



Corrigendum: miR156/SPL10 Modulates Lateral Root Development, Branching and Leaf Morphology in Arabidopsis by Silencing *AGAMOUS-LIKE 79*

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A Corrigendum on

miR156/SPL10 Modulates Lateral Root Development, Branching and Leaf Morphology in Arabidopsis by Silencing *AGAMOUS-LIKE 79*

by Gao, R., Wang, Y., Gruber, M. Y., and Hannoufa, A. (2018). *Front. Plant Sci.* 8:2226. doi: 10.3389/fpls.2017.02226

In the original article, there was a mistake in the legend for **Figure 4D**, **Figure 8A** as published. There was an error in our CRISPR-generated AGL79KD mutant. In the **Results**, subsection **Characterization of the AGL79KD Arabidopsis Mutant**, we showed Arabidopsis knockdown mutants with a CRISPR-mutated AGL79. In performing follow up research we discovered that the plants had no point mutations as claimed in the original manuscript. As the researcher who carried out this experiment left the lab about 18 months ago, we reviewed the original sequence traces, and found that when reading the sequence in one direction (as the researcher probably did) it seemed to indicate point mutations, but when we sequenced in the reverse direction we could find no such point mutations in the AGL79 locus, and determined that this was indeed an error in reading the sequencing data. Figure legends that included this mutant have been modified to omit ALG79KD (**Figures 4D** and **8A**).

Due to the same error detailed above, there was a mistake in Figures 6, 7, Supplementary Figure 1 and the Supplementary Material Document 2. Figures and their corresponding legends that were specific to AGL79KD have been removed (Figures 6, 7, Supplementary Figure 1, and Supplementary Material Document 2). Figures have also been renumbered to reflect these changes. In addition, primers related to the generation and validation of CRISPR plants have been removed from **Supplementary Table 2**.

Furthermore, in the original article, the following eight references, that are related to AGL79 KD mutant descriptions/discussions, have been removed.

- An, H., Roussot, C., Suarez-Lopez, P., Corbesier, L., Vincent, C., Pineiro, M., et al. (2004). CONSTANS acts in the phloem to regulate a systemic signal that induces photoperiodic flowering of *Arabidopsis*. *Development* 131, 3615–3626. doi: 10.1242/dev.01231
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time by ambient temperature in *Arabidopsis*. *Genes Dev.* 21, 397–402. doi: 10.1101/gad.1518407

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- Takada, S., and Goto, K. (2003). Terminal flower2, an *Arabidopsis* homolog of heterochromatin protein1, counteracts the activation of flowering locus T by constans in the vascular tissues of leaves to regulate flowering time. *Plant Cell* 15, 2856–2865. doi: 10.1105/tpc.016345

Lastly, corrections have been made to the **Methods**, **Results**, and **Discussion** of the original article and the AGL79KD mutant has been removed as detailed below.

In the **Methods**, subsection **CRISPR-Cas9 Design and Screening for AGL79 Gene Editing** has been removed.

Subsection **Characterization of AGL79 Overexpression Plants** has been corrected to:

“To investigate the role of AGL79 in *Arabidopsis* development, we generated transgenic plants with enhanced expression of AGL79. Compared to WT, the highest AGL79 overexpression plants flowered early and had fewer and

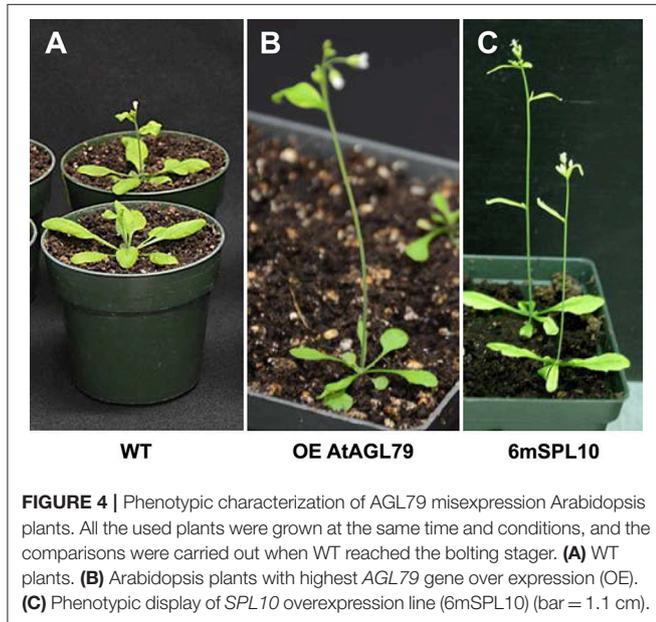


FIGURE 4 | Phenotypic characterization of AGL79 misexpression *Arabidopsis* plants. All the used plants were grown at the same time and conditions, and the comparisons were carried out when WT reached the bolting stager. **(A)** WT plants. **(B)** *Arabidopsis* plants with highest AGL79 gene over expression (OE). **(C)** Phenotypic display of *SPL10* overexpression line (6mSPL10) (bar = 1.1 cm).

