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Phytohormonal modulation of the drought stress in soybean: outlook, research progress, and cross-talk

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Phytohormones play vital roles in stress modulation and enhancing the growth of plants. They interact with one another to produce programmed signaling responses by regulating gene expression. Environmental stress, including drought stress, hampers food and energy security. Drought is abiotic stress that negatively affects the productivity of the crops. Abscisic acid (ABA) acts as a prime controller during an acute transient response that leads to stomatal closure. Under long-term stress conditions, ABA interacts with other hormones, such as jasmonic acid (JA), gibberellins (GAs), salicylic acid (SA), and brassinosteroids (BRs), to promote stomatal closure by regulating genetic expression. Regarding antagonistic approaches, cytokinins (CK) and auxins (IAA) regulate stomatal opening. Exogenous application of phytohormone enhances drought stress tolerance in soybean. Thus, phytohormoneproducing microbes have received considerable attention from researchers owing to their ability to enhance drought-stress tolerance and regulate biological processes in plants. The present study was conducted to summarize the role of phytohormones (exogenous and endogenous) and their corresponding microbes in drought stress tolerance in model plant soybean. A total of n=137 relevant studies were collected and reviewed using different research databases.

KEYWORDS

phytohormones, drought stress, soybean, microbes, endogenous



GRAPHICAL ABSTRACT

A schematic representation of the phytohormone modulates drought stress tolerance where (A) represents the endogamous phytohormone modulation, (B) represents the activation of phytohormones via application of the microorganism (C) represents the exogenous application of phytohormones regulates the drought stress tolerance.

1 Introduction

Environmental stress negatively affects plant productivity by up to 70%. Various ecological stresses, including biotic and abiotic stresses, hinder plant development (Mittler, 2006; Choudhury et al., 2017). The agricultural industry is under double pressure. One is environmental stress, and the other is population growth. It is predicted that the global population will reach 10 billion by 2050. Furthermore, by 2050, agricultural crops will have lost up to 30% of their production. In addition, the heat index will reach 52 degrees Celsius. Among abiotic stresses, drought stress poses an alarming risk to agronomical yield, minimizing plant yield and productivity. Moreover, it is a multidimensional stress that arrests plant biomass and energy at molecular and sub-molecular levels. Changes in the climate and landscape temperature, increasing population, and shortage of rain in a particular period enhance the intensity of drought stress (Hasanuzzaman et al., 2013; Shaffique et al., 2022a). One of the main abiotic stresses that has a negative impact on crop growth and production is water deficiency. These modifications are mostly linked to changed metabolic processes, such as reduced or absent photosynthetic pigment production, ion uptake and translocation, glucose biosynthesis, food metabolism, and growth promoter synthesis. The generation of reactive oxygen species (ROS) in response to plant stress is directly correlated with these modifications to metabolic processes and the creation of photosynthetic pigments. Reductions in fresh and dry biomass are a frequent detrimental outcome of water stress on crop plants

(Batool et al., 2022; Madouh and Quoreshi, 2023). Strong relationships exist between the mechanisms of dry matter partitioning and temporal biomass distribution and plant productivity under drought stress. Numerous biochemical mechanisms, such as the fluidity of plasma membranes, the production of osmolytes, lipid peroxidation, the generation of (ROS), the rigidity of cellular membranes, and the activation of various enzymes involved in the oxidative defense system, are all triggered by drought stress. In the past, the production of ROS in different crop species has caused serious harm to proteins, lipid peroxidation (LPO), and other cellular components (Ajithkumar and Panneerselvam, 2014). The lipid membrane and protein were catastrophically affected by drought stress-induced ROS production. The majority of ROS are created during photosynthesis by enzymatic or non-enzymatic mechanisms, including the superoxide radical (O⁻²), hydrogen peroxide (H_2O_2) , singlet oxygen $({}^1O_2)$, and hydroxyl radical (OH^-) . Additionally, they are produced by partial oxidation or reduction in the mitochondrial electron transport system components (Impa et al., 2012). The control of ROS homeostasis involves a number of cellular events that plants use to withstand oxidative stress. As byproducts of several metabolic processes in diverse cellular compartments like chloroplast, mitochondria, and peroxisomes, plants continuously produce a variety of free radicals. Their efficient scavenging by enzymatic and non-enzymatic cascades typically counter balances the formation of ROS in plant cells. ROS have partially reduced forms of ambient oxygen. Because ROS

can harm various biomolecules, including DNA, proteins, and lipids, it can result in oxidative injury, which inhibits plant growth and development. In response to the free radicals oxidation stress increases. However, plant cells reprogram the cellular event via activation of the phytohormones and antioxidant (Figure 1) to mitigate the stress up to a certain limit after that cell shows symptoms of drought stress such as the burning of the leaf, necrosis, fewer pods, lower weight pods, reduced productivity, lower yield crops, inhibit germination, reduced water potential and closure of the stomata to prevent the loss of water etc., (Xiong et al., 2020). As shown in Figure 2.

In 1970, scientists discovered certain chemicals that can optimize plant growth from seed germination to plant development, which were subsequently termed chemical messengers or phytohormones. The concentration of phytohormones required to optimize plant growth is low $(10^{-6}$ to 10^{-5} mol/L), thereby making it difficult to study these phytohormones (Tarakhovskaya et al., 2007; Su et al., 2017). After 1970, research involving phytohormones and their interactions and implications in plant physiology progressed. All phytohormones exhibit distinctive features, nature, and location (Lu and Xu, 2015; Pozo et al., 2015). Their roles have been extensively elucidated not only in plant growth and development but also in stress mitigation (Khan et al., 2012; Egamberdieva et al., 2017). Phytohormones are natural organic s molecules that coordinate with one another to regulate complex signaling pathways and ensure optimal functioning of cellular plant activities (Javid et al., 2011; Fenn and Giovannoni, 2021). Phytohormones exhibit diverse characteristics; for example, ethylene is an alkene, abscisic acid (ABA) is a sesquiterpenoid, gibberellins (GAs) are tetracyclic diterpenoid, jasmonic acid (JA) is a derivative of linoleic acid, and brassinosteroids (BRs) are steroids (Kefeli et al., 2003; Altmann et al., 2020). Endogenous plant hormones are important in the response to drought, in addition to supporting signaling pathways. Plants' responses to osmotic adjustment under stress are heavily mediated by phytohormones. Small signaling molecules known as phytohormones have a substantial impact on nearly every aspect of plant development (Sati et al., 2023; Shaffique et al., 2023b). Furthermore, it is generally understood that a single hormone can influence a wide range of cellular and developmental processes or that numerous hormones can regulate a single function at the same time. Plants are protected and controlled by phytohormones against biotic and abiotic stressors. As a result, the use of phytohormones seeks to broaden agricultural stress research in the future (Singh and Roychoudhury, 2023).

Exogenous phytohormone treatment is a more promising technique for dealing with the negative impacts of drought on sustainable agriculture production. Because of their multifunctionality against abiotic stressors, phytohormones are gaining popularity among plant researchers (Ozturk and Unal, 2023; Swain et al., 2023). However, their use in legume crops (Soybean) is still limited. As a result, the current work expands the use of phytohormones on model plant soybean under drought stress. Deep insights into the physiological, biochemical, and molecular basis of soybean adaptation to drought were also investigated.

Several studies have shown that microbes produce small amounts of phytohormones that enhance plant growth and stress tolerance in several plant species. Numerous studies have documented the usefulness of phytohormone-producing microorganisms in reducing abiotic stress in plants (Shaffique et al., 2022a; Shaffique



A holistic view of the mechanism of oxidation stress and drought stress tolerance represents the plant's cellular reprogramming via activation of the phytohormones to modulate the drought stress.



et al., 2023a). Table 1 lists some examples of phytohormoneproducing bacteria and their capacity to lessen abiotic stress. Numerous studies have documented the beneficial benefits of plant-associated bacteria and IAA generation on promoting plant growth under abiotic stress situations. For example, the bacterial isolate *Enterobacter ludwigii* SH-6 increased plant biomass and improved drought stress tolerance in maize (Shaffique et al., 2022b).

