Check for updates

OPEN ACCESS

EDITED AND REVIEWED BY Pung Pung Hwang, Academia Sinica, Taiwan

*CORRESPONDENCE Davide Seveso, adavide.seveso@unimib.it

RECEIVED 31 July 2024 ACCEPTED 09 August 2024 PUBLISHED 19 August 2024

CITATION

Seveso D, Louis YD, Bhagooli R, Downs CA and Dellisanti W (2024) Editorial: The cellular stress response and physiological adaptations of corals subjected to environmental stressors and pollutants, volume II. *Front. Physiol.* 15:1473792. doi: 10.3389/fphys.2024.1473792

COPYRIGHT

© 2024 Seveso, Louis, Bhagooli, Downs and Dellisanti. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: The cellular stress response and physiological adaptations of corals subjected to environmental stressors and pollutants, volume II

Davide Seveso^{1,2}*, Yohan D. Louis^{1,2}, Ranjeet Bhagooli^{3,4,5,6,7}, Craig A. Downs⁸ and Walter Dellisanti⁹

¹Department of Earth and Environmental Science, University of Milano Bicocca, Milano, Italy, ²MaRHE Center (Marine Research and High Education Center), Magoodhoo, Faafu, Maldives, ³Department of Biosciences and Ocean Studies, Faculty of Science and Pole of Research Excellence in Sustainable Marine Biodiversity, University of Mauritius, Réduit, Mauritius, ⁴The Biodiversity and Environment Institute, Réduit, Mauritius, ⁵Institute of Oceanography and Environment (INOS), University Malaysia Terengganu, Kuala Terengganu, Terengganu, Malaysia, ⁶Department of Marine Science, Faculty of Fisheries and Marine Science, Diponegoro University, Semarang, Indonesia, ⁷The Society of Biology (Mauritius), Réduit, Mauritius, ⁸Haereticus Environmental Laboratory, Clifford, VA, United States, ⁹Marine Biology Section, Department of Biology, University of Copenhagen, Helsingar, Denmark

KEYWORDS

cellular stress, corals, biomarkers, climate change, anthropogenic impact

Editorial on the Research Topic

The cellular stress response and physiological adaptations of corals subjected to environmental stressors and pollutants, volume II

There is substantial evidence that coral reefs are suffering worldwide due to global climate change, anthropogenic pressures, and local stressors, which led to their rapid decline over the last few decades with bleaching causing most of this loss (Hughes et al., 2017; Hughes et al., 2018). In order to more accurately predict the impacts of global changes and develop conservation and stress mitigation strategies, efforts have recently increased in elucidating the cellular and molecular mechanisms underlying coral bleaching and other coral responses to environmental stressors (Helgoe et al., 2024). As sessile and long-lived animals that experience variable conditions, corals rely mainly on their cellular stress responses for acclimatization and adaptation (Drury, 2020). Moreover, since changes at the cellular level are the first detectable responses to environmental perturbations, the analysis of cellular biomarkers represents a useful diagnostic tool reflecting variations in cellular integrity and pathways before larger-scale processes are affected (Downs, 2005; Louis et al., 2020; Montalbetti et al., 2021).

Although recent and substantial advances in omics technologies have made the study of coral molecular processes more efficient, rapid, and accessible (Weis, 2019; Cziesielski et al., 2020), our understanding of coral cell biology remains inadequate (Oakley and Davy, 2018; Weis, 2019). This Research Topic aimed to expand this knowledge and bridge existing gaps. The articles presented here demonstrate how various physiological and molecular approaches and techniques can be adopted to understand the responses of coral holobionts to a multitude of stressors.

