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RECEIVED 31 December 2024 ACCEPTED 21 April 2025 PUBLISHED 03 June 2025

#### CITATION

Xu J, Zhou M, Weng Q, Li M and Gan S (2025) Across-rotation genetic analysis and multitrait selection in a cloned cross of *Eucalyptus urophylla* × *E. tereticornis. Front. Plant Sci.* 16:1553819. doi: 10.3389/fpls.2025.1553819

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# Across-rotation genetic analysis and multitrait selection in a cloned cross of *Eucalyptus urophylla* × *E. tereticornis*

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Clonal testing is crucial for estimating genetic parameters and selecting elite clones. However, few studies have tested clones over two or more rotations. In this study, a clonal trial of the sibs from a Eucalyptus urophylla  $\times$  Eucalyptus tereticornis cross was used for genetic parameter analyses and multitrait selection based on traits such as across-rotation growth, wood properties, coppicing ability, and coppice chlorophyll fluorescence. Clonal repeatability  $(H^2)$  estimates for growth traits in the first rotation ranged from 0.73 (diameter at breast height at age 15) to 0.84 (single-tree volume at age 2.5) and were generally higher than those for growth traits in the second rotation (0.11-0.62).  $H^2$  estimates for coppicing and chlorophyll fluorescence traits ranged from 0.35 to 0.57 and 0.03 to 0.53, respectively, indicating low to medium genetic control of these traits. Phenotypic correlations  $(r_p)$  and additive genetic correlations  $(r_q)$ were generally high among growth traits within a single rotation and among coppicing traits but varied considerably across other trait pairs. In particular, second-rotation volume at 2.5 years showed a very weak  $r_p$  but a moderately positive  $r_a$  with early-stage volumes from the first rotation. Multitrait selection index (SI) and multitrait genotype-idiotype distance index (MGIDI) methods were applied at a selection intensity of 15% under four scenarios, each combining two to five traits. Relative genetic gain (RG) and selection efficiency (E) estimates were positive for all traits included in multitrait selection, indicating the usefulness of all selection scenarios. Compared to SI, the MGIDI method produced slightly lower RG and E values for volumes at ages 8 and 15 years, but higher RG and E values for wood basic density and cellulose content at age 8 in the first rotation. This study has important implications for eucalypt clonal breeding and management.

#### KEYWORDS

*Eucalyptus* hybrid, across-rotation traits, genetic parameters, multi-trait genetic selection, genetic gain, selection efficiency

## 1 Introduction

Clonal forestry, defined as the commercial deployment of identified, well-characterized forest tree clones (Burdon and Aimers-Halliday, 2006), has been successfully implemented to improve plantation productivity in many countries. In China, for example, over 60% of *Eucalyptus* L'Hérit. (family Myrtaceae Juss.) plantations are clonal (Wei, 2005), and the large-scale development of clonal forestry has increased productivity from less than 8 to over 15 m<sup>3</sup> ha<sup>-1</sup> year<sup>-1</sup> in around 30 years (Arnold et al., 2020). Also, the genetic uniformity of clonal ramets can lead to phenotypic uniformity, which enables the managers to plan and execute clonal forestry activities much more efficiently than seedling-based plantations (Kleinschmit et al., 1993; Soares et al., 2016). Moreover, the improved productivity and phenotypic uniformity can be extended over a couple of rotations through coppice regeneration.

There can sometimes be concerns about the risks of the deployment of clones in plantation forestry, such as reduced genetic diversity, vegetative propagation failure, cultivar decline, and biotic or climatic damage. However, appropriate choice of management regimes, such as the use of an optimal number of unrelated clones (or so-called clonal composites) and effective control of possible damages, can diminish such risks (Kleinschmit et al., 1993; Stelzer and Goldfarb, 1997; Burdon and Aimers-Halliday, 2006; Rezende et al., 2019). Nowadays, clonal forestry has been implemented for a multitude of broadleaved and coniferous trees, including pure species and interspecific hybrids from the genera *Eucalyptus, Populus L., Pinus L.*, and *Picea* Dietr (Wu, 2019).

Clonal testing is required for the estimation of genetic parameters and the selection of elite clones (White, 1996). Genetic parameters such as heritability (or repeatability) and trait-trait correlations are crucial for determining clonal breeding strategies and predicting genetic gains. Compared to seedling-based tests, clonal tests allow for more efficient dissection of genetic and environmental effects from general phenotypic variation due to the use of ramets of the same genotype (Foster and Shaw, 1988; Isik et al., 2003; Wu, 2019). This can improve the accuracy of breeding value prediction and elite genotype selection, thus contributing to greater genetic gains (Mullin and Park, 1992; Isik et al., 2005). In addition, simultaneous testing of clonal ramets and their ortets provides opportunities for assessing the correlation between clonal and seedling performance, which may have important implications in a breeding program (Shalizi et al., 2020). In forest trees, clonebased genetic parameters and elite selection have been reported in many species, e.g., Eucalyptus grandis Hill ex Maiden (Lambeth et al., 1994; Osorio et al., 2001, 2003; Amâncio et al., 2020), Pinus taeda L (Isik et al., 2003, 2005; Shalizi and Isik, 2019; Braga et al., 2020; Shalizi et al., 2020), and Picea abies (L.) H. Karst (Berlin et al., 2019; Chen et al., 2020; Zeltiņš et al., 2022). However, previous studies presented results only within a single rotation, and little is known about the corresponding information across two or more rotations.

In breeding practices, multiple traits of economic importance should be simultaneously considered for selection. Several methods exist for multitrait selection, including tandem selection, simultaneous culling, conventional index selection (Smith, 1936; Hazel, 1943), and single-environment multitrait genotype-idiotype distance index (MGIDI) selection (Olivoto and Nardino, 2021). Of these, conventional index selection involves the construction of a selection index that is a linear combination of multiple traits and their relative weights. This method has a long history of application to tree breeding (Cotterill and Dean, 1990) and is still the method of choice for selection in many woody species, e.g., Eucalyptus urophylla S. T. Blake × E. grandis (Bouvet et al., 2020) and Picea glauca (Moench) Voss (Rashidi-Jouybari et al., 2022). More recently, the MGIDI method was proposed as a novel approach to selection based on mean performances of multiple traits (Olivoto and Nardino, 2021). It has been used effectively in crops such as Zea mays L. (Singamsetti et al., 2023) forest trees such as P. abies (Alexandru et al., 2023) and Populus simonii Carr. × P. nigra L (Wang et al., 2024). However, its comparison to that of conventional index selection remains to be investigated.

Eucalyptus represents the most widely planted broadleaved tree genus in the world (there being more than 21 million ha of plantations globally; Midgley, 2013). With the development of clonal forestry, the vast majority of eucalypt planting stock is now clonal, particularly utilizing interspecific hybrid varieties. For instance, hybrid clones from E. urophylla, E. grandis, E. tereticornis Smith, and E. camaldulensis Dehnh. predominate in eucalypt plantations in China (Wei, 2005). As noted, E. urophylla × E. tereticornis clones have characteristic fast growth, high yields, and moderate resistance to typhoons and have been cultivated widely in coastal regions in southern China (Peng et al., 2013). Earlier reports have revealed significant variation in the growth and wood properties of E. urophylla × E. tereticornis clones (Gan et al., 2006; Yang et al., 2018). However, similar to those aforementioned species available for clone-based genetic parameter estimation and elite genotype selection, earlier reports are limited to one single rotation.

We hypothesized that genetic variations existed among clonal sibs even within a single cross in *Eucalyptus* and effective selection can be carried out across rotations. In the present study, a clonal trial of the sibs of an *E. urophylla* × *E. tereticornis* cross was used to conduct genetic parameter analysis and elite clone selection on such traits as across-rotation growth, wood properties of the first rotation, coppicing ability, and coppice chlorophyll fluorescence. Our objectives were to determine the clonal repeatability ( $H^2$ ), trait–trait, and age–age correlations; predict the relative genetic gains (*RG*, %) from multitrait selections; and compare the selection efficiency (*E*, %) of index selection versus MGIDI method.

## 2 Materials and methods

### 2.1 Plant material and experimental design

Rooted cuttings of 403 sibs of an interspecific cross between *E. urophylla* (maternal genotype UX-30) and *E. tereticornis* (paternal genotype T43-05) were planted in a common garden experiment in

April 2006. The experimental site was located at Gonghe Town ( $22^{\circ}34'$  24" N, 112°51'14" E), Heshan County, Guangdong Province, China (more details as described in Yang et al., 2018). The experiment was laid out in a design of randomized complete blocks, with single-tree plots, six blocks (replicates), and a spacing of 2 m × 3 m. The last two blocks were damaged by a fire accident that occurred at the age of around five years during the first rotation, so thereafter these blocks were excluded from investigation and analyses. Moreover, 309 ortets of the sibs were planted along with the clonal experiment.

In August 2021, all trees were felled to allow for coppice regeneration of the experiment. Stumps were cut at approximately 0.1 m above ground level. Nine months after the felling, when most stumps had sprouts exceeding 2.5 m, as commonly practiced in eucalypt coppice management in China, only the highest two sprouts were retained per stump.

### 2.2 Trait measurements and calculations

Growth traits of the first rotation (prior to felling) were measured for all trees at 10, 12, and 15 years of age, including tree height ( $HT_{10}$ ,  $HT_{12}$ , and  $HT_{15}$  in m, respectively) and diameter over bark at breast height ( $DBH_{10}$ ,  $DBH_{12}$ , and  $DBH_{15}$  in cm, respectively). Wood basic density at age 15 ( $BD_{15}$ , g m<sup>-3</sup>) was determined based on near-infrared spectroscopy predictions using a MPA spectrometer (Bruker Optik, Ettlingen, Germany), which was similar to the method of Yang et al. (2018).

Earlier growth traits and 8-year-old wood properties were investigated as per Yang et al. (2018), including tree height at ages 0.5, 1.5, 2.5, 4.5, 5.5, 6.5, and 8 years ( $HT_{0.5}$ ,  $HT_{1.5}$ ,  $HT_{2.5}$ ,  $HT_{4.5}$ ,  $HT_{5.5}$ ,  $HT_{6.5}$ , and  $HT_8$ , respectively) and diameter at breast height at 1.5, 2.5, 4.5, 5.5, 6.5, and 8 years ( $DBH_{1.5}$ ,  $DBH_{2.5}$ ,  $DBH_{4.5}$ ,  $DBH_{5.5}$ ,  $DBH_{6.5}$ , and  $DBH_8$ , respectively), as well as 8-year-old wood basic density ( $BD_8$ , g m<sup>-3</sup>), cellulose content ( $CC_8$ , %), hemicellulose content ( $HC_8$ , %), Klason lignin content ( $LC_8$ , %), and lignin guaiacyl-to-syringyl ratio ( $G/S_8$ ). Single-tree volumes at 1.5, 2.5, 4.5, 5.5, 6.5, 8, 10, 12, and 15 years ( $V_{1.5}$ ,  $V_{2.5}$ ,  $V_{4.5}$ ,  $V_{5.5}$ ,  $V_{6.5}$ ,  $V_8$ ,  $V_{10}$ ,  $V_{12}$ , and  $V_{15}$  in m<sup>3</sup>, respectively) were calculated as  $HT_n \times DBH_n^{2/}$ 30,000 (He et al., 2012), where  $HT_n$  (m) and  $DBH_n$  (cm) are height and diameter at breast height of a given year *n*, respectively.

