



Maternal Deprivation Increased Vulnerability to Depression in Adult Rats Through DRD2 Promoter Methylation in the Ventral Tegmental Area

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Specialty section:

This article was submitted to
Psychopathology,
a section of the journal
Frontiers in Psychiatry

Received: 02 December 2021

Accepted: 21 January 2022

Published: 02 March 2022

Citation:

Guo Z, Li S, Wu J, Zhu X and Zhang Y
(2022) Maternal Deprivation Increased
Vulnerability to Depression in Adult
Rats Through DRD2 Promoter
Methylation in the Ventral Tegmental
Area. *Front. Psychiatry* 13:827667.
doi: 10.3389/fpsy.2022.827667

Objective: Early life adversity is a risk factor for depression in adulthood; however, the underlying mechanisms are not well understood. This study aims to investigate the effect of DNA methylation of DRD2 gene on early life stress-induced depression in adult rats.

Methods: Newborn Sprague–Dawley rats were randomly assigned to four groups: maternal deprivation group (MD), chronic unpredictable stress (CUS) group, maternal deprivation plus chronic unpredictable stress (MD/CUS) group, and normal control group (NOR). Behaviors were measured by open field test (OFT), sucrose preference test (SPT), and Original Research Article forced swimming test (FST). Fecal CORT level was detected by ELISA. Bisulfite amplicon sequencing PCR was used to assess methylation levels of DRD2 promoter.

Results: CUS and MD/CUS rats had a significantly shorter total distance, longer immobility time, and higher CORT level, while MD and MD/CUS rats had a significantly lower percentage of central distance, more feces, lower rate of sucrose preference, and lower levels of DRD2 protein and mRNA in the VTA than NOR rats. CUS rats showed a significantly higher DRD2 mRNA and protein levels in the VTA than NOR rats. CUS, MD, and MD/CUS rats showed a significantly higher level of DRD2 promoter methylation than NOR rats. CORT level was significantly correlated with the sucrose preference rate in SPT, the immobility time in FST, the total distance, and the number of fecal pellets in OFT. DRD2 protein level was significantly correlated with the sucrose preference rate and the number of fecal pellets. DRD2 mRNA level was significantly correlated with the percentage of central distance and the number of fecal pellets in OFT. The level of DRD2 promoter methylation was significantly correlated with the sucrose preference rate, immobility time, total distance, the percentage of central distance, and the number of fecal pellets.

Conclusions: Early life MD increased vulnerability to stress-induced depressive-like behavior in adult rats. Enhanced DRD2 promoter methylation in the VTA may increase the susceptibility to depression.

Keywords: maternal deprivation, depression, DRD2, DNA methylation, ventral tegmental area

INTRODUCTION

Depressive disorder is one of the most prevalent psychiatric illnesses. Data from WHO demonstrated that the lifetime prevalence of depression is as high as 18.1% globally (1). The etiology of depression is complex with risk factors that can enhance the vulnerability being a family history of depression (2) and childhood traumatic experiences (3). The childhood or early life period is particularly sensitive to adverse events that increase the risk of depression (3, 4). However, this predisposition, or vulnerability to depression is poorly understood.

Dopamine (DA) is the most abundant monoamine neurotransmitter in the CNS and regulates motivation, emotions, reward circuits, cognition, and reinforcement behaviors (5, 6). It has been widely reported that dopaminergic neurotransmitter is deficient in depressive individuals (7, 8). Mesocorticolimbic DA circuitry, originating from the ventral tegmental area (VTA) neurons, is a key part of the brain's reward circuitry and plays an important role in mediating stress response. The VTA DA neurons have been suggested to be highly susceptible to stress and are involved in the pathophysiology of stress-induced depression (9, 10). The dopamine D2 receptor (DRD2) is one of the major DA receptor subtypes. In the VTA dopaminergic neurons, DRD2 expression in pre-synaptic (autoreceptors) can inhibit the excitability of dopaminergic neurons by regulating firing rate and plays a negative feedback regulation on dopamine neurons (11). Previous studies show that DRD2 plays a key role on the development of stress-related depression (11). The study of Chen et al. (12) found that CUS induces depression-like behaviors and upregulates the expression of DRD2 in the prefrontal cortex of rats. Depression-like behaviors were observed more pronounced in DRD2 knockout (DRD2^{-/-}) mice than wild-type mice following chronic stress (13). Our previous study also revealed that early life maternal deprivation (MD) induces aberrant expression of DRD2 gene in the mesocorticolimbic DA system of adult rats, especially elevates DRD2 gene expression in the striatum, and reduces DRD2 gene expression in the VTA (14–17). However, the molecular mechanisms regulating DRD2 gene expression have yet to be fully understood.

DNA methylation has been demonstrated to be a stable epigenetic imprint after stress and can consistently repress gene expression. DNA methylation in mammals currently occurs mainly on cytosines in the CpG islands of the gene promoter region. Hypermethylation in the promoter region can spatially prevent DNA from binding to transcription factor complexes and repress gene expression, while demethylation restores it (18). DNA methylation can be affected by early life stress with a long-lasting change, which can alter disease susceptibility later in life, including depression (19–21). For example, Tozzi et al. (22) found that childhood maltreatment can induce demethylation of the FK506 binding protein 5 (FKBP5, a regulator of the glucocorticoid receptor) may be associated with clinical symptoms of depression disorder. McGowan et al.'s (23) study in the postmortem of suicide victims found hypermethylation in the neuron-specific glucocorticoid receptor promoter in hippocampus of victims with early life adversity compared with that without early life adversity. It has been

shown that methylation of the dopamine D2 receptor (DRD2) gene promoter is elevated in women who were abused during childhood (24). However, it should be further clarified that early life stress may enhance vulnerability to depression through changing DNA methylation of DRD2 gene in adulthood.

In this study, maternal deprivation (MD) and chronic unpredictable stress (CUS) were used, as early life stress and adulthood stress paradigms, respectively, to establish animal models of depression. The level of DNA methylation and expression of DRD2 gene in the VTA were investigated.

MATERIALS AND METHODS

Animals and Design

Pregnant Sprague–Dawley (SD) rats (SLAC Laboratory Animal Inc., Shanghai, China) were housed in the Experimental Animal Center of the Second Xiangya Hospital, Central South University and checked daily for delivery. The date of birth was designated as postnatal day 0 (PND 0). On PND 1, litters were randomly divided into four groups: MD, CUS, MD plus CUS (MD/CUS), and normal control (NOR). Rats in MD and MD/CUS group were exposed to MD stress from PND 1 to PND 14. Rats in CUS and MD/CUS group were exposed to CUS for 28 days at the age of 10 weeks; the NOR rats received no MD and CUS. Only male rats were included and there were 10 rats in each group. Forty male rats were weaned on PND 21 and housed in separate cages (two rats/cage). The behaviors of all male rats were assessed at 14 weeks old and 24-h fecal corticosterone (CORT) was collected before behavioral tests. All male rats were decapitated within 24 h after the end of the behavioral tests and brain tissues were taken for biological experiments. The animal protocol was approved by the Animal Ethics Committee of the Second Xiangya Hospital, Central South University. During the experiment, the rats can drink and eat freely with a light and dark cycle of 12 h/12 h (lights on 8:00–20:00), room temperature of 21–23°C, and humidity of 50–55%.

Maternal Deprivation (MD)

The MD paradigm was performed according to a previous publication (25). The litters in MD and MD/CUS groups were moved from the dams to a single standard polycarbonate box (26 × 20 × 14 cm), paved the bottom of the box with wood chips at 9:00, and then moved back the dams at 15:00 from PND 1 to PND 14 for MD. Rats were weaned at PND 21 and housed in separate cages for every two rats until the 10th week.

Chronic Unpredictable Stress (CUS)

The CUS paradigm was established following a published protocol with a small modification (26). The protocol included seven stressors: crowding (5–6 rats/cage, 2 h), wet bedding (15 h), elevated open platform (10 × 10 cm, 160 cm in height, 2 h), restraint (2 h), water deprivation (16 h), food deprivation (16 h), and blank stimulus. CUS and MD/CUS rats received a stressor every day from 10 to 14 weeks old. The same stressor cannot occur for two consecutive days so that the rats could not predict the occurrence of the stressor.