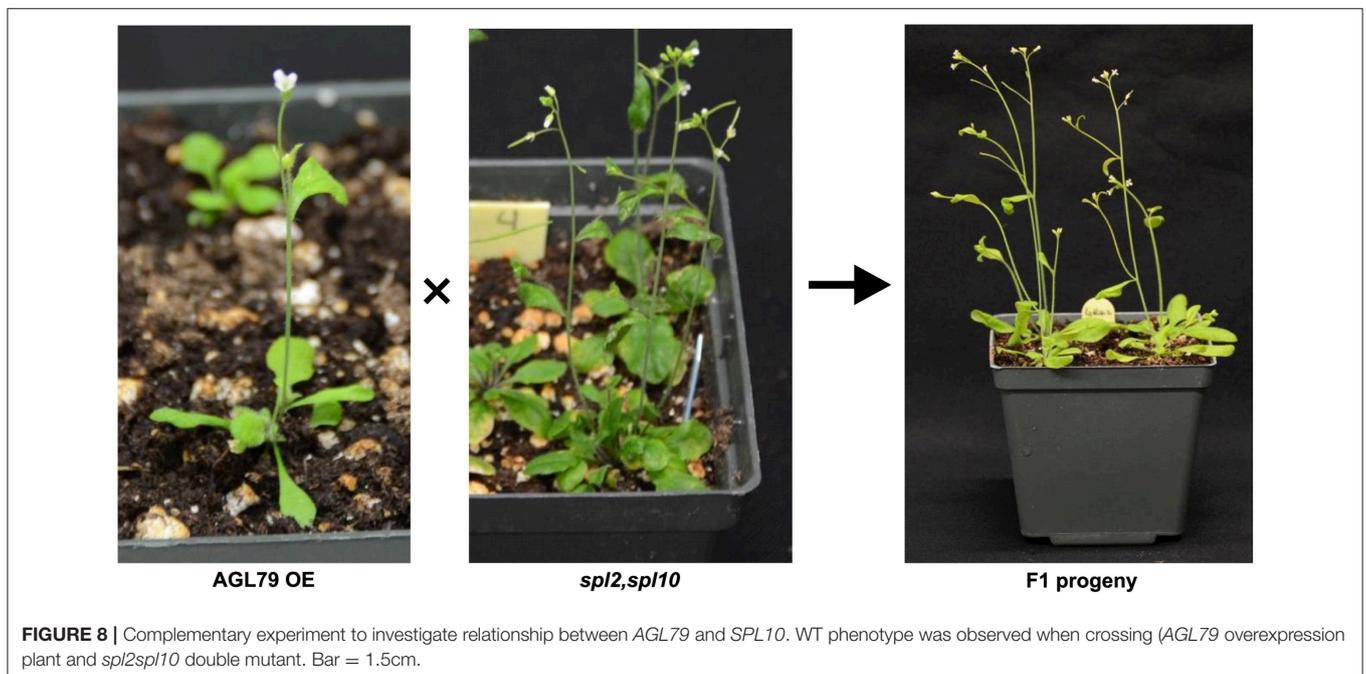


FIGURE 8 | Complementary experiment to investigate relationship between AGL79 and *SPL10*. WT phenotype was observed when crossing (*AGL79* overexpression plant and *spl2spl10* double mutant. Bar = 1.5cm.

