



## OPEN ACCESS

## EDITED BY

Fiona L. Goggin,  
University of Arkansas, United States

## REVIEWED BY

Sotiris Tjamos,  
Agricultural University of  
Athens, Greece  
Andreia Figueiredo,  
University of Lisbon, Portugal

## \*CORRESPONDENCE

Massimo Reverberi  
massimo.reverberi@uniroma1.it

## SPECIALTY SECTION

This article was submitted to  
Plant Pathogen Interactions,  
a section of the journal  
Frontiers in Plant Science

RECEIVED 26 November 2021

ACCEPTED 24 August 2022

PUBLISHED 16 September 2022

## CITATION

Beccaccioli M, Pucci N, Salustri M,  
Scortichini M, Zaccaria M, Momeni B,  
Loreti S, Reverberi M and Scala V  
(2022) Fungal and bacterial oxylipins  
are signals for intra- and inter-cellular  
communication within plant disease.  
*Front. Plant Sci.* 13:823233.  
doi: 10.3389/fpls.2022.823233

## COPYRIGHT

© 2022 Beccaccioli, Pucci, Salustri,  
Scortichini, Zaccaria, Momeni, Loreti,  
Reverberi and Scala. This is an  
open-access article distributed under  
the terms of the [Creative Commons  
Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use,  
distribution or reproduction in other  
forums is permitted, provided the  
original author(s) and the copyright  
owner(s) are credited and that the  
original publication in this journal is  
cited, in accordance with accepted  
academic practice. No use, distribution  
or reproduction is permitted which  
does not comply with these terms.

# Fungal and bacterial oxylipins are signals for intra- and inter-cellular communication within plant disease

Marzia Beccaccioli<sup>1</sup>, Nicoletta Pucci<sup>2</sup>, Manuel Salustri<sup>1</sup>,  
Marco Scortichini<sup>3</sup>, Marco Zaccaria<sup>4</sup>, Babak Momeni<sup>4</sup>,  
Stefania Loreti<sup>2</sup>, Massimo Reverberi<sup>1\*</sup> and Valeria Scala<sup>2</sup>

<sup>1</sup>Department of Environmental Biology, Sapienza University of Rome, Rome, Italy, <sup>2</sup>Research Centre for Plant Protection and Certification, Council for Agricultural Research and the Analysis of Agricultural Economics (CREA), Rome, Italy, <sup>3</sup>Research Centre for Olive, Fruit and Citrus Crops, Council for Agricultural Research and the Analysis of Agricultural Economics (CREA), Rome, Italy, <sup>4</sup>Department of Biology, Boston College, Newton, MA, United States

Lipids are central at various stages of host–pathogen interactions in determining virulence and modulating plant defense. Free fatty acids may act as substrates for oxidizing enzymes [e.g., lipoxygenases (LOXs) and dioxygenases (DOXs)] that synthesize oxylipins. Fatty acids and oxylipins function as modulators of several pathways in cell-to-cell communication; their structural similarity among plant, fungal, and bacterial taxa suggests potential in cross-kingdom communication. We provide a prospect of the known role of fatty acids and oxylipins in fungi and bacteria during plant–pathogen interactions. In the pathogens, oxylipin-mediated signaling pathways are crucial both in development and host infection. Here, we report on case studies suggesting that oxylipins derived from oleic, linoleic, and linolenic acids are crucial in modulating the pathogenic lifestyle in the host plant. Intriguingly, overlapping (fungi-plant/bacteria-plant) results suggest that different inter-kingdom pathosystems use similar lipid signals to reshape the lifestyle of the contenders and occasionally determine the outcome of the challenge.

## KEYWORDS

lipids, oxylipins, *Xylella fastidiosa*, *Fusarium* spp, *Aspergillus* spp, *Olea europaea* L., *Zea mays* (L)

## Host–pathogen communication through the oxylipin language

Host–pathogen recognition relies upon an elaborate multi-molecular communication. In this context, oxylipins—oxidized fatty acids (FAs)—produced in several *phyla* can regulate numerous events associated with physiological and pathological processes (Blée, 2002; Andreou et al., 2009; Mosblech et al., 2009; Ambaw et al., 2021; Liu et al., 2021; Fernandes and Ghag, 2022). The term oxylipin (Gerwick et al., 1993) refers to a broad family of secondary metabolites originated

from the oxidation of polyunsaturated and/or monounsaturated FAs (PUFAs and/or MUFAs) (Mosblech et al., 2009), which comprises a complex array of products: alcohols, aldehydes, ketones, acids, and hydrocarbon gases, generated *via* enzymatic and non-enzymatic processes. Oxylipins share significant structural and functional similarities across different mammal, plant, fungal, and bacterial taxa, including part of their biosynthetic pathway, structure, function, and modifications.

In plant, oxylipins are signaling molecules involved in the regulation of development and immunity. Jasmonates are the better-characterized oxylipins in plants and include jasmonic acid (JA), its precursor 12-oxo-phytodienoic acid (12-OPDA), and JA derivatives such as methyl jasmonate and JA-isoleucine. JA and its derivatives act as defense regulators, influencing the reproductive and pathogenetic processes during the interaction with both beneficial and pathogenic microorganisms (Andreou et al., 2009; Deboever et al., 2020); pathogen infection and pest wounding are the best-studied environmental triggers, in this context (Farmer et al., 2003; Block et al., 2005; Gorman et al., 2021; Shaban et al., 2021). The well-characterized JA pathway activation starts with conjugated lipids in the chloroplast membranes (e.g., monogalactosyldiacylglycerol). Lipase A1 mediates the release of  $\alpha$ -linolenic acid, and LOX enzyme catalyzes PUFA dioxygenation. Oxygenation may happen at several positions along the carbon chain. For JA, important PUFA precursors are  $\alpha$ -linolenic acid (Wasternack and Strnad, 2018) or, alternatively, the hexadecatrienoic acid (Chini et al., 2018). Local defenses and systemic acquired resistances (e.g., SAR, ISR) require JA involvement (Ryan and Moura, 2002; Yu et al., 2022) to express a distinct set of defense-related genes (Okada et al., 2015). Biosynthesis of other plant oxylipins is initiated by the 9 and 13-LOX and  $\alpha$ -dioxygenase ( $\alpha$ -DOX), or monooxygenases, which all catalyze the oxygenation of linoleic acid and linolenic acid (mainly) into reactive hydroperoxides, followed by a secondary modification by cytochrome P450 enzymes or peroxygenases (Blée, 2002; Hamberg et al., 2003; Andreou and Feussner, 2009). Oxylipins can also be produced nonenzymatically through free radical-mediated oxygenation (Oenel et al., 2017). Plant (or phyto-) oxylipins are produced under abiotic and biotic stress, as well as during beneficial interactions (Blée, 2002; Feussner and Wasternack, 2002; Prost et al., 2005). Their role spans from antimicrobial agents to signaling molecules. Recently, the employment of oxylipins

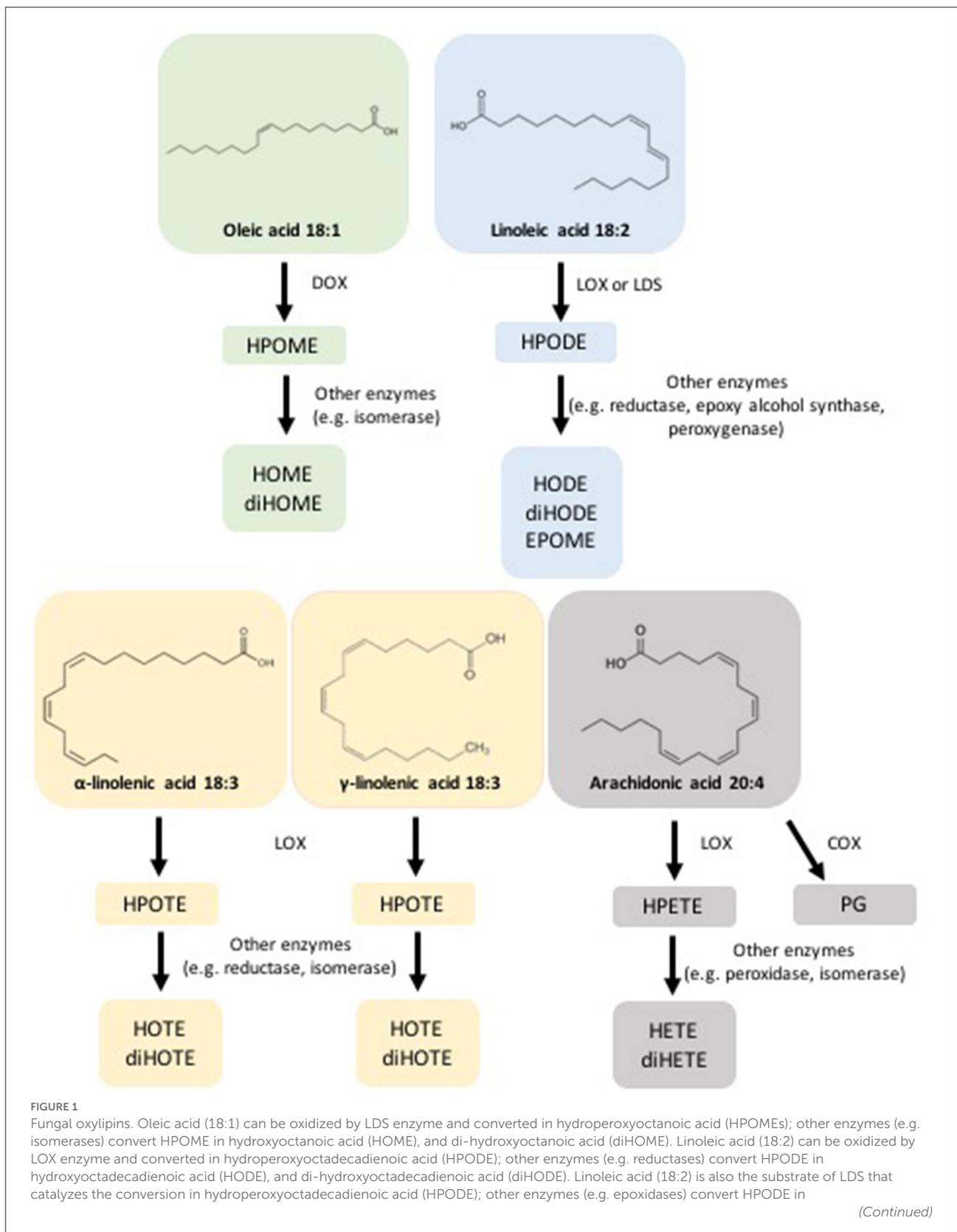
Abbreviations: 12-OH-JA, 12-hydroxy jasmonic acid; COX, cyclooxygenase; CYP450, cytochrome P450; DOX, dioxygenase; FA, fatty acid; GPCR, G protein-coupled receptors; HR, hypersensitive response; ISR, induced systemic resistance; JA, jasmonic acid; LDS, linoleate diol synthase; LOX, lipoxygenase; MUFA, monounsaturated fatty acid; ODS, oxylinpin-dependent quorum sensing system; OQDS, olive quick decline syndrome; PTO, *Pseudomonas syringae* pathovar tomato; PUFA, polyunsaturated fatty acid; QS, quorum sensing; ROS, reactive oxygen species; SAR, systemic acquired resistance.