Open Field Test (OFT)

The OFT was used to assess locomotor activity and anxiety-like behavior. All rats were tested and sequentially placed in the very center of a box (50 × 83 × 56 cm). A video camera was used to record activities of rats in the box for 5 min. The total distance (index for locomotor activity), vertical counts (index for exploration), the percentage of central distance, and the number of fecal pellets (both indexes for anxiety level) were automatically recorded with a computerized tracking system (Ethovision 1.50; Noldus IT, Wageningen, Netherlands). The percentage of central distance represented the percentage of central distance to the total distance during the test. The box was fully cleaned with ethanol (75%) after each trial.

Sucrose Preference Test (SPT)

The SPT was used to assess anhedonia of rodents and performed by a published protocol (26). The protocol included three procedures: (1) rats received two bottles of 1.5% sucrose solution for 24 h; (2) rats received one bottle of 1.5% sucrose solution and one bottle of plain water for the next 24 h; (3) rats were deprived of water and food for the next 18 h. Two pre-weighed bottles of solution (respectively 1.5% sucrose solution and plain water) were then provided to rats for 1 h. The position of two bottles were swapped. All rats were housed in a single cage throughout the test period and the weight of consumed solution of each rat (both 1.5% sucrose solution and plain water) was recorded. The sucrose preference rate was calculated (sucrose preference rate = consumption of sucrose solution/total consumption solution × 100%).

Forced Swimming Test (FST)

The FST was used to assess behavioral despair. The test was carried out in a quiet environment. On the first day, rats were individually placed in a forced swimming bucket (a transparent tempered glass cylinder with a diameter of 21 cm and a height of 46 cm with a water depth of 30 cm, 25°C) and then taken out after 15 min of acclimatization, dried with a hair dryer, and placed back in the cage. On the second day at the same time, rats were placed into the forced swimming bucket again. All rats were tested and the time of immobility of each rat (the rat was passively floating with only the tail, slightly swinging with the paws to maintain body balance, and exposing the head to the surface of the water) was recorded within 5 min as an indicator. The swimming bucket was flushed, and water was changed after each trial to avoid affecting the behavior of the next rat.

Enzyme-Linked Immunosorbent Assay (ELISA)

ELISA was used to detect the level of fecal corticosterone (CORT). The rats were housed singly, and feces of each rat were simultaneously collected for 24 h at the end of the 14th week (27–29). Feces fell freely through the bars of the wire cage and could be directly collected using the pan below each cage. The collected 24-h feces were homogenized in glacial acetic acid (2%). After being dried, ground, mixed, and large particles removed, feces were mixed with 80% methanol by high-speed vortex, followed by centrifugation at 2,500 × g for 10 min. The supernatant was

mixed with EIA buffer as the test sample. The concentration of CORT in feces was measured using a Corticosterone EIA Kit (Cayman, Germany) following the manual instructions.

Western Blot

The whole VTA tissue was dissected with coronal section rodent brain matrices (1 mm) according to the rat brain in stereotaxic coordinates and homogenized in ice-cold buffer (30). Western blot was performed as previously described. Anti-DRD2 antibody was purchased from Santa Cruz Biotechnology (San Diego, CA, USA). The horseradish peroxidase-conjugated anti-rabbit IgG antibody and anti-β-actin antibody were purchased from Sigma-Aldrich (St. Louis, MI, USA).

Real-Time Reverse Transcription Quantitative PCR (RT-PCR)

RT-PCR was used to detect the DRD2 mRNA expression in the VTA. After the behavioral experiments, the rats were decapitated and the VTA tissues were stripped off quickly and transferred to liquid nitrogen. Total RNA was isolated from the dissected VTA tissues using Trizol reagent (Invitrogen, Carlsbad, CA, USA). RT-PCR was conducted as previously described (15). The primers for DRD2 were 5'-CTTGATAGTCAGCCTTGCTGTG-3' and 5'-AGGGCACGTAGAATGAGACAAT-3'. The primers for β-actin were 5'-CACGATGGAGGGGCCGACTCATC-3' and 5'-TAAAGACCTCTATGCCAACACAGT-3'. The $\Delta\Delta C_t$ method was used to calculate the results, and $2^{-\Delta\Delta C_t}$ represented the expression level of DRD2 mRNA. β-Actin mRNAs were used as an internal control.

DNA Methylation Analysis

Genomic DNA was isolated from the VTA tissues by proteinase K/phenol-chloroform. DNA was quantified and then diluted to a working concentration of 10 ng/μl with TE buffer (Tris-EDTA). CpG islands locating in the promoter of DRD2 were selected for measurement according to the following criteria: (1) 200 bp minimum length; (2) 50% or higher GC content; (3) 0.60 or higher ratio of observed/expected dinucleotides CpG. One region from CpG islands (including 24 CpG sites) of DRD2 was selected and sequenced. Bisulfite amplicon sequencing PCR was used for quantitative methylation analysis by following a previously described protocol (31).

Statistical Analysis

Data were presented as mean ± SEM and analyzed using the statistical software SPSS 21.0. One-way ANOVA and the least significant difference (LSD) method were used to compare differences among groups. Pearson correlation analysis was used to analyze the correlations between biomarkers and behavioral indexes. A $p < 0.05$ was considered as significant.

RESULTS

Effect of MD on the Behaviors of Adulthood Rats

OFT showed significant differences in the total distance ($F_{(3,36)} = 11.67$, $p < 0.001$), the percentage of central distance

($F_{(3,36)} = 7.32, p = 0.001$), and the number of fecal pellets ($F_{(3,36)} = 6.59, p = 0.001$) among four groups. Specifically, CUS and MD/CUS rats had shorter total distance than NOR rats (post $p < 0.05$, **Figure 1A**). MD and MD/CUS rats showed a lower percentage of central distance (post $p < 0.05$, **Figure 1B**) and more feces (post $p < 0.05$, **Figure 1C**) than NOR rats. No significant difference in vertical counts was observed between four groups ($F_{(3,36)} = 0.742, p = 0.534$; **Figure 1D**).

SPT showed a significant difference in the sucrose preference rate among four groups ($F_{(3,36)} = 11.51, p < 0.001$). Specifically, MD and MD/CUS rats had significantly lower rate of sucrose preference than rats in the NOR group (post $p < 0.05$), whereas no significant difference in the sucrose preference rate was observed between CUS rats and NOR rats (post $p > 0.05$, **Figure 1E**).

FST showed significant differences in the immobility time among groups ($F_{(3,36)} = 9.07, p < 0.001$). Specifically, CUS and MD/CUS rats showed significantly longer immobility time than NOR rats (post $p < 0.05$), whereas there was no significant difference in the immobility time between MD rats and NOR rats (post $p > 0.05$, **Figure 1F**).

Effect of MD on Fecal CORT Concentration in Adult Rats

Significant difference in fecal CORT concentration was observed among four groups ($F_{(3,36)} = 22.81, p < 0.001$). Specifically, CUS and MD/CUS group showed a significantly higher CORT level in feces than NOR rats (post $p < 0.05$), whereas no significant difference in the fecal CORT concentration was observed between MD and NOR rats (post $p > 0.05$, **Figure 2A**).

Effect of MD on the Level of DRD2 Protein, MRNA, and Methylation

Significant differences in DRD2 protein level in the VTA were observed among four groups ($F_{(3,36)} = 26.96, p < 0.001$). Specifically, a significantly lower DRD2 protein level was observed in the VTA of MD and MD/CUS rats than NOR rats (post $p < 0.05$), whereas CUS rats showed significantly higher level of DRD2 protein in the VTA than NOR rats (post $p < 0.05$, **Figure 2B**).

The expression of DRD2 mRNA in the VTA was significantly different among four groups ($F_{(3,36)} = 16.09, p < 0.001$). Specifically, MD and MD/CUS rats showed significantly lower expression of DRD2 mRNA than NOR rats (post $p < 0.05$), whereas CUS rats showed significantly higher expression of DRD2 mRNA than NOR rats (post $p < 0.05$, **Figure 2C**).

Significant differences in the level of DRD2 promoter methylation in the VTA were observed among groups ($F_{(3,36)} = 151.834, p < 0.001$). Specifically, CUS, MD, and MD/CUS rats had a significantly higher level of DRD2 promoter methylation in the VTA than NOR rats (post $p < 0.05$). Moreover, MD/CUS rats had a significantly higher DRD2 promoter methylation level than MD and CUS rats (post $p < 0.05$, **Figure 2D**).

