



OPEN ACCESS

EDITED AND REVIEWED BY Giuseppe D'Antona, University of Pavia, Italy

SPECIALTY SECTION

This article was submitted to Exercise Physiology,

to Exercise Physiology, a section of the journal Frontiers in Physiology

RECEIVED 06 March 2023 ACCEPTED 09 March 2023 PUBLISHED 14 March 2023

CITATION

Verboven K and Vechetti IJ (2023), Editorial: Inter-organ crosstalk during exercise in health and disease: Extracellular vesicles as new kids on the block.

Front. Physiol. 14:1180972. doi: 10.3389/fphys.2023.1180972

COPYRIGHT

© 2023 Verboven and Vechetti. This is an open-access article distributed under the terms of the Creative Commons
Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: Inter-organ crosstalk during exercise in health and disease: Extracellular vesicles as new kids on the block

Kenneth Verboven^{1,2}* and Ivan J. Vechetti³

¹REVAL—Rehabilitation Research Center, Faculty of Rehabilitation Sciences, Hasselt University, Diepenbeek, Belgium, ²BIOMED—Biomedical Research Institute, Faculty of Medicine and Life Sciences, Hasselt University, Diepenbeek, Belgium, ³Department of Nutrition and Health Sciences, College of Education and Human Sciences, University of Nebraska-Lincoln, Lincoln, NE, United States

KEYWORDS

extracellular vesicle (EV), exercise, myokines, exosomes, microRNA (microRNA)

Editorial on the Research Topic

Inter-organ crosstalk during exercise in health and disease: Extracellular vesicles as new kids on the block

Within exercise physiology, the study of factors potentially mediating interorgan crosstalk during and after exercise is a fascinating field of research. As exercise activates a plethora of metabolic pathways in several tissues, organs and systems, examining the underlying biological mechanisms contributing to exercise related metabolic benefits is imperative. Since two decades, the skeletal muscle is known to secrete humoral factors into the circulation in response to exercise, originally described as "myokines" by Pedersen et al. (2003). These myokines are now well known and extensively studied in the field of exercise science (Pedersen and Febbraio, 2012). Interestingly, exercise also triggers other metabolic organs to release similar factors arising from the heart, liver, white and brown adipose tissue, and the nervous system (Chow et al., 2022). These "exerkines" (Safdar et al., 2016) have been recognized to comprise an extensive range of biologically active signalling molecules, including cytokines, lipids, metabolites and (noncoding) nucleic acids, as recently reviewed (Chow et al., 2022).

Extracellular vesicles (EVs) and their role as carrier particles for molecular signals became of specific interest in the exerkine field, as EVs are considered (co-)drivers of exercise-induced interorgan crosstalk (Whitham et al., 2018; Vechetti et al., 2021). Differentiated by both their size and nature of vesicular biogenesis, EVs can be primarily classified as exosomes, microvesicles and apoptotic bodies although some overlap does exist between these classifications. EVs may enclose plenty of material, including lipids, proteins and nucleic acids (Théry et al., 2018). Indeed, pioneering EV-related exercise studies have shown an increase in the circulating number of EVs after a single bout of exercise (Brahmer et al., 2019; Frühbeis et al., 2015; Oliveira et al., 2018; Whitham et al., 2018), with recent *in vivo* research estimating about 5% of circulating, tetraspanin-positive EVs to be muscle-derived (Estrada et al., 2022). However, the frequent lack of rigorous characterization, purification and/or quantification of EVs (which ideally requires a combination of multiple methodologies) makes the understanding of the role of EVs in exercise physiology rather hard (Darragh et al., 2021) and argues for standard approaches and reporting on EV-related exercise science. Nevertheless, many points need to be clarified, but as the interest in EVs research from an exercise and health

Verboven and Vechetti 10.3389/fphys.2023.1180972

perspective is still growing, many researchers are currently trying to understand the mechanisms involved in generation, cell-specific release and uptake of EVs in both health and disease. We therefore proposed a Frontiers Research Topic to present some novel research on the role of EVs in interorgan crosstalk during exercise in health and disease, which resulted in the current Research Topic of 3 original research papers and 1 review paper.

The review by Nederveen et al. discusses the current understanding of the effects of exercise on EVs. Based on existing literature, this review supports the ability of skeletal muscle tissue to secrete bioactive EVs, although tracking cell-specific origin of systemic EVs remains to be elucidated, as well as what population of EVs (either microvesicles or exosomes) are transporting these bioactive signalling molecules. Although the field of exercise EVs is still in its infancy, several factors potentially influencing EV dynamics during and following acute/chronic exercise have been summarized by Nederveen et al., including exercise intensity, training and health status, concomitantly taking into account methodological heterogeneity (with respect to sample Research Topic and handling, but also isolation, purification and quantification of EVs) among existing studies.

In relation to exercise intensity, Kobayashi et al. examined changes in circulating number and proteomic profile of high-intensity interval (HIIT) exercise-induced EVs in a small set of young healthy, physically active males. HIIT exercise rapidly augmented circulatory EVs in the post-exercise phase (i.e., 30 and 120 min after HIIT, respectively), originating from skeletal muscle, liver and adipose tissue, a finding which was suggested to be related to processing by their target cells or degradation. Proteomic analyses of these EVs indicated a prompt elevation of proteins implicated in coagulation cascades, acid-base homeostasis and antioxidative pathways.