as effective and commercial antimicrobial agents has been discussed (Deboever et al., 2020). Plants produce diversified oxylipins for different purposes, and oxylipins can act directly against the pathogen (such as *Fusarium* spp.) but also as attractors of biocontrol agents (such as *Trichoderma* spp.) (Gol et al., 2018; Lombardi et al., 2018; Wang et al., 2020). As signaling molecules, they have been implicated in several pathways, including morphogenesis, and as determinants of the hypersensitive response (HR) in incompatible plant–pathogen interactions (Mehta et al., 2021).

In fungi and bacteria, oxylipins control lifestyle and quorum sensing *inter alia* (Su et al., 1995; Tsitsigiannis et al., 2005b). The “oxylinpin regulation system” in fungi and bacteria can employ host oxylipins to improve virulence (e.g., enhancing toxin production or switching to biofilm stage) (Burow et al., 2000; Brodhagen et al., 2008; Martínez et al., 2019) and affect reproduction rate within host tissues by increasing the sporulation (Scarpari et al., 2014). Bacterial oxylipins impact the plant defense mechanisms, as in the case of coronatine production in *Pseudomonas syringae*, a toxin that mimics the plant hormone JA-isoleucine with the aim to induce the opening of stomata to enable bacterial ingress (Zheng et al., 2012). These features support the hypothesis that oxylipins may act as “words” in the lipid common language in host–pathogen communication with a paracrine activity, as suggested by Niu and colleagues (Tsitsigiannis and Keller, 2007; Christensen and Kolomiets, 2011; Niu et al., 2020).

Discriminating which oxylipins are produced by the host and which by the pathogen is not trivial, since the two actors will produce the same molecules at the same time. Oxylipins play a relevant ecological role for the producers (i.e., plants, fungi, and bacteria) and their interspecific interactions (Siebers et al., 2016; Beccaccioli et al., 2021b). In plant–pathogen interactions, plants (the host) produce oxylipins to systemically signal pathogen attack, mounting an efficient defense system and interfering with pathogen growth and reproduction (Burow and Nesbitt, 1997; Brodhagen et al., 2008; Andreou et al., 2009; Scarpari et al., 2014).

Numerous evidence shows that oxylipins mediate interspecies signaling among eukaryotes (Pohl and Kock, 2014). *Trichoderma virens* stimulates symbiont-induced systemic resistance in maize by promoting the release of 12-OPDA and  $\alpha$ -ketol of octadecadienoic in the xylem sap (Wang et al., 2020). In plants, 9-HPODE and 13-HPODE alter the secondary metabolism and conidiation in *Aspergillus flavus* (Calvo et al., 1999), *Colletotrichum gramminicola* (Gorman et al., 2021), *Verticillium dahliae* (Shaban et al., 2021), *Fusarium oxysporum* (Fernandes and Ghag, 2022), and *inter alia*. Fungal oxylipins are involved in the control of sexual and asexual structures formation, secondary metabolism, density-dependent growth, and in the interaction with hosts (Reverberi et al., 2010; Brodhun and Feussner, 2011; Christensen and Kolomiets, 2011). In bacteria, very recent studies in *Pseudomonas aeruginosa* and



**FIGURE 1 (Continued)**

hydroxyoctadecadienoic acid (HODE), di-hydroxyoctadecadienoic acid (diHODE), and epoxyoctadecenoic acids (EPOME). LOX enzyme acts upon  $\alpha/\delta$ -Linolenic acid (18:3) to generate hydroperoxyoctadecatrienoic acid (HPOTE); HPOTE is the substrate of other enzymes to generate hydroxyoctadecatrienoic acid (HOTE) and di-hydroxyoctadecatrienoic acid (diHOTE). Arachidonic acid (20:4) can be oxidized by LOX and converted in hydroperoxyeicosatetraenoic acid (HPETE); HPETE is the substrate of hydroxyeicosatetraenoic acid (HETE), di-hydroxyeicosatetraenoic acid (diHETE). COX enzyme convert the arachidonic acid (20:4) in prostaglandins (PGs).

*Xylella fastidiosa* suggest a role in mediating autocrine or paracrine signals in the communication with their hosts or vectors (Martínez et al., 2019; Niu et al., 2020; Scala et al., 2020).

During infection, host–pathogen communication determines the fate of the interaction. Here, we show how oxylipins represent a common language shared among plant and pathogens, both fungal and bacterial. Some case studies related to oxylipin-mediated plant–pathogen interaction regarding interaction between host plants and filamentous fungi or bacterial pathogens will be presented.

## Fungal oxylipins

From a chemical point of view, oxylipins are products of oxygenation of MUFAs or PUFAs. FAs are the main components of several complex lipids (acylglycerols, glycerophospholipids, glycolipids, sphingolipids, and sterol) from where they may be cleaved. Free FAs are diversified by length and degree of unsaturation. PUFA availability is crucial for the cell because, based on the degree of saturation, they alter cell membrane fluidity, the arrangement and availability of receptors, influencing signal transduction regulation. In this regard, fatty acids may act directly on the receptors (de Angelis et al., 2016).

Oxidation of FAs may be spontaneous in the presence of reactive oxygen species (ROS) or a consequence of enzymatic activity. The study of oxylipins in fungi followed the discovery of the role of lipid hydroperoxides in *Aspergillus parasiticus* (Fabbri et al., 1983) and of the *psi* factors (precocious sexual inducers), a series of fatty acid-derived oxylipins involved in the regulation of the development of spores and conidia (Champe et al., 1987). Different studies have elucidated the biosynthetic pathways of these molecules. Phospholipids and acylglycerides provide the substrate to oxylipin synthesis: phospholipases release oleic, linoleic, linolenic, and arachidonic acids, central elements for the formation of fungal oxylipins (Sakuradani et al., 2009; Beccaccioli et al., 2019).

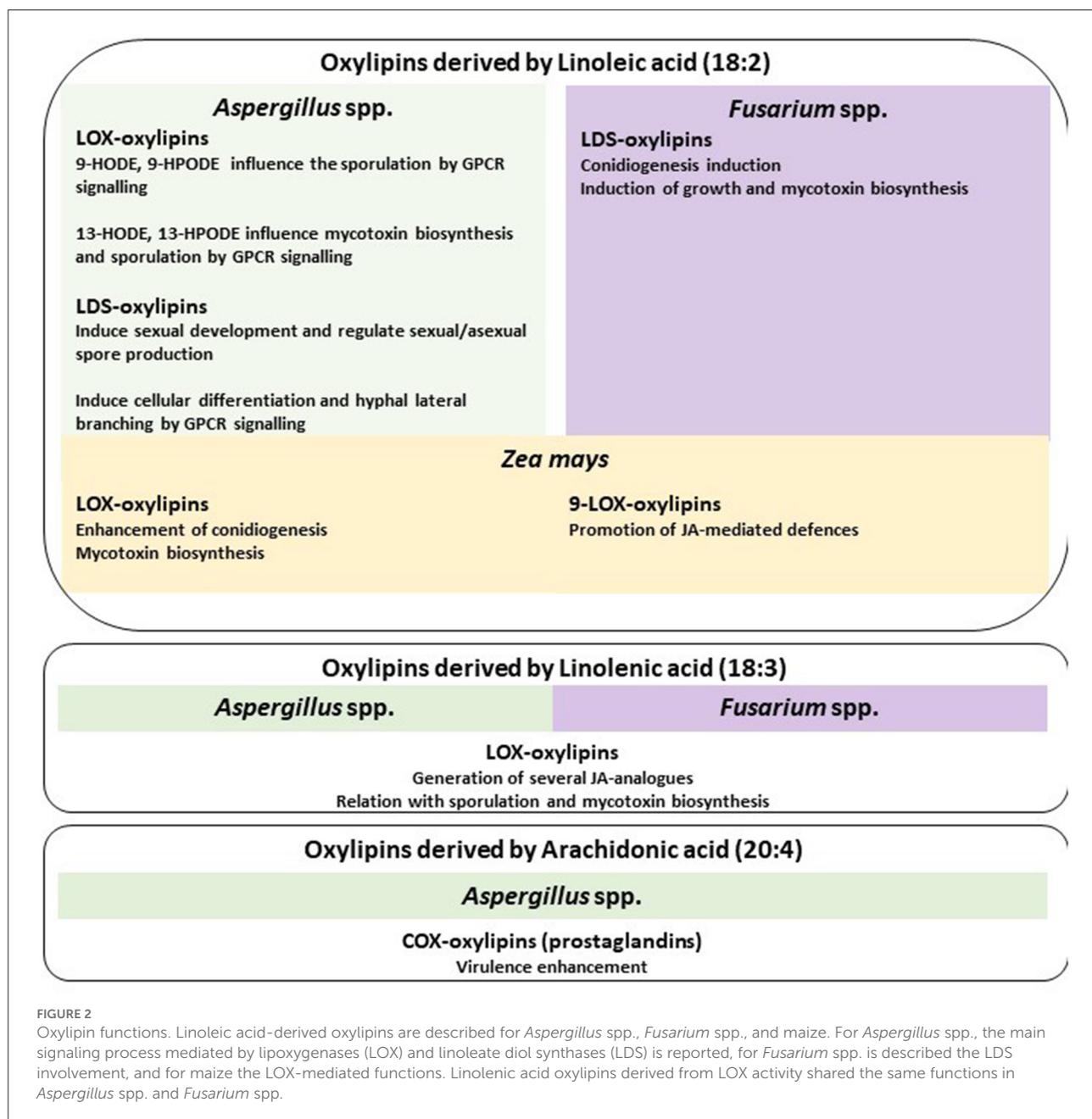
The first evidence of enzymatic formation of oxylipins in fungi was found in *Gaeumannomyces graminis*, the causal agent of root and crown rot diseases. *G. graminis* produces oxylipins *via* FA oxidation with LOXs, DOXs [including the transformation of linoleic acid into dihydroxy-linoleate by 8-DOX (also named linoleate diol synthase or LDS)], and cyclooxygenase (COX) (Su et al., 2000). More recently, fatty acid dioxygenase-cytochrome P450 fusion enzymes have been