The Correlations Between CORT, DRD2 Protein, DRD2 MRNA, DRD2 Methylation Levels, and Behavioral Indexes

CORT level in feces was significantly correlated with the total distance ($r = -0.490, p < 0.01$, **Figure 3A**), the number of fecal pellets ($r = 0.367, p < 0.05$, **Figure 3C**), the sucrose preference rate ($r = -0.551, p < 0.001$, **Figure 3D**), and immobility time ($r = 0.513, p < 0.01$, **Figure 3E**). While there was no significant correlation with CORT level and the percentage of central distance ($r = -0.188, p = 0.245$, **Figure 3B**).

DRD2 protein level in VTA showed a significant correlation with the number of fecal pellets ($r = -0.529, p < 0.001$, **Figure 4C**) and the sucrose preference rate ($r = 0.354, p < 0.05$, **Figure 4D**). But the level of DRD2 protein were not significantly correlated with the total distance ($r = -0.122, p = 0.454$, **Figure 4A**), the percentage of central distance ($r = 0.308, p = 0.053$, **Figure 4B**), and the immobility time ($r = 0.116, p = 0.475$, **Figure 4E**). DRD2 mRNA level in VTA was significantly correlated with the percentage of central distance ($r = 0.345, p < 0.05$, **Figure 5B**) and the number of fecal pellets ($r = -0.410, p < 0.01$, **Figure 5C**). While DRD2 mRNA level were not significantly correlated with the total distance ($r = -0.241, p = 0.134$, **Figure 5A**), the sucrose preference rate ($r = 0.287, p = 0.073$, **Figure 5D**), and immobility time ($r = -0.003, p = 0.984$, **Figure 5E**).

The level of DRD2 promoter methylation in VTA was significantly correlated with the total distance ($r = -0.562, p < 0.001$, **Figure 6A**), the percentage of central distance ($r = -0.473, p < 0.01$, **Figure 6B**), the number of fecal pellets ($r = 0.438, p < 0.01$, **Figure 6C**), the sucrose preference rate ($r = -0.657, p < 0.001$, **Figure 6D**), and immobility time ($r = 0.498, p < 0.01$, **Figure 6E**).

DISCUSSION

Maternal deprivation and chronic unpredictable stress paradigm are classical model for mimicking early life stress and stress experienced during adulthood, respectively. These stressors have been demonstrated to induce depression-like and anxiety-like behaviors in rodents (32–34). Our study revealed that maternal deprivation independently induces anxiety-like behavior (decreased percentage of central distance and increased number of fecal pellets in OFT), whereas chronic unpredictable stress alone induces despair-like behavior (increased immobility time in FST) and reduces locomotor activity (decreased total distance in OFT) in rats. It is worth noting that early life maternal deprivation plus chronic unpredictable stress can induce anhedonia-like behavior in adulthood rats, in addition to anxiety-like and despair-like behaviors (35). Previous studies demonstrated that individuals with high anxiety traits and anhedonia are more sensitive to stress-induced depression (36, 37). These findings suggest that early life maternal deprivation is generally regarded as a predictor of psychopathological vulnerability to depression in adulthood, but not a direct inducer of depression (38).

The dynamics of the hypothalamic–pituitary–adrenal (HPA) axis require balance of excitation and inhibition to adequately

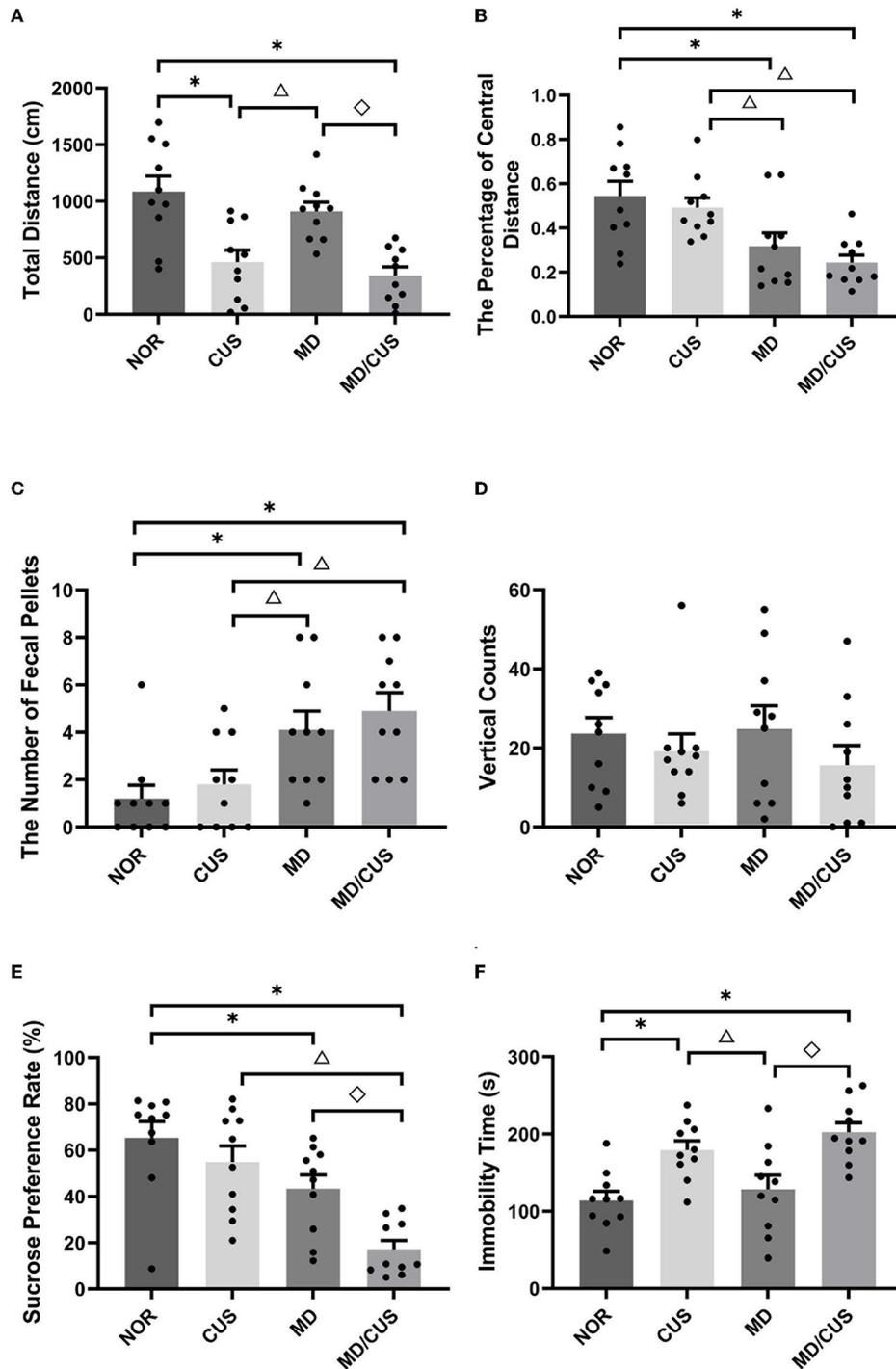


FIGURE 1 | Effect of maternal deprivation on rats' behaviors in adulthood. **(A)** Total distance in the open field test. **(B)** The percentage of central distance in the open field test (the percentage of central distance = central distance/total distance). **(C)** The number of fecal pellets in the open field test. **(D)** Vertical counts in the open field test. **(E)** Sucrose preference rate in the sucrose preference test. **(F)** Immobility time in the forced swimming test. NOR: normal control ($n = 10$); CUS: chronic unpredictable stress ($n = 10$); MD: maternal deprivation ($n = 10$); MD/CUS: maternal deprivation plus chronic unpredictable stress ($n = 10$). * $p < 0.05$ compared with NOR group. ^ $p < 0.05$ compared with CUS group. ◇ $p < 0.05$ compared with MD group.