Coppicing traits were observed for each stump six months after felling, including the number of sprouts ( $NS_{0.5}$ ), height ( $HTs1_{0.5}$  and  $HTs2_{0.5}$ , m), and ground diameter ( $GDs1_{0.5}$  and  $GDs2_{0.5}$ , cm) of the two highest sprouts, as well as coppicing potential ( $CP_{0.5}$ ). Specifically, coppicing potential was evaluated by combining the number and the growth condition of sprouts, and  $CP_{0.5}$  was scored using a six-grade scale based on the classification of  $NS_{0.5}$  values: grades 1, 2, 3, 4, 5, and 6 for 1–4, 5–8, 9–12, 13–16, 17–20, and  $\geq$  21 sprouts, respectively, with modification by increase or decrease of one grade in respect of the sprout growth condition.

Chlorophyll fluorescence and content measurements were performed in triplicate for three fully expanded young leaves per stump in August 2022, when the earliest sprouts were about one year old. A portable chlorophyll fluorometer PAM-2500 (Heinz Walz GmbH, Effeltrich, Germany) was used to measure chlorophyll fluorescence traits. Each leaf was dark-adapted for 30 min before determining the initial ( $F_0$ ), maximal ( $F_m$ ), and variable ( $F_v = F_m - F_0$ ) fluorescence, as well as the maximal quantum yield of photosystem II [ $Y(II) = F_v/F_m$ ]. After light-adapted conditions ( $\geq 800 \mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>) for at least 25 min, steady-light chlorophyll fluorescence ( $F_s$ ), maximal fluorescence ( $F_m'$ ), variable fluorescence ( $F_v' = F_m' - F_s$ ), maximal quantum yield of photosystem II [ $Y(II)' = F_v'/F_m'$ ], and photosynthetic electron transport rate (*ETR*) were recorded also using PAM-2500. Nonphotochemical quenching (*NPQ*) was computed as  $F_m/F_m' - 1$ . In addition, a hand-held chlorophyll meter SPAD-502 (Spectrum Technologies Inc., Plainfield, IL, USA) was used to detect leaf chlorophyll content as SPAD reading (*SPADR*).

Growth traits of the two sprouts retained per stump were measured when the coppice age was around 1.5 and 2.5 years. For the age of 1.5 years, the higher and lower sprouts per stump were measured for height (HTs11,5 and HTs21,5 in m, respectively) and diameter at breast height (DBHs11.5 and DBHs21.5 in cm, respectively). The individual volume of each of the two sprouts (Vs11.5 and Vs21.5, m<sup>3</sup>) was calculated similarly as mentioned above, and Vs11,5 and Vs21,5 were added up to represent the single-ramet volume ( $Vs_{1.5}$ , m<sup>3</sup>). Moreover, branch angle (BAs1.5, °) relative to the stem was determined by averaging those of the four branches, each being the biggest in the up, down, left, or right direction of a row. The height of the lowest live branch (HTLBs<sub>1.5</sub>, m) was observed across the two sprouts of a ramet, and number of branches (NBs1.5) was counted only for the higher sprout. Crown width was measured along row and column directions (CWs115 and CWs215, m), which were subsequently used for calculating crown projected area (CPAs<sub>1.5</sub> = CWs1<sub>1.5</sub> × CWs2<sub>1.5</sub> ×  $\pi$ / 4,  $m^2$ ) assuming an elliptical shape. Crown length (*CLs*<sub>1.5</sub>, m) or depth was estimated as the difference between the height of the higher sprout (HTs11.5 or HTs21.5) and HTLBs1.5. For the age of 2.5 years, sprout height (HTs12.5 and HTs22.5, m), diameter at breast height (DBHs12.5 and DBHs22,5, cm), and volume (Vs12,5 and Vs22,5, m<sup>3</sup>) as well as single-ramet volume (Vs2.5, m3) were measured or calculated similarly as conducted above for 1.5 years of age. In a few cases, only one sprout remained per stump and was consequently measured.

### 2.3 Statistical analyses

Statistical analyses were performed using RStudio (RStudio Team, 2021) in R version 4.1.3 (R Core Team, 2022). Trait means, along with their standard deviations (SD) and coefficients of variation (CV), were calculated. In addition, *t*-tests were conducted to evaluate the significance of differences between ramet means and ortet values.

# 2.3.1 Variance components and clonal repeatability

For each of the traits, variance components were estimated using the *lmer()* function of the lme4 statistical package (Bates et al., 2015) following the linear mixed model:

$$y_{ij} = \mu + G_i + B_j + E_{ij}$$

where  $Y_{ij}$  is the trait value of the *i*th genotype (clonal sib) in the *j*th block,  $\mu$  represents the overall mean,  $G_i$  is the additive genetic effect (random) of the *i*th genotype,  $B_j$  is the effect (fixed) of the *j*th block, and  $E_{ij}$  is the residual error. The genotypic variance component ( $\sigma_g^2$ ) and residual error variance component ( $\sigma_e^2$ ) were used to calculate the coefficient of genetic variation (CVg) as ( $\delta_g/\mu$ ) × 100 % and  $H^2$  as  $\sigma_g^2/(\sigma_g^2 + \sigma_e^2/r)$ , where  $\mu$  and *r* represents the overall mean and the number of blocks, respectively.

### 2.3.2 Phenotypic and additive genetic correlations

Phenotypic correlations  $(r_p)$  and additive genetic correlations  $(r_q)$  between trait pairs were estimated as:

$$r_{p} = \sigma_{p_{x}p_{y}} / \sqrt{\sigma_{p_{x}}^{2} \sigma_{p_{y}}^{2}}$$
$$r_{g} = \sigma_{g_{x}g_{y}} / \sqrt{\sigma_{g_{x}}^{2} \sigma_{g_{y}}^{2}}$$

where  $\sigma_{p_x p_y}$  and  $\sigma_{g_x g_y}$  are the estimated phenotypic and genotypic covariances between traits *x* and *y*,  $\sigma_{p_x}^2$  and  $\sigma_{p_y}^2$  represent phenotypic variance components of traits *x* and *y*, and  $\sigma_{g_x}^2$  and  $\sigma_{g_y}^2$  represent genotypic variance components of traits *x* and *y*, respectively. Genotypic and phenotypic covariances were computed using *asreml()* function, and  $r_p$ ,  $r_g$ , and their standard error (SE) were then calculated using *vpredict()* function in ASReml (Butler et al., 2017).

# 2.3.3 Multitrait index selection and multitrait genotype-idiotype distance index selection

Multitrait selections were conducted for ages 8 and 15 years of the first rotation, combining the two economic traits  $CP_{0.5}$  and  $Vs_{2.5}$ of the second rotation. In China, age 8 is the approximate rotation length of eucalypt plantations established for veneer and pulp industries, while age 15 is one of the rotation periods recommended for large-size eucalypt timber production (Chen et al., 2017). In respect of the traits of economic importance,  $V_8$ ,  $BD_8$ , and  $CC_8$  for age 8 and  $V_{15}$  and  $BD_{15}$  for age 15 were employed for the across-rotation multitrait selection. Also, these traits were subject to additional single-rotation multitrait selection. Moreover, single-trait selection for  $V_{1.5}$ ,  $V_{2.5}$ ,  $V_{4.5}$ ,  $V_{5.5}$ ,  $V_{6.5}$ ,  $V_8$ ,  $BD_8$ ,  $CC_8$ ,  $V_{10}$ ,  $V_{12}$ ,  $V_{15}$ ,  $DB_{15}$ ,  $CP_{0.5}$ ,  $V_{8.15}$ , and  $Vs_{2.5}$  was performed based on the best linear unbiased prediction (BLUP) of clonal breeding values (*BV*). For all the selections, a selection intensity of 15% was adopted.

Multitrait selection index (SI) was calculated following the method of Smith (1936) and Hazel (1943):

$$SI = \sum_{i=1}^{n} b_i x_i$$

where  $x_i$  is clonal phenotypic mean values of trait *i* and  $b_i$  is index coefficient for trait *i* derived from:

$$b=P^{-1}Gw$$

where P and G are the phenotypic and genetic variance–covariance matrices, respectively, and w is the vector of relative economic weights each defined for an objective trait using an equal-emphasis approach (Cotterill and Dean, 1990):

$$w_i = 1/\sigma_{pi}$$

where  $\sigma_{pi}$  is the phenotypic standard deviation of trait *i*.

For MGIDI selection, the MGIDI value of each clone was estimated for the objective traits based on BLUP (Olivoto and Nardino, 2021). Four main steps were carried out (Olivoto and Nardino, 2021), namely, rescaling the objective traits each into a 0–100 range, estimating the factorial score of each clone to group correlated traits into factors, planning the ideotype (clone) that has the highest rescaled value for each trait, and calculating the MGIDI of clone i (MGIDI<sub>i</sub>) as:

$$MGIDI_i = \left[\sum_{j=1}^{f} (\gamma_{ij} - \gamma_j)^2\right]^{0.5}$$

where  $\gamma_{ij}$  is the score of clone *i* in factor *j*, and  $\gamma_j$  is the score of factor *j* of the ideotype. The lower the MGIDI of a clone, the closer the clone is to the ideotype. The function *mgidi()* of metan 1.18.0 (Olivoto and Lúcio, 2020) was used for MGIDI calculation.

# 2.3.4 Relative genetic gain and selection efficiency

*RG* for a specific trait was calculated according to White et al. (2007):

$$RG(\%) = \frac{H^2(M_s - M_o)}{M_o} \times 100$$

where  $H^2$  is clonal repeatability,  $M_s$  is the phenotypic mean of selected clones, and  $M_o$  is the overall phenotypic mean of all the clones.

*E* of an earlier selection (usually on a single trait) relative to a later selection (usually on multiple traits) was expressed as:

$$E(\%) = \frac{H^2(M_{s1} - M_o)}{M_o} \div \frac{H^2(M_{s2} - M_o)}{M_o} \times 100 = \frac{M_{s1} - M_o}{M_{s2} - M_o} \times 100$$

where  $H^2$  is clonal repeatability of a trait in later selection,  $M_{s1}$  and  $M_{s2}$  are the later phenotypic means of earlier and later selected clones, respectively, and  $M_o$  is the later overall phenotypic mean of all the clones. Similarly, efficiency between selections at the same age was calculated.

## **3** Results and discussion

## 3.1 Trait variation

There were 318 (78.9%), 292 (72.5%), and 275 (67.0%) clones that survived at age 8, age 15 of the first rotation, and age 2.5 of the second rotation, respectively. Of the 54 traits investigated,  $BD_{15}$  and all the 15 growth traits of the first rotation, all the six coppicing traits, and nine of the 18 second-rotation growth traits (exclusive of  $HTs2_{1.5}$ ,  $DBHs2_{1.5}$ ,  $Vs2_{1.5}$ ,  $BAs_{1.5}$ ,  $NBs_{1.5}$ ,  $HTs1_{2.5}$ ,  $HTs2_{2.5}$ , and  $DBHs1_{2.5}$ ) showed significant differences ( $p \le 0.001$ , 0.01, or 0.05) among clones. These results indicate high progeny phenotypic variability for these traits, while most sprout leaf chlorophyllrelated traits (7/11 with  $F_0$ ,  $F_s$ , NPQ, and SPADR excluded) proved to be not significant (Supplementary Table 1). This corroborates our earlier finding of significant clonal difference within the same cross for earlier growth (tree height and diameter at breast height across 0.5–8 years) and 8-year-old wood chemical properties (Yang et al., 2018). Similarly, significant differences among clonal sibs within a single cross were noticed for growth and leaf traits in *Populus deltoides* Bartr. ex Marsh. × *P. nigra* (Marron and Ceulemans, 2006) and (*Populus pseudo-simonii* Kitag. × *P. nigra*) × *Populus beijingensis* W. Y. Hsu (Liao et al., 2016) for branching in *P. taeda* (Xiong et al., 2014), but not for leaf chlorophyll traits such as  $F_v/F_m$  [Y(II)] in (*P. pseudo-simonii* × *P. nigra*) × *P. beijingensis* (Liao et al., 2016).