Soybean constitutes an important legume crop that is a rich source of nutrition. Owing to its nutritional value, soybean is in high demand in developing countries. It is rich in protein, oils, and

TABLE 1 Phytohormone-producing bacteria and their action mechanism in drought-stress tolerance in soybean.

| Country, Year, and Ref- erence | Strain | Phytohormones | Mechanism of action (MOA) |
|---|--|-----------------|---|
| South Korea 2014 (Kang et al., 2014) | Pseudomonas putida H-2-3 | ABA GA | Modulating antioxidant defense system |
| India 2019 (Vaishnav and Choudhary, 2019) | Pseudomonas simiae AU | ABA | Genetic expression Overexpression of <i>DREB</i> , <i>PIP</i> , and <i>TIP</i> |
| South Korea 2019 (Bilal et al., 2020) | LHL10 and LHL06 | ABA GA | Over expression of gene <i>DREB</i> , Modulating antioxidant defense system SOD CAT Glutathione, |
| Brazil 2019 (Bulegon et al., 2019) | Azospirillum brasilense | СК | ↑RWC ↑Grain yield ↑Gas exchange |
| 2010 South Korea (Khan et al., 2011) | LH02 | ABA GA SA | Secondary metabolites↑ |
| Iran 2015 (Zahedi and Abbasi, 2015) | Rhizobium japonicum, Azotobacter chroococcum and Azospirillum brasilense | GA ABA | Growth promotion |

The \uparrow and \downarrow represents the increase and decrees of the specific response.

fibers (Specht et al., 2014; Novikova et al., 2022; Razgonova et al., 2022). The soybean (Glycine max) is a crop that is produced all over the globe and is considered an essential ingredient in cuisine due to the large amount of nutrients it contains. Even though there have been considerable improvements in production, environmental stress remains a persistent danger for the soybean crop. There has not been enough research on the soybean, particularly about the modulatory role of phytohormones modulation in the model plant soybean. There is a significant gap in studies between phytohormones and soybean. Environmental stresses, particularly drought stress, considerably affect soybean production (Li et al., 2019; Feng et al., 2020). Herein, we explored the research progress made regarding the effect of phytohormones in mitigating drought stress in soybean. The importance of phytohormones as plant growth regulators have led to a new era of research focusing on phytohormones producing biostimulants to enhance drought-stress tolerance. We further highlighted that phytohormone-producing microbes can mitigate drought stress. We searched the terms "phytohormone, drought, and soybean" on the search engine and downloaded and studied relevant articles for a comprehensive review. There were certain inclusion and exclusion criteria applied to the articles included in this review. All relevant data, including the nature and action mechanism of the phytohormones and gas chromatography-mass spectrometry (GCMS), liquidchromatography-mass spectrometry (LCMS), chromatographic quantification of hormone analysis and hormonal role, and exogenous application data, published until 2022 were included. Non-English data and conference papers were excluded.

2 Review

Phytohormones govern all cellular functions in higher plants and serve an important role in organizing various signal transduction pathways during plant stress response. Their critical function in promoting adaptability to ever-changing environments through growth and differentiation, changes in the source/sink ratio, and nutrient allocation has been well described (Paul et al., 2023; Wani et al., 2023). Abiotic stressors activate signal transduction cascades that are linked to baseline pathways transduced by plant hormones. Aside from the five conventional phytohormones auxins, cytokinins, gibberellins (GA), abscisic acid (ABA), and ethylene (ET), newly found phytohormones include salicylic acid (SA), brassinosteroids, jasmonate (JA), polyamines, and strigolactone. Abscisic acid, salicylic acid, jasmonate, and ethylene are well-known for their anti-inflammatory properties. According to (Zheng et al., 2023), abscisic acid, salicylic acid, jasmonate, and ethylene all contribute positively to plants' ability to withstand stress. For the control of plant defensive responses, auxins, cytokinin, and gibberellins interact with ABA, ethylene, salicylic acid, and jasmonate. The organization of various genes and their regulators involved in stress relief is made possible by hormonal cross-talk. Therefore, it is crucial to comprehend the intricate relationship between cross-talk among phytohormones. Cross-talk between jasmonate and salicylic acid exists, which can interact either negatively or positively by assisting in the development of specific defense responses. At various points along the signaling pathways that result in the control of salicylic acid and jasmonate (Ahmad et al., 2023; Singh and Roychoudhury, 2023)

2.1 Phytohormone abscisic acid triggers drought stress tolerance

ABA is a sesquiterpenoid with the molecular formula $C_{15}H_2O_4$. It has various biological functions and is present in organisms belonging to numerous kingdoms, such as mosses, algae, plants, cyanobacteria, and mammals (Seo and Koshiba, 2002; Kuromori et al., 2018). ABA constitutes an important growth regulator of plant cells. In addition, an important phytohormone controls plant productivity and stress tolerance (Muhammad Aslam et al., 2022). ABA is sometimes referred to as the stress hormone. Reactive oxygen species quickly rise when a plant is under stress, amplifying oxidative stress (Murtaza et al., 2016).

Endogenous ABA levels rise immediately after oxidative stress. ABA acts as a signaling molecule. The ABA involves the regulation of stress-responsive genes, for example, dehydration-responsive element-binding (DREB) proteins and basic leucine zipper (bZIP), during drought stress to improve plant productivity via a stress tolerance mechanism (Liang et al., 2011). A genome-wide association study (GWAS) of soybean plants under stress revealed that ABA either induces or suppresses more than half the genes. The ABA-induced genes included regulatory proteins, transcription factor (Tf), kinases, and phospholipase enzymes for signaling pathways. The ABA-suppressed genes promoted plant growth by regulating guard cells, stomatal conductance, root development, and photosynthesis (Hirayama and Shinozaki, 2007; Yoshida et al., 2014). Stomatal conductance via guard cell regulation is vital for drought stress mitigation in plants. When plants are exposed to drought stress, guard cells regulate osmosis by activating genes that encode proteins to prevent dehydration in all the plant cells (Raghavendra et al., 2010; Ma et al., 2018). Furthermore, ABA intervenes in the high root length density under osmotic stress conditions to reach water present at deeper levels. In addition, ABA modulates root architecture in various ways, including lateral root formation and adaptive morphological alterations, such as reduced xylem diameter to promote the axial hydraulic conductivity of soil under water scarcity stress (Danquah et al., 2014; Sakata et al., 2014).

Six independent studies included herein involved the application of ABA in soybean, and their findings confirmed that exogenous ABA application improved drought stress tolerance in the plant. Initially, independent studies were conducted in 2004 to investigate the effects of exogenous ABA application on soybean during the early reproductive stage. The results indicated that stress exposure increased the ABA levels up to 1.5-fold and decreased the plant's photosynthetic activity. Exogenous ABA application decreased the pod set owing to its direct effect on the metabolic processes in the ovary; however, further studies regarding the mechanism underlying this effect are warranted (Liu et al., 2004). The second independent study indicated that exogenous ABA

application rapidly overexpressed the $G_m RAV$ gene, which is involved in several plant biological processes, including the signaling pathway, to form a complex network in drought stress tolerance (Zhao S.-P. et al., 2017). Exogenous ABA application also improves plants' relative water content (RWC) and root-to-shoot ratio. For this study, two drought genotypes were selected: tolerant and susceptible genotypes. A comparison of the genotypes revealed that the drought-tolerant genotype was more likely to respond to ABA than the drought-susceptible genotype (Xing et al., 2016). Suinong 14 (Glycine max) was experimentally analyzed under different drought conditions to evaluate the effectiveness of ABA on drought-stress tolerance. Drought stress was induced naturally by withholding water from 69.0 MPa to 35.0 MPa at the flowering stage. Subsequently, 1.0-8.0 mg/L ABA was applied exogenously. All secondary metabolites were screened and excluded. The results indicated that up to 2.0 mg/L ABA enhanced soybean's antioxidant capability and drought tolerance (Ruan et al., 2012). This result was further confirmed by another study with similar findings. Furthermore, exogenous ABA application improved osmotic adjustment and leaf water potential but not plant yield (He et al., 2019).

Inoculating plants with plant growth-promoting rhizobacteria (PGPR) mediates ABA production and drought-stress mitigation by activating several relevant cascades. The proposed action mechanism involves ABA-mediated osmotic adjustment regulation and turgor pressure maintenance (Kour and Yadav, 2022).A scientific report by Kang et al. (2014) published in Korea revealed that Pseudomonas putida H-2-3 mitigated drought stress by endogenously producing ABA. Supposedly, ABA is a stressresponsive hormone that is upregulated during stress, and it serves as a signaling molecule to activate the antioxidant defense system in soybean (Kang et al., 2014). In 2019, Vaishnav et al. reported that the Pseudomonas simiae strain AU mitigated drought stress by regulating the endogenous hormone, ultimately activating various gene expressions, such as those of DREB and water transporter genes, including PIP and TIP, which are involved in osmotic stress regulation (Vaishnav and Choudhary, 2019). The results are given in Table 1.

