



Physiological and Multi-Omics Approaches for Explaining Drought Stress Tolerance and Supporting Sustainable Production of Rice

Sajad Majeed Zargar^{1*}, Rakeeb Ahmad Mir², Leonard Barnabas Ebinezer³, Antonio Masi³, Ammarah Hami¹, Madhiya Manzoor¹, Romesh K. Salgotra⁴, Najeebul Rehman Sofi⁵, Roohi Mushtaq⁶, Jai Singh Rohila⁷ and Randeep Rakwal^{8*}

OPEN ACCESS

Edited by:

Ravi Gupta, Kookmin University, South Korea

Reviewed by:

Sebastien Christian Carpentier, Bioversity International, Belgium Palak Chaturvedi, University of Vienna, Austria Tofazzal Islam, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Bangladesh

*Correspondence:

Sajad Majeed Zargar smzargar@skuastkashmir.ac.in Randeep Rakwal plantproteomics@gmail.com

Specialty section:

This article was submitted to Crop and Product Physiology, a section of the journal Frontiers in Plant Science

Received: 28 October 2021 Accepted: 14 December 2021 Published: 27 January 2022

Citation:

Zargar SM, Mir RA, Ebinezer LB, Masi A, Hami A, Manzoor M, Salgotra RK, Sofi NR, Mushtaq R, Rohila JS and Rakwal R (2022) Physiological and Multi-Omics Approaches for Explaining Drought Stress Tolerance and Supporting Sustainable Production of Rice. Front. Plant Sci. 12:803603. doi: 10.3389/fpls.2021.803603 ¹ Proteomics Laboratory, Division of Plant Biotechnology, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Srinagar, India, ² Department of Biotechnology, School of Biosciences and Biotechnology, BGSB University, Rajouri, India, ³ Department of Agronomy, Food, Natural Resources, Animals, and Environment, University of Padova, Padua, Italy, ⁴ School of Biotechnology, Sher-e-Kashmir University of Agricultural Sciences and Technology of Jammu, Jammu, India, ⁵ Division of Plant Breeding and Genetics, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Srinagar, India, ⁶ Department of Biotechnology and Bioinformatics, SP College, Cluster University Srinagar, Srinagar, India, ⁷ Dale Bumpers National Rice Research Center, United States Department of Agriculture (USDA)-Agricultural Research Service (ARS), Stuttgart, AR, United States, ⁸ Faculty of Health and Sport Sciences, University of Tsukuba, Ibaraki, Japan

Drought differs from other natural disasters in several respects, largely because of the complexity of a crop's response to it and also because we have the least understanding of a crop's inductive mechanism for addressing drought tolerance among all abiotic stressors. Overall, the growth and productivity of crops at a global level is now thought to be an issue that is more severe and arises more frequently due to climatic change-induced drought stress. Among the major crops, rice is a frontline staple cereal crop of the developing world and is critical to sustaining populations on a daily basis. Worldwide, studies have reported a reduction in rice productivity over the years as a consequence of drought. Plants are evolutionarily primed to withstand a substantial number of environmental cues by undergoing a wide range of changes at the molecular level, involving gene, protein and metabolite interactions to protect the growing plant. Currently, an in-depth, precise and systemic understanding of fundamental biological and cellular mechanisms activated by crop plants during stress is accomplished by an umbrella of -omics technologies, such as transcriptomics, metabolomics and proteomics. This combination of multi-omics approaches provides a comprehensive understanding of cellular dynamics during drought or other stress conditions in comparison to a single -omics approach. Thus a greater need to utilize information (big-omics data) from various molecular pathways to develop droughtresilient crop varieties for cultivation in ever-changing climatic conditions. This review article is focused on assembling current peer-reviewed published knowledge on the use of multi-omics approaches toward expediting the development of drought-tolerant rice plants for sustainable rice production and realizing global food security.

Keywords: QTL, global food security, multiomics, rice, drought, abiotic stress, proteome, metabolome

1

INTRODUCTION

Rice (Oryza sativa L.) is the topmost grain consumed as a staple food by humans worldwide, most prominently in Asian countries (Asia, 2021; Samal et al., 2021). It belongs to the family Poaceae and tribe Oryzeae and presently comprises 22 wild species with only two species cultivated at a global level. Rice, with a genome size of 430 Mb, is an important model crop plant, and it has been reported that Asia ranks the highest in global rice production (Manavalan et al., 2012; Asia, 2021). As rice requires more water, up to 4000 liters per kilogram production, the plant is more prone to drought stress than other environmental stressors. Among all these stressors, drought stress is a major factor that can hinders the growth, yield and productivity of rice crops (Mumtaz et al., 2020). Drought is a multi-faceted stress and may affect rice production in various ways. In rainfed areas, it affects directly by further drying of the paddy soils, but in irrigated areas it hits in the form of declining underground water tables and become an issue for water sustainability. In the field, drought effects can be compounded by interactions with other biotic and abiotic stresses. Further, conventional cultural practices in rice require a season-long flooding, but in today's circumstances, it is unsustainable, and there is competition for the utilization of surface water (rivers, etc.) with increased urbanization; additionally, low underground water availability due to the continually declining water tables of aquifers is also becoming common. All these combinations have increased a strong need for understanding and increasing drought tolerance in rice crop.

The negative implications of drought stress on the physiological functioning of plants are mainly due to its reduced water potential and turgor pressure that suppress plant growth and metabolism (Lisar et al., 2012). Drought is a period in crop's growing season during which the soil moisture reaches to a certain level that reduces its yield or quality resulting from either limited irrigations or a below average rate of precipitation and higher evapo-transpirations, which causes a decline in plant growth and productivity (Rollins et al., 2013; McClung et al., 2020). Drought affects plants in numerous ways by impairing normal molecular, metabolic and physiological networks to reduce growth and metabolism (Zu et al., 2017). A typical implication of drought includes a decline in the expansion of leaves and an overall decrease in the stomatal conductance and rate of photosynthesis (Avramova et al., 2016; Barnaby et al., 2019). Furthermore, a reduction in growth is also reported to be induced by the inhibition of cell elongation, cell expansion and impairment in mitosis (Potopova et al., 2016). Against these consequences, plants respond at the morphological, physiological and molecular levels. These responses include maximum uptake of water by a dense and deep root system, reduction in water loss by closure of stomata, adjustments in osmosis, reducing the leaf area, altering the elasticity of cell wall and several more physiological adjustments (Saha et al., 2016). Physiological traits are critical to selecting drought-tolerant germplasm of crop plants, even though the response to drought stress by crop plants is dependent on the environment and diverges across genotypes and genetic interactions (Lonbani and Arzani, 2011).

At present, freshwater shortage is major issue on global scale and could be more critical in the near future as per climate fluctuation projections. In developing sustainable solutions to overcome water scarcity, there is a constant need for scouting strategies that could be possible alternatives to ensure the availability and accessibility of freshwater to crop plants (Noemi et al., 2014). Rice, among other cereal crops, demands access to freshwater, i.e., water-intensive cultivation. Moreover, half of humankind relies on rice as a staple food, and at the global level, a total of 160 million hectares (Mha) of cultivated land comprises this crop, most of which (around 40%) lies in Asian countries (Prasad et al., 2017). The rice plant is cultivated in flooded paddy fields since it requires an adequate water supply of almost 2 to 3 times more than dryland cereals.

Consequently, researchers are now faced with a challenge to increase the adaptation of rice cultivars to water-scarce conditions and to reduce the dependency on a large quantity of water to support their cultivation in almost dry land cultivation systems (Sandhu et al., 2014). Major adaptations include adjustments in physiological, anatomical and morphological characteristics of root and shoot traits (Kadam et al., 2015; Sandhu et al., 2016) (Tables 1, 2). An irrigated rice system is the most commonly cultivated practice, comprising up to 55% of the 158 Mha cultivated land area, whereas 34% of the 54 Mha land cover is shared by rainfed lowland systems, and rice grown in flood-prone areas shares 7% of the 11 Mha land area (Bouman et al., 2007). Approximately 37% of global rice production is from South Asia, among which 50% is rainfed (Dawe et al., 2010). Moreover, rainfed rice is also cultivated in sub-Saharan Africa, accounting for up to 84% of the total area of rice cultivation (Gauchan and Pandey, 2012). Since the global production of rice is grossly dependent on rainfed ecosystems, drought stress becomes a major factor behind decreased productivity from 13 to 35%. Thus, one can witness an enormous global loss in rice productivity compared to other crops due to its higher dependency on water and hence proneness to drought stress. All the evidence suggests that climate induced fluctuating rainfall and drought result in heavy losses to rice farming systems (Fahad et al., 2019).

A sustainable approach for rice production is to minimize irrigation, which is being realized in commercial production by the introduction of alternate wetting and drying (AWD) irrigation management (de Avila et al., 2015; Chen M.-H. et al., 2021; see also¹). Currently, farmers are practicing safe-AWD levels because most current varieties are bred for seasonlong flood irrigation management under both lowland and upland rice cultivations. In AWD, there is a drying phase, and rice is susceptible to drying conditions, because they affect yield and grain quality (McClung et al., 2020). Shaibu et al. (2015) compared Nunkile and NERICA 4 rice varieties, which are adapted to upland and lowland irrigated conditions, under continuous flood and three different AWD schemes. The results showed that water productivity for continuous flood treatment was superior to all three AWD schemes. Hence,

 $^{^1}http://www.knowledgebank.irri.org/training/fact-sheets/water-management/saving-water-alternate-wetting-drying-awd$

TABLE 1 | Drought tolerance genes that have been tested on rice.

Sr. No	Cellular mechanism	Gene	Promoter	Genetic transformation method	Targeted phenotype	References
1	Abscisic acid metabolism	CAMV35SP	DSM2	Agrobacterium	Oxidative and drought stress tolerance	Du et al., 2010
2	ROS scavenging	OSSROIC	Ubi 1	Agrobacterium	Oxidative stress tolerance and stomata closure regulation	You et al., 2013
3	Protoporphyrinogen oxidase	PPO	_	Agrobacterium	Less oxidative damage and drought tolerance	Phung et al., 2011
4	Ubiquitin ligase	OSSDIR1	CAMV35 Ubi1	Agrobacterium	Stomata regulation under drought stress	Gao et al., 2011
5	Abscisic acid sensitivity	OSSAPK2	_	-	Abscisic acid sensitivity and drought tolerance	Lou et al., 2017
6	DNA damage repair and defense response	OsNAC14	OsRAD51A1		DNA damage repair and defense response resulting in improved tolerance to drought	Shim et al., 2018
7	Multiple stress tolerances in rice plants during both seedling and panicle development stages	OsAHL1	P _{AHL1}	-	Regulates root development under drought condition to enhance drought avoidance	Zhou et al., 2016
8	Enhanced resistance to a bacterial pathogen	OsWRKY11	CHIT 2	-	Enhanced resistance to a bacterial pathogen	Lee et al., 2018
9	Reactive oxygen species scavenging	OsLG3	OsLG3-OE	_	Reactive oxygen species scavenging and drought tolerance	Xiong et al., 2018
10	Increased moderate susceptibility to the pathogens	OsMADS26	ubiquitin1	_	Increased moderate susceptibility to the pathogens and drought tolerance	Khong et al., 2015
11	Induced a variety of environmental stresses and plant hormones	OsDRAP1	CaMV35S	Agrobacterium	High expression in response to drought	Huang et al., 2018
12	Increases ABA sensitivity and enhances osmotic tolerance in rice	OsEm1	LEA	-	ABA sensitivity and enhances osmotic tolerance promising for engineering drought tolerance in rice	Yu et al., 2016
13	Reactive oxygen species (ROS)-scavenging	OsCML4	CaMV35S	-	Reactive oxygen species (ROS)-scavenging and drought tolerance	Yin et al., 2015
14	Control of tiller outgrowth	OsIAA6	PGD1	Agrobacterium	Drought stress responses and the control of tiller outgrowth.	Jung et al., 2015
15	Conversion of aspartate amino acid to glutamate was found to be associated with drought tolerance	OsDREB1F	CaMV35S		The categorization of all the significant SNPs with H5 drought tolerant haplogroup supports their role in drought tolerance in rice	Singh et al., 2015

for significant water savings, farmers need improved varieties that can withstand drier soils without decreasing yield and grain quality. Thus, to develop improved rice cultivars that can produce profitable yields with significant water savings, researchers need a better understanding of drought tolerance mechanisms and of the tools available to them. Furthermore, the situation is compounded by the use of susceptible rice varieties by farmers in subtropical and tropical cultivated areas (Zu et al., 2017). Thus, it is important to develop new rice cultivars that possess traits such as higher yield and resilience to drought stress. The evolution and production of drought tolerant rice varieties are developed by understanding the molecular mechanisms and signal responses initiated by tolerant cultivars under water deficit conditions. This approach does not mean that the traditional germplasm or breeding line selection process does not play an important role in this process of developing new rice varieties, some of which could have desirable traits such as short duration with higher yield (Mehana et al., 2021). However, the underlying molecular mechanism must be understood at a holistic level, and therein omics tools are needed and are prominent.

The past few decades have witnessed extensive research based on application of omics technologies paving way to identify a large number of candidate genes, proteins and pathways to generate drought tolerant varieties of rice plants (Panda et al., 2021; Figure 1). Omics-based high throughput techniques facilitated unbiased studies on the genome, epigenome (epi)transcriptome, metabolome and proteome (Oliver et al., 1998; Hasin et al., 2017; Yu et al., 2021). Furthermore, technologies are employed to comprehend the underlying molecular mechanisms to resolve complex cellular responses and effects on the phenotypes of the crop (Manzoni et al., 2018; Upadhyaya and Panda, 2019). Considerable molecular data has been created by advancements in genomic and transcriptomic techniques but researchers are still behind in correlating the data with the proteomes due to limited depth of quantitative proteomic data inputs with respect to post-translational modifications especially (Molina et al., 2011; Ghatak et al., 2017). To date, gene expression analyses through applications of high-throughput technologies have revealed the transcription levels in cells. In addition, several investigations were conducted to correlate the transcriptomic

and proteomic information to highlight the critical impact of post-transcriptional modifications on cellular subtleties (Foss et al., 2011; Ghazalpour et al., 2011). Recent advancements in drought stress-related data support the role of omics technologies in understanding and determining the mechanistic make-up of plants, to develop crop varieties that are resilient to drought stress. Most of these studies have demonstrated the critical role of post-transcriptional and post-translational modifications of proteins in defensive mechanisms that allow the plant to adapt to a diverse range of abiotic stressors (Matsuura et al., 2010; Sormani et al., 2011; Juntawong and Bailey-Serres, 2012; Liu et al., 2012).

The quantification of biomolecules, such as genes and proteins, is now easier due to the advent of high-throughput technologies, for studying the transcriptomes and proteomes. For instance, the E3-Ubiquitin Ligases proteins abundantly expressed in rice help in modulation of abiotic stressors such as drought, salinity, nutrient deprivation and radiation (Melo et al., 2021). Drought tolerance genes and their cellular processes, such as signaling-controlled gene expression and cellular modifications, have been deeply investigated by multi-omics high-throughput technologies (Huynh et al., 2018; Yadav et al., 2019; Singh et al., 2021). Keeping in perspective the importance of a multiomics approach to better understand abiotic stress tolerance mechanisms in crops and developing novel stress mitigation strategies, the present article comprehensively summarizes the state-of -the-art knowledge on the molecular aspects of drought stress. Moreover, the current review also presents insights gleaned thus far on drought stress using various omics approaches with a special focus on rice, given its global significance as a major food crop.

APPROACHES IN USE FOR CONFERRING DROUGHT TOLERANCE IN RICE

The improvement of rice varieties could be accomplished by mining genes and superior alleles capable of better signal perception, signal transduction, and functional roles in drought tolerance. The repository of resilient genes and traditional donors of these genes includes wild accessions, landraces and varieties, such as Aus 276, Birsa gora, Dhagaddeshi, Dular, Kali Aus, Nagina 22 (or N22), and Vandana. Considering genetic improvement, several varieties have been developed through the selection of resilient landraces for a specific trait. For example, the landrace "Rajbhog" found in the foothills of Nepal was used to develop Nagina 22 through selection processes and is also a well-known drought tolerant variety of rice (Vikram et al., 2011). Furthermore, a drought tolerant Kataush landrace from the Nepal Tarai region was characterized (Puri et al., 2010). Landraces also resulted in the development of Laloo-14, a drought tolerant Indian rice variety cultivated in rainfed areas of Madhya Pradesh state of India, which validates the immense potential of landraces for improving rice cultivars for cultivation in rainfed areas (Akshaya et al., 2017). The rich repository of resilient genes found in landraces makes them suitable candidates

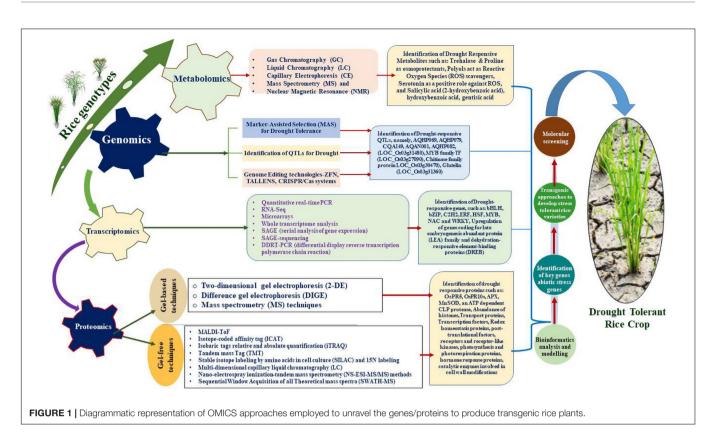
TABLE 2 | QTLs identified for drought tolerance related traits in rice.