Meanwhile, growth traits of both rotations presented generally larger coefficients of variation, ranging from 6.9% in  $BAs_{1.5}$  to 57.2% in  $Vs1_{1.5}$ , as compared to those of leaf chlorophyll traits and  $BD_{15}$ (between 2.6% in Y(II) and 27.7% in  $F_v'$  except the extremely high case of 121.1% in *NPQ*; Supplementary Table 1). For most traits, high phenotypic variability together with considerable coefficients of variation may suggest good potential for clone selection for future deployment programs.

Significant differences ( $p \le 0.001, 0.01, \text{ or } 0.05$ ) between ramet means and ortet values were found for most traits, including nine, six, three, and 10 of the 15 first-rotation growth, six coppicing, 11 chlorophyll, and 21 second-rotation growth traits, respectively (Supplementary Table 2). For the 28 traits with significant differences, ortets outperformed their ramet means in 17 traits, including eight, six, and three of the first-rotation growth, coppicing, and second-rotation growth traits, respectively (Supplementary Table 2). Several previous studies reported better growth of seedlings than rooted cutting progeny originating from the same full-sib families in Eucalyptus (Sasse and Sands, 1997; Costa e Silva et al., 2013; van den Berg et al., 2015) and Pinus (Antony et al., 2014; Quesada et al., 2017; Shalizi et al., 2020). These studies attributed the growth difference to the poor root structure of rooted cuttings, indicating the presence of propagation effects specific to some clones (C-effects; Borralho and Kanowski, 1995). Even so, significantly higher or nonsignificantly different performance of ramets as compared to seedlings was shown in the current study in a couple of the growth traits (e.g., those of ages 10 and 12 of the first rotation, and age 2.5 of the second rotation, except only HTs12.5; Supplementary Table 2). In forest trees, occasional cases of nonsignificantly different or better clonal performance were also reported for certain growth traits in E. grandis × E. urophylla hybrid (Sasse and Sands, 1997) and P. taeda full-sib progeny (Quesada et al., 2017).

# 3.2 Variance components and repeatability estimates

Estimates of  $\sigma_g^2$  were significant ( $p \le 0.001, 0.01, \text{ or } 0.05$ ) for all the traits of the first rotation, coppicing traits, 11 of the 21 second-rotation growth traits, and three of the 11 chlorophyll traits

(Table 1), which was almost coincident with the ANOVA results (Supplementary Table 1).  $\sigma_g^2$  varied largely with the trait, e.g.,  $CV_g$  being 2.4%, 9.4%–44.5%, 10.5%–20.0%, 0.9%–16.5%, and 1.3%–23.2% for  $BD_{15}$ , the first-rotation growth, coppicing, chlorophyll, and the second-rotation growth traits, respectively (Table 1).

 $H^2$  estimates for first-rotation growth traits ranged from 0.73 in  $DBH_{15}$  to 0.84 in  $V_{2.5}$ , which tended to remain relatively stable over time and were generally higher than those of wood density (0.65 in  $BD_{15}$ ) and second-rotation growth traits (0.11–0.62; Table 1). The  $H^2$  levels for the first rotation were in agreement with the previous report on earlier tree height (0.72–0.88 in  $H^2$ , except only  $HT_8$  at 0.40), diameter at breast height (0.76–0.84), and wood density (0.62 in  $BD_8$ ) of the same trial (Yang et al., 2018). According to Singh (2001), broad-sense heritability is considered low, medium, moderately high, and very high at values  $\leq 0.40$ , 0.40–0.59, 0.60–0.79, and  $\geq 0.80$ , respectively. Thus, the growth and wood density of the first rotation can be considered to be under moderately high or very high genetic control, while second-rotation growth traits tended to be under low to medium genetic control (except for  $HTLBs_{1.5}$ , which had a moderately high  $H^2$  of 0.62).

Despite the current study involving only a single cloned cross, the  $H^2$  estimates obtained were comparable to those reported previously for multiple cloned families or crosses of *Eucalyptus*. For example, Reis et al. (2011) reported a clonal repeatability of over 0.74 for a 2-year-old volume of the first rotation in three trials of cloned *E. grandis* pure-species families and of *E. grandis* × *E. urophylla* interspecific crosses. Somewhat later, Amâncio et al. (2020) reported clonal repeatabilities of between 0.19 and 0.28 for 5.5-year-old coppice growth of the second rotation in five trials of cloned *E. grandis, E. saligna* Smith, and *E. urophylla* families and of *E. urophylla* × *E. grandis* crosses. Also, a similar magnitude of  $H^2$ was found in a cloned single family of *P. taeda*, e.g., 0.79 and 0.75 for tree height and volume at the age of 6–7 years, respectively (Xiong et al., 2014).

Coppicing traits had  $H^2$  between 0.35 in  $GDs1_{0.5}$  and 0.57 in  $HTs2_{0.5}$  (Table 1), revealing low to medium genetic control of these traits. In plantation management, coppice systems involving regeneration from stump sprouts after tree felling can be more economical (Crous and Burger, 2015; Hardiyanto et al., 2022) and sustainable (Zhou et al., 2017) than planting new stands. Coppicing traits are therefore important for guaranteeing plantation productivity and management profit. However, their genetic basis has been explored by only a few studies. Amâncio et al. (2020) found low broad-sense heritabilities (0.205-0.334) in post-cut survival of clonal trials in E. grandis, E. saligna, E. urophylla, and *E. urophylla*  $\times$  *E. grandis.* Hernández et al. (2022) detected one and four quantitative trait loci related to numbers of post-fire basal resprouts and epicormic clusters, respectively, in E. globulus Labill. In combination with these studies, it can be highlighted that coppicing traits are characteristically under genetic control, at least in Eucalyptus, and the genetic effect may be low or medium depending on the trait.

Chlorophyll fluorescence and content traits fell in the range of 0.03-0.53 in  $H^2$  (Table 1), indicating low to medium genetic control

TABLE 1 Clonal additive genetic variance components ( $\sigma_g^2$ ), their standard errors (SE), coefficients of variation (CV<sub>g</sub>, %), and repeatabilities ( $H^2 \pm$  SE) for 54 traits investigated in a cloned *E. urophylla* × *E. tereticornis* cross.

Trait	$\sigma_{\!g}^{\;\;2}$ (± SE)	CV <sub>g</sub> (%)	H <sup>2</sup> ( <u>+</u> SE)	
Growth of the first rotation				
V <sub>1.5</sub> (m <sup>3</sup> )	$3.86 (\pm 0.39) \times 10^{-5***}$	31.5	0.82 (± 0.03)	
$V_{2.5} (m^3)$	$1.38 (\pm 0.13) \times 10^{-4_{***}}$	34.7	0.84 (± 0.04)	
V <sub>4.5</sub> (m <sup>3</sup> )	$1.38 (\pm 0.15) \times 10^{-3***}$	42.7	0.82 (± 0.03)	
$V_{5.5}$ (m <sup>3</sup> )	$2.65 (\pm 0.29) \times 10^{-3***}$	44.5	0.81 (± 0.03)	
$V_{6.5} (m^3)$	$3.41 (\pm 0.47) \times 10^{-3***}$	37.4	0.77 (± 0.03)	
$V_8 (m^3)$	$0.01 \ (\pm \ 0.68 \times 10^{-3})^{***}$	39.2	0.79 (± 0.03)	
<i>HT</i> <sub>10</sub> (m)	4.54 (± 0.68)***	9.4	0.77 (± 0.03)	
$DBH_{10}$ (cm)	10.18 (± 1.37)***	18.2	0.78 (± 0.03)	
V <sub>10</sub> (m <sup>3</sup> )	$0.01 \ (\pm \ 0.16 \times 10^{-2})^{***}$	36.5	0.75 (± 0.03)	
<i>HT</i> <sub>12</sub> (m)	4.96 (± 0.75)***	9.7	0.78 (± 0.04)	
$DBH_{12}$ (cm)	11.39 (± 1.56)***	18.6	0.78 (± 0.03)	
V <sub>12</sub> (m <sup>3</sup> )	$0.01 \ (\pm \ 0.20 \times 10^{-2})^{***}$	37.0	0.74 (± 0.03)	
<i>HT</i> <sub>15</sub> (m)	7.42 (± 1.21)***	11.1	0.81 (± 0.04)	
$DBH_{15}$ (cm)	9.67 (± 1.54)***	16.2	0.73 (± 0.03)	
V <sub>15</sub> (m <sup>3</sup> )	$0.02 \ (\pm \ 0.33 \times 10^{-2})^{***}$	36.1	0.74 (± 0.04)	
Wood density	of the first rotation			
$BD_{15} (g/cm^3)$	$1.90 (\pm 0.61) \times 10^{-4***}$	2.4	0.65 (± 0.03)	
Coppicing				
NS <sub>0.5</sub>	7.20 (± 1.84)***	20.0	0.54 (± 0.02)	
$HTs1_{0.5}$ (m)	0.06 (± 0.02)***	11.0	0.47 (± 0.01)	
<i>HTs2</i> <sub>0.5</sub> (m)	0.08 (± 0.02)***	13.3	0.57 (± 0.02)	
GDs1 <sub>0.5</sub> (cm)	4.48 (± 2.07) × $10^{-4_{\star}}$	10.5	0.35 (± 0.01)	
GDs2 <sub>0.5</sub> (cm)	$5.55 (\pm 1.72) \times 10^{-4***}$	13.7	0.48 (± 0.01)	
CP <sub>0.5</sub>	0.34 (± 0.09)***	13.1	0.54 (± 0.02)	
Chlorophyll fluorescence and concentration				
$F_0$	$8.19 (\pm 4.88) \times 10^{-5}$	4.1	0.30 (± 0.01)	
Fm	$1.14 (\pm 6.03) \times 10^{-4}$	1.2	$0.04~(\pm~0.96\times10^{-4})$	
$F_{\rm v}$	2.21 (± 3.66) × $10^{-4}$	2.2	0.12 (± 0.87 × $10^{-3}$ )	
Y(II)	4.49 (± 2.65) × $10^{-5}$	0.9	0.31 (± 0.01)	
Fs	$5.94 (\pm 2.70) \times 10^{-4*}$	6.9	0.38 (± 0.01)	
$F_{\rm m}'$	$1.59 (\pm 0.66) \times 10^{-3**}$	7.9	0.40 (± 0.01)	
$F_{\rm v}{}'$	7.37 (± 13.40) × $10^{-5}$	5.8	$0.11 \ (\pm \ 0.72 \times 10^{-3})$	
Y(II)'	$1.50 (\pm 1.16) \times 10^{-4}$	4.4	$0.25~(\pm~0.37\times10^{-2})$	
ETR	8.06 (± 51.30)	2.0	$0.03~(\pm~0.54 imes10^{-4})$	
NPQ	0.02 (± 0.01)*	16.3	0.37 (± 0.01)	

(Continued)