2.2 Jasmonic acid

JA, a chief plant growth regulator, modulates biological processes in plants. It is a derivative of α -linoleic acid, and jasmonate is an active derivative of JA. In response to abiotic stresses, such as drought, heat, chilling and heavy metals, JA enhances stress tolerance in plants. Furthermore, it promotes plant growth (Muñoz-Espinoza et al., 2015; Ghorbel et al., 2021; Kim et al., 2021). The primary features of JA that distinguish it from other hormones include its involvement in fruit ripening, pollen production, tendril coiling, and root hydraulic pressure (Ruan et al., 2019; Wang et al., 2020). The complete genomic analysis of soybean demonstrated that various JA-regulated genes confer stress tolerance. The action mechanism of JA is similar to that of ABA, as both hormones induce stomatal closure. Furthermore, various studies have described that exogenous JA application induces

stomatal closure in soybean (Stintzi et al., 2001; Du et al., 2013; Ghorbel et al., 2021; Wang et al., 2021).

Stress-induced endogenous phytohormone production is a multifarious spectacle that involves several enzymatic pathways (Kaur and Asthir, 2017). Free radical accumulation under drought stress conditions converts unsaturated fatty acid into 12oxo-phytodienoic acid (12-OPDA) and deoxymethylated vegetable dienic acid in peroxisomes. Furthermore, free radicals convert endogenous JA into their corresponding molecules in peroxisomes. Following the conversion, JA and its derivatives, including MeJA, JA-Ile,12-hydroxyjasmonic acid, and cisjasmone, enter the cytoplasm to further confer free radicals (Koo et al., 2009; Savchenko et al., 2014; Kim et al., 2017).12-OPDA is a JA precursor, and various studies have confirmed that its concentration is inversely proportional to the stomatal aperture and stress tolerance of plants. Stress prevents the conversion of 12-OPDA to jasmonate. This delayed conversion might be related to ABA, which induces stress tolerance via stomatal closure (Hu et al., 2009; Ali and Baek, 2020). The findings of several studies have evidenced that exogenous JA application induces stomatal closure, further enhancing the production of antioxidant molecules such as dehydroascorbate acid reductase (DHAR), ascorbate, glutathione reductase (GR), and monodehydroascorbate reductase (MDAR), and consequently, plant stress tolerance. In addition, JA accumulation in roots increases ABA concentration (de Ollas et al., 2015). Therefore, ABA and JA function synergistically to enhance plant stress tolerance (Dathe et al., 1981; Seo et al., 2001; de Ollas et al., 2013).

A pot experiment was performed to elucidate the effectiveness of methyl jasmonate on the stress tolerance of soybean. The exogenous application of 50 µM methyl jasmonate inhibited leaf gas exchange and plant growth and reduced chlorophyll content under extreme drought stress (Anjum S. A.et al., 2011). Jasmonate is involved in a natural molecular signaling pathway that regulates plant progress under stress conditions. Furthermore, it enhances ABA production to control plants' stomatal conductance and water status (Ghassemi-Golezani and Farhangi-Abriz, 2021). Conversely, some studies have reported that exogenous JA application enhances secondary metabolite production, inhibits trypsin, and induces protective genes against plant environmental stresses (Anjum S. et al., 2011). Another research report suggested that 50 µM methyl jasmonate progresses drought-stress tolerance by minimizing lipid peroxidation (LPO) and enhancing the free radicals scavenging system (Hao L. et al., 2013).

In 2014, Kyungpook National University, a leading university in Korea considered the hub of investigating plant microbial interactions, informed that inoculating soybean with *P. putida* H-2-3 enhances the levels of JA and downregulates those of ABA and SA. The results indicated that *P. putida* H-2-3 reprograms chlorophyll and improves hormonal regulation, thereby mitigating drought stress in plants (Kang et al., 2014) In 2019, entophytic fungi LHL10 and LHL06 were reported to synergistically improve drought-stress tolerance by downregulating JA levels synergistically, overexpressing the Gm*DREB* gene, and modulating the Intrinsic defense system by production of antioxidant enzymes as shown in Table 1 (Bilal et al., 2020).

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2.3 Cytokinins

Like other hormones, cytokinins (CKs) are biologically involved in the process of plant growth, development, and stress acclimatization (Werner et al., 2001; Kieber and Schaller, 2014). The chief distinguishing feature of CKs is that it promotes stress acclimatization and adaptation alongside tolerance. Cytokinins play numerous roles in plant growth and morphogenesis. They regulate cell division and interact with auxins to control apical dominance, lateral branching, and the root-shoot ratio in intact plants and tissue culture. They slow leaf senescence and promote dark-grown seedlings' light-independent deetiolation response, including greening. Plants naturally produce a number of cytokinins. They have an adenine base and an isopentenyl side chain of five carbons. Zeatin, specifically trans-zeatin, is the most abundant of them. The concreted functions of isopentenvl transferase, CK oxidase, and CK dehydrogenase maintain the CK hemostatic level in plant cells. Plants are multicellular organisms with complex networks of interactions among its hormones. Recently, it was established that plants exhibit a sophisticated coping mechanism in response to stress involving the CK signal transduction pathway (Ha et al., 2012; O'Brien and Benková, 2013). CKs play a dual role in the stress signaling pathway (Pospíšilová et al., 2000; Kakimoto, 2003); they serves as a negative regulator by inducing the expressions of certain genes such as CKX1, CKX2, CKX3, and CKX4, thereby acclimatizing the plant during stress (Brugière et al., 2003; Niemann et al., 2018), and a positive regulatory effect by increasing the CK levels and delaying senescence in plants (Mok, 2019). When CKs (zeatin riboside) were exogenously applied, they restored the germination potential in Glycine max seeds (Gidrol et al., 1994; Gupta et al., 2000).The inoculation of Azospirillum brasilense in soybean mitigates drought stress by inducing the production of endogenous phytohormones, for example CKs. The results indicated that CKs improve the RWC by 76.96%, gas exchange by up to 860.43%, and grain yield by 19% (Bulegon et al., 2019).

2.4 Gibberellins

GAs are tetracyclic diterpenoid carboxylic acids essential for plant biological functions and stress tolerance. The exact mechanism underlying GA-mediated drought-stress tolerance modulation remains unknown (Takahashi et al., 2012; Hedden and Sponsel, 2015). However, reportedly, GAs exhibit a positive effect on plants throughout their life cycle, promoting cell elongation and division during the juvenile and adult stages (Tanimoto, 2002; Yamaguchi, 2008). GA may also negatively regulate plant stress tolerance by inhibiting its biosynthesis. Furthermore, previous studies have reported that GA content was reduced under water scarcity stress (Fleta-Soriano et al., 2015; Omena-Garcia et al., 2019). Many studies have found that gibberellins significantly improve plant drought tolerance (Iqbal et al., 2022). Drought-responsive element-binding protein (DREB) improves drought tolerance by lowering the expression of gibberellin biosynthesis genes. Reduced GA levels in plants are claimed to improve drought tolerance (Wang et al., 2017; Yang et al., 2021). An environmentally friendly plant growth regulator used all around the world is mepiquat chloride (DPC). DPC is frequently employed to regulate plant geometry in addition to maintaining a balance between nutrition and reproductive growth. The biosynthesis and signal transduction of other plant hormones, such as zeatin and brassinolide, as well as the production and signal transduction of GA were altered by DPC. Additionally, DPC encouraged the formation of flavonoids, increasing drought resistance (Wang et al., 2022).

Similar to other hormones, the exogenous application of GA also recovers the physio-metabolic features of plants and enhances their stress tolerance. Numerous studies have reported that the application of GA hormones enhances plant productivity and mitigates drought stress (Cohen et al., 2009).

In 2011, (Khan et al., 2011) described that the Inoculation of LH02 in soybean improved its stress tolerance by inducing GA and secondary metabolite production. In 2014, (Kang et al., 2014) experimentally observed the effect of P. putida H-2-3 strain on the drought-stress tolerance of soybean. The results revealed that this microbial strain enhanced the GA levels by modulating the antioxidant status and enhancing the plant's stress tolerance by up to 15%.In 2015, three-plant growth-promoting rhizobacteria, namely Rhizobium japonicum, Azotobacter chroococcum, and A. brasilense, were inoculated into the soybean plant. Drought stress was induced naturally via water scarcity. The hormonal status and physio-morphological features of the plant were studied. The results showed that inoculation enhanced drought-stress tolerance by modulating the levels of GAs and ABA (Zahedi and Abbasi, 2015). In 2019, (Bilal et al., 2020) reported that the entophytic fungi LH10 and LH06 synergistically improved drought-stress tolerance by producing up to 300 ng/g GAs. Moreover, the application of these strains downregulated the DRE-binding Tf and decreased ABA levels and oxidation stress in plants.