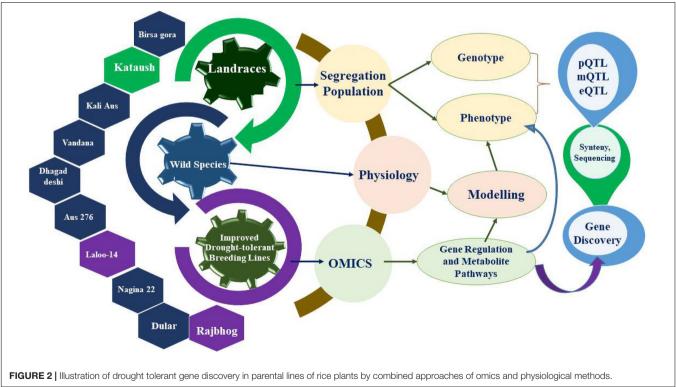
S. No	Targeted trait	Number of QTL'S	References
1	Grain yield	1 (Qdty2.1)	Mishra et al., 2013
2	Grain yield	1 (Qdty3.2)	Yadaw et al., 2013
3	Grain yield	14	Wang et al., 2014
4	Filled grain number per panicle	23	Wang et al., 2014
5	Panicle number per plant	14	Wang et al., 2014
6	Grain yield	1 (Qdty2.3)	Palanog et al., 2014
7	Grain yield	1 (Qdty2.2)	Palanog et al., 2014
8	Grain yield	4	Saikumar et al., 2014
9	Grain yield	7	Singh et al., 2015
10	Grain yield	1 (qDTY 12.1)	Swamy et al., 2018
11	Flowering time	1	Prince et al., 2015
12	Flowering time	5	Saikumar et al., 2014
13	Flowering time	1	Sandhu et al., 2014
14	Canopy temperature	6	Prince et al., 2015
15	Biomass	8	Prince et al., 2015
16	Biomass	4	Saikumar et al., 2014
17	Drought index	3	Prince et al., 2015
18	Grain weight	2	Zhou et al., 2013
19	Grain yield	24	Verma et al., 2014
20	Seed setting rate	6	Prince et al., 2015

to look for adaptation related genes or superior alleles important for resiliency for production in limited irrigation management. Various drought tolerant Indian landraces have been reported to possess drought tolerant genes (Vanniarajan et al., 2012). For further improvement of rice crop, it is pertinent to also identify and characterize drought tolerant genes by modern molecular breeding technologies.

Apart from the landrace varieties and wild germplasm accessions, natural hybrids and genetic stocks also act as rich genetic sources for the improvement of rice cultivars (Biswas et al., 2020). This wide range of rice landraces can be used to identify genes by diverse approaches, including omics-based techniques (**Figure 2**), for their subsequent transfer into elite breeding lines.

Only a few traits that pertain to roles in water use efficiency (WUE) and tolerance to drought have been implemented in the field, even though numerous crop species have been screened by mining drought tolerant genes (Langridge and Reynolds, 2015). A wide range of techniques, such as omics tools, molecular breeding and precise phenotyping, augment the identification of candidate genes that pertain to metabolic and signaling pathways and that are pivotal for drought tolerance in crops. Besides, the workflows for initiating the in-depth cellular phenotyping of diverse crop plants through applications of multi-omics technologies will further augment the quest of unraveling the resilience mechanisms (Zivy et al., 2015). In addition, various studies, such as functional genomics and expression profiling of genes, are available to comprehend the complexity of drought tolerance mechanisms in crop plants (Delphine et al., 2010; Li et al., 2018). Differential gene expression analysis has also been used to identify drought responsive genes in crop plants (Guo et al., 2008; Gray et al., 2010).





Even though realistic progress through omics technologies such as metabolomics, proteomics, and genomic methods compared with epigenomics methodologies is scant, these approaches have provided much needed and useful information pertaining to genetic and physiological attributes linked to drought tolerance. We aim to combine efforts in the application of omics approaches

in conjunction with plant breeding methods to extend the development of resilient rice cultivars aimed at cultivation in rainfed ecological niches or save significant amounts of water under irrigated field managements.

Plant Traits

Physiological adjustments established by crop plants against drought stress enable them to regulate their water requirements for normal physiology, growth and metabolism (Figure 3). The following sub-sections detail the important physiological adjustments accomplished by crop plants against drought stress:

Shoot Traits

Reductions in growth and metabolism are primarily induced by drought in rice plants. Moreover, the initiation of new aerial organs, such as the stems, leaves and pre-existing organs, in aerial parts of plants are considerably affected by drought stress (Chaves et al., 2002). Central to the reduced growth of these aerial parts is the decline in cell division in plants and modulation of the physio-chemical properties of cell walls due to an increase in cell wall rigidity, to reduce expansion and hence the growth (Granier et al., 2000; Cosgrove, 2005). A significant decline in the leaf expansion rate was observed when water stress of 80% transpirable water soil moisture for 20 days was observed in the vegetative phase in rice (Farooq et al., 2009). Thus, adaptations result in limited loss of water by transpiration through leaf surfaces. A reduction in the rate of photosynthesis in rice prevails based on the change in leaf color and conductance of mesophyll layer to CO2 during the drought stress period (Lauteri et al., 2014). The change in green color is due to the relative chlorophyll content in rice leaves and can be measured by SPAD meter under drought conditions in comparison to the irrigated conditions (Ndjiondjop et al., 2010; Barnaby et al., 2019). Zinolabedin et al. (2008) reported contradictory observations regarding water and nutrients uptake by the plant root system, which in turn caused a reduction in the chlorophyll concentration and subsequent yellowing of the leaves. Under drought stress, plants roll their leaves to reduce the transpiration rate, to maintain water levels (Sié et al., 2008). Leaf rolling capacity is now considered to be one of the major physiological adaptation strategies by crop plants to maintain their water status during drought stress periods (Fukai and Cooper, 2002). Moreover, a direct correlation was observed between grain yield and leaf rolling attributes of crop plants. Henderson et al. (1995) reported that leaf tip drying acts as a good indicator of the level of drought stress. The rolling of leaves is a reversible physiological adaptation, even though leaf tip rolling induced by drought stress is irreversible. Other abiotic stress factors, such as freezing and salinity, are also linked to a decline in the availability of water to plants. The decline in water availability is quantified as a decrease in the water potential. The indicator of dehydration avoidance is recognized by the leaf water potential (LWP) (Pantuwan et al., 2002). The stomata are closed when the water level goes beyond the water deficit in leaves to lower the rate of transpiration by minimizing the damage caused by water loss. Moreover, the higher LWP is regulated by varietal differences in the stomatal response and the closure of stomata (Jongdee et al., 1998). Higher

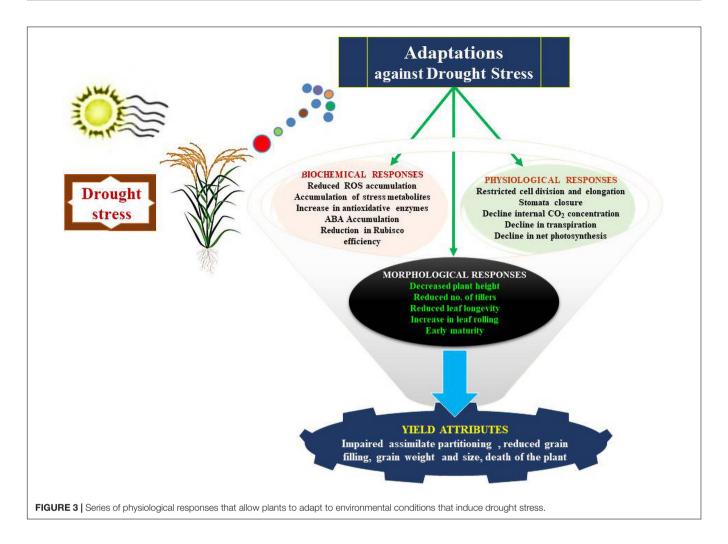
photosynthetic capability possessed by certain genotypes helps plants to protect from the onset of senescence under stress conditions. This so-called stay green trait possessed by plants also helps in assimilating nitrogen and helps in optimizing the photosynthetic capability of plants under drought stress (Borrell et al., 2001). Other physiological adaptations possessed by plants include modifications in water uptake, loss of water potential, enhancing accumulation of compatible solutes, overexpression of tolerant proteins, prevention of cell damage and enhancing repairing by cell division.

Root Traits

Imperative to the growth and metabolism of plants is the absorption of water and nutrients, which is grossly dependent on modifications in the morphological and physiological characteristics of roots (Ghosh and Xu, 2014). Various characteristics such as, type of root system, its properties, structures and wide distribution of roots helps to uptake water from soil to maintain productivity under water stress conditions (Comas et al., 2013). Under drought stress, several root traits such as, long specific root length; root length density and small fine root diameters maintain the productivity and regulate water levels in rice crops (Comas et al., 2013). For example, under drought stress periods, it was reported that rice tolerant varieties such as, Chuanguyou208 and Deyou4727 showed larger length in roots, higher root number and weight under 30-50 cm depth soil layer and in addition these varieties also displayed higher activities of peroxidase (POD) and superoxide dismutase (SOD) (Wang X. et al., 2019). Moreover, the hydraulic properties of roots in rice under drought stress were enhanced by developing deep and thick root systems (Lipiec et al., 2013). These attributes helped rice plants to develop tolerance to drought through absorption of water from deep layers of soil (Gowda et al., 2011; Uga et al., 2013a). Moreover, supplementation of rice seedlings with NH₄₊ resulted in higher root growth and an increased number of root tips to enhance water uptake under water stress conditions (Jia et al., 2020). In addition, rice seedlings supplemented with NO³⁻ induced restriction in water uptake resulted in the induction of root aerenchyma formation (Yang et al., 2012). Later studies revealed that NH4⁺ helped in higher water uptake compared with NO³ due to enhanced root growth.

Inflorescence Traits

Rice plant is susceptible to water stress during reproductive stage, and the severity of yield and quality losses depend on magnitude and duration of the stress (Yang et al., 2019; Chen M.-H. et al., 2021). Shaibu et al. (2015) compared continuous flood with two AWD schemes focused on two different flowering stages: AWD up to the start of flowering (AWD1), and AWD up to the start of grain filling (AWD2). The results showed that both AWD schemes had significant yield loss compared with the continuous flood treatment. The results emphasize the need for robust inflorescence traits for drought tolerance. Under drought conditions, it is interesting to note that generally early morning period is cooler compared to late morning and afternoons. Hirabayashi et al. (2015) identified a novel QTL, qEMF3, for early-morning flowering trait from wild rice, *Oryza*



officinalis. The qEMF3 shifted flower opening time by 2 h earlier and improved spikelet fertility under stress environment. Similarly, studying pistil's role is another interesting inflorescence trait that has not been explored in detail to unravel molecular basis of stress tolerance (Wang et al., 2021). Rice is one of the best models to study the development of inflorescence through molecular analysis of inflorescence mutants for investigating architecture of inflorescence (Zhang and Yuan, 2014). The critical factors to decipher the complexity of rice traits include the total number of fertile tillers, and the development and architecture of the inflorescence (Ikeda et al., 2004). The rachis, primary and secondary branches, including the spikelet, are critical physiological features of rice inflorescences. The spikelet consists of one fertile floret, two sterile florets and two rudimentary glumes. Spikelet differentiation, the determining factor of seeds per panicle, depends on the establishment, transition and activity of axillary and apical meristems. The development of rice inflorescences is well explained by the ABCDE model for the specification of floral organs in addition to the role of genes such as OsMADS3, OsMADS14, OsMADS15, and OsMADS58 (Yoshida and Nagato, 2011). The inflorescence in rice is controlled by cytokinin and auxin signaling pathways, CLV-WUS signaling and expression of MADS box genes. In rice,

the Drought and Salt Tolerance (DST) protein, a member of zincfinger transcription factor, induces overexpression of OsCKX2 in the reproductive meristem for induction of inflorescence (Li et al., 2013). The depth in understanding the molecular mechanism underlying the regulatory responses against drought and that influence the inflorescence development will further reveal deep comprehensions regarding enhancing yield and thus food security in the changing climatic scenario.

OMICS APPROACHES FOR UNDERSTANDING DROUGHT TOLERANCE

The methodologies studied under the omics approaches are categorized as genomics, transcriptomics, proteomics, metabolomics, epigenomics and comparative genomics.

Genomic Approaches

Genomic studies are technical approaches for investigating the gene structure and functional dynamics of coding and noncoding sequences applicable to augmenting crop improvement. Important resources in structural and genomics studies include

mutant libraries, cDNAs, expression profiles, sequence data sets and quantitative trait loci (QTLs) (Jiang et al., 2011). For instance, Vikram et al. (2012) cloned several genes and identified QTLs that were important for drought tolerance in rice. Large numbers of QTLs were identified to resolve the complex nature of drought stress and alleviating mechanism for tolerating drought stress (Fleury et al., 2010). In addition, several QTLs are identified to be responsible for enhanced grain yield and for other secondary characteristics that allow withstanding drought stress in rainfed uplands and lowlands (Bernier et al., 2007; Venuprasad et al., 2009). Differential performance of QTLs has been observed in upland and lowland ecosystems under drought conditions. Finally, the best suitable QTLs are selected as per the ecosystems, genetic background and environmental conditions. Moreover, for developing high yielding varieties, marker assisted back crossing (MABC) was efficaciously applied experimentally in crop plants. For example, MABC has been employed to increase the productivity of KDML105 rice variety in north-eastern regions of Thailand (Kanjoo et al., 2012). It is expected in near future that applications of molecular breeding techniques such as, marker-assisted selection (MAS), SNP marker applications and genome wide assisted selection (GWAS) will pave way to understanding the molecular mechanisms underlying the resilience in crop plants against wide range of environmental challenges such as drought stress.

Comparative Genomics

The evolutionary history of organisms is clearly revealed by carrying out comparative account on structure and functions of genes across the species. For example, the phylogenetic relationships of Poaceae family (members of grasses) are now well understood by investigated species-specific DNA markers. Further, the comparative studies of genomes have led to identify the syntenic regions of different species. For instance, it is found that two thirds of genes located on chromosome number 11 in wheat are distributed in 06 homologues of grass genomes (Singh et al., 2004). The QTL1.1 is classical example of syntenic relationship; this QTL is found in chromosome 3 in maize, chromosome number 4B in wheat and chromosome 6 in barely (Swamy et al., 2011). The rice drought grain yield QTL was shown to be homologues of with maize counterparts (Swamy et al., 2011). Comparative analyses showed the role of OsCPK9 and AtCPK10 genes in augmenting the ABA dependent drought responsive mechanism (Zou et al., 2010; Wei et al., 2014). Whereas, OsCPK10 played pivotal role in defending cellular membranes against ROS and drought stress (Kumar et al., 2014; Nakabayashi et al., 2014; Fang et al., 2015; Yin et al., 2015). Large numbers of stress responsive genes are reported to play critical role in development and growth of plants. In addition, a network of genes pertaining to seed development and stress response has been reported in rice (Cooper et al., 2003). Molecular chaperons have been found to play critical role in circumventing the drought response in crop plants. Upon 2DE-MS and GC/MS based proteomic and metabolomic techniques, it was reported that Bip (molecular chaperon binding protein) was overexpressed in transgenic soybean plant (Coutinho et al., 2021). Moreover, transgenic rice over expressing OsSKIPa gene

performed well under drought conditions due to its critical role in expression of series of stress related genes including, PP2C, CBF2, RD22 and SNAC1 and also enhancing the ROS scavenging ability in rice plants (Hou et al., 2009). Furthermore, *OsbZIP46CA1* transcription factor gene, a modified product of OsbZIP46 was found to be overexpressed in transgenic rice under drought stress, was found to enhance the expression of stress related genes, most of which in turn downregulate the ABF/AREBs (Ning et al., 2012). The transgenic approaches thus lead to unraveling the repository of genes critical for regulating plant metabolism under drought and other stress conditions, thus paving a way toward global food security (Details regarding genes conferring drought tolerance in rice have been presented in **Table 1**).

STRESS-INDUCED REGULATORY GENES AND QUANTITATIVE TRAIT LOCI FOR DROUGHT TOLERANCE

Against the stress induction, crop plants initiate a coordinated series of cellular and signaling processes. A great deal of physiological and morphological changes is actually the outcome of cellular responses regulated by large number of genes. It is reported that in rice, about 6,000 genes are downregulated and 5000 genes are upregulated under drought stress conditions (Maruyama et al., 2014). In addition, genome wide expression analysis led to identification of about 5,284 differentially expressed genes under drought conditions (Wang et al., 2011). For instance, the CO-like gene, *Ghd2* (grain number, plant height, and heading date2) is attributed to enhance the yield potential in rice. The Ghd2 is almost prototypical to Ghd7, which helps in leaf senescence and drought tolerance. Contrary to this, it is reported that Ghd2 overexpression considerably decreased the drought tolerance and once knocked-out it resulted in opposite effects (Liu et al., 2016). Another gene OsbZIP42 encoding a transcription factor is positive regulator of ABA dependent signaling pathway, hence critical for drought tolerance was found activated by stress or SAPK4 (ABA-activated protein kinase 4) (Joo et al., 2019). The OsbZIP42 genes, a member of EbZIP are found to be critical for established tolerance against the abiotic stress. In addition, large number of signaling cascades have been reported to be activated by ABA to activate adaptive response against abiotic stress conditions (Jiang and Zhang, 2002; Salazar et al., 2015; Sah et al., 2016).

In rice, SNAC1 and DST transcription factors (TFs) are activated by ABA mediated signaling to regulate stomatal movement (Hu et al., 2006; Huang et al., 2009). Furthermore, SNAC1 (stress responsive NAC1), a TF, is a member of the NAC family of TFs, which includes ATAF, CUC, and NAM, and approximately 149 members have been identified in rice plants. SNAC1 in particular is abundantly expressed in guard cells under drought stress conditions. Apart from ABA mediated activation of drought tolerant pathways, DREB (drought responsive element binding) is activated to act as frontline TFs to withstand drought stress (Yamaguchi-Shinozaki and Shinozaki, 2005). Moreover, the TF C₂-H₂-type zinc fingercontaining protein helps in drought and salt tolerance and

mediates H_2O_2 induced pathways to mediate stomatal closure (Huang et al., 2009).

The ABA phytohormone is activated by drought stress, which in turn causes stomatal closure and expression of diverse genes. The concentration of ABA in guard cells directly influences the closure of stomata, and its concentration is increased during drought stress. The pyrabactin tolerance (PYR)/PYL (PYR1like)/regulatory receptors that belong to ABA response receptors sense ABA to initiate the downstream signaling cascade (Ma et al., 2009; Park et al., 2009). Upon ABA binding, its receptor undergoes conformational changes, which enable binding and inactivation of PP2Cs (protein phosphatase 2C), a negative regulator, including ABA insensitive 2 (ABI2), ABA insensitive 1 (ABI1), PP2CA and homology to ABI1 (HAB1) (Geiger et al., 2009). Since the negative regulator of the signaling pathway PP2C is inhibited, downstream signaling molecules are released their by switching the signaling cascade. The Ser/Thr kinase OST1 (open stomata1/SnRK2.6/SRK2E) is the main target of PP2C and is an important junction for the regulation of transcriptional responses, including ABA and CO2 responses. Upon release of OST1, it activates several TFs and phosphorylation of several plasma membrane proteins, which results in the closure of stomata through promoters such as AtRBOHF and SLAC1 (Fuji et al., 2009; Sirichandra et al., 2009; Klingler et al., 2010). In rice, protein complexes such as RCAR5/OsPYL, OsPP2C30, OREB1, and SAPK2 act as signaling units regulated by ABA (Kim et al., 2012). Among these proteins, OsPYL/RCAR5 were reported to be positive regulators of abiotic stress and are attributed to enhanced drought tolerance when overexpressed in transgenic rice plants. In rice, both reverse genetics and forward genetics have led to the identification of hundreds of stress responsive genes and respective QTLs (Wang et al., 2016).

