



OPEN ACCESS

EDITED BY

Johannes W. Von den Hoff,
Radboud University Medical Centre,
Netherlands

REVIEWED BY

Emily L. Durham,
Children's Hospital of Philadelphia,
United States
Jeremie Oliver Piña,
National Institutes of Health (NIH),
United States

*CORRESPONDENCE

Ana Prates Soares,
✉ ana.prates-soares@charite.de

SPECIALTY SECTION

This article was submitted to Craniofacial
Biology and Dental Research,
a section of the journal
Frontiers in Physiology

RECEIVED 27 January 2023

ACCEPTED 07 March 2023

PUBLISHED 17 March 2023

CITATION

Soares AP, Fischer H, Aydin S, Steffen C,
Schmidt-Bleek K and Rendenbach C
(2023), Uncovering the unique
characteristics of the mandible to
improve clinical approaches to
mandibular regeneration.
Front. Physiol. 14:1152301.
doi: 10.3389/fphys.2023.1152301

COPYRIGHT

© 2023 Soares, Fischer, Aydin, Steffen,
Schmidt-Bleek and Rendenbach. This is
an open-access article distributed under
the terms of the [Creative Commons
Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use,
distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication
in this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Uncovering the unique characteristics of the mandible to improve clinical approaches to mandibular regeneration

Ana Prates Soares^{1,2*}, Heilwig Fischer^{1,3,4}, Sabrin Aydin¹,
Claudius Steffen¹, Katharina Schmidt-Bleek^{2,5} and
Carsten Rendenbach¹

¹Department of Oral and Maxillofacial Surgery, Charité—Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin, and Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany, ²Julius Wolff Institute for Biomechanics and Musculoskeletal Regeneration, Berlin Institute of Health at Charité—Universitätsmedizin Berlin, Berlin, Germany, ³Centrum für Muskuloskeletale Chirurgie, Charité—Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin, and Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany, ⁴BIH Biomedical Innovation Academy, BIH Charité Clinician Scientist Program, Berlin Institute of Health at Charité—Universitätsmedizin Berlin, Berlin, Germany, ⁵Berlin Institute of Health Centre for Regenerative Therapies (BCRT), Berlin Institute of Health at Charité—Universitätsmedizin Berlin, Berlin, Germany

The mandible (lower jaw) bone is aesthetically responsible for shaping the lower face, physiologically in charge of the masticatory movements, and phonetically accountable for the articulation of different phonemes. Thus, pathologies that result in great damage to the mandible severely impact the lives of patients. Mandibular reconstruction techniques are mainly based on the use of flaps, most notably free vascularized fibula flaps. However, the mandible is a craniofacial bone with unique characteristics. Its morphogenesis, morphology, physiology, biomechanics, genetic profile, and osteoimmune environment are different from any other non-craniofacial bone. This fact is especially important to consider during mandibular reconstruction, as all these differences result in unique clinical traits of the mandible that can impact the results of jaw reconstructions. Furthermore, overall changes in the mandible and the flap post-reconstruction may be dissimilar, and the replacement process of the bone graft tissue during healing can take years, which in some cases can result in postsurgical complications. Therefore, the present review highlights the uniqueness of the jaw and how this factor can influence the outcome of its reconstruction while using an exemplary clinical case of pseudoarthrosis in a free vascularized fibula flap.

KEYWORDS

mandible, morphophysiology, free tissue flap, mandibular reconstruction, morphogenesis, bone regeneration

1 Introduction

Various pathologies can affect mandibular health, such as congenital deformities, cysts, tumors, infections, necrosis, as well as trauma (Slootweg, 2010; Pickrell et al., 2017). In large tumors or osteonecrosis of the jaws, a significant portion of the mandible may need to be surgically resected. The resulting discontinuity of the mandible alters its morphology and function, which severely impacts the lives of patients physiologically, aesthetically, and

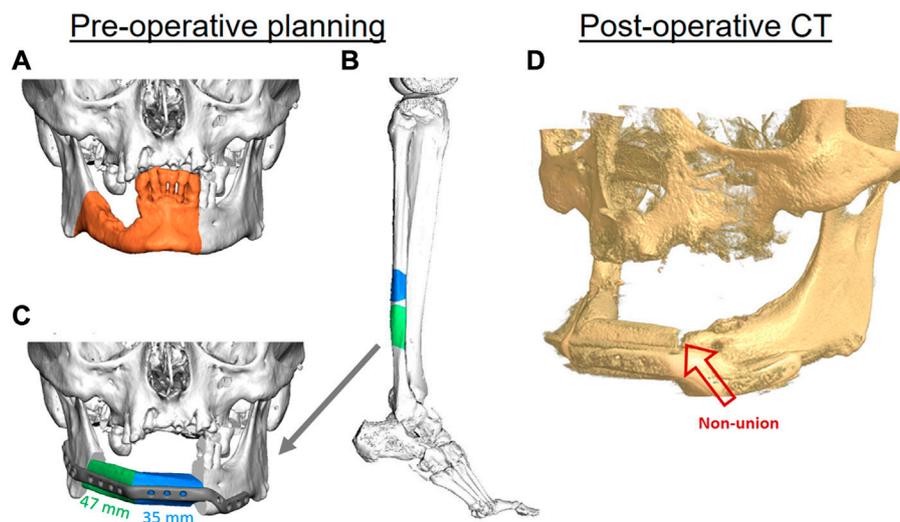


FIGURE 1

The virtual planning steps for mandibular reconstruction. Three-dimensional (3D) reconstruction of a cone beam computed tomography (CBC) scan, showing the area of affected bone including a safety margin of the right-sided mandible (orange area) (A). A two-segmental fibula free flap (FFF) from the right leg (green and blue parts) was virtually planned for the reconstruction of the mandibular defect using computed tomography (CT) (B). The segments were virtually fitted, and fixation was planned using a patient-specific 3D-printed titanium reconstruction plate (C). The postoperative CBCT scan of the patient after 11 months demonstrates a non-union between the FFF and the mandible in the anterior region (red arrow) (D).

psychologically. The intricate geometry, sizeable dimensions, and complex mechanics of the mandible complicate the search for bone engineering alternatives for mandibular reconstruction. Another difficulty in this endeavor is the fact that most bone replacement materials and most reconstructive techniques are designed for and tested in appendicular bones (Fernandez de Grado et al., 2018).

Current useful regenerative treatments for segmental mandibular resections rely on autologous bone flaps from the iliac crest, scapula, and most favorably the fibula. Due to the fibula's limited weight-bearing function in the leg, its ample length, and location, allowing for a two-team surgical procedure (simultaneous resection and flap harvesting), the free vascularized fibula flap (FFF) has become the main surgical approach for mandibular reconstruction (Antúnez-Conde et al., 2021). The clinical use of the FFF technique has allowed for reasonably positive results in dental and prosthetic rehabilitation, speech intelligibility, deglutition function, the aesthetic outcome, and the subjective overall wellbeing of tumor patients post-rehabilitation (Attia et al., 2019). However, even the use of FFF results in several severe complications in 12.4%–20% of cases (Verhelst et al., 2019; Knitschke et al., 2021). Flap postoperative issues can range from early- and late-onset plate-related abscess formation, fistula, plate exposure, and delayed bone healing, to non-union. Consequently, the patient can lose proper mandibular function.

The mandible is unlike any other bone used for its reconstruction. There are differences between the mandible and other bones (appendicular and most flat bones) in morphology, biomechanics, and physiology, as well as morphogenesis, cell genotype and phenotype, the osteoimmune environment, and several clinically relevant aspects. Therefore, the present work aims to investigate the possible influence of the characteristics that distinguish the mandible from other bones on clinical

reconstructive work and the development of new regenerative alternatives, while using a pseudoarthrosis case to illustrate the problem.

2 Clinical case example of osseous non-union after FFF reconstruction

As an example of a typical pseudoarthrosis (non-union) case of FFF, a partial union between the fibula and mandible can occur. In this case, a 70-year-old male patient was initially diagnosed and later treated at the department of Oral Maxillofacial Surgery, Charité—Universitätsmedizin Berlin, for oral squamous cell carcinoma without associated metastases. Secondary diagnoses were hypertension, type 2 diabetes, carotid artery stenosis, and alcohol and nicotine abuse. Consequently, tumor resection including segmental mandibular resection, neck dissection, tracheotomy, and mandibular reconstruction were virtually planned. A patient-specific titanium plate (Karl Leibinger Medizintechnik GmbH & Co. KG, Mühlheim an der Donau, Germany) and a microvascular free fibula flap (FFF) with two segments were virtually designed using the computer-aided design and manufacturing (CAD/CAM) workflow to ensure surgical precision and suitable reconstruction (Figure 1).

Postoperatively, the patient received adjuvant radiotherapy performed over a period of 6 weeks with the application of 56 Gy. Postoperative controls using computed tomography (CT) and clinical examination did not reveal any signs of recurrence within the first 2 years after radiotherapy. Osseous union at the posterior (fibula vs. mandible) and intermediate gaps (fibula vs. fibula) showed satisfactory results; However, there were still signs of incomplete osseous union in the anterior gap connecting the

mandible with the FFF after 12 months (Figure 1D). The removal of fixation plates and screws is generally necessary before dental implant placement for oral rehabilitation. In this case, oral rehabilitation with dental implants was not possible within the first 2 years after surgery and radiotherapy due to the incomplete osseous union. To try to solve the incomplete union, bone grafting from the iliac crest was performed 3.5 years after the initial reconstruction. However, the incomplete union persisted, the plate had to be kept in place, and oral rehabilitation with dental implants still could not be performed. The result was a limited number of teeth in the patient's mouth, which impaired his masticatory and speech capacities.

In the presented case, many factors may have contributed to the flap complication that resulted in an impaired functional outcome, such as age, radiotherapy, and co-morbidities. Despite the patient's overall health issues and habits, the distal area of the fibula did bridge with the mandible. While there was sufficient healing between the two fibula segments in the anterior area, there was diminished bone healing between the fibula and mandible, even though both intersegmental gaps were located correspondingly in the mandible and thus underwent similar biomechanics. This begs the question as to whether the differences in healing are caused by biological differences between the mandible and fibula bone.

3 Uncovering the differences

3.1 Morphogenesis

The most noticeable difference between craniofacial bones, like the mandible, and hard tissues in the rest of the body is their embryonic origin. While the mandible stems from the cranial neural crest, bones from the limbs have a mesodermal origin (Yuan and Chai, 2019). In contrast to mesodermal cells, cranial neural crest cells originate from a broader variety of tissues (Chai and Maxson, 2006) with ectodermal and mesenchymal natures (Zalc et al., 2021), such as smooth muscles, teeth, sensory neurons, and craniofacial bones.

The developmental mechanism of the mandible also differs from other skeletal bones. Appendicular bones, for example, are solely formed through endochondral ossification, a multistage process that first generates a cartilage template, that is, then converted into bone. On the other hand, the lower jaw is formed as a mosaic (Hinton et al., 2017). The most posterior (condyle and angle) and the most anterior (symphysis) areas of the mandible are formed through endochondral ossification, while the bulk of the mandible is formed by intramembranous ossification, a process in which an osteoid tissue is deposited, then mineralized and does not require a cartilage template (Hinton et al., 2017; Yuan and Chai, 2019).

3.2 Morphophysiology and biomechanics

Like other craniofacial bones, the mandible has a complex shape. The mandible is arch-shaped in its anterior portion, called the body, and has two vertical extensions on its posterior ends, the rami. The lower part of the body is the base of the mandible, while the superior part of the body is the alveolar process, which holds the lower teeth.

