



Biofertilizers as Strategies to Improve Photosynthetic Apparatus, Growth, and Drought Stress Tolerance in the Date Palm

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Rainfall regimes are expected to shift on a regional scale as the water cycle intensifies in a warmer climate, resulting in greater extremes in dry versus wet conditions. Such changes are having a strong impact on the agro-physiological functioning of plants that scale up to influence interactions between plants and microorganisms and hence ecosystems. In (semi)-arid ecosystems, the date palm (*Phoenix dactylifera* L.) -an irreplaceable tree- plays important socio-economic roles. In the current study, we implemented an adapted management program to improve date palm development and its tolerance to water deficit by using single or multiple combinations of exotic and native arbuscular mycorrhizal fungi (AMF1 and AMF2 respectively), and/or selected consortia of plant growth-promoting rhizobacteria (PGPR: B1 and B2), and/or composts from grasses and green waste (C1 and C2, respectively). We analyzed the potential for physiological functioning (photosynthesis, water status, osmolytes, mineral nutrition) to evolve in response to drought since this will be a key indicator of plant resilience in future environments. As result, under water deficit, the selected biofertilizers enhanced plant growth, leaf water potential, and electrical conductivity parameters. Further, the dual-inoculation of AMF/PGPR amended with composts alone or in combination boosted the biomass under water deficit conditions to a greater extent than in non-inoculated and/or non-amended plants. Both single and dual biofertilizers improved physiological parameters by elevating stomatal conductance, photosynthetic pigments (chlorophyll and carotenoids content), and photosynthetic efficiency. The dual inoculation and compost significantly enhanced, especially under drought stress, the concentrations of sugar and protein content, and antioxidant enzymes (polyphenoloxidase and peroxidase) activities as a defense strategy as compared with controls. Under water stress, we demonstrated that phosphorus was improved in the inoculated and

amended plants alone or in combination in leaves (AMF2: 807%, AMF1+B2: 657%, AMF2+C1+B2: 500%, AMF2+C2: 478%, AMF1: 423%) and soil (AMF2: 397%, AMF1+B2: 322%, AMF2+C1+B2: 303%, AMF1: 190%, C1: 188%) in comparison with controls under severe water stress conditions. We summarize the extent to which the dual and multiple combinations of microorganisms can overcome challenges related to drought by enhancing plant physiological responses.

Keywords: arbuscular mycorrhizal fungi, climate change, compost, PGPR, plant fitness, photosynthesis, agro-physiological responses, water deficit

INTRODUCTION

Owing to rapid climate change, drought is becoming one of the most important environmental stresses that is outside of plants' physiological limits and is causing a substantial decline in crop productivity (Shaw and Etterson, 2012; Sarwat and Tuteja, 2017). Drought influences transport and availability of soil nutrients (Vurukonda et al., 2016), and it affects morphological, physiological, and nutritional traits of plants especially water content, leaf water potential, photosynthetic pigment, stomatal conductance and phosphorus (P) and nitrogen (N) absorption (Jaleel et al., 2009; Augé et al., 2014; Baslam et al., 2014; Meddich et al., 2015a, 2018; Symanczik et al., 2018). Drought also affects antioxidant defense leading to oxidative stress owing to the accumulation of reactive oxygen species (ROS) such as hydrogen peroxide (Maheshwari et al., 2012; Vurukonda et al., 2016; Abdel Latef et al., 2019a,b). On a higher scale, when plants are exposed to drought that alters their photosynthesis machinery, this can shift source/sink relationships of photosynthate, symbiotic interactions, plant growth, and fitness (Becklin et al., 2016; Sever et al., 2018). The inhibition and degradation of chlorophyll synthesis through ROS over-accumulation leads to a decrease of photosynthetic activity (Abd El-Mageed et al., 2018; Duo et al., 2018).

The date palm (*Phoenix dactylifera* L.) is a dioecious evergreen tree with major ecological and socio-economic roles in many countries, mainly arid zones (Chao and Krueger, 2007). Indeed, the economic importance of the palm family of plants (Arecaceae) ranks second only to the grass family (Poaceae) among monocotyledons and the third in the world (after the Gramineae and Leguminosae families) (FAO¹). The economic utility of these palms is based on their fruits and derived beverages as a staple food, their wood, as palms are used in building and the artisanal sector, where they are used for ornamental purposes (Arias et al., 2016). In addition, in oasis ecosystems, date palms create a microclimate that is essential for the cultivation of underlying crops such as fruit trees, vegetables and forage species (Meddich et al., 2019). However, date palm groves are subjected to biotic (primary *fusarium* wilt) and abiotic (mainly drought and salinity) constraints as well as low soil fertility and management that have decimated this crop thus deteriorating oasis ecosystems, hindering agricultural production under this environmental instability, and driving higher levels of migration (Oihabi, 1991;

Meddich et al., 2015a, 2018; Arias et al., 2016; Meddich and Boumezzough, 2017; FAO, 2018; Whitman, 2019). Factors such as drought, exacerbated by climate change, will affect plant physiology by altering plant-organism interactions since plant species evolve in complex environments with networks of interacting species. Although studies have examined various ways of handling stress inducers to increase plant survival and performance, little has been done on the integrative aspect of this improvement approach to protecting and empowering plants to resist and grow better under drought conditions. In this regard, management practices and strategies that allow plants to resist abiotic and biotic stresses are urgently required and should be exploited to improve agricultural production (Meddich et al., 2018, 2019), reduce the use of pesticides and chemical fertilizers, and protect crops and soil quality (Shen et al., 2013). It should be noted that the plant nursery industry – being trees and fruit plants the most important nursery crops – constitutes a large and growing sector of agriculture. The regular practice of cultivating *in vitro* plants by farmers under nursery conditions prior to field transplantation involves growing the plants only in sandy soil without any amendments. At the time of planting is especially apparent when outplants face stresses and/or strong competition from other vegetation. At this stage, the growth potential of planting stock plays a primordial role for the transplanting success. The application of biofertilizers such as organic fertilizers and beneficial soil microorganisms has emerged as a potential solution to promote plant fitness, boost yield, and tolerance to environmental constraints (Abdel Latef and Chaoping, 2011; Meddich et al., 2015b; Abdel Latef et al., 2016; Padilla et al., 2017; Júnior et al., 2018; Symanczik et al., 2018; Ait-El-Mokhtar et al., 2019; Ben-Laouane et al., 2019; Raklami et al., 2019). The appropriate management of plant nutrition, growth, and tolerance to drastic constraints such as drought, salinity and soil poverty is becoming a key component in increasing crop yield under changing environmental conditions (Zou and Wu, 2011; Baslam and Goicoechea, 2012; Maheshwari et al., 2012; Hidri et al., 2016; De Pascale et al., 2017; Meddich et al., 2018). In addition to regulating nutrient acquisition, inoculation with Arbuscular Mycorrhizal Fungi (AMF) and Plant Growth Promoting Rhizobacteria (PGPR) are successfully being seen to be an effective substitute to ensure stable, safe and sustainable agricultural and biomass production (Wu and Zou, 2017; Zhang et al., 2018; Kumar et al., 2019; Kumari et al., 2019; Abdel Latef et al., 2020). Indeed, the beneficial effects of the inoculation by the rhizosphere microbial communities are linked with nutrient recycling,

¹<http://www.fao.org>

mineral nutrition. solubilisation of nutrients such as P, potassium (K) and iron (Fe), biodegradation of soil organic matter, phytohormone and antibiotics productions, improvements in soil structure and aggregation, and enhancement of plant resistance to pests and diseases (Al-Karaki, 2000, 2006; Al-Karaki et al., 2004; Nadeem et al., 2014; Rillig et al., 2014; Grobelak et al., 2015; Kumar et al., 2015; Meddich et al., 2018, 2019; Symanczik et al., 2018; Raklami et al., 2019; Wu et al., 2019). The (co)-inoculants of PGPR and/or AMF can advance the nutrient use efficiency of fertilizers (Adesemoye et al., 2009). Moreover, AMF and/or PGPR inoculation could mitigate the detrimental effect of stresses through the enhancement of photosynthetic efficiency, overproduction of antioxidant enzymes and/or non-enzymatic antioxidants, and/or activation of the mycorrhizal induced resistance (MIR) mechanism by bypassing plant defense (Pozo and Azcón-Aguilar, 2007; Bompadre et al., 2014; Nadeem et al., 2014; Pieterse et al., 2014; Gusain et al., 2015; Lenoir et al., 2016; Duo et al., 2018; Javan Gholiloo et al., 2019). Although PGPR and AMF are commonly applied as beneficial microorganisms in agriculture and several studies have been devoted to studying the influence of this symbiosis on the plant response to abiotic stress, the mechanisms responsible for increased plant tolerance to stress have yet to be fully elucidated. Also, little is known about the agro-physiological changes occurring in the plants when these microorganisms are applied together. It has been suggested that the combined application of both microorganisms has positive effects on the nutritional composition of several plant species (Nadeem et al., 2014; Ben-Laouane et al., 2019).

Furthermore, the exogenous introduction of beneficial organic substances such as compost has been evaluated previously (Nikitas et al., 2008) as a soil conditioner in agriculture or a substitute to chemical fertilizers to improve resilience, yield, and tolerance of plants to the toxicity of these stress-imposed conditions. The use of local composts represents an eco-friendly alternative for plant growth, mineral nutrition, soil organic matter content, and soil properties such as water retention capacity and soil suppressiveness (Shen et al., 2013; Luciens et al., 2014; Mehta et al., 2014; Kwey et al., 2015; Ning et al., 2017). Previous studies have suggested that compost application in soil increased the supply of organic carbon and N for microbial communities and improved soil health and plant yield (Shen et al., 2013; Liu et al., 2018). Further, compost application triggers plant resistance to different environmental stresses (Tartoura et al., 2014; Ortuño et al., 2018) by increasing the photosynthetic activity (Abd El-Mageed et al., 2019; Khosravi Shakib et al., 2019). In the current research, we explored the agro-physiological and biochemical responses involved in drought adaptation in date palms, and the functionality of the single and dual-use of selected strains of PGPR and native and exotic AMF with or without the addition of two composts. The objective of this study was to evaluate the morpho-physiological basis of drought responses in date palms under implementation of eco-friendly cultivation practices. The results obtained here will provide a deeper understanding of the mechanisms of date palm tolerance to long-term drought stress as well as paving the way for identification of the best factors that led to successful outcomes in the biofertilization experiments for other crops.

MATERIALS AND METHODS

Biofertilizer Materials

Two types of AMF inoculants were used in our experiment: (i) an exogenous AMF strain (*Rhizoglyphus irregularis*, DAOM 197198) provided by the Plant Biotechnology Institute of Montreal (Canada) and (ii) an indigenous consortium of AMF isolated from the Tafilalet palm grove located 500 km southeast of Marrakesh and containing a mixture of native species: (i) *Glomus* sp. (15 spores/g of substrate), (ii) *Sclerocystis* sp. (9 spores/g substrate), and (iii) *Acaulospora* sp. (1 spore/g of substrate) (Meddich et al., 2015a). The inoculum was enriched in propagules by co-cultivation with *Zea mays* L. as the host plant under controlled greenhouse conditions. Corn roots containing hyphae, vesicles, and spores were harvested, cut into small pieces and used as the inoculum. Inoculation of date palm was performed by adding 40 g of the inoculum (roots and substrate containing spores) to the date palm root system. Non-mycorrhizal (NM) treatments received an equal quantity of both non-inoculated (and non-mycorrhizal) *Z. mays* roots to match 'organic matter' in the pots and filtered inoculum in an attempt to restore other soil free-living microorganisms accompanying the AMF. The filtrate for each pot was obtained by passing the mycorrhizal inoculum in 20 mL of distilled water through a layer of 15- to 20- μ m filter papers (Whatman, GE Healthcare, Buckinghamshire, United Kingdom).

The bacterial inocula used in our study consisted of four PGPR isolates (Z1, Z2, Z4, and ER21 strains) isolated from the date palm groves rhizosphere (Tafilalet, Morocco). The inocula were prepared by growing the strains in Tryptic Soy Broth (TSB) liquid culture at 28°C to an optical density of 1 at 600 nm (about 10⁹ CFU/mL). The plant inoculations were carried out by adding 4 mL of the bacterial suspension formed from the four abovementioned strains into equal volumes closer to the roots. After 15 days, a second inoculation (booster) was carried out by placing another 4 mL of the bacterial suspension next to the plant roots to increase the bacterial rate in the soil and ensure the infection of the newly formed roots.