A large number of kinases, such as OsMAPK5, which are functionally characterized as stress responsive MAPK (mitogenactivated protein kinase), have been found to be involved in regulating a diverse range of abiotic stresses, such as drought, cold and salt stress, even though they have been found to negatively regulate biotic stress, including bacterial and fungal infections (Xiong and Yang, 2003; Rohila and Yang, 2007). It has been reported that overexpression of TFs such as, SKIP, bZIP, and NAC helps to enhance drought tolerance in rice. In particular, NAC TFs, such as OsNAC5, OsNAC6, OsNAC9, OsNAC10, OsbZIP16, OsbZIP23, OsbZIP46, and OsbZIP71 are involved in enhancing drought tolerance in crop plants (Fukao and Xiong, 2013). In addition, WRKY TFs are also involved in drought tolerance and a large number of growth and developmental processes. For example, 97 WRKY genes identified in O. nivara (OnWRKY) and 89 WRKY genes in Japonica were mapped and identified in plants that have diverse range of functions (Sahebi et al., 2018).

A large number of QTLs have been identified in different populations linked to drought tolerance (Khowaja and Price, 2008). Large effect QTLs have also been identified for grain yield under drought conditions (Bernier et al., 2007). Approximately, 77 QTLs for grain yield and other aspects important for drought tolerance were identified by crossing two rice cultivars (Lanceras et al., 2004). These QTLs were identified to possess several physiological attributes, such as 05 QTLs for days to flowering, 6 QTLs for harvest index, 7 QTLs for grain yield, 7 QTLs for

percent spikelet sterility, 8 QTLs for biological yield, 10 QTLs for total spikelet number, 11 QTLs for plant height and 23 QTLs for panicle number. Approximately 10 QTL components and high grain yield were identified by using a recombinant inbred population of rice, such as IR64 and Cabacu, under drought stress (Trijatmiko et al., 2014). The QTL, qDTY12.1 was the first report linked to grain yield in the upland reproductive stage under drought conditions. This QTL was identified from the 436 F3 populations derived from Vandana and Way Raren (Bernier et al., 2007). The qDTY2.1 and qDTY3.1 identified in the backcross between Swarna and Apo are two other large effect QTLs known to affect grain yield under drought conditions at the lowland reproductive stage. Both QTLs showed pleiotropic effects on a few traits, such as PHT (plant height) and DTF (days to flowering) (Venuprasad et al., 2009). Moreover, Venuprasad et al. (2012a) reported qDTY6.1 QTL has had a strong effect on aerobic drought stress. Grain yield under severe drought stress in lowland reproductive stage QTLs was identified in F3 populations derived from the crosses of Swarna × N22, IR64 × N22 and MTU1010 × N22 (Vikram et al., 2011). Later, QTLs were also identified in the Apo/IR64 and CT9993-5-10-1-M/IR62266-42-6-2 populations (Kumar et al., 2008; Venuprasad et al., 2012b). Other QTLs reported include *qDTY2.2*, *qDTY4.1*, *qDTY9.1*, and qDTY10.1, which were linked to grain yield and identified in populations obtained from backcrosses between IR64 and Aday Sel rice varieties (Swamy et al., 2013) (Details related to QTLs conferring drought tolerance in rice have been presented in Table 2).

ROOT ARCHITECTURE FOR DROUGHT TOLERANCE

Water deficit is overcome by a series of genes responsible for root architecture and hence higher yields are observed in crop plants (Jeong et al., 2010; Iwata et al., 2013; Cheng et al., 2015). Specifically, the NAC family of TFs was found to be overexpressed, to regulate root architectures (Zheng et al., 2009). For instance, OsNAC9 is a member of the NAC TF that is responsible for root architecture under drought tolerance and grain yield (Redillas et al., 2012). Later, this gene was reported to be overexpressed when controlled by a constitutive or root specific promoter in transgenic rice under both drought and wellwatered conditions. It was reported that recombinant lines of maize have 144% more yield than control plants under drought conditions (Zhan et al., 2015). In rice, the overexpression of the OsDHODH1 gene, encoding a putative cytosolic dihydroorotate dehydrogenase (DHODH), enhanced drought and salt tolerance (Liu et al., 2009).

Enormous putative genes controlling physio-morphological traits confer drought tolerance in rice (Deivanai et al., 2010). The appropriate water and nutrient status of plants is largely supported by the root architecture (Kato et al., 2006). The rooting depth, root thickness, root density and distribution pattern of roots are various traits studied under root system architecture to regulate water and nutrient uptake to address adverse conditions of drought (Lilley and Fukai, 1994; Fukai and Cooper, 1995; Pantuwan et al., 1996; Wade et al., 1996). Furthermore, in

rice populations, various QTLs related to morphology and root index penetration have been identified (Champoux et al., 1995; Ray et al., 1996; Zhang et al., 2001; Henry et al., 2014; Kijoji et al., 2014). Important traits adapted by crop pants to retain water and nutrients during drought stress include the ratio of deep rooting (RDR), root growth angle (RGA) and direction of root elongation. In rice plants, overexpression of EDT1/HDG11 genes resulted in enhanced root development and reduced stomatal density under drought conditions (Yu et al., 2013). For instance, squalene synthase (SQS), a key enzyme located in the endoplasmic reticulum is capable of catalyzing first reaction, i.e., conversion of two farnesyl pyrophosphates into squalene, a step in the isoprenoid metabolic pathway finally directed to synthesize sterols in plants (Tansley and Shechter, 2001). Under drought conditions, it was reported that the disruption of squalene synthase (SQS) function by RNAi led to enhanced root length, a higher number of lateral roots and a decline in the stomatal conductance (Manavalan et al., 2012).

Approximately 675 QTLs for root traits were identified by meta-analysis of 12 populations of crop plants (Courtois et al., 2009). For deep rooting traits, only 05 major QTLs were mapped as root traits (Kitomi et al., 2015; Uga et al., 2011, 2015). A cross between the deep rooting cultivar Kinandang Patong and the shallow-rooting cultivar IR64 resulted in recombinant inbred lines (IK-RILs) leading to the identification of the DRO1 gene located on chromosome 9 (Uga et al., 2011). It was reported that DRO1 considerably encodes the RGA and grain yield. The DRO2 gene found on chromosome 4 is another major QTL that encodes the RGA and was obtained by crossing Kinandang Patong and shallow-rooting cultivars (ARC5955, Pinulupot1 and Tupa729) (Uga et al., 2013b). Furthermore, the DRO3 gene found on chromosome 7 also had a great impact on the RGA due to its involvement in the DRO1 genetic pathway (Uga et al., 2015). Genome wide expression profiling also led to the identification of OsAHL1, a novel gene involved in drought tolerance and avoidance in rice plants. Later, genes were found to regulate root development, oxidative stress and chlorophyll content under drought stress conditions (Zhou et al., 2016). Norton and Price (2009) identified 04 QTLs for seminal root morphology and 2 QTLs for root gravitropic responses. By using 124 recombinant inbred lines, QTLs such as Soil Surface Rooting 1 (qSOR1) were found on chromosome 7 in rice (Uga et al., 2012). In addition, 2 QTLs for canopy temperature and 6 for leaf water potential in RILs were identified in crop plants (Liu et al., 2005). Lou et al. (2015) identified 6 QTLs by using 1,019,883 SNPs. In drought stress, 1 QTL for leaf drying, 1 for SPAD and 2 QTLs for canopy temperature were identified to manage stress (Prince et al., 2015). It is concluded that the introduction of traits contributes to drought tolerance and avoidance in rice to enhance yield (Fukai and Cooper, 1995; Nguyen et al., 1997). Moreover, a considerable number of QTLs have been mapped for osmotic adjustment in crop plants, even though very few loci have a major impact during stress conditions (Lilley et al., 1996). On the other hand, DRO1 was found to enhance yield under drought stress (Uga et al., 2013a).

Genomics plays a pivotal role in deciphering the new genome sites that code for critical traits and apprehending the nucleotide

variations linked to specific variant phenotypes. Specific Rice SNP-seek database was updated to gain insights into nucleotide and phenotypic variants (Locedie et al., 2017). The nucleotide variants were curated by databases of rice such as, Gramene (Tello-Ruiz et al., 2016), NCBI (Sherry et al., 2001), RiceVarMap (Zhao et al., 2015), RMBreeding (Zheng et al., 2015), and IC4R (The IC4R Consortium, 2015).

TRANSCRIPTOMICS

Transcriptomic studies involve studying total transcripts found in cells/tissues or organisms. Through transcriptome-wide studies, it has been revealed that a large number of TFs have helped to confer drought tolerance in rice. Broadly, two classes of TFs have been identified to mediate signaling and physiological responses in plants such as, ABA independent and ABA dependant activation of TFs for drought tolerance. ABA dependant TFs includes (1) basic leucine zipper (bZIP) and (2) NAM, ATAF, and CUC2 (NAC). The bZIP class of TFs plays a pivotal role in dehydration induced by ABA signaling under drought conditions (Yang et al., 2010). Moreover, ABF-3 mediated drought tolerance was more efficient than DREB1A/CBF3 triggered drought stress tolerance (Oh et al., 2005). NAC TFs are another potential member that belongs to the ABA dependant group, which consists of a DNA binding NAC domain (Palaniswamy et al., 2006; Guo et al., 2008). The SNAC1 is typical of the NAC family and is overexpressed in transgenic rice lines showing a higher seed setting rate and spikelet fertility under severe drought stress (Hu et al., 2006).

Two families of TFs are discussed in ABA-independent signaling pathways, zinc fingers and AP2/ERFs. Former TFs consist of Zn ions coordinated to motifs that stabilize the protein folds. These TFs include *WRKY*, *ZFP252*, and *Zat10/STZ* genes (Xu et al., 2008; Wu et al., 2009; Xiao et al., 2009). Among these, STZ was reported to enhance the fertility of spikelets and grain yield under drought stress (Xiao et al., 2009). Under the control of the drought inducible promoter *OsHVA22P*, *AtDREB1A/CBF3* gene overexpression enhanced drought tolerance in rice plants to increase the yield and spikelet fertility. Large numbers of TFs are deployed to impart drought tolerance in crop plants, but their successful introduction in crop plants is still in its infancy.

Tian et al. (2015) assembled the *de-novo* transcriptome of common wild rice (*Oryza rufipogon* Griff.) and identified several drought responsive and related genes in root tissue based on transcriptomic data, thus, providing critical data pertaining to genetic and genomic studies in rice. Analyzing the transcript and metabolic responses of two rice cultivars with contrasting drought-tolerance to long-term drought, Ma et al. (2016) showed that a large number of DEGs related to photosynthesis were upregulated in the drought tolerant variety and concluded that well-maintained photosynthesis under drought is a critical aspect for improved drought-tolerance in rice. An RNA-seq based transcriptomic profile between OsMIOX-overexpressing (OE) and wild-type (WT) rice plants revealed that this unique monooxygenase plays an essential role in drought tolerance. Significantly, upregulated DEGs in

OE lines were associated with TFs, plant hormone transduction and sugar metabolism (Shi et al., 2020). Luo et al. (2020) examined the morphological differences between upland and lowland rice ecotypes. Furthermore, genetic and transcriptomic divergences between these contrasting rice ecotypes under well-watered and drought conditions were observed using RNA-seq. The results revealed that the expression divergences were higher in upland rice than in lowland rice, and it was concluded that this transcriptomic divergence contributed to their morphological differences in drought tolerance.

Xia et al. (2020) investigated the temporal transcriptomic data of 12 rice genotypes varying in naturally induced drought in field conditions. They highlighted that the drought tolerant varieties had higher proportions of upregulated DEGs related to fucose, trehalose, and raffinose metabolic processes that were specifically induced. In addition, the DEGs related to proteins and their modifications such as protein peptidyl-prolyl isomerization, histone deacetylation, transcriptional attenuation and ferric iron transport, were induced earlier in drought tolerant genotypes. An integrated phenotypic and transcriptomic landscape of 61 rice (Oryza sativa) varieties grown in an upland field with highly diverse below-ground traits under mild drought stress was reported (Kawakatsu et al., 2020). Rice accessions were classified into four admixture groups based on phenotypic variation and the transcriptomic analysis showed admixture group-specific enrichment of stress-related genes. Signaling network and key TFs were identified by co-expression network analysis of DNA affinity purification followed by sequencing (DAP-seq) datasets and were found to negatively regulate crown root diameter. Tarun et al. (2020) carried out co-expression network analysis of the transcriptomes obtained from emerging panicle tissues and flagleaf in drought-tolerant yield recurrent parent and introgression line under drought stress. Results indicated that plants scavenge ROS and enhanced protein turnover as pivotal mechanisms induced in both tissues during drought stress conditions.

Liang et al. (2021) conducted comparative RNA-seq analysis of a moderately tolerant breeding line and drought susceptible elite varieties during the grain-filling stage and observed that the tolerant line had much earlier responses at the transcriptomic level. Genes and gene families related to TFs, drought tolerance genes, and reactive oxygen species (ROS) scavengers were significantly altered. More recently, Tiwari et al. (2021) carried out time-based comparative transcriptomic profiling of two varieties that showed contrasting drought tolerance. Comparatively, a significant proportion of the identified DEGs related to phytohormone signaling, stressresponse, photosynthesis, anti-oxidative mechanisms and TFs were higher in the drought tolerant variety. Furthermore, QTL mapping could distinguish drought-responsive traits at the chromosomal level, because a very high number of DEGs associated with drought stress was observed in the drought tolerant variety.

Studies have revealed that to unravel the complexity of expression levels, it is important to investigate the transcriptomes as a function of development and environmental conditions (Dayan et al., 2015; Liu et al., 2015; Gould et al., 2018). Furthermore, the transcriptomic investigation of rice lines under

two different drought stress conditions could help to elucidate the possible molecular switches adapted by rice crops (Gould et al., 2018). The variation induced by stress tolerant and stress sensitive cultivars of rice at the molecular level was identified by microarray analysis (Walia et al., 2005; Lenka et al., 2011; Ray et al., 2011). The studies evaluated the differential expression of genes by both tolerant and sensitive cultivars of rice under stress conditions (Walia et al., 2005; Lenka et al., 2011). Therefore, it is important to gain in-depth knowledge about the complexity of transcriptional regulation in rice during drought and salinity stresses using advanced technologies.

PROTEOMICS

This approach involves studying the proteomes of a cell/organ/tissue and organism for experimental purposes (Rakwal and Agrawal, 2003). Proteomes are largely influenced by the environmental conditions that occur in the immediate vicinity of biological systems. Therefore, it is pertinent that genomes can produce a large number of proteomes according to the prevailing conditions. Little progress has been made in rice proteomics compared to genomic studies. Proteome maps of rice seeds, roots and leaves investigated at different developmental stages augment the development of stress tolerant rice varieties (Komatsu et al., 1999; Koller et al., 2002; Tanaka et al., 2005; Nozu et al., 2006). Approximately 31 drought responsive proteins have been identified in crop plants (Muthurajan et al., 2010). The preliminary Rice Proteome Database was available (Komatsu et al., 2004; National Institute of Agrobiological Sciences) and the data might be used to process the comparative proteomic analysis of drought tolerant and control varieties of rice.

A tandem mass tag (TMT)-based proteomic approach was applied to compare the roots, flag leaves and spikelets of a wild type and its near-isogenic line (NIL) harboring QTL qDTY 12.1, a large effect QTL for rice yield under drought (Raorane et al., 2015). The proteomic data correlated with drought-specific morpho-physiological responses and tissue-specific differences in protein abundance were observed. The DAPs were related to respiration, photosynthesis, energy generation and carbonnitrogen acquisition/remobilization largely through the pathways of sugar/starch and amino acid metabolism. Paul et al. (2015) carried out comparative proteomic analysis to understand the metabolic networks regulated by the proteomes of roots in transgenic and wild type rice plants. Later studies led to the identification of DREB1A overexpression in rice plants by employing two-dimensional gel electrophoresis (2DE)-coupled with MALDI TOF MS/MS cultivated under drought stress. Most DAPs were linked to carbohydrate metabolism and defense, and a novel protein, R40C1, significantly accumulated only in the roots of transgenic plants. Agrawal et al. (2016) employed the 2DE-MALDI-TOF-MS/MS approach to compare the root cytoplasmic proteome of a drought tolerant rice variety for drought tolerant rice grown under normal and PEG-induced drought conditions. Major proportion of the identified differentially abundant proteins (DAPs) were involved in bioenergy, metabolism, cell defense and rescue, protein

biogenesis, protein storage and cell signaling, and the authors proposed that protein biogenesis, cell defense, and efficient homeostasis could render better drought-adaptation. Wu et al. (2016) carried out comparative label-free shotgun proteomics and TMT labeling proteomics of two rice cultivars with contrasting genetic backgrounds and levels of tolerance to drought. Proteomics results indicated alteration in multiple stress and defense response related proteins. Notably, a ClpD1 protease was upregulated several folds only in the drought tolerant variety, and porphyrin and chlorophyll biosynthesis pathways were down-regulated. A comparative proteomic analysis of a susceptible rice cultivar and its stress-resistant somaclonal mutant line showed that a large proportion of DAPs primarily related to retrotransposons were predominantly identified in the resistant line, suggesting that the gene expression associated with drought tolerance mechanisms is under strong epigenetic regulation. It was also suggested based on proteomic analysis that photosynthetic adaptation through NADP(H) homeostasis contributes to drought tolerance in rice (Chintakovid et al., 2017).

Wang et al. (2017) performed physiological measurements and conducted 2DE-based proteomic analysis of rice flag leaves at the flowering and milk stages to understand the drought responsive mechanism. Based on the proteomic analysis, it was postulated that at the flowering stage, CO₂ assimilation and ATP synthesis were disrupted, while at the milk stage, both CO₂ assimilation and photosynthesis were impaired. However, there was an increased abundance of DAPs related to defense and antioxidant machinery, which suggests redox imbalance and activation of the ROS scavenging system in drought stressed plants (Saini et al., 2021).