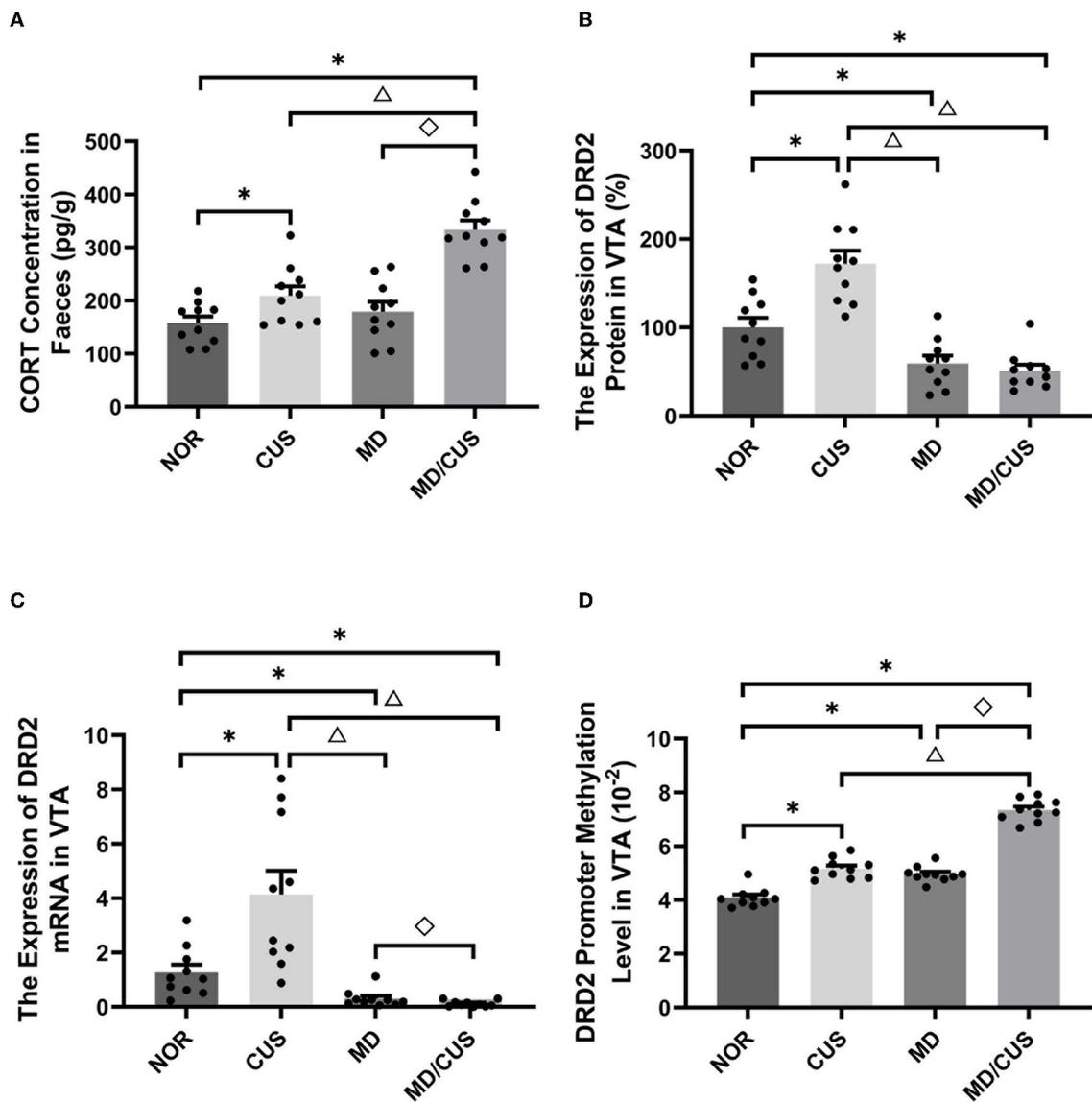


FIGURE 2 | Effect of maternal deprivation on biomarker of rats in adulthood. **(A)** CORT concentration in feces. **(B)** The DRD2 protein level in the ventral tegmental area (VTA) tissues. **(C)** The DRD2 mRNA level in the VTA tissues. **(D)** The DRD2 promoter methylation level in the VTA tissues. NOR: normal control ($n = 10$); CUS: chronic unpredictable stress ($n = 10$); MD: maternal deprivation ($n = 10$); MD/CUS: maternal deprivation plus chronic unpredictable stress ($n = 10$). * $p < 0.05$ compared with NOR group. $^{\Delta} p < 0.05$ compared with CUS group. $^{\diamond} p < 0.05$ compared with MD group.

respond to stress and adapt to environmental demand (39). CORT has been identified as a biological marker of HPA axis activity, which relates to chronic stress and psychopathological vulnerability. Compared with traditional plasma samples, the quantification of CORT in feces offers the advantage of long-term monitoring of stress responses as a non-invasive method (40). In this study, chronic unpredictable stress and maternal deprivation plus chronic unpredictable stress both increased CORT level in feces. CORT level in feces was significantly correlated with the sucrose preference rate, immobility time, the total distance, and the number of fecal pellets. These findings are consistent with a previous study showing elevation of basal CORT levels

and susceptibility to depression or anxiety after chronic stress (41, 42).

The dopaminergic system fully matures until young adulthood after development in the embryonic period (43). This prolonged development provides an extensive time window for the influence of early adverse events on vulnerability to psychopathology (43, 44). This study focused on the changes of DRD2 expression in the VTA in stress-induced depression and found that maternal deprivation reduced the expression of DRD2 gene. The reduction was more pronounced in maternal deprivation rats re-exposed to chronic unpredictable stress in adulthood. However, chronic unpredictable stress enhanced