The quantification *in vitro* of plant growth-promoting traits of the strains used was examined by standard protocols: phosphate solubilization was performed by the production of halo on agar medium as described by Alikhani et al. (2006) and the tolerance to water deficiency was tested by the resistance to polyethylene glycol. A confrontation assay was carried out to confirm the absence of inhibition between the four strains. The PGPR characteristics of the four strains are listed in **Table 1**.

The composts used were prepared from grass (C1) and a mixture of green (C2) waste as described by Meddich et al. (2016). The composts (5% W/W with respect to culture soil) were added

TABLE 1 | Phosphate solubilization and resistance to polyethylene glycol (tolerance to water deficiency) of the four tested PGPR strains (Z1, Z2, Z4, and ER21).

Activity	Z1	Z2	Z4	ER21
Phosphate solubilization	+	+	+	+
Resistance to polyethylene glycol 6000	+	+	-	-

to the corresponding pots at date palm vitroplants transplanting (Anli et al., 2020). The physicochemical and microbiological properties of the two composts are presented in **Table 2**.

Experimental Design

Date palm vitroplants (*Phoenix dactylifera* L.) of variety Boufgouss -an elite variety with high commercial importance- were transplanted at the two leaf stage into 2.4 L plastic buckets filled (4/5) with sterilized soil (at 180°C for 3 h on 3 consecutive days) alone or mixed with compost at 5%. The soil used (bulk density: $1.32 \pm 0.01 \text{ g cm}^{-3}$) had the following characteristics: sand, 51%; clay, 19%; loam, 30%, available phosphorus, 11 ppm; organic matter, 1%; total organic carbon, 0.58%; nitrogen, 0.84 mg/g; EC 0.19 mS/cm; and pH, 8.6. The soil bulk density did not vary in every treatment. The finely textured soil to successfully grow date palm, instead of sand, has been used to achieve fast equilibrium rates during drying. The plants were watered and maintained at 75% field capacity (FC). During the pre-sowing period, irrigation was applied to FC to ensure full stand establishment in all treatments. Soil moisture was measured randomly in untreated and treated pots in each treatment using a TDR meter (Delta UK Ltd., Clacton-on-Sea, United Kingdom) in the morning and evening of each day. According to the measured soil water content, soil bulk density, soil moisture maximum field capacity and soil weight, the amount of needed water under different water conditions was calculated. Plants were grown in the greenhouse at 25.5°C (16/8 h light/dark) with fluorescent lighting ($500 \mu\text{mol m}^{-2} \text{ s}^{-1}$) and average relative humidity of 68.5%.

The experiment was carried out in a fully randomized design with 10 biological replicates for each treatment (in total 54 treatments) and all plants were placed randomly in the greenhouse (**Table 3**).

Three months after experiment start, two water regimes were imposed 75 and 25% FC (Baslam et al., 2014; Meddich et al., 2015a, 2018).

Chlorophyll Fluorescence and Stomatal Conductance Determinations

Chlorophyll fluorescence was measured by a fluorometer (OPTI-SCIENCE, OS30p). Dark adaptation was made on the upper side of the second fully developed leaf from the apex by obscuring for 20 min. This parameter was measured by transmission at 650 nm on a leaf area of 12.5 mm². The fluorescence signal was recorded for a second at an acquisition speed of 10 μs (Strasser and Strasser, 1995). Stomatal conductance (g_s) was determined as described by Harley et al. (1992).

Photosynthetic Pigments Quantification

The concentration of chlorophyll *a*, *b*, total chlorophyll, and carotenoids was determined according to the method described by Arnon (1949). Photosynthetic pigments were extracted from the frozen leaf powder subsample using cold acetone 80%. Following centrifugation at $10,000 \times g$ for 10 min, supernatant absorbance was read at 480, 645, and 663 nm using a UV-visible spectrophotometer (UV-3100PC spectrophotometer, VWR).

Leaf Water Potential

Leaf water potential (Ψ_w) was measured using a pressure chamber (Model 600-EXP Super Pressure Chamber, PMS instrument, Albany, OR, United States) at predawn (06:00–08:00 h). The measurements were taken on mature fully expanded leaves from the upper part of the stem. Cutting leaves water potential were measured over the same days and immediately after gas exchange measurements.

Growth Assessment and Mineral Analysis

The growth performance of date palm plants was assessed by measuring the number of leaves, shoot height, root length, leaf area, and total dry matters (DM; obtained after drying samples at 80°C until the weight remained constant). The first fully expanded leaf of date palms in each treatments was harvested at the end of the light period, snap-frozen, ground to a fine powder in liquid N using a pestle and mortar, and kept at -80°C for the subsequent biochemical analyses.

For mineral analyses, the dried shoots were grounded using a coffee mill. Shoot N concentration was measured according to the method described by Rodier (1984). Shoot P concentration was estimated using the Olsen method (Olsen and Sommers, 1982) by incinerating the shoot powder (500 mg) in a muffle furnace before acid extraction.

Mycorrhization Assessment

Root samples were washed with distilled water and cleaned with 10% of KOH at 90°C for 30 min. Then, they were washed again and acidified with 2% HCl for 10 min and stained with Trypan blue at 90°C for 20 min according to Phillips and Hayman (1970). The microscopic assessment of mycorrhizal root colonization rates was performed according to the method of Trouvelot et al. (1986).

Total Soluble Sugars Quantification

Total soluble sugars (TSS) were determined in 0.1 g of the frozen leaf powder in 80% (v/v) ethanol. The quantity of TSS was determined according to Dubois et al. (1956) in

TABLE 2 | Physico-chemical and microbiological properties of the composts used in this study.

Composts	pH	EC (mS/cm)	COT (%)	NTK (%)	C/N	P (mg/g)	Bacterial population (CFU/g)	Fungal population (CFU/g)
Compost (C1)	7.86	7.10	30.65	2.19	14.00	0.270	1.65×10^8	4.30×10^5
Compost (C2)	7.80	8.50	27.24	1.32	20.64	0.266	2.12×10^5	9.75×10^4

EC, electrical conductivity; TOC, total organic carbon; TKN, total Kjeldahl-nitrogen; C/N, carbon-to-nitrogen ratio; P, phosphorous; CFU, colony-forming unit.

TABLE 3 | Different treatments (and their nomenclature) applied in this study.

Treatments	Water regime	
Control	75% FC	Plants non-amended with compost and no-inoculated with AMF/PGPR
B1	25% FC	Plants non-amended with compost, no-inoculated with AMF, and inoculated with PGPR consortia B1 (Z1+Z2)
B2		Plants non-amended with compost, no-inoculated with AMF, and inoculated with PGPR consortia B2 (Z1+Z2+Z4+ER21)
C1		Plants amended with compost C1 (grass waste), no-inoculated with AMF/PGPR
C1 + B1		Plants amended with compost C1, no-inoculated with AMF, and inoculated with PGPR consortia B1
C1 + B2		Plants amended with compost C1, no-inoculated with AMF, and inoculated with PGPR consortia B2
C2		Plants amended with compost C2 (mixture of green waste) and no-inoculated with AMF/PGPR
C2 + B1		Plants amended with compost C2, no-inoculated with AMF, and inoculated with PGPR consortia B1
C2 + B2		Plants amended with compost C2, no-inoculated with AMF and inoculated with PGPR consortia B2
AMF1		Plants inoculated with AMF1 (exogenous <i>R. irregulare</i>), non-amended with compost, and no-inoculated with PGPR
AMF1 + B1		Plants inoculated with AMF1, inoculated with PGPR consortia B1, and non-amended with compost
AMF1 + B2		Plants inoculated with AMF1, inoculated with PGPR consortia B2, and non-amended with compost
AMF1 + C1		Plants inoculated with AMF1, amended with C1, and no-inoculated with PGPR
AMF1 + C1 + B1		Plants inoculated with AMF1, inoculated with PGPR consortia B1, and amended with C1
AMF1 + C1 + B2		Plants inoculated with AMF1, inoculated with PGPR consortia B2, and amended with C1
AMF1 + C2		Plants inoculated with AMF1, amended with C2, and no-inoculated with PGPR
AMF1 + C2 + B1		Plants inoculated with AMF1, inoculated with PGPR consortia B1, and amended with C2
AMF1 + C2 + B2		Plants inoculated with AMF1, inoculated with PGPR consortia B2, and amended with C2
AMF2		Plants inoculated with AMF2 (indigenous consortium of AMF), non-amended with compost, and no-inoculated with PGPR
AMF2 + B1		Plants inoculated with AMF2, inoculated with PGPR consortia B1, and non-amended with compost
AMF2 + B2		Plants inoculated with AMF2, inoculated with PGPR consortia B2, and non-amended with compost
AMF2 + C1		Plants inoculated with AMF2, amended with C1, and no-inoculated with PGPR
AMF2 + C1 + B1		Plants inoculated with AMF2, inoculated with PGPR consortia B1, and amended with C1
AMF2 + C1 + B2		Plants inoculated with AMF2, inoculated with PGPR consortia B2, and amended with C1
AMF2 + C2		Plants inoculated with AMF2, amended with C2, and no-inoculated with PGPR
AMF2 + C2 + B1		Plants inoculated with AMF2, inoculated with PGPR consortia B1, and amended with C2
AMF2 + C2 + B2		Plants inoculated with AMF2, inoculated with PGPR consortia B2, and amended with C2

0.2 mL of the supernatant mixed with 0.2 mL of phenol and 1 mL of concentrated sulfuric acid. After 15 min, TSS content was determined by measuring the absorbance at 485 nm and calculated using the standard glucose curve.

Total Soluble Proteins and Antioxidant Enzymes

Frozen leaf powder subsamples (0.1 g) were homogenized in a cold mortar with 4 mL of 1 M phosphate buffer (pH 7) containing 5% polyvinylpyrrolidone (PVPP). The homogenate was centrifuged at $18,000 \times g$ for 15 min at 4°C and the supernatant was used to measure antioxidant enzyme activities (Tejera García et al., 2004). Total soluble proteins were determined according to the technique described by Bradford (1976). Peroxidase (POX) activity was measured as described previously (Hori et al., 1997). The reaction mixture (3 mL) contained 1 M phosphate buffer (pH 7.0), 20 mM guaiacol, 40 mM H₂O₂, and 0.1 mL of the enzymatic extract which was added to start the reaction. POX activity was determined at 470 nm by its ability to convert guaiacol to tetraguaiacol ($\epsilon = 26.6 \text{ mM}^{-1} \cdot \text{cm}^{-1}$). One unit of POX activity was defined as an absorbance change of 0.01 unit min⁻¹. Polyphenol oxidase (PPO) was estimated by the method of Hori et al. (1997). The assay solution contained 20 mM catechol in 0.1 M phosphate buffer (pH 7). The reaction was started by addition of 100 μL of the enzymatic extract. PPO activity was

expressed in enzyme unit mg⁻¹ protein. One unit of PPO activity was defined as the amount of enzyme causing an increase in the absorbance of 0.001/min at 420 nm.

Malondialdehyde and Hydrogen Peroxide Content

Malondialdehyde (MDA) content in leaves was estimated by homogenizing the frozen leaf powder subsamples (0.25 g) in 10 mL of 0.1% (w/v) trichloroacetic acid (TCA) and centrifuging at 18,000 g for 10 min as described by Madhava Rao and Sresty (2000). Two milliliters of supernatant were mixed with 2 mL of 20% TCA containing 0.5% Thiobarbituric acid (TBA). The mixture was then heated in a water bath at 100°C for 30 min and immediately cooled in an ice bath. The absorbance was read at 532 nm. The nonspecific turbidity was corrected by subtracting A₆₀₀ from A₅₃₂, and the MDA content was calculated as follows: $[\text{MDA}] = 6.45 (A_{532} - A_{600}) - 0.56A_{450}$.

Hydrogen peroxide (H₂O₂) concentration in leaves was determined by the method described by Velikova et al. (2000). Briefly, 0.25 g of the frozen leaf powder were homogenized with 5 mL 10% (w/v) TCA and then centrifuged at 15,000 $x g$ for 15 min at 4°C. The supernatant (0.5 mL) was recovered to determine the content of H₂O₂ and 0.5 mL of potassium phosphate buffer (10 mM, pH 7) and 1 mL of iodine potassium (1 M) was added. After 1 h of incubation, the absorbance at 390 nm

was recorded and plotted against a standard H_2O_2 curve. The blank was made by replacing the sample extract by 10% TCA.