identified in several pathogenic fungi, along with the implication of several oxylipins such as 8-, 9-, or 10-hydroperoxy metabolite biosynthesis (Oliw, 2021). Each fatty acid may generate different oxylipins based on enzymatic activity and spontaneous reactions available in the oxidant environment. We list known fungal oxylipins (Figure 1) and present examples of their function in relation to their chemical origin based on the class of oxygenase (LOX, DOX or LDS, and COX) (Figure 2), with a focus on the role in plant–fungus interaction.

## Enzymatic biosynthesis of oxylipins

### LOX-derived oxylipins in fungi

LOX-mediated oxygenation may occur on linoleic or linolenic FAs. Early evidence was reported in *Aspergillus nidulans*; activity on linoleic acid generates oxylipins 9-HODE and 9-HPODE that are involved in sporulation control (Calvo et al., 1999). LOX activity on linoleic acid may also form 13-HODE and 13-HPODE, inhibiting mycotoxin production in *Aspergillus* spp. (Burow and Nesbitt, 1997) and promoting sporulation (Calvo et al., 1999). Both 9-LOX and 13-LOX, which differ in the position at which they cause oxygenation, stimulate cAMP production by fungal G protein-coupled receptor (GPCR) signaling (Affeldt et al., 2012), suggesting its activation to transduce autocrine signaling. In the mycoparasite *Trichoderma atroviride*, LOX1 is indispensable for 6-pentyl-2H-pyran-2-one production and to parasitize and antagonize host fungi, for conidiation in darkness, response to injury (i.e., 9-HODE, 13-HODE, 9-oxoOTrE, and 13-HOTrE), and production of volatile organic compounds. LOX1 in *T. atroviride* is required during the interaction with the host plant, particularly in *Arabidopsis thaliana*, to induce systemic resistance against the plant-pathogenic fungus *Botrytis cinerea* (Speckbacher et al., 2020). Fungal oxylipins derived from linolenic acid by LOX-oxidation may generate several JA analogs borrowed by the plant and act as precursors for JA, methyl-JA, and 12-OH-JA formation. These compounds were first identified in *Fusarium oxysporum* and *Aspergillus niger* (Miersch et al., 1999a,b). In plants, 12-OH-JA represents the inactive form of JA. It is likely to prevent the inhibitive effects of accumulation of JA on plant development and growth. In *Ganoderma lucidum*, the presence of 12-OH-JA also regulates host colonization by inhibiting methyl-JA formation, important for plant defense (Patkar et al., 2015). More recently, research has shown that



*Magnaporthe oryzae*, the causal agent of the rice blast, can autonomously produce analogs of JA to control its pathogenic development (Liu et al., 2021).

### DOX/LDS-derived oxylin in fungi

Several fungi possess genes encoding for DOX which is involved in oxylin biosynthesis. DOX enzymes contain a catalytic domain often fused to a functional cytochrome P450 at the C-terminal end. Therefore, they constitute a peculiar group within the peroxidase-cyclooxygenase

superfamily. Their oxidizing activity toward linoleic acid produces five different hydroperoxides. The CYP450 domain can then transform these hydroperoxides into diols, epoxy alcohols, and allene oxides which all support sporulation, hyphal branching, and development in filamentous fungi. Fungal pathogens with biotrophic [e.g., *Blumeria graminis* (Rupasinghe et al., 2006), *Ustilago maydis* (Huber et al., 2002)], hemibiotrophic [e.g., *Fusarium verticillioides* (Scala et al., 2014)], and necrotrophic [e.g., *Botrytis cinerea* (Niu et al., 2020)] phenotypes express these “fusion enzymes” during pathogenesis to support virulence.

LDS-derived oxylipins have different roles across fungal species. In *F. verticillioides*, linoleic acid is the substrate of LDS from which 8-HPODE and 8,13-diHODE are derived. These oxylipins can influence fungal growth and mycotoxin production (Scala et al., 2013, 2014). LDS oxylipins also promote pathogenesis and modulate the expression of maize oxylipins which, in turn, produce 9-LOX oxylipins to trigger JA-mediated defense (Battilani et al., 2018). In *A. nidulans*, 5,8-diHODE, produced from linoleic acid, regulates sexual development and the conidiation/sporulation ratio (Champe et al., 1987; Mazur et al., 1990; EbrahimáEl-Zayat, 1991; Tsitsigiannis and Keller, 2007; Brown et al., 2008). In *Aspergillus fumigatus* and *Aspergillus flavus*, 5,8-diHODE and 7,8-diHODE induce cell differentiation and lateral branching through GPCRs. Psi-producing oxidase (essentially a group of oxygenases such as LDS) is largely conserved among filamentous fungi; the synthesis of diol-containing oxylipins, primarily 5,8- and 7,8-diHODE, might represent a crucial step in the organization of fungal “morphology” (namely hyphal branching and polarity) and even “escape” from antifungals (Niu et al., 2020). For instance, in the pathogenic ascomycete *Magnaporthe grisea*, 5,8-diHODE accumulates during appressoria formation (Niu et al., 2020) paving the way for host infection. Apparently, these species of diol-oxylipins generate a signaling network in the mycelial mat to tightly adapt hyphal morphology to the environment.

### COX-derived oxylipins in fungi

COX produces active mediators of inflammatory response from arachidonic acid oxygenation. The main findings on fungal development focus on oxylipins deriving from linoleic acid involved in sporulation and mycotoxin biosynthesis in *A. flavus*, *A. parasiticus* (Calvo et al., 1999), and *F. verticillioides* (Gao et al., 2007). Recently, research has also focused on fungal production of jasmonate analogs (Eng et al., 2021). Information is more scarce on oxylipins generated from arachidonic acid, a fatty acid mainly present in extremophile fungi such as *Mortierella alpina* (Kikukawa et al., 2018). In *A. fumigatus*, arachidonic acid is oxygenated by COX-like enzymes to produce prostaglandins to enhance virulence (Tsitsigiannis et al., 2005a). In *Cryptococcus neoformans* and *Candida albicans*, several studies show that fungal prostaglandins promote colonization and infection (Noverr et al., 2001, 2003).

### Oxylipin signaling in plant–fungus interactions

Mechanisms of oxylipin biosynthesis and signaling are similar in plants and fungi. Here, we report examples on how plant and fungus exploit the oxylipins to exchange messages involved in the outcome of the interaction.

The structural similarity between plant and fungal oxylipins can be exploited to reciprocally manipulate each other's signaling. In *A. flavus* on maize, thanks to this structural similarity, the pathogen's oxylipins can act on the plant's oxylipinogenic pathways, and *vice versa*. This cross talk among host and pathogen was demonstrated in different experimental settings, as reported below. In *Aspergillus* cultures, the exogenous application of plant oxylipins (i.e., 9-HPODE naturally produced by maize) increases sporulation and mycotoxin production. In *Aspergillus* deletion mutants for the DOX *ppoA* and *ppoC*, the wild-type phenotype can be restored through the insertion of maize lipoxygenase, suggesting a complementing activity. Inoculation of peanut seeds with *Aspergillus ppo*-mutants reduces LOX expression in seed, suggesting that fungal oxylipins are related to the plant LOX expression (Brodhagen et al., 2008).

The hydroperoxide 9-HODE generated by LOX activity offers an example of a shared signaling molecule. In plants, it induces programmed cell death, as observed in tomato protoplasts (Knight et al., 2001); in fungi, it induces sporulation and the cAMP-G protein-coupled pathway-mediated signaling (Calvo et al., 1999; Affeldt et al., 2012). Furthermore, during the interaction between maize and specific fungal pathogens, the disruption of 9-LOX from maize promotes increased resistance to the fungal pathogen and decreased mycotoxin contamination (Gao et al., 2007). 9-HODE seems to be very important for the progression of infection.

The *F. verticillioides*-maize pathosystem provides numerous examples of oxylipins as mediators of fungus–plant interaction. A close correlation exists between the fumonisin B1 accumulation and oxylipin signaling. When fumonisin is produced, maize increases the release of 9-HODE, suggesting an active role in infection development (Dall'Asta et al., 2014; Beccaccioli et al., 2021a). Further evidence shows that, in maize, the 9-LOX (*ZmLOX3*) deletion alters *F. verticillioides* fitness, decreasing conidiation and fumonisin B1 synthesis (Gao et al., 2007). In addition, when *FvLDS1* is deleted in the pathogen and *ZmLOX3* is mutagenized in the host, resistance to the infection increased, *ZmLOX4*, *ZmLOX5*, and *ZmLOX12* expression was upregulated, and the fungus showed decreased fumonisin production. The *ZmLOX4* and *ZmLOX5* mutants were more susceptible to *F. verticillioides* and showed reduced levels of JA, suggesting the relevance of JA-mediated defense signaling. Intriguingly, *F. verticillioides* infecting the *ZmLOX3* mutant is less effective at accumulating oxylipins from linoleate diol synthase and properly infecting maize kernels; this suggests that the oxylipins derived by *ZmLOX3* activity are central to fungal virulence (Battilani et al., 2018).