#### TABLE 1 Continued

Trait	$\sigma_{\!g}^{2}$ (± SE)	CV <sub>g</sub> (%)	H <sup>2</sup> ( <u>+</u> SE)	
Chlorophyll fluorescence and concentration				
SPADR	1.68 (± 0.53)***	3.1	0.53 (± 0.02)	
Growth of the	e second rotation			
<i>HTs1</i> <sub>1.5</sub> (m)	0.30 (± 0.18)	7.1	0.33 (± 0.01)	
<i>HTs2</i> <sub>1.5</sub> (m)	0.33 (± 0.19)	7.5	0.37 (± 0.01)	
DBHs1 <sub>1.5</sub> (cm)	0.43 (± 0.18)	12.0	0.41 (± 0.01)	
<i>DBHs2</i> <sub>1.5</sub> (cm)	0.32 (± 0.19)	10.4	0.35 (± 0.01)	
<i>Vs1</i> <sub>1.5</sub> (m <sup>3</sup> )	$4.64 (\pm 2.39) \times 10^{-6_{\star}}$	22.3	0.37 (± 0.01)	
$Vs2_{1.5} (m^3)$	$2.95 (\pm 2.13) \times 10^{-6}$	17.6	0.30 (± 0.01)	
Vs <sub>1.5</sub> (m <sup>3</sup> )	$1.61 (\pm 0.66) \times 10^{-5_{**}}$	22.2	0.44 (± 0.01)	
BAs <sub>1.5</sub> (°)	0.67 (± 1.29)	1.3	$0.11 (\pm 0.73 \times 10^{-3})$	
$HTLBs_{1.5}$ (m)	0.25 (± 0.06)***	16.3	0.62 (± 0.02)	
NBs <sub>1.5</sub>	4.46 (± 3.57)	7.5	$0.25~(\pm~0.38\times10^{-2})$	
<i>CWs1</i> <sub>1.5</sub> (m)	0.06 (± 0.03)*	8.5	0.37 (± 0.01)	
<i>CWs2</i> <sub>1.5</sub> (m)	0.09 (± 0.03)**	9.8	0.46 (± 0.01)	
$CPAs_{1.5} (m^2)$	1.51 (± 0.56)**	16.6	0.46 (± 0.01)	
<i>CLs</i> <sub>1.5</sub> (m)	0.25 (± 0.10)**	10.1	0.45 (± 0.01)	
<i>HTs1</i> <sub>2.5</sub> (m)	0.38 (± 0.41)	5.7	$0.19 (\pm 2.23 \times 10^{-3})$	
<i>HTs2</i> <sub>2.5</sub> (m)	0.39 (± 0.45)	5.7	$0.21 (\pm 2.81 \times 10^{-3})$	
<i>DBHs1</i> <sub>2.5</sub> (cm)	0.41 (± 0.28)	8.6	$0.27 (\pm 4.48 \times 10^{-3})$	
<i>DBHs2</i> <sub>2.5</sub> (cm)	0.74 (± 0.33)*	11.7	0.44 (± 0.01)	
Vs1 <sub>2.5</sub> (m <sup>3</sup> )	2.79 (± 1.48) × 10 <sup>-5</sup> *	21.8	$0.35 (\pm 7.64 \times 10^{-3})$	
<i>Vs2</i> <sub>2.5</sub> (m <sup>3</sup> )	$3.13 (\pm 1.54) \times 10^{-5_{\star}}$	23.1	0.42 (± 0.01)	
Vs <sub>2.5</sub> (m <sup>3</sup> )	9.73 (± 3.72) × $10^{-5*}$	23.2	0.44 (± 0.01)	

Trait abbreviations are provided in the section of Glossary. Significance level: \*\*\* $p \le 0.001$ ; \*\* $p \le 0.01$ ; \* $p \le 0.05$ ; nonsignificant for all other *F*-values.

of the traits. Similar chlorophyll traits were investigated in earlier studies on forest trees under stress, e.g., high temperature in Populus euphratica Oliv. (Zhou et al., 2010) and elevated CO<sub>2</sub> in E. tereticornis (Wujeska-Klausea et al., 2019), heritability including  $H^2$  has rarely been reported. In other plants, broad-sense heritabilities for chlorophyll fluorescence traits have varied with trait and drought conditions from 0.40 to 0.74 in Triticum turgidum L. ssp. durum Desf. (Santos et al., 2021) and from 0.05 to 0.54 in Sorghum bicolor L (Ortiz and Salas-Fernandez, 2022). Also,  $H^2$ fluctuations with light level have been reported for the quantum yield of photosystem II in Arabidopsis thaliana (L.) Heynh (Flood et al., 2016). Therefore, the magnitude of  $H^2$  estimates for chlorophyll-related traits may depend on species, population, and/or environmental conditions. In this respect, the present work provides new information about the genetic control of photosynthetic traits for woody plants.

# 3.3 Phenotypic and additive genetic correlations

Trait-trait correlations  $r_p$  and  $r_g$  are presented in Supplementary Table 3. The first-rotation growth traits showed consistently favorable and positive  $r_p$  and  $r_g$ , ranging from 0.31 to 0.99 and from 0.66 to 1.00, respectively, whereas wood properties had  $r_p$  and  $r_q$  between -0.46 and 0.43 and between -0.80 and 0.82, respectively. In terms of age-age correlations, the earlier growth traits appeared to correlate increasingly with time with that of the final age. For instance,  $r_p$  and  $r_q$  of  $HT_{15}$ ,  $DBH_{15}$ , and  $V_{15}$  increased from 0.48 and 0.72 (with  $HT_{0.5}$ ) to 0.95 and 1.00 (with  $HT_{12}$ ), 0.70 and 0.91 (with DBH<sub>1.5</sub>) to 0.99 and 1.00 (with DBH<sub>12</sub>), and 0.63 and 0.88 (with  $V_{1.5}$ ) to 0.98 and 0.99 (with  $V_{12}$ ), respectively, and remained relatively high with the respective counterpart traits after age 2.5 (more than 0.75 and 0.93 in  $r_p$  and  $r_q$ , respectively; Figure 1). Meanwhile, moderate age-age correlations were observed in wood basic density, with  $r_p$  and  $r_g$  of 0.23 and 0.68 between  $BD_{15}$ and BD<sub>8</sub>, respectively. Moreover, for correlations between growth and wood properties, growth traits showed generally positive  $r_p$  and  $r_g$  with  $BD_8$ ,  $CC_8$ , and  $BD_{15}$ , negative  $r_p$  and  $r_g$  with  $HC_8$  and  $S/G_8$ , but negative  $r_p$  and positive  $r_q$  with  $LC_8$ . Similar correlation patterns were reported earlier for the same cross over three environments at age 8 (Yang et al., 2018).

The high correlations between growth traits obtained in the current study generally concur with some earlier studies on Eucalyptus clones, e.g., genotypic correlations being 0.85-0.90 and 0.94-0.96 between height and diameter in clones of E. camaldulensis (ages 3 and 5; Kien et al., 2010) and E. globulus (ages 4 and 12; Costa e Silva et al., 2013). Though correlations between wood traits for Eucalyptus clones were available in a couple of reports (Kien et al., 2010; Gallo et al., 2018; Bouvet et al., 2020), only Gallo et al. (2018) analyzed merely a pair of wood traits comparable to this study, who also found a negative genotypic correlation (-0.54) between wood basic density and Klason lignin content in E. dunnii Maiden clones. Moreover, similar positive and negative genetic correlations were frequently observed between wood property traits in seedling-based family tests in Eucalyptus, e.g., 0.36 between seedling wood basic density and cellulose content in E. nitens Deane & Maiden (age 9; Hamilton et al., 2009) and -0.31 between S/G and lignin content in E. globulus (age 16; Stackpole et al., 2010). This may indicate a small effect of propagule type (seedling versus vegetatively propagated clone) on trait-trait genetic correlations (Costa e Silva et al., 2013). Furthermore, weak genetic correlations of clonal growth with lignin content detected here (0.03-0.06 at age 8) are in accordance with those observed in clones of E. dunnii (- 0.19 to - 0.13 at age 3.5; Gallo et al., 2018) and E. urophylla × E. grandis (0.238 at 55 months of age; Bouvet et al., 2020). However, the relatively high positive correlations between growth and basic density (0.67-0.82 and 0.44-0.67 at ages 8 and 15, respectively) are opposite to those reported for E. grandis (- 0.06 to 0.06 and -0.08 to 0.16 in  $r_{\sigma}$  of basic density with height and mean annual increment, respectively, across three sites at age 6; Osorio et al., 2003), E. camaldulensis (0.17 and - 0.16 between height and basic density and 0.21 and 0.07 between *DBH* and basic density for two sites at age 5; Kien et al., 2010), and *E. dunnii* (-0.04, -0.01 and 0.02 in  $r_g$  of basic density with *DBH*, height, and mean annual increment, respectively, at age 3.5; Gallo et al., 2018). This may suggest the variation of such correlations depends on both population and environment.

For coppicing traits, trait-trait correlations were positive and generally high, with  $r_p$  of 0.17–0.91 and  $r_g$  of 0.65–1.00. Moreover, coppicing traits  $NS_{0.5}$  and  $CP_{0.5}$  had moderate positive  $r_p$  and  $r_g$  with the first-rotation final growth, namely, 0.28–0.37 for  $r_p$  and 0.47–0.67 for  $r_g$  with  $HT_{15}$ ,  $DBH_{15}$ , and  $V_{15}$ . Meanwhile,  $HTs1_{0.5}$ ,  $HTs2_{0.5}$ ,  $GDs1_{0.5}$ , and  $GDs2_{0.5}$  had very weak  $r_p$  (– 0.01 to 0.05) but moderately low positive  $r_g$  (0.18–0.43) with those growth traits. Coppicing traits were weakly correlated with the wood trait  $BD_{15}$ , with  $r_p$  and  $r_g$  being – 0.03 to 0.13 and – 0.33 to 0.15, respectively. This study represents by far the only attempt in *Eucalyptus* to survey the correlations between coppicing ability and earlier-rotation traits, which could be helpful for understanding the relationship between these traits.

Chlorophyll fluorescence and content traits were correlated variably with each other, with  $r_p$  and  $r_q$  being – 0.83 to 0.98 and - 0.96 to 0.99, respectively. Their  $r_p$  estimates with the traits observed earlier were consistently very weak while those of  $r_{g}$ varied notably with trait, e.g., - 0.17 to 0.28, - 0.12 to 0.10, and -0.14 to 0.12 for  $r_p$  and - 0.98 to 1.00, - 0.41 to 0.99, and - 0.90 to 1.00 for  $r_g$  with first-rotation growth, wood property, and coppicing traits, respectively. The majority of  $r_g$  estimates were accompanied by large standard errors, suggesting non-significance of these correlations. Specifically, the negative  $r_p$  between  $F_0$  and Y(II)(-0.48) and between Y(II)' and NPQ (-0.30) is in line with the observation that decline of  $F_v/F_m$  [Y(II)] and  $F_v'/F_m'$  [Y(II)'] was associated with increased  $F_0$  and NPQ in two eucalypt clones under Cd stress (Pietrini et al., 2015). On the other hand, the very weak  $r_p$ observed between Y(II) and Y(II)' (-0.03) was in sharp contrast to that of a highly significant correlation (0.42, p < 0.001) in S. bicolor under normal conditions (Ortiz and Salas-Fernandez, 2022). Nevertheless, correlations between chlorophyll fluorescence, chlorophyll content, and other trait types have rarely been reported from forest trees, and our results will provide valuable information in this context.