2.5 Salicylic acid

SA is an essential endogenous phytohormone that regulates protein expression and contributes to the plant defense system. Similar to the other hormones, including ABA and JA, SA is also involved in plant stress tolerance. SA overexpression in response to drought stress is due to two inducible genes, namely PR1 and PR2 (Hayat and Ahmad, 2007; Chen et al., 2009). Salicylic acid (SA) is a key regulator of immunity and programmed cell death in plants. According to early research, greater SA accumulation during resistance gene-mediated defense responses is linked to the initiation of the hypersensitive reaction. In lesion-mimic mutants, SA was also discovered to accumulate to high levels, and in certain cases, this accumulation is necessary for the phenotype of spontaneous cell death. High amounts of SA have been demonstrated to inhibit plant cell death during effector-triggered immunity, indicating that SA has two roles in the regulation of cell death. The drought stress-induced activation of such genes produces a protective effect in plants. The levels of SA may increase up to many folds in response to stress (Shah, 2003; Rao et al., 2012); particularly, exposure to drought stress increases the

SA levels and promotes stomatal closure by generating free radicals (Ilyas et al., 2017; Noreen et al., 2017). However, the effect of SA on drought-stress tolerance in plants remains debatable. Some researchers have reported that SA positively regulates drought-stress tolerance, whereas others have claimed the opposite, thereby rendering its actual effect controversial (Janda et al., 2007; Khan et al., 2015). A small concentration (0.5 mM) of SA induces drought tolerance, whereas a higher concentration (2–3 mM) enhances stress (Ben Ahmed et al., 2009; Zhao P. et al., 2017). SA application causes ROS generation in the chloroplast, thereby reducing stress tolerance via a cascade of events, including antioxidant activation and hormonal modulation. Thus, SA is an important hormone involved in plant stress tolerance. The findings of various studies have evidenced that SA application enhances stress tolerance (Khan et al., 2018).

When inoculated into *G. max*, the *Pseudomonas simiae* strain AU mitigates drought stress by producing phytohormones, particularly SA, and upregulating the Tf of *DREB*, osmoprotactans, and water transporter genes (Vaishnav and Choudhary, 2019).

2.6 Brassinosteroids

BRs are mainly plant steroids based phytohormones involved in regulating plant physiological development. They regulate photosynthesis, photo-morphogenesis, seed germination, fertility, flowering, fruit ripening, grain filling, and leaf senescence (Krishna, 2003; Zhu et al., 2013). They also play an important role in stress tolerance. 24-epibrasonolide EBR is a BR derivative. The action mechanism of BR involves increasing the efficiency of light consumption by the photosystem II (Divi and Krishna, 2009; Hao J. et al., 2013).

Exogenous BR application improves plant photosynthetic characteristics, RWC, and antioxidant enzyme production by reducing hydrogen peroxide and monoaldehyde contents (Shahbaz and Ashraf, 2007; Soares et al., 2020). However, to the best of our knowledge, no studies have implicated microbial BR production in enhancing drought-stress tolerance in soybean. Only a few studies have suggested that BRs are emerging growth regulators that can be used in sustainable agricultural practices. However, although they exhibit a promising effect on plant growth, the role of BRs in stress tolerance remains debatable (Nolan et al., 2017; Alhaithloul and Soliman, 2021).

2.7 Auxins

Auxin, indole acetic acid (IAA), is an important plant hormone that regulates several biological processes, from seed dormancy to development. It plays a primitive role in stress mitigation. Drought stress in plants induces a rapid overexpression of *YUCCA* (YUC) (Woodward and Bartel, 2005; Zhao, 2010), which belongs to the flavin monooxygenase protein family responsible for auxin biosynthesis (Overvoorde et al., 2010; Sauer et al., 2013). The overexpression of this gene family is attributed to apical dormancy, which tall the stem like slender and increases plant drought tolerance. Auxins promote drought tolerance by modulating root architecture (Zarea, 2019; Fadiji et al., 2022). Furthermore, they function in synergy with ABA. Thus, the overexpression of ABA-responsive genes causes ROS accumulation and auxin activation (Van Ha et al., 2013; Wu and Zhang, 2019; Li et al., 2022). Auxins are considered eco-friendly biofertilizer and can be applied in sustainable agricultural practices. Reportedly, auxins exhibit an ameliorative effect on plants that are under stress conditions (Van Ha et al., 2013).

In 2019, (Bulegon et al., 2019) reported that *A. brasilense* inoculation improved drought-stress tolerance in plants by inducing the production of phytohormones, such as auxins, which improved crop yield by up to 19%. In 2021, three bacterial strains, namely AKAD A1-1, AKAD A1-2, and AKAD A1-16, belonging to the *Bacillus* family, were reported to produce auxins. Thus, inoculating these strains can mitigate drought stress in a soybean variety via the modulation of phytohormones (Dubey et al., 2021).

2.8 Ethylene

Ethylene (C_2H_4) is an important gaseous hormone implicated in germination, flowering, fruit ripening, senescence, and stress tolerance. It also plays a dual role in regulating stomatal conductance (Arraes et al., 2015). Osmotic stress induces ROS accumulation in plants, thereby activating ABA-induced stomatal closure. When stress is over, ethylene inhibits the ABA-induced stomatal closure (Sharp, 2002; Yan et al., 2016). Furthermore, ethylene promotes stomatal closure via NADPH oxidase accumulation (Husain et al., 2020). Under stress conditions, ROS accumulation mediates NADPH oxidase production, which promotes stomatal closure (Acharya and Assmann, 2009; Wahab et al., 2022).

Beneficial microbes mitigate drought stress by modulating certain plant hormones, such as ethylene, which is responsible for the plant stress mitigation mechanism (Rehrig, 2010; Ma et al., 2019). The downregulation of ethylene improves abiotic stress tolerance in various plants (Kim et al., 2012; Chandra et al., 2018).

3 Conclusion and future prospective

Plants have evolved to develop sophisticated mechanisms involving phytohormones and phytohormone-producing microbes to combat drought stress. The endogenous modulation, exogenous application of phytohormones and phytohormone-producing microbes strengthens the defense mechanism of plants, as shown in the graphical abstract. Moreover, plants synthesize diverse signaling molecules in response to drought stress. Furthermore, this review highlights that phytohormone-producing microbes enhance drought-stress tolerance in soybean, thereby providing a platform for introducing microbes that can mitigate drought stress by inducing the production of phytohormones and activating their molecular action mechanism. Microbial mediation has led to considerable progress in plant drought-stress tolerance. Although the relevant literature is encouraging in sustainable agronomy, future studies are warranted to elucidate the fundamental mechanism underlying microbe-mediated enhancement of drought-stress tolerance in plants. This review will help scientists develop sustainable agriculture production via phytohormones and their corresponding microbes. Strigolactones are sparse in the scientific literature for soybean growth under drought stress, thus scientists should pay more attention to and conduct more research on strigolactone-generating microorganisms.

Author contributions

SS conceptualized and wrote the original draft and drew the figures. SH did the critical review editing. MI, S-MK and MK did the formatting. Prof I-JL supervised and validated the results. All authors contributed to the article and approved the submitted version.

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References

Acharya, B. R., and Assmann, S. M. (2009). Hormone interactions in stomatal function. *Plant Mol. Biol.* 69, 451-462. doi: 10.1007/s11103-008-9427-0

Ahmad, N., Jiang, Z., Zhang, L., Hussain, I., and Yang, X. (2023). Insights on phytohormonal crosstalk in plant response to nitrogen stress: A focus on plant root growth and development. *Int. J. Mol. Sci.* 24, 3631. doi: 10.3390/ijms24043631

Ajithkumar, I. P., and Panneerselvam, R. (2014). ROS scavenging system, osmotic maintenance, pigment and growth status of Panicum sumatrense roth. under drought stress. *Cell Biochem. biophysics* 68, 587–595. doi: 10.1007/s12013-013-9746-x

Alhaithloul, H., and Soliman, M. H. (2021). "Methyl jasmonate and brassinosteroids: emerging plant growth regulators in plant abiotic stress tolerance and environmental changes," in *Plant growth regulators* (Springer), 173–195.