The coexistence of a common language among hosts and pathogens supports the idea that a common receptor may also exist, and it could be represented by the GPCRs (Brown et al., 2018). Oxylipin perception has been explored only partially. In mammals, oxylipins are perceived by GPCRs (Funk, 2001;

Noverr et al., 2003) in the plasma membrane (Funk, 2001). G2A is a GPCR receptor, characterized in the lymphoid tissues, and is able to recognize oxylipins derived from linoleic and arachidonic acid (Obinata et al., 2005). GPCR promotes root growth and ROS scavenging probably in the context of oxylipins pathway perception of abiotic stress in *Arabidopsis* and cotton (Lu et al., 2019). In fungi, several research studies showed the GPCR involvement in numerous functions among which oxylipin sensing (Affeldt et al., 2012). G proteins have a crucial role in sensing external ligands such as nutrients, hormones, proteins, pheromones and other peptides, ions, hydrophobic surfaces, and light (Kochman, 2014). Several works suggest that fungal GPCR-mediated signaling is linked to pathogenesis and could be considered a target for disease control (Brown et al., 2018). Secondary metabolism seems to be related to the G protein pathway as well through the activity of transcription factors related to toxin biosynthesis (Gao et al., 2021).

## Bacterial oxylipins

The “story” of oxylipins in bacteria is far more recent than in fungi. Only recently, this class of lipid compounds was found to be significant in regulating different aspects of the bacterial lifestyle. The molecules from which oxylipins originate, that is, lipids, have a quite vast background. Many lipid molecules in bacterial cells, such as hopanoids and ornitholipids, are absent in eukaryotes, while others are shared (e.g., phospholipids). Lipids play important roles in bacterial cell-to-cell communication by regulating quorum sensing (QS) and in the interactions with the host and the vector (Siebers et al., 2016). In phytopathogenic bacteria, different types of lipids can determine compatibility with the host. Numerous research papers describe the role of FAs as diffusible signal factors (DSF) acting as modulators of different pathways in cell-to-cell communication to modulate QS and virulence (Wang and Qian, 2019). In phytopathogenic bacteria such as *X. fastidiosa* responsive of olive quick decline syndrome (OQDS), DSF-based QS model promotes biofilm formation and stickiness, determining (a) degradation of pit membranes to enable cross-vessel diffusion in the xylem; (b) twitching motility of bacterial cells; and (c) adhesion to the xylem surface and the switch from the planktonic endophytic lifestyle to the sessile insect-acquisition stage (Chatterjee et al., 2008; Beaulieu et al., 2013; Ionescu et al., 2016). The QS regulation is based on a delicate balance of several DSFs [e.g., MUFAs: lauroleic acid (C12:1), myristoleic acid (14:1), palmitoleic acid (16:1), oleic acid (18:1)] (Lindow et al., 2014; Ionescu et al., 2016).

As reported in Figure 1, oleic acid is the major substrate for the DOX-mediated synthesis of 10-HPOME and 7,10-diHOME (Martínez and Campos-Gómez, 2016). The role of oxylipins in mediating autocrine or paracrine signaling in

communication is reported in several recent papers (Martínez et al., 2019; Niu et al., 2020; Scala et al., 2020). Despite their importance in eukaryotes and in plant–fungi interactions, the role of oxylipins is overlooked in phytopathogenic bacteria. Only recently, Martínez and colleagues demonstrate that unsaturated FAs can act as substrates for oxidizing enzymes (e.g., LOX and DOX) to form oxylipins that, in the opportunistic bacterial pathogen *Pseudomonas aeruginosa*, may transform into mono- and di-hydroxylated derivatives during the interaction with the host (e.g., *Drosophila*, lettuce) (Martínez and Campos-Gómez, 2016).

## Enzymatic biosynthesis of oxylipins

### LOX-derived oxylipins in bacteria

Lipoxygenase LOXA, which was first identified in the human parasite *P. aeruginosa*, was the first prokaryotic lipoxygenase ever to be characterized. LOXA transforms arachidonic acid into 15-hydroxyheicosatetraenoic acid (15-HETE) (Vance et al., 2004). LOXA is secreted by *P. aeruginosa* in the lungs and oxidizes the PUFAs. Several biological roles, including interference with the host lipid signaling, and modulation of bacterial invasion have been hypothesized (Morello et al., 2019).

In liquid culture, *X. fastidiosa* accumulates different oxylipins, in particular, 10-HPOME, 10-HOME, and epoxyoctadecamonoenoic acids (EpOMEs), with different intra/extra-cellular distribution. LOX enzyme-derived oxylipins (i.e., 13-HODE, 9-HODE; 8,13-diHODE, 13HOTrE, and methyl jasmonic acid) are less represented (Christensen and Kolomiets, 2011). The plant stress hormone methyl jasmonate is secreted by *X. fastidiosa in vitro* and, overall, in large amounts in artificially infected plant tissues (Nomura et al., 2005). These results indicate that *X. fastidiosa* can synthesize and secrete oxylipins suggesting that, although oxygenation may occur inside the cell, oxylipins are transported through the outer membrane and accumulate in the medium (Martínez and Campos-Gómez, 2016; Scala et al., 2018). Recent *in vitro* studies on *X. fastidiosa* subsp. *pauca* demonstrate that the 9-LOX-derived oxylipins promote biofilming, whereas DOX-derived oxylipins stimulate planktonic growth and inhibit biofilm formation (Scala et al., 2020).

### DOX-derived oxylipins in bacteria

In *P. aeruginosa*, DOXs including diol synthase catalyze the stereospecific oxygenation of oleic acid (Martínez and Campos-Gómez, 2016) to synthesize 10-HOME and 7,10-diHOME. These oleic acid-derived oxylipins are involved in regulating motility, biofilm formation, and virulence. In the QS regulating system, they inhibit type III pili-induced-motility by stimulating

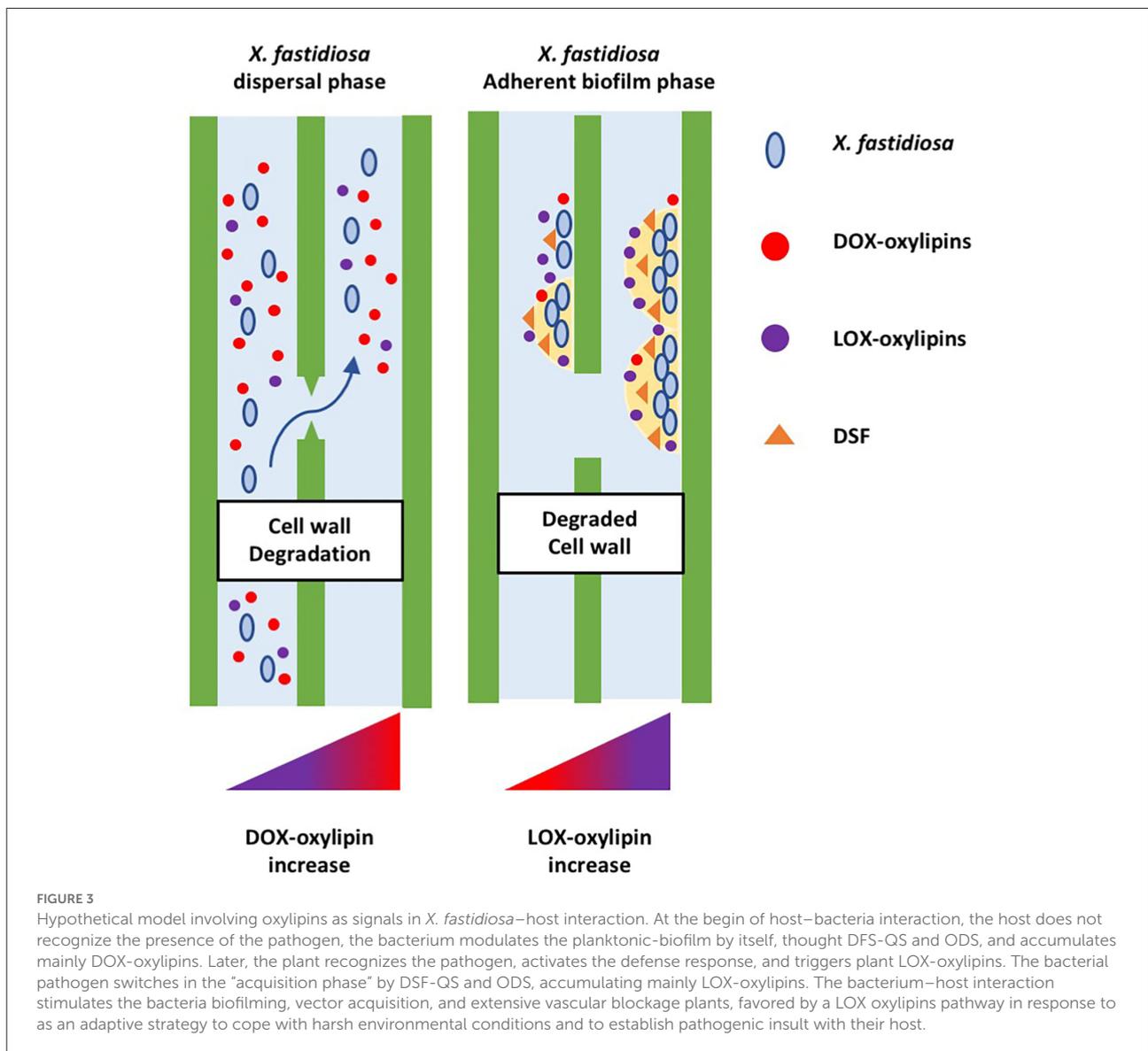
the expression of type IV pili, thus promoting twitching and aggregation in micro-colonies and biofilm formation, *in vitro*. These oxylipins are promoters of virulence on *Drosophila melanogaster* and lettuce (Martínez and Campos-Gómez, 2016). *P. aeruginosa* can co-opt host oxylipins to let them operate as environment-specific QS signals. A recent study (Martínez et al., 2019) highlighted a new oxylipin-dependent quorum sensing system (ODS) and the role of oxylipins produced from the host's oleic acid as auto-inducers of lifestyle switch in *P. aeruginosa* (Martínez et al., 2019).