To understand the role of jasmonic acid under drought stress, studies were conducted to compare the morpho-physiological traits and the root proteome of a wild type (WT) rice plant with its jasmonic acid biosynthesis mutant coleoptile photomorphogenesis 2 (cpm2), disrupted in the allene oxide cyclase (AOC) gene. Studies revealed that roots of cpm2 mutants compared to wild type had higher water use efficacy, high ABA concentration in shoots, and higher stomatal conductance under drought conditions. Moreover, the TMT technique was used to analyze the root proteome to better understand this difference at the molecular level. Identification of other DAPs proposed increased energy metabolism (i.e., increased mobilization of resources) and ROS scavenging in cpm2 under drought. In addition, it was revealed that there was an abundance of proteins pertaining to cell growth, secondary metabolism and cell wall synthesis in cpm2 mutants grown under drought stress. A clear understanding of adaptation and responses in roots of cpm2 mutants under drought stress was elucidated by proteome-guided metabolites, transcripts and histological investigations (Dhakarey et al., 2017). By conducting comparative morphological and proteomic analysis of two rice genotypes, 5 DAPs, including chitinase, were found to be abundant under drought stress (Anupama et al., 2019).

Cunha et al. (2019) investigated APX found in rice thylakoids in response to mild drought stress by carrying out physiological measurements and leaf proteomic analysis of thylakoidal APX knockdown rice plants (apx8) and non-transformed control plants. A correlation was observed between the sensitivity of plants to mild drought stress and the lower accumulation of DAPs related to several metabolic processes, especially photosynthesis, photorespiration and redox metabolism, in apx8 plants. Although apx8 effectively induces other compensatory antioxidant mechanisms in well-watered conditions, the apx8 plants could not maintain H2O2 homeostasis and accordingly avoid adverse conditions under mild drought conditions. To carry label-free proteomic studies, Hamzelou et al. (2020) exposed eight genotypes of upland and lowland rice plants to drought stress at the late vegetative stage. The majority of the identified DAPs under drought conditions were related to photosynthesis, oxidative stress response, proteolysis, and translation of stress-responsive proteins, such as heat shock, and LEA proteins (late embryogenesis-associated proteins) sharply increased under drought stress. Du et al. (2020) investigated the physiology of rice grown under two nitrogen management modes by proteomics and metabolomics approaches to investigate their yield formation and the mechanism of nitrogen regulation for drought tolerance. It was evident from the proteomic analysis that the most altered biological processes and pathways were related to the biosynthesis of primary and secondary metabolites important for the growth and metabolism of crop plants.

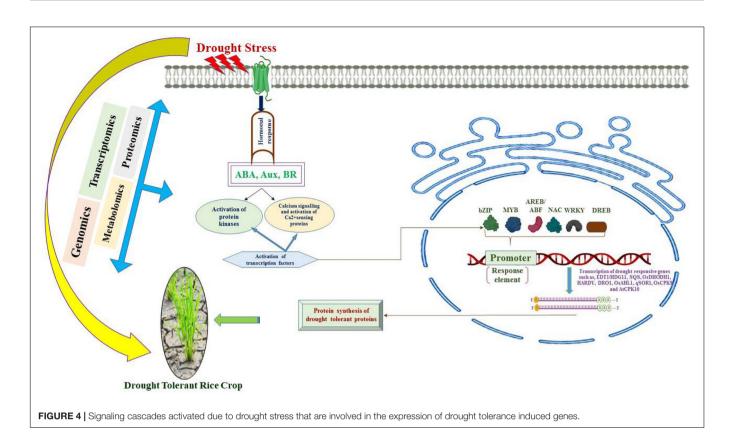
The rice proteomes are assessed by several gel and off-gel based proteomics techniques. For instance, proteome changes are analyzed by a combination of proteomic techniques such as 2-DE matrix-assisted laser desorption ionization-tandem time of flight (MALDI-TOF/TOF) and 1-DE/LC-Fourier transform-ion cyclotron resonance (FT-ICR) MS based analyses (Wan and Liu, 2008). Moreover, several proteomic databases for rice have been established and updated to curate the sequences for desirable traits. For example, the Rice Yield-related Database (RicyerDB) was created to curate the research related to yield related traits in rice by assessing the genomic and proteomic data inputs, and it provides a vast literature source related to where we can browse, investigate and analyze the desirable genes related to yield (Jiang et al., 2018). Systemic proteome analysis through a wide range of proteomic techniques is a powerful tool to investigate the complete proteomes of rice plants at different levels, such as the organelle, cell, tissue, organ and organism levels, and further provides insights into assessing proteomes once plants are exposed to stressful conditions.

METABOLOMICS OF RICE PLANTS

Metabolomic studies involve comprehensive and systematic identification and quantification of endogenous metabolites from biological samples and have rapidly evolved and advanced in recent years (Ryan and Robards, 2006; Kumar et al., 2017).

Some Major Breakthroughs in Metabolomics-Related Articles on Rice From 2015 to 2021

These include: Nam et al. (2016) – studies carried in transgenic rice to unravel the metabolomic changes in grains of well-watered



and drought conditions; Ma et al. (2016) – studied the key pathways by transcriptomic and metabolomic studies to maintain photosynthesis under the drought and the consequent drought-tolerance in rice; Xiong et al. (2019) – Analysis of grain yield reduction in rice by comprehensive proteomic, metabolomic, and physiological analyses of grain under abrupt drought-flood alternation stress; and, Du et al. (2020) – Combinatorial proteomic, metabolomic and physiological studies of rice growth and grain yield with heavy nitrogen application before and after drought.

Metabolomics analysis in plant systems is rapidly advancing because it measures total, or groups of metabolites expressed in few samples in specific time-periods. Metabolomes of higher plants could comprise hundreds of thousands of metabolites out of that number, so little is known (Wang S. et al., 2019). Qualitative and quantitative measures of plant metabolomes mirror their responses to organic phenomenon and abiotic stimuli, genome, and physiological standing, serving as a connecting link between genotypes and phenotypes. They provide a considerable contribution to stress biology by deciphering numerous compounds such as by-products of stress metabolism, signaling molecules, and compounds, that square measure a part of the plant acclimatization method (Lanzinger et al., 2015; Ramalingam et al., 2015). Metabolites such as amino acids, fatty acids, soluble sugars, nucleotides, organic acids, phenolics, peptides, cofactors, and secondary metabolites act as cellular measurements that changed during drought stress (Das et al., 2017; Arora et al., 2018). Various metabolomic approaches elucidated the wide range of metabolite

markers expressed in response to abiotic stressors (Ghatak et al., 2018). Several of these metabolites square measure crucial parts of the plant's weapons system. Polyamines and phenoplast compounds are major substantial plant secondary metabolites that impart tolerance, and square measures are interpreted as replacements for biostimulants underneath environmental stress, especially drought stress conditions (Aninbon et al., 2016; Chen D. et al., 2019). Underneath environmental stresses, metabolic changes could assert some necessary metabolic products, which illustrates that the metabolic pathways are regulated at different levels. Additionally, plants can also change metabolic pathways to accumulate giant amounts of energy, which is a prerequisite to resist environments (Ullah et al., 2017). Metabolic analyses have shown that changes in metabolites under drought responses in various plant species are pivotal for developing adaptations. In Arabidopsis, most amino acid intermediates (such as from proline, glutamine, tryptophan, alanine, aspartate, ornithine, isoleucine, leucine, and valine) from the TCA cycle (such as 2oxoglutarate, cis-aconitate, and succinate), flavonoids (such as quercetin and cyanidin) and lipids accumulate under drought stress (Tarazon et al., 2015; Pires et al., 2016). Similarly, in rice the metabolomic data suggest that in response to increasing water stress, rice cultivars that were drought tolerant accumulated higher levels of carbohydrates (fructose, glucose, and myo-inositol) compared with the susceptible cultivars (Barnaby et al., 2019).

Both quantitative and qualitative rice metabolomics analyses help to investigate primary and secondary metabolites and

their differential expression patterns during biotic and abiotic stress conditions (Khakimov et al., 2014). Major metabolic techniques, such as nuclear magnetic resonance (NMR), GC/MS, and LC/MS in combination with Fourier transform ion cyclotron resonance (FT-ICR), have been employed to decipher the metabolite profiles of organisms in conjunction with databases, including Kyoto Encyclopedia of Genes and Genomes (KEGG) and MetaCyc (Kanehisa et al., 2012; Morreel et al., 2014; Caspi et al., 2018; Chen L. et al., 2021). The database specific to rice, including RiceCyc, is constricted based on metabolic networking and MetaCyc (Chae et al., 2007). To comprehend the basic mechanism of the response to abiotic stress, in particular drought, comparative metabolomics approaches provide a solid framework of gene expression, protein expression and other metabolites expressed temporally. Metabolomics approaches are promising technical interventions that overall serve as frameworks to obtain an in-depth biochemical and genetic portrait of organisms to pave the way for molecular breeding under stress conditions (Bino et al., 2004; Fernie and Schauer, 2009; Kusano et al., 2011).

EPIGENOMICS FOR DROUGHT TOLERANCE

The epigenome could be outlined as the summation of all of the biochemical changes in nuclear polymer, simple proteins and tiny non-coding ribonucleic acid biogenesis of a cell. Studies on the epigenetic changes in and around polymers that regulate ordination activity are outlined as epigenetics, and the branch of genetic science that addresses epigenomic studies is named epigenomics. Plants have evolved non-heritable, extremely subtle systems to deal with varied environmental stresses. The last decade has witnessed considerable progress in understanding the signaling and metabolic pathways that are dominant in plant responses to stresses, which has been summarized in previous reviews (Ku et al., 2018; Kumar et al., 2019). Activation of signaling pathways typically results in transcriptional adjustments to initiate the expression of stress responsive proteins (Kim et al., 2019; Shahid, 2019; Wu et al., 2019). A large number of adaptations have been attributed to crop plants in response to stress factors (Thomashow, 1999). The temporal responses to stress include the expression of stressinduced proteins, RNA molecules and metabolites. These stress adaptions could be longer if phenological and morphological adjustments have been accomplished. The transcriptional reprogramming and regulation of stress-responsive genes are critical to numerous epigenetic processes and components, such as DNA methylation, protein modifications and non-coding RNAs based regulations (Deleris et al., 2016; Kim et al., 2017; Chen R. et al., 2019). Drought stress conditions primarily tend to extend demethylation. It is conjointly discovered that DNA methylation shows tissue specificity. A variation of up to 1% methylation occurs across cells, tissues, genotype and organic process stages. The DNA methylation level is reported to be lower in roots than in leaves, which indicates

an important role of roots in addressing water scarcity (Suji and John, 2010). The correlation between drought stress and DNA methylation is shown in lowland and drought-tolerant rice cultivars. The drought inclined IR20 variety showed hypomethylation under drought conditions, whereas the tolerant varieties "Paiyur" and "PMK3" showed hyper-methylation. These changes in methylation patterns were found to be responsible for the differential expression of stress responsive genes (Gayacharan, 2019). In another study conducted in rice, it was illustrated that hypomethylation has an important role within the drought tolerance attributes of rice genotypes. Epigenomic studies involve a range of techniques, such as chromatin immunoprecipitation (ChiP), ChiP-sequencing, methylated-DNA immunoprecipitation and shotgun bisulfite sequencing. For the development of resilient rice cultivars, it is pertinent to focus on the profile and functioning of epigenomic profiles, such as histone modifications, DNA methylation, diverse classes of regulatory non-coding RNAs and especially the 3D genomic structure of rice (Lu et al., 2020). The role of epigenomics studies has considerably contributed to studying the changing dynamics of expression patterns under stress conditions, thus unraveling the complexity of crop plants in response to a wide range of stress factors. In addition, a detailed account on rice epigenomics variations and their identification is critical to the characterization of phenotypes fitting to agronomic traits for providing framework for enhancing productivity and traits in rice crops (Chen and Zhou, 2013).

CONCLUSION

The mechanism of drought tolerance is an important quantitative trait that is accompanied by several phenotypic adjustments and adaptations. A large number of stressors are supposed to be tolerated by crop plants, such as high temperatures, high irradiance, toxicities, and nutrient deficiencies due to the induction of drought stress. The past few decades have witnessed a higher prevalence of stresses due to abiotic factors imparting hindrances to growth and productivity in rice crop. Globally, scientists are extensively investigating strategies to cope with ever increasing climatic change and drought stress factors. The emergence of innovation in omics technologies has led to the deciphering of a large amount of molecular machinery to adapt to a large number of environmental factors. Besides, multiomics data integration and molecular modeling protocols can be employed to understand the complex traits to unearth the tolerance mechanism and yield of major legumes and cereal crops (Pazhamala et al., 2021; Yang et al., 2021). It is pertinent that enhancing tolerance to drought in rice requires the establishment of research programs to investigate complex networks of molecular interactions. Investigations based on transcriptomics, proteomics and metabolic studies should be initiated in unison to unravel the drought tolerance mechanisms in rice. These techniques will further widen our understanding of molecular signaling cascades and a wide range of stress responsive proteins and intracellular adjustments under stress factors (Figure 4).

Moreover, the cited literature validates the role of investigating QTLs and SNPs that play pivotal role of genes in drought tolerance in rice crops. This investigation at large could possibly augment our efforts to develop drought tolerant crop plants such as rice to ensure food security. The applications of markers and genomic selection are an efficient way to improve crop plants. Moreover, the development of high throughput phenomics techniques will help to augment the in-depth understanding of the mechanisms of tolerance to stressors. A low level of progress has been made with respect to proteomics, epigenomics and metabolomics in conjunction with genomic approaches. It is an important prerequisite to use the data obtained from these approaches to understand the genetic and physiological basis of drought tolerance. In addition, efforts aimed at combinatorial omics approaches and practical plant-breeding applications can accelerate progress in producing rice cultivars that are suitable for rainfed environments or to realize significant freshwater savings. Against this background, a PANOMICS platform that integrates mathematical and statistical tool boxes with omics data to facilitate improvement of crops through discovering pathways and target genes critical for resilience and creation of elite lines (Weckwerth et al., 2020). Finally, physiological and morphological adaptations accomplished by molecular mechanisms traced in resistant parent lines can be utilized to transfer the resilient genes to mainstream crop plants to become very adept to a particular environment, to establish a solid base for food security at a global level.

REFERENCES

- Agrawal, L., Gupta, S., Mishra, S. K., Pandey, G., Kumar, S., Chauhan, P. S., et al. (2016). Elucidation of complex nature of PEG induced drought-stress response in rice root using comparative proteomics approach. *Front. Plant Sci.* 7:1466. doi: 10.3389/fpls.2016.01466
- Akshaya, K. B., Shamim, Md, Cruzado, K., Soriano, G., Ghatak, A., Toleco, M. R., et al. (2017). "Role of biotechnology in rice production," in *Rice Production Worldwide*, eds B. Chauhan, K. Jabran, and G. Mahajan (Cham: Springer), 487–547. doi: 10.1007/978-3-319-47516-5_18
- Aninbon, C., Jogloy, S., Vorasoot, N., Nuchadomrong, S., Senawong, T., Holbrook, C. C., et al. (2016). Effect of mid-season drought on phenolic compounds in peanut genotypes with different levels of tolerance to drought. *Field Crops Res.* 187, 127–134.
- Anupama, A., Bhugra, S., Lall, B., Chaudhury, S., and Chugh, A. (2019). Morphological, transcriptomic and proteomic responses of contrasting rice genotypes towards drought stress. *Environ. Exp. Bot.* 166:103795. doi: 10.1016/j.envexpbot.2019.06.008
- Arora, N., Dubey, D., Sharma, M., Patel, A., Guleria, A., Pruthi, P. A., et al. (2018). NMR-based metabolomic approach to elucidate the differential cellular responses during mitigation of arsenic (III, V) in a green microalga. ACS Omega. 3, 11847–11856. doi: 10.1021/acsomega.8b01692
- Asia (2021). Available online at: https://ricepedia.org/rice-around-the-world/asia. (accessed September 09 16, 2021).
- Avramova, V., Nagel, K. A., AbdElgawad, H., Bustos, D., DuPlessis, M., Fiorani, F., et al. (2016). Screening for drought tolerance of maize hybrids by multi-scale analysis of root and shoot traits at the seedling stage. *J. Exp. Bot.* 67, 2453–2466. doi: 10.1093/jxb/erw055
- Barnaby, J. Y., Rohila, J. S., Henry, C. G., Sicher, R. C., Reddy, V. R., and McClung, A. M. (2019). Physiological and metabolic responses of rice to reduced soil moisture: relationship of water stress tolerance and grain production. *Agronomy* 20:1846. doi: 10.3390/ijms20081846

AUTHOR CONTRIBUTIONS

SZ and RR conceived the idea, framed outline and contributed in writing and final editing of the manuscript. RAM contributed in writing genomics part and prepared the figures. LE prepared transcriptomics part of the manuscript. AM edited the manuscript and also contributed in preparing the proteomics and transcriptomics sections. AH, MM, and RM prepared the tables and also contributed in preparing genomics section. RS and NS contributed in writing introduction and physiological section of the manuscript. JR revised the original draft and carried out the final editing with SZ and RR. All authors contributed to the article and approved the submitted version.

ACKNOWLEDGMENTS

SZ and AM acknowledges European Union for funding Erasmus Programme (2018–2020 and 2020–2023) that helped in interaction among the authors for preparing this manuscript. JR acknowledge the NSF funding (award# 1849708). SZ, RR, and AM also acknowledge INPPO (International Plant Proteomics Organization) for helping in establishing the collaboration among scientists from various institutions and countries. SZ also acknowledges Hon'ble Vice Chancellor, SKUAST-Kashmir (Prof. Nazir A. Ganai) for his support and encouragement for the international collaboration.