Ali, M. S., and Baek, K.-H. (2020). Jasmonic acid signaling pathway in response to abiotic stresses in plants. *Int. J. Mol. Sci.* 21, 621. doi: 10.3390/ijms21020621

Altmann, M., Altmann, S., Rodriguez, P. A., Weller, B., Elorduy Vergara, L., Palme, J., et al. (2020). Extensive signal integration by the phytohormone protein network. *Nature* 583, 271–276. doi: 10.1038/s41586-020-2460-0

Anjum, S., Wang, L., Farooq, M., Khan, I., and Xue, L. (2011). Methyl jasmonateinduced alteration in lipid peroxidation, antioxidative defence system and yield in soybean under drought. J. Agron. Crop Sci. 197, 296–301. doi: 10.1111/j.1439-037X.2011.00468.x

Anjum, S. A., Xie, X., Farooq, M., Wang, L., Xue, L., Shahbaz, M., et al. (2011). Effect of exogenous methyl jasmonate on growth, gas exchange and chlorophyll contents of soybean subjected to drought. *Afr. J. Biotechnol.* 10, 9647–9656. doi: 10.5897/ AJB10.2641

Arraes, F. B. M., Beneventi, M. A., Lisei De Sa, M. E., Paixao, J. F. R., Albuquerque, E. V. S., Marin, S. R. R., et al. (2015). Implications of ethylene biosynthesis and signaling in soybean drought stress tolerance. *BMC Plant Biol.* 15, 1–20. doi: 10.1186/s12870-015-0597-z

Batool, M., El-Badri, A. M., Hassan, M. U., Haiyun, Y., Chunyun, W., Zhenkun, Y., et al. (2022). Drought stress in Brassica napus: effects, tolerance mechanisms, and management strategies. *J. Plant Growth Regul.*, 1–25.

Ben Ahmed, H., Abidi, F., Manaa, A., Mimouni, H., and Zid, E. (2009). Salicylic acid induced changes on some physiological parameters in tomato grown under salinity.

Bilal, S., Shahzad, R., Imran, M., Jan, R., Kim, K. M., and Lee, I.-J. (2020). Synergistic association of endophytic fungi enhances Glycine max L. resilience to combined abiotic stresses: Heavy metals, high temperature and drought stress. *Ind. Crops Products* 143, 111931.

Brugière, N., Jiao, S., Hantke, S., Zinselmeier, C., Roessler, J. A., Niu, X., et al. (2003). Cytokinin oxidase gene expression in maize is localized to the vasculature, and is induced by cytokinins, abscisic acid, and abiotic stress. *Plant Physiol.* 132, 1228–1240. doi: 10.1104/pp.102.017707

Bulegon, L. G., Guimarães, V. F., Battistus, A. G., Inagaki, A. M., and Costa, N. V. D. (2019). Mitigation of drought stress effects on soybean gas exchanges induced by Azospirillum brasilense and plant regulators. *Pesquisa Agropecuária Trop.* 49. doi: 10.1590/1983-40632019v4952807 2022R1A2C1008993) Funding : Republic of Korea and Korea Basic Science Institute (National Research Facilities and Equipment Center) grant funded by the Ministry of Education (NRF-2021R1A6C101A416).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Chandra, D., Srivastava, R., Glick, B. R., and Sharma, A. K. (2018). Drought-tolerant Pseudomonas spp. improve the growth performance of finger millet (Eleusine coracana (L.) Gaertn.) under non-stressed and drought-stressed conditions. *Pedosphere* 28, 227– 240. doi: 10.1016/S1002-0160(18)60013-X

Chen, Z., Zheng, Z., Huang, J., Lai, Z., and Fan, B. (2009). Biosynthesis of salicylic acid in plants. *Plant Signaling Behav.* 4, 493–496. doi: 10.4161/psb.4.6.8392

Choudhury, F. K., Rivero, R. M., Blumwald, E., and Mittler, R. (2017). Reactive oxygen species, abiotic stress and stress combination. *Plant J.* 90, 856–867. doi: 10.1111/tpj.13299

Cohen, A. C., Travaglia, C. N., Bottini, R., and Piccoli, P. N. (2009). Participation of abscisic acid and gibberellins produced by endophytic Azospirillum in the alleviation of drought effects in maize. *Botany* 87, 455–462. doi: 10.1139/B09-023

Danquah, A., De Zélicourt, A., Colcombet, J., and Hirt, H. (2014). The role of ABA and MAPK signaling pathways in plant abiotic stress responses. *Biotechnol. Adv.* 32, 40–52. doi: 10.1016/j.biotechadv.2013.09.006

Dathe, W., Rönsch, H., Preiss, A., SChade, W., Sembdner, G., and Schreiber, K. (1981). Endogenous plant hormones of the broad bean, Vicia faba L.(-)-jasmonic acid, a plant growth inhibitor in pericarp. *Planta* 153, 530–535. doi: 10.1007/BF00385537

de Ollas, C., Arbona, V., and Gómez-Cadenas, A. (2015). Jasmonic acid interacts with abscisic acid to regulate plant responses to water stress conditions. *Plant Signaling Behav.* 10, e1078953. doi: 10.1080/15592324.2015.1078953

de Ollas, C., Hernando, B., Arbona, V., and Gómez-Cadenas, A. (2013). Jasmonic acid transient accumulation is needed for abscisic acid increase in citrus roots under drought stress conditions. *Physiologia plantarum* 147, 296–306. doi: 10.1111/j.1399-3054.2012.01659.x

Divi, U. K., and Krishna, P. (2009). Brassinosteroid: a biotechnological target for enhancing crop yield and stress tolerance. *New Biotechnol.* 26, 131–136. doi: 10.1016/ j.nbt.2009.07.006

Du, H., Liu, H., and Xiong, L. (2013). Endogenous auxin and jasmonic acid levels are differentially modulated by abiotic stresses in rice. *Front. Plant Sci.* 4, 397. doi: 10.3389/fpls.2013.00397

Dubey, A., Saiyam, D., Kumar, A., Hashem, A., Abd_Allah, E. F., and Khan, M. L. (2021). Bacterial root endophytes: Characterization of their competence and plant growth promotion in soybean (Glycine max (L.) Merr.) under drought stress. *Int. J. Environ. Res. Public Health* 18, 931.

Egamberdieva, D., Wirth, S. J., Alqarawi, A. A., Abd_Allah, E. F., and Khan, M. L. (2017). Phytohormones and beneficial microbes: essential components for plants to balance stress and fitness. *Front. Microbiol.* 8, 2104. doi: 10.3389/fmicb.2017.02104

Fadiji, A. E., Santoyo, G., Yadav, A. N., and Babalola, O. O. (2022). Efforts towards overcoming drought stress in crops: Revisiting the mechanisms employed by plant growth-promoting bacteria. *Front. Microbiol.* 13. doi: 10.3389/fmicb.2022.962427

Feng, Z., Ding, C., Li, W., Wang, D., and Cui, D. (2020). Applications of metabolomics in the research of soybean plant under abiotic stress. *Food Chem.* 310, 125914. doi: 10.1016/j.foodchem.2019.125914

Fenn, M. A., and Giovannoni, J. J. (2021). Phytohormones in fruit development and maturation. *Plant J.* 105, 446–458. doi: 10.1111/tpj.15112

Fleta-Soriano, E., Pintó-Marijuan, M., and Munné-Bosch, S. (2015). Evidence of drought stress memory in the facultative CAM, Aptenia cordifolia: possible role of phytohormones. *PloS One* 10, e0135391. doi: 10.1371/journal.pone.0135391

Ghassemi-Golezani, K., and Farhangi-Abriz, S. (2021). "Plant responses to exogenous salicylic and jasmonic acids under drought stress," in *Jasmonates and salicylates signaling in plants*. (Springer), 65–85.

Ghorbel, M., Brini, F., Sharma, A., and Landi, M. (2021). Role of jasmonic acid in plants: The molecular point of view. *Plant Cell Rep.* 40, 1471–1494. doi: 10.1007/s00299-021-02687-4

Gidrol, X., Lin, W. S., Dégousée, N., Yip, S. F., and Kush, A. (1994). Accumulation of reactive oxygen species and oxidation of cytokinin in germinating soybean seeds. *Eur. J. Biochem.* 224, 21–28. doi: 10.1111/j.1432-1033.1994.tb19990.x

Gupta, N., Gupta, S., and Kumar, A. (2000). Exogenous cytokinin application increases cell membrane and chlorophyll stability in wheat (Triticum aestivum L.). *Cereal Res. Commun.* 28, 287–291. doi: 10.1007/BF03543606

Ha, S., Vankova, R., Yamaguchi-Shinozaki, K., Shinozaki, K., and Tran, L.-S. P. (2012). Cytokinins: metabolism and function in plant adaptation to environmental stresses. *Trends Plant Sci.* 17, 172–179. doi: 10.1016/j.tplants.2011.12.005

Hao, L., Wang, Y., Zhang, J., Xie, Y., Zhang, M., Duan, L., et al. (2013). Coronatine enhances drought tolerance via improving antioxidative capacity to maintaining higher photosynthetic performance in soybean. *Plant Sci.* 210, 1–9. doi: 10.1016/j.plantsci.2013.05.006

Hao, J., Yin, Y., and Fei, S.-Z. (2013). Brassinosteroid signaling network: implications on yield and stress tolerance. *Plant Cell Rep.* 32, 1017–1030. doi: 10.1007/s00299-013-1438-x

Hasanuzzaman, M., Nahar, K., Gill, S. S., and Fujita, M. (2013). "Drought stress responses in plants, oxidative stress, and antioxidant defense," in *Climate change and plant abiotic stress tolerance*, 209–250.