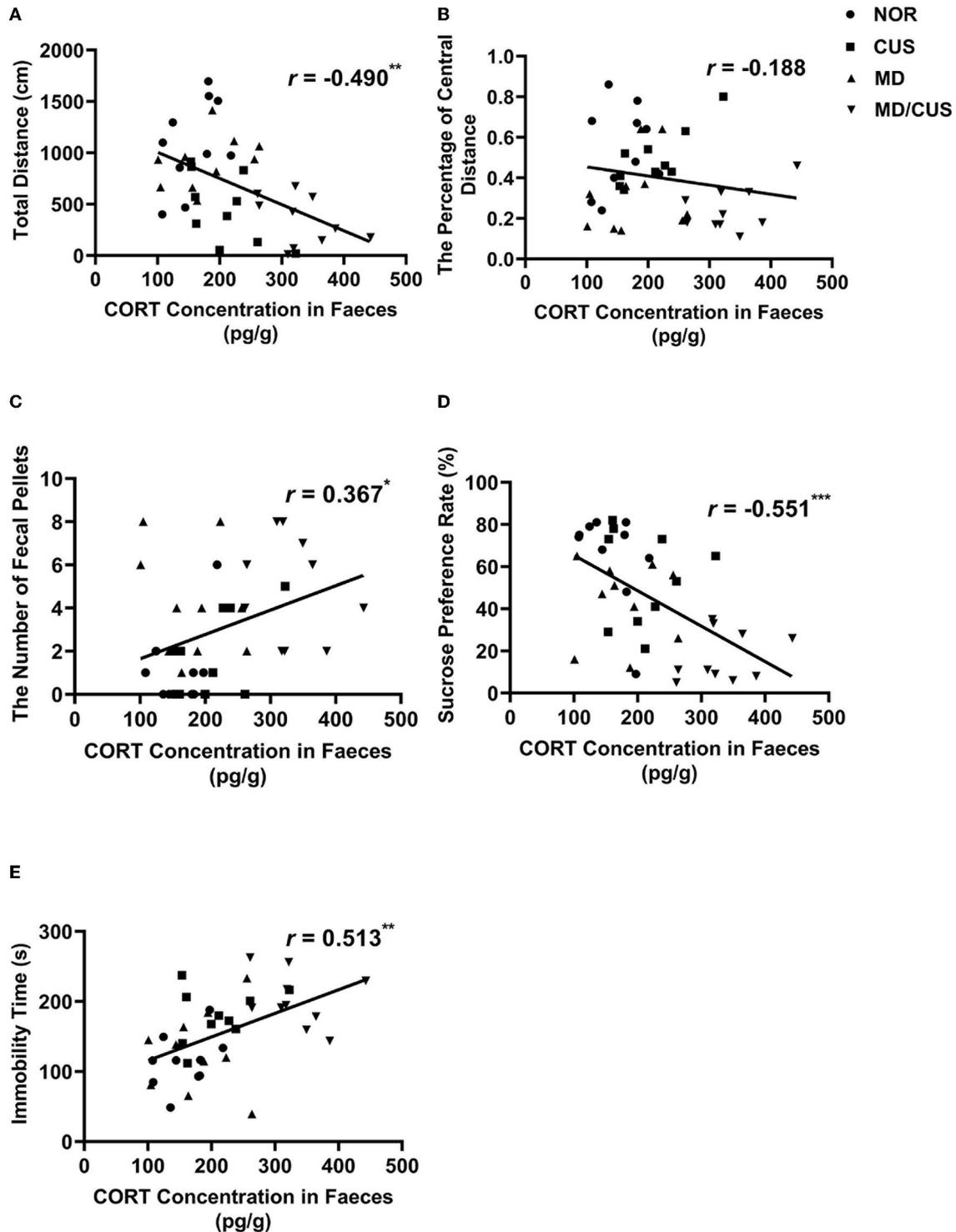


FIGURE 3 | The correlations between CORT level in feces and behavioral indexes. **(A)** The correlation between CORT level in feces and the total distance in the open field test. **(B)** The correlation between CORT level in feces and the percentage of central distance in the open field test. **(C)** The correlation between CORT level in feces and the number of fecal pellets in the open field test. **(D)** The correlation between CORT level in feces and sucrose preference rate in the sucrose preference test. **(E)** The correlation between CORT level in feces and immobility time in the forced swimming test. Correlations were performed in all animals ($N = 40$). $^{*}p < 0.05$. $^{**}p < 0.01$. $^{***}p < 0.001$.

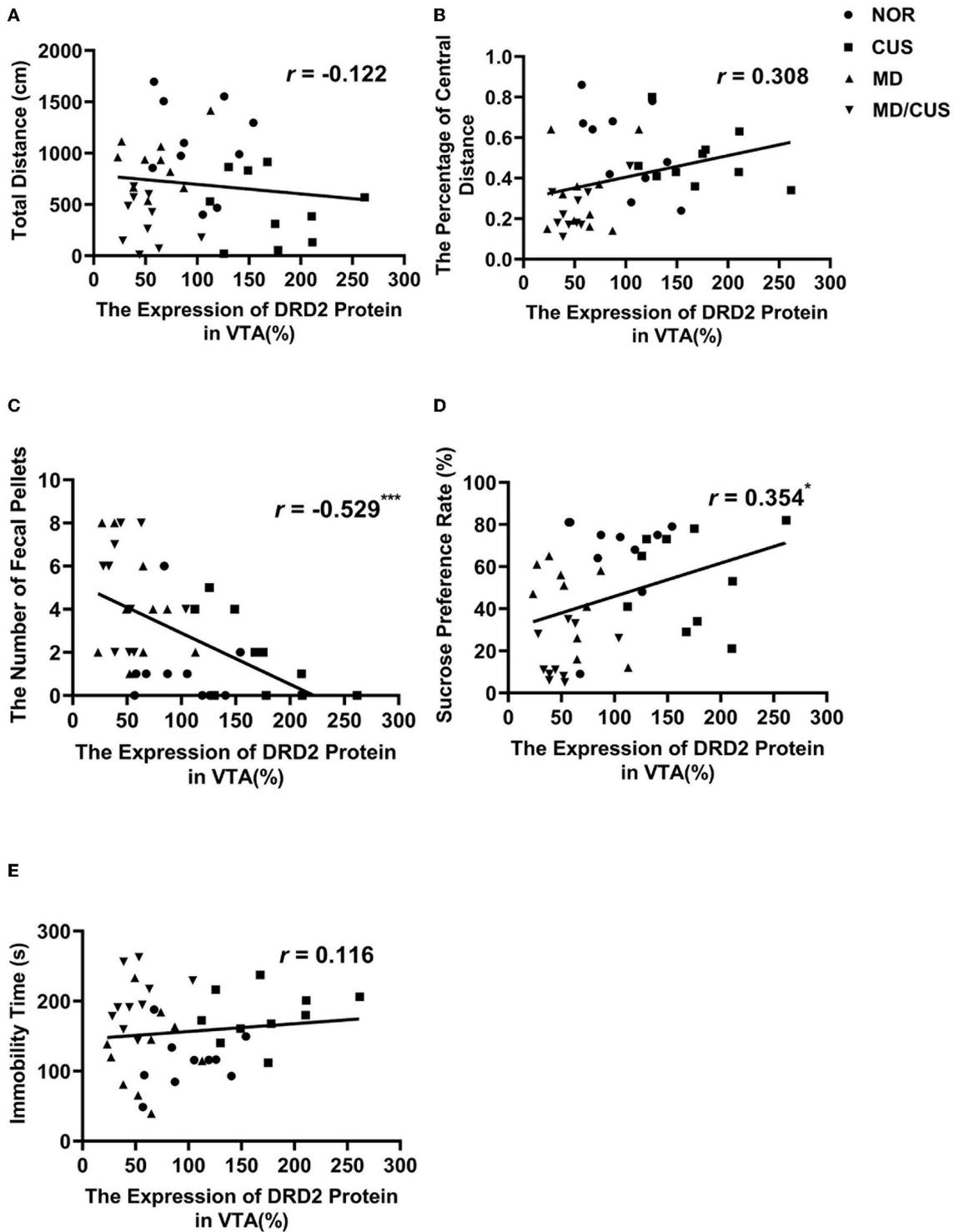


FIGURE 4 | The correlations between DRD2 protein level in the VTA and behavioral indexes. **(A)** The correlation between DRD2 protein level in the VTA and the total distance in the open field test. **(B)** The correlation between DRD2 protein level in the VTA and the percentage of central distance in the open field test. **(C)** The correlation between DRD2 protein level in the VTA and the number of fecal pellets in the open field test. **(D)** The correlation between DRD2 protein level in the VTA and sucrose preference rate in the sucrose preference test. **(E)** The correlation between DRD2 protein level in the VTA and immobility time in the forced swimming test. Correlations were performed in all animals ($N = 40$). * $p < 0.05$. ** $p < 0.01$. *** $p < 0.001$.