Except for  $BAs_{1.5}$  and  $HTLBs_{1.5}$ , 1.5- and 2.5-year-old growth traits of the second rotation showed generally high positive  $r_p$  and  $r_g$ with each other, demonstrating the strong correlations between growth traits such as sprout height, diameter at breast height, volume, and crown projected area. For instance, irrespective of  $BAs_{1.5}$  and  $HTLBs_{1.5}$ ,  $Vs_{1.5}$  had  $r_p$  ranging from 0.25 (with  $NBs_{1.5}$ ) to 0.91 (with  $Vs_{2.5}$ ) and  $r_g$  ranging from 0.47 (with  $NBs_{1.5}$ ) to 1.00 (with  $VsI_{1.5}$  and  $CPAs_{1.5}$ ). In contrast,  $BAs_{1.5}$  had consistently weak  $r_p$  (0.02–0.14) and  $r_g$  (– 0.66 to 0.70 with high stand errors) with the other traits, while  $HTLBs_{1.5}$  had low to intermediate  $r_p$  (– 0.11 to 0.54) and  $r_g$  (– 0.51 to 0.38 with relatively high stand errors). Similar weak  $r_g$  of branch angle with tree height (– 0.24) and volume (– 0.02) were noted in *P. taeda* clones (Xiong et al., 2014). In particular, the final volume  $Vs_{2.5}$  had very weak  $r_p$  but moderate positive  $r_g$  with earlier volume of the first rotation such as  $V_8$  and



Phenotypic correlations  $(r_p)$  and additive genetic correlations  $(r_g)$  for growth at ages 8 and 15 years in the first rotation and age 2.5 years in the second rotation with earlier respective growth traits. (A) Tree height (*HT*). (B) Diameter at breast height (*DBH*). (C) Volume (*V*). In the legend, numerals following the trait abbreviation indicate the tree or sprout age. For growth at age 2.5 years in the second rotation, *HT* and *DBH* of the higher sprout and total *V* of two sprouts (only one in a few cases) per stump were used for calculating correlations. Trait abbreviations are provided in the section of Glossary.

 $V_{15}$  (0.03 and – 0.06 for  $r_p$  and 0.31 and 0.32 for  $r_g$ , respectively). The very weak  $r_p$  in volume between the first and second rotations was in discrepancy with those observed for 5.5-year-old volume across rotations in *Eucalyptus* clones (more than 0.71; Amâncio et al., 2020). Such a discrepancy may reflect that across-rotation correlations are population and/or age-specific.

# 3.4 Multitrait selection, relative genetic gain, and selection efficiency

Multitrait SI calculations were accomplished with the following four scenarios:  $10.90V_8 + 11.96BD_8 + 0.50CC_8$  (scenario 1),  $2.12V_{15} + 10.33BD_{15}$  (scenario 2),  $11.27V_8 + 18.48BD_8 + 0.22CC_8 + 0.50CP_{0.5} + 0.50CP_{0.5} + 0.50CP_{0.5}$ 

22.31 $Vs_{2.5}$  (scenario 3), and  $3.50V_{15}$  +  $4.61BD_{15}$  +  $0.18CP_{0.5}$  + 12.01Vs<sub>2.5</sub> (scenario 4). This indicates that the relative economic weight of an objective trait can vary with the traits involved. A total of 48, 44, 41, and 41 clones were selected under scenarios 1-4, resulting in selection differentials of 4.14, 3.62, 7.00, and 3.40, respectively (Supplementary Table 4). For MGIDI selection, scenario 1', 2', 3', and 4' involved the same traits as the counterpart scenarios 1, 2, 3, and 4, respectively. Only factors with eigenvalues of more than 1.00 were included, leading to one, one, two, and two factors with an accumulated variance of 56.5%, 60.0%, 60.1%, and 66.8% in scenarios 1'-4', respectively (Table 2). Selection differentials ranged from - 1.55 to - 1.33, with the smaller value being the better. Scenarios 1'-4' shared 35 (72.9%), 39 (88.6%), 24 (58.5%), and 35 (85.4%) selected clones, respectively, with the counterpart SI scenario (Supplementary Table 4). In addition, single-trait selection for  $V_{1.5}$ , V2.5, V4.5, V5.5, V6.5, V8, BD8, CC8, V10, V12, V15, DB15, CP0.5, V\$1.5, and Vs<sub>2.5</sub> resulted in 0.003-0.52 of selection differential in BV, with 60-41 clones selected (Supplementary Tables 5, 6).

RG estimates for all the traits involved in multitrait selection scenarios were positive (Figure 2), indicating the usefulness of all the scenarios for the combined selection of these traits. Of the trait types, growth of the first rotation ( $V_8$  and  $V_{15}$ ) showed consistently the highest RG (50.6%-68.7%) as compared to wood properties (*BD*<sub>8</sub>, *CC*<sub>8</sub>, and *BD*<sub>15</sub>; 0.46%–4.2%), coppicing (*CP*<sub>0.5</sub>; 6.4%–13.1%), and growth of the second rotation (Vs<sub>2.5</sub>; 14.0-17.7%). These estimates are extremely high for growth but comparable for wood properties relative to genetic gains reported by others for the selection of Eucalyptus clones, e.g., 7.5%-20.4% for volume and -0.3% to 0.4% for cellulose content in E. urophylla × E. grandis clones selected using an equal emphasis SI method (Bouvet et al., 2020). In terms of RG comparison between selection methods, SI was always preferred for a higher RG in the first-rotation growth  $V_8$  and  $V_{15}$ whereas MGIDI was more favorable for wood properties BD<sub>8</sub>, CC<sub>8</sub>, and  $BD_{15}$  except for  $BD_{15}$  with slightly lower RG in scenario 4'. In addition, single-trait selections resulted in RG of 45.1%-79.1% in the first-rotation growth, 3.4%-4.6% in wood properties, 16.1% in coppicing potential, and 38.8% in Vs<sub>2.5</sub> (Supplementary Table 5).

In accordance with RG estimates, E values for all the multitrait selection scenarios relative to each single-trait selection were also positive (Figure 3) but varied largely from 10.6% (BD15 with MGIDI) to 97.9% ( $V_8$  with SI). As compared to the SI method, MGIDI showed somewhat lower E in first-rotation growth  $V_8$  and  $V_{15}$  but higher E in wood properties  $BD_8$ ,  $CC_8$ , and  $BD_{15}$  except for BD<sub>15</sub> with slightly lower E in scenario 4'. Similarly, in Avena strigosa S., desirable selection efficiency varied with the selection method (SI or MGIDI) for the same trait (Klein et al., 2023). In addition, E of earlier single-trait selections in volume was relatively high for  $V_8$  (70.7%–95.8%), low to moderately high for  $V_{15}$  (33.0%– 76.9%), BD<sub>8</sub> (31.4%-51.0%), and CP<sub>0.5</sub> (27.3%-46.5%), very low to low for  $CC_8$  (- 0.3% to 0.3%),  $Vs_{1.5}$  (- 16.1% to 7.4%), and  $Vs_{2.5}$  (-5.1% to 12.8% with the only exception of 88.5% by  $Vs_{1.5}$ ), and extremely low for  $BD_{15}$  (- 250.9% to - 69.7%; Supplementary Table 5). Specifically, single-trait selections in  $V_8$ ,  $V_8$ , and  $V_{15}$  gave rise to E of 42.1%, 0.04%, and – 120.6% for wood properties BD<sub>8</sub>, CC<sub>8</sub>, and BD<sub>15</sub>, respectively (Supplementary Table 5), being somewhat less than those of multitrait selections. Meanwhile, single-trait selections in volume at ages 2.5 and 6.5 could have brought about *E* more than 80% and 70% for  $V_8$  and  $V_{15}$ , respectively, indicating the possibility of effective volume selection at less than half-rotation age.

## 3.5 Implications for eucalypt clonal breeding

Clonal tests usually focus on the first rotation performance in forest trees such as eucalypts (Lambeth et al., 1994; Osorio et al., 2001, 2003; Reis et al., 2011; Amâncio et al., 2020; Romão et al., 2023) and poplars (Marron and Ceulemans, 2006; Rönnberg-Wästljung et al., 2022). This can be meaningful for estimating genetic parameters and evaluating clonal performance within a single rotation, especially for those plantation trees regenerated merely through means of replanting. However, in regard to across-rotation trait variability and management regime shifts, information only on the first rotation may not be enough for trees in plantation systems amenable to coppice regeneration. Here, the weak or moderate  $r_p$  and  $r_q$  in volume between rotations (e.g.,  $r_p$  and  $r_g$  between  $V_{15}$  and  $Vs_{2.5}$ being - 0.06 and 0.32, respectively; Figure 1C, Supplementary Table 3), coupled with remarkably poor selection efficiency in volume across rotations (e.g., 0.7% of V15 on Vs2.5; Supplementary Table 5) and relatively low proportion of common clones between single- and across-rotation multitrait selection scenarios (e.g., 56.1% between scenarios 1' and 4'; Supplementary Table S4), may justify the acrossrotation growth variability and the necessity of across-rotation investigation. Also, across-rotation growth variability was noted in clones of other eucalypt taxa, which may be related to management regime shifts such as retention of more than one sprout per tree (Amâncio et al., 2020).

Both SI and MGIDI methods are effective in guaranteeing desirable positive RG and E for all the selection traits (Figures 2, 3),

demonstrating the potential for simultaneous improvement of the traits involved. Nevertheless, the two methods differed more or less in selected clones (Supplementary Table 4). Such difference in selected genotypes was also noticed between SI and MGIDI in Zea mays L. (Yue et al., 2022) and Avena strigosa S (Klein et al., 2023). As MGIDI uses a factor analysis process free from such economic coefficient weightings and multicollinearity issues that SI relies on, it brings more balanced genetic gains (Klein et al., 2023). This seems to be the case for E.  $urophylla \times E.$  tereticornis clones, as each of the MGIDI selection scenarios shows a narrower range of RG (also E) values than the respective SI scenarios (Figures 2, 3). Therefore, neither of the two methods can fit all multitrait selection circumstances in our eucalypt hybrid clones, and the choice of method depends on the tradeoff of expected genetic gains among the traits involved. If a higher RG (also E) is expected for a specific target trait, the SI method with a higher weighting of the trait would be preferred. Moreover, as a factor analysis tends to group positively and negatively correlated variables into the same factor (Joliffe and Morgan, 1992), MGIDI is limited in selection for two traits in opposite selection gains (Olivoto et al., 2022).

Early selection is attractive in tree breeding because selection prior to rotation age can capture genetic gains sooner and therefore increase gains achievable per unit of time (White et al., 2007). The efficiency of early selection is affected by the heritability of a measured trait and its genetic correlation with a target trait (Jansson et al., 2003; White et al., 2007). In the present study, like many previous reports for eucalypts (e.g. Osorio et al., 2001, 2003; He et al., 2012; Araujo et al., 2021; Miranda et al., 2024), the heritability estimates of volume are similar magnitude over ages within the first rotation, and age–age genetic correlations should have a more pronounced influence on selection efficiency. In consideration of those strong correlations between earlier and rotation ages (age 8 for veneer and pulping and 15 for large-size timber purposes), together with the favorable E estimates (Supplementary Table 5), half-rotation or somewhat earlier age

TABLE 2 Factors retained in the multitrait genotype-idiotype distance index (MGIDI) analysis along with their eigenvalues, explained variances, and accumulative variances.