Hayat, S., and Ahmad, A. (2007). Salicylic acid-a plant hormone (Springer Science & Business Media).

He, J., Jin, Y., Palta, J. A., Liu, H.-Y., Chen, Z., and Li, F.-M. (2019). Exogenous ABA induces osmotic adjustment, improves leaf water relations and water use efficiency, but not yield in soybean under water stress. *Agronomy* 9, 395. doi: 10.3390/agronomy9070395

Hedden, P., and Sponsel, V. (2015). A century of gibberellin research. J. Plant Growth Regul. 34, 740–760. doi: 10.1007/s00344-015-9546-1

Hirayama, T., and Shinozaki, K. (2007). Perception and transduction of abscisic acid signals: keys to the function of the versatile plant hormone ABA. *Trends Plant Sci.* 12, 343–351. doi: 10.1016/j.tplants.2007.06.013

Hu, X., Wansha, L., Chen, Q., and Yang, Y. (2009). Early signals transduction linking the synthesis of jasmonic acid in plant. *Plant Signaling Behav.* 4, 696–697. doi: 10.4161/ psb.4.8.9181

Husain, T., Fatima, A., Suhel, M., Singh, S., Sharma, A., Prasad, S. M., et al. (2020). A brief appraisal of ethylene signaling under abiotic stress in plants. *Plant Signaling Behav.* 15, 1782051. doi: 10.1080/15592324.2020.1782051

Ilyas, N., Gull, R., Mazhar, R., Saeed, M., Kanwal, S., Shabir, S., et al. (2017). Influence of salicylic acid and jasmonic acid on wheat under drought stress. *Commun. Soil Sci. Plant Anal.* 48, 2715–2723. doi: 10.1080/00103624.2017.1418370

Impa, S., Nadaradjan, S., and Jagadish, S. (2012). "Drought stress induced reactive oxygen species and anti-oxidants in plants," in *Abiotic stress responses in plants: metabolism, productivity and sustainability*, 131–147.

Iqbal, S., Wang, X., Mubeen, I., Kamran, M., Kanwal, I., Díaz, G. A., et al. (2022). Phytohormones trigger drought tolerance in crop plants: outlook and future perspectives. *Front. Plant Sci.* 12, 3378. doi: 10.3389/fpls.2021.799318

Janda, T., Horváth, E., Szalai, G., and Paldi, E. (2007). "Role of salicylic acid in the induction of abiotic stress tolerance," in *Salicylic acid: A plant hormone*. (Springer), 91–150.

Javid, M. G., Sorooshzadeh, A., Moradi, F., Modarres Sanavy, S. A. M., and Allahdadi, I. (2011). The role of phytohormones in alleviating salt stress in crop plants. *Aust. J. Crop Sci.* 5, 726–734.

Kakimoto, T. (2003). Perception and signal transduction of cytokinins. Annu. Rev. Plant Biol. 54, 605–627. doi: 10.1146/annurev.arplant.54.031902.134802

Kang, S.-M., Radhakrishnan, R., Khan, A. L., Kim, M.-J., Park, J.-M., Kim, B.-R., et al. (2014). Gibberellin secreting rhizobacterium, Pseudomonas putida H-2-3 modulates the hormonal and stress physiology of soybean to improve the plant growth under saline and drought conditions. *Plant Physiol. Biochem.* 84, 115–124. doi: 10.1016/ j.plaphy.2014.09.001

Kaur, G., and Asthir, B. (2017). Molecular responses to drought stress in plants. *Biol. Plantarum* 61, 201–209. doi: 10.1007/s10535-016-0700-9

Kefeli, V. I., Kefeli, V., Kalevitch, M., and Borsari, B. (2003). Natural growth inhibitors and phytohormones in plants and environment. (Springer Science & Business Media).

Khan, M. I. R., Fatma, M., Per, T. S., Anjum, N. A., and Khan, N. A. (2015). Salicylic acid-induced abiotic stress tolerance and underlying mechanisms in plants. *Front. Plant Sci.* 6, 462. doi: 10.3389/fpls.2015.00462

Khan, A. L., Hamayun, M., Kim, Y.-H., Kang, S.-M., Lee, J.-H., and Lee, I.-J. (2011). Gibberellins producing endophytic Aspergillus fumigatus sp. LH02 influenced endogenous phytohormonal levels, isoflavonoids production and plant growth in salinity stress. *Process Biochem.* 46, 440–447. doi: 10.1016/j.procbio.2010.09.013

Khan, N. A., Nazar, R., Iqbal, N., and Anjum, N. A. (2012). *Phytohormones and abiotic stress tolerance in plants*. (Springer Science & Business Media).

Khan, N., Zandi, P., Ali, S., Mehmood, A., Adnan Shahid, M., and Yang, J. (2018). Impact of salicylic acid and PGPR on the drought tolerance and phytoremediation potential of Helianthus annus. *Front. Microbiol.* 9, 2507. doi: 10.3389/fmicb.2018.02507

Kieber, J. J., and Schaller, G. E. (2014). Cytokinins. Arabidopsis Book/American Soc. Plant Biologists 12. doi: 10.1199/tab.0168

Kim, Y.-C., Glick, B. R., Bashan, Y., and Ryu, C.-M. (2012). "Enhancement of plant drought tolerance by microbes," in *Plant responses to drought stress*. (Springer), 383–413.

Kim, H., Seomun, S., Yoon, Y., and Jang, G. (2021). Jasmonic acid in plant abiotic stress tolerance and interaction with abscisic acid. *Agronomy* 11, 1886. doi: 10.3390/ agronomy11091886

Kim, J.-M., To, T. K., Matsui, A., Tanoi, K., Kobayashi, N. I., Matsuda, F., et al. (2017). Acetate-mediated novel survival strategy against drought in plants. *Nat. Plants* 3, 1–7.

Koo, A. J., Gao, X., Daniel Jones, A., and Howe, G. A. (2009). A rapid wound signal activates the systemic synthesis of bioactive jasmonates in Arabidopsis. *Plant J.* 59, 974–986. doi: 10.1111/j.1365-313X.2009.03924.x

Kour, D., and Yadav, A. N. (2022). Bacterial mitigation of drought stress in plants: Current perspectives and future challenges. *Curr. Microbiol.* 79, 1–19. doi: 10.1007/ s00284-022-02939-w

Krishna, P. (2003). Brassinosteroid-mediated stress responses. J. Plant Growth Regul. 22, 289–297. doi: 10.1007/s00344-003-0058-z

Kuromori, T., Seo, M., and Shinozaki, K. (2018). ABA transport and plant water stress responses. *Trends Plant Sci.* 23, 513–522. doi: 10.1016/j.tplants.2018.04.001

Li, M., Cao, L., Mwimba, M., Zhou, Y., Li, L., Zhou, M., et al. (2019). Comprehensive mapping of abiotic stress inputs into the soybean circadian clock. *Proc. Natl. Acad. Sci.* 116, 23840–23849. doi: 10.1073/pnas.1708508116

Li, G., Wang, Q., Lu, L., Wang, S., Chen, X., Khan, M. H. U., et al. (2022). Identification of the soybean small auxin upregulated RNA (SAUR) gene family and specific haplotype for drought tolerance. *Biologia* 77, 1197–1217. doi: 10.1007/s11756-022-01010-0

Liang, F.-S., Ho, W. Q., and Crabtree, G. R. (2011). Engineering the ABA plant stress pathway for regulation of induced proximity. *Sci. Signaling* 4, rs2–rs2.

Liu, F., Jensen, C. R., and Andersen, M. N. (2004). Pod set related to photosynthetic rate and endogenous ABA in soybeans subjected to different water regimes and exogenous ABA and BA at early reproductive stages. *Ann. Bot.* 94, 405–411. doi: 10.1093/aob/mch157

Lu, Y., and Xu, J. (2015). Phytohormones in microalgae: a new opportunity for microalgal biotechnology? *Trends Plant Sci.* 20, 273–282.

Ma, Y., Cao, J., He, J., Chen, Q., Li, X., and Yang, Y. (2018). Molecular mechanism for the regulation of ABA homeostasis during plant development and stress responses. *Int. J. Mol. Sci.* 19, 3643. doi: 10.3390/ijms19113643

Ma, Y., Vosátka, M., and Freitas, H. (2019). Beneficial microbes alleviate.

Madouh, T. A., and Quoreshi, A. M. (2023). The function of arbuscular mycorrhizal fungi associated with drought stress resistance in native plants of arid desert ecosystems: A review. *Diversity* 15, 391. doi: 10.3390/d15030391

Mittler, R. (2006). Abiotic stress, the field environment and stress combination. *Trends Plant Sci.* 11, 15–19. doi: 10.1016/j.tplants.2005.11.002

Mok, M. C. (2019). "Cytokinins and plant development—an overview," in *Cytokinins*, 155–166.