The anterior area of the body forms a triangle, called the mental protuberance, and it is where the fusion of the two lateral halves of the mandible occurs in early infancy (Nyström and Ranta, 2003). The bilateral most posterior parts of the body form the angles of the mandible with the rami. The rami extend vertically upwards, and each ramus forms a condyle at its most posterior ends. Each condyle (left and right) articulates with its respective temporal bone to form the temporomandibular joints (Figure 2).

It is through the articulation of the temporomandibular joint that the body of the mandible works as a cantilever, while both joints move in an intricate and coordinated three-dimensional (3D) manner. Normal masticatory and speech functions result in frequent loading of the mandible with variable amplitude and force (Grigoriadis et al., 2014). The complexity of the mandibular movements results in the deformation of its structure. Most impressively, during the opening, protrusion (frontal excursion), and lateralization (lateral excursion) movements, the mandible arch becomes narrower, only returning to its original form during centric mouth closing movements (El-Sheikh et al., 2007). Therefore, the shape and constant 3D deformation of the mandible make its biomechanical environment different from that of appendicular bones.

Likewise, there are daily higher, more rhythmic, and predictable compressive strains in the lower jaw bone than in appendicular bones (de Jong et al., 2010). Because bone regeneration is sensitive to loading cycles and the strain rate (Rubin and Lanyon, 1984), the differences between the mandible and the original mechanical environment of the flaps may play a role in bone healing after mandibular reconstructive surgeries. Furthermore, the masticatory muscle fibers have a different profile from those of the trunk and limb muscles. Directly related to the constant motion of the mandible either during function (e.g., chewing, speaking, and swallowing) or parafunction (e.g., grinding, clenching, and thumb suction), masticatory muscles have higher fatigue resistance and increased muscle force (Sciote et al., 2003). This indicates an added difference in the mechanical load at the mandibular site, which will demand changes in the macro- and microstructure of the flaps, as there is a direct link between morphology and biomechanics in bone (Glatt et al., 2017).

For its biomechanical demands, the mandibular structure must be more flexible to withstand daily load-bending movements. The unique constant multidirectional movements of the mandible lead to differences between this and other axial and appendicular bones. The collagen content of the mandible has been shown to be higher and has less post-translational modification (Lys hydroxylation) than that of humeral and femoral bones (Sasaki et al., 2010), which makes the lower jaw bone more flexible. The higher collagen content and lower hydroxylation are also connected to a higher bone toughness, or ability to deform without fracture. Furthermore, bone mineral density (BMD), which looks at the mean mass of a bone in a certain area to assess the fragility or strength (quality) of a bone, seems to be higher in the lower jaw bone body and symphysis areas than in the femur, hip, or spine (Drage et al., 2007).

The microstructure of the mandible also differs from that of other bones. Compared with the iliac, the cortical bone of the mandible has a smaller vessel surface to bone volume ratio, and therefore a lower vessel porosity (Rothweiler et al., 2022). Bone mineralization density distribution (BMDD) is used to measure

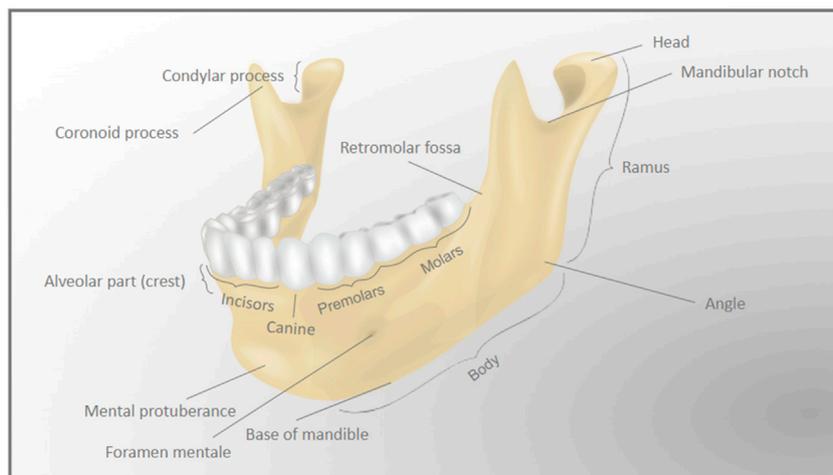


FIGURE 2
Detailed anatomy of the mandible. The complex anatomy of the mandible (lower jaw) bone is represented by the differently labeled anatomic regions.

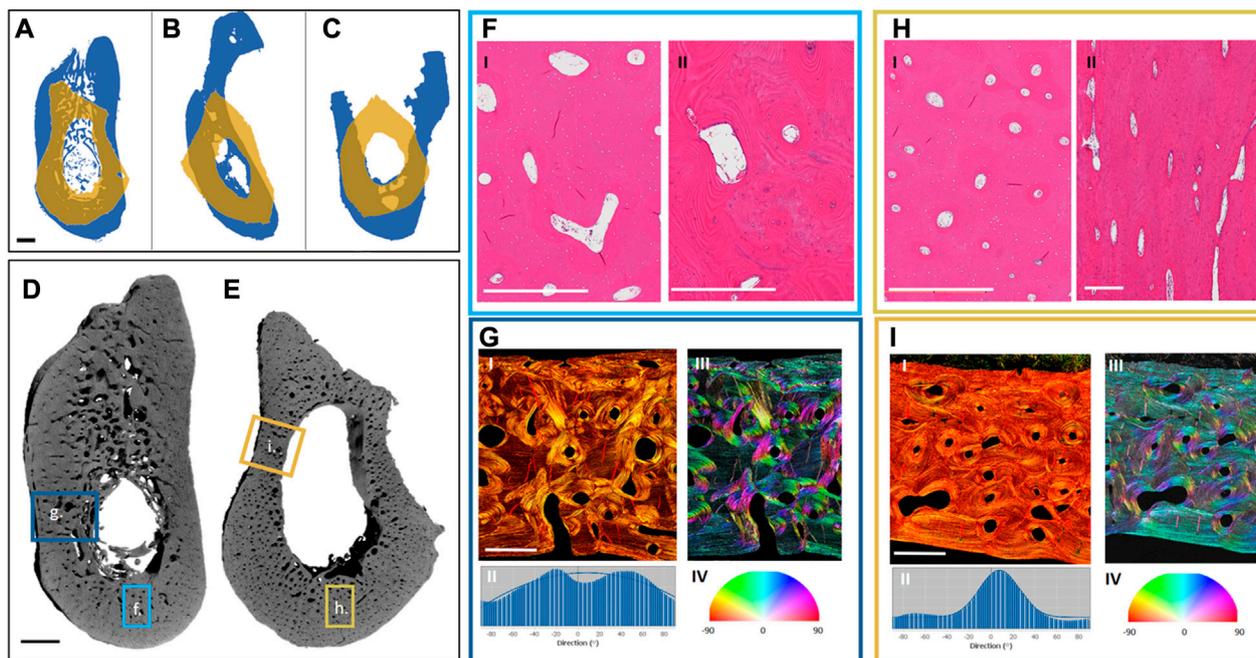


FIGURE 3
Preliminary analysis of differences between the mandible and the fibula. Projections of the exemplary cross-sections of three different patients (A–C): The mandibular projections in blue and the overlaying projections of the corresponding fibulas in yellow, showing the differences in the shape and size of the bones. Three different fibula shapes can be noted: Irregular (A), triangular (B), and quadrilateral (C). Renderings of one of the patient samples reveal the different complexity in the morphology of the mandible (D) and the fibula (E). Highlighted hematoxylin and eosin (H, E)-stained regions from the mandible (F) and fibula (H)—Cross-sections of the mandible (F, I) and fibula (H, I)—Reveal a more homogeneous size and distribution of the Haversian canals in the fibula, and a larger vascular area in the mandible. Further, regional longitudinal sections in the mandible (F, II) and fibula (H, II) again show a more regular longitudinally aligned Haversian system in the fibula. Highlighted Picrosirius red-stained regions from the mandible (G, I) and fibula (I, I) were analyzed for the directionality of their fiber bundles (G, II, I, II), showing higher multi-directionality in the mandible (G, II, G, III, G, IV) than in the fibula (I, II, I, III, I, IV). The black scale bar is 2 mm and the white scale bar is 500 μ m.

bone mass distribution on the microscale. BMDD indicates areas of higher and lower mineral apposition. Moreover, the lacunar-canalicular network (LCN) is the group of small cavities (lacuna

in the bone that house osteocytes and their minute dendritic extensions that form interconnecting channels (canaliculi), through which neighboring cells can communicate. The BMDD

and lacuna distribution have a more heterogeneous spatial distribution in the mandible than in the tibia or femur (Hesse et al., 2014). There is a higher bone density surrounding the mandibular LCN (Hesse et al., 2015). Furthermore, the total lacunar volume is larger in mandibular samples (Hesse et al., 2014), and there is a higher canalicular volume and canalicular length, and more nodes in the younger mandibular LCN (Bortel et al., 2022). However, the impact of this greater cellular communication in the mandible compared with possible flaps is still uncertain.

3.3 Direct comparison between the mandible and the fibula

The free vascularized fibula flap (FFF) is the main surgical approach for mandibular reconstruction; it is chosen due to its ample length and mostly triangular cross-section, similarly to the mandible (Ide et al., 2015). However, geometrically, the fibula presents major disadvantages for mandibular rehabilitation, as its lower height can hinder implant-prosthetic rehabilitation, and its linear shape cannot mimic the round angles of the lower jaw (Antúñez-Conde et al., 2021) (Figures 3A–C).

To better understand the underlying differences in the bones which we aim to join together by using the FFF technique, a preliminary look at the morphological patterns of mandible and fibula bones was undertaken. Exemplary mandibular and fibular cross-sections were harvested from three different patients (under Charité Universitätsmedizin Ethical committee approval EA1/062/21) undergoing mandibular reconstruction using FFF. The samples were harvested from the edges of the resection during surgery. They were then imaged using a laboratory micro-CT (Skyscan 1172, Bruker, pixel size: 9 µm) and histologically processed and stained with hematoxylin and eosin as well as Picosirius red.

The macroscopic examination of the samples revealed that the fibular sections have a thicker cortical area, no trabecular bone, and corresponded to between 1/2 and 2/3 of the mandibular height (Figures 3D, E). Each fibula sample could be distinctively classified based on its shape as irregular (Figure 3A), triangular (Figure 3B), or quadrilateral (Figure 3C) (Ide et al., 2015). Microscopically, there is a similar osteocyte density (number of osteocytes per bone area) in the mandibular and fibular samples and a higher vascularity (vessel area per bone area) in the cortex of the mandible. Furthermore, the Haversian system distribution is different between samples (Figures 3F, H). There is greater homogeneity in the fibula with a dominance of longitudinally aligned Haversian canals (Figure 3F). In contrast to the fibula, the Haversian system in the mandible presents a more irregular pattern in both orientation and shape (Figure 3H). Furthermore, the organization of the collagen fiber bundles revealed by Picosirius red staining is different between the bones (Figures 3G, I). While the fibula possesses unidirectional wide layers of collagen fibers on the outer and inner circumference of the cortical bone (Figures 3L,I–IV), the mandible shows a more irregular collagen pattern (Figures 3G,I–IV). These morphological differences are most likely connected to the biomechanical environment of each sample.