Sea surface temperature increase and heat waves are the primary drivers of coral bleaching and reef degradation worldwide (Hughes et al., 2018; Eakin et al., 2019). Therefore, mesophotic habitats often represent potential refugia for corals (Bongaerts et al., 2010; Muir et al., 2017). In their study, Tavakoli-Kolour et al. analyzed the photosynthetic efficiency (maximum quantum yield at photosystem II) and the bleaching conditions, via symbiotic microalgal density and chlorophyll concentrations, of mesophotic and shallow coral species subjected to different temperature scenarios reproducing different Degree Heating Weeks (DHWs). Their results indicated that mesophotic corals have a threshold temperature slightly lower or equal to that of shallow corals, suggesting that, although they can survive thermal stress below 4 DHWs, mass bleaching can occur above this threshold. Coral reefs at relatively high latitudes could also be potential refuges for corals (Camp et al., 2018; Dellisanti et al. , 2023). However, corals living in such habitat could suffer from lowtemperature stress, inducing bleaching (Tracey et al., 2003; Marangoni et al., 2021). Wei et al. explore the response of Porites lutea from a high-latitude coral reef in the South China Sea under acute (1-2 weeks) and chronic (6-12 weeks) low-temperature stress, by analyzing maximum quantum yields and transcriptomic profiles. Low temperatures inhibited photosynthetic efficiency and reduced energy production and calcification by down-regulating sugar metabolism and calcification-related genes. However, this was particularly observed during a short acute treatment, suggesting a possible coral acclimation to chronic low temperature.

Although thermal stress is recognized as the main cause of coral bleaching, high solar irradiance can also play a central role in this process by exacerbating the production of reactive oxygen species (ROS) (Roth, 2014; Courtial et al., 2018). Shading-based management interventions could therefore reduce coral bleaching risk. Butcherine et al. examined the effectiveness of intermittent shade on two coral species held at either optimum or high temperatures. The analysis of coral health condition through the bleaching assessment (chlorophyll *a*, and symbiont density), the photochemistry, and the use of antioxidant enzymes (SOD and CAT) as cellular stress biomarkers, suggested that intermittently shading corals for 4 h can mitigate the impact of thermal stress.

However, even extremely low light levels, mainly related to high sedimentation rate and turbidity, can induce coral bleaching and negatively impact coral metabolism (DeSalvo et al., 2012; Bollati et al., 2021; Tuttle and Donahue, 2022). Using transcriptomics, Lock et al. identified gene expression patterns and molecular pathways that may allow the massive coral *Porites lobata* to tolerate and persist to chronic and severe sedimentation in the turbid Fouha Bay (Guam), providing important insights into coral metabolic plasticity and acclimation to this stressor. In particular, alternative energy generation pathways may help to counteract low light and oxygen levels, the upregulation of apoptosis genes may maintain colony integrity, and increased expression of cellular communication genes may help corals respond to sedimentassociated pathogens.

Molecular biomarkers are also used as a proxy for water quality and anthropogenic pollution. Tisthammer et al. employed enzymelinked immunosorbent assays to evaluate stress responses in *P. lobata* along an environmental gradient in Maunalua Bay (Hawaii), revealing distinct protein expression patterns, especially those of ubiquitin and Hsp70, which correlate with anthropogenic stressor levels across the bay. Nardi et al. analyzed the ecotoxicological response of the Mediterranean coral Madracis polycyclic aromatic hydrocarbons (PAHs) pharensis to bioaccumulated from chronic oil leakage from a shipwreck in Cyprus. The high ROS scavenging capacity and the low functionality of detoxification processes associated with the glutathione-S-transferase enzyme suggested that M. pharensis has the capability to develop cellular and physiological adaptations to chemical-mediated stress. Morgan et al. focused on the synergistic, antagonistic, or additive effect of oxybenzone BP-3, the active ingredient in sunscreen, and ocean acidification (OA), on the expression profiles of 22 genes of interest (GOIs) in sea the anemone Exaiptasia diaphana. The collective antagonistic responses of GOIs associated with collagen synthesis suggested their role as candidate biomarkers of stress, while GOIs with synergistic and additive responses, such as serotransferrin-like (TF) and monocarboxylate transporters (MCTs) genes. respectively, were also identified.