Trait	Scenario 1': V <sub>8</sub> +BD <sub>8</sub> +CC <sub>8</sub>	Scenario 2': <i>V</i> <sub>15</sub> + <i>BD</i> <sub>15</sub>	Scenario 3': $V_8+BD_8$ + $CC_8+CP_{0.5}+Vs_{2.5}$		Scenario 4': V <sub>15</sub> +BD <sub>15</sub> +CP <sub>0.5</sub> +Vs <sub>2.5</sub>	
	Factor 1	Factor 1	Factor 1	Factor 2	Factor 1	Factor 2
$V_8$	- 0.76	-	- 0.79	- 0.14	-	-
$BD_8$	- 0.87	-	- 0.85	0.07	-	-
CC <sub>8</sub>	- 0.60	-	- 0.47	- 0.16	-	-
$V_{15}$	-	0.77	-	-	0.79	0.24
BD <sub>15</sub>	-	0.77	-	-	0.30	0.78
CP <sub>0.5</sub>	-	-	- 0.46	- 0.60	0.79	- 0.18
Vs <sub>2.5</sub>	-	-	0.04	- 0.90	0.41	- 0.69
Eigenvalue	1.69	1.20	1.97	1.03	1.50	1.17
Variance explained (%)	56.5	60.0	39.4	20.7	37.5	29.3
Accumulative variance (%)	-	-	-	60.1	-	66.8

Trait abbreviations are provided in the section of Glossary.



can be determined for early growth selection, that is, age 4.5 or 2.5 for veneer and pulp usages and age 8.5 or 6.5 for large-diameter timber production. Similar early selection timelines have previously been proposed for *Eucalyptus* clones (Osorio et al., 2003; Massaro et al., 2010; He et al., 2012; Moraes et al., 2014; Yang et al., 2018).

The finding of significant differences between ramet means and

ortet values for most traits, especially those involved in multitrait

selection such as  $V_8$ ,  $V_{15}$ , and  $CP_{0.5}$  ( $p \le 0.001$  or 0.01; Supplementary Table 2), can also have implications for clonal breeding. Firstly, seedling-based genetic information such as heritability (or repeatability), genetic correlation, and breeding value cannot be used for clonal selection. This is true particularly for low-heritable traits, as seedling-clone correlation depends largely on the magnitude of genetic control of the trait concerned (Costa e Silva et al., 2013).

в А 86.7 120 Scenario 1 90 Scenario 2 80.5 Scenario 2 Scenario 1 90 97.9 8100 86.6 70 Selection efficiency 76.3 58.4 80 60 53.6 66 4 50 60 45.8 40 40 30 20 12.5 20 10 0 n BD<sub>15</sub>  $V_{s}$ BD. CC.  $V_{_{15}}$ С D <sup>100</sup> [ 88.1 Scenario 3 100 Scenario 4 90.2 89.4 Scenario 3' 90 Scenario 4 90 81.4 Selection efficiency (%) 78.9 78.3 777 80 80 69.3 70 70 61.7 60 60 45.8 45.5 45.2 50 50 39.9 39.0 36.0 40 40 30 30 19.1 18.0 20 20 10.6 10 10 Λ Λ  $V_{s}$  $BD_8$  $CC_8$ CP<sub>0.5</sub> Vs<sub>2.5</sub> V<sub>15</sub> BD<sub>15</sub> CP<sub>0.5</sub> Vs<sub>2.5</sub> FIGURE 3

Selection efficiency (*E*, %) of multitrait selection scenarios relative to each single-trait selection in a cloned *E. urophylla* × *E. tereticornis* cross. Trait abbreviations are provided in the section of Glossary.

Secondly, direct clonal selection may be the desirable option rather than the inclusion of prior seedling progeny test data. Also, van den Berg et al. (2015) questioned the feasibility of using prior seedling selection for *E. grandis* and *E. urophylla* clonal testing. Thirdly, the gap between seedling and clonal selections may be bridged by molecular marker-assisted tools such as genomic selection (GS). GS offers an efficient tool to screen a large number of genotypes, reduce the time for selection of an operational clone, and consequently increase genetic gains (Balocchi et al., 2023). GS has been recently conducted for *Eucalyptus* clone selection in a number of eucalypt breeding programs (Torres-Dini et al., 2016; Durán et al., 2017; Balocchi et al., 2023). If a GS model is validated efficiently for predicting

performance in a clonal population, it can be applied to screen much larger seedling (also clonal) populations of the kind at a considerably low selection intensity. Consequently, only a small number of clones (say, 30 to 50) identified by GS need to be field validated, and the genetic gain will increase in the clonal deployment population (Balocchi et al., 2023).

# 4 Conclusion

This study provides novel information on across-rotation genetic parameters and multitrait selection methods based on a cloned E. urophylla × E. tereticornis cross. The high progeny phenotypic variability suggests the potential of clone selection within a single cross. The relatively high  $H^2$  estimates for some traits reveal high genetic control of such traits. The generally weak or moderate correlations in growth between rotations may justify the acrossrotation growth variability and the necessity of across-rotation investigation. Neither the SI nor the MGIDI method can fit all multitrait selection circumstances, and the choice of method depends on the tradeoff of expected genetic gains among the traits involved. Halfrotation or somewhat earlier age can be used for the early selection of growth traits. Direct clonal selection may be desirable rather than the inclusion of prior seedling progeny tests, and seedling-based selection may be integrated using molecular marker-assisted approaches. The results can have important implications for eucalypt clone breeding and management, including the necessity of across-rotation investigation, evaluation of multitrait selection options, determination of selection ages, and the possible use of seedling deployment population.

## Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

## Author contributions

JX: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Visualization, Writing – original draft, Writing – review & editing. MZ: Formal analysis, Investigation, Writing – review & editing. QW: Data curation, Investigation, Resources, Writing – review & editing. ML: Conceptualization, Data curation, Investigation, Methodology, Project administration, Writing – review & editing. SG: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Supervision, Writing – original draft, Writing – review & editing.

## Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was financially supported by the Forestry Administration of Guangdong Province, China (2022KJCX024).

## Acknowledgments

The authors thank Juying Wu and Fagen Li for their valuable cooperation in establishing and maintaining the plant material, as well as for their assistance with trait observation. We are also grateful to Roger Arnold for his critical reading and valuable suggestions that improved the manuscript. We appreciate the assistance of Zhaoyuan Zhang, Jingquan Lin, Yong Guo, Changpin Zhou, Xiaoli Yu, and Heyu Yang in the field trial investigation and wood sample collection.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2025.1553819/ full#supplementary-material

#### 10.3389/fpls.2025.1553819

## References

Alexandru, A.-M., Mihai, G., Stoica, E., and Curtu, A. L. (2023). Multi-trait selection and stability in Norway spruce (*Picea abies*) provenance trials in Romania. *Forests* 14, 456. doi: 10.3390/f14030456

Amâncio, M. R., Pereira, F. B., Paludeto, J. G. Z., Vergani, A. R., Bison, O., Peres, F. S. B., et al. (2020). Genetic control of coppice regrowth in Eucalyptus spp. *Silvae Genet.* 69, 6–12. doi: 10.2478/sg-2020-0002

Antony, F., Schimleck, L. R., Jordan, L., Hornsby, B., Dahlen, J., Daniels, R. F., et al. (2014). Growth and wood properties of genetically improved loblolly pine: propagation type comparison and genetic parameters. *Can. J. For. Res.* 44, 263–272. doi: 10.1139/ cjfr-2013-0163

Araujo, M. J., Lee, D. J., Tambarussi, E. V., Paula, R. C., and Silva, P. H. M. (2021). Initial productivity and genetic parameters of three *Corymbia* species in Brazil: designing a breeding strategy. *Can. J. For. Res.* 51, 25–30. doi: 10.1139/cjfr-2019-0438

Arnold, R. J., Xie, Y. J., Luo, J. Z., Wang, H. R., and Midgley, S. J. (2020). A tale of two genera: Exotic *Eucalyptus* and *Acacia* species in China. 2. Plantation resource development. *Int. For. Rev.* 22, 153–168. doi: 10.1505/146554820829403441

Balocchi, C., Durán, R., Nuñez, P., Ordoñez, J., Ramírez, M., and Zapata-Valenzuela, J. (2023). Genomic selection: an effective tool for operational *Eucalyptus globulus* clonal selection. *Tree Genet. Genomes* 19, 32. doi: 10.1007/s11295-023-01609-7

Bates, D., Mächler, M., Bolker, B., and Walker, S. (2015). Fitting linear mixed-effects models using lme4. J. Stat. Software 67, 1–48. doi: 10.18637/jss.v067.i01

Berlin, M., Jansson, G., Högberg, K.-A., and Helmersson, A. (2019). Analysis of nonadditive genetic effects in Norway spruce. *Tree Genet. Genomes* 15, 42. doi: 10.1007/ s11295-019-1350-9

Borralho, N. M. G., and Kanowski, P. J. (1995). Correspondence of performance between genetically related clones and seedlings. *Can. J. For. Res.* 25, 500–506. doi: 10.1139/x95-05

Bouvet, J.-M., Ekomono, C. G. M., Brendel, O., Laclau, J.-P., Bouillet, J.-P., and Epron, D. (2020). Selecting for water use efficiency, wood chemical traits and biomass with genomic selection in a Eucalyptus breeding program. *For. Ecol. Manage.* 465, 118092. doi: 10.1016/j.foreco.2020.118092

Braga, R. C., Paludeto, J. G. Z., Souza, B. M., Aguiar, A. V., Pollnow, M. F. M., Carvalho, A. G. M., et al. (2020). Genetic parameters and genotype × environment interaction in *Pinus taeda* clonal tests. *For. Ecol. Manage.* 474, 118342. doi: 10.1016/j.foreco.2020.118342

Burdon, R. D., and Aimers-Halliday, J. (2006). "Managing risk in clonal forestry," in CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources 1, No. 035 (CABI Publishing, Delémont, Switzerland). doi: 10.1079/ PAVSNNR20061035

Butler, D. G., Cullis, B. R., Gilmour, A. R., Gogel, B. G., and Thompson, R. (2017). ASReml-R Reference Manual Version 4 (Hemel Hempstead, U.K: VSN International Ltd).

Chen, S., Xie, Y., Zhou, Q., Li, H., Li, T., Liu, X., et al. (2017). "Technical regulations on eucalypt cultivation for large-size timber," in *Forestry Standard LY/T 2909-2017, People's Republic of China* (National Forestry Administration, Beijing, China).

Chen, Z.-Q., Hai, H. N. T., Helmersson, A., Liziniewicz, M., Hallingbäck, H. R., Fries, A., et al. (2020). Advantage of clonal deployment in Norway spruce (*Picea abies* (L.) H. Karst). *Ann. For. Sci.* 77, 14. doi: 10.1007/s13595-020-0920-1

Costa e Silva, J., Potts, B. M., and Tilyard, P. (2013). Stability of genetic effects across clonal and seedling populations of *Eucalyptus globulus* with common parentage. *For. Ecol. Manage.* 291, 427–435. doi: 10.1016/j.foreco.2012.11.005

Cotterill, P. P., and Dean, C. A. (1990). *Successful tree breeding with index selection* (East Melbourne, Victoria, Australia: CSIRO).