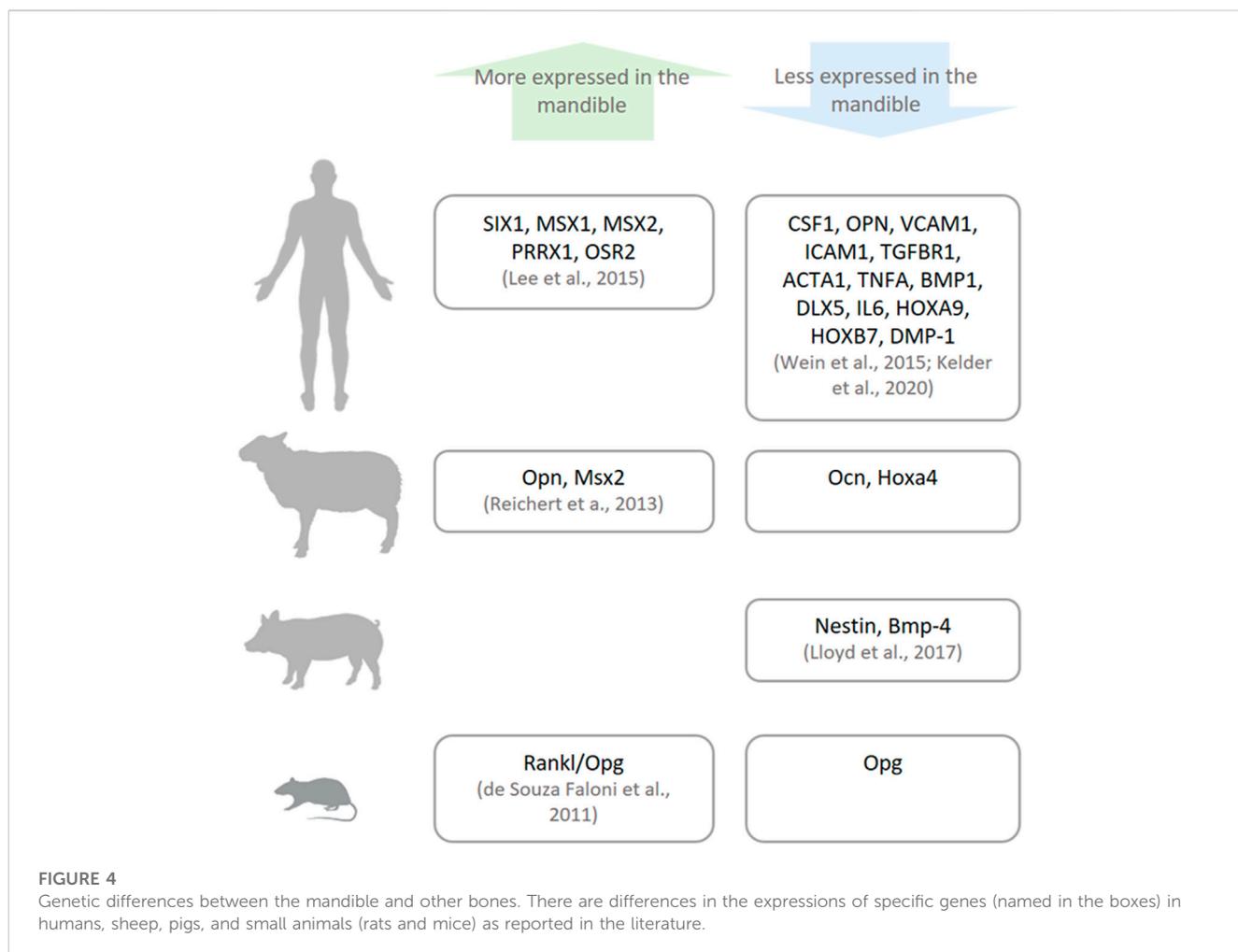
Because this is a preliminary comparison of the morphology of both bones, a more ample assessment is needed with multiple samples and their surgical follow-up to assess the impact of different morphologies on the clinical results. Further description of the methodology used for the morphological assessment can be found in the [Supplementary Material](#).

3.4 Gene expression

The gene expression profile differs between bone sites (Isaac et al., 2017; Youten et al., 2021) and at times between species (Figure 4), most notably, the expression of genes that function as morphogenesis regulators and cell differentiation, called homeobox genes (Mark et al., 1997). Homeobox genes are divided into two subfamilies: clustered (Hox) genes, which provide cells with regional information along the body axis and are highly expressed in appendicular bones, and non-clustered (Hox negative) genes (e.g., MSX, PAX, and DLX), which are present in different tissues and possess multiple functions including craniofacial morphology (Leucht et al., 2008). Hox stem cells harvested from appendicular bones and grafted into a mandibular defect are able to keep their Hox profile (Leucht et al., 2008). This discrepancy between the Hox status of the mandible and grafted cells lingers throughout the bone healing process, and as result, the transplanted cells differentiate into chondrocytes instead of osteoblasts, locally ensuring an atypical endochondral ossification instead of an intramembranous one (Leucht et al., 2008). On the other hand, when Hox-negative cells from the mandible are transplanted into tibial defects, they can change their Hox profile (Leucht et al., 2008). This higher plasticity of mandibular cells can be a result of their embryonic origin that stems from broader multipotent cranial neural crest cells.

Besides the difference in morphogenic genes, the mandible and appendicular bones in the human body can also diverge in their genetic profile. Lee et al. (2015) confirmed higher expression of craniofacial morphogenesis-related genes (non-clustered Homeobox genes: SIX1, MSX1, MSX2, PRRX1, and OSR2) in the mandible compared with the iliac bone. Moreover, the iliac bone and tibia show higher expression of genes related to the proliferation and differentiation of stem cells into the osteolineage (Hox genes: HOXB7 and HOXA9); osteoblast differentiation (DLX5 and DMP-1; Wein et al., 2015); cartilage formation and maintenance (BMP1 and TGFBR1; Wein et al., 2015); osteoclast differentiation, activation, and their attachment to the extracellular matrix (CSF1, ICAM1, VCAM1, OPN; Wein et al., 2015; Kelder et al., 2020); and structural cytoskeletal component of osteocytes (ACTA1; Kelder et al., 2020). Taken together, the difference in gene expression signals the more mature and differentiated character of cells from appendicular bones. The dissimilarity in gene profiles between bone cells may influence their divergent response to stimuli.

A well-researched clinical example of the difference between the response of the mandible and appendicular bones to stimuli is the reaction to chronic consumption of bisphosphonates, a medication used to prevent bone loss. In the mandible, bisphosphonates induce the suppression of the MSX1 and OPN genes and a subsequent increase in DLX5 expression (Wehrhan et al., 2011). MSX1 suppression limits the proliferation of pre-osteoblasts (Roybal et al., 2010; Wehrhan et al., 2011) and also causes an



increase in the expression of DLX5, which in turn induces osteoblast terminal differentiation and further extracellular matrix mineralization (Zhang et al., 2003; Wehrhan et al., 2015). OPN depletion impairs osteoclast attachment to the bone matrix (Wehrhan et al., 2015) as well as the migration of endothelial cells and, therefore, the early development of blood vessels (Wein et al., 2019). Moreover, although both the mandible and appendicular bones express OPN, only one type of RNA splice variant is expressed in the mandible (OPNa), while two others are expressed in appendicular bones (OPNb and OPNc) (Wein et al., 2019). This difference might be one of the reasons that bisphosphonates specifically impact the blood perfusion of the mandible. Together, MSX1 and OPN suppression generates a highly mineralized extracellular matrix, with a lack of efficient bone turnover and low blood perfusion, therefore resulting in tissue death characteristically present in the pathomorphology of bisphosphonate-related osteonecrosis of the jaws (BRONJ) (Koerdt et al., 2014). Perhaps due to the lack of deleterious impact on appendicular bones, FFF can be used in mandibular reconstruction of osteonecrosis cases with a >90% success rate (Sacco et al., 2018).

In vertebrates used as animal models in bone research, gene expression also varies between the mandible and appendicular bones, although not necessarily in the same pattern as their

human orthologs. Specifically, the mandible from murine, porcine, and ovine origins have been harvested and their gene expression evaluated and compared with their appendicular bone counterparts. In rodents, the Rankl/Opg ratio, which positively regulates osteoclast activation, is higher in mandibular marrow-derived cells (de Souza Faloni et al., 2011). In porcine bone marrow cells, there is higher expression of genes related to bone development (Nestin) and limb formation (Bmp-4) in appendicular bones (Lloyd et al., 2017). On the other hand, in ovine bones, genes related to craniofacial morphogenesis (Msx2) and osteoclast attachment to the extracellular matrix (Opn) are more expressed, while genes related to bone remodeling (Ocn) and limb morphogenesis (Hoxa4) are less expressed in the lower jaw (Reichert et al., 2013). The differences between species and their gene expression may be caused by evolutionary epigenetic regulation (Chan et al., 2018).

3.5 *In vitro* dynamics and *in vivo* applications

Numerous studies have tested and compared the characteristics of cells harvested from the mandible of various species to corresponding cells from appendicular or flat bones (non-craniofacial bones). As shown in Figure 5, the outcome from *in vitro* and *in vivo* assessments of skeletal cells can vary, which