Vann et al. used myobundles, a three-dimensional tissue-engineered model of human skeletal muscle, to investigate changes in sarcoplasmic and secretory microRNA expression (miRNA sequencing), the latter assessed in culture medium derived EVs, in response to exercise-mimetic contractile activity *in vitro*. Their findings showed a differential microRNA expression profile (n=152 microRNAs) between myobundles and myobundle derived EVs, of which some were differentially responsive to chronic low frequency stimulation (miRNA-543, -487b-3p, and -6511-3p) or intermittent high frequency stimulation (miRNA-6511-3p, and -543) between myobundles and EVs. As such, Vann et al. provide novel microRNA targets to which the effects of exercise (training) could be explored in future (human) research.

As exercise is a potent modifier of skeletal muscle EV secretion and content in both health and disease, Vechetti et al. found circulating EVs showed similar morphology, but lower concentration, in individuals with cerebral palsy (CP) compared

to typically developed individuals, both at rest and following acute aerobic exercise. Aside from congruent morphology, the skeletal muscle-specific microRNAs miRNA-486 was upregulated in CP, irrespective of the exercise stimulus. miRNA-486 might be an important regulator of satellite cells, thereby affecting extracellular matrix and sarcomerogenesis related genes (such as *Pax-7*) and thus contributing to the known CP-related skeletal muscle alterations.

By proposing this Research Topic, our initial goal was to attract manuscripts focusing on the role of the EVs and their responsiveness to the physiological and metabolic adaptations during exercise (training). Ultimately, despite the body of work collected, we only managed to scratch the surface of this rapidly evolving and interesting field in exercise science. Future studies will bring us closer to unravelling the function of the exercise induced EVs in health and diseases.

Author contributions

KV and IV conceived, wrote, and approved the editorial for publication. All authors contributed to the article and approved the submitted version.

Acknowledgments

We sincerely thank all authors and reviewers who participated in the Research Topic.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Brahmer, A., Neuberger, E., Esch-Heisser, L., Haller, N., Jorgensen, M. M., Baek, R., et al. (2019). Platelets, endothelial cells and leukocytes contribute to the exercise-triggered release of extracellular vesicles into the circulation. *J. Extracell. Vesicles* 8 (1), 1615820. doi:10.1080/20013078.2019.1615820

Chow, L. S., Gerszten, R. E., Taylor, J. M., Pedersen, B. K., van Praag, H., Trappe, S., et al. (2022). Exerkines in health, resilience and disease. *Nat. Rev. Endocrinol.* 18 (5), 273–289. doi:10.1038/s41574-022-00641-2

Darragh, I. A. J., O'Driscoll, L., and Egan, B. (2021). Exercise training and circulating small extracellular vesicles: Appraisal of methodological approaches and current knowledge. *Front. Physiol.* 12, 738333. doi:10.3389/fphys.2021.738333

Estrada, A. L., Valenti, Z. J., Hehn, G., Amorese, A. J., Williams, N. S., Balestrieri, N. P., et al. (2022). Extracellular vesicle secretion is tissue-dependent *ex vivo* and skeletal muscle myofiber extracellular vesicles reach the circulation *in vivo*. *Am. J. Physiol. Cell Physiol.* 322 (2), C246–C259. doi:10.1152/ajpcell.00580.2020

Frühbeis, C., Helmig, S., Tug, S., Simon, P., and Krämer-Albers, E. M. (2015). Physical exercise induces rapid release of small extracellular vesicles into the circulation. *J. Extracell. Vesicles* 4, 28239. doi:10.3402/jev.v4.28239

Oliveira, G. P., Jr, Porto, W. F., Palu, C. C., Pereira, L. M., Petriz, B., Almeida, J. A., et al. (2018). Effects of acute aerobic exercise on rats serum extracellular vesicles diameter, concentration and small RNAs content. *Front. Physiol.* 9, 532. doi:10.3389/fphys.2018.00532

Verboven and Vechetti 10.3389/fphys.2023.1180972

Pedersen, B. K., and Febbraio, M. A. (2012). Muscles, exercise and obesity: Skeletal muscle as a secretory organ. *Nat. Rev. Endocrinol.* 8 (8), 457–465. doi:10.1038/nrendo.2012.49

Pedersen, B. K., Steensberg, A., Fischer, C., Keller, C., Keller, P., Plomgaard, P., et al. (2003). Searching for the exercise factor: Is IL-6 a candidate? *J. Muscle Res. Cell Motil.* 24 (2-3), 113–119. doi:10.1023/a:1026070911202

Safdar, A., Saleem, A., and Tarnopolsky, M. A. (2016). The potential of endurance exercise-derived exosomes to treat metabolic diseases. *Nat. Rev. Endocrinol.* 12 (9), 504–517. doi:10.1038/nrendo.2016.76

Théry, C., Witwer, K. W., Aikawa, E., Alcaraz, M. J., Anderson, J. D., Andriantsitohaina, R., et al. (2018). Minimal information for studies of extracellular

vesicles 2018 (MISEV2018): A position statement of the international society for extracellular vesicles and update of the MISEV2014 guidelines. *J. Extracell. Vesicles* 7 (1), 1535750. doi:10.1080/20013078.2018.1535750

Vechetti, I. J., Jr, Valentino, T., Mobley, C. B., and McCarthy, J. J. (2021). The role of extracellular vesicles in skeletal muscle and systematic adaptation to exercise. *J. Physiol.* 599 (3), 845–861. doi:10.1113/JP278929

Whitham, M., Parker, B. L., Friedrichsen, M., Hingst, J. R., Hjorth, M., Hughes, W. E., et al. (2018). Extracellular vesicles provide a means for tissue crosstalk during exercise. *Cell Metab.* 27 (1), 237–251. doi:10.1016/j.cmet.2017.12.001