Soil Analyses

At plant harvest, soil physicochemical properties were analyzed on samples taken near the roots. The pH and electrical conductivity (EC) were measured in a diluted soil suspension 1/5 (v/v) using a pH meter HI 9025 and a conductivity meter HI-9033 (Hanna Instruments, Padua, Italy), respectively. Total organic carbon (TOC) and organic matter (OM) were measured according to the method described by Aubert (1978), which consists of the oxidation of organic matter by potassium dichromate in the presence of sulfuric acid. Available P was determined according to Olsen and Sommers method (1982). The amount mineral N available in soil was measured according to the method described by Rodier (1984).

Statistical Analysis

Data are presented as mean \pm SE (standard error) of six independent biological replicates. Data were analyzed by employing one-way analysis of variance (ANOVA) followed by Tukey's honest significant difference test using a significance level of 5% ($p \leq 0.05$). Normality of residuals was tested using the Shapiro-Wilk test. Mycorrhizal root colonization rates were arcsin-square root transformed to fit the assumption of normal distribution. Multivariate analysis of variance (MANOVA) was performed using SPSS 10.0 software to determine the interaction among the tested factors (AMF \times Bacteria \times Compost \times Drought). Different lower cases indicate significant differences among treatments at $p \leq 0.05$. In order to integrate all the data, a complete dataset comprising all growth, physiological, and biochemical data was subjected to Principal Components Analysis (PCA). The PCA was performed using XLSTAT v. 2014.

RESULTS

Mycorrhization Parameters

Our results showed that no mycorrhizal structure was observed in the roots of non-treatment controls. The frequency and intensity of AMF in date palm roots was significantly decreased by drought stress (**Supplementary Table 1**). The plants inoculated with AMF, especially for AMF1, without compost and PGPR showed the higher root colonization intensity compared to plants treated with compost and PGPR (**Figures 1A,B**). AMF infection frequency and intensity showed no significant difference between date palm inoculated with AMF alone or combined with PGPR and/or composts (bi- and tripartite combinations) under drought stress conditions (**Figures 1A,B**). The interactions between AMF and drought were significant for these two parameters (**Supplementary Tables 1A and B**).

Growth Assessment and Mineral Nutrition

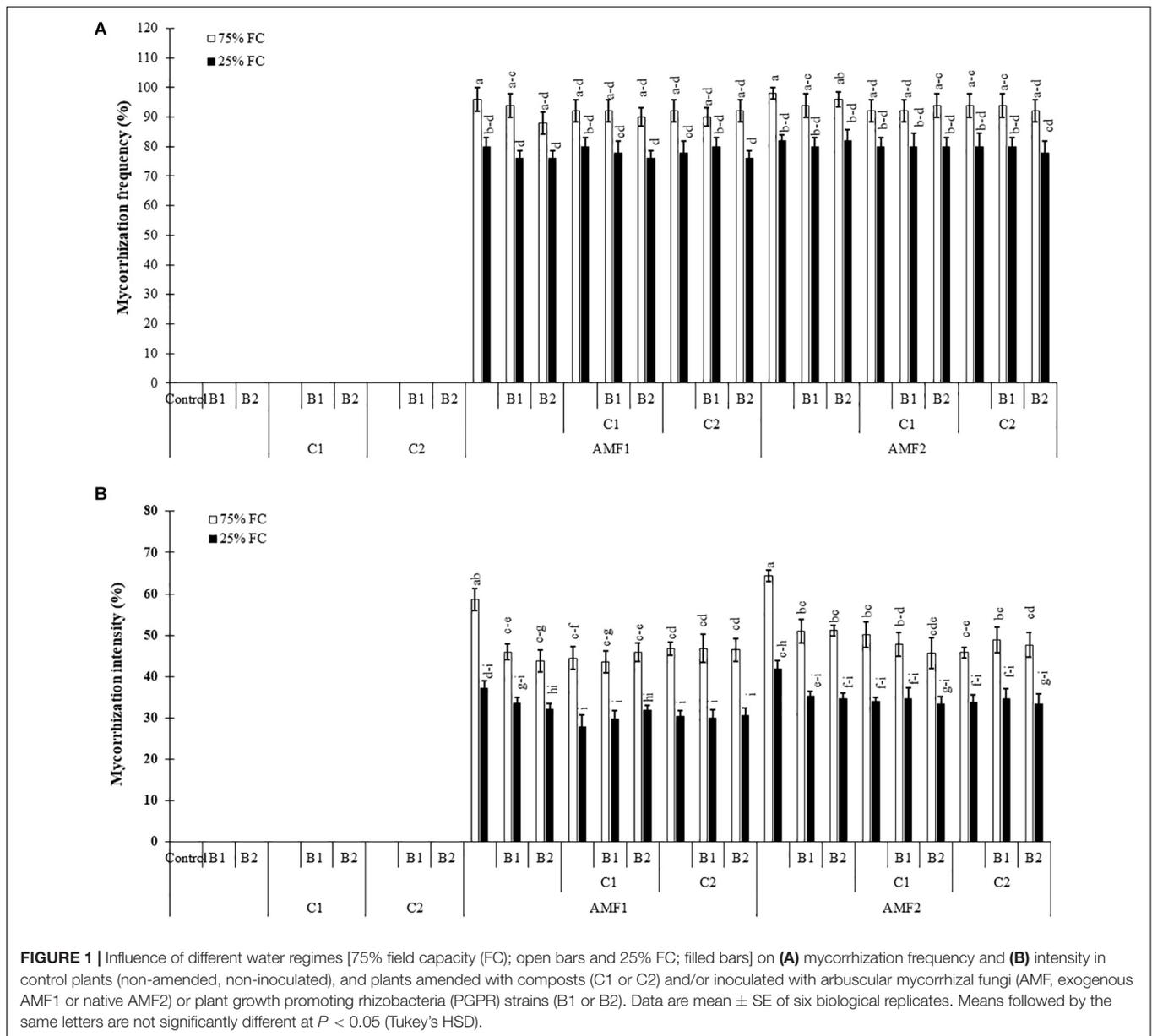
Drought caused a significant decrease ($P < 0.001$) (**Supplementary Table 1**) in all the growth parameters such

as leaf number, plant height, root length, leaf area (**Table 4**), and total dry weight (**Figure 2**). Our results showed that the un-inoculated and un-amended control performed very weak response in all these parameters compared to the treated plants under both well-watered and drought stress conditions (**Figure 2**, **Table 4**, and **Supplementary Figure 1**). Under drought stress, however, the application of bi- and tripartite combinations of biofertilizers (AMF1+C1, AMF1+C1+B1, AMF2+C2+B1, and AMF2+C1+B2) showed positive effects by promoting date palm shoot height and root length to a greater extent than in non-inoculated and non-amended plants. Moreover, the compost alone, bi and tripartite combinations (AMF1+C2+B1, AMF2+C2, C2) increased the number of leaves as compared to non-inoculated and non-amended date palm plants under well-watered and water deficit conditions. The plants treated with AMF1+C1, AMF2+B2, and AMF2+C2 improved the leaf area compared to non-inoculated and non-amended vitroplants under water deficit. A positive effect on the total dry weight of vitroplants subjected to water stress was recorded after application of biofertilizers (**Figure 2**). Indeed, the AMF and compost alone, bi and tripartite combinations (5 g in AMF2+C2 and C2, 4.8 g in AMF1+C1, and 4.6 g in AMF2, AMF1+B1, and AMF2+C1+B1) showed the highest values of this parameter under water deficit in comparison with non-inoculated and non-amended vitroplants (ca. 2.4 g).

We assayed the P and N content in shoots of date palm plants under drought and different biofertilizers, since the degree of stress and growth depend on their uptake and translocation. Under the control condition (75% FC), shoot P was significantly increased in plants treated with AMF (**Figure 3A**) as compared to non-amended and non-inoculated control plants, whereas under drought stress, it was decreased. Under water deficit, shoot P content was significantly increased by C1+B1, AMF1, AMF1+B2, AMF1+C1+B1, AMF2, AMF1+B1+C1, AMF2+C1+B2, and AMF2+C2 in comparison with non-treated plants (**Figure 3A**). Under 75% FC, N levels in leaves of all treated plants remained significantly higher than in control conditions. Drought stress decreased N content in all treatments, and all the biofertilizer treatments were able to maintain higher content than non-amended and non-inoculated control plants (**Figure 3B**). The interaction between drought \times C1 \times B1 (**Supplementary Table 1A**), drought \times B2, and drought \times C1 \times B2 (**Supplementary Table 1B**) had a significant effect ($P < 0.01$) on P, while N showed a significant effect between drought \times AMF2 (**Supplementary Table 1B**).

Physiological Traits

All physiological parameters were significantly decreased by drought ($P < 0.001$) (**Supplementary Table 1**). Under water scarcity, the leaf water potential values were decreased in non-inoculated and non-amended control plants. Plant inoculation with AMF and/or PGPR amended or not with the compost yielded an improvement in leaf water potential under water deficit, especially AMF1 (-1 MPa), AMF1+C1+B1 (-1.15 MPa), and AMF2+C2+B2 (-1.20 MPa) versus non-inoculated and non-amended plants (-2.18 MPa) (**Figure 4A**).



Under water control condition, there was obvious stomatal conductance differences between non-amended/non-inoculated and treated plants with AMF and/or PGPR. Under water stress, stomatal conductance values decreased in date palm plants. However, the application of biofertilizers increased stomatal conductance, with AMF1 alone, the bi- (C2+B1, AMF1+B1, AMF1+B2, AMF1+C2, and AMF2+B1) and tripartite (AMF1+C2+B1, AMF1+C2+B2, AMF2+C1+B2, and AMF2+C2+B2) combinations being the most effective in improving this parameter compared to control plants (**Figure 4B**).

As shown in **Figure 4C**, the chlorophyll fluorescence (F_v/F_m) was only slightly affected by drought stress. Biofertilizer application improved F_v/F_m in date palm plants under water shortage. The single (AMF1, PGPR B2 and compost C1),

bi- (AMF1+C2), and tripartite (AMF2+C1+B2) combinations presented the most effective treatments to increase chlorophyll fluorescence under water deficit conditions compared to non-inoculated and non-amended plants.

In response to drought stress and inoculation with AMF, PGPR and compost application, chlorophyll *a*, *b*, total chlorophyll, and carotenoid content are shown in **Figure 5**. Under water deficit, the photosynthetic pigment content was reduced. However, the application of AMF, compost, and PGPR especially the combination C2+B1, AMF2+C1+B2, AMF2+C2+B1, AMF1, and AMF2+C2+B2 increased pigments contents compared to control plants, under water stress conditions. As for carotenoid content, this was positively affected by biofertilizers applied alone (B1, B2, C2, C1, AMF2, and AMF1) or in combination (AMF2+C1, AMF2+C1+B1, AMF1+B2,

TABLE 4 | Influence of different water regimes on growth parameters of non-amended and non-inoculated plants (control), and plants amended and inoculated date palm plants with composts (C1 or C2) and/or arbuscular mycorrhizal fungi (AMF, exogenous AMF1 and native AMF2), and/or plant growth promoting rhizobacteria (PGPR) (B1 or B2).