Research on *X. fastidiosa* highlights similar trends (Scala et al., 2018, 2020), suggesting that oxylipins' role in autocrine bacterial cell communication (Niu et al., 2020) could be common across bacterial families (Martínez et al., 2019).

### Oxylipin signaling in plant–bacteria interactions

Although studies on oxylipins in plant–bacteria interactions are few and primarily focused plant oxylipins, nevertheless, a pivotal study by Martínez and Campos-Gómez (2016) suggests that oxylipins may also be crucial in bacterial communication. This section focuses on several recent studies that suggest that bacterial oxylipins may also be important signals in the intricate signaling network between plant hosts and pathogenic bacteria.

The example of *Arabidopsis thaliana* inoculated with *Pseudomonas syringae* pv. *tomato* (Pto) represents one of the first studies based on the cross talk mediated by the oxylipins during plant–bacteria interaction. The pre-treatment of plant



with 9-LOX- and  $\alpha$ -DOX-oxylipins from linoleic acid activated SAR and protected plant tissues against infection through a JA-independent signaling pathway (Hamberg et al., 2003; Prost et al., 2005; Truman et al., 2007; Vellosillo et al., 2007; Jung et al., 2009; Xia et al., 2009; Chanda et al., 2011; Vincent et al., 2012). In plant, the presence of 9-LOX-derived oxylipins induces brassinosteroid synthesis (hormones important for plant development and growth), SAR, and cell wall-based defense such as callose deposition (Hamberg et al., 2003; Vellosillo et al., 2007; Marcos et al., 2015). Among 9-oxylipins, 9-ketooctadecatrienoic acid pre-treatment modifies hormone homeostasis during infection and interferes with the hormonal changes caused by bacterial effectors (Vellosillo et al., 2007). JA, generated within 13-LOX pathway of linolenic acid, participates in the establishment of SAR after infection by biotrophic bacteria (Truman et al., 2007). 13-LOX derivatives participate in plant defense as regulators of gene expression, cell death, and antimicrobials (Stintzi, 2000; Vollenweider et al., 2000; Montillet et al., 2004; Prost et al., 2005).

Details of the oxylipin-mediated communication between plant and phytopathogenic bacteria were provided by several studies on *X. fastidiosa*. The infection caused by *X. fastidiosa* was evaluated in plant model systems. *A. thaliana* induces the expression of ethylene/JA pathway to limit oxidative damage and represses the salicylic acid pathway (Rogers, 2012; Pereira et al., 2019); *Nicotiana tabacum* leads to a differential accumulation of specific lipid entities, including the oxylipins (Scala et al., 2018). Oxylipins emerged as hallmarks of pathogenic invasion in host tissues: Plants infected with *X. fastidiosa* accumulate oleic acid- and linoleic acid-derived oxylipins (e.g., 7,10-diHOME and 13-HODE). The study of oxylipin metabolism in the *X. fastidiosa* pathosystem was investigated also in *Olea europaea*. Symptomatic trees of the susceptible olive cultivar Ogliarola salentina accumulate ten lipid compounds that should be defined as hallmarks of OQDS (Scala et al., 2019). Identified hallmarks include 10-HOME (DOX-oxylipins); oleic and linoleic acid; LOX-derived 9- and 13-oxylipins (9-HODE, 9-OXODE, 9-HOTRE, 13-HODE, 13-OXODE, and 13-HOTRE). All the identified compounds were tested *in vitro* to unveil their effect on planktonic or biofilming state of *X. fastidiosa* subsp. *pauca*. The 7,10-diHOME, a DOX-derived oxylipin, downregulates biofilm formation, while LOX oxylipins from linoleic acid stimulate it. Recently, combined mass spectrometry/machine learning approach demonstrated that 13-HODE is a hallmark of OQDS and a susceptibility factor for olive tree toward *X. fastidiosa* (Scala et al., 2022). Considering the lipidome profile of the bacteria alone, the host–pathogen pathosystem, and available literature (Chatterjee et al., 2008; Ropicavoli et al., 2018; Roper et al., 2019; Scala et al., 2020), we argue that in plant tissue infected by *X. fastidiosa*, oxylipin involvement occurs from the early stage of infection just before host–pathogen recognition. At this

stage, the pathogen modulates the planktonic-biofilm through DFS-QS and ODS; DOX-oxylipins are mainly accumulated for extensive vessel colonization, leading the planktonic state. At a later stage of infection, the host identifies the pathogen, activates the defense response, and triggers plant LOX-oxylipins. The accumulation of LOX oxylipins leads to a switch to the “acquisition phase” in the pathogen, a step of the infection cycle that culminates with the bacteria acquisition from the xylem sap to the insect vector (Figure 3). LOX accumulation is induced by the bacterium–host recognition and stimulates biofilm formation, vector acquisition, and extensive vascular blockage in plants, in accordance with the results obtained in the *A. thaliana*-Pto interaction. The activation of the LOX oxylipin pathway, in response to the plant–pathogen interaction, seems to be an adaptive strategy for the bacteria to cope with harsh environmental conditions and establish pathogenic insult with the host (Mosblech et al., 2009; Wasternack, 2014). A reversible cycle is *de facto* in place among the dual state of bacteria up to the host immune-system activation. Arguably, the pathogen regulates its behavior through FAs (as the DSFs) and oxylipins to trigger systemic invasion, limiting biofilm formation and acquisition by insect vectors.

These studies suggest that oxylipins have a significant role in determining the fate of the interactions: Resistance vs. susceptibility and that plant on one side and bacteria on the other try to adapt to the oxylipin signature formed during their interaction to exploit the weaknesses of the opponent.

## Conclusion

Lipid-mediated signal communication is an issue ongoing with numerous implications in plant–fungi and plant–bacteria disease control. This review focused primarily on oxidized lipids, the oxylipins, implicated in host–pathogen interactions, notably when the plant is exposed to fungal or bacterial disease. In plants, the oxylipins—jasmonates apart—are mainly involved in the defense against pathogens; in pathogens, a dual role emerges: oxylipins are produced as autocrine signals to mediate the development and differentiation (i.e., sexual sporulation in fungi; phase transition in bacteria), and paracrine signals to entertain a communication during plant diseases. The LOX pathway is the best characterized in plants and seems to be crucial for communicating with both fungi and bacteria, probably antagonizing and modulating the pathogen’s response. LDS pathway seems to be crucial for the pathogen invasion (i.e., fungi and bacteria). The identification of oxylipins common or structurally similar in plants, fungi, and bacteria consolidates the theory of cross-kingdom communication. We can suggest that the

ability to “recognize and react” to specific oxylipins may drive the fate of the interaction for the host: susceptibility vs. resistance.

Despite numerous findings regarding the specific oxylipins that are exchanged during the interactions, many questions remain open. One of these concerns the understanding of the receptor and transduction system of oxylipins that remains still unclear.

## Author contributions

MB, MR, and VS: conceptualization. MSa, MB, and NP: methodology. MR, MSc, and SL: investigation and funding acquisition. MB and VS: writing—original draft. MB, VS, BM, MZ, MSc, NP, and SL: writing—reviewing and editing. MB: visualization. MR and VS: supervision. All authors contributed to the article and approved the submitted version.

## Funding

This study was funded by MIPAAFT, Project Oli.Di.X.I.It (Olive growing and defense against *Xylella fastidiosa* and vector insects in Italy), D.M. 23773 of 6/09/2017, Project SALVAOLIVI (Safeguard and enhancement of the Italian olive-growing

heritage with research actions in the phytosanitary defense sector), D.M.33437 of 12/21/20. MIUR National Operational Program Research and Innovation 2014–2020 (PON RI 2014–2020) Development of Nutraceuticals from Natural Sources - ARS01\_01166. PUT National Operational Program “Enterprises and Competitiveness” 2014–2020 ERDF, UR - Development of new technologies in precision agriculture for the sustainable production of potato genotypes with high nutritional qualities (Acronym SOS TATA), no. F / 200088 / 01-03 / X45.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher’s note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## References