Finally, cellular stress mechanisms are also known to be involved in coral response to biotic interactions (Seveso et al., 2012; Seveso et al., 2017). For example, using gel-filtration chromatography and liquid chromatography-tandem mass spectrometry, Suzuki et al. identified and characterized red fluorescent proteins (RFPs) and chromoproteins (CPs) in inflammatory pink lesions of *Porites* colonies subjected to the pink pigmentation response (PPR). The results suggested a possible differential role of these proteins in coral immunity despite their coexistence. Additionally, CPs, which are specifically expressed in PPR lesions, may serve as an antioxidant protection, providing new insights into the role of CPs in the coral immune response.

Considering that understanding how corals can genetically or physiologically adapt to environmental changes has become a global research priority, we believe that this Research Topic provides a more comprehensive view of the cellular mechanisms involved. It may encourage future advancements in this field and support strategies and tools to potentially reduce or mitigate the impacts of cellular stress in corals.

Author contributions

DS: Writing-original draft. YL: Writing-review and editing. RB: Writing-review and editing. CD: Writing-review and editing. WD: Writing-review and editing.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

Bollati, E., Rosenberg, Y., Simon-Blecher, N., Tamir, R., Levy, O., and Huang, D. (2021). Untangling the molecular basis of coral response to sedimentation. *Mol. Ecol.* 2021, 884–901. doi:10.1111/mec.16263

Bongaerts, P., Ridgway, T., Sampayo, E. M., and Hoegh-Guldberg, O. (2010). Assessing the 'deep reef refugia' hypothesis: focus on Caribbean reefs. *Coral Reefs* 29, 309–327. doi:10.1007/s00338-009-0581-x

Camp, E. F., Schoepf, V., Mumby, P. J., Hardtke, L. A., Rodolfo-Metalpa, R., Smith, D. J., et al. (2018). The future of coral reefs subject to rapid climate change: lessons from natural extreme environments. *Front. Mar. Sci.* 5. doi:10.3389/fmars.2018.00004

Courtial, L., Planas Bielsa, V., Houlbreque, F., and Ferrier-Pagès, C. (2018). Effects of ultraviolet radiation and nutrient level on the physiological response and organic matter release of the scleractinian coral *Pocillopora damicornis* following thermal stress. *PLoS One* 13 (10), e0205261. doi:10.1371/journal.pone.0205261

Cziesielski, M. J., Schmidt-Roach, S., and Aranda, M. (2020). The past, present, and future of coral heat stress studies. *Ecol. Evol.* 9, 10055–10066. doi:10.1002/ece3. 5576

Dellisanti, W., Chung, J. T., Yiu, S. K., Tsang, R. H. L., Ang, J. P., Yeung, Y. H., et al. (2023). Seasonal drivers of productivity and calcification in the coral *Platygyra carnosa* in a subtropical reef. *Front. Mar. Sci.* 10, 994591. doi:10.3389/fmars.2023.994591

DeSalvo, M. K., Estrada, A., Sunagawa, S., and Medina, S. M. (2012). Transcriptomic responses to darkness stress point to common coral bleaching mechanisms. *Coral Reefs* 31, 215–228. doi:10.1007/s00338-011-0833-4

Downs, C. A. (2005). "Cellular diagnostics and its application to aquatic and marine toxicology," in *Techniques in aquatic toxicology*. Editor G. K. Ostrander (Boca Raton: CRC Press), Vol. 2, 181–208. doi:10.1201/9780203501597.sec2

Drury, C. (2020). Resilience in reef-building corals: the ecological and evolutionary importance of the host response to thermal stress. *Mol. Ecol.* 29, 448–465. doi:10.1111/ mec.15337

Eakin, C. M., Sweatman, H. P., and Brainard, R. E. (2019). The 2014–2017 global-scale coral bleaching event: insights and impacts. *Coral Reefs* 38 (4), 539–545. doi:10.1007/s00338-019-01844-2