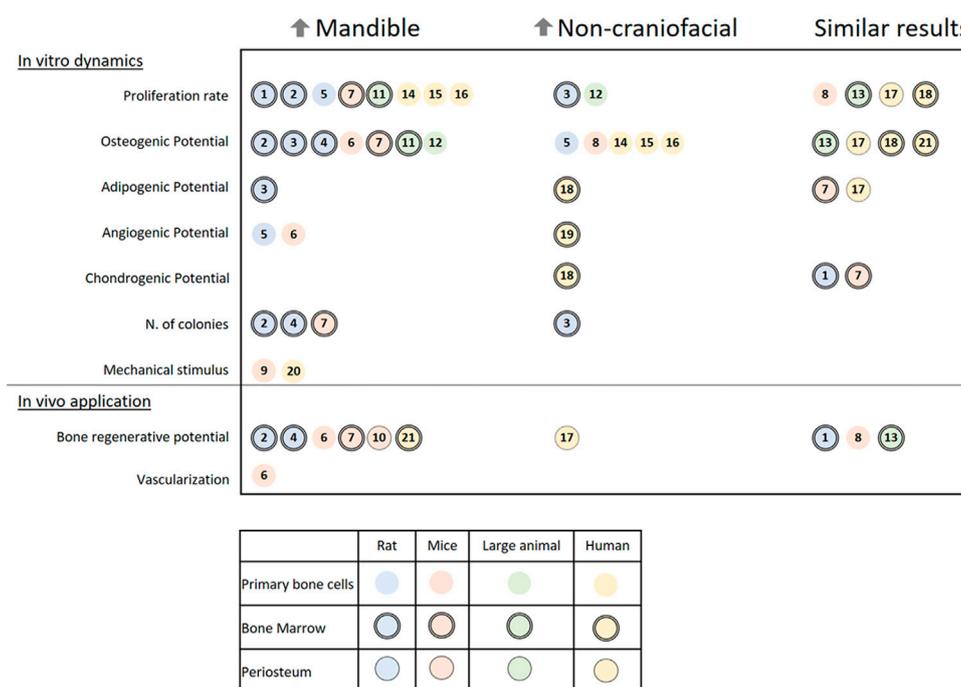


FIGURE 5

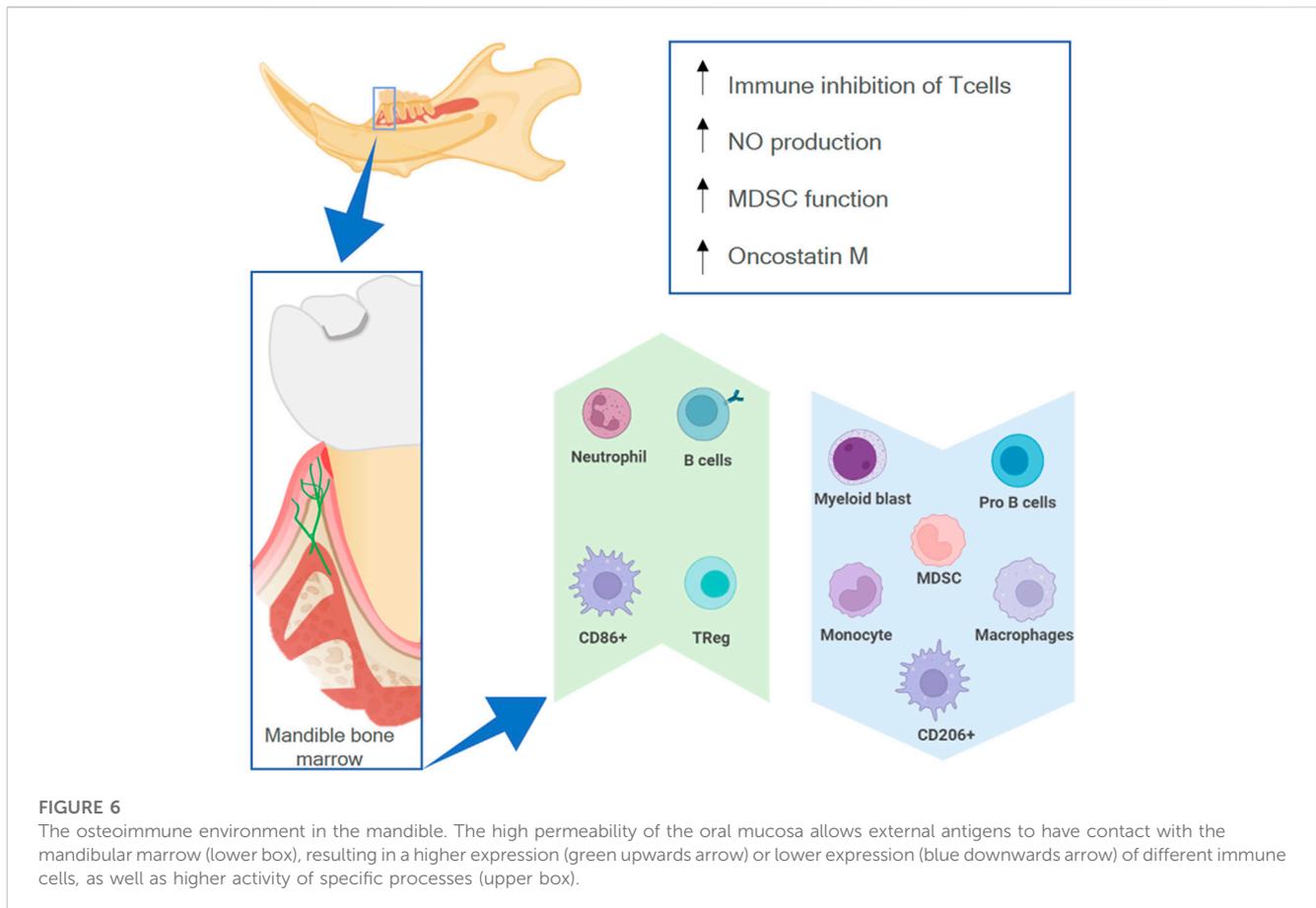
A literature review on the *in vitro* dynamics and *in vivo* applications of different cells harvested from the mandible and non-craniofacial bones. The results are separated into three columns referencing papers (a total of 21 papers) in which cells from the mandible of rats, mice, large animals, or humans presented higher activity (↑Mandible) or lower activity (↑Non-craniofacial) or similar activity (Similar results) when harvested from the same individual and compared in the same conditions to non-craniofacial bones cells. Symbols indicate the origin of the cells: Rats (blue), mice (orange), large animal models (green), humans (yellow), primary bone cells (open circle), bone marrow cells (double contoured circle), and periosteum (single contoured circle). Numbered references: 1) 10.1177/2041731419830427, 2) 10.1007/s00441-014-1927-4, 3) 10.1089/scd.2019.0256, 4) 10.1177/0022034510378427, 5) 10.1007/s10735-019-09810-6, 6) 10.1016/j.jbo.2020.100346, 7) 10.1177/0022034510387796, 8) 10.1002/jbm4.10382, 9) 10.1016/j.yexcr.2011.07.015, 10) 10.1111/jre.12229 11) 10.1016/j.archoralbio.2017.01.012, 12) 10.1016/j.gene.2013.04.026, 13) 10.1177/0022034518772283, 14) 10.1016/j.jcms.2015.07.030, 15) 10.1007/s00784-014-1353-8, 16) 10.3390/jjms21145072, 17) 10.3389/fcell.2020.554984, 18) 10.1359/JBMR.041117, 19) 10.1016/j.acthis.2015.02.006, 20) 10.1016/j.jobcr.2019.09.005, 21) 10.1111/j.1601-0825.2007.01402.x.

further demonstrates the need to standardize methods to examine possible reasons for the distinct results. For better understanding in this review, the results from different papers have been classified by cell origins as primary bone cells, periosteum cells, or bone marrow cells, and by the studied species divided into rats, mice, large animal models (comprising sheep, dogs, and pigs), or humans.

The first characteristic that most researchers examine is the proliferation rate of the isolated cells. In the majority of papers, cells harvested from the mandible present a higher proliferation rate than of any other bone (Yamaza et al., 2011; Lee et al., 2015, Lee et al., 2019; Wein et al., 2015; Lloyd et al., 2017; Yang et al., 2019; Kelder et al., 2020). Important exceptions are connected to the difference in medium formulation (Li et al., 2020) and the comparison between cells from juvenile bone (Reichert et al., 2013). A colony-forming unit assay has also been used in a few studies and has shown that mandibular cells have a higher cloning ability (Aghaloo et al., 2010; Yamaza et al., 2011; Dong et al., 2014).

When assessed, the osteogenic potential of cells harvested from the mandible varies among the studied species. The majority of small (rats and mice) and large animal models have shown a higher osteogenic potential in cells harvested from the mandible (Aghaloo et al., 2010; Yamaza et al., 2011; Reichert et al., 2013; Dong et al., 2014; Lloyd et al., 2017; Li et al., 2020; Eber et al., 2021). Meanwhile,

in humans, cells harvested from appendicular and flat bones mostly have a higher osteogenic potential (Wein et al., 2015; Kelder et al., 2020), although it is sometimes similar to that of cells harvested from the mandible (Groeneveldt et al., 2020). Only one report showed higher osteogenic potential in human mandibular bone marrow stem cells (BMSCs; Stefanik et al., 2008); Note that compared with other studies, these authors used a higher concentration of fetal bovine serum (FBS; 20%) in the growth medium. Not much can be said about the comparison of the adipogenic, chondrogenic, or angiogenic potentials of the cells, as only some of the studies have presented data about it (Matsubara et al., 2004; Yamaza et al., 2011; Marini et al., 2015; Lee et al., 2019; Groeneveldt et al., 2020; Li et al., 2020; Eber et al., 2021) with confounding results. Furthermore, when mechanically stimulated by orbital shear stress, there is an increase in osteogenesis, pro-angiogenic cytokines, and mineralization of cells harvested from the mandible *in vitro* (Pravitharangul et al., 2019). When stimulated by ultrasound, the same type of cells from mice increase their receptor activator of nuclear factor kappa-B ligand (RANKL) expression, a gene related to the increase of osteoclastic activity (Watabe et al., 2011). The results of both mechanical *in vitro* studies indicate that local remodeling of the mandible is mechanically modulated, maybe even more so than in other bones.



The expression of distinct genes and proteins can vary at different time points between the mandible and non-craniofacial bone tissue *in vitro*. Their expression dynamics reveal that the cells diverge in response to osteogenic induction. Importantly, there is a greater increase in alkaline phosphatase (ALP) activity in cells harvested from the mandible of small and large animals (Yamaza et al., 2011; Reichert et al., 2013; Dong et al., 2014; Lloyd et al., 2017). This enzyme is one of the most used markers for osteogenic differentiation. Moreover, osteocalcin (OCN), which encodes an essential protein for apatite crystallites alignment and bone strength (Moriishi et al., 2020), has higher expression in *in vitro* cells harvested from the rat mandible (Aghaloo et al., 2010; Dong et al., 2014; Lee et al., 2019; Li et al., 2020).

All the differences between cells harvested from the mandible and non-craniofacial bones *in vitro* can impact their application in a tissue engineering context. The characteristics that distinguish the mandible might even require site-specific cell choice (Reichert et al., 2013). Researchers have used small animal models to test the bone regenerative potential of the isolated cells. They have mixed the cells with various materials (e.g., gelatin and β -tricalcium phosphate) and then applied the mixtures to the animals either subcutaneously (Aghaloo et al., 2010; Yamaza et al., 2011; Groeneveldt et al., 2020; Son et al., 2020; Eber et al., 2021) or into bone defects (Stefanik et al., 2008; Dong et al., 2014; Ichikawa et al., 2015; Wang et al., 2018; Lee et al., 2019). The results revealed that cells harvested from the mandible produce more mineralized tissue content than cells harvested from non-craniofacial bone (Aghaloo et al., 2010;

Yamaza et al., 2011; Dong et al., 2014; Ichikawa et al., 2015; Eber et al., 2021).

3.6 Osteoimmunology

Interactions between immune and bone systems, termed the osteoimmune environment, play an essential role in regulating bone health. Both innate and adaptive immune cells are associated with the dynamic processes of bone turnover and can be classified through their cluster of differentiation (CD), meaning the expression of different surface molecules (Actor, 2019). Most remarkably, macrophages, which are highly plastic innate immune cells, give rise to essential bone cells: osteoclasts. Macrophages are also present in all bone healing phases, having either a more pro-inflammatory profile (so-called M1 macrophages) and expressing CD86⁺ on its surface, or a more bone-regenerating profile (also called M2 macrophages) with CD206⁺ surface expression (Schlundt et al., 2021). T cells with different CD play a dynamic role in bone homeostasis (Pacifci, 2016). While activated CD4⁺ CD8⁺ T cells and CD4⁺ T helper 17 (Th17) can increase bone resorption, regulatory CD4⁺ T cells (Tregs) can inhibit bone resorption. Activated B cells are also implicated in increased bone resorption (Horowitz et al., 2010).

The mandible has a different osteoimmune microenvironment from that of non-craniofacial bones (Figure 6). Anatomically, the existence of a permeable barrier between the oral cavity and the

periodontal ligament (called junctional epithelial attachment) as well as the presence of lymphatic vessels circulating from the oral mucosa—Colonized by a wide range of microbes—To the bone marrow allow microbial products to constantly reach the mandibular marrow space (Jensen and Folke, 1974; Barker, 1982). Therefore, the bone marrow of the mandible is constantly exposed to antigens from the oral microbiota and has a more active immune environment than non-craniofacial bones, which only have contact with bacterial products during infections or when transported by the vasculature (Hathaway-Schrader et al., 2022).

Due to the environment in the oral cavity, there are differences between the myeloid cell population in the mandible and in non-oral bones. There are fewer monocyte progenitor cells like myeloid lineage-early blasts and myeloid blasts as well as fewer monocytes, myeloid-derived suppressor cells (MDSCs), and macrophages (Cd11b + F4/80+) (de Souza Faloni et al., 2011; Kwack et al., 2021) in the mandibular marrow. Furthermore, BMSCs from the mandible of mice show high nitric oxide production (Yamaza et al., 2011). There is a higher inhibitory capacity of CD4⁺ CD8⁺ T cells, more Tregs, an increase in MDSC function, and more B cells (Kwack et al., 2021). These features demonstrate a higher immunosuppressive potential and more adaptive humoral response in the mandibular environment. This might be necessary because commensal microbiota can provoke a higher pro-inflammatory response in alveolar bone marrow cells than on appendicular bone marrow cells, increasing the amount of activated dendritic cells, CD4⁺ T cells, and Th17 cells (Hathaway-Schrader et al., 2022). Moreover, there are more CD86⁺ and fewer CD206⁺ macrophages in the mandible, which also signals a more pro-inflammatory response to microbial products (Lin et al., 2021).