Treatments	Leaf number		Shoot height (cm)		Root length (cm)		Leaf area (cm ²)	
	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC
Control	4.6 ± 0.2 fg	3.8 ± 0.2 g	23.6 ± 0.8 lm	21.1 ± 0.6 m	21.9 ± 0.6 n-p	18.5 ± 0.9 p	29.8 ± 1.4 g-j	21.7 ± 0.9 j
B1	5.2 ± 0.2 c-f	4.6 ± 0.2 fg	25.7 ± 0.5 c-m	23.4 ± 0.7 lm	25.2 ± 0.7 d-n	23.2 ± 1.6 i-o	35.0 ± 1.8 c-h	22.7 ± 1.4 ij
B2	6.4 ± 0.2 a-c	5.4 ± 0.2 b-f	27.2 ± 0.4 b-l	23.0 ± 0.7 lm	26.3 ± 1.1 a-m	22.4 ± 1.4 m-p	34.0 ± 1.8 d-h	26.3 ± 1.6 h-j
C1	6.4 ± 0.2 a-c	5.4 ± 0.2 b-f	28.8 ± 1.2 a-j	24.1 ± 0.6 j-m	26.1 ± 0.8 a-m	22.5 ± 1.0 l-p	39.8 ± 1.9 a-f	28.3 ± 1.0 g-j
C1+B1	6.2 ± 0.2 a-d	4.6 ± 0.2 fg	27.5 ± 0.9 b-l	25.0 ± 0.8 f-m	26.7 ± 1.1 a-k	21.9 ± 0.9 n-p	42.5 ± 0.9 a-d	32.7 ± 0.9 f-h
C1+B2	6.0 ± 0.3 a-e	5.0 ± 0.3 d-g	29.8 ± 0.8 a-f	26.6 ± 1.0 b-l	26.6 ± 0.5 a-l	23.4 ± 1.3 i-o	43.3 ± 2.1 a-c	29.3 ± 1.3 g-j
C2	6.8 ± 0.2 a	5.6 ± 0.2 a-f	30.1 ± 0.6 a-d	25.7 ± 0.5 c-m	27.1 ± 0.5 a-j	22.8 ± 0.8 k-o	45.3 ± 1.5 a	34.5 ± 0.8 c-h
C2+B1	6.2 ± 0.2 a-d	5.4 ± 0.2 b-f	27.6 ± 0.9 a-l	25.1 ± 0.9 e-m	24.5 ± 0.2 g-o	21.0 ± 0.4 op	42.0 ± 1.2 a-e	32.0 ± 0.7 f-h
C2+B2	5.8 ± 0.2 a-f	5.4 ± 0.2 b-f	31.0 ± 1.1 ab	26.7 ± 0.9 b-l	28.7 ± 0.9 a-f	23.4 ± 0.5 i-o	48.5 ± 1.7 a	33.0 ± 1.6 f-h
AMF1	6.4 ± 0.2 a-c	5.4 ± 0.2 b-f	30.0 ± 1.3 a-d	27.2 ± 0.5 b-l	27.9 ± 1.3 a-h	23.1 ± 0.4 j-o	43.3 ± 1.7 a-c	31.5 ± 0.7 f-i
AMF1+B1	6.2 ± 0.3 a-d	5.2 ± 0.2 c-f	30.2 ± 0.7 a-c	25.6 ± 0.9 c-m	29.3 ± 0.8 a-d	25.3 ± 0.5 d-n	44.0 ± 1.6 ab	33.0 ± 1.3 f-h
AMF1+B2	6.6 ± 0.2 ab	5.4 ± 0.2 b-f	29.6 ± 0.9 a-g	26.4 ± 0.6 b-l	29.2 ± 1.3 a-d	24.4 ± 0.5 g-o	43.0 ± 1.1 a-c	34.0 ± 1.1 d-h
AMF1+C1	5.6 ± 0.2 a-f	4.8 ± 0.2 e-g	29.9 ± 1.0 a-e	27.6 ± 0.7 a-l	30.0 ± 1.1 a	25.0 ± 0.5 e-o	46.0 ± 1.9 a	34.7 ± 1.5 c-h
AMF1+C1+B1	6.8 ± 0.2 a	5.2 ± 0.2 c-f	30.4 ± 1.3 a-c	27.5 ± 1.1 a-l	29.5 ± 0.5 a-c	27.1 ± 0.5 a-j	45.0 ± 2.2 a	31.3 ± 0.9 f-i
AMF1+C1+B2	5.8 ± 0.2 a-f	5.2 ± 0.2 c-f	29.5 ± 0.5 a-h	26.8 ± 0.9 b-l	30.0 ± 0.8 a	24.9 ± 0.6 e-o	42.0 ± 2.0 a-e	33.0 ± 1.3 f-h
AMF1+C2	6.4 ± 0.2 a-c	5.4 ± 0.2 b-f	28.6 ± 1.2 a-j	24.7 ± 0.7 h-m	28.2 ± 1.2 a-g	26.2 ± 0.6 a-m	45.3 ± 1.7 a	31.7 ± 1.1 f-h
AMF1+C2+B1	5.8 ± 0.2 a-f	5.8 ± 0.2 a-f	27.2 ± 0.8 b-l	24.4 ± 0.7 j-m	27.3 ± 0.4 a-i	24.7 ± 0.5 f-o	44.3 ± 1.9 ab	31.7 ± 1.6 f-h
AMF1+C2+B2	6.2 ± 0.2 a-d	5.0 ± 0.3 d-g	30.2 ± 0.6 a-c	24.8 ± 0.7 g-m	29.0 ± 0.7 a-e	25.6 ± 0.7 b-n	47.8 ± 1.7 a	32.6 ± 1.5 f-h
AMF2	6.2 ± 0.2 a-d	5.4 ± 0.2 b-f	28.9 ± 0.7 a-j	24.8 ± 0.6 g-m	28.8 ± 0.9 a-f	24.4 ± 0.5 g-o	47.8 ± 2.4 a	32.3 ± 1.1 f-h
AMF2+B1	6.0 ± 0.3 a-e	5.2 ± 0.2 c-f	29.5 ± 1.2 a-h	25.9 ± 0.6 c-m	27.9 ± 0.6 a-h	25.2 ± 0.3 d-n	46.3 ± 1.4 a	33.7 ± 1.7 d-h
AMF2+B2	6.2 ± 0.2 a-d	5.4 ± 0.2 b-f	28.5 ± 0.7 a-k	24.6 ± 0.6 i-m	28.8 ± 0.9 a-f	25.4 ± 0.5 c-n	47.5 ± 1.5 a	35.0 ± 0.9 c-h
AMF2+C1	5.8 ± 0.2 a-f	5.0 ± 0.0 d-g	32.3 ± 1.2 a	27.0 ± 0.7 b-l	26.0 ± 0.4 a-n	22.6 ± 0.7 k-p	43.3 ± 1.2 a-c	32.8 ± 1.1 f-h
AMF2+C1+B1	6.2 ± 0.2 a-d	5.4 ± 0.2 b-f	29.4 ± 0.9 a-i	23.8 ± 0.7 k-m	26.1 ± 0.5 a-m	24.0 ± 0.7 h-o	44.5 ± 2.0 ab	34.0 ± 1.4 d-h
AMF2+C1+B2	5.8 ± 0.2 a-f	5.2 ± 0.2 c-f	31.0 ± 0.6 ab	28.4 ± 0.8 a-k	29.6 ± 1.2 ab	27.1 ± 0.7 a-j	45.3 ± 2.2 a	33.5 ± 1.6 e-h
AMF2+C2	6.2 ± 0.2 a-d	5.6 ± 0.2 a-f	29.9 ± 1.3 a-e	26.8 ± 0.4 b-l	26.2 ± 0.8 a-m	23.5 ± 0.5 i-o	46.0 ± 2.1 a	36.0 ± 1.1 b-g
AMF2+C2+B1	6.0 ± 0.3 a-e	4.8 ± 0.2 e-g	32.3 ± 1.1 a	27.3 ± 0.9 b-l	28.5 ± 0.4 a-g	24.8 ± 0.6 f-o	45.0 ± 1.9 a	32.3 ± 1.4 f-h
AMF2+C2+B2	6.0 ± 0.0 a-e	4.8 ± 0.0 e-g	29.9 ± 0.6 a-d	25.3 ± 0.5 d-m	26.5 ± 0.6 a-m	22.7 ± 0.7 k-o	44.5 ± 0.8 ab	33.8 ± 1.6 d-h

Means followed by the same letters are not significantly different $P < 0.05$ (Tukey's HSD).

AMF2+C2+B2, AMF2+C1+B2, and AMF2+C2+B1) as compared with non-inoculated with AMF/PGPR and non-amended with composts, under water deficit.

Biochemical Traits

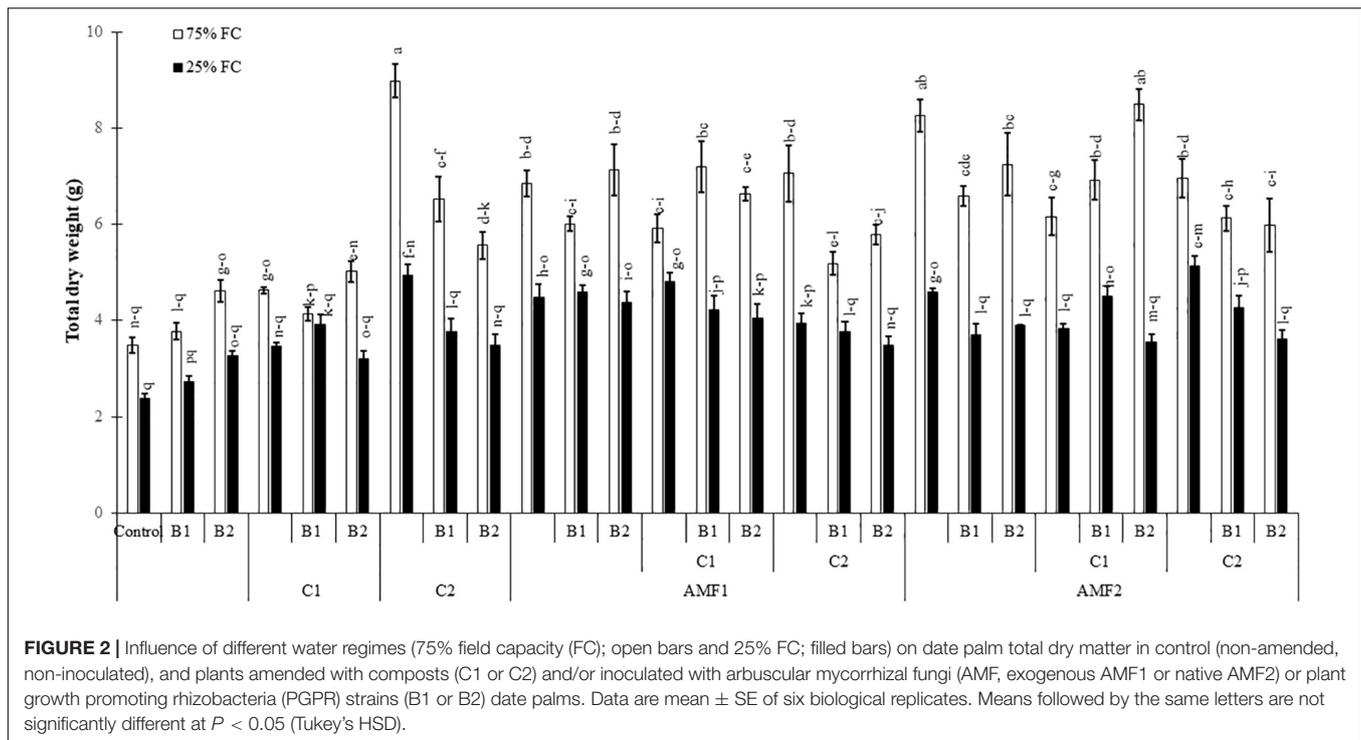
Treatment effects on biochemical traits were significantly decreased by drought ($P < 0.001$) (Supplementary Table 1). Results related to the effect of drought stress and biofertilizer applications on sugar and protein content and POX and PPO activities in date palm plants are presented in Figure 6. Under normal water conditions, both compost and AMF increased sugar and protein content. Exposure to water deficit caused a significant decrease in sugar and protein content (Figures 6A,B). The addition of biofertilizers yielded a significant increase in sugar and protein compared to stressed control plants. Under 75% FC conditions, POX and PPO did not differ significantly among the biofertilizers treatments (Figures 6C,D) Exposure to drought stress led to a considerable increase in the POX and PPO specific activities as compared to non-treated control plants.

To characterize damage caused by drought stress, we carried out MDA and H₂O₂ analyses (Figure 7). The exposure of date

palm plants to severe water deficit resulted in an increase in MDA and H₂O₂ content. Under water stress, in contrast, the application of single or combined biofertilizers showed reduced MDA and H₂O₂ content compared to non-inoculated and non-amended controls. The interactions AMF1 × C2 × Drought, and AMF1 × B1 × Drought (Supplementary Table 1A), AMF2 × C2 × Drought, B1 × Drought, and B1 × Drought (Supplementary Table 1B) had a significant effect on H₂O₂ content.

Soil Analysis

We assayed the pH, electrical conductivity, total organic carbon, total organic matter, N, and P content in the soil just after harvesting the plants, since the degree of soil quality depends on their values after the culture (Table 5). Under control conditions, the addition of composts increased the soil quality more than controls. Under drought stress, soil analyses at harvest time showed that biofertilizers application - mainly composts - improved soil quality as compared to the controls (Table 5). As a result, relative to control plants, the soil pH was decreased following the application of biofertilizers



after prolonged drought. Under these conditions, EC, TOC, and OM were improved by biofertilizers. Moreover, under drought stress, both total N and P content in the soil were improved when biofertilizers were applied, especially in the treatments AMF1+B2, AMF1+C2+B1, AMF2, AMF2+B2, AMF2+C1+B2, and AMF2+C2.

