2018; Mederos et al., 2018; Tønnesen et al., 2018; Nguyen et al., 2020; Klimczak et al., 2021). The elucidation of the evolutionary reason for the divergence in gene expression patterns in the cerebral cortex, and the features that determine neuronal diversity and specialization in humans are important (Hodge et al., 2019, 2020; Kalmbach et al., 2021). Regarding the latter, some features of human cortical pyramidal neurons include: (1) larger dendritic length and branch complexity than macaque and mice (Mohan et al., 2015; Benavides-Piccione et al., 2020); (2) a class of calcium-mediated graded dendritic action potentials that would classify linearly non-separable inputs (Gidon et al., 2020); and (3) membrane properties that significantly enhance synaptic charge-transfer from dendrites to soma and spike propagation along the axon (Eyal et al., 2016). This indicates that extrapolations on some neuronal features from other species to the human brain have to be done carefully. Human dendritic spines are systematically larger and longer and exist at higher densities than in the mouse cortex (Benavides-Piccione et al., 2020), likely increasing our capacity of synaptic processing and plasticity (DeFelipe, 2011). Human spines, also, show a high diversity of size and shapes (Dall’Oglio et al., 2015; Vásquez et al., 2018; Rasia-Filho et al., 2021). The functional implication of long “silent” spines (Yuste, 2013) and those of convoluted shapes, observed from subcortical to cortical human neurons (Dall’Oglio et al., 2015; Fuentealba-Villarroel et al., 2021), need additional studies. Multiform spines likely indicate the existence of multisynaptic sites for signaling compartmentalization and further computational possibilities within functional microdomains (Chen and Sabatini, 2012; Dall’Oglio et al., 2015; Reberger et al., 2018).

The postsynaptic density (PSD) is a dense area behind the postsynaptic membrane, as seen by EM. It consists of many proteins, including receptors, ion channels, and adhesion proteins, shared with the membrane, cytoskeletal proteins, and scaffolding proteins, all arranged in a hierarchical fashion (Cohen, 2013). Like dendritic spines, PSDs can display morphological alterations with various physiological and behavioral inputs. PSD area can relate to spine head diameter (Arellano et al., 2007) depending on an NMDA receptor-mediated long-term potentiation plasticity (Borczyk et al., 2019). Stubby and mushroom spines show similar average protein copy number and topology for PSD composition identified after summing EM, stimulated emission depletion microscopy, mass spectrometry, fluorescence microscopy, and 3D reconstruction procedures in cultured hippocampal neurons of rats (Helm et al., 2021; **Figure 1D**). However, proteins related to synaptic strength, spine dynamics, ion channels, endocytosis cofactors, cytoskeletal structure, signaling and trafficking, secretory proteins, and ribosomes are more evident in mushroom spines (Helm et al., 2021). These findings open the possibility to test different spines also in neuropathological conditions (Forrest et al., 2018; Chidambaram et al., 2019; Runge et al., 2020; Baczyńska et al., 2021; Montero-Crespo et al., 2021). From this perspective,

dendritic spines are sexually dimorphic and/or affected by gonadal steroids (Woolley and McEwen, 1993; Rasia-Filho et al., 2012; Luine and Frankfurt, 2020), sexual experience and motherhood (Rasia-Filho et al., 2004; Zancan et al., 2018). These phenomena are relevant to sexual differentiation in healthy brain connectivity and as a biological variable in neuropsychiatric research (Joel and McCarthy, 2017; Rubinow and Schmidt, 2019; Arnold, 2020; Hidalgo-Lopez et al., 2021).

Lastly, the human cerebral cortex shows a highly polygenic architecture (Grasby et al., 2020) and ~16 billion neurons (Herculano-Houzel et al., 2014). One cortical pyramidal neuron can form ~30,000 synapses, 90% of them being excitatory (DeFelipe, 2011). From ~100 trillion spines in the human cortex (Kasai et al., 2021), ~99.5% of all spines lie in pyramidal neurons (Kubota et al., 2016; see also Foggetti et al., 2019) for the organization of the ongoing synaptic transmission from multiple neurochemical circuits (Palomero-Gallagher and Zilles, 2019). This complexity is exemplified by the huge spine density and shape variation in a human CA1 pyramidal neuron related to circuits for memory modulation and self-identity (see Figure 9 in Rasia-Filho et al., 2021). On a spine-by-spine basis (Oray et al., 2006), there can be a high degree of synaptic processing arising from spatiotemporal and functional heterogeneity among individual synapses on the same dendrite, between different neurons, and across and between brain regions (Grant and Fransén, 2020). Synaptic diversity and strength are finely adjusted to code information (Grant and Fransén, 2020), enabling coincidence detection (Chabrol et al., 2015) and merging multimodal inputs from parallel pathways (Soltész and Losonczy, 2018). The integrated synaptic processing and complex plasticity linked to the role of an increasing number of specialized neurons and glia cells within circuits may ultimately lead to the emergence of multiple sensorimotor, cognitive, emotional, abstract, creative and conscious elaborations, visceral reactions, and behavioral displays (for a parallel discussion see Timo-Iaria and Valle, 1995; DeFelipe, 2011; Jezek et al., 2011; Hodge et al., 2019; Freiwald, 2020; Grant and Fransén, 2020; Rasia-Filho et al., 2021).