Macrophages/monocytes can also elicit a greater modulatory effect on mesenchymal stem cells (MSCs) in the mandibular context. Macrophage conditioned medium has a greater effect on the proliferation, colony formation, and osteogenic potential of MSCs harvested from the mandible (Lin et al., 2021). Furthermore, macrophages harvested from the mandible have a higher *in vitro* capacity for recruiting MSCs (Dong et al., 2014) as well as a higher expression of oncostatin M (Osm) (Lin et al., 2021). Osm is a cytokine that can regulate the osteogenic fate of MSCs (Guihard et al., 2015) and stimulate osteoblasts to secrete RANKL, which in turn results in more active osteoclasts in the mandible (Lin et al., 2021).

The osteoclastic population of the mandible also differs from that of non-craniofacial bones. Morphologically, osteoclasts from the mandible are larger (de Souza Faloni et al., 2011), longer, and have a smaller amount of nuclei (Azari et al., 2011; de Souza Faloni et al., 2011; Goldberg, 2016). *In vitro*, when compared with appendicular bones, the osteoclastogenesis from mouse mandibular marrow cells is slower (Goldberg, 2016), while cells from the bone marrow of human mandibles have a lower osteoclastogenic potential (Kelder et al., 2020). For rats and mice, there is a similar resorptive activity of mandibular and non-craniofacial bone osteoclasts (de Souza Faloni et al., 2011; Goldberg, 2016); however, osteoclasts from human mandibles have a lower resorptive potential (Kelder et al., 2020). The substrate can modulate osteoclastic activity, and when mandibular osteoclasts are seeded in dentin they grow larger in size and number (de Souza Faloni et al., 2011). Furthermore, osteoclasts from the mandible can respond differently to hormonal changes. In culture under the influence of parathyroid

hormone (PTH), osteoclastogenesis of mandibular bone marrow cells is lower than that of non-craniofacial bones (Chaichanasakul et al., 2014), and *in vivo*, the loss of ovarian hormones does not appear to affect the osteoclastic content (Goldberg, 2016).

The assembled data on osteoclasts seem to underline the difference between bones as well as between osteoclastic activity in humans and other species. As reviewed by Sculean et al. (2019), mandibular bone remodeling after tooth extraction in large animal models (e.g., dogs and monkeys) is faster than in humans. Additional data from the literature also reveal a lower remodeling potential of human mandibles at the cellular level compared with appendicular bones (Kelder et al., 2020). Moreover, although it is customary to state that bone turnover of the mandible is higher than of appendicular bones due to the results of animal studies (Huja et al., 2006; Inoue et al., 2019), this does not seem to be applicable to a broad human population, as there is high variability between individuals (Trombelli et al., 2008). As shown in different clinical studies, bone turnover of the mandible can be similar to that of different appendicular bones (Ristow et al., 2014a; Ristow et al., 2014b).

3.7 Clinical aspects

Due to the mandible's role in the physiology of multiple body functions and the morphology of the lower third of the face, rehabilitating this bone is of the utmost importance. To achieve good surgical results, understanding the susceptibility of the mandible to different ailments and how its healing process works is essential. Furthermore, knowing the differences and similarities between the mandible and the bones used for its regeneration is important for a better long-term outcome of mandibular reconstruction.

Several researchers have investigated, and some have compared, appendicular bone and mandibular healing using small animal models. Overall, these papers have shown similarities between the mandible and appendicular bones regarding their healing processes. Bone fractures in both the mandible and appendicular bones respond to unstable fixation by secondary healing and cartilage formation (Wong et al., 2021; Ito et al., 2022), while stable fixation leads to primary healing and greater intramembranous ossification (DeConde et al., 2014; Wong et al., 2021). To mimic the same physical conditions of tooth extraction in appendicular bone, drill holes can be made with the removal or preservation of the periosteum (Liu et al., 2020; Ito et al., 2022). In rats as in humans, the extraction socket consistently heals through intramembranous ossification as the remaining periodontal cells go through osteoblastic differentiation (Cardaropoli et al., 2003; Trombelli et al., 2008; Ito et al., 2022); However, the ossification speed of the socket is highly variable and individual (Trombelli et al., 2008). Furthermore, drill holes seem to heal faster in the rat tibia than in the rat mandible (Liu et al., 2020). Although drill holes in appendicular bones with and without periosteum mainly heal through intramembranous ossification, there is associated cartilaginous formation in the wound when the periosteum is present, which demonstrates a higher chondrogenic potential in long bones (Ito et al., 2022).

Different techniques can be used for mandibular bone augmentation. Distraction osteogenesis is a surgical technique

used to induce changes in bone morphology (height or thickness) by gradual separation and controlled bone formation within the interval space of surgically severed bones. Although used in the mandible and appendicular bones with a similar protocol, distraction osteogenesis has a higher complication rate in appendicular bones (Shah et al., 2021). The reason might be that the distraction osteogenesis process stimulates the focal adhesion kinase (FAK) signaling pathway that is naturally active in cranial neural crest cells during development, and this reversion to a more primary state of the cells can in turn facilitate regeneration in the mandible (Ransom et al., 2018). Another bone augmentation technique is bone grafting. It can be used for horizontal (width) or vertical (thickness) mandibular augmentation. Bone grafts can be harvested from the mandible itself (angle or symphysis) or most commonly from the iliac bone. When mandibular grafts are used, there is a slower vertical loss of the graft; furthermore, they have a lower incidence of peri-implantitis (Kang et al., 2015) and a higher implant survival rate (Sbordone et al., 2009). However, when grafts larger than 5 mm are needed, iliac bone grafts can lead to better results, as they have a structure that allows greater blood perfusion, a necessary trait for larger grafts (Troeltzsch et al., 2016; Rothweiler et al., 2022).

Many changes occur in the organism throughout the aging process that affect bone morphophysiology. Although aging greatly impacts appendicular and flat bones, it does not have the same effect on the mandible. For example, protein-energy malnutrition is a common condition in elderly people; it is mainly caused by appetite loss and ingestion difficulties connected to different age-related ailments (Price, 2008). Protein undernutrition can have an impact on human bone health, leading to bone mass loss and an increase in fractures (Rizzoli and Bonjour, 2004). Appendicular bones are more affected by undernutrition than the mandible in both adult and newborn rats (Nakamoto and Miller, 1977; Nakamoto and Miller, 1979; Mavropoulos et al., 2007). To our knowledge, there is no report on the undernutrition effect on the mandibular structure in humans (Algra et al., 2021).

Another age-related ailment is osteoporosis. The impact of osteoporosis on the mandibular structure of patients is not yet understood. There is no clear evidence that estrogen deficiency induces osteoporotic changes in human mandibles (Nicolielo et al., 2017). Meanwhile, small animal ovariectomized models have shown that in the same individuals, the femur and the tibia are more affected, and thus have a greater decrease in the bone volume fraction and BMD as a consequence of ovariectomy (Mavropoulos et al., 2007; Hsu et al., 2016). Adipogenesis in bone marrow cavities and the formation of bone marrow adipose tissue (BMAT) are also associated with aging and osteoporosis. There is a markedly higher BMAT increase in appendicular bones of ovariectomized rats than in the mandible (Coutel et al., 2019). Furthermore, titanium screws implanted in the mandibles of ovariectomized rats present higher osseointegration than screws implanted in their femurs (Liu et al., 2020). Independent of gender, in humans affected by osteoporosis, there is a moderate correlation between changes in the BMD of reference bones (femur, forearm, and lumbar bones) and the mandible (Horner et al., 1996; Esfahanizadeh et al., 2013). However, the impact of osteoporosis in the mandible is less than in appendicular and flat bones (Horner et al., 1996; Mavropoulos et al., 2007; Esfahanizadeh et al., 2013; Hsu

et al., 2016; Nicolielo et al., 2017; Liu et al., 2020). Tooth loss more than aging impacts the morphology of the mandible (Oetlé et al., 2016), without any alterations to the mandible's BMD (Sprunge et al., 2014).

The most documented adverse effect that specifically affects oral bones is medication-related osteonecrosis of the jaws (MRONJ). Although MRONJ is the topic of diverse literature reviews, its pathophysiology is not yet fully understood. It is considered a rare occurrence, but many clinical reports have demonstrated the development of MRONJ in patients using antiresorptive medications and angiogenic inhibitors. The most prevalent triggering factors to the development of MRONJ are tooth extraction and spontaneous onset of osteonecrosis (Fliefel et al., 2015). There are a few *in vitro* and *in vivo* reports directly comparing the effects of such medication on the mandible and appendicular bones. *In vivo* reports have shown that the use of zoledronic acid, a potent bisphosphonate applied in numerous diseases to decrease bone resorption, has a targeted deleterious effect in rat and mouse mandibles. Serial injections of zoledronic acid result in three deleterious effects: 1) A decrease in the number of mandibular marrow cells (Wang et al., 2019); 2) Lower expression of mandibular bone turnover signaling promoters (RANKL/OPG and Wnt-3); and 3) Overall suppression of alveolar bone remodeling (Gong et al., 2017), more so in the mandible than in the maxilla (Wang et al., 2019). In contrast, zoledronic acid enhances bone quality during bone remodeling in appendicular bones (Gong et al., 2017). *In vitro* tests using human cells taken from the mandible, iliac, and other appendicular bones and placed under the influence of pamidronate, another potent bisphosphonate, have shown that BMSCs more than bone-derived primary cells can be influenced by this medication (Marolt et al., 2012). The influence of pamidronate *in vitro* leads to a decrease in the survival and metabolic activity of mandibular BMSCs as well as an increase in osteoclastic recruitment compared with iliac BMSCs (Stefanik et al., 2008). After implanting these cells into bone defects in mice, the authors found greater bone formation but with a less organized structure when compared with BMSCs from the iliac bone (Stefanik et al., 2008). These results show the difficulty in ascertaining the cause of MRONJ. The human body, the complex interaction between different systems, and the differences at the individual level are hard to mimic in experimental settings. However, as mentioned earlier in this review, there are genetic clues as to why MRONJ develops particularly in oral bones.

Differently from non-oral bones, the mandible can be affected by odontogenic tumors, which originate from the tooth and periodontal tissues. Moreover, the incidence of distinct bone tumors is different in the mandible. The occurrence of osteochondroma and intraosseous lipoma, for example, is lower than in appendicular bones (Xi et al., 2008; Cakarer et al., 2009; Angiero et al., 2011), and osteochondroma presents a better prognosis in the mandible.

All the aforementioned clinical differences can lead to dissimilar responses from the host and harvested bone to the mechanical and biological environment of the mandible. Observing and differentiating the two bone areas can be important for clinical follow-up, to understand and diagnose their changes. It is especially important to have these clinical differences in mind, because creeping substitution, or

the replacement process of the bone graft tissue by new locally formed bone tissue, can take years (Roberts and Rosenbaum, 2012), and cells harvested from long bone can keep their genetic profile at the graft-receiving site (Leucht et al., 2008). Although the clinical success rate of FFF is high, changes in both bone areas post-reconstruction may be dissimilar. More research is needed to investigate microstructural changes and how the mandible and the graft respond to aging and comorbidities over time.

4 Conclusion

As this review has shown, there is a large assembly of data pointing to the uniqueness of the physiology, pathology, and clinical needs of the mandible. The main differences between the mandible and other bones can be summarized as follows.