Muhammad Aslam, M., Waseem, M., Jakada, B. H., Okal, E. J., Lei, Z., Saqib, H. S. A., et al. (2022). Mechanisms of abscisic acid-mediated drought stress responses in plants. *Int. J. Mol. Sci.* 23, 1084. doi: 10.3390/ijms23031084

Muñoz-Espinoza, V. A., López-Climent, M. F., Casaretto, J. A., and Gómez-Cadenas, A. (2015). Water stress responses of tomato mutants impaired in hormone biosynthesis reveal abscisic acid, jasmonic acid and salicylic acid interactions. *Front. Plant Sci.* 6, 997.

Murtaza, G., Rasool, F., Habib, R., Javed, T., Sardar, K., Ayub, M. M., et al. (2016). A review of morphological, physiological and biochemical responses of plants under drought stress conditions. *Imp. J. Interdiscip. Res.* 2, 1600–1606.

Niemann, M. C., Weber, H., Hluska, T., Leonte, G., Anderson, S. M., Novák, O., et al. (2018). The cytokinin oxidase/dehydrogenase CKX1 is a membrane-bound protein requiring homooligomerization in the endoplasmic reticulum for its cellular activity. *Plant Physiol.* 176, 2024–2039. doi: 10.1104/pp.17.00925

Nolan, T., Chen, J., and Yin, Y. (2017). Cross-talk of Brassinosteroid signaling in controlling growth and stress responses. *Biochem. J.* 474, 2641–2661. doi: 10.1042/BCJ20160633

Noreen, S., Fatima, K., Athar, H., Ahmad, S., and Hussain, K. (2017). Enhancement of physio-biochemical parameters of wheat through exogenous application of salicylic acid under drought stress. *JAPS: J. Anim. Plant Sci.* 27.

Novikova, L., Seferova, I., Matvienko, I., Shchedrina, Z., and Vishnyakova, M. (2022). Photoperiod and temperature sensitivity in early soybean accessions from the VIR

collection in Leningrad Province of the Russian Federation. *Turkish J. Agric. Forestry* 46, 947–954. doi: 10.55730/1300-011X.3055

O'Brien, J. A., and Benková, E. (2013). Cytokinin cross-talking during biotic and abiotic stress responses. *Front. Plant Sci.* 4, 451.

Omena-Garcia, R. P., Martins, A. O., Medeiros, D. B., Vallarino, J. G., Ribeiro, D. M., Fernie, A. R., et al. (2019). Growth and metabolic adjustments in response to gibberellin deficiency in drought stressed tomato plants. *Environ. Exp. Bot.* 159, 95–107. doi: 10.1016/j.envexpbot.2018.12.011

Overvoorde, P., Fukaki, H., and Beeckman, T. (2010). Auxin control of root development. Cold Spring Harbor Perspect. Biol. 2, a001537. doi: 10.1101/ cshperspect.a001537

Ozturk, M., and Unal, B. T. (2023). "Exogenous application of phytohormones and phytometabolites to plants to alleviate the effects of drought stress," in *Phytohormones and stress responsive secondary metabolites* (Elsevier), 1–12.

Paul, P., Sharma, S., and Pandey, R. (2023). Phosphorus scavenging and remobilization from root cell walls under combined nitrogen and phosphorus stress is regulated by phytohormones and nitric oxide cross-talk in wheat. *J. Plant Growth Regul.* 42, 1614–1630. doi: 10.1007/s00344-022-10646-w

Pospíšilová, J., Synková, H., and Rulcová, J. (2000). Cytokinins and water stress. *Biol.* plantarum 43, 321–328. doi: 10.1023/A:1026754404857

Pozo, M. J., López-Ráez, J. A., Azcón-Aguilar, C., and García-Garrido, J. M. (2015). Phytohormones as integrators of environmental signals in the regulation of mycorrhizal symbioses. *New Phytol.* 205, 1431–1436. doi: 10.1111/nph.13252

Raghavendra, A. S., Gonugunta, V. K., Christmann, A., and Grill, E. (2010). ABA perception and signalling. *Trends Plant Sci.* 15, 395-401. doi: 10.1016/j.tplants.2010.04.006

Rao, S., Qayyum, A., Razzaq, A., Ahmad, M., Mahmood, I., and Sher, A. (2012). Role of foliar application of salicylic acid and l-tryptophan in drought tolerance of maize. *J. Anim. Plant Sci.* 22, 768–772.

Razgonova, M. P., Zinchenko, Y. N., Kozak, D. K., Kuznetsova, V. A., Zakharenko, A. M., Ercisli, S., et al. (2022). Autofluorescence-based investigation of spatial distribution of phenolic compounds in soybeans using confocal laser microscopy and a high-resolution mass spectrometric approach. *Molecules* 27, 8228. doi: 10.3390/molecules27238228

Rehrig, E. M. (2010). The role of ERF transcription factors in defenses against specialist and generalist herbivores in Arabidopsis thaliana (University of Missouri-Columbia).

Ruan, Y., Dong, S., Liu, L., Sun, C., Wang, L., Guo, Q., et al. (2012). Effect of exogenous abscisic acid (ABA) on the physiological characteristics during soybean flowering under drought stress. *Soybean Sci.* 31, 385–394.

Ruan, J., Zhou, Y., Zhou, M., Yan, J., Khurshid, M., Weng, W., et al. (2019). Jasmonic acid signaling pathway in plants. *Int. J. Mol. Sci.* 20, 2479. doi: 10.3390/ijms20102479

Sakata, Y., Komatsu, K., and Takezawa, D. (2014). "ABA as a universal plant hormone," in *Progress in botany*. (Springer), 57-96.

Sati, D., Pande, V., Pandey, S. C., and Samant, M. (2023). Recent advances in PGPR and molecular mechanisms involved in drought stress resistance. *J. Soil Sci. Plant Nutr.* 23, 106–124. doi: 10.1007/s42729-021-00724-5

Sauer, M., Robert, S., and Kleine-Vehn, J. (2013). Auxin: simply complicated. J. Exp. Bot. 64, 2565–2577. doi: 10.1093/jxb/ert139

Savchenko, T., Kolla, V. A., Wang, C.-Q., Nasafi, Z., Hicks, D. R., Phadungchob, B., et al. (2014). Functional convergence of oxylipin and abscisic acid pathways controls stomatal closure in response to drought. *Plant Physiol.* 164, 1151–1160. doi: 10.1104/pp.113.234310

Seo, M., and Koshiba, T. (2002). Complex regulation of ABA biosynthesis in plants. *Trends Plant Sci.* 7, 41–48. doi: 10.1016/S1360-1385(01)02187-2

Seo, H. S., Song, J. T., Cheong, J.-J., Lee, Y.-H., Lee, Y.-W., Hwang, I., et al. (2001). Jasmonic acid carboxyl methyltransferase: a key enzyme for jasmonate-regulated plant responses. *Proc. Natl. Acad. Sci.* 98, 4788–4793. doi: 10.1073/pnas.081557298

Shaffique, S., Hussain, S., Kang, S.-M., Imran, M., Kwon, E.-H., Khan, M. A., et al. (2023a). Recent progress on the microbial mitigation of heavy metal stress in soybean: overview and implications. *Front. Plant Sci.* 14, 1188856. doi: 10.3389/fpls.2023.1188856

Shaffique, S., Kang, S.-M., Hoque, M. I. U., Imran, M., Aaqil Khan, M., and Lee, I.-J. (2023b). Research progress in soybean by phytohormone modulation and metal chelation over the past decade. *Agriculture* 13, 1325. doi: 10.3390/agriculture13071325

Shaffique, S., Khan, M. A., Imran, M., Kang, S.-M., Park, Y.-S., Wani, S. H., et al. (2022a). Research progress in the field of microbial mitigation of drought stress in plants. *Front. Plant Sci.* 13, 870626. doi: 10.3389/fpls.2022.870626

Shaffique, S., Khan, M. A., Wani, S. H., Imran, M., Kang, S.-M., Pande, A., et al. (2022b). Biopriming of maize seeds with a novel bacterial strain SH-6 to enhance drought tolerance in South Korea. *Plants* 11, 1674. doi: 10.3390/plants11131674

Shah, J. (2003). The salicylic acid loop in plant defense. Curr. Opin. Plant Biol. 6, 365–371. doi: 10.1016/S1369-5266(03)00058-X

Shahbaz, M., and Ashraf, M. (2007). Influence of exogenous application of brassinosteroid on growth and mineral nutrients of wheat (Triticum aestivum L.) under saline conditions. *Pakistan J. Bot.* 39, 513.