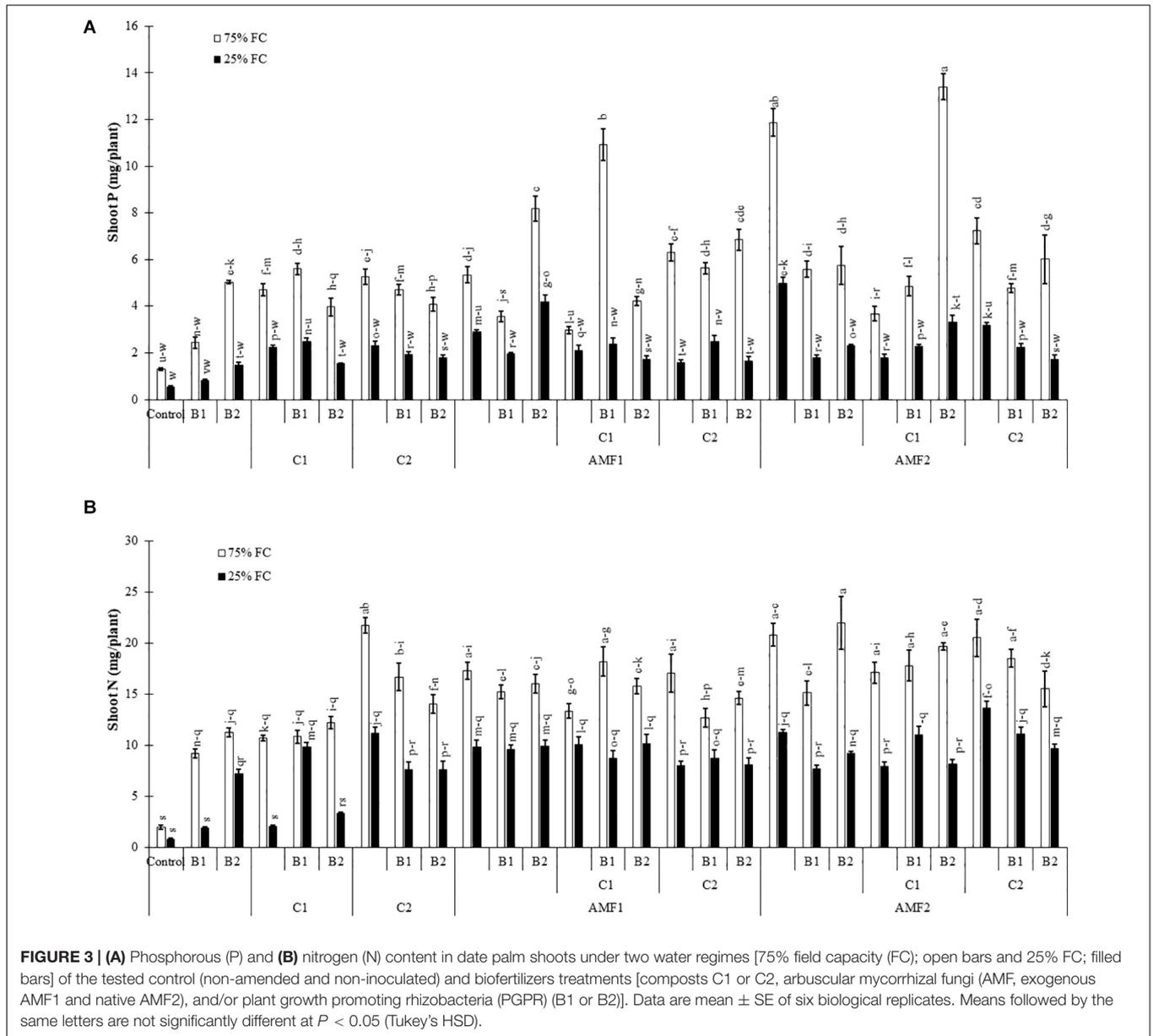
Principal Component Analysis (PCA)

The PCA showed that AMF alone or combined with compost and/or PGPR were the most effective treatments to improve growth, nutrition, osmolytes and antioxidant traits (shown in the right panel of **Figure 8A**) under drought stress (**Figure 8A**). PC1 explained 40.8% and PC2 explained 11.9% of the total variance. **Figure 8B** showed that all biofertilizer treatments, single or combined (right panel of **Figure 8B**), were separate from the control. In **Figure 8A**, we observed in the right lower panel of the PC1 component, that the traits PPO, F_v/F_m , OM, TOC, Total Chlorophyll, chlorophyll *a* and *b*, shoot height, carotenoid, and leaf water potential were closely related to soil P and N concentration, shoot P and N content, sugar, root length, leaf area, leaf number, POX, protein, mycorrhizal frequency and intensity, and EC. In contrast, relative to biofertilizer treatments, the non-treatment control separated in the left of the PC1 component (**Figure 8B**) and was related to H_2O_2 , MDA and pH traits (**Figure 8A**).

DISCUSSION

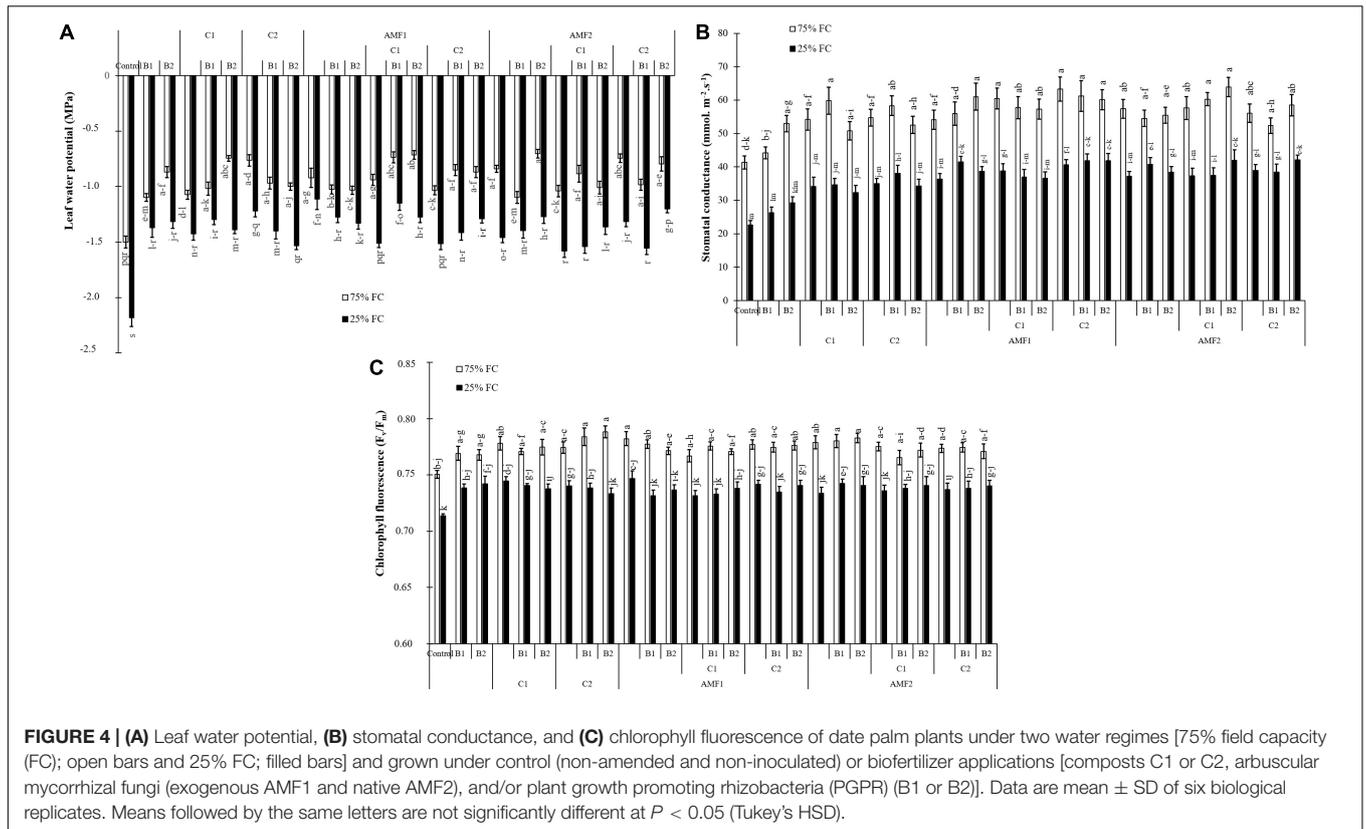
In the present study, the application of composts together with inoculation of the exotic and native AMF and PGPR strains

inoculations either as single or combined treatments were very effective in helping date palm plants to attenuate the detrimental effects of drought stress on growth, photosynthetic apparatus, nutrient uptake, physiological traits, and oxidative stress. The treated biofertilizers assayed showed a high level of osmotic stress tolerance under water deficit. Our results show that no mycorrhizal structure was observed in the roots of non-treatment controls, but the plants inoculated with AMF were successfully infected by the native or exogenous mycorrhizal consortium under both well-watered and drought conditions. In the presence of water stress, mycorrhization intensity decreased in the AMF treatments alone or in combination with compost and/or PGPR. Our results are in line with several studies showing that mycorrhizal infection decreased when the host plants were exposed to drought stress (Baslam and Goicoechea, 2012; Meddich et al., 2015a; Paymaneh et al., 2019). Under drought stress, however, our findings show an increase of the mycorrhization intensity in date palms treated by compost and PGPR alone or in combination. Cavagnaro (2014) and Kohler et al. (2015) reported that the application of compost at a low dose (2 and 12.5%) increased AMF infection in the root system. Other studies have shown a positive effect of PGPR on enhancing root infection by AMF (Ben-Laouane et al., 2019; Raklami et al., 2019). On the other hand, Sghir et al. (2014) noted that mycorrhizal frequencies and arbuscular content decreased significantly in palm trees inoculated with the combination AMF-PGPR *Trichoderma harzianum* as compared to plants inoculated only with AMF because it colonizes the same space as AMF. However, dual-inoculation AMF-*Trichoderma* led to the best date palm growth (Sghir et al., 2014). The application of different biofertilizers (alone or in combination) benefited plant growth



(mainly leaf number, shoot height, root length, leaf area, and total dry biomass production). The beneficial effect of mycorrhizal and PGPR associations and compost amendment on growth of date palms, under water deficit, could be explained by the greater uptake of nutrients with low mobility such as P and N contained in the substrate. Previous studies demonstrated that date palm and other plants inoculated with AMF accumulated more N and P in leaves than non-mycorrhizal plants when subjected to drought stress (Meddich et al., 2015c; Hao et al., 2019). Javan Gholiloo et al. (2019) showed that the application of biofertilizers (AMF and PGPR) improve the P and N nutrition and consequently enhances date palm plant growth under deficit conditions. Nadeem and colleagues (2014) showed that AMF and PGPR can regulate mineral nutrition by solubilizing nutrients in the soil and producing plant growth regulators (i.e.,

hormones). Several studies have indicated that compost, AMF and PGPR improve plant growth through the assimilation of immobile soil nutrients such as N and P (Yadav et al., 2013; Baslam et al., 2014; Al-Karaki, 2016; Barje et al., 2016; Frosi et al., 2016; Yang et al., 2018; Raklami et al., 2019; Yu et al., 2019). Here we showed that inoculated and amended date palm plants had considerably higher mineral nutrient content (P and N) as compared to controls under water deficit conditions allowing higher plant performance. This resulted from the better absorption of the surface area provided by extensive fungal hyphae (Wu and Zou, 2017; Zhang et al., 2018) and/or a direct uptake from compost to plant root (Kohler et al., 2015) and/or the mobilization and absorption of various nutrients from soil to plants by PGPR (Gobelak et al., 2015). AMF-colonization results in the establishment of extensive hyphal networks and glomalin



secretion, fueling plants with water and nutrient, and thereby, enhancing soil structure (Pagano, 2014). Recently, Volpe et al. (2016) reported the role of PT family genes as components of the Pi-sensing machinery in root tips, which is up-regulated in AMF colonized plants. In addition to P and N, evidence of the role of AMF and PGPR symbiosis in the “transportome” of several mineral nutrients has been obtained in studies on several plant species (Hogekamp et al., 2011; Casieri et al., 2013).