1. While the mandible stems from the cranial neural crest, bones from the limbs have a mesodermal origin.
2. The morphophysiology of the mandible and its associated muscles are different from appendicular bones mainly due to daily higher, more rhythmic, and predictable compressive strains and thus unique biomechanics.
3. The gene expression profile differs between bone sites, especially between morphogenic genes.
4. Cells harvested from the mandible of different species can behave in a dissimilar manner when tested *in vivo* and applied *in vivo*.
5. The mandible has a more active osteoimmune microenvironment than that of non-craniofacial bones.
6. Aging, comorbidities, and pathologies affect the mandible in a dissimilar manner compared with other bones.

Although researchers have uncovered differences between the mandible and other bones, there are still essential questions concerning mandibular biology and its response to stimuli. The underlying differences between the fibula and mandible could have an impact on osseous healing in cases of pseudarthrosis after segmental mandibular reconstruction. Additional studies are needed to closely investigate the causes that lead to diminished healing after osseous reconstructions.

Author contributions

AS, HF, CR, and KS-B conceived the paper's idea. AS, HF, KS-B, and SA searched the literature and extracted the data. CR and CS verified the analyzed data. CS and CR analyzed and wrote the clinical case reported in the review. AS performed the

morphological analysis of patient samples presented in the review. CR encouraged the investigation of FFF complications stemming from differences between the mandible and the bone flap and supervised the findings of this work. All authors discussed the results and contributed to writing the final manuscript.

Funding

The authors acknowledge the support of the European Union (EU) Horizon 2020 project InterLynk (grant agreement: H2020-NMBP-TR-IND-2020, project ID: 953169) for the present work. HF is a participant in the BIH-Charité Clinician Scientist Programme funded by the Charité—Universitäts-Medizin Berlin and the Berlin Institute of Health. We acknowledge financial support from the Open Access Publication Fund of Charité—Universitätsmedizin Berlin and the German Research Foundation (DFG).

Acknowledgments

We would like to thank Sabine Stumpp for the histological workup. Figures 2, 4 were created with the help of Biorender.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fphys.2023.1152301/full#supplementary-material>

References

- Actor, J. K. (2019). "A functional overview of the immune system and immune components," in *Introductory immunology*. Editor J. K. Actor (Cambridge, MA: Academic Press), 1–16. doi:10.1016/B978-0-12-816572-0.00001-2
- Aghaloo, T. L., Chaichanasakul, T., Bezouglaia, O., Kang, B., Franco, R., Dry, S. M., et al. (2010). Osteogenic potential of mandibular vs. long-bone marrow stromal cells. *J. Dent. Res.* 89, 1293–1298. doi:10.1177/0022034510378427
- Algra, Y., Haverkort, E., Kok, W., Etten-Jamaludin, F. V., Schoot, L. V., Hollaar, V., et al. (2021). The association between malnutrition and oral health in older people: A systematic review. *Nutrients* 13, 3584. doi:10.3390/nu13103584
- Angiero, F., Moltrasio, F., Cattoretti, G., and Valente, M. G. (2011). Clinical and histopathological profile of primary or secondary osteosarcoma of the jaws. *Anticancer Res.* 31, 4485–4489.

- Antúnez-Conde, R., Salmerón, J. I., Díez-Montiel, A., Agea, M., Gascón, D., Sada, Á., et al. (2021). Mandibular reconstruction with fibula flap and dental implants through virtual surgical planning and three different techniques: Double-barrel flap, implant dynamic navigation and CAD/CAM mesh with iliac crest graft. *Front. Oncol.* 11, 719712. doi:10.3389/fonc.2021.719712
- Attia, S., Wiltfang, J., Streckbein, P., Wilbrand, J.-F., El Khassawna, T., Mausbach, K., et al. (2019). Functional and aesthetic treatment outcomes after immediate jaw reconstruction using a fibula flap and dental implants. *J. Cranio-Maxillofac. Surg.* 47, 786–791. doi:10.1016/j.jcms.2018.12.017
- Azari, A., Schoenmaker, T., de Souza Faloni, A. P., Everts, V., and de Vries, T. J. (2011). Jaw and long bone marrow derived osteoclasts differ in shape and their response to bone and dentin. *Biochem. Biophys. Res. Commun.* 409, 205–210. doi:10.1016/j.bbrc.2011.04.120
- Barker, J. H. (1982). Lymphatic vessels in human alveolar bone. *Lymphology* 15, 1–13.
- Bortel, E., Grover, L. M., Eisenstein, N., Seim, C., Suhonen, H., Pacureanu, A., et al. (2022). Interconnectivity explains high canalicular network robustness between neighboring osteocyte lacunae in human bone. *Adv. NanoBiomed Res.* 2, 2100090. doi:10.1002/anbr.202100090
- Cakarar, S., Selvi, F., Isler, S. C., Soluk, M., Olgac, V., and Keskin, C. (2009). Intraosseous lipoma of the mandible: A case report and review of the literature. *Int. J. Oral Maxillofac. Surg.* 38, 900–902. doi:10.1016/j.ijom.2009.03.712
- Cardaropoli, G., Araújo, M., and Lindhe, J. (2003). Dynamics of bone tissue formation in tooth extraction sites. An experimental study in dogs. *J. Clin. Periodontol.* 30, 809–818. doi:10.1034/j.1600-051x.2003.00366.x
- Chai, Y., and Maxson, R. E. (2006). Recent advances in craniofacial morphogenesis. *Dev. Dyn.* 235, 2353–2375. doi:10.1002/dvdy.20833
- Chaichanasakul, T., Kang, B., Bezougliaia, O., Aghaloo, T. L., and Tetradis, S. (2014). Diverse osteoclastogenesis of bone marrow from mandible versus long bone. *J. Periodontol.* 85, 829–836. doi:10.1902/jop.2013.130376
- Chan, C. K. F., Gulati, G. S., Sinha, R., Tompkins, J. V., Lopez, M., Carter, A. C., et al. (2018). Identification of the human skeletal stem cell. *Cell* 175, 43–56. doi:10.1016/j.cell.2018.07.029
- Coutel, X., Delattre, J., Marchandise, P., Falgayrac, G., Béhal, H., Kerckhofs, G., et al. (2019). Mandibular bone is protected against microarchitectural alterations and bone marrow adipose conversion in ovariectomized rats. *Bone* 127, 343–352. doi:10.1016/j.bone.2019.06.031
- de Jong, W. C., Korfage, J. A. M., and Langenbach, G. E. J. (2010). Variations in habitual bone strains *in vivo*: Long bone versus mandible. *J. Struct. Biol.* 172, 311–318. doi:10.1016/j.jsb.2010.06.013
- de Souza Faloni, A. P., Schoenmaker, T., Azari, A., Katchburian, E., Cerri, P. S., de Vries, T. J., et al. (2011). Jaw and long bone marrows have a different osteoclastogenic potential. *Calcif. Tissue Int.* 88, 63–74. doi:10.1007/s00223-010-9418-4
- DeConde, A. S., Lee, M. K., Sidell, D., Aghaloo, T., Lee, M., Tetradis, S., et al. (2014). Defining the critical-sized defect in a rat segmental mandibulectomy model. *JAMA Otolaryngol. Neck Surg.* 140, 58–65. doi:10.1001/jamaoto.2013.5669
- Dong, W., Ge, J., Zhang, P., Fu, Y., Zhang, Z., Cheng, J., et al. (2014). Phenotypic characterization of craniofacial bone marrow stromal cells: Unique properties of enhanced osteogenesis, cell recruitment, autophagy, and apoptosis resistance. *Cell Tissue Res.* 358, 165–175. doi:10.1007/s00441-014-1927-4
- Drage, N. A., Palmer, R. M., Blake, G., Wilson, R., Crane, F., and Fogelman, I. (2007). A comparison of bone mineral density in the spine, hip and jaws of edentulous subjects. *Clin. Oral Implants Res.* 18, 496–500. doi:10.1111/j.1600-0501.2007.01379.x
- Eber, M. R., Park, S. H., Contino, K. F., Patel, C. M., Hsu, F.-C., and Shiozawa, Y. (2021). Osteoblasts derived from mouse mandible enhance tumor growth of prostate cancer more than osteoblasts derived from long bone. *J. Bone Oncol.* 26, 100346. doi:10.1016/j.jbo.2020.100346
- El-Sheikh, A. M., Abdel-Latif, H. H., Howell, P. G., and Hobkirk, J. A. (2007). Midline mandibular deformation during nonmasticatory functional movements in edentulous subjects with dental implants. *Int. J. Oral Maxillofac. Implants* 22, 243–248.
- Esfahanizadeh, N., Davaie, S., Rokn, A. R., Daneshparvar, H. R., Bayat, N., Khondi, N., et al. (2013). Correlation between bone mineral density of jaws and skeletal sites in an Iranian population using dual X-ray energy absorptiometry. *Dent. Res. J.* 10, 460–466.
- Fernandez de Grado, G., Keller, L., Idoux-Gillet, Y., Wagner, Q., Musset, A.-M., Benkirane-Jessel, N., et al. (2018). Bone substitutes: A review of their characteristics, clinical use, and perspectives for large bone defects management. *J. Tissue Eng.* 9, 2041731418776819. doi:10.1177/2041731418776819
- Fliedel, R., Tröltzsch, M., Kühnisch, J., Ehrenfeld, M., and Otto, S. (2015). Treatment strategies and outcomes of bisphosphonate-related osteonecrosis of the jaw (BRONJ) with characterization of patients: A systematic review. *Int. J. Oral Maxillofac. Surg.* 44, 568–585. doi:10.1016/j.ijom.2015.01.026
- Glatt, V., Evans, C. H., and Tetsworth, K. (2017). A concert between biology and biomechanics: The influence of the mechanical environment on bone healing. *Front. Physiol.* 7, 678. doi:10.3389/fphys.2016.00678
- Goldberg, S., Grynbas, M. D., and Glogauer, M. (2016). Heterogeneity of osteoclast activity and bone turnover in different skeletal sites. *Arch. Oral Biol.* 71, 134–143. doi:10.1016/j.archoralbio.2016.06.026
- Gong, X., Yu, W., Zhao, H., Su, J., and Sheng, Q. (2017). Skeletal site-specific effects of zoledronate on *in vivo* bone remodeling and *in vitro* BMSCs osteogenic activity. *Sci. Rep.* 7, 36129. doi:10.1038/srep36129
- Grigoriadis, A., Johansson, R. S., and Trulsson, M. (2014). Temporal profile and amplitude of human masseter muscle activity is adapted to food properties during individual chewing cycles. *J. Oral Rehabil.* 41, 367–373. doi:10.1111/joor.12155
- Groeneveldt, L. C., Herpelink, T., Maréchal, M., Politis, C., van IJcken, W. F. J., Huylebroeck, D., et al. (2020). The bone-forming properties of periosteum-derived cells differ between harvest sites. *Front. Cell Dev. Biol.* 8, 554984. doi:10.3389/fcell.2020.554984
- Guihard, P., Boutet, M.-A., Brounais-Le Royer, B., Gamblin, A.-L., Amiaud, J., Renaud, A., et al. (2015). Oncostatin M, an inflammatory cytokine produced by macrophages, supports intramembranous bone healing in a mouse model of tibia injury. *Am. J. Pathol.* 185, 765–775. doi:10.1016/j.ajpath.2014.11.008
- Hathaway-Schrader, J. D., Aartun, J. D., Poulides, N. A., Kuhn, M. B., McCormick, B. E., Chew, M. E., et al. (2022). Commensal oral microbiota induces osteoimmunomodulatory effects separate from systemic microbiome in mice. *JCI Insight* 7, e140738. doi:10.1172/jci.insight.140738
- Hesse, B., Langer, M., Varga, P., Pacureanu, A., Dong, P., Schrof, S., et al. (2014). Alterations of mass density and 3D osteocyte lacunar properties in bisphosphonate-related osteonecrotic human jaw bone, a synchrotron μ CT study. *PLoS One* 9, e88481. doi:10.1371/journal.pone.0088481
- Hesse, B., Varga, P., Langer, M., Pacureanu, A., Schrof, S., Männicke, N., et al. (2015). Canalicular network morphology is the major determinant of the spatial distribution of mass density in human bone tissue: Evidence by means of synchrotron radiation phase-contrast nano-CT. *J. Bone Min. Res.* 30, 346–356. doi:10.1002/jbmr.2324
- Hinton, R. J., Jing, Y., Jing, J., and Feng, J. Q. (2017). Roles of chondrocytes in endochondral bone formation and fracture repair. *J. Dent. Res.* 96, 23–30. doi:10.1177/0022034516668321
- Horner, K., Devlin, H., Alsop, C. W., Hodgkinson, I. M., and Adams, J. E. (1996). Mandibular bone mineral density as a predictor of skeletal osteoporosis. *Br. J. Radiol.* 69, 1019–1025. doi:10.1259/0007-1285-69-827-1019
- Horowitz, M. C., Fretz, J. A., and Lorenzo, J. A. (2010). How B cells influence bone biology in health and disease. *Bone* 47, 472–479. doi:10.1016/j.bone.2010.06.011
- Hsu, P.-Y., Tsai, M.-T., Wang, S.-P., Chen, Y.-J., Wu, J., and Hsu, J.-T. (2016). Cortical bone morphological and trabecular bone microarchitectural changes in the mandible and femoral neck of ovariectomized rats. *PLoS One* 11, e0154367. doi:10.1371/journal.pone.0154367
- Huja, S. S., Fernandez, S. A., Hill, K. J., and Li, Y. (2006). Remodeling dynamics in the alveolar process in skeletally mature dogs. *Anat. Rec. A Discov. Mol. Cell. Evol. Biol.* 288A, 1243–1249. doi:10.1002/ar.a.20396
- Ichikawa, Y., Watahiki, J., Nampo, T., Nose, K., Yamamoto, G., Irie, T., et al. (2015). Differences in the developmental origins of the periosteum may influence bone healing. *J. Periodontol. Res.* 50, 468–478. doi:10.1111/jre.12229
- Ide, Y., Matsunaga, S., Harris, J., Connell, D. O., Seikaly, H., and Wolfaardt, J. (2015). Anatomical examination of the fibula: Digital imaging study for osseointegrated implant installation. *J. Otolaryngol. Head. Neck Surg.* 44, 1. doi:10.1186/s40463-015-0055-9
- Inoue, M., Ono, T., Kameo, Y., Sasaki, F., Ono, T., Adachi, T., et al. (2019). Forceful mastication activates osteocytes and builds a stout jawbone. *Sci. Rep.* 9, 4404. doi:10.1038/s41598-019-40463-3
- Isaac, J., Lignon, G., Jacques, J., and Bernal, A. (2017). “Ameloblastin as biomarker of bone,” in *Biomarkers in bone disease. Biomarkers in disease: Methods, discoveries and applications*. Editors V. B. Patel and V. R. Preedy (Dordrecht: Springer Netherlands), 267–300. doi:10.1007/978-94-007-7693-7_19
- Ito, S., Kasahara, N., Kitamura, K., Matsunaga, S., Mizoguchi, T., Htun, M. W., et al. (2022). Pathological differences in the bone healing processes between tooth extraction socket and femoral fracture. *Bone Rep.* 16, 101522. doi:10.1016/j.bonr.2022.101522
- Jensen, R. L., and Folke, L. E. A. (1974). The passage of exogenous tritiated thymidine into gingival tissues. *J. Periodontol.* 45, 786–791. doi:10.1902/jop.1974.45.11.786
- Kang, Y.-H., Kim, H.-M., Byun, J.-H., Kim, U.-K., Sung, I.-Y., Cho, Y.-C., et al. (2015). Stability of simultaneously placed dental implants with autologous bone grafts harvested from the iliac crest or intraoral jaw bone. *BMC Oral Health* 15, 172. doi:10.1186/s12903-015-0156-x
- Kelder, C., Kleverlaan, C. J., Gilijamse, M., Bakker, A. D., and de Vries, T. J. (2020). Cells derived from human long bone appear more differentiated and more actively stimulate osteoclastogenesis compared to alveolar bone-derived cells. *Int. J. Mol. Sci.* 21, 5072. doi:10.3390/ijms21145072
- Knitschke, M., Sonnabend, S., Bäcker, C., Schmermund, D., Böttger, S., Howaldt, H.-P., et al. (2021). Partial and total flap failure after fibula free flap in head and neck reconstructive surgery: Retrospective analysis of 180 flaps over 19 years. *Cancers* 13, 865. doi:10.3390/cancers13040865