smaller rosette leaves [Figures 4B, 5B (WT and Group 1)] much like the *SPL10* overexpression plants (6mSLP10) (Figure 4C, Supplementary Figure 1). Transgenic *Arabidopsis* plants harboring the *AGL79* overexpression construct were divided into three groups depending on *AGL79* expression. Group 1 (lines L1, L2, and L3) had the highest *AGL79* transcript levels in both the leaf and root tissues (Figure 5A), with lower expression in roots relative to leaves. Group 2 (lines L11, L12, and L20) had intermediate *AGL79* expression, with variable expression levels between leaf and root (Figure 5A). Group 3 (lines L16, L18, and L27) displayed the lowest *AGL79* gene transcripts, and there were no obvious differences in *AGL79* transcript levels between the leaf and root (Figure 5A). Different phenotypes could be observed in these *AGL79* overexpression plants depending on *AGL79* expression levels (Figure 5B). Compared to WT (3 weeks after seed germination), Group 1 plants displayed fewer rosette leaves and early flowering time (Figure 5B). Group 2 plants displayed a phenotype similar to WT (Figure 5B). Group 3 plants showed more lateral shoot branches and a higher number of rosette leaves, as well as a significant delay in flowering (Figure 5B). In addition, the transcript level of *SPL10* gene was also investigated in both the leaves and roots of the above-mentioned plants. Although changes in *SPL10* expression could be detected in three groups of *AGL79*OE plants (Figure 5C), these changes did not follow any consistent trend, as found for *AGL79* (Figure 5C), suggesting that *AGL79* could be a downstream gene regulated by *SPL10*, and hence fluctuations in *AGL79* expression would not affect the expression of the upstream *SPL10* gene.”

Subsection **Characterization of the *AGL79*KD *Arabidopsis* Mutant**, has been removed.

Subsection **Regulatory Relationship between *AGL79* and *SPL10***, has been corrected to:

“As all the evidence derived from molecular and biological analysis (Figures 2A,B, 4C) revealed that *AGL79* is likely regulated through the miR156-SPL pathway, we investigated whether a linear regulatory relationship exists between *SPL10* and *AGL79*. Crossing *AGL79*OE plants and *spl2spl10* double mutant produced F1 progeny showing WT-like phenotype (Figure 8B). The selected genotyping results of the double mutant (*spl2spl10*) and *AGL79* OE plants are shown in Supplementary Figure 3. These results suggest a direct linear relationship between *AGL79* and *SPL10* genes.”

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- Okushima, Y., Fukaki, H., Onoda, M., Theologis, A., and Tasaka, M. (2007). ARF7 and ARF19 regulate lateral root formation via direct activation of LBD/ASL genes in *Arabidopsis*. *Plant Cell* 19, 118–130. doi: 10.1105/tpc.106.047761
- Yu, N., Niu, Q. W., Ng, K. H., and Chua, N. H. (2015). The role of miR156/SPLs modules in *Arabidopsis* lateral

In the **Discussion**, paragraph two has been corrected to:

“The discovery that *AGL79* is regulated by *SPL10* may provide insight into how the latter regulates lateral root development in *Arabidopsis* (Yu et al., 2015). Currently lateral root formation in *Arabidopsis* is known to be regulated by two related *AUXIN RESPONSE FACTORS* (*ARF7* and *ARF19*) via direct activation of *LATERAL ORGAN BOUNDARIES DOMAIN* and *ASYMMETRIC LEAVES-LIKE* (*LBD/ASLs*) (Okushima et al., 2007). In addition, lateral root formation in *Arabidopsis* is also redundantly regulated by cytokinin biosynthesis genes *IPT3* and *IPT5* and all three cytokinin histidine kinase receptor genes (*AHK2*, *AHK3*, and *CRE1/AHK4*) (Chang et al., 2013). The plant hormones (auxin, cytokinins, gibberellins, abscisic acid, ethylene, jasmonic acid, strigolactones, brassinosteroids, and salicylic acid) also regulate normal root growth and mediate root morphological responses to abiotic stress (Chang et al., 2013). Morphological analysis of *Arabidopsis* plants with enhanced expression of *AGL79* revealed *AGL79* to be involved in controlling shoot branching.”

and paragraph four should be removed and the last paragraph has been corrected to:

“In summary, our results suggest that the miR156/*SPL10* regulatory pathway is involved in regulating plant lateral root growth by directly targeting and activating the expression of *AGL79*. By investigating the gain- of function of *AGL79* transgenic plants, we also found *AGL79* to be involved in regulating plant leaf shape, shoot branching, and flowering time. Further characterization of the *AGL79* gene in other plant species, especially in major crops, will determine how conserved *AGL79* is in plants. It can also be tested in crop improvement efforts to enhance resilience and productivity.”

The authors apologize for these errors and state that this does not change the scientific conclusions of the article in any way. The original article has been updated.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2019.00515/full#supplementary-material>

root development. *Plant J.* 83, 673–685. doi: 10.1111/tpj.12919

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