Physiological traits constitute an important tool to study the effect of drought stress on many plants. Our results showed an increase in leaf water potential, stomatal conductance, F_v/F_m , and chlorophyll pigment synthesis in plants inoculated with AMF and PGPR and/or amended by composts. This improvement of physiological traits can lead to an increase in CO_2 assimilation for photosynthesis. Similarly, several studies have demonstrated the capacity of AMF inoculation to reduce the negative effect of drought stress on date palm growth by improving photosynthesis, water status, and antioxidant activity (Baslam et al., 2014; Meddich et al., 2015a). In our research under drought stress, treated plants reduced the degradation of total chlorophyll *a* and *b* and carotenoid. A higher photosynthetic pigment under drought stress conditions suggests a better performance of the photosynthetic apparatus. Our data show that biofertilizers not only increase water and nutrient uptake to mitigate the negative effect of drought but also improve stomatal conductance. Several studies have reported the existence of a positive correlation between photosynthetic efficiency maintenance and tolerance to drought stress in plants amended with compost and/or

inoculated with AMF/PGPR (Wu et al., 2006; Sandhya et al., 2010; Tartoura, 2010; Abd El-Mageed et al., 2018, 2019; Duo et al., 2018; Khosravi Shakib et al., 2019). In response to drought stress, plants treated with biofertilizers showed lower levels of potentials and higher water content, allowing the inoculated and amended plants to sustain high organ hydration and turgor level which maintain overall physiological activities of the cells, especially those linked to the photosynthetic apparatus. Another study showed that the positive effect of compost on soil was related to the improvement of water retention (Tartoura, 2010). The association with AMF amends the plants' water regulation by triggering hormonal signaling such as ABA-mediated stomatal conductance or by stimulating osmolytes. Other studies showed, under drought stress, the development of microorganisms-mediated mechanisms including modifications in the content of plant hormones (e.g., strigolactones, jasmonic acid, and abscisic acid) and improvement in plant water status by increasing hydraulic conductivity (Chaumont and Tyerman, 2014; Fernández-Lizarazo and Moreno-Fonseca, 2016). The increase in root hydraulic conductivity can be related to an enhanced expression in fungal or plant AQPs (Sánchez-Romera et al., 2016). Chitarra et al. (2016) showed an enhancement in the water transport capacity of treated roots, which correlated with overexpression of the NIP AQP-encoding gene (*LeNIP3;1*). Recently, Xie et al. (2018) found up-regulation of the root AQP gene *PIP* expression under moderate water deficit in AMF roots.

Soil organic matter and total organic carbon were improved by the biofertilizers used. This improvement could be explained

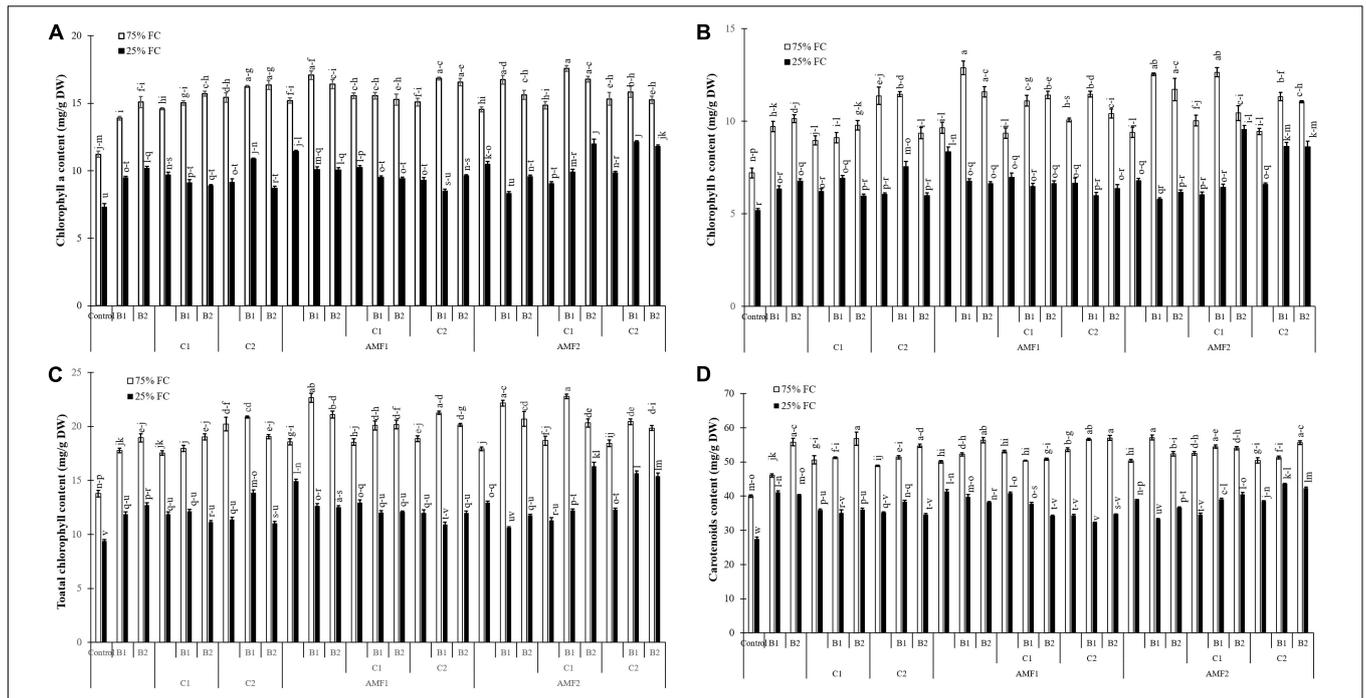


FIGURE 5 | (A) Chlorophyll a, **(B)** chlorophyll b, **(C)** total chlorophyll, and **(D)** carotenoid content in leaves of date palm plants under two water regimes [75% field capacity (FC); open bars and 25% FC; filled bars] and further grown without (control; non-amended and non-inoculated) or with biofertilizers [composts C1 or C2, arbuscular mycorrhizal fungi (AMF, exogenous AMF1 and native AMF2), and/or PGPR (B1 or B2)]. Data are mean \pm SE of six independent biological replicates. Means followed by the same letters are not significantly different at $P < 0.05$ (Tukey's HSD).

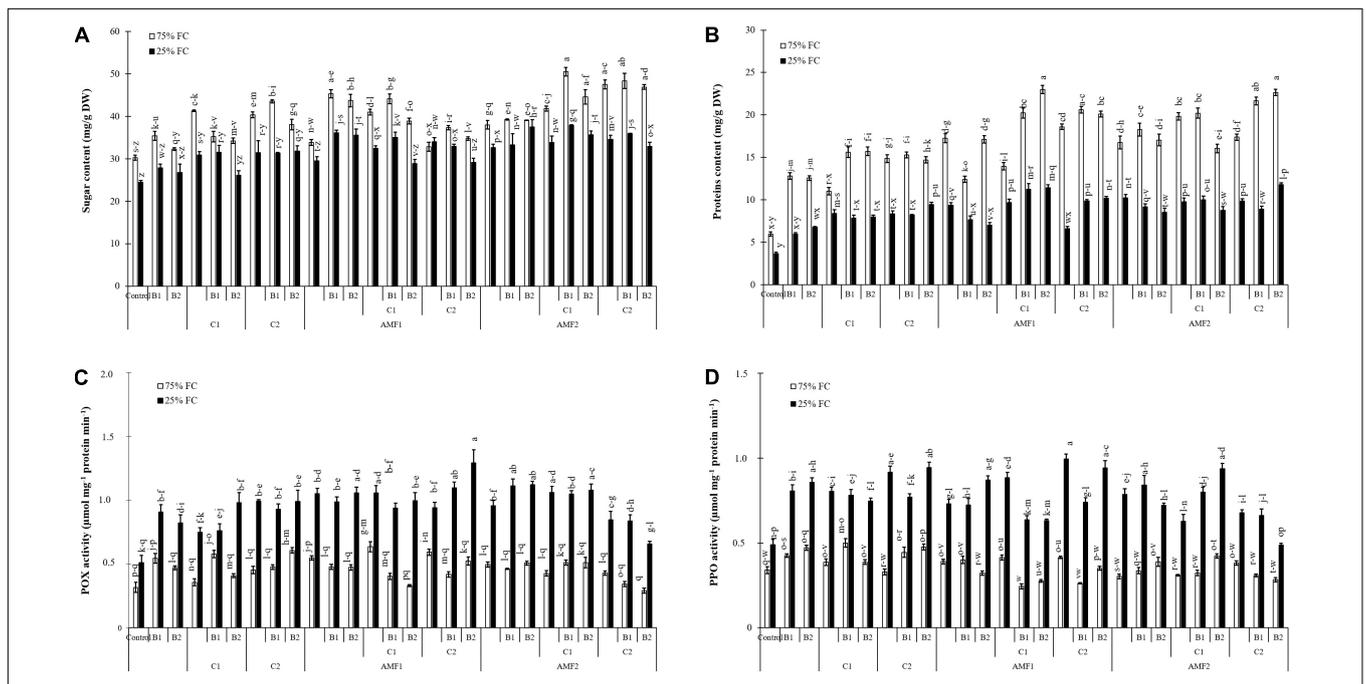
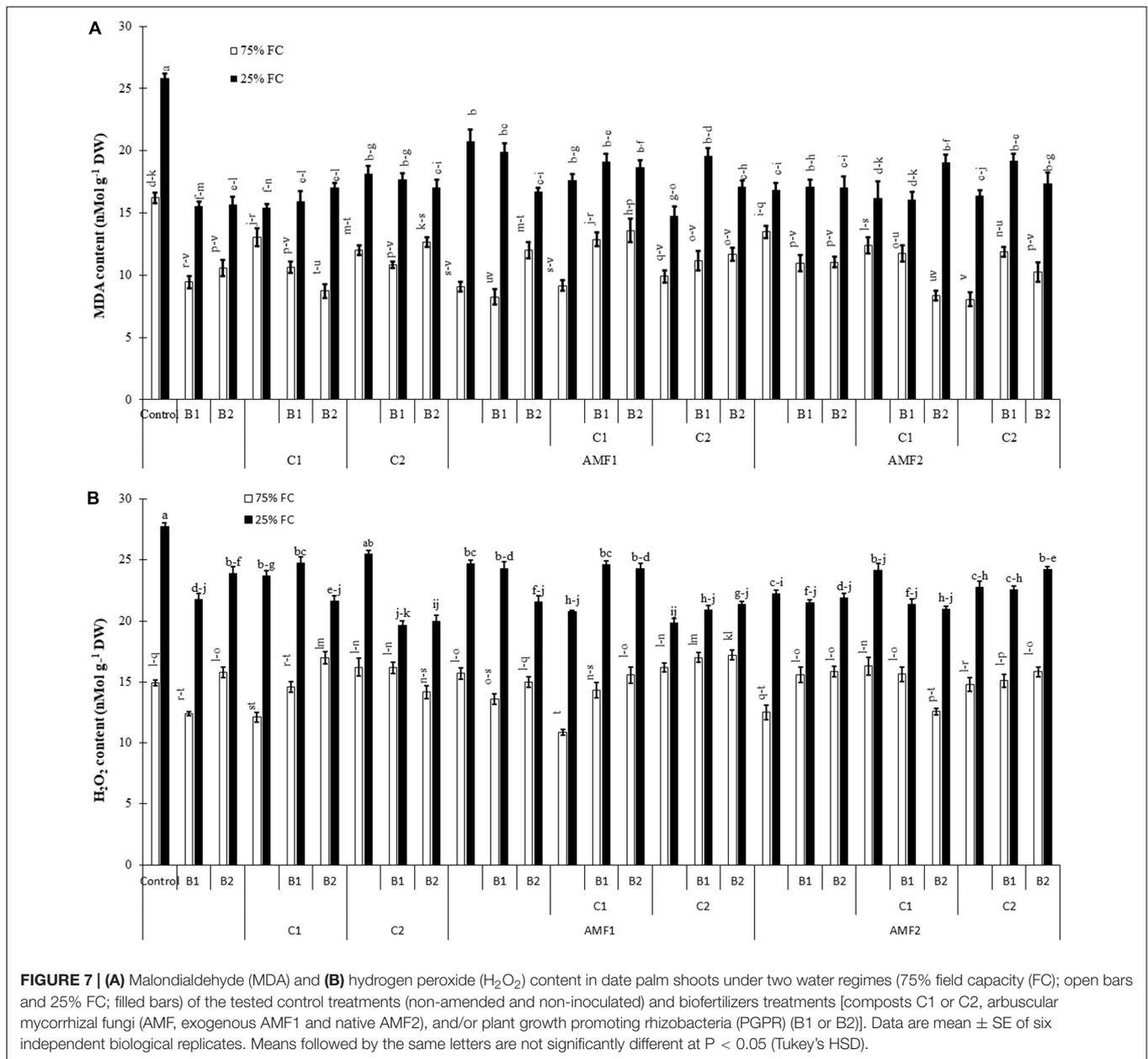