CONCLUSION

Dendritic spines are key elements for innovative research in integrative physiology. Various approaches can expand our knowledge on spines studying them at both network-scale and synapse-scale in the brain. For example, we still do not know the implications of dendritic spines in different subpopulations of neurons for the cytoarchitectonics, rich networks connections, and complex information processing in the insular cortex. This is an interesting cortical area that is strongly activated when “you see the person you are in love with, try to listen to your own heartbeat, suffer from a headache, or crave for a chocolate cookie” (Gogolla, 2017; see also Benarroch, 2019). As mentioned by Mancuso et al. (2014): “Anatomical changes occur on a full range of scales from the trafficking of individual proteins, to alterations in synaptic morphology both individually and on a systems level, to reductions in long-distance connectivity and brain volume.”

Dendritic spines relate to all these processes in the brain and with a notable integrative complexity in humans. Unraveling the dynamic role of dendritic spines for synaptic processing is a task that needs multiple complementary approaches. The level of complexity for this endeavor resides in the fact we are looking for representative data that is likely on the astonishing scale of 10^{15} ongoing connections in the human brain.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

REFERENCES

- Arellano, J. I., Benavides-Piccione, R., DeFelipe, J., and Yuste, R. (2007). Ultrastructure of dendritic spines: correlation between synaptic and spine morphologies. *Front. Neurosci.* 1:131–143. doi: 10.3389/neuro.01.1.1.010.2007
- Arnold, A. P. (2020). Sexual differentiation of brain and other tissues: five questions for the next 50 years. *Horm. Behav.* 120, 104691. doi: 10.1016/j.yhbeh.2020.104691
- Assem, M., Glasser, M. F., Van Essen, D. C., and Duncan, J. (2020). A domain-general cognitive core defined in multimodally parcellated human cortex. *Cereb. Cortex* 30, 4361–4380. doi: 10.1093/cercor/bhaa023
- Baczyńska, E., Pels, K. K., Basu, S., Włodarczyk, J., and Ruszczycki, B. (2021). Quantification of dendritic spines remodeling under physiological stimuli and in pathological conditions. *Int. J. Mol. Sci.* 22, 4053. doi: 10.3390/ijms22084053
- Barnett, S. C., Parr-Brownlie, L. C., Perry, B. A. L., Young, C. K., Wicky, H. E., Hughes, S. M., et al. (2021). Anterior thalamic nuclei neurons sustain memory. *Curr. Res. Neurobiol.* 2, 100022. doi: 10.1016/j.crneur.2021.100022
- Benarroch, E. E. (2019). Insular cortex: functional complexity and clinical correlations. *Neurology* 93, 932–938. doi: 10.1212/WNL.0000000000008525
- Benavides-Piccione, R., Regaldo-Reyes, M., Fernaud-Espinosa, I., Kastanauksaitė, A., Tapia-González, S., León-Espinosa, G., et al. (2020). Differential structure of hippocampal CA1 pyramidal neurons in the human and mouse. *Cereb. Cortex* 30, 730–752. doi: 10.1093/cercor/bhz122
- Borczyk, M., Sliwińska, M. A., Caly, A., and Bernas, T., Radwanska, K. (2019). Neuronal plasticity affects correlation between the size of dendritic spine and its postsynaptic density. *Sci. Rep.* 9, 1693. doi: 10.1038/s41598-018-38412-7
- Bourne, J., and Harris, K. M. (2007). Do thin spines learn to be mushroom spines that remember? *Curr. Opin. Neurobiol.* 17, 381–386. doi: 10.1016/j.conb.2007.04.009
- Bourne, J. N., and Harris, K. M. (2009). “Ultrastructural analysis of spine plasticity,” in *Encyclopedia of Neuroscience*, ed L. R. Squire (New York, NY: Elsevier), 11–17. doi: 10.1016/B978-008045046-9.01771-X
- BRAIN Initiative Cell Census Network (BICCN). (2021). A multimodal cell census and atlas of the mammalian primary motor cortex. *Nature* 598, 86–102. doi: 10.1038/s41586-021-03950-0
- Bruner, E., Preuss, T. M., Chen, X., and Rilling, J. K. (2017). Evidence for expansion of the precuneus in human evolution. *Brain Struct. Funct.* 222, 1053–1060. doi: 10.1007/s00429-015-1172-y