- Koerdt, S., Dax, S., Grimaldi, H., Ristow, O., Kuebler, A. C., and Reuther, T. (2014). Histomorphologic characteristics of bisphosphonate-related osteonecrosis of the jaw. *J. Oral Pathol. Med.* 43, 448–453. doi:10.1111/jop.12156
- Kwack, K. H., Lamb, N. A., Bard, J. E., Kramer, E. D., Zhang, L., Abrams, S. I., et al. (2021). Discovering myeloid cell heterogeneity in mandibular bone – cell by cell analysis. *Front. Physiol.* 12, 731549. doi:10.3389/fphys.2021.731549
- Lee, D. J., Kwon, J., Current, L., Yoon, K., Zalal, R., Hu, X., et al. (2019). Osteogenic potential of mesenchymal stem cells from rat mandible to regenerate critical sized calvarial defect. *J. Tissue Eng.* 10, 2041731419830427. doi:10.1177/2041731419830427
- Lee, J.-T., Choi, S.-Y., Kim, H.-L., Kim, J.-Y., Lee, H.-J., and Kwon, T.-G. (2015). Comparison of gene expression between mandibular and iliac bone-derived cells. *Clin. Oral Investig.* 19, 1223–1233. doi:10.1007/s00784-014-1353-8
- Leucht, P., Kim, J.-B., Amasha, R., James, A. W., Girod, S., and Helms, J. A. (2008). Embryonic origin and Hox status determine progenitor cell fate during adult bone regeneration. *Development* 135, 2845–2854. doi:10.1242/dev.023788
- Li, C., Wang, F., Zhang, R., Qiao, P., and Liu, H. (2020). Comparison of proliferation and osteogenic differentiation potential of rat mandibular and femoral bone marrow mesenchymal stem cells *in vitro*. *Stem Cells Dev.* 29, 728–736. doi:10.1089/scd.2019.0256
- Lin, W., Li, Q., Zhang, D., Zhang, X., Qi, X., Wang, Q., et al. (2021). Mapping the immune microenvironment for mandibular alveolar bone homeostasis at single-cell resolution. *Bone Res.* 9, 17–12. doi:10.1038/s41413-021-00141-5
- Liu, L.-N., Zhang, X.-H., Liu, H.-H., Li, K.-H., Wu, Q.-H., Liu, Y., et al. (2020). Osteogenesis differences around titanium implant and in bone defect between jaw bones and long bones. *J. Craniofac. Surg.* 31, 2193–2198. doi:10.1097/SCS.00000000000006795
- Lloyd, B., Tee, B. C., Headley, C., Emam, H., Mallery, S., and Sun, Z. (2017). Similarities and differences between porcine mandibular and limb bone marrow mesenchymal stem cells. *Arch. Oral Biol.* 77, 1–11. doi:10.1016/j.archoralbio.2017.01.012
- Marini, M., Bertolai, R., Ambrosini, S., Sarchielli, E., Vannelli, G. B., and Sgambati, E. (2015). Differential expression of vascular endothelial growth factor in human fetal skeletal site-specific tissues: Mandible versus femur. *Acta Histochem* 117, 228–234. doi:10.1016/j.acthis.2015.02.006
- Mark, M., Rijli, F. M., and Chambon, P. (1997). Homeobox genes in embryogenesis and pathogenesis. *Pediatr. Res.* 42, 421–429. doi:10.1203/00006450-199710000-00001
- Marolt, D., Cozin, M., Vunjak-Novakovic, G., Cremers, S., and Landesberg, R. (2012). Effects of pamidronate on human alveolar osteoblasts *in vitro*. *J. Oral Maxillofac. Surg.* 70, 1081–1092. doi:10.1016/j.joms.2011.05.002
- Matsubara, T., Suardita, K., Ishii, M., Sugiyama, M., Igarashi, A., Oda, R., et al. (2004). Alveolar bone marrow as a cell source for regenerative medicine: Differences between alveolar and iliac bone marrow stromal cells. *J. Bone Min. Res.* 20, 399–409. doi:10.1359/JBMR.041117
- Mavropoulos, A., Rizzoli, R., and Ammann, P. (2007). Different responsiveness of alveolar and tibial bone to bone loss stimuli. *J. Bone Min. Res.* 22, 403–410. doi:10.1359/jbmr.061208
- Moriishi, T., Ozasa, R., Ishimoto, T., Nakano, T., Hasegawa, T., Miyazaki, T., et al. (2020). Osteocalcin is necessary for the alignment of apatite crystallites, but not glucose metabolism, testosterone synthesis, or muscle mass. *PLoS Genet.* 16, e1008586. doi:10.1371/journal.pgen.1008586
- Nakamoto, T., and Miller, S. A. (1977). Effect of protein-energy malnutrition on the growth of mandible and long bone in newborn male and female rats. *J. Nutr.* 107, 983–989. doi:10.1093/jn/107.6.983
- Nakamoto, T., and Miller, S. A. (1979). Physical and biochemical changes of the mandible and long bone in protein-energy malnourished newborn rats. *J. Nutr.* 109, 1477–1482. doi:10.1093/jn/109.8.1477
- Nicolielo, L. F. P., Jacobs, R., Ali Albdour, E., Hoste, X., Abeloos, J., Politis, C., et al. (2017). Is oestrogen associated with mandibular condylar resorption? A systematic review. *Int. J. Oral Maxillofac. Surg.* 46, 1394–1402. doi:10.1016/j.ijom.2017.06.012
- Nyström, M., and Ranta, H. (2003). Tooth formation and the mandibular symphysis during the first five postnatal months. *J. Forensic Sci.* 48, 2002274–2002378. doi:10.1520/jfs2002274
- Oettlé, A. C., Ehlers, R., and Steyn, M. (2016). Changes in the mandibular angle during adulthood in South Africans: Changes in the mandibular angle in adults. *Am. J. Hum. Biol.* 28, 681–686. doi:10.1002/ajhb.22850
- Pacifici, R. (2016). T cells, osteoblasts, and osteocytes: Interacting lineages key for the bone anabolic and catabolic activities of parathyroid hormone. *Ann. N. Y. Acad. Sci.* 1364, 11–24. doi:10.1111/nyas.12969
- Pickrell, B., Serebrakian, A., and Maricevich, R. (2017). Mandible fractures. *Semin. Plast. Surg.* 31, 100–107. doi:10.1055/s-0037-1601374
- Pravitharangul, A., Suttapreyasri, S., and Leethanakul, C. (2019). Mandible and iliac osteoblasts exhibit different Wnt signaling responses to LMHF vibration. *J. Oral Biol. Craniofacial Res.* 9, 355–359. doi:10.1016/j.jobcr.2019.09.005
- Price, D. M. (2008). Protein-energy malnutrition among the elderly: Implications for nursing care. *Holist. Nurs. Pract.* 22, 355–360. doi:10.1097/01.HNP.0000339347.03629.41
- Ransom, R. C., Carter, A. C., Salhotra, A., Leavitt, T., Marecic, O., Murphy, M. P., et al. (2018). Mechanoresponsive stem cells acquire neural crest fate in jaw regeneration. *Nature* 563, 514–521. doi:10.1038/s41586-018-0650-9
- Reichert, J. C., Gohlke, J., Friis, T. E., Quent, V. M. C., and Huttmacher, D. W. (2013). Mesodermal and neural crest derived ovine tibial and mandibular osteoblasts display distinct molecular differences. *Gene* 525, 99–106. doi:10.1016/j.gene.2013.04.026
- Ristow, O., Gerngroß, C., Schwaiger, M., Hohlweg-Majert, B., Kehl, V., Jansen, H., et al. (2014a). Effect of antiresorptive drugs on bony turnover in the jaw: Denosumab compared with bisphosphonates. *Br. J. Oral Maxillofac. Surg.* 52, 308–313. doi:10.1016/j.bjoms.2014.01.021
- Ristow, O., Gerngroß, C., Schwaiger, M., Hohlweg-Majert, B., Kehl, V., Jansen, H., et al. (2014b). Is bone turnover of jawbone and its possible over suppression by bisphosphonates of etiologic importance in pathogenesis of bisphosphonate-related osteonecrosis? *J. Oral Maxillofac. Surg.* 72, 903–910. doi:10.1016/j.joms.2013.11.005
- Rizzoli, R., and Bonjour, J.-P. (2004). Dietary protein and bone health. *J. Bone Min. Res.* 19, 527–531. doi:10.1359/JBMR.040204
- Roberts, T. T., and Rosenbaum, A. J. (2012). Bone grafts, bone substitutes and orthobiologics: The bridge between basic science and clinical advancements in fracture healing. *Organogenesis* 8, 114–124. doi:10.4161/org.23306
- Rothweiler, R., Gross, C., Bortel, E., Früh, S., Gerber, J., Boller, E., et al. (2022). Comparison of the 3D-microstructure between alveolar and iliac bone for enhanced bioinspired bone graft substitutes. *Front. Bioeng. Biotechnol.* 10, 862395. doi:10.3389/fbioe.2022.862395
- Roybal, P. G., Wu, N. L., Sun, J., Ting, M., Schafer, C. A., and Maxson, R. E. (2010). Inactivation of Msx1 and Msx2 in neural crest reveals an unexpected role in suppressing heterotopic bone formation in the head. *Dev. Biol.* 343, 28–39. doi:10.1016/j.ydbio.2010.04.007
- Rubin, C. T., and Lanyon, L. E. (1984). Regulation of bone formation by applied dynamic loads. *J. Bone Jt. Surg. Am.* 66, 397–402. doi:10.2106/00004623-198466030-00012
- Sacco, R., Sacco, N., Hamid, U., Ali, S. H., Singh, M., and Blythe, J. St. J. (2018). Microsurgical reconstruction of the jaws using vascularised free flap technique in patients with medication-related osteonecrosis: A systematic review. *Biomed. Res. Int.* 2018, 9858921–9859012. doi:10.1155/2018/9858921
- Sasaki, M., Matsuura, T., Katafuchi, M., Tokutomi, K., and Sato, H. (2010). Higher contents of mineral and collagen but lower of hydroxylysine of collagen in mandibular bone compared with those of humeral and femoral bones in human. *J. Hard Tissue Biol.* 19, 175–180. doi:10.2485/jhtb.19.175
- Sbordone, L., Toti, P., Menchini-Fabris, G., Sbordone, C., and Guidetti, F. (2009). Implant survival in maxillary and mandibular osseous onlay grafts and native bone: A 3-year clinical and computerized tomographic follow-up. *Int. J. Oral Maxillofac. Implants* 24, 695–703.
- Schlundt, C., Fischer, H., Bucher, C. H., Rendenbach, C., Duda, G. N., and Schmidt-Bleek, K. (2021). The multifaceted roles of macrophages in bone regeneration: A story of polarization, activation and time. *Acta Biomater.* 133, 46–57. doi:10.1016/j.actbio.2021.04.052
- Sciote, J. J., Horton, M. J., Rowleson, A. M., and Link, J. (2003). Specialized cranial muscles: How different are they from limb and abdominal muscles? *Cells Tissues Organs* 174, 73–86. doi:10.1159/000070576
- Sculean, A., Stavropoulos, A., and Bosshardt, D. D. (2019). Self-regenerative capacity of intra-oral bone defects. *J. Clin. Periodontol.* 46 (21), 70–81. doi:10.1111/jcpe.13075
- Shah, H. N., Jones, R. E., Borrelli, M. R., Robertson, K., Salhotra, A., Wan, D. C., et al. (2021). Craniofacial and long bone development in the context of distraction osteogenesis. *Plast. Reconstr. Surg.* 147, 54e–65e. doi:10.1097/PRS.00000000000007451
- Slootweg, P. J. (2010). Bone diseases of the jaws. *Int. J. Dent.* 2010, 702314–702317. doi:10.1155/2010/702314
- Son, C., Choi, M. S., and Park, J. (2020). Different responsiveness of alveolar bone and long bone to epithelial-mesenchymal interaction-related factor. *JBMR Plus* 4, e10382. doi:10.1002/jbmr.4.10382
- Springe, B., Slaidina, A., Soboleva, U., and Lejniaks, A. (2014). Bone mineral density and mandibular residual ridge resorption. *Int. J. Prosthodont.* 27, 270–276. doi:10.11607/ijp.3283
- Stefanik, D., Sarin, J., Lam, T., Levin, L., Leboy, P. S., and Akintoye, S. O. (2008). Disparate osteogenic response of mandible and iliac crest bone marrow stromal cells to pamidronate. *Oral Dis.* 14, 465–471. doi:10.1111/j.1601-0825.2007.01402.x
- Troeltzsch, M., Troeltzsch, M., Kauffmann, P., Gruber, R., Brockmeyer, P., Moser, N., et al. (2016). Clinical efficacy of grafting materials in alveolar ridge augmentation: A systematic review. *J. Cranio-Maxillofac. Surg.* 44, 1618–1629. doi:10.1016/j.jcms.2016.07.028