- Bernier, J., Kumar, A., Venuprasad, R., Spaner, D., and Atlin, G. N. (2007). A large-effect QTL for grain yield under reproductive-stage drought stress in upland rice. Crop Sci. 47, 507–516.
- Bino, R. J., Hall, R. D., Fiehn, O., Kopka, J., Saito, K., Draper, J., et al. (2004). Potential of metabolomics as a functional genomics tool. *Trends Plant Sci.* 9, 418–425. doi: 10.1016/j.tplants.2004.07.004
- Biswas, S., Tian, J., Li, R., Chen, X., Luo, Z., Chen, M., et al. (2020). Investigation of CRISPR/Cas9-induced SD1 rice mutants highlights the importance of molecular characterization in plant molecular breeding. *J. Genet. Genom.* 47, 273–280. doi: 10.1016/j.jgg.2020.04.004
- Borrell, A., Hammer, G., and Oosterom, E. (2001). Stay-green: a consequence of the balance between supply and demand for nitrogen during grain filling. *Ann. Appl. Biol.* 138, 91–95.
- Bouman, B., Barker, R., Humphreys, E., Tuong, T. P., Atlin, G., Bennett, J., et al. (2007). "Rice: feeding the billions," in *Water for Food, Water for Life: A Comprehensive Assessment of Water Management in Agriculture*, ed. D. Molden (London: IWMI), 515–549.
- Caspi, R., Billington, R., Fulcher, C. A., Keseler, I. M., Kothari, A., Krummenacker, M., et al. (2018). The MetaCyc database of metabolic pathways and enzymes. Nucleic Acids Res. 46, 633–639.
- Chae, M. J., Lee, J. S., Nam, M. H., Cho, K., Hong, J. Y., Yi, S. A., et al. (2007).
 A rice dehydration-inducible SNF1-related protein kinase 2 phosphorylates an abscisic acid responsive element-binding factor and associates with ABA signalling. *Plant Mol. Biol.* 63, 151–169. doi: 10.1007/s11103-006-9 079-x
- Champoux, M. C., Wang, G., Sarkarungm, S., Mackill, D. J., O'Toole, J. C., Huang, et al. (1995). Locating genes associated with root morphology and drought avoidance in rice via linkage to molecular markers. Theor. Appl. Genet. 90, 969–981. doi: 10.1007/BF00222910
- Chaves, M. M., Pereira, J. S., Maroco, J., Rodrigues, M. L., Ricardo, C. P. P., Osório, M. L., et al. (2002). How plants cope with water stress in the field photosynthesis and growth. *Ann. Bot.* 89, 907–916. doi: 10.1093/aob/mcf105

Chen, D., Shao, Q., Yin, L., Younis, A., and Zheng, B. (2019). Polyamine function in plants: metabolism, regulation on development, and roles in abiotic stress responses. Front. Plant Sci. 9:1975. doi: 10.3389/fpls.2018.01945

- Chen, L., Lu, W., Wang, L., Xing, X., Chen, Z., Teng, X., et al. (2021). Metabolite discovery through global annotation of untargeted metabolomics data. *Nat. Methods* 18, 1377–1385. doi: 10.1038/s41592-021-01303-3
- Chen, M.-H., McClung, A. M., Rohila, J. S., and Barnaby, J. (2021). Effects of alternate wetting and drying irrigation management and air temperature during grain fill on rice grain physicochemical and functionality traits of US inbred varieties. *Cereal Chemistry* 98, 980–993. doi: 10.1002/cche.10440
- Chen, R., Li, M., Zhang, H. Y., Duan, L. J., Sun, X. J., Jiang, Q. Y., et al. (2019). Continuous salt stress-induced long non-coding RNAs and DNA methylation patterns in soybean roots. *BMC Genom*. 20:730. doi: 10.1186/s12864-019-6 101-7
- Chen, X., and Zhou, D. X. (2013). Rice epigenomics and epigenetics: challenges and opportunities. *Curr. Opin. Plant Biol.* 16, 164–169. doi: 10.1016/j.pbi.2013.
- Cheng, X., Liu, X., Wang, H., Ji, X., Wang, K., Wei, M., et al. (2015). Effect of emamectin benzoate on root-knot nematodes and tomato yield. PLoS One 10:e0141235. doi: 10.1371/journal.pone.0141235
- Chintakovid, N., Maipoka, M., Phaonakrop, N., Mickelbart, M. V., Roytrakul, S., and Chadchawan, S. (2017). Proteomic analysis of drought-responsive proteins in rice reveals photosynthesis-related adaptations to drought stress. Acta Physiol. Plantar. 39, 1–13.
- Comas, L. H., Becker, S. R., Cruz, V. M., Byrne, P. F., and Dierig, D. A. (2013). Root traits contributing to plant productivity under drought. Front. Plant Sci. 4:442. doi: 10.3389/fpls.2013.00442
- Cooper, B., Clarke, J. D., Budworth, P., Kreps, J., Hutchison, D., Park, S., et al. (2003). A network of rice genes associated with stress response and seed development. *Proc. Natl. Acad. Sci. U.S.A.* 100, 4945–4950. doi: 10.1073/pnas. 0737574100
- Cosgrove, D. J. (2005). Growth of the plant cell wall. Nat. Rev. Mol. Cell Biol. 6, 850–861.
- Courtois, B., Ahmadi, N., Khowaja, F., Price, A. H., Rami, J. F., Frouin, J., et al. (2009). Rice root genetic architecture: meta-analysis from a drought QTL database. *Rice* 2, 115–128.
- Coutinho, F. S., Rodrigues, J. M., Lima, L. L., Mesquita, R. O., Carpinetti, P. A., Machado, J. P. B., et al. (2021). Remodeling of the cell wall as a droughttolerance mechanism of a soybean genotype revealed by global gene expression analysis. 3Biotech 2, 14–31. doi: 10.1007/s42994-021-00043-4
- Cunha, A. P., Zeri, M., Deusdará Leal, K., Costa, L., Cuartas, L. A., Marengo, J. A., et al. (2019). Extreme drought events over Brazil from 2011 to 2019. Atmosphere 10:642. doi: 10.3390/atmos10110642
- Das, A., Rushton, P. J., and Rohila, J. S. (2017). Metabolomic profiling of soybeans (*Glycine max* L.) reveals the importance of sugar and nitrogen metabolism under drought and heat stress. *Plants* 6:21. doi: 10.3390/plants602 0021
- Dawe, D., Pandey, S., and Nelson, A. (2010). "Emerging trends and spatial patterns of rice production," in *Rice in the Global Economy: Strategic Research and Policy Issues for Food Security*, eds S. Pandey, D. Byerlee, D. Dawe, A. Dobermann, S. Mohanty, S. Rozelle, et al. (Los Baños, Philippines: International Rice Research Institute), 15–35.
- Dayan, D. I., Crawford, D. L., and Oleksiak, M. F. (2015). Phenotypic plasticity in gene expression contributes to divergence of locally adapted populations of Fundulus heteroclitus. Mol. Ecol. 24, 3345–3359. doi: 10.1111/mec.13188
- de Avila, L. A., Martini, L. F. D., Mezzomo, R. F., Refatti, J. P., Campos, R., and Cezimbra, D. M. (2015). Rice water use efficiency and yield under continuous and intermittent irrigation. *Agronomy J.* 107, 442–448. doi: 10.2134/agronj14. 0080
- Deivanai, S., Devi, S. S., and Rengeswari, P. S. (2010). Physiochemical traits as potential indicators for determining drought tolerance during active tillering stage in rice (*Oryza sativa L.*). *Pertanika J. Trop. Agric. Sci.* 33, 61–70.
- Deleris, A., Halter, T., and Navarro, L. (2016). DNA methylation and demethylation in plant immunity. Annu. Rev. Phytopathol. 54, 579–603. doi: 10.1146/annurevphyto-080615-100308
- Delphine, F., Jefferies, S., Kuchel, H., and Langridge, P. (2010). Genetic and genomic tools to improve drought tolerance in wheat. J. Exp. Bot. 61, 3211– 3222. doi: 10.1093/jxb/erq152

- Dhakarey, R., Raorane, M. L., Treumann, A., Peethambaran, P. K., Schendel, R. R., Sahi, V. P., et al. (2017). Physiological and proteomic analysis of the rice mutant cpm2 suggests a negative regulatory role of jasmonic acid in drought tolerance. Front. Plant Sci. 8:1903.
- Du, H., Wang, N., Cui, F., Li, X., Xiao, J., and Xiong, L. (2010). Characterization of the β-carotene hydroxylase gene DSM2 conferring drought and oxidative stress tolerance by increasing xanthophylls and abscisic acid synthesis in rice. *Plant Physiol.* 154, 1304–1318. doi: 10.1104/pp.110.163741
- Du, J., Shen, T., Xiong, Q., Zhu, C., Peng, X., He, X., et al. (2020). Combined proteomics, metabolomics and physiological analyses of rice growth and grain yield with heavy nitrogen application before and after drought. *BMC Plant Biol*. 20:1–17. doi: 10.1186/s12870-020-02772-y
- Fahad, S., Rehman, A., Shahzad, B., Tanveer, M., Saud, S., Kamran, M., et al. (2019).
 "Rice responses and tolerance to metal/metalloid toxicity," in *Advances in Rice Research for Abiotic Stress Tolerance*, (Woodhead Publishing), 299–312.
- Fang, Y., Liao, K., Du, H., Xu, Y., Song, H., Li, X., et al. (2015). A stress-responsive NAC transcription factor SNAC3 confers heat and drought tolerance through modulation of reactive oxygen species in rice. *J. Exp. Bot.* 66, 6803–6817. doi: 10.1093/jxb/erv386
- Farooq, M., Basra, S. M. A., Wahid, A., Ahmad, N., and Saleem, B. A. (2009). Improving the drought tolerance in rice (*Oryza sativa* L.) by exogenous application of salicylic acid. *J. Agron. Crop Sci.* 195, 237–246. doi: 10.1111/j. 1439-037x.2009.00365.x
- Fernie, A. R., and Schauer, N. (2009). Metabolomics-assisted breeding: a viable option for crop improvement. *Trends Genet.* 25, 39–48. doi: 10.1016/j.tig.2008. 10.010
- Fleury, D., Jefferies, S., Kuchel, H., and Langridge, P. (2010). Genetic and genomic tools to improve drought tolerance in wheat. *J. Exp. Bot.* 61, 3211–3222.
- Foss, E. J., Radulovic, D., Shaffer, S. A., Goodlet, D. R., Kruglyak, L., and Bedalov, A. (2011). Genetic variation shapes protein networks mainly through nontranscriptional mechanisms. *PLoS Biol.* 9:e1001144. doi: 10.1371/journal.pbio. 1001144
- Fuji, H., Chinnusamy, V., Rodrigues, A., Rubio, S., Antoni, R., Park, S. Y., et al. (2009). In vitro reconstitution of an ABA signaling pathway. *Nature* 462, 660–664.
- Fukai, S., and Cooper, M. (1995). Development of drought-resistant cultivars using physiomorphological traits in rice. *Field Crops Res.* 40, 67–86.
- Fukai, S., and Cooper, M. (2002). "Field screening of adaptability in drought prone rainfed lowland rice: ACIAR experience in thailand and laos, in: international workshop on field screening for drought tolerance in rice," in *Proceedings of An International Workshop on Field Screening for Drought Tolerance in Rice, 11-14 December 2000*, eds N. P. Saxena and J. C. O'Toole (Patancheru: International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, India), 61–32. doi: 10.1016/s0378-4290(99)00051-9
- Fukao, T., and Xiong, L. (2013). Genetic mechanisms conferring adaptation to submergence and drought in rice: simple or complex? Curr. Opin. Plant Biol. 16, 196–204. doi: 10.1016/j.pbi.2013.02.003
- Gao, T., Wu, Y., Zhang, Y., Liu, L., Ning, Y., Wang, D., et al. (2011). OsSDIR1 overexpression greatly improves drought tolerance in transgenic rice. *Plant Mol. Biol.* 76, 145–156. doi: 10.1007/s11103-011-9775-z
- Gauchan, D., and Pandey, S. (2012). "Synthesis of key results and implications," in Patterns of Adoption of Improved Rice Varieties and Farm-Level Impacts in Stress-Prone Rainfed Areas in South Asia, eds S. Pandey, D. Gauchan, M. Malabayabas, M. Bool-Emerick, and B. Hardy (Los Baños, Philippines: International Rice Research Institute), 3.
- Gayacharan, A. J. (2019). Epigenetic responses to drought stress in rice (Oryza sativa L.). Physiol. Mol. Biol. Plants 19, 379–387. doi: 10.1007/s12298-013-0 176-4
- Geiger, D., Scherzer, S., Mumm, P., Stange, A., Marten, I., Bauer, H., et al. (2009). Activity of guard cell anion channel SLAC1 is controlled by drought-stress signaling kinase-phosphatase pair. *Proc. Natl. Acad. Sci. U.S.A.* 106, 21425– 21430. doi: 10.1073/pnas.0912021106
- Ghatak, A., Chaturvedi, P., and Weckwerth, W. (2017). Cereal crop proteomics: systemic analysis of crop drought stress responses towards markerassisted selection breeding. Front. Plant Sci. 8:757. doi: 10.3389/fpls.2017.0 0757
- Ghatak, A., Chaturvedi, P., and Weckwerth, W. (2018). "Metabolomics in plant stress physiology," in *Plant Genetics and Molecular Biology. Advances in*

Biochemical Engineering/Biotechnology, Vol. 164, eds R. Varshney, M. Pandey, and A. Chitikineni (Cham: Springer). doi: 10.1007/10_2017_55

- Ghazalpour, A., Bennett, B., Petyuk, V. A., Orozco, L., Hagopian, R., Mungrue, I. N., et al. (2011). Comparative analysis of proteome and transcriptome variation in mouse. *PLoS Genet.* 7:e1001393. doi: 10.1371/journal.pgen. 1001393
- Ghosh, D., and Xu, J. (2014). Abiotic stress responses in plant roots: a proteomics perspective. *Front. Plant Sci.* 5:6. doi: 10.3389/fpls.2014.00006
- Gould, B. A., Chen, Y., and Lowry, D. B. (2018). Gene regulatory divergence between locally adapted ecotypes in their native habitats. *Mol. Ecol.* 27, 4174– 4188. doi: 10.1111/mec.14852
- Gowda, V. R. P., Henry, A., Yamauchi, A., Shashidhar, H. E., and Serraj, R. (2011).
 Root biology and genetic improvement for drought avoidance in rice. Field Crop. Res. 122, 1–13. doi: 10.1016/j.fcr.2011.03.001
- Granier, C., Inzé, D., and Tardieu, F. (2000). Spatial distribution of cell division rate can be deduced from that of p34(cdc2) kinase activity in maize leaves grown at contrasting temperatures and soil water conditions. *Plant Physiol*. 124, 1393–1402. doi: 10.1104/pp.124.3.1393
- Gray, S. B., Dermody, O., and DeLucia, E. H. (2010). Spectral reflectance from a soybean canopy exposed to elevated CO2 and O3. J. Exp. Bot. 61, 4413–4422. doi: 10.1093/jxb/erq244
- Guo, A.-Y., Chen, X., Gao, G., Zhang, H., Zhu, Q.-H., Liu, X.-C., et al. (2008).
 PlantTFDB: a comprehensive plant transcription factor database. *Nucleic Acids Res*, 36, 966–969.
- Hamzelou, S., Pascovici, D., Kamath, K. S., Amirkhani, A., McKay, M., Mirzaei, M., et al. (2020). Proteomic responses to drought vary widely among eight diverse genotypes of rice (*Oryza sativa*). *Int. J. Mol. Sci.* 21:363. doi: 10.3390/ijms21010363
- Hasin, Y., Seldin, M., and Lusis, A. (2017). Multi-omics approaches to disease. Geno. Biol. 18, 1–15. doi: 10.1186/s13059-017-1215-1
- Henderson, S., Kamboonruang, V., and Cooper, M. (1995). "Evaluation of a glasshouse screening method to select for drought tolerance in rainfed lowland rice, in: N fed Lowland rice, in: fragile lives in fragile ecosystems," in *Proceedings* of the International Rice Research Conference, 13–17 February 1995 (Los Banos: International Rice Research Institute (IRRI)), 783–806.
- Henry, A., Dixit, S., Mandal, N. P., Anantha, M. S., Torres, R., and Kumar, A. (2014). Grain yield and physiological traits of rice lines with the drought yield QTL qDTY12.1 showed different responses to drought and soil characteristics in upland environments. Funct. Plant Biol. 41, 1066–1077. doi: 10.1071/FP13324
- Hirabayashi, H., Sasaki, K., Kambe, T., Gannaban, R. B., Miras, M. A., Mendioro, M. S., et al. (2015). qEMF3, a novel QTL for the early-morning flowering trait from wild rice, *Oryza officinalis*, to mitigate heat stress damage at flowering in rice, *O. sativa. J. Exp. Bot.* 66, 1227–1236. doi: 10.1093/jxb/eru474
- Hou, X., Xie, K., Yao, J., Qi, Z., and Xiong, L. (2009). A homolog of human skiinteracting protein in rice positively regulates cell viability and stress tolerance. *Proc. Natl. Acad. Sci. U.S.A.* 106, 6410–6415. doi: 10.1073/pnas.0901940106
- Hu, H., Dai, M., Yao, J., Xiao, B., Li, X., Zhang, Q., et al. (2006). Overexpressing a NAM, ATAF, and CUC (NAC) transcription factor enhances drought tolerance and salt tolerance in rice. *Proc. Natl. Acad. Sci. U.S.A.*. 103, 12987–12992.
- Huang, L., Wang, Y., Wang, W., Zhao, X., Qin, Q., Sun, F., et al. (2018). Characterization of transcription factor gene OsDRAP1 conferring drought tolerance in rice. Front. Plant Sci. 9:94. doi: 10.3389/fpls.2018.00094
- Huang, X.-Y., Chao, D.-Y., Gao, J.-P., Zhu, M. Z., Shi, M., and Lin, H. X. (2009).
 A previously unknown zinc finger protein, DST, regulates drought and salt tolerance in rice via stomatal aperture control. Genes Dev. 23, 1805–1817.
 doi:10.1101/gad.1812409
- Huynh, B. L., Ehlers, J. D., Huang, B. E., Muñoz-Amatriaín, M., Lonardi, S., Santos, J., et al. (2018). A multi-parent advanced generation inter-cross (MAGIC) population for genetic analysis and improvement of cowpea (Vigna unguiculata L. walp.). Plant J. Cell Mol. Biol. 93, 1129–1142. doi: 10.1111/tpj.13827
- Ikeda, K., Sunohara, H., and Nagato, Y. (2004). Developmental course of inflorescence and spikelet in rice. *Breed. Sci.* 54, 147–156. doi: 10.3390/ ijms22126610
- Iwata, S., Miyazawa, Y., Fujii, N., and Takahashi, H. (2013). MIZ1-regulated hydrotropism functions in the growth and survival of *Arabidopsis thaliana* under natural conditions. *Ann. Bot.* 112, 103–114. doi: 10.1093/aob/mct098