- Afeldt, K. J., Brodhagen, M., and Keller, N. P. (2012). *Aspergillus* oxylipin signaling and quorum sensing pathways depend on G protein-coupled receptors. *Toxins* 4, 695–717. doi: 10.3390/toxins4090695
- Ambaw, Y. A., Pagac, M. P., Irudayaswamy, A. S., Raida, M., Bendt, A. K., Torta, F. T., et al. (2021). Host/malassezia interaction: a quantitative, non-invasive method profiling oxylipin production associates human skin eicosanoids with *Malassezia*. *Metabolites* 11, 700. doi: 10.3390/metabo11100700
- Andreou, A., Brodhun, F., and Feussner, I. (2009). Biosynthesis of oxylipins in non-mammals. *Prog. Lipid Res.* 48, 148–170. doi: 10.1016/j.plipres.2009.02.002
- Andreou, A., and Feussner, I. (2009). Lipoxygenases - structure and reaction mechanism. *Phytochemistry* 70, 1504–1510. doi: 10.1016/j.phytochem.2009.05.008
- Battilani, P., Lanubile, A., Scala, V., Reverberi, M., Gregori, R., Falavigna, C., et al. (2018). Oxylipins from both pathogen and host antagonize jasmonic acid-mediated defence via the 9-lipoxygenase pathway in *Fusarium verticillioides* infection of maize. *Mol. Plant Pathol.* 19, 2162–2176. doi: 10.1111/mpp.12690
- Beaulieu, E. D., Ionescu, M., Chatterjee, S., Yokota, K., Trauner, D., and Lindow, S. (2013). Characterization of a diffusible signaling factor from *Xylella fastidiosa*. *MBio*. 4, e00539–12. doi: 10.1128/mBio.00539-12
- Beccaccioli, M., Reverberi, M., and Scala, V. (2019). Fungal lipids: biosynthesis and signalling during plant-pathogen interaction. *Front. Biosci.* 24, 172–185. doi: 10.2741/4712
- Beccaccioli, M., Salustri, M., Scala, V., Ludovici, M., Cacciotti, A., D’Angeli, S., et al. (2021a). The effect of fusarium verticillioides fumonisins on fatty acids, sphingolipids, and oxylipins in maize germlings. *Int. J. Mol. Sci.* 22, 2435. doi: 10.3390/ijms22052435
- Beccaccioli, M., Scala, V., and Massimo, R. (2021b). “Communication with plants,” In *Encyclopedia of Mycology* (Amsterdam: Oliver Walter), 114–122.
- Blée, E. (2002). Impact of phyto-oxylipins in plant defense. *Trends Plant Sci.* 7, 315–321. doi: 10.1016/S1360-1385(02)02290-2
- Block, A., Schmelz, E., Jones, J. B., and Klee, H. J. (2005). Coronatine and salicylic acid: the battle between *Arabidopsis* and *Pseudomonas* for phytohormone control. *Mol. Plant Pathol.* 6, 79–83. doi: 10.1111/j.1364-3703.2004.00265.x
- Brodhagen, M., Tsitsigiannis, D. I., Hornung, E., Goebel, C., Feussner, I., and Keller, N. P. (2008). Reciprocal oxylipin-mediated cross-talk in the *Aspergillus*-seed pathosystem. *Mol. Microbiol.* 67, 378–391. doi: 10.1111/j.1365-2958.2007.06045.x
- Brodhun, F., and Feussner, I. (2011). Oxylipins in fungi. *FEBS J.* 278, 1047–1063. doi: 10.1111/j.1742-4658.2011.08027.x
- Brown, N. A., Schrevens, S., Van Dijck, P., and Goldman, G. H. (2018). Fungal G-protein-coupled receptors: mediators of pathogenesis and targets for disease control. *Nat. Microbiol.* 3, 402–414. doi: 10.1038/s41564-018-0127-5
- Brown, S. H., Zarnowski, R., Sharpee, W. C., and Keller, N. P. (2008). Morphological transitions governed by density dependence and lipoxygenase activity in *Aspergillus flavus*. *Appl. Environ. Microbiol.* 74, 5674–5685. doi: 10.1128/AEM.00565-08
- Burow, G. B., Gardner, H. W., and Keller, N. P. (2000). A peanut seed lipoxygenase responsive to *Aspergillus* colonization. *Plant Mol. Biol.* 42, 689–701. doi: 10.1023/A:1006361305703
- Burow, G. B., and Nesbitt, T. C. (1997). Seed lipoxygenase products modulate *Aspergillus mycotoxin* biosynthesis. *Mol. Plant Microbe Interact.* 10, 380–387. doi: 10.1094/MPML1997.10.3.380
- Calvo, A. M., Hinze, L. L., Gardner, H. W., and Keller, N. P. (1999). Sporogenic effect of polyunsaturated fatty acids on development of *Aspergillus* spp. *Appl. Environ. Microbiol.* 65, 3668–3673. doi: 10.1128/AEM.65.8.3668-3673.1999

- Champe, S. P., Rao, P., and Chang, A. (1987). An endogenous inducer of sexual development in *Aspergillus nidulans*. *Microbiology* 133, 1383–1387. doi: 10.1099/00221287-133-5-1383
- Chanda, B., Xia, Y. E., Mandal, M. K., Yu, K., Sekine, K., Gao, Q., et al. (2011). Glycerol-3-phosphate is a critical mobile inducer of systemic immunity in plants. *Nat. Genet.* 43, 421–427. doi: 10.1038/ng.798
- Chatterjee, S., Wistrom, C., and Lindow, S. E. (2008). A cell–cell signaling sensor is acquired for virulence and insect transmission of *Xylella fastidiosa*. *Proc. Natl. Acad. Sci.* 105, 2670–2675. doi: 10.1073/pnas.0712236105
- Chini, A., Monte, I., Zamarreño, A. M., Hamberg, M., Lassueur, S., Reymond, P., et al. (2018). An OPR3-independent pathway uses 4, 5-didehydrojasmonate for jasmonate synthesis. *Nat. Chem. Biol.* 14, 171. doi: 10.1038/nchembio.2540
- Christensen, S. A., and Kolomiets, M. V. (2011). The lipid language of plant–fungal interactions. *Fungal Genet. Biol.* 48, 4–14. doi: 10.1016/j.fgb.2010.05.005
- Dall'Asta, C., Giorni, P., Cirilini, M., Reverberi, M., Gregori, R., Ludovici, M., et al. (2014). Maize lipids play a pivotal role in the fumonisin accumulation. *World Mycotoxin J.* 8, 87–97. doi: 10.3920/WMJ14.1754
- de Angelis, L., Rinaldi, T., Cirigliano, A., Bello, C., Reverberi, M., Amaretti, A., et al. (2016). Functional roles of the fatty acid desaturases encoded by KLOLE1, FAD2 and FAD3 in the yeast *Kluyveromyces lactis*. *Microbiology* 162, 1435–1445. doi: 10.1099/mic.0.000315
- Deboever, E., Deleu, M., Mongrand, S., Lins, L., and Fauconnier, M.-L. (2020). Plant–pathogen interactions: underestimated roles of phyto-oxylipins. *Trends Plant Sci.* 25, 22–34. doi: 10.1016/j.tplants.2019.09.009
- EbrahimiEl-Zayat, A. A. (1991). Structure and synthesis of sporogenic psi factors from *Aspergillus nidulans*. *J. Chem. Soc. Chem. Commun.* 1486–87. doi: 10.1039/c39910001486
- Eng, F., Marin, J. E., Zienkiewicz, K., Gutiérrez-Rojas, M., Favela-Torres, E., and Feussner, I. (2021). Jasmonic acid biosynthesis by fungi: derivatives, first evidence on biochemical pathways and culture conditions for production. *PeerJ* 9, e10873. doi: 10.7717/peerj.10873
- Fabbri, A., Fanelli, C., Panfili, G., Passi, S., and Fasella, P. (1983). Lipoperoxidation and aflatoxin biosynthesis by *Aspergillus parasiticus* and *Aspergillus falvus*. *Microbiology* 129, 3447–3452. doi: 10.1099/00221287-129-11-3447
- Farmer, E. E., Alméras, E., and Krishnamurthy, V. (2003). Jasmonates and related oxylipins in plant responses to pathogenesis and herbivory. *Curr. Opin. Plant Biol.* 6, 372–378. doi: 10.1016/S1369-5266(03)00045-1
- Fernandes, L. B., and Ghag, S. B. (2022). Molecular insights into the jasmonate signaling and associated defense responses against wilt caused by *Fusarium oxysporum*. *Plant Physiol. Biochem.* 174, 22–34. doi: 10.1016/j.plaphy.2022.01.032
- Feussner, I., and Wasternack, C. (2002). The lipoxygenase pathway. *Annu. Rev. Plant Biol.* 53, 275–297. doi: 10.1146/annurev.arplant.53.100301.135248