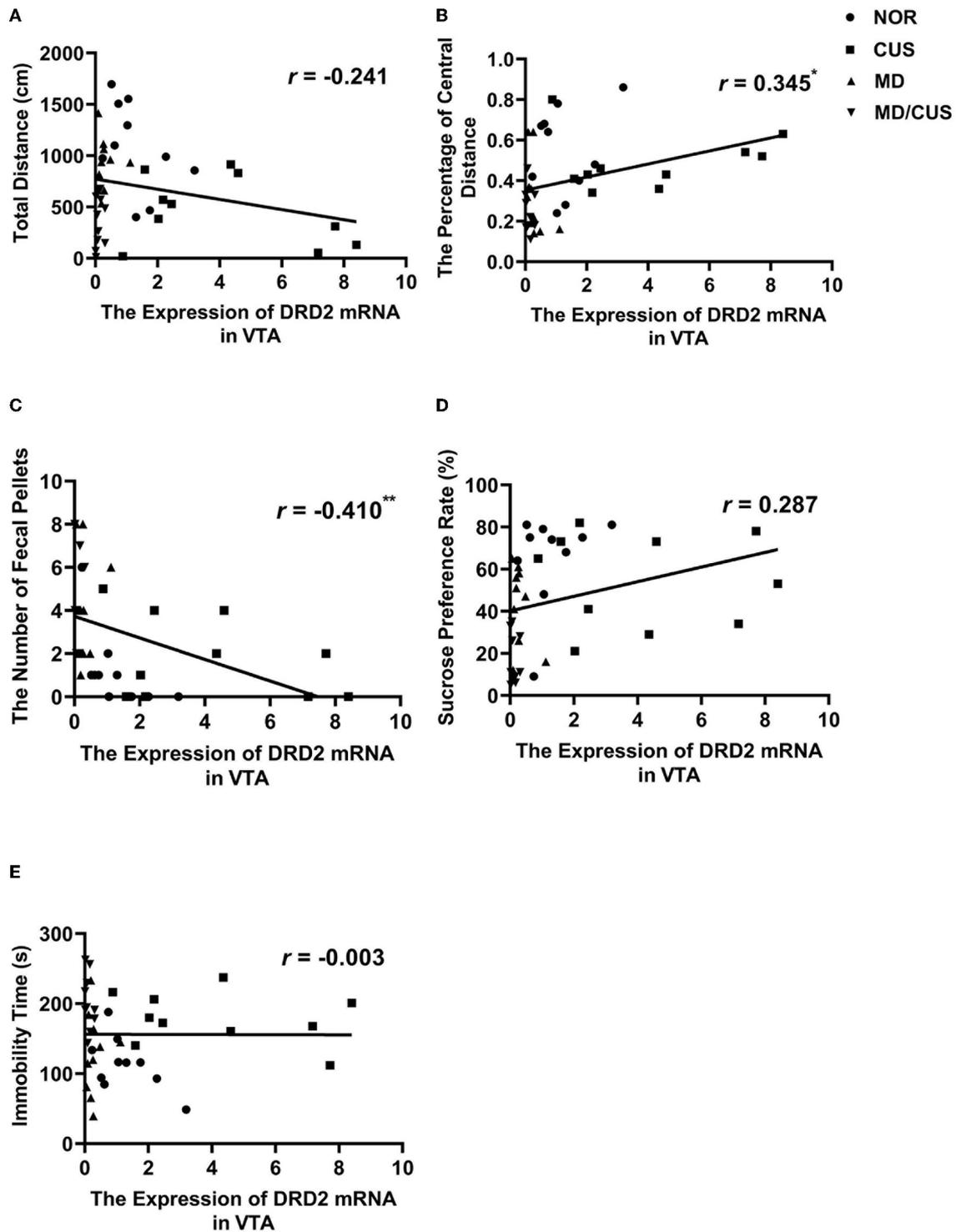


FIGURE 5 | The correlations between DRD2 mRNA level in the VTA and behavioral indexes. **(A)** The correlation between DRD2 mRNA level in the VTA and the total distance in the open field test. **(B)** The correlation between DRD2 mRNA level in the VTA and the percentage of central distance in the open field test. **(C)** The correlation between DRD2 mRNA level in the VTA and the number of fecal pellets in the open field test. **(D)** The correlation between DRD2 mRNA level in the VTA and sucrose preference rate in the sucrose preference test. **(E)** The correlation between DRD2 mRNA level in the VTA and immobility time in the forced swimming test. Correlations were performed in all animals ($N = 40$). $^*p < 0.05$. $^{**}p < 0.01$. $^{***}p < 0.001$.

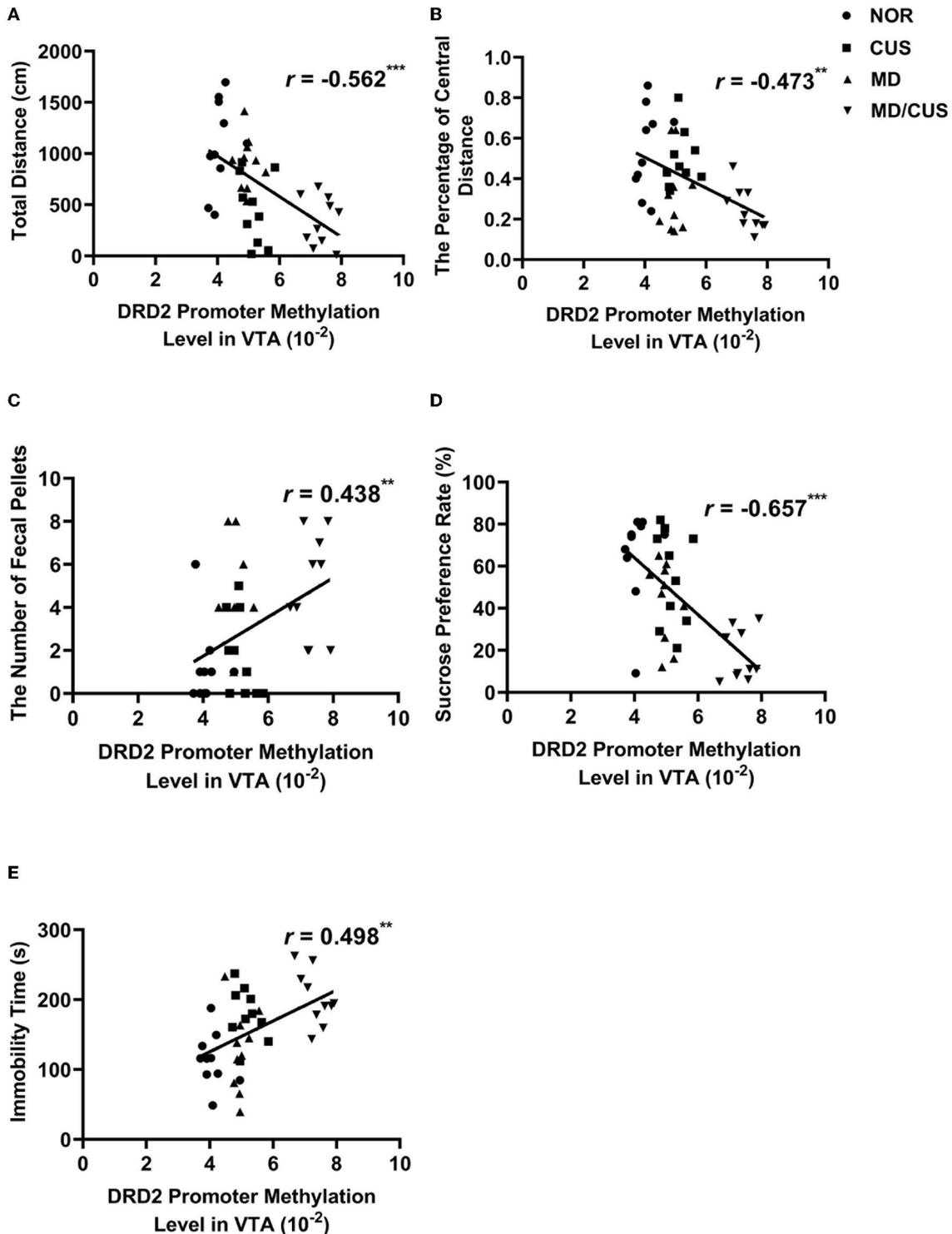


FIGURE 6 | The correlations between DRD2 promoter methylation level in VTA and behavioral indexes. **(A)** The correlation between DRD2 promoter methylation level in VTA and the total distance in the open field test. **(B)** The correlation between DRD2 promoter methylation level in VTA and the percentage of central distance in the open field test. **(C)** The correlation between DRD2 promoter methylation level in VTA and the number of fecal pellets in the open field test. **(D)** The correlation between DRD2 promoter methylation level in VTA and sucrose preference rate in the sucrose preference test. **(E)** The correlation between DRD2 promoter methylation level in VTA and immobility time in the forced swimming test. Correlations were performed included all animals ($N = 40$). $*p < 0.05$. $**p < 0.01$. $***p < 0.001$.

the expression of DRD2 gene in the VTA in adult rats without MD. It can also be seen that the correlation trends between DRD2 expression and behavioral changes induced by chronic unpredictable stress are different from other stresses, which could explain why the overall correlations between DRD2 expression and some behavioral indicators were not significant. Nevertheless, the influence of abnormal DRD2 expression cannot be ignored. In the VTA, DRD2 is a type of autoreceptor and expresses on the somatodendrites of dopaminergic neurons, which can inhibit the excitability of dopaminergic neurons by modulating the firing rate. The firing rate of dopaminergic neurons in the VTA was observed to increase in rodents that experienced chronic social defeated stress and chronic restraint stress (45–47), and decrease in chronic mild stress-induced rodents (48, 49), which are associated with increased vulnerability to depression (50). Authement et al.'s (51) study using a single 24-h episode of MD revealed that MD shifts spike timing-dependent plasticity toward long-term depression at GABAergic synapses onto VTA DA neurons and proposed that MD can promote DA dysregulation of VTA DA neurons through an epigenetic impairment of synaptic plasticity.