- Jeong, J. S., Kim, Y. S., Baek, K. H., Jung, H., Ha, S. H., Do Choi, Y., et al. (2010). Root-specific expression of OsNAC10 improves drought tolerance and grain yield in rice under field drought conditions. *Plant Physiol.* 153, 185–197. doi: 10.1104/pp.110.154773
- Jia, L., Xie, Y., Wang, Z., Luo, L., Zhang, C., Pélissier, P. M., et al. (2020). Rice plants respond to ammonium stress by adopting a helical root growth pattern. *Plant J.* 104, 1023–1037. doi: 10.1111/tpj.14978
- Jiang, J., Xing, F., Zeng, X., and Zou, Q. (2018). RicyerDB: a database for collecting rice yield-related genes with biological analysis. *Int. J. Biol. Sci.* 14, 965–970. doi: 10.7150/ijbs.23328
- Jiang, M., and Zhang, J. (2002). Water stress-induced abscisic acid accumulation triggers the increased generation of reactive oxygen species and up-regulates the activities of antioxidant enzymes in maize leaves. *J. Exp. Bot.* 53, 2401–2410. doi: 10.1093/ixb/erf090
- Jiang, Y., Cai, Z., Xie, W., Long, T., Yu, H., and Zhang, Q. (2011). Rice functional genomics research: progress and implications for crop genetic improvement. *Biotechnol. Adv.* 30, 1059–1070. doi: 10.1016/j.biotechadv.2011.08.013
- Jongdee, B., Fukai, S., and Cooper, M. (1998). "Genotypic variation for grain yield of rice under water-deficit conditions," in *Proceeding of the Agronomy, Growing a Greener Future. Presented at the Proceedings of 9th Australian Agronomy Conference*, eds D. L. Michalk and J. E. Pratley 403–406. doi: 10.1007/s11033-020-06029-7
- Joo, J., Lee, Y. H., and Song, S. I. (2019). OsbZIP42 is a positive regulator of ABA signaling and confers drought tolerance to rice. *Planta* 249, 1521–1533. doi: 10.1007/s00425-019-03104-7
- Jung, H., Lee, D. K., Do Choi, Y., and Kim, J. K. (2015). OsIAA6, a member of the rice Aux/IAA gene family, is involved in drought tolerance and tiller outgrowth. *Plant Sci.* 236, 304–312. doi: 10.1016/j.plantsci.2015.04.018
- Juntawong, P., and Bailey-Serres, J. (2012). Dynamic light regulation of translation status in *Arabidopsis thaliana*. Front. Plant Sci. 3:66. doi: 10.3389/fpls.2012. 00066
- Kadam, N. N., Yin, X., Bindraban, P. S., Struik, P. C., and Jagadish, K. S. (2015). Does morphological and anatomical plasticity during the vegetative stage make wheat more tolerant of water deficit stress than rice? *Plant Physiol.* 167, 1389–1401. doi: 10.1104/pp.114.253328
- Kanehisa, M., Goto, S., Sato, Y., Furumichi, M., and Tanabe, M. (2012). KEGG for integration and interpretation of large-scale molecular data sets. *Nucleic Acids Res.* 40, 109–114. doi: 10.1093/nar/gkr988
- Kanjoo, V., Punyawaew, K., Siangliw, J. L., Jearakongman, S., Vanavichit, A., and Toojinda, T. (2012). Evaluation of agronomic traits in chromosome segment substitution lines of KDML105 containing drought tolerance QTL under drought stress. *Rice Sci.* 19, 117–124.
- Kato, Y., Abe, J., Kamoshita, A., and Yamagishi, J. (2006). Genotypic variation in root growth angle in rice (*Oryza sativa* L.) and its association with deep root development in upland fields with different water regimes. *Plant Soil*. 287, 117–129. doi: 10.1007/s11104-006-9008-4
- Kawakatsu, T., Teramoto, S., Takayasu, S., Maruyama, N., Nishijima, R., Kitomi, Y., et al. (2020). The transcriptomic landscapes of diverse rice cultivars grown under mild drought conditions. bioRxiv [preprint]. doi: 10.1101/2020.12.11. 421685
- Khakimov, B., Jespersen, B., and Engelsen, S. (2014). Comprehensive and comparative metabolomic profiling of wheat, barley, oat and rye using gas chromatography mass spectrometry and advanced chemometrics. *Foods* 3, 569–585. doi: 10.3390/foods3040569
- Khong, G. N., Pati, P. K., Richaud, F., Parizot, B., Bidzinski, P., Mai, C. D., et al. (2015). OsMADS26 negatively regulates resistance to pathogens and drought tolerance in rice. *Plant Physiol.* 169, 2935–2949. doi: 10.1104/pp.15.01192
- Khowaja, F. S., and Price, A. H. (2008). QTL mapping rolling, stomatal conductance and dimension traits of excised leaves in the Bala × azucena recombinant inbred population of rice. *Field Crops Res.* 106, 248–257.
- Kijoji, A. A., Nchimbi-Msolla, S., Kanyeka, Z. L., Serraj, R., and Henry, A. (2014). Linking root traits and grain yield for rainfed rice in sub-Saharan Africa: response of *Oryza sativa*× *Oryza glaberrima* introgression lines under drought. *Field Crops Res.* 165, 25–35.
- Kim, H., Lee, K., Hwang, H., Yoon, I. S., Kim, D. Y., Kwon, T., et al. (2012). The orthologues of ABA receptors and ABA signaling components in rice. *Plant Stress* 6, 18–28.

Kim, H., Shim, D., Moon, S., Lee, J., Bae, W., Choi, H., et al. (2019). Transcriptional network regulation of the brassinosteroid signaling pathway by the BES1-TPL-HDA19 co-repressor complex. *Planta* 250, 1371–1377. doi: 10.1007/s00425-019-03233-z

- Kim, J. M., To, T. K., Matsui, A., Tanoi, K., Kobayashi, N. I., Matsuda, F., et al. (2017). Acetate-mediated novel survival strategy against drought in plants. *Nat. Plants* 3:17097.
- Kitomi, Y., Kanno, N., Kawai, S., Mizubayashi, T., Fukuoka, S., and Uga, Y. (2015).
 QTLs underlying natural variation of root growth angle among rice cultivars with the same functional allele of deeper rooting. *Rice* 8:16. doi: 10.1186/s12284-015-0049-2
- Klingler, J. P., Batelli, G., and Zhu, J.-K. (2010). ABA receptors: the START of a new paradigm in phytohormone signalling. J. Exp. Bot. 61, 3199–3210. doi: 10.1093/jxb/erq151
- Koller, A., Washburn, M. P., Lange, B. M., Andon, N. L., Deciu, C., Haynes, P. A., et al. (2002). Proteomic survey of metabolic pathways in rice. *Proc. Natl. Acad. Sci. U.S.A.* 99, 11969–11974. doi: 10.1073/pnas.172183199
- Komatsu, S., Kojima, K., Suzuki, K., Ozaki, K., and Higo, K. (2004). Rice proteome database based on two-dimensional polyacrylamide gel electrophoresis: its status in 2003. Nucleic Acids Res. 32, 388–392. doi: 10.1093/nar/gkh020
- Komatsu, S., Muhammad, A., and Rakwal, R. (1999). Separation and characterization of proteins from green and etiolated shoots of rice (*Oryza sativa* L.): towards a rice proteome. *Electrophoresis* 20, 630–636. doi: 10.1002/ (SICI)1522-2683(19990301)20:3<630::AID-ELPS630>3.0.CO;2-Z
- Ku, Y. S., Sintaha, M., Cheung, M. Y., and Lam, H. M. (2018). Plant hormone signaling crosstalks between biotic and abiotic stress responses. *Int. J. Mol. Sci.* 19:3206. doi: 10.3390/ijms19103206
- Kumar, A., Bernier, J., Verulkar, S., Lafitte, H. R., and Atlin, G. N. (2008). Breeding for drought tolerance: direct selection for yield, response to selection and use of drought-tolerant donors in upland and lowland-adapted populations. *Field Crops Res.* 107, 221–231.
- Kumar, M., Kesawat, M. S., Ali, A., Lee, S. C., Gill, S. S., and Kim, H. U. (2019). Integration of abscisic acid signaling with other signaling pathways in plant stress responses and development. *Plants* 8:592. doi: 10.3390/plants8120592
- Kumar, M., Lee, S. C., Kim, J. Y., Kim, S. J., Aye, S. S., and Kim, S. R. (2014). Over-expression of dehydrin gene, OsDhn1, improves drought and salt stress tolerance through scavenging of reactive oxygen species in rice (*Oryza sativa* L.). J. Plant Biol. 57, 383–393. doi: 10.1007/s12374-014-0487-1
- Kumar, R., Bohra, A., Pandey, A. K., Pandey, M. K., and Kumar, A. (2017). Metabolomics for plant improvement: status and prospects. Front. Plant Sci. 8:1302. doi: 10.3389/fpls.2017.01302
- Kusano, M., Redestig, H., Hirai, T., Oikawa, A., and Matsuda, F. (2011). Covering chemical diversity of genetically modified tomatoes using metabolomics for objective substantial equivalence assessment. *PLoS One* 6:e16989. doi: 10.1371/ journal.pone.0016989
- Lanceras, J. C., Pantuwan, G., Jongdee, B., and Toojinda, T. (2004). Quantitative trait loci associated with drought tolerance at reproductive stage in rice. *Plant Physiol.* 135, 384–399. doi: 10.1104/pp.103.035527
- Langridge, P., and Reynolds, M. P. (2015). Genomic tools to assist breeding for drought tolerance. Curr. Opin. Biotechnol. 32, 130–135. doi: 10.1016/j.copbio. 2014.11.027
- Lanzinger, A., Frank, T., Reichenberger, G., Herz, M., and Engel, K.-H. (2015). Metabolite profiling of barley grain subjected to induced drought stress: responses of free amino acids in differently adapted cultivars. J. Agric. Food Chem. 63, 4252–4261. doi: 10.1021/acs.jafc.5b01114
- Lauteri, M., Haworth, M., Serraj, R., Monteverdi, M. C., and Centritto, M. (2014).
 Photosynthetic diffusional constraints affect yield in drought stressed rice cultivars during flowering. PLoS One 9:e109054.
- Lee, H., Cha, J., Choi, C., Choi, N., Ji, H. S., Park, S. R., et al. (2018). Rice WRKY11 plays a role in pathogen defense and drought tolerance. *Rice* 11, 1–12. doi: 10.1186/s12284-018-0199-0
- Lenka, S. K., Katiyar, A., Chinnusamy, V., and Bansal, K. C. (2011). Comparative analysis of drought-responsive transcriptome in Indica rice genotypes with contrasting drought tolerance. *Plant Biotechnol. J.* 9, 315–327. doi: 10.1111/j. 1467-7652.2010.00560.x
- Li, S., Zhao, B., Yuan, D., Duan, M., Qian, Q., Tang, L., et al. (2013). Rice zinc finger protein DST enhances grain production through controlling Gn1a/OsCKX2

- expression. Proc. Natl. Acad. Sci. U.S.A. 110, 3167–3172. doi: 10.1073/pnas. 1300359110
- Li, Y., Xiao, J., Chen, L., Huang, X., Cheng, Z., Han, B., et al. (2018). Rice functional genomics research: past decade and future. *Mol. Plant* 11, 359–380. doi: 10.1016/j.molp.2018.01.007
- Liang, Y., Tabien, R. E., Tarpley, L., Mohammed, A. R., and Septiningsih, E. M. (2021). Transcriptome profiling of two rice genotypes under mild field drought stress during grain-filling stage. AoB Plants 13:lab043. doi: 10.1093/aobpla/ plab043
- Lilley, J. M., and Fukai, S. (1994). Effect of timing and severity of water deficit on four diverse rice cultivars III. phenological development, crop growth and grain yield. Field Crops Res. 37, 225–234. doi: 10.1016/0378-4290(94)90101-5
- Lilley, J. M., Ludlow, M. M., McCouch, S. R., and O'Toole, J. C. (1996). Locating QTL for osmotic adjustment and dehydration tolerance in rice. J. Exp. Bot. 47, 1427–1436. doi: 10.1093/jxb/47.9.1427
- Lipiec, J., Doussan, C., Nosalewicz, A., and Kondracka, K. (2013). Effect of drought and heat stresses on plant growth and yield: a review. *Int. Agrophys.* 27, 463–477. doi: 10.2478/intag-2013-0017
- Lisar, S. Y. S., Motafakkerazad, R., Hossain, M. M., and Rahman, I. M. M. (2012). Water Stress in Plants: Causes, Effects and Responses. Available online at: https://www.intechopen.com/books/water-stress/water-stress-inplants-causes-effects-and-responses (accessed December 5, 2021).
- Liu, H., Zou, G., Liu, G., Hu, S., Li, M., Yu, X., et al. (2005). Correlation analysis and QTL identification for canopy temperature, leaf water potential and spikelet fertility in rice under contrasting moisture regimes. *Chin. Sci. Bull.* 50, 317–326. doi: 10.1360/982004-619
- Liu, J., Shen, J., Xu, Y., Li, X., Xiao, J., and Xiong, L. (2016). Ghd2, a CONSTANS like gene, confers drought sensitivity through regulation of senescence in rice. J. Exp. Bot. 67, 5785–5798. doi: 10.1093/jxb/erw344
- Liu, M. J., Wu, S. H., and Chen, H. M. (2012). Widespread translational control contributes to the regulation of *Arabidopsis photomorphogenesis*. Mol. Syst. Biol. 8:566. doi: 10.1038/msb.2011.97
- Liu, W. Y., Wang, M. M., Huang, J., Tang, H. J., Lan, H. X., and Zhang, H. S. (2009). The OsDHODH1 gene is involved in salt and drought tolerance in rice. J. Integrat. Plant Biol. 51, 825–833. doi: 10.1111/j.1744-7909.2009.00853.x
- Liu, Z., Xin, M., Qin, J., Peng, H., Ni, Z., Yao, Y., et al. (2015). Temporal transcriptome profiling reveals expression partitioning of homeologous genes contributing to heat and drought acclimation in wheat (*Triticum aestivum L.*). BMC Plant Biol. 15, 1–20. doi: 10.1186/s12870-015-0511-8
- Locedie, M., Roven, R. F., Frances, N. B., Jeffery, D., Juan, M. A.-S., Dmytro, C., et al. (2017). Rice SNP-seek database update: new SNPs, indels, and queries. Nucleic Acids Res. 45, D1075–D1081. doi: 10.1093/nar/gkw1135
- Lonbani, M., and Arzani, A. (2011). Morpho-physiological traits associated with terminal drought stress tolerance in triticale and wheat. Agron. Res. 9, 315–329.
- Lou, D., Wang, H., Liang, G., and Yu, D. (2017). OsSAPK2 confers abscisic acid sensitivity and tolerance to drought stress in rice. Front. Plant Sci. 8:993. doi: 10.3389/fpls.2017.00993
- Lou, Q., Chen, L., Mei, H., Wei, H., Feng, F., Wang, P., et al. (2015). Quantitative trait locus mapping of deep rooting by linkage and association analysis in rice. J. Exp. Bot. 66, 4749–4757. doi: 10.1093/jxb/erv246
- Lu, Y., Zhou, D. X., and Zhao, Y. (2020). Understanding epigenomics based on the rice model. *Theor. Appl. Genet.* 133, 1345–1363. doi: 10.1007/s00122-019-03518-7
- Luo, Z., Xiong, J., Xia, H., Ma, X., Gao, M., Wang, L., et al. (2020). Transcriptomic divergence between upland and lowland ecotypes contributes to rice adaptation to a drought-prone agroecosystem. *Evol. Appl.* 13, 2484–2496. doi: 10.1111/eva. 13054
- Ma, X., Xia, H., Liu, Y., Wei, H., Zheng, X., Song, C., et al. (2016). Transcriptomic and metabolomic studies disclose key metabolism pathways contributing to well-maintained photosynthesis under the drought and the consequent drought-tolerance in rice. Front. Plant Sci. 7:1886. doi: 10.3389/fpls.2016.01886
- Ma, Y., Szostkiewicz, I., Korte, A., Moes, D., Yang, Y., Christmann, A., et al. (2009).Regulators of PP2C phosphatase activity function as abscisic acid sensors.Science 324, 1064–1068. doi: 10.1126/science.1172408
- Manavalan, L. P., Chen, X., Clarke, J., Salmeron, J., and Nguyen, H. T. (2012).RNAi-mediated disruption of squalene synthase improves drought tolerance and yield in rice. J. Exp. Bot. 63, 163–175. doi: 10.1093/jxb/err258

Manzoni, C., Kia, D. A., Vandrovcova, J., Hardy, J., Wood, N. W., Lewis, P. A., et al. (2018). Genome, transcriptome and proteome: the rise of omics data and their integration in biomedical sciences. *Brief. Bioinform.* 19, 286–302. doi: 10.1093/bib/bbw114