- Funk, C. D. (2001). Prostaglandins and leukotrienes: advances in eicosanoid biology. *Science* 294, 1871–1875. doi: 10.1126/science.294.5548.1871
- Gao, J., Xu, X., Huang, K., and Liang, Z. (2021). Fungal G-protein-coupled receptors: a promising mediator of the impact of extracellular signals on biosynthesis of ochratoxin A. *Front. Microbiol.* 12, 193. doi: 10.3389/fmicb.2021.631392
- Gao, X., Shim, W.-B., Göbel, C., Kunze, S., Feussner, I., Meeley, R., et al. (2007). Disruption of a maize 9-lipoxygenase results in increased resistance to fungal pathogens and reduced levels of contamination with mycotoxin fumonisin. *Mol. Plant Microbe Interact.* 20, 922–933. doi: 10.1094/MPMI-20-8-0922
- Gerwick, W. H., Proteau, P. J., Nagle, D. G., Wise, M. L., Jiang, Z. D., Bernart, M. W., et al. (1993). Biologically active oxylipins from seaweeds. *Hydrobiologia* 260, 653–665. doi: 10.1007/BF00049085
- Gol, S., Pena, R. N., Rothschild, M. F., Tor, M., and Estany, J. (2018). A polymorphism in the fatty acid desaturase-2 gene is associated with the arachidonic acid metabolism in pigs. *Sci. Rep.* 8, 1–9. doi: 10.1038/s41598-018-32710-w
- Gorman, Z., Tolley, J. P., Koiwa, H., and Kolomiets, M. V. (2021). The synthesis of pentyl leaf volatiles and their role in resistance to anthracnose leaf blight. *Front. Plant Sci.* 12, 1760. doi: 10.3389/fpls.2021.719587
- Hamberg, M., Sanz, A., Rodriguez, M. J., Calvo, A. P., and Castresana, C. (2003). Activation of the fatty acid  $\alpha$ -dioxygenase pathway during bacterial infection of tobacco leaves: formation of oxylipins protecting against cell death. *J. Biol. Chem.* 278, 51796–51805. doi: 10.1074/jbc.M310514200
- Huber, S., Lottspeich, F., and Kämper, J. (2002). A gene that encodes a product with similarity to dioxygenases is highly expressed in teliospores of *Ustilago maydis*. *Mol. Genet. Genomics* 267, 757–771. doi: 10.1007/s00438-002-0717-y
- Ionescu, M., Yokota, K., Antonova, E., Garcia, A., Beaulieu, E., Hayes, T., et al. (2016). Promiscuous diffusible signal factor production and responsiveness of the *Xylella fastidiosa* Rpf system. *MBio* 7, e01054–e01016. doi: 10.1128/mBio.01054-16
- Jung, H. W., Tschaplinski, T. J., Wang, L., Glazebrook, J., and Greenberg, J. T. (2009). Priming in systemic plant immunity. *Science* 324, 89–91. doi: 10.1126/science.1170025
- Kikukawa, H., Sakuradani, E., Ando, A., Shimizu, S., and Ogawa, J. (2018). Arachidonic acid production by the oleaginous fungus *Mortierella alpina* 1S-4: A review. *J. Adv. Res.* 11, 15–22. doi: 10.1016/j.jare.2018.02.003
- Knight, V. I., Wang, H., Lincoln, J. E., Lulai, E. C., Gilchrist, D. G., and Bostock, R. M. (2001). Hydroperoxides of fatty acids induce programmed cell death in tomato protoplasts. *Physiol. Mol. Plant Pathol.* 59, 277–286. doi: 10.1006/mpmp.2001.0366
- Kochman, K. (2014). Superfamily of G-protein coupled receptors (GPCRs)-extraordinary and outstanding success of evolution. *Postepy Hig Med. Dosw.* 68, 1225–1237. doi: 10.5604/17322693.1127326
- Lindow, S., Newman, K., Chatterjee, S., Baccari, C., Iavarone, A. T., and Ionescu, M. (2014). Production of *Xylella fastidiosa* diffusible signal factor in transgenic grape causes pathogen confusion and reduction in severity of Pierce's disease. *Mol. Plant Microbe Interact.* 27, 244–254. doi: 10.1094/MPMI-07-13-0197-FI
- Liu, Y., Pagac, M., Yang, F., Patkar, R. N., and Naqvi, N. I. (2021). Fungal jasmonate as a novel morphogenetic signal for pathogenesis. *J. Fungi* 7, 693. doi: 10.3390/jof7090693
- Lombardi, N., Vitale, S., Turrà, D., Reverberi, M., Fanelli, C., Vinale, F., et al. (2018). Root exudates of stressed plants stimulate and attract *Trichoderma* soil fungi. *Mol. Plant Microbe Interact.* 31, 982–994. doi: 10.1094/MPMI-12-17-0310-R
- Lu, P., Magwanga, R. O., Kirungu, J. N., Dong, Q., Cai, X., Zhou, Z., et al. (2019). Genome-wide analysis of the cotton G-coupled receptor proteins (GPCR) and functional analysis of GTOM1, a novel cotton GPCR gene under drought and cold stress. *BMC Genom.* 20, 1–17. doi: 10.1186/s12864-019-5972-y
- Marcos, R., Izquierdo, Y., Velloso, T., Kulasekaran, S., Cascón, T., Hamberg, M., et al. (2015). 9-Lipoxygenase-derived oxylipins activate brassinosteroid signaling to promote cell wall-based defense and limit pathogen infection. *Plant Physiol.* 169, 2324–2334. doi: 10.1104/pp.15.00992
- Martínez, E., and Campos-Gómez, J. (2016). Oxylipins produced by *Pseudomonas aeruginosa* promote biofilm formation and virulence. *Nat. Commun.* 7, 13823. doi: 10.1038/ncomms13823
- Martínez, E., Cosnahan, R. K., Wu, M., Gadila, S. K., Quick, E. B., Mobley, J. A., et al. (2019). Oxylipins mediate cell-to-cell communication in *Pseudomonas aeruginosa*. *Commun. Biol.* 2, 1–10. doi: 10.1038/s42003-019-0310-0
- Mazur, P., Meyers, H. V., Nakanishi, K., and Champe, S. P. (1990). Structural elucidation of sporogenic fatty acid metabolites from *Aspergillus nidulans*. *Tetrahedron Lett.* 31, 3837–3840. doi: 10.1016/S0040-4039(00)97482-3
- Mehta, S., Chakraborty, A., Roy, A., Singh, I. K., and Singh, A. (2021). Fight hard or die trying: current status of lipid signaling during plant–pathogen interaction. *Plants* 10, 1098. doi: 10.3390/plants10061098
- Miersch, O., Bohlmann, H., and Wasternack, C. (1999a). Jasmonates and related compounds from *Fusarium oxysporum*. *Phytochemistry* 50, 517–523. doi: 10.1016/S0031-9422(98)00596-2
- Miersch, O., Porzel, A., and Wasternack, C. (1999b). Microbial conversion of jasmonates-hydroxylations by *Aspergillus niger*. *Phytochemistry* 50, 1147–1152. doi: 10.1016/S0031-9422(98)00698-0
- Montillet, J., Cacas, J., Garnier, L., Montané, M., Douki, T., Bessoule, J., et al. (2004). The upstream oxylipin profile of *Arabidopsis thaliana*: a tool to scan for oxidative stresses. *Plant J.* 40, 439–451. doi: 10.1111/j.1365-313X.2004.02223.x
- Morello, E., Pérez-Berezo, T., Boisseau, C., Baranek, T., Guillon, A., Bréa, D., et al. (2019). *Pseudomonas aeruginosa* lipoxygenase LoxA contributes to lung infection by altering the host immune lipid signaling. *Front. Microbiol.* 10, 1826. doi: 10.3389/fmicb.2019.01826
- Mosblech, A., Feussner, I., and Heilmann, I. (2009). Oxylipins: structurally diverse metabolites from fatty acid oxidation. *Plant Physiol. Biochem.* 47, 511–517. doi: 10.1016/j.plaphy.2008.12.011
- Niu, M., Steffan, B. N., Fischer, G. J., Venkatesh, N., Raffa, N. L., Wettstein, M. A., et al. (2020). Fungal oxylipins direct programmed developmental switches in filamentous fungi. *Nat. Commun.* 11, 1–13. doi: 10.1038/s41467-020-18999-0
- Nomura, K., Melotto, M., and He, S.-Y. (2005). Suppression of host defense in compatible plant–*Pseudomonas syringae* interactions. *Curr. Opin. Plant Biol.* 8, 361–368. doi: 10.1016/j.pbi.2005.05.005
- Noverr, M. C., Erb-Downward, J. R., and Huffnagle, G. B. (2003). Production of eicosanoids and other oxylipins by pathogenic eukaryotic microbes. *Clin. Microbiol. Rev.* 16, 517–533. doi: 10.1128/CMR.16.3.517-533.2003