Crous, J. W., and Burger, L. (2015). A comparison of planting and coppice regeneration of *Eucalyptus grandis* × *Eucalyptus urophylla* clones in South Africa. *South. For.* 77, 277–285. doi: 10.2989/20702620.2015.1063031

Durán, R., Isik, F., Zapata-Valenzuela, J., Balocchi, C., and Valenzuela, S. (2017). Genomic predictions of breeding values in a cloned *Eucalyptus globulus* population in Chile. *Tree Genet. Genomes* 13, 74. doi: 10.1007/s11295-017-1158-4

Flood, P. J., Kruijer, W., Schnabel, S. K., van der Schoor, R., Jalink, H., Snel, J. F. H., et al. (2016). Phenomics for photosynthesis, growth and reflectance in Arabidopsis thaliana reveals circadian and long-term fluctuations in heritability. Plant Methods 12, 14. doi: 10.1186/s13007-016-0113-y

Foster, G. S., and Shaw, D. V. (1988). Using clonal replicates to explore genetic variation in a perennial plant species. *Theor. Appl. Genet.* 76, 788–794. doi: 10.1007/BF00303527

Gallo, R., Pantuza, I. B., Santos, G. A., Resende, M. D. V., Xavier, A., Simiqueli, G. F., et al. (2018). Growth and wood quality traits in the genetic selection of potential *Eucalyptus dunnii* Maiden clones for pulp production. *Ind. Crops Prod.* 123, 434–441. doi: 10.1016/j.indcrop.2018.07.016

Gan, S., Li, M., Li, F., Wu, K., Wu, J., Lu, G., et al. (2006). Analysis on cutting and growth traits of clones of *Eucalyptus urophylla* × *E. tereticornis. For. Res.* 19, 135–140.

Hamilton, M. G., Raymond, C. A., Harwood, C. E., and Potts, B. M. (2009). Genetic variation in *Eucalyptus nitens* pulpwood and wood shrinkage traits. *Tree Genet. Genomes* 5, 307–316. doi: 10.1007/s11295-008-0179-4

Hardiyanto, E. B., Inail, M. A., Mendham, D. S., Thaher, E., and Sitorus, B. K. (2022). *Eucalyptus pellita* coppice vs. seedlings as a re-establishment method in South Sumatra, Indonesia. *Forests* 13, 1017. doi: 10.3390/f13071017

Hazel, L. N. (1943). The genetic basis for constructing selection indexes. *Genetics* 28, 476–490. doi: 10.1093/genetics/28.6.476

He, X., Li, F., Li, M., Weng, Q., Shi, J., Mo, X., et al. (2012). Quantitative genetics of cold hardiness and growth in *Eucalyptus* as estimated from *E. urophylla* × *E. tereticornis* hybrids. *New For.* 43, 383–394. doi: 10.1007/s11056-011-9287-3

Hernández, M. A., Butler, J. B., Ammitzboll, H., Freeman, J. S., O'Reilly–Wapstra, J., Vaillancourt, R. E., et al. (2022). Genetic variation in fire recovery and other fire-related traits in a global eucalypt species. *Tree Genet. Genomes* 18, 42. doi: 10.1007/s11295-022-01572-9

Isik, F., Goldfarb, B., LeBude, A., Li, B., and McKeand, S. (2005). Predicted genetic gains and testing efficiency from two loblolly pine clonal trials. *Can. J. For. Res.* 35, 1754–1766. doi: 10.1139/x05-064

Isik, F., Li, B., and Frampton, J. (2003). Estimates of additive, dominance and epistatic genetic variances from a clonally replicated test of loblolly pine. *For. Sci.* 49, 77–88. doi: 10.1093/forestscience/49.1.77

Jansson, G., Li, B., and Hannrup, B. (2003). Time trends in genetic parameters for height and optimal age for parental selection in Scots pine. *For. Sci.* 49, 696–705. doi: 10.1093/forestscience/49.5.696

Joliffe, I., and Morgan, B. (1992). Principal component analysis and exploratory factor analysis. *Stat. Methods Med. Res.* 1, 69–95. doi: 10.1177/096228029200100105

Kien, N. D., Jansson, G., Harwood, C., and Almqvist, C. (2010). Clonal variation and genotype by environment interactions in growth and wood density in *Eucalyptus camaldulensis* at three contrasting sites in Vietnam. *Silvae Genet.* 59, 17–28. doi: 10.1515/sg-2010-0003

Klein, L. A., Marchioro, V. S., Toebe, M., Olivoto, T., Meira, D., Meier, C., et al. (2023). Selection of superior black oat lines using the MGIDI index. *Crop Breed. Appl. Biotechnol.* 23, e45112332. doi: 10.1590/1984-70332023v23n3a25

Kleinschmit, J., Khurana., D. K., Gerhold, H. D., and Libby, W. J. (1993). "Past, present and anticipated applications of clonal forestry," in Clonal Forestry II, Conservation and Application. Eds. M. R. Ahuja and W. J. Libby (Springer-Verlag, Berlin, Germany), 9–41. doi: 10.1007/978-3-642-84813-1\_2

Lambeth, C., Endo, M., and Wright, J. (1994). Genetic analysis of 16 clonal trials of *Eucalyptus grandis* and comparisons with seedling checks. *For. Sci.* 40, 397–411. doi: 10.1093/forestscience/40.3.397

Liao, T., Cheng, S., Zhu, X., Min, Y., and Kang, X. (2016). Effects of triploid status on growth, photosynthesis, and leaf area in *Populus. Trees* 30, 1137–1147. doi: 10.1007/s00468-016-1352-2

Marron, N., and Ceulemans, R. (2006). Genetic variation of leaf traits related to productivity in a *Populus deltoides* × *Populus nigra* family. *Can. J. For. Res.* 36, 390–400. doi: 10.1139/X05-245

Massaro, R. A. M., Bonine, C. A. V., Scarpinati, E. A., and Paula, R. C. (2010). Early selection viability in *Eucalyptus* spp clonal tests. *Ciênc. Florest.* 20, 597–609. doi: 10.5902/198050982418

Midgley, S. J. (2013). Making a difference: celebrating success in Asia. Aust. For. 76, 73–75. doi: 10.1080/00049158.2013.790098

Miranda, L., Estopa, R. A., Paludeto, J. G. Z., and Tambarussi, E. V. (2024). Genetic control and early selection of three *Corymbia* species. *Can. J. For. Res.* 54, 31–44. doi: 10.1139/cjfr-2023-0038

Moraes, C. B., Brizolla, T. F., Teixeira, L. G., Zimback, L., Tambarussi, E. V., Chaves, R., et al. (2014). Estimates of genetic parameters for selection of trees in *Eucalyptus. Sci. For.* 42, 623–629.

Mullin, T. J., and Park, Y. S. (1992). Estimating genetic gains from alternative breeding strategies for clonal forestry. *Can. J. For. Res.* 22, 14–23. doi: 10.1139/x92-003

Olivoto, T., Diel, M. I., Schmidt, D., and Lúcio, A. D. (2022). MGIDI: a powerful tool to analyze plant multivariate data. *Plant Methods* 18, 121. doi: 10.1186/s13007-022-00952-5

Olivoto, T., and Lúcio, A. D. (2020). metan: An R package for multi-environment trial analysis. *Methods Ecol. Evol.* 11, 783-789. doi: 10.1111/2041-210X.13384

Olivoto, T., and Nardino, M. (2021). MGIDI: Toward an effective multivariate selection in biological experiments. *Bioinformatics* 37, 1383–1389. doi: 10.1093/bioinformatics/btaa981

Ortiz, D., and Salas-Fernandez, M. G. (2022). Dissecting the genetic control of natural variation in sorghum photosynthetic response to drought stress. *J. Exp. Bot.* 73, 3251–3267. doi: 10.1093/jxb/erab502

Osorio, L. F., White, T. L., and Huber, D. A. (2001). Age trends of heritabilities and genotype-by-environment interactions for growth traits and wood density from clonal trials of *Eucalyptus grandis* Hill ex Maiden. *Silvae Genet.* 50, 108–116.

Osorio, L. F., White, T. L., and Huber, D. A. (2003). Age-age and trait-trait correlations for *Eucalyptus grandis* Hill ex Maiden and their implications for optimal selection age and design of clonal trials. *Theor. Appl. Genet.* 106, 735–743. doi: 10.1007/s00122-002-1124-9

Peng, S., Xu, J., Li, G., and Chen, Y. (2013). Growth and genetic analysis of 42 *Eucalyptus urophylla* × *E. tereticornis* clones in Leizhou peninsula in China. *J. Cent.* South Univ. For. Tech. 33, 23–27. doi: 10.14067/j.cnki.1673-923x.2013.04.018

Pietrini, F., Iori, V., Bianconi, D., Mughini, G., Massacci, A., and Zacchini, M. (2015). Assessment of physiological and biochemical responses, metal tolerance and accumulation in two eucalypt hybrid clones for phytoremediation of cadmium-contaminated waters. *J. Environ. Manage.* 162, 221–231. doi: 10.1016/j.jenvman.2015.07.053

Quesada, T., Parisi, L. M., Huber, D. A., Gezan, S. A., Martin, T. A., Davis, J. M., et al. (2017). Genetic control of growth and shoot phenology in juvenile loblolly pine (*Pinus taeda* L.) clonal trials. *Tree Genet. Genomes* 13, 65–80. doi: 10.1007/s11295-017-1143-y

Rashidi-Jouybari, I., Lenz, P., Beaulieu, J., Nadeau, S., Jean Bousquet, J., and Achim, A. (2022). Multi-trait selection for improved solid wood physical and flexural properties in white spruce. *Forestry* 95, 492–503. doi: 10.1093/forestry/cpac006

R Core Team (2022). R: A language and environment for statistical computing (Vienna: R Foundation for Statistical Computing). Available at: https://cran.r-project. org/bin/windows/base.

Reis, C. A. F., Gonçalves, F. M. A., Rosse, L. N., Costa, R. R. G. F., and Ramalho, M. A. P. (2011). Correspondence between performance of *Eucalyptus* spp trees selected from family and clonal tests. *Genet. Mol. Res.* 10, 1172–1179. doi: 10.4238/vol10-2gmr1078

Rezende, G. D. S. P., Lima, J. L., Dias, D. C., Lima, B. M., Aguiar, A. M., Bertolucci, F. L. G., et al. (2019). Clonal composites: An alternative to improve the sustainability of production in eucalypt forests. *For. Ecol. Manage.* 449, 117445. doi: 10.1016/j.foreco.2019.06.042

Romão, K. C. S., Nunes, A. C. P., Santos, A. P., Santos, O. P., and Valente, B. M. R. T. (2023). Multi-site comparison of clonal arrangements for tropical and subtropical hybrids of *Eucalyptus. Ind. Crops Prod.* 197, 116536. doi: 10.1016/j.indcrop.2023.116536

Rönnberg-Wästljung, A. C., Adler, A., Karacic, A., Liepins, K., Richards, T. J., Ingvarsson, P. K., et al. (2022). Phenotypic plasticity in *Populus trichocarpa* clones across environments in the Nordic–Baltic region. Scand. *J. For. Res.* 37, 1–5. doi: 10.1080/02827581.2022.2039279

RStudio Team (2021). *RStudio: Integrated development for R* (Boston, MA, USA). Available at: http://www.rstudio.com/.