- Brusco, J., Merlo, S., Ikeda, É. T., Petralia, R. S., Kachar, B., Rasia-Filho, A. A., et al. (2014). Inhibitory and multisynaptic spines, and hemispherical synaptic specialization in the posterodorsal medial amygdala of male and female rats. *J. Comp. Neurol.* 522, 2075–2088. doi: 10.1002/cne.23518
- Cano-Astorga, N., DeFelipe, J., and Alonso-Nanclares, L. (2021). Three-dimensional synaptic organization of layer III of the human temporal neocortex. *Cereb. Cortex* 31, 4742–4764. doi: 10.1093/cercor/bhz120
- Cembrowski, M. S., and Spruston, N. (2019). Heterogeneity within classical cell types is the rule: lessons from hippocampal pyramidal neurons. *Nat. Rev. Neurosci.* 20, 193–204. doi: 10.1038/s41583-019-0125-5
- Chabrol, F. P., Arenz, A., Wiechert, M. T., Margrie, T. W., and DiGregorio, D. A. (2015). Synaptic diversity enables temporal coding of coincident multisensory inputs in single neurons. *Nat. Neurosci.* 18, 718–727. doi: 10.1038/nn.3974
- Chelini, G., Pantazopoulos, H., Durnig, P., and Berretta, S. (2018). The tetrapartite synapse: a key concept in the pathophysiology of schizophrenia. *Eur. Psychiatry* 50, 60–69. doi: 10.1016/j.eurpsy.2018.02.003
- Chen, X., Leischner, U., Rochefort, N. L., Nelken, I., and Konnerth, A. (2011). Functional mapping of single spines in cortical neurons *in vivo*. *Nature* 475, 501–505. doi: 10.1038/nature10193
- Chen, Y., and Sabatini, B. L. (2012). Signaling in dendritic spines and spine microdomains. *Curr. Opin. Neurobiol.* 22, 389–396. doi: 10.1016/j.conb.2012.03.003
- Chidambaram, S. B., Rathipriya, A. G., Bolla, S. R., Bhat, A., Ray, B., Mahalakshmi, A. M., et al. (2019). Dendritic spines: revisiting the physiological role. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 92, 161–193. doi: 10.1016/j.pnpbp.2019.01.005
- Close, J. L., Long, B. R., and Zeng, H. (2021). Spatially resolved transcriptomics in neuroscience. *Nat. Methods* 18, 23–25. doi: 10.1038/s41592-020-01040-z
- Cohen, R. S. (2013). “The postsynaptic density,” in *Neuroscience in the 21st Century*, ed D. W. Pfaff (New York, NY: Springer), 403–437. doi: 10.1007/978-1-4614-1997-6_17
- Dall’Oglio, A., Dutra, A. C., Moreira, J. E., and Rasia-Filho, A. A. (2015). The human medial amygdala: structure, diversity, and complexity of dendritic spines. *J. Anat.* 227, 440–459. doi: 10.1111/joa.12358
- Dall’Oglio, A., Xavier, L. L., Hilbig, A., Ferme, D., Moreira, J. E., Achaval, M., et al. (2013). Cellular components of the human medial amygdaloid nucleus. *J. Comp. Neurol.* 521, 589–611. doi: 10.1002/cne.23192
- de Boer, A., Van Buel, E. M., and Ter Horst, G. J. (2012). Love is more than just a kiss: a neurobiological perspective on love and affection. *Neuroscience* 201, 114–124. doi: 10.1016/j.neuroscience.2011.11.017
- DeFelipe, J. (2011). The evolution of the brain, the human nature of cortical circuits, and intellectual creativity. *Front. Neuroanat.* 5, 29. doi: 10.3389/fnana.2011.00029
- Demas, J., Manley, J., Tejera, F., Barber, K., Kim, H., Traub, F. M., et al. (2021). High-speed, cortex-wide volumetric recording of neuroactivity at cellular resolution using light beads microscopy. *Nat. Methods* 18, 1103–1111. doi: 10.1038/s41592-021-01239-8
- Diano, M., Tamietto, M., Celeghin, A., Weiskrantz, L., Tatu, M. K., Bagnis, A., et al. (2017). Dynamic changes in amygdala psychophysiological connectivity reveal distinct neural networks for facial expressions of basic emotions. *Sci. Rep.* 7, 1–13. doi: 10.1038/srep45260
- Eyal, G., Verhoog, M. B., Testa-Silva, G., Deitcher, Y., Lodder, J. C., Benavides-Piccione, R., et al. (2016). Unique membrane properties and enhanced signal processing in human neocortical neurons. *eLife* 5:e16553. doi: 10.7554/eLife.16553
- Eze, U. C., Bhaduri, A., Haeussler, M., Nowakowski, T. J., and Kriegstein, A. R. (2021). Single-cell atlas of early human brain development highlights heterogeneity of human neuroepithelial cells and early

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- radial glia. *Nat. Neurosci.* 24, 584–594. doi: 10.1038/s41593-020-00794-1
- Fiala, J. C., and Harris, K. M. (1999). “Dendrite structure,” in *Dendrites*, eds G. Stuart, N. Spruston, and M. Häusser (New York, NY: Oxford University Press), 1–34.