FIGURE 6 | (A) Total soluble sugar content, **(B)** protein content, **(C)** peroxidase (POX) activity, and **(D)** polyphenol oxidase (PPO) activity in date palm shoots under two water regimes [75% field capacity (FC); open bars and 25% FC; filled bars] of the tested control treatments (non-amended and non-inoculated) and biofertilizers treatments [composts C1 or C2, arbuscular mycorrhizal fungi (exogenous AMF1 and native AMF2), and/or plant growth promoting rhizobacteria (PGPR) (B1 or B2)]. Data are mean \pm SE of six biological replicates. Means followed by the same letters are not significantly different at $P < 0.05$ (Tukey's HSD).



by a direct contribution from compost or by the ability of AMF and PGPR to metabolize different compounds produced by plant roots mainly carbohydrates and organic acids. Shen et al. (2013) and Ning et al. (2017) showed that inoculation with microorganisms and compost application to plants was very effective in improving soil quality especially in organic matter, water retention, and mineral nutrition. The results obtained showed that biological treatments had an important effect on N and P in soil. However, a high amount of N and P in the rhizosphere soil of plants amended with compost and inoculated with AMF and PGPR could be due to a direct absorption via compost or by the fact that AMF and PGPR efficiently and directly take up from the soil to the plant nutrients such as N and P (Grobela et al., 2015; Liu et al., 2018). In order to

tolerate drought stress, plant adaptation is associated with high concentration of solutes such as soluble sugars and protein to regulate the osmotic potential of cells which, in turn, induce an improvement in water absorption under unfavorable condition (Zhang et al., 2010; Liao et al., 2019). Our data indicated that the concentrations of soluble sugars and protein in leaves increased during drought stress in treated plants as compared to microbes-free controls. These results are in agreement with previous reports using AMF or PGPR (Abbaspour et al., 2012; Vurukonda et al., 2016; Javan Gholiloo et al., 2019). In fact, PGPR were shown to secrete osmolytes to mitigate drought stress, which act synergistically with plants internal osmolytes boosting plant growth (Paul et al., 2008). Bano et al. (2013) and Sandhya et al. (2010) reported that plants inoculated with PGPR and

TABLE 5 | Soil physico-chemical analysis at harvest time of date palm grown under two water regimes (75 and 25% Field Capacity (FC)) of the tested control (non-amended and non-inoculated) and biofertilizers (composts C1 or C2, arbuscular mycorrhizal fungi (AMF, exogenous AMF1 and native AMF2), and/or plant growth promoting rhizobacteria (PGPR) (B1 or B2). Data are mean \pm SE of six biological replicates.

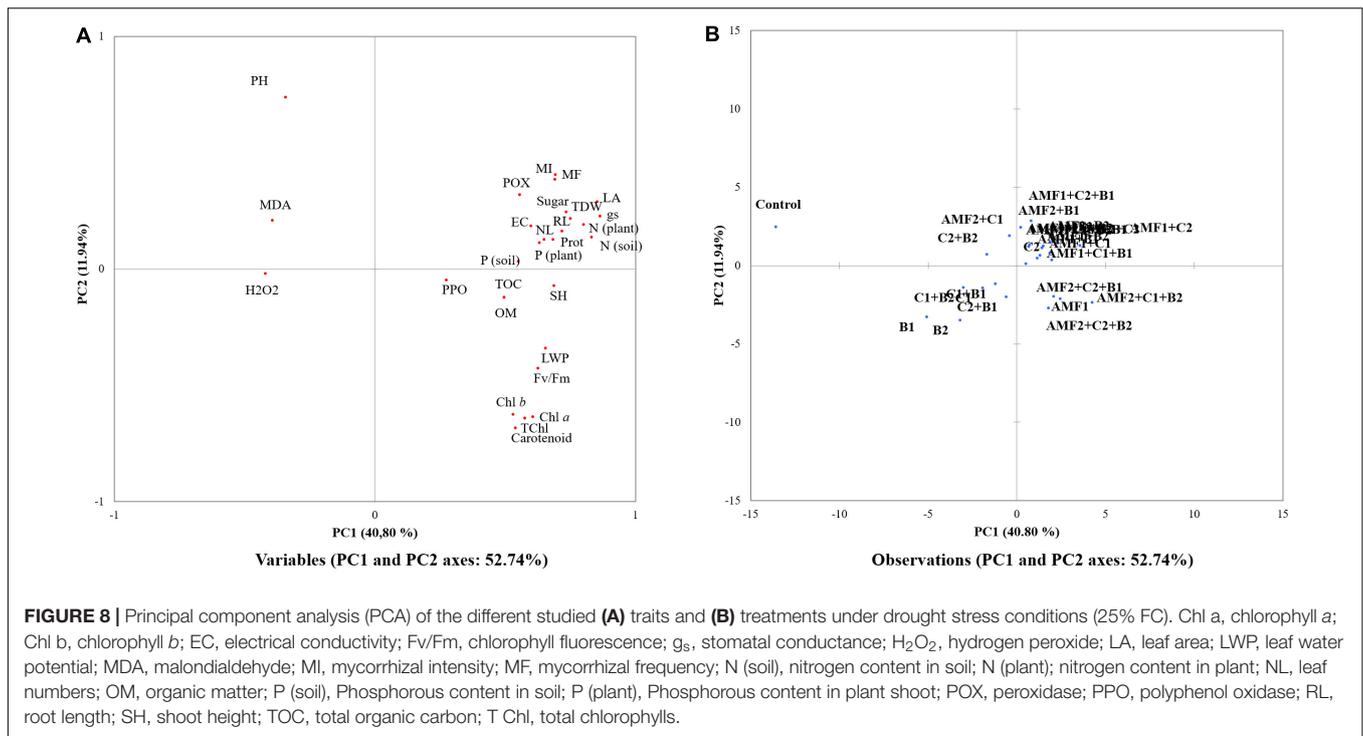
Treatments	pH		EC (mS/cm)		TOC (%)		OM (%)		N (mg/g of soil)		P (ppm)	
	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC
Control	7.96 \pm 0.05 a	7.88 \pm 0.06 ab	0.28 \pm 0.01 v-x	0.20 \pm 0.01 A	0.40 \pm 0.02 v	0.52 \pm 0.01 u	0.70 \pm 0.03 v	0.89 \pm 0.02 u	0.16 \pm 0.02 lm	0.06 \pm 0.02 m	51.97 \pm 1.47 u-w	30.05 \pm 1.44 w
B1	7.50 \pm 0.07 c-e	7.46 \pm 0.02 c-e	0.23 \pm 0.00 y-A	0.21 \pm 0.01 zA	0.75 \pm 0.01 n-s	0.71 \pm 0.01 p-t	1.29 \pm 0.02 n-s	1.22 \pm 0.02 p-t	0.30 \pm 0.03 e-k	0.22 \pm 0.02 j-l	86.53 \pm 5.37 m-t	41.34 \pm 1.64 vw
B2	7.38 \pm 0.04 c	7.42 \pm 0.02 de	0.25 \pm 0.01 x-z	0.21 \pm 0.01 zA	0.68 \pm 0.06 st	0.74 \pm 0.02 o-s	1.16 \pm 0.10 st	1.28 \pm 0.04 o-s	0.30 \pm 0.03 e-k	0.22 \pm 0.02 j-l	151.48 \pm 7.02 c-e	57.97 \pm 3.46 s-w
C1	7.46 \pm 0.05 c-e	7.56 \pm 0.02 c-e	0.30 \pm 0.01 s-w	0.31 \pm 0.01 r-v	0.87 \pm 0.03 e-l	0.83 \pm 0.01 h- o	1.51 \pm 0.05 e-l	1.43 \pm 0.03 h- o	0.24 \pm 0.02 j-l	0.34 \pm 0.04 b-k	140.04 \pm 11.28 c-g	86.59 \pm 3.88 m-s
C1+B1	7.44 \pm 0.05 c-e	7.54 \pm 0.02 c-e	0.39 \pm 0.01 h- n	0.39 \pm 0.01 h- n	0.94 \pm 0.02 b-g	0.90 \pm 0.00 e-k	1.62 \pm 0.03 b-g	1.55 \pm 0.01 e-k	0.34 \pm 0.04 b-k	0.22 \pm 0.02 j-l	190.18 \pm 9.77 b	80.89 \pm 1.68 n-u
C1+B2	7.52 \pm 0.04 c-e	7.48 \pm 0.04 c-e	0.38 \pm 0.01 i-o	0.21 \pm 0.01 zA	0.92 \pm 0.01 d-i	0.84 \pm 0.01 g- n	1.58 \pm 0.03 d-i	1.46 \pm 0.02 g- n	0.32 \pm 0.02 c-k	0.25 \pm 0.01 j-l	105.63 \pm 8.57 h-n	64.06 \pm 1.36 q-v
C2	7.42 \pm 0.04 de	7.48 \pm 0.04 c-e	0.38 \pm 0.01 h- o	0.37 \pm 0.01 j-p	0.94 \pm 0.02 b-g	0.87 \pm 0.02 e-l	1.62 \pm 0.03 b-g	1.50 \pm 0.03 e-l	0.47 \pm 0.02 ab	0.28 \pm 0.02 g- l	79.43 \pm 3.91 n-u	61.90 \pm 2.57 r-v
C2+B1	7.52 \pm 0.05 c-e	7.58 \pm 0.02 c-e	0.32 \pm 0.00 q-v	0.31 \pm 0.00 r-v	0.95 \pm 0.02 b-f	0.84 \pm 0.02 g- o	1.64 \pm 0.03 b-f	1.44 \pm 0.03 g- o	0.35 \pm 0.02 a-k	0.21 \pm 0.02 kl	96.76 \pm 3.17 k-p	71.07 \pm 3.07 o-u
C2+B2	7.50 \pm 0.04 c-e	7.58 \pm 0.04 c-e	0.35 \pm 0.01 n-r	0.33 \pm 0.01 p-u	0.91 \pm 0.02 e-j	0.68 \pm 0.01 r-t	1.57 \pm 0.03 e-j	1.16 \pm 0.03 r-t	0.32 \pm 0.02 c-k	0.26 \pm 0.02 i-l	100.34 \pm 2.42 j-o	69.50 \pm 4.30 p-v
AMF1	7.56 \pm 0.02 c-e	7.52 \pm 0.04 c-e	0.25 \pm 0.01 x-z	0.23 \pm 0.00 y-A	0.75 \pm 0.01 n-s	0.68 \pm 0.01 r-t	1.29 \pm 0.02 n-s	1.17 \pm 0.03 r-t	0.29 \pm 0.02 f-l	0.32 \pm 0.02 c-k	114.85 \pm 5.74 g-m	87.29 \pm 4.24 m-s
AMF1+B1	7.64 \pm 0.02 b-d	7.56 \pm 0.02 c-e	0.25 \pm 0.01 x-z	0.27 \pm 0.01 w-y	0.70 \pm 0.03 p-t	0.78 \pm 0.00 l-q	1.21 \pm 0.05 p-t	1.35 \pm 0.01 l-q	0.39 \pm 0.03 a-i	0.34 \pm 0.02 b-k	80.54 \pm 2.33 n-u	57.87 \pm 2.25 s-w
AMF1+B2	7.68 \pm 0.04 bc	7.56 \pm 0.02 c-e	0.30 \pm 0.01 t-w	0.29 \pm 0.01 u-x	0.70 \pm 0.01 q-t	0.64 \pm 0.02 t	1.20 \pm 0.01 q-t	1.10 \pm 0.04 t	0.32 \pm 0.02 c-k	0.28 \pm 0.02 g- l	156.47 \pm 8.83 cd	126.84 \pm 1.00 e-j
AMF1+C1	7.48 \pm 0.04 c-e	7.54 \pm 0.02 c-e	0.39 \pm 0.01 h- o	0.31 \pm 0.01 r-w	0.90 \pm 0.00 e-j	0.80 \pm 0.01 k-p	1.56 \pm 0.00 e-j	1.38 \pm 0.02 k-p	0.41 \pm 0.03 a-g	0.31 \pm 0.01 d-k	68.55 \pm 1.05 p-v	58.47 \pm 3.04 r-w
AMF1+C1+B1	7.50 \pm 0.09 c-e	7.48 \pm 0.04 c-e	0.45 \pm 0.01 d-g	0.34 \pm 0.01 o-t	0.97 \pm 0.01 a-e	0.77 \pm 0.02 m-r	1.67 \pm 0.01 a-e	1.32 \pm 0.03 m-r	0.31 \pm 0.03 d-k	0.25 \pm 0.01 j-l	207.15 \pm 3.30 ab	74.39 \pm 3.45 o-u
AMF1+C1+B2	7.64 \pm 0.02 b-d	7.68 \pm 0.04 bc	0.41 \pm 0.01 g- k	0.32 \pm 0.01 q-v	1.07 \pm 0.00 a	0.89 \pm 0.02 e-k	1.85 \pm 0.00 a	1.54 \pm 0.04 e-k	0.36 \pm 0.03 a-j	0.31 \pm 0.01 d-k	86.99 \pm 1.47 m-s	57.06 \pm 2.51 t-w
AMF1+C2	7.65 \pm 0.05 c-e	7.68 \pm 0.04 bc	0.49 \pm 0.01 cd	0.41 \pm 0.01 f-j	0.90 \pm 0.01 e-k	1.03 \pm 0.01 a-c	1.55 \pm 0.02 e-k	1.78 \pm 0.01 a-c	0.44 \pm 0.02 a-e	0.31 \pm 0.02 d-k	127.90 \pm 12.23 d-j	58.22 \pm 3.26 s-w
AMF1+C2+B1	7.66 \pm 0.06 b-d	7.68 \pm 0.04 bc	0.55 \pm 0.01 b	0.42 \pm 0.01 e-i	1.05 \pm 0.02 ab	0.93 \pm 0.00 c-h	1.80 \pm 0.03 ab	1.60 \pm 0.01 c-h	0.32 \pm 0.02 c-k	0.29 \pm 0.02 f-l	149.41 \pm 8.46 c-f	87.94 \pm 3.18 m-r
AMF1+C2+B2	7.48 \pm 0.07 c-e	7.68 \pm 0.04 bc	0.45 \pm 0.00 d-f	0.40 \pm 0.01 h- m	1.02 \pm 0.01 a-d	0.82 \pm 0.01 i-o	1.77 \pm 0.02 a-d	1.41 \pm 0.02 i-o	0.31 \pm 0.02 a-d	0.29 \pm 0.02 f-l	157.48 \pm 7.11 c	66.08 \pm 3.09 q-v
AMF2	7.60 \pm 0.03 c-e	7.66 \pm 0.04 b-d	0.35 \pm 0.01 m-r	0.32 \pm 0.01 q-v	0.53 \pm 0.00 u	0.64 \pm 0.01 t	0.91 \pm 0.00 u	1.10 \pm 0.02 t	0.35 \pm 0.03 a-k	0.34 \pm 0.03 b-k	198.64 \pm 8.48 ab	149.41 \pm 8.35 c-f