- Maruyama, K., Urano, K., Yoshiwara, K., Morishita, Y., Sakurai, N., Suzuki, H., et al. (2014). Integrated analysis of the effects of cold and dehydration on rice metabolites, phytohormones, and gene transcripts. *Plant Physiol.* 164, 1759–1771. doi: 10.1104/pp.113.231720
- Matsuura, H., Ishibashi, Y., Shinmyo, A., Kanaya, S., and Kato, K. (2010). Genome-wide analyses of early translational responses to elevated temperature and high salinity in *Arabidopsis thaliana*. *Plant Cell Physiol.* 51, 448–462. doi: 10.1093/pcp/pcq010
- McClung, A. M., Rohila, J. S., Henry, C. G., and Lorence, A. (2020). Responses of U.S. rice cultivars grown under non-flooded irrigation management. *Agronomy* 10:55. doi: 10.3390/agronomy10010055
- Mehana, M., Abdelrahman, M., Emadeldin, Y., Rohila, J. S., and Karthikeyan, R. (2021). Impact of genetic improvements of rice on its water use and effects of climate variability in Egypt. Agriculture 11:865. doi: 10.3390/ agriculture11090865
- Melo, F. V., Oliveira, M. M., Saibo, N. J. M., and Lourenço, T. F. (2021). Modulation of abiotic stress responses in rice by E3-ubiquitin ligases: a promising way to develop stress-tolerant crops. Front. Plant Sci. 12:640193. doi: 10.3389/fpls. 2021.640193
- Mishra, K. K., Vikram, P., Yadaw, R. B., Swamy, B. M., Dixit, S., Cruz, M. T. S., et al. (2013). qDTY 12.1: a locus with a consistent effect on grain yield under drought in rice. BMC Genet. 14:1–10. doi: 10.1186/1471-2156-14-12
- Molina, C., Zaman-Allah, M., Khan, F., Fatnassi, N., Horres, R., Rotter, B., et al. (2011). The salt–responsive transcriptome of chickpea roots and nodules via deep super SAGE. BMC Plant Biol. 11:31. doi: 10.1186/1471-2229-11-31
- Morreel, K., Saeys, Y., Dima, O., Lu, F., Van de Peer, Y., Vanholme, R., et al. (2014). Systematic structural characterization of metabolites in Arabidopsis via candidate substrate-product pair networks. Plant Cell 26, 929–945. doi: 10.1105/tpc.113.122242
- Mumtaz, M. Z., Saqib, M., Abbas, G., Akhtar, J., and Ul-Qamar, Z. (2020). Drought stress impairs grain yield and quality of rice genotypes by impaired photosynthetic attributes and K nutrition. *Rice Sci.* 27, 5–9. doi: 10.1016/j.rsci. 2019.12.001
- Muthurajan, R., Shobbar, Z.-S., Jagadish, S. V. K., Bruskiewich, R., Ismail, A., Leung, H., et al. (2010). Physiological and proteomic responses of rice peduncles to drought stress. *Mol. Biotechnol.* 48, 173–182. doi: 10.1007/s12033-010-9 358-2
- Nakabayashi, R., Yonekura-Sakakibara, K., Urano, K., Suzuki, M., Yamada, Y., Nishizawa, T., et al. (2014). Enhancement of oxidative and drought tolerance in arabidopsis by overaccumulation of antioxidant flavonoids. *Plant J.* 77, 367–379. doi: 10.1111/tpj.12388
- Nam, K. H., Park, K. W., Han, S. M., Kim, S. W., Lee, J. H., and Kim, C. G. (2016). Compositional analysis of protoporphyrinogen oxidase-inhibiting herbicide-tolerant rice and conventional rice. *Int. J. Food Sci. Technol.* 51, 1010–1017. doi: 10.1111/ijfs.13060
- Ndjiondjop, M. N., Cissé, F., Futakuchi, K., Lorieux, M., Manneh, B., Bocco, R., et al. (2010). "Effect of drought on rice (*Oryza* spp.) genotypes according to their drought tolerance level," in *Proceedings of the Second Africa Rice Congress, Bamako, Mali, 22–26 March 2010: Innovation and Partnerships to Realize Africa's Rice Potential* (Bamako: Africarice), 5–7.
- Nguyen, H. T., Babu, R. C., and Blum, A. (1997). Breeding for drought tolerance in rice: physiology and molecular genetics considerations. *Crop Sci.* 37, 1426–1434. doi: 10.2135/cropsci1997.0011183x003700050002x
- Ning, T., Hua, Z., Xianghua, L., Jinghua, X., and Lizhong, X. (2012). Constitutive activation of transcription factor OsbZIP46 improves drought tolerance in rice. *Plant Physiol.* 158, 1755–1768. doi: 10.1104/pp.111.190389
- Noemi, D. P., Darryl, H. B. C., and Christine, L. (2014). Chichioco-hernandez, anti-ulcer activity of leguminosae plants. Arq. Gastroenterol. 51:1. doi: 10.1590/ s0004-28032014000100013
- Norton, G. J., and Price, A. H. (2009). Mapping of quantitative trait loci for seminal root morphology and gravitropic response in rice. *Euphytica* 166, 229–237. doi: 10.1007/s10681-008-9833-z

Nozu, Y., Tsugita, A., and Kamijo, K. (2006). Proteomic analysis of rice leaf, stem and root tissues during growth course. *Proteomics* 6, 3665–3670. doi: 10.1002/ pmic.200600043

- Oh, S. J., Song, S. I., Kim, Y. S., Jang, H. J., Kim, S. Y., Kim, M., et al. (2005). Arabidopsis CBF3/DREB1A and ABF3 in transgenic rice increased tolerance to abiotic stress without stunting growth. *Plant Physiol.* 138, 341–351. doi: 10.1104/pp.104.059147
- Oliver, S. G., Winson, M. K., Kell, D. B., and Baganz, F. (1998). Systematic functional analysis of the yeast genome. *Trends Biotechnol.* 16, 373–378. doi: 10.1016/s0167-7799(98)01214-1
- Palaniswamy, S. K., James, S., Sun, H., Lamb, R. S., Davuluri, R. V., and Grotewold, E. (2006). AGRIS and AtRegNet: a platform to link cis regulatory elements and transcription factors into regulatory networks. *Plant Physiol.* 140, 818–829. doi: 10.1104/pp.105.072280
- Palanog, A. D., Swamy, B. M., Shamsudin, N. A. A., Dixit, S., Hernandez, J. E., Boromeo, T. H., et al. (2014). Grain yield QTLs with consistent-effect under reproductive-stage drought stress in rice. Field Crops Res. 161, 46–54. doi: 10.1016/j.fcr.2014.01.004
- Panda, D., Mishra, S. S., and Behera, P. K. (2021). Drought tolerance in rice: focus on recent mechanisms and approaches. *Rice Sci.* 28, 119–132. doi: 10.1016/j.rsci. 2021.01.002
- Pantuwan, G., Fukai, S., Cooper, M., Rajatasereekul, S., and O'Toole, J. C. (2002). Yield response of rice (*Oryza sativa* L.) genotypes to drought under rainfed lowlands: 2. selection of drought resistant genotypes. *Field Crops Res.* 73, 169–180. doi: 10.1016/s0378-4290(01)00195-2
- Pantuwan, G., Ingram, K., and Sharma, P. (1996). "Rice root system development under rainfed conditions," in *Proceedings of the Thematic Conference on Stress Physiology, Rainfed Lowland Rice Research Consortium* (Manila, Philippines, Lucknow: International Rice Research Centre), 198–206.
- Park, S.-Y., Fung, P., Nishimura, N., Jensen, D. R., Fujii, H., Zhao, Y., et al. (2009).
 Abscisic acid inhibits type 2C protein phosphatases via the PYR/PYL family of START proteins. Science 324, 1068–1071. doi: 10.1126/science.1173041
- Paul, S., Gayen, D., Datta, S. K., and Datta, K. (2015). Dissecting root proteome of transgenic rice cultivars unravels metabolic alterations and accumulation of novel stress responsive proteins under drought stress. *Plant Sci.* 234, 133–143. doi: 10.1016/j.plantsci.2015.02.006
- Pazhamala, L. T., Kudapa, H., Weckwerth, W., Millar, A. H., and Varshney, R. K. (2021). Systems biology for crop improvement. *Plant Genome*. 14:e20098. doi: 10.1002/tpg2.20098
- Phung, T. H., Jung, H. I., Park, J. H., Kim, J. G., Back, K., and Jung, S. (2011). Porphyrin biosynthesis control under water stress: sustained porphyrin status correlates with drought tolerance in transgenic rice. *Plant Physiol.* 157, 1746– 1764. doi: 10.1104/pp.111.188276
- Pires, M. V., Pereira, A. A. Jr., Medeiros, D. B., Daloso, D. M., Pham, P. A., Barros, K. A., et al. (2016). The influence of alternative pathways of respiration that utilize branched-chain amino acids following water shortage in arabidopsis. Plant Cell Environ. 39, 1304–1319. doi: 10.1111/pce.12682
- Potopova, V., Boroneant, C., Boincean, B., and Soukup, J. (2016). Impact of agricultural drought on main crop yields in the republic of moldova. *Int. J. Climatol.* 36, 2063–2082. doi: 10.1002/joc.4481
- Prasad, R., Shivay, Y. S., and Kumar, D. (2017). Current Status, Challenges, and Opportunities in Rice Production. In Rice Production Worldwide. Cham: Springer, 1–32.
- Prince, S. J., Beena, R., Gomez, S. M., Senthivel, S., and Babu, R. C. (2015). Mapping consistent rice (*Oryza sativa* L.) yield QTLs under drought stress in target rainfed environments. *Rice* 8, 1–13. doi: 10.1186/s12284-015-0053-6
- Puri, R. R., Khadka, K., and Paudyal, A. (2010). Separating climate resilient crops through screening of drought tolerant rice land races in Nepal. Aronomy J. Nepal. 1, 80–84. doi: 10.3126/ajn.v1i0.7546
- Rakwal, R., and Agrawal, G. K. (2003). Rice proteomics: current status and future perspectives. Electrophoresis 24, 3378–3389. doi: 10.1002/elps.200305586
- Ramalingam, A., Kudapa, H., Pazhamala, L. T., Weckwerth, W., and Varshney, R. K. (2015). Proteomics and metabolomics: two emerging areas for legume improvement. Front. Plant Sci. 6:1116. doi: 10.3389/fpls.2015.01116
- Raorane, M. L., Pabuayon, I. M., Varadarajan, A. R., Mutte, S. K., Kumar, A., Treumann, A., et al. (2015). Proteomic insights into the role of the large-effect QTL qDTY 12.1 for rice yield under drought. *Mol. Breed.* 35, 1–14.

Ray, J. D., Yu, L., McCouch, S. R., Champoux, M. C., Wang, G., and Nguyen, H. T. (1996). Mapping quantitative trait loci associated with root penetration ability in rice (*Oryza sativa L.*). Theor. Appl. Genet. 92, 627–636. doi: 10.1007/ BF00226082

- Ray, S., Dansana, P. K., Giri, J., Deveshwar, P., Arora, R., Agarwal, P., et al. (2011). Modulation of transcription factor and metabolic pathway genes in response to water-deficit stress in rice. *Funct. Integr. Genom.* 11, 157–178. doi: 10.1007/ s10142-010-0187-v
- Redillas, M. C. F. R., Jeong, J. S., Kim, Y., Jung, H., Bang, S. W., Choi, Y. D., et al. (2012). The overexpression of OsNAC9 alters the root architecture of rice plants enhancing drought tolerance and grain yield under field conditions. *Plant Biotechnol. J.* 10, 792–805. doi: 10.1111/j.1467-7652.2012.00697.x
- Rohila, J. S., and Yang, Y. (2007). Rice mitogen-activated protein kinase gene family and its role in biotic and abiotic stress response. J. Integrat. Plant Biol. 49, 751–759. doi: 10.1111/j.1744-7909.2007.00501.x
- Rollins, J. A., Habte, E., Templer, S. E., Colby, T., Schmidt, J., and Von Korff, M. (2013). Leaf proteome alterations in the context of physiological and morphological responses to drought and heat stress in barley (*Hordeum vulgare* L.). J. Exp. Bot. 64, 3201–3212. doi: 10.1093/jxb/ert158
- Ryan, D., and Robards, K. (2006). Metabolomics: the greatest omics of them all? Analy. Chem. 78, 7954–7958. doi: 10.1021/ac0614341
- Sah, S. K., Reddy, K. R., and Li, J. (2016). Abscisic acid and abiotic stress tolerance in crop plants. Front. Plant Sci. 7:571. doi: 10.3389/fpls.2016.00571
- Saha, P., Sade, N., Arzani, A., Rubio Wilhelmi, M. D. M., Coe, K. M., Li, B., et al. (2016). Effects of abiotic stress on physiological plasticity and water use of *Setaria viridis* (L.). *Plant Sci.* 251, 128–138. doi: 10.1016/j.plantsci.2016. 06.011
- Sahebi, M., Hanafi, M. M., Rafii, M. Y., Mahmud, T., Azizi, P., Osman, M., et al. (2018). Improvement of drought tolerance in rice (*Oryza sativa* L.): genetics, genomic tools, and the WRKY gene family. *BioMed. Res. Int.* 2018;3158474. doi: 10.1155/2018/3158474
- Saikumar, S., Gouda, P. K., Saiharini, A., Varma, C. M. K., Vineesha, O., Padmavathi, G., et al. (2014). Major QTL for enhancing rice grain yield under lowland reproductive drought stress identified using an O. sativa/O. glaberrima introgression line. Field Crops Res. 163, 119–131. doi: 10.1016/j.fcr.2014.03.011
- Saini, A., Rohila, J. S., Govindan, G., Li, Y. F., and Sunkar, R. (2021). Splice variants of superoxide dismutases in rice and their expression profiles under abiotic stresses. *BioMed Res. Int.* 22:3997. doi: 10.3390/ijms22083997
- Salazar, C., Hernández, C., and Pino, M. T. (2015). Plant water stress: associations between ethylene and abscisic acid response. *Chilean J. Agric. Res.* 75, 71–79. doi: 10.4067/s0718-58392015000300008
- Samal, A. C., Bhattacharya, P., Biswas, P., Maity, J. P., Bundschuh, J., and Santra, S. C. (2021). Variety-specific arsenic accumulation in 44 different rice cultivars (O. sativa L.) and human health risks due to co-exposure of arseniccontaminated rice and drinking water. J. Hazardous Mater. 407:124804. doi: 10.1016/j.jhazmat.2020.124804
- Sandhu, N., Raman, K. A., Torres, R. O., Audebert, A., Dardou, A., Kumar, A., et al. (2016). Rice root architectural plasticity traits and genetic regions for adaptability to variable cultivation and stress conditions. *Plant Physiol.* 171, 2562–2576. doi: 10.1104/pp.16.00705
- Sandhu, N., Singh, A., Dixit, S., Cruz, M. T. S., Maturan, P. C., Jain, R. K., et al. (2014). Identification and mapping of stable QTL with main and epistasis effect on rice grain yield under upland drought stress. *BMC Genet.* 15:63. doi: 10.1186/1471-2156-15-63
- Shahid, S. (2019). To be or not to be pathogenic: transcriptional reprogramming dictates a fungal pathogen's response to different hosts. *Plant Cell* 32:289. doi: 10.1105/tpc.19.00976
- Shaibu, Y., Mloza Banda, H., Makwiza, C., and Malunga, J. (2015). Grain yield performance of upland and lowland rice varieties under water saving irrigation through alternate wetting and drying in sandy clay loams of southern Malawi. Exp. Agric. 51, 313–326. doi: 10.1017/S0014479714000325
- Sherry, S. T., Ward, M. H., Kholodov, M., Baker, J., Phan, L., Smigielski, E. M., et al. (2001). dbSNP: the NCBI database of genetic variation. *Nucleic Acids Res.* 29, 308–311. doi: 10.1093/nar/29.1.308
- Shi, F., Dong, Y., Wang, M., and Qiu, D. (2020). Transcriptomics analyses reveal that OsMIOX improves rice drought tolerance by regulating the expression of plant hormone and sugar related genes. *Plant Biotechnol. Rep.* 14, 607–617.

- Shim, J. S., Oh, N., Chung, P. J., Kim, Y. S., Choi, Y. D., and Kim, J. K. (2018). Overexpression of OsNAC14 improves drought tolerance in rice. Front. Plant Sci. 9:310. doi: 10.3389/fpls.2018.00310
- Sié, M., Futakuchi, K., Mande, H., Manneh, B., Ndjiondjop, M. N., Efisue, A., et al. (2008). "Drought research at WARDA: current situation and prospects," in *Drought Frontiers in Rice: Crop Improvement for Increased Rainfed Production*, eds R. Serraj, J. Bennett, and B. Hardy (Los Baños, Singapore/Philippines: World Scientific Publishing, International Rice Research Institute).
- Singh, B. P., Jayaswal, P. K., Singh, B., Singh, P. K., Kumar, V., Mishra, S., et al. (2015). Natural allelic diversity in OsDREB1F gene in the Indian wild rice germplasm led to ascertain its association with drought tolerance. *Plant Cell Rep.* 34, 993–1004. doi: 10.1007/s00299-015-1760-6
- Singh, N. K., Raghuvanshi, S., Srivastava, S. K., Gaur, A., Pal, A. K., Dalal, V., et al. (2004). Sequence analysis of the long arm of rice chromosome 11 for rice-wheat synteny. Funct. Integrat. Genom. 4, 102–117. doi: 10.1007/s10142-004-0109-y
- Singh, R. K., Sood, P., Prasad, A., and Prasad, M. (2021). Advances in omics technology for improving crop yield and stress resilience. *Plant Breed.* 140, 719–731.
- Sirichandra, C., Gu, D., Hu, H.-C., Davanture, M., Lee, S., Djaoui, M., et al. (2009). Phosphorylation of the arabidopsis atrbohf NADPH oxidase by OST1 protein kinase. FEBS Lett. 583, 2982–2986. doi: 10.1016/j.febslet.2009.08.033
- Sormani, R., Delannoy, E., Lageix, S., Bitton, F., Lanet, E., Saez-Vasquez, J., et al. (2011). Sublethal cadmium intoxication in *Arabidopsis thaliana* impacts translation at multiple levels. *Plant Cell Physiol.* 52, 436–447. doi: 10.1093/pcp/pcr001
- Suji, K. K., and John, A. (2010). An epigenetic change in rice cultivars underwater stress conditions. *Electr. J. Plant Breed.* 1, 1142–1143.
- Swamy, B. P. M., Ahmed, H. U., Henry, A., Mauleon, R., Dixit, S., Vikram, P., et al. (2013). Genetic, physiological, and gene expression analyses reveal that multiple QTL enhance yield of rice mega-variety IR64 under drought. *PLoS One* 8:e62795. doi: 10.1371/journal.pone.0062795
- Swamy, B. P. M., Kaladhar, K., Anuradha, K., Batchu, A. K., Longvah, T., and Sarla, N. (2018). QTL analysis for grain iron and zinc concentrations in two O. nivara derived backcross populations. *Rice Sci.* 25, 197–207. doi: 10.1016/j.rsci.2018. 06.003
- Swamy, B. P. M., Vikram, P., Dixit, S., Ahmed, H. U., and Kumar, A. (2011). Metaanalysis of grain yield QTL identified during agricultural drought in grasses showed consensus. BMC Genom. 12:319. doi: 10.1186/1471-2164-12-319
- Tanaka, N., Mitsui, S., Nobori, H., Yanagi, K., and Komatsu, S. (2005). Expression and function of proteins during development of the basal region in rice seedlings. Mol. Cell. Prot. 4, 796–808. doi: 10.1074/mcp.M400211-MCP200
- Tansley, T. R., and Shechter, I. (2001). Squalene synthase: structure and regulation. Prog. Nucleic Acid Res. Mol. Biol. 65, 157–195.
- Tarazon, P., Feussner, K., and Feussner, I. (2015). An enhanced plant lipidomics method based on multiplexed liquid chromatography-mass spectrometry reveals additional insights into cold- and drought-induced membrane remodeling. Plant J. 84, 621–633. doi: 10.1111/tpj.13013
- Tarun, J. A., Mauleon, R., Arbelaez, J. D., Catausan, S., Dixit, S., Kumar, A., et al. (2020). Comparative transcriptomics and co-expression networks reveal tissueand genotype-specific responses of qDTYs to reproductive-stage drought stress in rice (*Oryza sativa* L.). *Genes* 11:1124. doi: 10.3390/genes11101124
- Tello-Ruiz, M. K., Stein, J., Wei, S., Preece, J., Olson, A., Naithani, S., et al. (2016).
 Comparative plant genomics and pathway resources. *Nucleic Acids Res.* 44, 1133–1140.
- The IC4R Consortium. (2015). Information commons for rice (IC4R). Nucleic Acids Res. 44, 1172–1180.
- Thomashow, M. F. (1999). Plant cold acclimation: freezing tolerance genes and regulatory mechanisms. Ann. Rev. Plant Physiol. Plant Mol. Biol. 50, 571–599. doi: 10.1146/annurev.arplant.50.1.571
- Tian, X. J., Long, Y., Wang, J., Zhang, J. W., Wang, Y. Y., Li, W. M., et al. (2015). De novo transcriptome assembly of common wild rice (*Oryza rufipogon* griff.) and discovery of drought-response genes in root tissue based on transcriptomic data. *PLoS One* 10:e0131455. doi: 10.1371/journal.pone.0131455
- Tiwari, P., Srivastava, D., Chauhan, A. S., Indoliya, Y., Singh, P. K., Tiwari, S., et al. (2021). Root system architecture, physiological analysis and dynamic transcriptomics unravel the drought-responsive traits in rice genotypes. *Ecotoxicol. Environ. Safety* 207:111252. doi: 10.1016/j.ecoenv.2020.111252