- Finkelstein, A., Fontolan, L., Economo, M. N., Li, N., Romani, S., et al. (2021). Attractor dynamics gate cortical information flow during decision-making. *Nat. Neurosci.* 24, 843–850. doi: 10.1038/s41593-021-0840-6
- Fogazzi, D. V., Neary, J. P., Sonza, A., Reppold, C. T., Kaiser, V., Scassola, C. M., et al. (2020). The prefrontal cortex conscious and unconscious response to social/emotional facial expressions involve sex, hemispheric laterality, and selective activation of the central cardiac modulation. *Behav. Brain Res.* 393, 112773. doi: 10.1016/j.bbr.2020.112773
- Foggetti, A., Baccini, G., Arnold, P., Schifffelholz, T., and Wulff, P. (2019). Spiny and non-spiny parvalbumin-positive hippocampal interneurons show different plastic properties. *Cell Rep.* 27, 3725–3732.e5. doi: 10.1016/j.celrep.2019.05.098
- Forrest, M. P., Parnell, E., and Penzes, P. (2018). Dendritic structural plasticity and neuropsychiatric disease. *Nat. Rev. Neurosci.* 19, 215–234. doi: 10.1038/nrn.2018.16
- Foster, N. N., Barry, J., Korobkova, L., Garcia, L., Gao, L., Becerra, M., et al. (2021). The mouse cortico–basal ganglia–thalamic network. *Nature* 598, 188–194. doi: 10.1038/s41586-021-03993-3
- Freiwald, W. A. (2020). Social interaction networks in the primate brain. *Curr. Opin. Neurobiol.* 65, 49–58. doi: 10.1016/j.conb.2020.08.012
- Fuentelba-Villarroel, F. J., Renner, J., Hilbig, A., Bruton, O. J., and Rasia-Filho, A. A. (2021). Spindle-shaped neurons in the human posteromedial (precuneus) cortex. *Front. Synaptic Neurosci.* 13, 769228. doi: 10.3389/fnsyn.2021.769228
- Fuzik, J., Zeisel, A., Máté, Z., Calvignoni, D., Yanagawa, Y., Szabó, G., et al. (2016). Integration of electrophysiological recordings with single-cell RNA-seq data identifies neuronal subtypes. *Nat. Biotech.* 34, 175–183. doi: 10.1038/nbt.3443
- Gal, E., Amsalem, O., Schindel, A., London, M., Schürmann, F., Markram, H., et al. (2021). The role of hub neurons in modulating cortical dynamics. *Front. Neural Circuits* 15, 718270. doi: 10.3389/fncir.2021.718270
- García-López, P., García-Marín, V., and Freire, M. (2010). Dendritic spines and development: towards a unifying model of spinogenesis - a present day review of Cajal's histological slides and drawings. *Neural Plast.* 2010, 769207. doi: 10.1155/2010/769207
- Gendron, M., and Barrett, L. F. (2009). Reconstructing the past: A century of ideas about emotion in psychology. *Emotion Rev.* 1, 316–339. doi: 10.1177/1754073909338877
- Gidon, A., Zolnik, T. A., Fidzinski, P., Bolduan, F., Papoutsi, A., Poirazi, P., et al. (2020). Dendritic action potentials and computation in human layer 2/3 cortical neurons. *Science* 367, 83–87. doi: 10.1126/science.aax6239
- Girkis, K. M., Stergachis, A. B., DeGennaro, E. M., Doan, R. N., Qian, X., Johnson, M. B., et al. (2021). Rewiring of human neurodevelopmental gene regulatory programs by human accelerated regions. *Neuron* 109, 239–325.e7. doi: 10.1016/j.neuron.2021.08.005
- Glasser, M. F., Coalson, T. S., Robinson, E. C., Hacker, C. D., Harwell, J., Yacoub, E., et al. (2016). A multi-modal parcellation of human cerebral cortex. *Nature* 536, 171–178. doi: 10.1038/nature18933
- Gogolla, N. (2017). The insular cortex. *Curr. Biol.* 27, R580–586. doi: 10.1016/j.cub.2017.05.010
- Grant, S. G., and Fransén, E. (2020). The synapse diversity dilemma: molecular heterogeneity confounds studies of synapse function. *Front. Synaptic Neurosci.* 12:590403. doi: 10.3389/fnsyn.2020.590403
- Grasby, K. L., Jahanshad, N., Painter, J. N., Colodro-Conde, L., Bralten, J., Hibar, D. P., et al. (2020). The genetic architecture of the human cerebral cortex. *Science* 367, eaay6690. doi: 10.1126/science.aya6690
- Hayashi-Takagi, A., Yagishita, S., Nakamura, M., Shirai, F., Wu, Y. I., Loshbaugh, A. L., et al. (2015). Labeling and optical erasure of synaptic memory traces in the motor cortex. *Nature* 525, 333–338. doi: 10.1038/nature15257