- Noverri, M. C., Phare, S. M., Toews, G. B., Coffey, M. J., and Huffnagle, G. B. (2001). Pathogenic yeasts *Cryptococcus neoformans* and *Candida albicans* produce immunomodulatory prostaglandins. *Infect. Immun.* 69, 2957–2963. doi: 10.1128/IAI.69.5.2957-2963.2001
- Obinata, H., Hattori, T., Nakane, S., Tatei, K., and Izumi, T. (2005). Identification of 9-hydroxyoctadecadienoic acid and other oxidized free fatty acids as ligands of the G protein-coupled receptor G2A. *J. Biol. Chem.* 280, 40676–40683. doi: 10.1074/jbc.M507787200
- Oenel, A., Fekete, A., Krischke, M., Faul, S. C., Gresser, G., Havaux, M., et al. (2017). Enzymatic and non-enzymatic mechanisms contribute to lipid oxidation during seed aging. *Plant Cell Physiol.* 58, 925–933. doi: 10.1093/pcp/pcx036
- Okada, K., Abe, H., and Arimura, G. (2015). Jasmonates induce both defense responses and communication in monocotyledonous and dicotyledonous plants. *Plant Cell Physiol.* 56, 16–27. doi: 10.1093/pcp/pcu158
- Oliw, E. H. (2021). Fatty acid dioxygenase-cytochrome P450 fusion enzymes of filamentous fungal pathogens. *Fungal Genet. Biol.* 157, 103623. doi: 10.1016/j.fgb.2021.103623
- Patkar, R. N., Benke, P. I., Qu, Z., Chen, Y. Y. C., Yang, F., Swarup, S., et al. (2015). A fungal monoxygenase-derived jasmonate attenuates host innate immunity. *Nat. Chem. Biol.* 11, 733–740. doi: 10.1038/nchembio.1885
- Pereira, W. E. L., Ferreira, C. B., Caserta, R., Melotto, M., and de Souza, A. A. (2019). *Xylella fastidiosa* subsp. pauca and fastidiosa colonize arabidopsis systemically and induce anthocyanin accumulation in infected leaves. *Phytopathology* 109, 225–232. doi: 10.1094/PHYTO-05-18-0155-FI
- Pohl, C. H., and Kock, J. L. F. (2014). Oxidized fatty acids as inter-kingdom signaling molecules. *Molecules* 19, 1273–1285. doi: 10.3390/molecules19011273
- Prost, I., Dhondt, S., Rothe, G., Vicente, J., Rodriguez, M. J., Kift, N., et al. (2005). Evaluation of the antimicrobial activities of plant oxylipins supports their involvement in defense against pathogens. *Plant Physiol.* 139, 1902–1913. doi: 10.1104/pp.105.066274
- Rapicavoli, J. N., Blanco-Ulate, B., Muszyński, A., Figueroa-Balderas, R., Morales-Cruz, A., Azadi, P., et al. (2018). Lipopolysaccharide O-antigen delays plant innate immune recognition of *Xylella fastidiosa*. *Nat. Commun.* 9, 1–12. doi: 10.1038/s41467-018-02861-5
- Reverberi, M., Punelli, F., Scarpari, M., Camera, E., Zjalic, S., Ricelli, A., et al. (2010). Lipoperoxidation affects ochratoxin A biosynthesis in *Aspergillus ochraceus* and its interaction with wheat seeds. *Appl. Microbiol. Biotechnol.* 85, 1935–1946. doi: 10.1007/s00253-009-2220-4
- Rogers, E. E. (2012). Evaluation of arabidopsis thaliana as a model host for *Xylella fastidiosa*. *Mol. Plant Microbe Interact.* 25, 747–754. doi: 10.1094/MPMI-11-10-0270
- Roper, C., Castro, C., and Ingel, B. (2019). *Xylella fastidiosa*: bacterial parasitism with hallmarks of commensalism. *Curr. Opin. Plant Biol.* 50, 140–147. doi: 10.1016/j.pbi.2019.05.005
- Rupasinghe, S., Schuler, M. A., Kagawa, N., Yuan, H., Lei, L., Zhao, B., et al. (2006). The cytochrome P450 gene family CYP157 does not contain EXXR in the K-helix reducing the absolute conserved P450 residues to a single cysteine. *FEBS Lett.* 580, 6338–6342. doi: 10.1016/j.febslet.2006.10.043
- Ryan, C. A., and Moura, D. S. (2002). Systemic wound signaling in plants: a new perception. *Proc. Natl. Acad. Sci.* 99, 6519–6520. doi: 10.1073/pnas.112196499
- Sakuradani, E., Ando, A., Ogawa, J., and Shimizu, S. (2009). Improved production of various polyunsaturated fatty acids through filamentous fungus *Mortierella alpina* breeding. *Appl. Microbiol. Biotechnol.* 84, 1–10. doi: 10.1007/s00253-009-2076-7
- Scala, V., Camera, E., Ludovici, M., Dall'Asta, C., Cirilini, M., Giorni, P., et al. (2013). *Fusarium verticillioides* and maize interaction *in vitro*: relationship between oxylipin cross-talk and fumonisin synthesis. *World Mycotoxin J.* 6, 343–351. doi: 10.3920/WMJ2012.1527
- Scala, V., Giorni, P., Cirilini, M., Ludovici, M., Visentin, I., Cardinale, F., et al. (2014). LDS1-produced oxylipins are negative regulators of growth, conidiation and fumonisin synthesis in the fungal maize pathogen *Fusarium verticillioides*. *Front. Microbiol.* 5, 669. doi: 10.3389/fmicb.2014.00669
- Scala, V., Pucci, N., Salustri, M., Modesti, V., L'Aurora, A., Scortichini, M., et al. (2019). Bacterial and plant produced lipids can exacerbate the olive quick decline syndrome caused by *xylella*. *bioRxiv*. 867523. doi: 10.1101/867523
- Scala, V., Pucci, N., Salustri, M., Modesti, V., L'Aurora, A., Scortichini, M., et al. (2020). *Xylella fastidiosa* subsp. pauca and olive produced lipids moderate the switch adhesive versus non-adhesive state and viceversa. *PLoS ONE* 15, e0233013. doi: 10.1371/journal.pone.0233013
- Scala, V., Reverberi, M., Salustri, M., Pucci, N., Modesti, V., Lucchesi, S., et al. (2018). Lipid profile of *Xylella fastidiosa* Subsp. pauca associated with the olive quick decline syndrome. *Front. Microbiol.* 9, 1839. doi: 10.3389/fmicb.2018.01839
- Scala, V., Salustri, M., Loreti, S., Pucci, N., Cacciotti, A., Tatulli, G., et al. (2022). Mass spectrometry-based targeted lipidomics and supervised machine learning algorithms in detecting disease, cultivar and treatment biomarkers in *Xylella fastidiosa* subsp. pauca-Infected Olive Trees. *Front. Plant Sci.* 13, 833245. doi: 10.3389/fpls.2022.833245
- Scarpari, M., Punelli, M., Scala, V., Zaccaria, M., Nobili, C., Ludovici, M., et al. (2014). Lipids in *Aspergillus flavus*-maize interaction. *Front. Microbiol.* 5, 74. doi: 10.3389/fmicb.2014.00074
- Shaban, M., Khan, A. H., Noor, E., Malik, W., Ali, H. M. W., Shehzad, M., et al. (2021). A 13-Lipoxygenase, GhLOX2, positively regulates cotton tolerance against *Verticillium dahliae* through JA-mediated pathway. *Gene* 796, 145797. doi: 10.1016/j.gene.2021.145797
- Siebers, M., Brands, M., Wewer, V., Duan, Y., Hölzl, G., and Dörmann, P. (2016). Lipids in plant-microbe interactions. *Biochim. Biophys. Acta Mol. Cell Biol. Lipids* 1861, 1379–1395. doi: 10.1016/j.bbalip.2016.02.021
- Speckbacher, V., Ruzsanyi, V., Martinez-Medina, A., Hinterdobler, W., Doppler, M., Schreiner, U., et al. (2020). The lipoxygenase *lox1* is involved in light- and injury-response, conidiation, and volatile organic compound biosynthesis in the mycoparasitic fungus *trichoderma atroviride*. *Front. Microbiol.* 11, 2004. doi: 10.3389/fmicb.2020.02004
- Stintzi, A. (2000). The Arabidopsis male-sterile mutant, *opr3*, lacks the 12-oxophytodienoic acid reductase required for jasmonate synthesis. *Proc. Natl. Acad. Sci.* 97, 10625–10630. doi: 10.1073/pnas.190264497
- Su, C., Brodowsky, I. D., and Oliw, E. H. (1995). Studies on linoleic acid 8R-dioxygenase and hydroperoxide isomerase of the fungus *Gaeumannomyces graminis*. *Lipids* 30, 43–50. doi: 10.1007/BF02537040
- Su, C., Sahlin, M., and Oliw, E. H. (2000). Kinetics of manganese lipoxygenase with a catalytic mononuclear redox center. *J. Biol. Chem.* 275, 18830–18835. doi: 10.1074/jbc.M001408200
- Truman, W., Bennett, M. H., Kubigstellig, I., Turnbull, C., and Grant, M. (2007). Arabidopsis systemic immunity uses conserved defense signaling pathways and is mediated by jasmonates. *Proc. Natl. Acad. Sci.* 104, 1075–1080. doi: 10.1073/pnas.0605423104
- Tsitsigiannis, D. I., Bok, J. W., Andes, D., Nielsen, K. F., Frisvad, J. C., and Keller, N. P. (2005a). *Aspergillus cyclooxigenase*-like enzymes are associated with prostaglandin production and virulence. *Infect. Immun.* 73, 4548–4559. doi: 10.1128/IAI.73.8.4548-4559.2005
- Tsitsigiannis, D. I., and Keller, N. P. (2007). Oxylipins as developmental and host-fungal communication signals. *Trends Microbiol.* 15, 109–118. doi: 10.1016/j.tim.2007.01.005
- Tsitsigiannis, D. I., Kowieski, T. M., Zarnowski, R., and Keller, N. P. (2005b). Three putative oxylipin biosynthetic genes integrate sexual and asexual development in *Aspergillus nidulans*. *Microbiology* 151, 1809–1821. doi: 10.1099/mic.0.27880-0
- Vance, R. E., Hong, S., Gronert, K., Serhan, C. N., and Mekalanos, J. J. (2004). The opportunistic pathogen *Pseudomonas aeruginosa* carries a secretable arachidonate 15-lipoxygenase. *Proc. Natl. Acad. Sci.* 101, 2135–2139. doi: 10.1073/pnas.0307308101
- Vellosillo, T., Martinez, M., López, M. A., Vicente, J., Cascón, T., Dolan, L., et al. (2007). Oxylipins produced by the 9-lipoxygenase pathway in Arabidopsis regulate lateral root development and defense responses through a specific signaling cascade. *Plant Cell* 19, 831–846. doi: 10.1105/tpc.106.046052
- Vincent, D., Du Fall, L. A., Livk, A., Mathesius, U., Lipscombe, R. J., Oliver, R. P., et al. (2012). A functional genomics approach to dissect the mode of action of the *Stagonospora nodorum* effector protein SnToxA in wheat. *Mol. Plant Pathol.* 13, 467–482. doi: 10.1111/j.1364-3703.2011.00763.x
- Vollenweider, S., Weber, H., Stolz, S., Chételat, A., and Farmer, E. E. (2000). Fatty acid ketodienes and fatty acid ketotrienes: michael addition acceptors that accumulate in wounded and diseased Arabidopsis leaves. *Plant J.* 24, 467–476. doi: 10.1046/j.1365-313x.2000.00897.x
- Wang, F.-F., and Qian, W. (2019). The roles of histidine kinases in sensing host plant and cell-cell communication signal in a phytopathogenic bacterium. *Philos. Trans. R. Soc. B* 374, 20180311. doi: 10.1098/rstb.2018.0311
- Wang, K.-D., Borrego, E. J., Kenerley, C. M., and Kolomiets, M. V. (2020). Oxylipins other than jasmonic acid are xylem-resident signals regulating systemic resistance induced by *Trichoderma virens* in maize. *Plant Cell* 32, 166–185. doi: 10.1105/tpc.19.00487
- Wasternack, C. (2014). Action of jasmonates in plant stress responses and development—applied aspects. *Biotechnol. Adv.* 32, 31–39. doi: 10.1016/j.biotechadv.2013.09.009
- Wasternack, C., and Strnad, M. (2018). Jasmonates: news on occurrence, biosynthesis, metabolism and action of an ancient group of signaling compounds. *Int. J. Mol. Sci.* 19, 2539. doi: 10.3390/ijms19092539

Xia, X.-J., Huang, L.-F., Zhou, Y.-H., Mao, W.-H., Shi, K., Wu, J.-X., et al. (2009). Brassinosteroids promote photosynthesis and growth by enhancing activation of Rubisco and expression of photosynthetic genes in *Cucumis sativus*. *Planta* 230, 1185–1196. doi: 10.1007/s00425-009-1016-1

Yu, Y., Gui, Y., Li, Z., Jiang, C., Guo, J., and Niu, D. (2022). Induced systemic resistance for improving plant immunity

by beneficial microbes. *Plants* 11, 386. doi: 10.3390/plants11030386

Zheng, X., Spivey, N. W., Zeng, W., Liu, P.-P., Fu, Z. Q., Klessig, D. F., et al. (2012). Coronatine promotes *Pseudomonas syringae* virulence in plants by activating a signaling cascade that inhibits salicylic acid accumulation. *Cell Host Microbe* 11, 587–596. doi: 10.1016/j.chom.2012.04.014