Trijatmiko, K., Supriyanta, R., Prasetiyono, J., Thomson, M. J., Vera Cruz, C. M., Moeljopawiro, S., et al. (2014). Meta-analysis of quantitative trait loci for grain yield and component traits under reproductive-stage drought stress in an upland rice population. *Mol. Breed.* 34, 283–295. doi: 10.1007/s11032-013-0012-0

- Uga, Y., Hanzawa, E., Nagai, S., Sasaki, K., Yano, M., and Sato, T. (2012).
 Identification of qSOR1, a major rice QTL involved in soil-surface rooting in paddy fields. *Theor. Appl. Genet.* 124, 75–86. doi: 10.1007/s00122-011-1688-3
- Uga, Y., Kitomi, Y., Yamamoto, E., Kanno, N., Kawai, S., Mizubayashi, T., et al. (2015). A QTL for root growth angle on rice chromosome 7 is involved in the genetic pathway of deeper rooting. *Rice* 8:8. doi: 10.1186/s12284-015-0044-7
- Uga, Y., Okuno, K., and Yano, M. (2011). Dro1, a major QTL involved in deep rooting of rice under upland field conditions. J. Exp. Bot. 62, 2485–2494. doi: 10.1093/jxb/erq429
- Uga, Y., Sugimoto, K., Ogawa, S., Rane, J., Ishitani, M., Hara, N., et al. (2013a). Control of root system architecture by DEEPER ROOTING 1 increases rice yield under drought conditions. *Nat. Genet.* 45, 1097–1102. doi: 10.1038/ng. 2725
- Uga, Y., Yamamoto, E., Kanno, N., Kawai, S., Mizubayashi, T., and Fukuoka, S. (2013b). A major QTL controlling deep rooting on rice chromosome 4. Sci. Rep. 3:3040. doi: 10.1038/srep03040
- Ullah, N., Yüce, M., Gökçe, Z. N. Ö, and Budak, H. (2017). Comparative metabolite profiling of drought stress in roots and leaves of seven triticeae species. BMC Genom. 18:969. doi: 10.1186/s12864-017-4321-2
- Upadhyaya, H., and Panda, S. K. (2019). "Drought stress responses and its management in rice," in Advances in Rice Research for Abiotic Stress Tolerance, eds M. Hasanuzzaman, M. Fujita, K. Nahar, and J. K. Biswas (Elsevier), 177–200. doi: 10.1016/b978-0-12-814332-2.00009-5
- Vanniarajan, C., Vinod, K. K., and Pereira, A. (2012). Molecular evaluation of genetic diversity and association studies in rice (*Oryza sativa L.*). J. Genet. 91, 9–19. doi: 10.1007/s12041-012-0146-6
- Venuprasad, R., Bool, M. E., Quiatchon, L., and Atlin, G. N. (2012a). A QTL for rice grain yield in aerobic environments with large effects in three genetic backgrounds. *Theor. Appl. Genet.* 124, 323–332. doi: 10.1007/s00122-011-1707-4
- Venuprasad, R., Bool, M. E., Quiatchon, L., Cruz, M. T. S., Amante, M., and Atlin, G. N. (2012b). A large-effect QTL for rice grain yield under upland drought stress on chromosome 1. *Mol. Breed.* 30, 535–547.
- Venuprasad, R., Dalid, C. O., Del Valle, M., Zhao, D., Espiritu, M., Sta Cruz, M. T., et al. (2009). Identification and characterization of large-effect quantitative trait loci for grain yield under lowland drought stress in rice using bulk-segregant analysis. *Theor. Appl. Genet.* 120, 177–190. doi: 10.1007/s00122-009-1168-1
- Verma, S. K., Saxena, R. R., Saxena, R. R., Xalxo, M. S., and Verulkar, S. B. (2014).
 QTL for grain yield under water stress and non-stress conditions over years in rice (Oryza sativa L.). Austral. J. Crop Sci. 8, 916–926.
- Vikram, P., Kumar, A., Singh, A. K., and Singh, N. K. (2012). "Rice: genomics-assisted breeding for drought tolerance," in *Improving Crop Tolerance to Abiotic Stress*, eds S. S. Tuteja, A. F. Gill, and R. Tiburico (Germany: Wiley-VCH Verlag GmbH & Co. KGaA), 715–731.
- Vikram, P., Swamy, B. P. M., Dixit, S., Sta Cruz, M. T., Ahmed, H. U., Singh, A. K., et al. (2011). qDTY1.1, a major QTL for rice grain yield under reproductive-stage drought stress with a consistent effect in multiple elite genetic backgrounds. BMC Genet. 12:89. doi: 10.1186/1471-2156-12-89
- Wade, L. J., McLaren, C., Regmi, K., and Sarkarung, S. (1996). "The importance of site characterization for understanding genotype by environment interactions," in *Plant Adaptation and Crop Improvement*, eds M. Cooper and G. L. Hammer (Wallingford: CABI), 549–562.
- Walia, H., Wilson, C., Condamine, P., Liu, X., Ismail, A. M., Zeng, L., et al. (2005). Comparative transcriptional profiling of two contrasting rice genotypes under salinity stress during the vegetative growth stage. *Plant Physiol.* 139, 822–835. doi: 10.1104/pp.105.065961
- Wan, X. Y., and Liu, J. Y. (2008). Comparative proteomics analysis reveals an intimate protein network provoked by hydrogen peroxide stress in rice seedling leaves. Mol. Cell. Proteomics 7, 1469–1488. doi: 10.1074/mcp. M700488-MCP200

Wang, H., Wang, H., Shao, H., and Tang, X. (2016). Recent advances in utilizing transcription factors to improve plant abiotic stress tolerance by transgenic technology. Front. Plant Sci. 7:67. doi: 10.3389/fpls.2016.00067

- Wang, J., Yang, J., Jia, Q., Zhu, J., Shang, Y., Hua, W., et al. (2014). A new QTL for plant height in barley (*Hordeum vulgare* L.) showing no negative effects on grain yield. *PLos One* 9:e90144. doi: 10.1371/journal.pone.0090144
- Wang, S., Alseekh, S., Fernie, A. R., and Luo, J. (2019). The structure and function of major plant metabolite modifications. *Mol. Plant* 12, 899–919. doi: 10.1016/j.molp.2019.06.001
- Wang, W.-S., Pan, Y.-J., Zhao, X.-Q., Dwivedi, D., Zhu, L.-H., Ali, J., et al. (2011). Drought-induced site-specific DNA methylation and its association with drought tolerance in rice (*Oryza sativa L.*). J. Exp. Bot. 62, 1951–1960. doi: 10.1093/jxb/erq391
- Wang, X., Samo, N., Li, L., Wang, M., Qadir, M., Jiang, K., et al. (2019). Root distribution and its impacts on the drought tolerance capacity of hybrid rice in the sichuan basin area of China. Agronomy 9:79. doi: 10.3390/ agronomy9020079
- Wang, Y., Impa, S. M., Sunkar, R., and Jagadish, S. V. K. (2021). The neglected other half - role of the pistil in plant heat stress responses. *Plant Cell Environ*. 44, 2200–2210. doi: 10.1111/pce.14067
- Wang, Y., Xu, C., Zhang, B., Wu, M., and Chen, G. (2017). Physiological and proteomic analysis of rice (*Oryza sativa* L.) in flag leaf during flowering stage and milk stage under drought stress. *Plant Growth Regulat*. 82, 201–218. doi: 10.1007/s10725-017-0252-9
- Weckwerth, W., Ghatak, A., Bellaire, A., Chaturvedi, P., and Varshney, R. K. (2020). PANOMICS meets germplasm. *Plant Biotechnol. J.* 18, 1507–1525. doi: 10.1111/pbi.13372
- Wei, S., Hu, W., Deng, X., Zhang, Y., Liu, X., Zhao, X., et al. (2014). A rice calcium-dependent protein kinase OsCPK9 positively regulates drought stress tolerance and spikelet fertility. BMC Plant Biol. 14:1–13. doi: 10.1186/1471-2229-14-133
- Wu, M. S., Ding, X., Fu, X., and Lozano-Duran, R. (2019). Transcriptional reprogramming caused by the geminivirus tomato yellow leaf curl virus in local or systemic infections in nicotiana benthamiana. *BMC Genom.* 20:542. doi: 10.1186/s12864-019-5842-7
- Wu, X., Shiroto, Y., Kishitani, S., Ito, Y., and Toriyama, K. (2009). Enhanced heat and drought tolerance in transgenic rice seedlings overexpressing OsWRKY11 under the control of HSP101 promoter. *Plant Cell Rep.* 28, 21–30. doi: 10.1007/ s00299-008-0614-x
- Wu, Y., Mirzaei, M., Pascovici, D., Chick, J. M., Atwell, B. J., and Haynes, P. A. (2016). Quantitative proteomic analysis of two different rice varieties reveals that drought tolerance is correlated with reduced abundance of photosynthetic machinery and increased abundance of ClpD1 protease. J. Proteomics 143, 73–82.
- Xia, H., Ma, X., Xu, K., Wang, L., Liu, H., Chen, L., et al. (2020). Temporal transcriptomic differences between tolerant and susceptible genotypes contribute to rice drought tolerance. BMC Geno. 21:1–18. doi: 10.1186/s12864-020-07193-7
- Xiao, B. Z., Chen, X., Xiang, C. B., Tang, N., Zhang, Q. F., and Xiong, L. Z. (2009). Evaluation of seven function-known candidate genes for their effects on improving drought tolerance of transgenic rice under field conditions. *Mol. Plant* 2, 73–83. doi: 10.1093/mp/ssn068
- Xiong, H., Yu, J., Miao, J., Li, J., Zhang, H., Wang, X., et al. (2018). Natural variation in OsLG3 increases drought tolerance in rice by inducing ROS scavenging. *Plant Physiol.* 178, 451–467. doi: 10.1104/pp.17.01492
- Xiong, L., and Yang, Y. (2003). Disease tolerance and abiotic stress tolerance in rice are inversely modulated by an abscisic acid-inducible mitogen-activated protein kinase. *Plant Cell* 15, 745–759. doi: 10.1105/tpc.008714
- Xiong, Q., Cao, C., Shen, T., Zhong, L., He, H., and Chen, X. (2019). Comprehensive metabolomic and proteomic analysis in biochemical metabolic pathways of rice spikes under drought and submergence stress. *Biochim. Biophys. Acta Proteins Proteom.* 1867, 237–247. doi: 10.1016/j.bbapap.2019.01.
- Xu, D. Q., Huang, J., Guo, S. Q., Yang, X., Bao, Y. M., Tang, H. J., et al. (2008). Overexpression of a TFIIIA-type zinc finger protein gene ZFP252 enhances drought and salt tolerance in rice (*Oryza sativa L.*). FEBS Lett. 582, 1037–1043. doi: 10.1016/j.febslet.2008.02.052

Yadav, S., Sandhu, N., Singh, V. K., Catolos, M., and Kumar, A. (2019). Genotypingby-sequencing based QTL mapping for rice grain yield under reproductive stage drought stress tolerance. Sci. Rep. 9:14326. doi: 10.1038/s41598-019-50880-z

- Yadaw, R. B., Dixit, S., Raman, A., Mishra, K. K., Vikram, P., Swamy, B. M., et al. (2013). A QTL for high grain yield under lowland drought in the background of popular rice variety sabitri from nepal. *Field Crops Res.* 144, 281–287.
- Yamaguchi-Shinozaki, K., and Shinozaki, K. (2005). Organization of cis acting regulatory elements in osmotic- and cold-stress-responsive promoters. *Trends Plant Sci.* 10, 88–94. doi: 10.1016/j.tplants.2004.12.012
- Yang, J. C., Zhang, H., and Zhang, J. H. (2012). Root morphology and physiology in relation to the yield formation of rice. J. Integrat. Agric. 11, 920–926. doi: 10.1016/s2095-3119(12)60082-3
- Yang, S., Vanderbeld, B., Jiangxin, W., and Huang, Y. (2010). Narrowing down the targets: towards successful genetic engineering of drought-tolerant crops. *Mol. Plant* 3, 469–490. doi: 10.1093/mp/ssq016
- Yang, X., Wang, B., Chen, L., Li, P., and Cao, C. (2019). The different influences of drought stress at the flowering stage on rice physiological traits, grain yield, and quality. Sci. Rep. 9:3742. doi: 10.1038/s41598-019-40161-0
- Yang, Y., Saand, M. A., Huang, L., Abdelaal, W. B., Zhang, J., Wu, Y., et al. (2021). Applications of multi-omics technologies for crop improvement. Front. Plant Sci. 12:563953. doi: 10.3389/fpls.2021.563953
- Yin, X. M., Huang, L. F., Zhang, X., Wang, M. L., Xu, G. Y., and Xia, X. J. (2015). OsCML4 improves drought tolerance through scavenging of reactive oxygen species in rice. J. Plant Biol. 58, 68–73. doi: 10.1016/j.plantsci.2012.08.003
- Yoshida, H., and Nagato, Y. (2011). Flower development in rice. *J. Exp. Bot.* 62, 4719–4730. doi: 10.1093/jxb/err272
- You, J., Zong, W., Li, X., Ning, J., Hu, H., Li, X., et al. (2013). The SNAC1-targeted gene OsSRO1c modulates stomatal closure and oxidative stress tolerance by regulating hydrogen peroxide in rice. J. Exp. Bot. 64, 569–583. doi: 10.1093/ jxb/ers349
- Yu, J., Lai, Y., Wu, X., Wu, G., and Guo, C. (2016). Overexpression of OsEm1 encoding a group I LEA protein confers enhanced drought tolerance in rice. Biochem. Biophys. Res. Commun. 478, 703–709. doi: 10.1016/j.bbrc.2016.08.010
- Yu, L., Chen, X., Wang, Z., Wang, S., Wang, Y., Zhu, Q., et al. (2013). Arabidopsis enhanced drought tolerance1/HOMEODOMAIN GLABROUS11 confers drought tolerance in transgenic rice without yield penalty. *Plant Physiol*. 162, 1378–1391. doi: 10.1104/pp.113.217596
- Yu, L., Liu, S., Yu, L., Xiao, Y., Zhang, S., Wang, X., et al. (2021). RNA demethylation increases the yield and biomass of rice and potato plants in field trials. *Nat. Biotechnol.* 39, 1581–1588. doi: 10.1038/s41587-021-00982-9
- Zhan, A., Schneider, H., and Lynch, J. (2015). Reduced lateral root branching density improves drought tolerance in maize. *Plant Physiol.* 168, 1603–1615. doi:10.1104/pp.15.00187
- Zhang, D., and Yuan, Z. (2014). Molecular control of grass inflorescence development. Annu. Rev. Plant Biol. 65, 553–578. doi: 10.1146/annurevarplant-050213-040104
- Zhang, J., Zheng, H. G., Aarti, A., Pantuwan, G., Nguyen, T. T., Tripathy, J. N., et al. (2001). Locating genomic regions associated with components of drought tolerance in rice: comparative mapping within and across species. *Theor. Appl. Genet.* 103, 19–29. doi: 10.1007/s001220000534

- Zhao, H., Yao, W., Ouyang, Y., Yang, W., Wang, G., Lian, X., et al. (2015).
 RiceVarMap: a comprehensive database of rice genomic variations. *Nucleic Acids Res.* 43, 1018–1022. doi: 10.1093/nar/gku894
- Zheng, T., Yu, H., Zhang, H., Wu, Z., Wang, W., and Tai, S. (2015). Rice functional genomics and breeding database (RFGB)-3K-rice SNP and InDel sub-database. *Chinese Sci. Bull.* 60, 367. doi: 10.1360/n972014-01231
- Zheng, X., Chen, B., Lu, G., and Han, B. (2009). Overexpression of a NAC transcription factor enhances rice drought and salt tolerance. *Biochem Biophys Res. Commun.* 379, 985–989. doi: 10.1016/j.bbrc.2008.12.163
- Zhou, L., Liu, Z., Liu, Y., Kong, D., Li, T., Yu, S., et al. (2016). A novel gene OsAHL1 improves both drought avoidance and drought tolerance in rice. *Sci. Rep.* 6:30264. doi: 10.1038/srep30264
- Zhou, S., Zhu, M., Wang, F., Huang, J., and Wang, G. (2013). Mapping of QTLs for yield and its components in a rice recombinant inbred line population. *Pakistan I. Bot.* 45, 183–189.
- Zinolabedin, T. S., Hemmatollah, P., Seyed, A. M. M., and Hamidreza, B. (2008). Study of water stress effects in different growth stages on yield and yield components of different rice (*Oryza sativa* L.) cultivars. *Pakistan J. Biol. Sci.* 11, 1303–1309. doi: 10.3923/pjbs.2008.1303.1309
- Zivy, M., Wienkoop, S., Renaut, J., Pinheiro, C., Goulas, E., and Carpentier, S. (2015). The quest for tolerant varieties: the importance of integrating "omics" techniques to phenotyping. Front. Plant Sci. 6:448. doi: 10.3389/fpls.2015.0 0448
- Zou, J. J., Wei, F. J., Wang, C., Wu, J. J., Ratnasekera, D., Liu, W. X., et al. (2010). Arabidopsis calcium-dependent protein kinase CPK10 functions in abscisic acid-and Ca2+-mediated stomatal regulation in response to drought stress. Plant Physiol. 154, 1232–1243. doi: 10.1104/pp.110.157545
- Zu, X., Lu, Y., Wang, Q., Chu, P., Miao, W., Wang, H., et al. (2017). A new method for evaluating the drought tolerance of upland rice cultivars. *Crop J.* 5, 488–498. doi: 10.3390/genes11090994

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The handling editor declared a past co-authorship with one of the authors RR.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zargar, Mir, Ebinezer, Masi, Hami, Manzoor, Salgotra, Sofi, Mushtaq, Rohila and Rakwal. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.