- Helm, M. S., Dankovich, T. M., Mandad, S., Rammner, B., Jähne, S., Salimi, V., et al. (2021). A large-scale nanoscopy and biochemistry analysis of postsynaptic dendritic spines. *Nat. Neurosci.* 24, 1151–1162. doi: 10.1038/s41593-021-00874-w
- Herculano-Houzel, S., Manger, P. R., and Kaas, J. H. (2014). Brain scaling in mammalian evolution as a consequence of concerted and mosaic changes in numbers of neurons and average neuronal cell size. *Front. Neuroanat.* 8:77. doi: 10.3389/fnana.2014.00007
- Hidalgo-López, E., Zeidman, P., Harris, T., Razi, A., and Pletzer, B. (2021). Spectral dynamic causal modeling in healthy women reveals brain connectivity changes along the menstrual cycle. *Commun. Biol.* 4:954. doi: 10.1038/s42003-021-02447-w
- Hodge, R. D., Bakken, T. E., Miller, J. A., Smith, K. A., Barkan, E. R., Graybuck, L. T., et al. (2019). Conserved cell types with divergent features in human versus mouse cortex. *Nature* 573, 61–68. doi: 10.1038/s41586-019-1506-7
- Hodge, R. D., Miller, J. A., Novotny, M., Kalmbach, B. E., Ting, J. T., Bakken, T. E., et al. (2020). Transcriptomic evidence that von Economo neurons are regionally specialized extratelencephalic-projecting excitatory neurons. *Nat. Commun.* 11, 1172. doi: 10.1038/s41467-020-14952-3
- Iino, Y., Sawada, T., Yamaguchi, K., Tajiri, M., Ishii, S., Kasai, H., et al. (2020). Dopamine D2 receptors in discrimination learning and spine enlargement. *Nature* 579, 555–560. doi: 10.1038/s41586-020-2115-1
- Jezek, K., Henriksen, E. J., Treves, A., Moser, E. I., and Moser, M.-B. (2011). Theta-paced flickering between place-cell maps in the hippocampus. *Nature* 478, 246–249. doi: 10.1038/nature10439
- Joel, D., and McCarthy, M. M. (2017). Incorporating sex as a biological variable in neuropsychiatric research: where are we now and where should we be? *Neuropsychopharmacology* 42, 379–385. doi: 10.1038/npp.2016.79
- Kalmbach, B. E., Hodge, R. D., Jorstad, N. L., Owen, S., de Frates, R., Yanny, A. M., et al. (2021). Signature morpho-electric, transcriptomic, and dendritic properties of human layer 5 neocortical pyramidal neurons. *Neuron* 109, 2914–2927.e5. doi: 10.1016/j.neuron.2021.08.030
- Kasai, H., Ziv, N. E., Okazaki, H., Yagishita, S., and Toyoizumi, T. (2021). Spine dynamics in the brain, mental disorders and artificial neural networks. *Nat. Rev. Neurosci.* 22, 407–422. doi: 10.1038/s41583-021-00467-3
- Klimczak, P., Rizzo, A., Castillo-Gómez, E., Perez-Rando, M., Gramuntell, Y., Beltran, M., et al. (2021). Parvalbumin interneurons and perineuronal nets in the hippocampus and retrosplenial cortex of adult male mice after early social isolation stress and perinatal NMDA receptor antagonist treatment. *Front. Synaptic Neurosci.* 13:733989. doi: 10.3389/fnsyn.2021.733989
- Knafo, S., Libersat, F., and Barkai, E. (2005). Olfactory learning-induced morphological modifications in single dendritic spines of young rats. *Eur. J. Neurosci.* 21, 2217–2226. doi: 10.1111/j.1460-9568.2005.04041.x
- Kubota, Y., Karube, F., Nomura, M., and Kawaguchi, Y. (2016). The diversity of cortical inhibitory synapses. *Front. Neural Circuits* 10:27. doi: 10.3389/fncir.2016.00027
- Lavenex, P., Lavenex, P. B., Bennett, J. L., and Amaral, D. G. (2009). Postmortem changes in the neuroanatomical characteristics of the primate brain: hippocampal formation. *J. Comp. Neurol.* 512, 27–51. doi: 10.1002/cne.21906
- LeDoux, J. E., and Brown, R. (2017). Emotions as higher-order states of consciousness. *Proc. Natl. Acad. Sci. USA* 114, E2016–E2025. doi: 10.1073/pnas.1619316114
- Leopold, D. A., Strick, P. L., Bassett, D. S., Bruno, R. M., Cuntz, H., Harris, K. M., et al. (2019). “Functional architecture of the cerebral cortex,” in *The Neocortex*, ed W. Singer (Cambridge: MA: MIT Press), 141–164.