(Continued)

TABLE 6 | Continued

Treatments	pH		EC (mS/cm)		TOC (%)		OM (%)		N (mg/g of soil)		P (ppm)	
	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC	75% FC	25% FC
AMF2+B1	7.56 ± 0.05 c-e	7.68 ± 0.04 bc	0.33 ± 0.01 p-u	0.28 ± 0.01 v-x	0.66 ± 0.01 st	0.78 ± 0.01 l-q	1.14 ± 0.02 st	1.35 ± 0.01 l-q	0.45 ± 0.03 a-d	0.31 ± 0 01 d-k	121.65 ± 2.58 f-k	65.12 ± 2.55 q-v
AMF2+B2	7.66 ± 0.04 b-d	7.60 ± 0.03 c-e	0.37 ± 0.01 k-p	0.36 ± 0.01 l-q	0.85 ± 0.02 f-m	0.84 ± 0.01 g- o	1.47 ± 0.04 f-m	1.44 ± 0.03 g- o	0.35 ± 0.03 a-k	0.36 ± 0.03 a-j	108.20 ± 5.69 h-n	83.21 ± 0.96 n-t
AMF2+C1	7.58 ± 0.04 c-e	7.64 ± 0.02 b-d	0.43 ± 0.01 e-h	0.38 ± 0.01 i-o	1.05 ± 0.01 ab	0.75 ± 0.01 n-s	1.80 ± 0.02 ab	1.30 ± 0.02 n-s	0.45 ± 0.02 a-d	0.29 ± 0.02 f-l	82.05 ± 1.52 n-t	62.10 ± 3.00 r-v
AMF2+C1+B1	7.64 ± 0.05 b-d	7.56 ± 0.04 c-e	0.42 ± 0.01 e-i	0.34 ± 0.01 n-s	0.92 ± 0.02 d-i	0.69 ± 0.01 q-t	1.59 ± 0.03 d-i	1.19 ± 0.01 q-t	0.40 ± 0.03 a-h	0.32 ± 0.02 c-k	92.03 ± 2.27 l-q	64.32 ± 1.73 q-v
AMF2+C1+B2	7.66 ± 0.04 b-d	7.52 ± 0.04 c-e	0.48 ± 0.01 cd	0.40 ± 0.01 h-l	0.81 ± 0.01 i-o	0.92 ± 0.02 d-i	1.40 ± 0.01 i-o	1.58 ± 0.04 d-i	0.48 ± 0.02 a	0.32 ± 0.02 c-k	223.33 ± 3.07 a	121.35 ± 6.35 f-l
AMF2+C2	7.54 ± 0.04 c-e	7.58 ± 0.05 c-e	0.65 ± 0.01 a	0.51 ± 0.01 bc	0.85 ± 0.02 f-m	0.90 ± 0.01 e-j	1.47 ± 0.03 f-m	1.55 ± 0.02 e-j	0.43 ± 0.03 a-f	0.32 ± 0.04 c-k	134.55 ± 6.93 c-h	81.90 ± 3.92 n-t
AMF2+C2+B1	7.64 ± 0.02 b-d	7.52 ± 0.04 c-e	0.51 ± 0.01 bc	0.42 ± 0.01 e-i	0.83 ± 0.01 h- o	0.84 ± 0.01 g- n	1.42 ± 0.02 h- o	1.45 ± 0.03 g- n	0.46 ± 0.02 a-c	0.27 ± 0.02 h- l	104.17 ± 3.09 i-n	70.97 ± 4.55 o-u
AMF2+C2+B2	7.68 ± 0.06 bc	7.54 ± 0.04 c-e	0.62 ± 0.01 a	0.46 ± 0.01 de	1.07 ± 0.00 a	0.90 ± 0.02 e-k	1.84 ± 0.01 a	1.55 ± 0.03 e-k	0.36 ± 0.03 a-j	0.26 ± 0.01 i-l	132.43 ± 9.97 c-i	61.90 ± 2.82 r-v

Means followed by the same letters are not significantly different at $P < 0.05$ (Tukey's HSD). EC, electrical conductivity; TOC, total organic carbon; OM, organic matter, N, nitrogen concentration; P, phosphorous concentration.



grown under drought stress improved their growth by soluble sugars and protein accumulations compared to non-treated plants. However, our results suggest that the application of compost and inoculation with AMF and PGPR were propitious to carbohydrate accumulation, mainly soluble sugars, in drastic conditions resulting in reduced osmotic potentials in host cells. Indeed, the osmotic stress induced by drought is tolerated by the host plant by altering biochemical responses via the enhancement of metabolite biosynthesis (e.g., sugars and proline) that function as osmolytes, and thereby maintaining the water potential, hydration and turgor level which maintain overall physiological activities under harsh environments. Previous studies showed that sugar metabolism-related genes tend to be enriched in plants treated with beneficial microbes under drought stress (Ahanger et al., 2014; Bárzana et al., 2015). Our results demonstrated that levels of MDA and H_2O_2 in leaves were lower under drought stress in treated plants compared to beneficial microbe-free control plants. To explain the low lipid peroxidation damage in AMF-treated plants, two possibilities were suggested by Abbaspour et al. (2012): (1) either inoculated plants with AMF suffered less drought stress owing to a primary drought avoidance effect by symbiosis (e.g., direct water uptake by fungal hyphae from the soil to the host plant) or (2) AMF colonization improved the activities of antioxidant enzymes as a defense to eliminate the ROS. Our results suggest that the application of compost and inoculation with AMF and PGPR could improve the defense against drought stress by reducing and eliminating ROS diffusion and production. Plants treated with AMF/PGPR counteract water deficit-induced oxidative stress by upregulating ROS-scavenging antioxidant compounds and antioxidant enzymatic activities. It is well known that plants protect against the damage caused by this

oxidative stress by mechanisms that detoxify ROS which can be enzymatic (superoxide dismutase, catalase, ascorbate peroxidase, glutathione reductase, and monodehydroascorbate reductase) and non-enzymatic (flavanones, anthocyanins, carotenoids, and ascorbic acid). Our results showed a significant increase in POX and PPO in plants subjected to water deficit and inoculated with beneficial microbes and/or amended with compost than controls. Duo et al. (2018) revealed that nano-compost alone or combined with bacterial strains minimized the effects of drought stress by increasing antioxidant enzymes and decreasing MDA. Other studies suggested that drought tolerance is acquired by bacteria through the improvement of plant cell membrane stability and elasticity by activating the antioxidant defense system (Dimkpa et al., 2009; Gusain et al., 2015).

The results highlight a physiological and biochemical switching mechanism in microbe association and provide additional confirmation of the hypothesis that, as illustrated in **Figure 9**, microbial association and compost operate at multiple (including photosynthesis machinery, antioxidant system, osmolytes biosynthesis, gene regulation) levels. Our study showed an improvement in the parameters studied in date palms growing in soil treated by the autochthonous biofertilizers mainly the consortium AMF2 alone and its combination with compost (C2) and/ or PGPR (B2) under drought stress, especially AMF2+C2 and AMF2+C2+B2 treatments. This improvement in growth, mineral uptake, and physio-biochemical traits together with the decrease in MDA and H_2O_2 could be due to the synergy between AMF, compost, and PGPR: (1) the compost (C2) with low dose 5% allows good mycorrhizal infectivity, the presence of essential mineral elements such as N, P, and K for plant growth, and improves the soil quality by

mainly owing to the several nutrition-related benefits that this class of soil beneficial microbes, symbionts and compost are able to provide to their host-plants. Furthermore, our data suggest that biofertilizers develop drought-adaptive strategies through the influence of plant mechanisms, such as photosynthetic machinery, better efficiency of PSII, root hydraulic conductivity, osmolyte accumulation, antioxidant enzyme production, higher membrane stability, and lower lipid peroxidation. Our findings are a first step toward encouraging farmers to autonomously produce their AMF inocula, starting from native soils. Further, this work makes biofertilization technology more likely to be affordable for farmers in harsh areas, and also those in developing countries for a sustainable crop growing system. For these reasons, the next significant step (an on-going study) toward the stable use of biofertilizers in agriculture and to better understand the potential effects of indigenous biofertilizers is carrying out field trials.

DATA AVAILABILITY STATEMENT

The datasets generated for this study are available on request to the corresponding author.

AUTHOR CONTRIBUTIONS

MA and MB designed and supervised the research. AM performed the experiments and carried out the analysis. AT, AR, AB, MA-E-M, RB-L, ST, and YA contributed analytic tools. MA, MB, and AM interpreted the data. MA, KO, MH, SS, and TM contributed to the conception and design of the work. AM and MB wrote the manuscript. AM, MB, and AM revised and finalized the manuscript. All authors read and approved the final version of the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2020.516818/full#supplementary-material>

Supplementary Figure 1 | Phenotypic comparison in control date palm plants (non-amended, non-inoculated), and plants amended with composts (C1 or C2) and/or inoculated with arbuscular mycorrhizal fungi (AMF; exogenous AMF1 or native AMF2) or plant growth-promoting rhizobacteria (PGPR) strains (B1 or B2) under two water regimes [75 and 25% field capacity (FC)].

Supplementary Table 1A | Result of multivariate analysis of variance MANOVA test for independent variables including Drought treatment, Composts (C1 and C2) amendment and exotic AMF (AMF1) and Bacteria (B1 and B2) inoculation and interaction among them.

Supplementary Table 1B | Result of multivariate analysis of variance MANOVA test for independent variables including Drought treatment, Composts (C1 and C2) amendment and autochthonous AMF (AMF2) and Bacteria (B1 and B2) inoculation and interaction among them.

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