Sharp, R. (2002). Interaction with ethylene: changing views on the role of abscisic acid in root and shoot growth responses to water stress. *Plant Cell Environ.* 25, 211–222. doi: 10.1046/j.1365-3040.2002.00798.x

Singh, A., and Roychoudhury, A. (2023). Abscisic acid in plants under abiotic stress: crosstalk with major phytohormones. *Plant Cell Rep.*, 1–14. doi: 10.1007/s00299-023-03013-w

Soares, T. F. S. N., Dos Santos Dias, D. C. F., Oliveira, A. M. S., Ribeiro, D. M., and Dos Santos Dias, L. A. (2020). Exogenous brassinosteroids increase lead stress tolerance in seed germination and seedling growth of Brassica juncea L. *Ecotoxicology Environ. Saf.* 193, 110296. doi: 10.1016/j.ecoenv.2020.110296

Specht, J. E., Diers, B. W., Nelson, R. L., De Toledo, J. F. F., Torrion, J. A., and Grassini, P. (2014). "Soybean," in Yield gains in major US field crops, vol. 33., 311–355.

Stintzi, A., Weber, H., Reymond, P., Browse, J., and Farmer, E. E. (2001). Plant defense in the absence of jasmonic acid: the role of cyclopentenones. *Proc. Natl. Acad. Sci.* 98, 12837–12842. doi: 10.1073/pnas.211311098

Su, Y., Xia, S., Wang, R., and Xiao, L. (2017). Phytohormonal quantification based on biological principles. *Hormone Metab. Signaling Plants* 13, 431–470. doi: 10.1016/B978-0-12-811562-6.00013-X

Swain, R., Sahoo, S., Behera, M., and Rout, G. R. (2023). Instigating prevalent abiotic stress resilience in crop by exogenous application of phytohormones and nutrient. *Front. Plant Sci.* 14, 1104874. doi: 10.3389/fpls.2023.1104874

Takahashi, N., Phinney, B. O., and Macmillan, J. (2012). *Gibberellins*. (Springer Science & Business Media).

Tanimoto, E. (2002). "Gibberellins," in Plant roots (CRC Press), 638-655.

Tarakhovskaya, E., Maslov, Y. I., and Shishova, M. (2007). Phytohormones in algae. Russian J. Plant Physiol. 54, 163–170. doi: 10.1134/S1021443707020021

Vaishnav, A., and Choudhary, D. K. (2019). Regulation of drought-responsive gene expression in Glycine max l. Merrill is mediated through Pseudomonas simiae strain AU. *J. Plant Growth Regul.* 38, 333–342. doi: 10.1007/s00344-018-9846-3

Van Ha, C., Le, D. T., Nishiyama, R., Watanabe, Y., Sulieman, S., Tran, U. T., et al. (2013). The auxin response factor transcription factor family in soybean: genome-wide identification and expression analyses during development and water stress. *DNA Res.* 20, 511–524. doi: 10.1093/dnares/dst027

Wahab, A., Abdi, G., Saleem, M. H., Ali, B., Ullah, S., Shah, W., et al. (2022). Plants' physio-biochemical and phyto-hormonal responses to alleviate the adverse effects of drought stress: A comprehensive review. *Plants* 11, 1620.

Wang, Y., Mostafa, S., Zeng, W., and Jin, B. (2021). Function and mechanism of jasmonic acid in plant responses to abiotic and biotic stresses. *Int. J. Mol. Sci.* 22, 8568. doi: 10.3390/ijms22168568

Wang, H., Pan, Y., and Chen, Y. (2017). Comparison of three drought indices and their evolutionary characteristics in the arid region of northwestern China. *Atmospheric Sci. Lett.* 18, 132–139. doi: 10.1002/asl.735

Wang, J., Song, L., Gong, X., Xu, J., and Li, M. (2020). Functions of jasmonic acid in plant regulation and response to abiotic stress. *Int. J. Mol. Sci.* 21, 1446. doi: 10.3390/ ijms21041446

Wang, X., Zhou, Q., Wang, X., Song, S., Liu, J., and Dong, S. (2022). Mepiquat chloride inhibits soybean growth but improves drought resistance. *Front. Plant Sci.* 13, 982415. doi: 10.3389/fpls.2022.982415

Wani, A. K., Akhtar, N., Garg, S. S., Rahayu, F., Prakash, A., and Singh, R. (2023). "Strigolactones in plants: web networking with phytohormonal crosstalk," in *Strigolactones, alkamides and karrikins in plants.* (CRC Press), 59–73.

Werner, T., Motyka, V., Strnad, M., and Schmülling, T. (2001). Regulation of plant growth by cytokinin. *Proc. Natl. Acad. Sci.* 98, 10487–10492. doi: 10.1073/pnas.171304098

Woodward, A. W., and Bartel, B. (2005). Auxin: regulation, action, and interaction. *Ann. Bot.* 95, 707–735. doi: 10.1093/aob/mci083

Wu, Z.-L., and Zhang, Y.-Z. (2019). Effects of exogenous auxin on physiological and biochemical characteristics of soybean under PEG simulated drought stress. *Hubei Agric. Sci.* 58, 16.

Xing, X., Jiang, H., Zhou, Q., Xing, H., Jiang, H., and Wang, S. (2016). Improved drought tolerance by early IAA-and ABA-dependent H2O2 accumulation induced by α -naphthaleneacetic acid in soybean plants. *Plant Growth Regul.* 80, 303–314. doi: 10.1007/s10725-016-0167-x

Xiong, C., Zhao, S., Yu, X., Sun, Y., Li, H., Ruan, C., et al. (2020). Yellowhorn drought-induced transcription factor XsWRKY20 acts as a positive regulator in drought stress through ROS homeostasis and ABA signaling pathway. *Plant Physiol. Biochem.* 155, 187–195. doi: 10.1016/j.plaphy.2020.06.037

Yamaguchi, S. (2008). Gibberellin metabolism and its regulation. Annu. Rev. Plant Biol. 59, 225–251. doi: 10.1146/annurev.arplant.59.032607.092804

Yan, Q., Cui, X., Lin, S., Gan, S., Xing, H., and Dou, D. (2016). GmCYP82A3, a soybean cytochrome P450 family gene involved in the jasmonic acid and ethylene signaling pathway, enhances plant resistance to biotic and abiotic stresses. *PloS One* 11, e0162253. doi: 10.1371/journal.pone.0162253

Yang, X., Lu, M., Wang, Y., Wang, Y., Liu, Z., and Chen, S. (2021). Response mechanism of plants to drought stress. *Horticulturae* 7, 50. doi: 10.3390/horticulturae7030050

Yoshida, T., Mogami, J., and Yamaguchi-Shinozaki, K. (2014). ABA-dependent and ABA-independent signaling in response to osmotic stress in plants. *Curr. Opin. Plant Biol.* 21, 133–139. doi: 10.1016/j.pbi.2014.07.009

Zahedi, H., and Abbasi, S. (2015). Effect of plant growth promoting rhizobacteria (PGPR) and water stress on phytohormones and polyamines of soybean. *Indian J. Agric. Res.* 49, 427-431. doi: 10.18805/ijare.v49i5.5805

Zarea, M. J. (2019). "Applications of beneficial microbe in arid and semiarid agroecosystem: IAA-producing bacteria," in *Microbiome in plant health and disease* (Springer), 105–118.

Zhao, Y. (2010). Auxin biosynthesis and its role in plant development. Annu. Rev. Plant Biol. 61, 49. doi: 10.1146/annurev-arplant-042809-112308

Zhao, P., Lu, G. H., and Yang, Y. H. (2017). "Salicylic acid signaling and its role in responses to stresses in plants," in *Mechanism of plant hormone signaling under stress*, 413–441.

Zhao, S.-P., Xu, Z.-S., Zheng, W.-J., Zhao, W., Wang, Y.-X., Yu, T.-F., et al. (2017). Genome-wide analysis of the RAV family in soybean and functional identification of GmRAV-03 involvement in salt and drought stresses and exogenous ABA treatment. *Front. Plant Sci.* 8, 905. doi: 10.3389/fpls.2017.00905

Zheng, Y., Wang, X., Cui, X., Wang, K., Wang, Y., and He, Y. (2023). Phytohormones regulate the abiotic stress: An overview of physiological, biochemical, and molecular responses in horticultural crops. *Front. Plant Sci.* 13, 1095363. doi: 10.3389/fpls.2022.1095363

Zhu, J.-Y., Sae-Seaw, J., and Wang, Z.-Y. (2013). Brassinosteroid signalling. Development 140, 1615–1620. doi: 10.1242/dev.060590