- Li, Z., Okamoto, K., Hayashi, Y., and Sheng, M. (2004). The importance of dendritic mitochondria in the morphogenesis and plasticity of spines and synapses. *Cell* 119, 873–887. doi: 10.1016/j.cell.2004.11.003
- Luine, V., and Frankfurt, M. (2020). Estrogenic regulation of memory: the first 50 years. *Horm. Behav.* 121:104711. doi: 10.1016/j.yhbeh.2020.104711
- Mancuso, J. J., Cheng, J., Yin, Z., Gilliam, J. C., Xia, X., Li, X., et al. (2014). Integration of multi scale dendritic spine structure and function data into systems biology models. *Front. Neuroanat.* 8:130. doi: 10.3389/fnana.2014.00130
- Mederos, S., González-Arias, C., and Perea, G. (2018). Astrocyte–neuron networks: a multilane highway of signaling for homeostatic brain function. *Front. Synaptic Neurosci.* 10:45. doi: 10.3389/fnsyn.2018.00045
- Miermans, C. A., Kusters, R. P., Hoogenraad, C. C., and Storm, C. (2017). Biophysical model of the role of actin remodeling on dendritic spine morphology. *PLoS ONE* 12, e0170113. doi: 10.1371/journal.pone.0170113
- Mohan, H., Verhoog, M. B., Doreswamy, K. K., Eyal, G., Aardse, R., Lodder, B. N., et al. (2015). Dendritic and axonal architecture of individual pyramidal

- neurons across layers of adult human neocortex. *Cereb. Cortex* 25, 4839–4853. doi: 10.1093/cercor/bhv188
- Montero-Crespo, M., Domínguez-Álvaro, M., Alonso-Nanclares, L., DeFelipe, J., and Blazquez-Llorca, L. (2021). Three-dimensional analysis of synaptic organization in the hippocampal CA1 field in Alzheimer's disease. *Brain* 144, 553–573. doi: 10.1093/brain/awaa406
- Morishima, M., and Kawaguchi, Y. (2006). Recurrent connection patterns of corticostriatal pyramidal cells in frontal cortex. *J. Neurosci.* 26, 4394–4405. doi: 10.1523/JNEUROSCI.0252-06.2006
- Nguyen, P. T., Dorman, L. C., Pan, S., Vainchtein, I. D., Han, R. T., Nakao-Inoue, H., et al. (2020). Microglial remodeling of the extracellular matrix promotes synapse plasticity. *Cell* 182, 388–403.e15. doi: 10.1016/j.cell.2020.05.050
- Obashi, K., Taraska, J. W., and Okabe, S. (2021). The role of molecular diffusion within dendritic spines in synaptic function. *J. Gen. Physiol.* 153, e202012814. doi: 10.1085/jgp.202012814
- Okabe, S. (2020). Regulation of actin dynamics in dendritic spines: nanostructure, molecular mobility, and signaling mechanisms. *Mol. Cell. Neurosci.* 109, 103564. doi: 10.1016/j.mcn.2020.103564
- Oray, S., Majewska, A., and Sur, M. (2006). Effects of synaptic activity on dendritic spine motility of developing cortical layer V pyramidal neurons. *Cereb. Cortex* 16, 730–741. doi: 10.1093/cercor/bhj019
- Ortiz, C., Carlén, M., and Meletis, K. (2021). Spatial transcriptomics: molecular maps of the mammalian brain. *Ann. Rev. Neurosci.* 44, 547–562. doi: 10.1146/annurev-neuro-100520-082639
- Palomero-Gallagher, N., and Zilles, K. (2019). Cortical layers: cyto-, myelo-, receptor- and synaptic architecture in human cortical areas. *NeuroImage* 197, 716–741. doi: 10.1016/j.neuroimage.2017.08.035
- Pchitskaya, E., and Bezprozvanny, I. (2020). Dendritic spines shape analysis – classification or clusterization? *Perspective. Front. Synaptic Neurosci.* 12, 31. doi: 10.3389/fnsyn.2020.00031
- Quirk, G. J., Repa, J. C., and LeDoux, J. E. (1995). Fear conditioning enhances short-latency auditory responses of lateral amygdala neurons: parallel recordings in the freely behaving rat. *Neuron* 15, 1029–1039. doi: 10.1016/0896-6273(95)90092-6
- Ramaswamy, S., and Markram, H. (2015). Anatomy and physiology of the thick-tufted layer 5 pyramidal neuron. *Front. Cell. Neurosci.* 9:233. doi: 10.3389/fncel.2015.00233
- Ramón y Cajal (1909–1911). *Histologie du système nerveux de l'homme et des vertébrés*, Paris: Maloine. doi: 10.5962/bhl.title.48637
- Rasia-Filho, A. A. (2006). Is there anything “autonomous” in the nervous system? *Adv. Physiol. Educ.* 30, 9–12. doi: 10.1152/advan.00022.2005
- Rasia-Filho, A. A., Andrejew, R., and Belló-Klein, A. (2018). “Integrating concepts of resilience from cellular functioning to human behavior,” in *Amygdala: Mechanisms, Structure and Role in Disease*, ed. A. Manu (Hauppauge: Nova Science Publishers), 1–30.