Santos, N. Z., Piepho, H. P., Condorelli, G. E., Groli, E. L., Newcomb, M., Ward, R., et al. (2021). High-throughput field phenotyping reveals genetic variation in photosynthetic traits in durum wheat under drought. *Plant Cell Environ.* 44, 2858–2878. doi: 10.1111/pce.14136

Sasse, J., and Sands, R. (1997). Configuration and development of root systems of cuttings and seedlings of *Eucalyptus globulus*. *New For*. 14, 85–105. doi: 10.1023/A:1006536512910

Shalizi, M. N., Gezan, S. A., McKeand, S. E., Sherrill, J. R., Cumbie, W. P., Whetten, R. W., et al. (2020). Correspondence between breeding values of the same *Pinus taeda* L. genotypes from clonal trials and half-sib seedling progeny trials. *For. Sci.* 66, 600–611. doi: 10.1093/forsci/fxaa016

Shalizi, M. N., and Isik, F. (2019). Genetic parameter estimates and G×E interaction in a large cloned population of *Pinus taeda L. Tree Genet. Genomes* 15, 46. doi: 10.1007/ s11295-019-1352-7

Singamsetti, A., Zaidi, P. H., Seetharam, K., Vinayan, M. T., Olivoto, T., Mahato, A., et al. (2023). Genetic gains in tropical maize hybrids across moisture regimes with multi-trait based index selection. *Front. Plant Sci.* 14. doi: 10.3389/fpls.2023.1147424

Singh, B. D. (2001). Plant Breeding: Principles and Methods (New Delhi, India: Kalyani).

Smith, H. F. (1936). A discriminant function of plant selection. Ann. Eugenics 7, 240–250. doi: 10.1111/j.1469-1809.1936.tb02143.x

Soares, A. A. V., Leite, H. G., Souza, A. L., Silva, S. R., Lourença, H. M., and Forrester, D. I. (2016). Increasing stand structural heterogeneity reduces productivity in Brazilian *Eucalyptus* monoclonal stands. *For. Ecol. Manage.* 373, 26–32. doi: 10.1016/j.foreco.2016.04.035

Stackpole, D. J., Vaillancourt, R. E., Aguigar, M., and Potts, B. (2010). Age trends in genetic parameters for growth and wood density in *Eucalyptus globulus*. *Tree Genet. Genomes* 6, 179–193. doi: 10.1007/s11295-009-0239-4

Stelzer, H. E., and Goldfarb, B. (1997). Implementing clonal forestry in south eastern United States: SRIEG satellite workshop summary remarks. *Can. J. For. Res.* 27, 442–446. doi: 10.1139/x96-200

Torres-Dini, D., Nunes, A. C. P., Aguiar, A., Nikichuk, N., Centurión, C., Cabrera, M., et al. (2016). Clonal selection of *Eucalyptus grandis × Eucalyptus globulus* for productivity, adaptability, and stability, using SNP markers. *Silvae Genet.* 65, 30–38. doi: 10.1515/sg-2016-0014

van den Berg, G. J., Verryn, S. D., Chirwa, P. W., and van Deventer, F. (2015). Genetic parameters of interspecific hybrids of *Eucalyptus grandis* and *E. urophylla* seedlings and cuttings. *Silvae Genet.* 64, 291–308. doi: 10.1515/sg-2015-0027

Wang, T., Ren, J., Huang, Q., and Li, J. (2024). Genetic parameters of growth and leaf traits and genetic gains with MGIDI in three *Populus simonii* × *P. nigra* families at two spacings. *Front. Plant Sci.* 15. doi: 10.3389/fpls.2024.1483580

Wei, R. P. (2005). "Genetic diversity and sustainable productivity of eucalypt plantations in China," in *Changing Patterns: Tree Introduction and Phytogeography*. Eds. H. Wang, Z. Jiang and Y. Li (China Forestry Publishing House, Beijing, China), 19–27.

White, T. L. (1996). "Genetic parameter estimates and breeding value predictions: issues and implications in tree improvement programs," in *Proceedings of QFRI-IUFRO Conference on Tree Improvement for Sustainable Tropical Forestry*. Eds. M. J. Dieters, A. C. Matheson, D. G. Nikles, C. E. Harwood and S. M. Walker (QFRI-IUFRO, Caloundra, Queensland, Australia).

White, T. L., Adams, W. T., and Neale, D. B. (2007). *Forest Genetics* (England: CABI).

Wu, H. X. (2019). Benefits and risks of using clones in forestry – a review. Scand. J. For. Res. 34, 352–359. doi: 10.1080/02827581.2018.1487579

Wujeska-Klausea, A., Crous, K. Y., Ghannouma, O., and Ellsworth, D. S. (2019). Leaf age and eCO<sub>2</sub> both influence photosynthesis by increasing light harvesting in mature *Eucalyptus tereticornis* at EucFACE. *Env. Exp. Bot.* 167, 103857. doi: 10.1016/j.envexpbot.2019.103857

Xiong, J. S., McKeand, S. E., Whetten, R. W., and Isik, F. T. (2014). Genetics of stem forking and ramicorn branches in a cloned loblolly pine family. *For. Sci.* 60, 360–366. doi: 10.5849/forsci.12-018

Yang, H., Weng, Q., Li, F., Zhou, C., Li, M., Chen, S., et al. (2018). Genotypic variation and genotype-by-environment interactions in growth and wood properties in a cloned *Eucalyptus urophylla × E. tereticornis* family in southern China. *For. Sci.* 64, 225–232. doi: 10.1093/forsci/fxx011

Yue, H., Wei, J., Xie, J., Chen, S., Peng, H., Cao, H., et al. (2022). A study on genotypeby-environment interaction analysis for agronomic traits of maize genotypes across Huang-Huai-Hai region in China. *Phyton* 91, 57-81. doi: 10.32604/ phyton.2022.017308

Zeltiņš, P., Kangur, A., Katrevičs, J., and Jansons, A. (2022). Genetic parameters of diameter growth dynamics in Norway spruce clones. *Forests* 13, 679. doi: 10.3390/f13050679

Zhou, H. H., Chen, Y. N., Li, W. H., and Chen, Y. P. (2010). Photosynthesis of *Populus euphratica* in relation to groundwater depths and high temperature in arid environment, northwest China. *Photosynthetica* 48, 257–268. doi: 10.1007/s11099-010-0032-5

Zhou, X. G., Ye, D., Zhu, H. G., Li, X. Q., Su, Y., Lan, J., et al. (2017). Effects of second rotation seedlings and coppice on understory vegetation and timber production of *Eucalyptus* plantations. *J. Trop. For. Sci.* 29, 54–68. Available at: https://www.jstor.org/stable/44028275 (Accessed April 27, 2024).

### 10.3389/fpls.2025.1553819

Glossary				
MGIDI	multitrait genotype-idiotype distance index	ETR	photosynthetic electron transport rate under light-adaptation	
$H^2$	clonal repeatability	NPQ	nonphotochemical quenching	
RG	relative genetic gain (%)	SPADR	SPAD (Spectrum Technologies Inc.) reading	
Ε	selection efficiency (%)	HTs1 <sub>1.5</sub>	HTs1 <sub>2.5</sub> , DBHs1 <sub>1.5</sub> , DBHs1 <sub>2.5</sub> , Vs1 <sub>1.5</sub> , and Vs1 <sub>2.5</sub> , 1.5- and 2.5-	
HT <sub>n</sub>	$HT_{0.5}, HT_{1.5}, HT_{2.5}, HT_{4.5}, HT_{5.5}, HT_{6.5}, HT_8, HT_{10}, HT_{12}$ , and $HT_{15}$ , tree height (m) at age of around 0.5, 1.5, 2.5, 4.5, 5.5, 6.5, 8, 10, 12, and 15 years of the first rotation, respectively		year-old height (m), diameter at breast height (cm), and volume $(m^3)$ of the highest sprout for the second rotation, respectively	
DBH <sub>n</sub>	<i>DBH</i> <sub>1.5</sub> , <i>DBH</i> <sub>2.5</sub> , <i>DBH</i> <sub>4.5</sub> , <i>DBH</i> <sub>5.5</sub> , <i>DBH</i> <sub>6.5</sub> , <i>DBH</i> <sub>8</sub> , <i>DBH</i> <sub>10</sub> , <i>DBH</i> <sub>12</sub> , and <i>DBH</i> <sub>15</sub> , diameter at breast height (cm) at age of around 1.5, 2.5, 4.5, 5.5, 6.5, 8, 10, 12, and 15 years of the first rotation, respectively	HTs2 <sub>1.5</sub>	$HTs2_{2.5}$ , $DBHs2_{1.5}$ , $DBHs2_{2.5}$ , $Vs2_{1.5}$ , and $Vs2_{2.5}$ , $1.5$ - and $2.5$ - year-old height (m), diameter at breast height (cm), and volume (m <sup>3</sup> ) of the second highest sprout for the second rotation, respectively	
BD15	15-year-old wood basic density (g/cm <sup>3</sup> ) of the first rotation	Vs <sub>1.5</sub>	the sum of $Vs1_{1.5}$ and $Vs2_{1.5}$	
BD <sub>8</sub>	$CC_8$ , $HC_8$ , $LC_8$ , and $S/G_8$ , 8-year-old wood basic density (g/ cm <sup>3</sup> ), cellulose content (%), hemicellulose content (%), lignin content (%), and syringyl-to-guaiacyl lignin ratio of the first rotation, respectively	Vs <sub>2.5</sub>	the sum of Vs1 <sub>2.5</sub> and Vs2 <sub>2.5</sub>	
		BAs <sub>1.5</sub>	$HTLBs_{1.5}$ , $NBs_{1.5}$ , $CWs1_{1.5}$ , $CWs2_{1.5}$ , $CLs_{1.5}$ , and $CPAs_{1.5}$ , branch angle (°), height of the lowest live branch (m), number of branches, crown width of the highest sprout (m),	
V <sub>n</sub>	$V_{1.5}$ , $V_{2.5}$ , $V_{4.5}$ , $V_{5.5}$ , $V_{6.5}$ , $V_8$ , $V_{10}$ , $V_{12}$ , and $V_{15}$ , volume (m <sup>3</sup> ) at age of around 1.5, 2.5, 4.5, 5.5, 6.5, 8, 10, 12, and 15 years of the first rotation, respectively		crown width of the second highest sprout (m), crown leng (m), and crown projected area (m <sup>2</sup> ) of the 1.5-year-o sprouts per stump for the second rotation, respectively	
${\rm NS}_{0.5}$ $F_0F_{\rm m},$ and $F_{\rm v}~(=F_{\rm m}{-}F_0)$	$HTs1_{0.5}$ , $HTs2_{0.5}$ , $GDs1_{0.5}$ , $GDs2_{0.5}$ , and $CP_{0.5}$ , number of sprouts, height of the highest sprout (m), height of the second highest sprout (m), ground diameter of the highest sprout (cm), ground diameter of the second highest sprout (cm), and coppicing potential measured at 0.5 year after	SD	standard deviation	
		CV	coefficient of variation (%)	
		ANOVA	analysis of variance	
		CVg	coefficient of genetic variation (%)	
	telling, respectively	r <sub>p</sub>	phenotypic correlation	
	after dark adapting, respectively	r <sub>g</sub>	additive genetic correlation	
$Y(II)(=F_v/F_m)$	maximal quantum vield of photosystem II after	SE	standard error	
	dark adaptation	BLUP	best linear unbiased prediction	
$F_{\rm s}F_{\rm m}{'},$ and $F_{\rm v}{'}$ (= $\!F_{\rm s}\!)$	steady-light, maximal, and variable leaf chlorophyll	BV	breeding value	
	fluorescence after light adaptation, respectively	SI	selection index	
$\Upsilon(II)^{\prime} (= F_{\rm v}^{\prime}/F_{\rm m}^{\prime})$	maximai quantum yield of photosystem II after light adaptation	GS	genomic selection.	