- Trombelli, L., Farina, R., Marzola, A., Bozzi, L., Liljenberg, B., and Lindhe, J. (2008). Modeling and remodeling of human extraction sockets. *J. Clin. Periodontol.* 35, 630–639. doi:10.1111/j.1600-051X.2008.01246.x
- Verhelst, P.-J., Dons, F., Van Bever, P.-J., Schoenaers, J., Nanhekhan, L., and Politis, C. (2019). Fibula free flap in head and neck reconstruction: Identifying risk factors for flap failure and analysis of postoperative complications in a low volume setting. *Craniomaxillofacial Trauma Reconstr.* 12, 183–192. doi:10.1055/s-0038-1651515
- Wang, F., Zhou, Y., Zhou, J., Xu, M., Zheng, W., Huang, W., et al. (2018). Comparison of intraoral bone regeneration with iliac and alveolar BMSCs. *J. Dent. Res.* 97, 1229–1235. doi:10.1177/0022034518772283
- Wang, J. Y., Huo, L., Yu, R. Q., Rao, N. J., Lu, W. W., and Zheng, L. W. (2019). Skeletal site-specific response of jawbones and long bones to surgical interventions in rats treated with zoledronic acid. *Biomed. Res. Int.* 2019, 5138175. doi:10.1155/2019/5138175
- Watabe, H., Furuhashi, T., Tani-Ishii, N., and Mikuni-Takagaki, Y. (2011). Mechanotransduction activates $\alpha\beta_1$ integrin and PI3K/Akt signaling pathways in mandibular osteoblasts. *Exp. Cell Res.* 317, 2642–2649. doi:10.1016/j.yexcr.2011.07.015
- Wehrhan, F., Amann, K., Möbius, P., Weber, M., Preidl, R., Ries, J., et al. (2015). BRONJ-Related jaw bone is associated with increased *dlx-5* and suppressed osteopontin—Implication in the site-specific alteration of angiogenesis and bone turnover by bisphosphonates. *Clin. Oral Investig.* 19, 1289–1298. doi:10.1007/s00784-014-1354-7
- Wehrhan, F., Hyclel, P., Amann, K., Ries, J., Stockmann, P., Schlegel, K., et al. (2011). *Msx-1* is suppressed in bisphosphonate-exposed jaw bone analysis of bone turnover-related cell signalling after bisphosphonate treatment: ONJ and *msx-1* expression. *Oral Dis.* 17, 433–442. doi:10.1111/j.1601-0825.2010.01778.x
- Wein, M., Fretwurst, T., Nahles, S., Duttonhoefer, F., Tomakidi, P., Steinberg, T., et al. (2015). Pilot investigation of the molecular discrimination of human osteoblasts from different bone entities. *J. Craniomaxillofac. Surg.* 43, 1487–1493. doi:10.1016/j.jcms.2015.07.030
- Wein, M., Huelter-Hassler, D., Nelson, K., Fretwurst, T., Nahles, S., Finkenzeller, G., et al. (2019). Differential osteopontin expression in human osteoblasts derived from iliac crest and alveolar bone and its role in early stages of angiogenesis. *J. Bone Min. Metab.* 37, 105–117. doi:10.1007/s00774-017-0900-1
- Wong, S. A., Hu, D. P., Slocum, J., Lam, C., Nguyen, M., Miclau, T., et al. (2021). Chondrocyte-to-osteoblast transformation in mandibular fracture repair. *J. Orthop. Res.* 39, 1622–1632. doi:10.1002/jor.24904
- Xi, Q., Liang, L. M., and Bu, R. F. (2008). Styloid process osteochondroma. *Int. J. Oral Maxillofac. Surg.* 37, 587–589. doi:10.1016/j.ijom.2008.01.003
- Yamaza, T., Ren, G., Akiyama, K., Chen, C., Shi, Y., and Shi, S. (2011). Mouse mandible contains distinctive mesenchymal stem cells. *J. Dent. Res.* 90, 317–324. doi:10.1177/0022034510387796
- Yang, X., Jiang, J., Zhou, L., Wang, S., He, M., Luo, K., et al. (2019). Osteogenic and angiogenic characterization of mandible and femur osteoblasts. *J. Mol. Histol.* 50, 105–117. doi:10.1007/s10735-019-09810-6
- Youtlen, S. E., Kemp, J. P., Logan, J. G., Ghirardello, E. J., Sergio, C. M., Dack, M. R. G., et al. (2021). Osteocyte transcriptome mapping identifies a molecular landscape controlling skeletal homeostasis and susceptibility to skeletal disease. *Nat. Commun.* 12, 2444. doi:10.1038/s41467-021-22517-1
- Yuan, Y., and Chai, Y. (2019). Regulatory mechanisms of jaw bone and tooth development. *Curr. Top. Dev. Biol.* 133, 91–118. doi:10.1016/bs.ctdb.2018.12.013
- Zalc, A., Sinha, R., Gulati, G. S., Wesche, D. J., Daszczuk, P., Swigut, T., et al. (2021). Reactivation of the pluripotency program precedes formation of the cranial neural crest. *Science* 371, eabb4776. doi:10.1126/science.abb4776
- Zhang, Z., Song, Y., Zhang, X., Tang, J., Chen, J., and Chen, Y. (2003). *Msx1/Bmp4* genetic pathway regulates mammalian alveolar bone formation via induction of *Dlx5* and *Cbfa1*. *Mech. Dev.* 120, 1469–1479. doi:10.1016/j.mod.2003.09.002