Previous studies proposed that epigenetic control of gene expression may be a bridge between early life stress and neurobehavioral outcomes in adulthood (52–54). For example, Godfrey et al.'s study suggested that epigenetic fine-tuning of the expression of genes can enhance the risk for depression in later life (55). This study showed that early life maternal deprivation increased the methylation of DRD2 promoter in the VTA. Indeed, the effect of maternal deprivation was more pronounced when rats were re-exposed to chronic unpredictable stress in adulthood. Furthermore, the level of DRD2 promoter methylation in the VTA was significantly correlated with the total distance, the percentage of central distance, the number of fecal pellets, the sucrose preference rate, and immobility time. Previous studies have demonstrated that the effect of maternal deprivation on DNA methylation of genes is associated with the susceptibility to depression (31, 56, 57). Collectively, long-term changes in the methylation of DRD2 promoter and its expression in the VTA induced by maternal deprivation may be related to the increased risk of developing depression later in life.

It should be known that many non-dopaminergic neurons exist in the VTA and D2 receptors also expressed on these non-dopaminergic cells (58, 59). For example, postsynaptic D2 receptors on the VTA GABAergic neurons can provide local inhibitory inputs onto dopaminergic neurons (60, 61) and knockdown of D2 receptors in GABAergic neurons can increase inhibitory tone (62). A previous study demonstrated that neuronal subpopulations are enriched in the VTA and other subpopulations of neurons may also express the DRD2 gene (63). Therefore, the expression of DRD2 in different types of neurons requires further investigations in the future.

We acknowledge that this study has several limitations. First, DRD2 expression on different subpopulations of neurons was not identified. In the future study, the cell-type specificity of DRD2 expression could be addressed using either immunohistochemistry or RNAscope staining. Second, what DNMT or DNMTs are responsible for the DRD2 gene methylation in the VTA is not identified. Thus, it is unknown

whether DNMTs mediated DNA methylation in the different conditions. Previous studies demonstrated that quetiapine could reverse depressive-like behavior and reduce methylation induced by maternal deprivation through increasing DNMT activities in the hippocampus and NAc (64). Treatment of unstressed control rats with DNMT inhibitors recapitulated the effect of chronic unpredictable stress on decreased AMPAR expression and function in PFC. In contrast, overexpression of Dnmt3a in PFC of stressed animals prevented the loss of glutamatergic responses. Chronic stress-induced behavioral abnormalities could be partially attenuated by Dnmt3a expression in PFC (65). Third, this study only used male rats. Both sexes should be used to address whether there is a difference between males and females although most previous publications only used male animals. Fourth, in this study, only CpG islands locating in the promoter of DRD2 were selected for measurement. Although DNA methylation of the gene promoter is negatively correlated with gene expression, the differentially methylated CpG sites in the gene are positively and negatively correlated with gene expression, suggesting a bidirectional functional output on the enhancer and repressor regions of gene (66, 67). Data of this study are not able to answer why CUS and MD produce exhibited different effects on DRD2 expression. Whether differentially methylated CpG sites of the DRD2 gene, or other genes, and even other epigenetic mechanisms could account for the difference between CUS and MD should be addressed in future studies.

In conclusion, early life maternal deprivation increases vulnerability to stress-induced depressive-like behavior in adulthood. Enhanced DRD2 promoter methylation and subsequent downregulation of DRD2 expression in the ventral tegmental area may be involved in the increase in susceptibility to depression.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary materials, further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The animal study was reviewed and approved by the Animal Ethics Committee of the Second Xiangya Hospital, Central South University.

AUTHOR CONTRIBUTIONS

ZG, SL, and JW: performed experiments, data analysis, and prepared article. ZG, YZ, and XZ: study design, article revision, and final approval. SL and YZ: response to reviewers and article revision. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by National Natural Science Foundation of China (Nos. 81701333 and 81671341).