- Rasia-Filho, A. A., Dalpian, F., Menezes, I. C., Brusco, J., and Moreira, J. E., Cohen, R. S. (2012). Dendritic spines of the medial amygdala: plasticity, density, shape, and subcellular modulation by sex steroids. *Histol. Histopathol.* 8, 985–1011. doi: 10.14670/HH-27.985
- Rasia-Filho, A. A., Fabian, C., Rigoti, K. M., and Achaval, M. (2004). Influence of sex, estrous cycle and motherhood on dendritic spine density in the rat medial amygdala revealed by the Golgi method. *Neuroscience* 126, 839–847. doi: 10.1016/j.neuroscience.2004.04.009
- Rasia-Filho, A. A., Guerra, K. T. K., Vásquez, C. E., Dall'Oglio, A., Reberger, R., Jung, C. R., et al. (2021). The subcortical-allocortical-neocortical continuum for the emergence and morphological heterogeneity of pyramidal neurons in the human brain. *Front. Synaptic Neurosci.* 13:616607. doi: 10.3389/fnsyn.2021.616607
- Real, R., Peter, M., Trabalza, A., Khan, S., Smith, M. A., Dopp, J., et al. (2018). *In vivo* modeling of human neuron dynamics and Down syndrome. *Science* 362, eaau1810. doi: 10.1126/science.aau1810
- Reberger, R., DallOglio, A., Jung, C. R., and Rasia-Filho, A. A. (2018). Structure and diversity of human dendritic spines evidenced by a new three-dimensional reconstruction procedure for Golgi staining and light microscopy. *J. Neurosci. Methods* 293, 27–36. doi: 10.1016/j.jneumeth.2017.09.001
- Reza-Zaldivar, E. E., Hernández-Sápiens, M. A., Minjarez, B., Gómez-Pinedo, U., Sánchez-González, V. J., Márquez-Aguirre, A. L., et al. (2020). Dendritic spine and synaptic plasticity in Alzheimer's disease: a focus on microRNA. *Front. Cell Dev. Biol.* 8:255. doi: 10.3389/fcell.2020.00255
- Rigotti, M., Barak, O., Warden, M., Wang, X.-J., Daw, N. D., Miller, E. K., et al. (2013). The importance of mixed selectivity in complex cognitive tasks. *Nature* 497, 585–590. doi: 10.1038/nature12160
- Rollenhagen, A., Walkenfort, B., Yakoubi, R., Klauke, S. A., Schmuhl-Giesen, S. F., Heinen-Weiler, J., et al. (2020). Synaptic organization of the human temporal lobe neocortex as revealed by high-resolution transmission, focused ion beam scanning, and electron microscopic tomography. *Int. J. Mol. Sci.* 21, 5558. doi: 10.3390/ijms21155558
- Rubinow, D. R., and Schmidt, P. J. (2019). Sex differences and the neurobiology of affective disorders. *Neuropsychopharmacol.* 44, 111–128. doi: 10.1038/s41386-018-0148-z
- Runge, K., Cardoso, C., and de Chevigny, A. (2020). Dendritic spine plasticity: function and mechanisms. *Front. Synaptic Neurosci.* 12, 36. doi: 10.3389/fnsyn.2020.00036
- Ruszczyczyk, B., Szepesi, Z., Wilczynski, G. M., Bijata, M., Kalita, K., Kaczmarek, L., et al. (2012). Sampling issues in quantitative analysis of dendritic spines morphology. *BMC Bioinformatics* 13, 213. doi: 10.1186/1471-2105-13-213
- Sala, C., and Segal, M. (2014). Dendritic spines: the locus of structural and functional plasticity. *Physiol. Rev.* 94, 141–188. doi: 10.1152/physrev.00012.2013
- Saygin, Z. M., Kliemann, D., Iglesias, J. E., van der Kouwe, A. J. W., Boyd, E., Reuter, M., et al. (2017). High-resolution magnetic resonance imaging reveals nuclei of the human amygdala: manual segmentation to automatic atlas. *NeuroImage* 155, 370–382. doi: 10.1016/j.neuroimage.2017.04.046
- Scheperjans, F., Eickhoff, S. B., Hömke, L., Mohlberg, H., Hermann, K., Amunts, K., et al. (2008). Probabilistic maps, morphometry, and variability of cytoarchitectonic areas in the human superior parietal cortex. *Cereb. Cortex* 18, 2141–2157. doi: 10.1093/cercor/bhm241
- Shapson-Coe, A., Januszewski, M., Berger, D. R., Pope, A., Wu, Y., Blakely, T., et al. (2021). A connectomic study of a petascale fragment of human cerebral cortex. *bioRxiv*. 2021.05.29.446289. doi: 10.1101/2021.05.29.446289
- Šimić, G., Tkalcic, M., Vukić, V., Mulc, D., Španić, E., Šagud, M., et al. (2021). Understanding emotions: Origins and roles of the amygdala. *Biomolecules* 11, 823. doi: 10.3390/biom11060823
- Soltész, I., and Losonczy, A. (2018). CA1 pyramidal cell diversity enabling parallel information processing in the hippocampus. *Nat. Neurosci.* 21, 484–493. doi: 10.1038/s41593-018-0118-0
- Spruston, N., Häusser, M., and Stuart, G. (2013). “Information processing in dendrites and spines,” in *Fundamental Neuroscience*, eds L. R. Squire, D. Berg, F. E. Bloom, S. du Lac, A. Ghosh, and N. C. Spitzer, (Waltham: Academic Press), 231–260. doi: 10.1016/B978-0-12-385870-2.00011-1
- Stewart, M. G., Popov, V. I., Kraev, I. V., Medvedev, N., and Davies, H. A. (2014). “Structure and complexity of the synapse and dendritic spine,” in *The Synapse*, eds V. Pickel and M. Segal (New York, NY: Academic Press), 1–20. doi: 10.1016/B978-0-12-418675-0.00001-8
- Tazerart, S., Mitchell, D. E., Miranda-Rottmann, S., and Araya, R. (2020). A spike-timing-dependent plasticity rule for dendritic spines. *Nat. Commun.* 11:4276. doi: 10.1038/s41467-020-17861-7
- Timo-Iaria, C., and Valle, A. C. (1995). The functional role of the conscious process. *Ciencia e Cultura J. Braz. Assoc. Adv. Sci.* 47, 221–234.