Helgoe, J., Davy, S. K., Weis, V. M., and Rodriguez-Lanetty, M. (2024). Triggers, cascades, and endpoints: connecting the dots of coral bleaching mechanisms. *Biol. Rev.* 99, 715–752. doi:10.1111/brv.13042

Hughes, T. P., Kerry, J., Álvarez-Noriega, M., Álvarez-Romero, J. G., Anderson, K. D., Baird, A. H., et al. (2017). Global warming and recurrent mass bleaching of corals. *Nature* 543, 373–377. doi:10.1038/nature21707 organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Hughes, T. P., Anderson, K. D., Connolly, S. R., Heron, S. F., Kerry, J. T., Lough, J. M., et al. (2018). Spatial and temporal patterns of mass bleaching of corals in the Anthropocene. *Science* 359, 80–83. doi:10.1126/science.aan8048

Louis, Y. D., Bhagooli, R., Seveso, D., Maggioni, D., Galli, P., Vai, M., et al. (2020). Local acclimatisation-driven differential gene and protein expression patterns of Hsp70 in *Acropora muricata*: implications for coral tolerance to bleaching. *Mol. Ecol.* 29, 4382-4394. doi:10.1111/mec.15642

Marangoni, L., Rottier, C., and Ferrier-Pagès, C. (2021). Symbiont regulation in *Stylophora pistillata* during cold stress: an acclimation mechanism against oxidative stress and severe bleaching. *J. Exp. Biol.* 224, jeb235275. doi:10.1242/jeb.235275

Montalbetti, E., Biscéré, T., Ferrier-Pagès, C., Houlbrèque, F., Orlandi, I., Forcella, M., et al. (2021). Manganese benefits heat-stressed corals at the cellular level. *Front. Mar. Sci.* 8, 681119. doi:10.3389/fmars.2021.681119

Muir, P. R., Marshall, P. A., Abdulla, A., and Aguirre, J. D. (2017). Species identity and depth predict bleaching severity in reef-building corals: shall the deep inherit the reef? *Proc. R. Soc. B* 284, 20171551. doi:10.1098/rspb.2017.1551

Oakley, C. A., and Davy, S. K. (2018). "Cell biology of coral bleaching," in *Coral bleaching. Ecological studies (analysis and synthesis)*. Editors M. van Oppen and J. Lough (Cham: Springer), 233, 189–211. doi:10.1007/978-3-319-75393-5_8

Roth, M. S. (2014). The engine of the reef: photobiology of the coral-algal symbiosis. *Front. Microbiol.* 5, 422. doi:10.3389/fmicb.2014.00422

Seveso, D., Montano, S., Reggente, M. A. L., Maggioni, D., Orlandi, I., Galli, P., et al. (2017). The cellular stress response of the scleractinian coral *Goniopora columna* during the progression of the black band disease. *Cell Stress Chap* 22 (2), 225–236. doi:10.1007/ s12192-016-0756-7

Seveso, D., Montano, S., Strona, G., Orlandi, I., Vai, M., and Galli, P. (2012). Upregulation of Hsp60 in response to skeleton eroding band disease but not by algal overgrowth in the scleractinian coral *Acropora muricata*. *Mar. Environ. Res.* 78, 34–39. doi:10.1016/j.marenvres.2012.03.008

Tracey, S., William, C. D., and Ove, H.-G. (2003). Photosynthetic responses of the coral *Montipora digitata* to cold temperature stress. *Mar. Ecol. Prog. Ser.* 248, 85–97. doi:10.3354/meps248085

Tuttle, L. J., and Donahue, M. J. (2022). Effects of sediment exposure on corals: a systematic review of experimental studies. *Environ. Evid.* 11 (1), 4–33. doi:10.1186/ s13750-022-00256-0

Weis, V. M. (2019). Cell biology of coral symbiosis: foundational study can inform solutions to the coral reef crisis. *Integr. Comp. Biol.* 59, 845–855. doi:10.1093/icb/icz067