- Tønnesen, J., Inavalli, V. V. G. K., and Nägerl, U. V. (2018). Super-Resolution Imaging of the extracellular space in living brain tissue. *Cell* 172, 1108–1121.e15. doi: 10.1016/j.cell.2018.02.007
- Tønnesen, J., and Nägerl, U. V. (2016). Dendritic spines as tunable regulators of synaptic signals. *Front. Psychol.* 7, 101. doi: 10.3389/fpsyg.2016.00101
- Turcotte, R., Liang, Y., Tanimoto, M., Zhang, Q., Li, Z., Koyama, M., et al. (2019). Dynamic super-resolution structured illumination imaging in the living brain. *Proc. Natl. Acad. Sci. USA* 116, 9586–9591. doi: 10.1073/pnas.1819965116
- Vásquez, C. E., Reberger, R., Dall'Oglio, A., Calcagnotto, M. E., and Rasia-Filho, A. A. (2018). Neuronal types of the human cortical amygdaloid nucleus. *J. Comp. Neurol.* 526, 2776–2801. doi: 10.1002/cne.24527
- Viscardi, L. H., Imparato, D. O., Bortolini, M. C., and Dalmolin, R. J. S. (2021). Ionotropic receptors as a driving force behind human synapse establishment. *Mol. Biol. Evol.* 38, 735–744. doi: 10.1093/molbev/msaa252

- Vogt, B. A. (2015). "Mapping cingulate subregions," in *Brain Mapping: An Encyclopedic Reference*, ed A. W. Toga, (Oxford: Academic Press), 325–339. doi: 10.1016/B978-0-12-397025-1.00230-X
- von Bohlen und Halbach, O. (2009). Structure and function of dendritic spines within the hippocampus. *Ann. Anat.* 191, 518–531. doi: 10.1016/j.aanat.2009.08.006
- Wen, Q., Stepanyants, A., Elston, G. N., Grosberg, A. Y., and Chklovskii, D. B. (2009). Maximization of the connectivity repertoire as a statistical principle governing the shapes of dendritic arbors. *Proc. Natl. Acad. Sci. USA* 106, 12536–12541. doi: 10.1073/pnas.0901530106
- Winnubst, J., Bas, E., Ferreira, T. A., Wu, Z., Economo, M. N., Edson, P., et al. (2019)... Reconstruction of 1,000 projection neurons reveals new cell types and organization of long-range connectivity in the mouse brain. *Cell* 179, 268–281.e13. doi: 10.1016/j.cell.2019.07.042
- Woolley, C. S., and McEwen, B. S. (1993). Roles of estradiol and progesterone in regulation of hippocampal dendritic spine density during the estrous cycle in the rat. *J. Comp. Neurol.* 336, 293–306. doi: 10.1002/cne.903360210
- Wu, Y. E., Dang, J., Kingsbury, L., Zhang, M., Sun, F., Hu, R. K., et al. (2021). Neural control of affiliative touch in prosocial interaction. *Nature* 599, 262–267. doi: 10.1038/s41586-021-03962-w
- Yang, L., Yang, Y., Yuan, J., Sun, Y., Dai, J., and Su, B. (2019). Transcriptomic landscape of von Economo neurons in human anterior cingulate cortex revealed by microdissected-cell RNA sequencing. *Cereb. Cortex* 29, 838–851. doi: 10.1093/cercor/bhy286
- Yuste, R. (2010). *Dendritic Spines*. Cambridge: MIT Press. doi: 10.7551/mitpress/9780262013505.001.0001
- Yuste, R. (2013). Electrical compartmentalization in dendritic spines. *Annu. Rev. Neurosci.* 36, 429–449. doi: 10.1146/annurev-neuro-062111-150455
- Yuste, R., Hawrylycz, M., Aalling, N., Aguilar-Valles, A., Arendt, D., Armañanzas, R., et al. (2020). A community-based transcriptomics classification and nomenclature of neocortical cell types. *Nat. Neurosci.* 23, 1456–1468. doi: 10.1038/s41593-020-0685-8
- Zaccard, C. R., Shapiro, L., Martin-de-Saavedra, M. D., Pratt, C., Myczek, K., Song, A., et al. (2020). Rapid 3D enhanced resolution microscopy reveals diversity in dendritic spine dynamics, regulation, and function. *Neuron* 107, 522–537. doi: 10.1016/j.neuron.2020.04.025
- Zancan, M., da Cunha, R. S. R., Schroeder, F., Xavier, L. L., and Rasia-Filho, A. A. (2018). Remodeling of the number and structure of dendritic spines in the medial amygdala: from prepupal sexual dimorphism to puberty and effect of sexual experience in male rats. *Eur. J. Neurosci.* 48, 1851–1865. doi: 10.1111/ejn.14052
- Zebarjadi, N., Adler, E., Kluge, A., Jääskeläinen, I. P., Sams, M., and Levy, J. (2021). Rhythmic neural patterns during empathy to vicarious pain: beyond the affective-cognitive empathy dichotomy. *Front. Hum. Neurosci.* 15:708107. doi: 10.3389/fnhum.2021.708107
- Zeki, S. (2007). The neurobiology of love. *FEBS Lett.* 581, 2575–2579. doi: 10.1016/j.febslet.2007.03.094

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