REFERENCES

- Fedko IO, Hottenga JJ, Helmer Q, Mbarek H, Huider F, Amin N, et al. Measurement and genetic architecture of lifetime depression in the Netherlands as assessed by LIDAS (Lifetime Depression Assessment Self-report). *Psychol Med.* (2021) 51:1345–54. doi: 10.1017/s0033291720000100
- Sullivan PF, Neale MC, Kendler KS. Genetic epidemiology of major depression: Review and meta-analysis. *Am J Psychiatry.* (2000) 157:1552–62. doi: 10.1176/appi.ajp.157.10.1552
- Nemeroff CB. Paradise lost: the neurobiological and clinical consequences of child abuse and neglect. *Neuron.* (2016) 89:892–909. doi: 10.1016/j.neuron.2016.01.019
- Heim C, Shugart M, Craighead WE, Nemeroff CB. Neurobiological and psychiatric consequences of child abuse and neglect. *Dev Psychobiol.* (2010) 52:671–90. doi: 10.1002/dev.20494
- Wise RA. Dopamine, learning and motivation. *Nat Rev Neurosci.* (2004) 5:483–94. doi: 10.1038/nrn1406
- Perkins KA, Lerman C, Grottenhaler A, Ciccocioppo MM, Milanak M, Conklin CA, et al. Dopamine and opioid gene variants are associated with increased smoking reward and reinforcement owing to negative mood. *Behav Pharmacol.* (2008) 19:641–9. doi: 10.1097/FBP.0b013e32830c367c
- Dunlop BW, Nemeroff CB. The role of dopamine in the pathophysiology of depression. *Arch Gen Psychiatry.* (2007) 64:327–37. doi: 10.1001/archpsyc.64.3.327
- Li ZC, He Y, Tang JS, Zong XF, Hu ML, Chen XG. Molecular imaging of striatal dopamine transporters in major depression—A meta-analysis. *J Affect Disord.* (2015) 174:137–43. doi: 10.1016/j.jad.2014.11.045
- Baik J-H. Dopamine signaling in reward-related behaviors. *Front Neural Circuits.* (2013) 7:152. doi: 10.3389/fncir.2013.00152
- Holly EN, Miczek KA. Ventral tegmental area dopamine revisited: effects of acute and repeated stress. *Psychopharmacology.* (2016) 233:163–86. doi: 10.1007/s00213-015-4151-3
- Zurawek D, Faron-Gorecka A, Kusmider M, Kolasa M, Gruca P, Papp M, et al. Mesolimbic dopamine D-2 receptor plasticity contributes to stress resilience in rats subjected to chronic mild stress. *Psychopharmacology.* (2013) 227:583–93. doi: 10.1007/s00213-013-2990-3
- Chen C, Yang JM, Hu TT, Xu TJ, Xu WP, Wei W. Elevated dopamine D2 receptor in prefrontal cortex of CUMS rats is associated with downregulated cAMP-independent signaling pathway. *Can J Physiol Pharmacol.* (2013) 91:750–8. doi: 10.1139/cjpp-2012-0399
- Sim H, Choi TY, Lee HJ, Kang EY, Yoon S, Han PL, et al. Role of dopamine D2 receptors in plasticity of stress-induced addictive behaviours. *Nat Commun.* (2013) 4. doi: 10.1038/ncomms2598
- Bai M, Zhu XZ, Zhang L, Zhang Y, Xue L, Wang YT, et al. Divergent anomaly in mesocorticolimbic dopaminergic circuits might be associated with different depressive behaviors, an animal study. *Brain Behav.* (2017) 7. doi: 10.1002/brb3.808
- Zhang Y, Wang YT, Wang L, Bai M, Zhang XW, Zhu XZ. Dopamine receptor D2 and associated microRNAs are involved in stress susceptibility and resistance to escitalopram treatment. *Int J Neuropsychopharmacol.* (2015) 18. doi: 10.1093/ijnp/pyv025
- Zhu XZ, Peng SF, Zhang S, Zhang XW. Stress-induced depressive behaviors are correlated with Par-4 and DRD2 expression in rat striatum. *Behav Brain Res.* (2011) 223:329–35. doi: 10.1016/j.bbr.2011.04.052
- Zhu XZ, Li T, Peng SF, Ma XL, Chen XG, Zhang XW. Maternal deprivation-caused behavioral abnormalities in adult rats relate to a non-methylation-regulated D2 receptor levels in the nucleus accumbens. *Behav Brain Res.* (2010) 209:281–8. doi: 10.1016/j.bbr.2010.02.005
- Lisoway AJ, Zai CC, Tiwari AK, Kennedy JL. DNA methylation and clinical response to antidepressant medication in major depressive disorder: A review and recommendations. *Neurosci Lett.* (2018) 669:14–23. doi: 10.1016/j.neulet.2016.12.071
- Provençal N, Binder EB. The effects of early life stress on the epigenome: From the womb to adulthood and even before. *Exp Neurol.* (2015) 268:10–20. doi: 10.1016/j.expneurol.2014.09.001
- Park C, Rosenblat JD, Brietzke E, Pan ZH, Lee Y, Cao B, et al. Stress, epigenetics and depression: A systematic review. *Neurosci Biobehav Rev.* (2019) 102:139–52. doi: 10.1016/j.neubiorev.2019.04.010
- Wheater ENW, Stoye DQ, Cox SR, Wardlaw JM, Drake AJ, Bastin ME, et al. DNA methylation and brain structure and function across the life course: A systematic review. *Neurosci Biobehav Rev.* (2020) 113:133–56. doi: 10.1016/j.neubiorev.2020.03.007
- Tozzi L, Farrell C, Booij L, Doolin K, Nemoda Z, Szyf M, et al. Epigenetic changes of FKBP5 as a link connecting genetic and environmental risk factors with structural and functional brain changes in major depression. *Neuropsychopharmacology.* (2018) 43:1138–45. doi: 10.1038/npp.2017.290
- McGowan PO, Sasaki A, D'Alessio AC, Dymov S, Labonte B, Szyf M, et al. Epigenetic regulation of the glucocorticoid receptor in human brain associates with childhood abuse. *Nat Neurosci.* (2009) 12:342–8. doi: 10.1038/nn.2270
- Groleau P, Joobar R, Israel M, Zeramdini N, DeGuzman R, Steiger H. Methylation of the dopamine D2 receptor (DRD2) gene promoter in women with a bulimia-spectrum disorder: Associations with borderline personality disorder and exposure to childhood abuse. *J Psychiatr Res.* (2014) 48:121–7. doi: 10.1016/j.jpsychires.2013.10.003
- Ploj K, Roman E, Nylander I. Long-term effects of maternal separation on ethanol intake and brain opioid and dopamine receptors in male wistar rats. *Neuroscience.* [2003] 121:787–99. doi: 10.1016/s0306-4522(03)00499-8
- Zhang Y, Wang YT, Lei H, Wang L, Xue L, Wang X, et al. Optimized animal model to mimic the reality of stress-induced depression in the clinic. *BMC Psychiatry.* (2017) 17. doi: 10.1186/s12888-017-1335-x
- Dantzer B, McAdam AG, Palme R, Fletcher QE, Boutin S, Humphries MM, et al. Fecal cortisol metabolite levels in free-ranging North American red squirrels: Assay validation and the effects of reproductive condition. *Gen Comp Endocrinol.* (2010) 167:279–86. doi: 10.1016/j.ygcen.2010.03.024
- Touma C, Sachser N, Mostl E, Palme R. Effects of sex and time of day on metabolism and excretion of corticosterone in urine and feces of mice. *Gen Comp Endocrinol.* (2003) 130:267–78. doi: 10.1016/s0016-6480[02]00620-2
- Kroll T, Kornadt-Beck N, Oskamp A, Elmenhorst D, Touma C, Palme R, et al. Additional assessment of fecal corticosterone metabolites improves visual rating in the evaluation of stress responses of laboratory rats. *Animals (Basel).* (2021) 11. doi: 10.3390/ani11030710
- De Deurwaerdere P, Ramos M, Bharatiya R, Pugnier E, Chagraoui A, Manem J, et al. Lorcaserin bidirectionally regulates dopaminergic function site-dependently and disrupts dopamine brain area correlations in rats. *Neuropharmacology.* (2020) 166. doi: 10.1016/j.neuropharm.2019.107915
- Zhang Y, Wang L, Wang X, Wang YT, Li CT, Zhu XZ. Alterations of DNA methylation at GDNF gene promoter in the ventral tegmental area of adult depression-like rats induced by maternal deprivation. *Front Psychiatry.* (2019) 9. doi: 10.3389/fpsy.2018.00732
- Kuniishi H, Ichisaka S, Yamamoto M, Ikubo N, Matsuda S, Futora E, et al. Early deprivation increases high-leaning behavior, a novel anxiety-like behavior, in the open field test in rats. *Neurosci Res.* (2017) 123:27–35. doi: 10.1016/j.neures.2017.04.012
- Antoniuk S, Bijata M, Ponimaskin E, Włodarczyk J. Chronic unpredictable mild stress for modeling depression in rodents: Meta-analysis of model reliability. *Neurosci Biobehav Rev.* (2019) 99:101–16. doi: 10.1016/j.neubiorev.2018.12.002
- Wang D, Levine JLS, Avila-Quintero V, Bloch M, Kaffman A. Systematic review and meta-analysis: effects of maternal separation on anxiety-like behavior in rodents. *Transl Psychiatry.* (2020) 10. doi: 10.1038/s41398-020-0856-0
- Shen CK, Cao KR, Cui SN, Cui YF, Mo HX, Wen WH, et al. SiNiSan ameliorates depression-like behavior in rats by enhancing synaptic plasticity via the CaSR-PKC-ERK signaling pathway. *Biomed Pharmacother.* (2020) 124. doi: 10.1016/j.biopha.2019.109787
- Neumann ID, Wigger A, Kromer S, Frank E, Landgraf R, Bosch OJ. Differential effects of periodic maternal separation on adult stress coping in a rat model of extremes in trait anxiety. *Neuroscience.* (2005) 132:867–77. doi: 10.1016/j.neuroscience.2005.01.034
- Chasson GS, Bello MS, Luxon AM, Graham TAA, Leventhal AM. Transdiagnostic emotional vulnerabilities linking obsessive-compulsive and depressive symptoms in a community-based sample of adolescents. *Depress Anxiety.* (2017) 34:761–9. doi: 10.1002/da.22669
- Friedman MJ, Resick PA, Bryant RA, Strain J, Horowitz M, Spiegel D, et al. DSM-5. *Depress Anxiety.* (2011) 28:737–49. doi: 10.1002/da.20845

39. Herman JP, McKlveen JM, Ghosal S, Kopp B, Wulsin A, Makinson R, et al. Regulation of the hypothalamic-pituitary-adrenocortical stress response. *Compr Physiol.* (2016) 6:603–21. doi: 10.1002/cphy.c150015
40. Palme R. Non-invasive measurement of glucocorticoids: Advances and problems. *Physiol Behav.* (2019) 199:229–43. doi: 10.1016/j.physbeh.2018.11.021
41. Thakur AK, Soni UK, Rai G, Chatterjee SS, Kumar V. Protective effects of andrographis paniculata extract and pure andrographolide against chronic stress-triggered pathologies in rats. *Cell Mol Neurobiol.* (2014) 34:1111–21. doi: 10.1007/s10571-014-0086-1
42. Sakhri FZ, Adachi N, Zerizer S, Ohashi Y, Ikemoto H, Tsukada M, et al. Behavioral and neurological improvement by *Cydonia oblonga* fruit extract in chronic immobilization stress rats. *Phytother Res.* (2021) 35:2074–84. doi: 10.1002/ptr.6953
43. Money KM, Stanwood GD. Developmental origins of brain disorders: roles for dopamine. *Front Cell Neurosci.* (2013) 7. doi: 10.3389/fncel.2013.00260
44. Gatzke-Kopp LM. The canary in the coalmine: The sensitivity of mesolimbic dopamine to environmental adversity during development. *Neurosci Biobehav Rev.* (2011) 35:794–803. doi: 10.1016/j.neubiorev.2010.09.013
45. Krishnan V, Han MH, Graham DL, Berton O, Renthal W, Russo SJ, et al. Molecular adaptations underlying susceptibility and resistance to social defeat in brain reward regions. *Cell.* (2007) 131:391–404. doi: 10.1016/j.cell.2007.09.018
46. Anstrom KK, Woodward DJ. Restraint increases dopaminergic burst firing in awake rats. *Neuropsychopharmacology.* (2005) 30:1832–40. doi: 10.1038/sj.npp.1300730
47. Cao JL, Covington HE, Friedman AK, Wilkinson MB, Walsh JJ, Cooper DC, et al. Mesolimbic dopamine neurons in the brain reward circuit mediate susceptibility to social defeat and antidepressant action. *J Neurosci.* (2010) 30:16453–8. doi: 10.1523/jneurosci.3177-10.2010
48. Tye KM, Mirzabekov JJ, Warden MR, Ferenczi EA, Tsai HC, Finkelstein J, et al. Dopamine neurons modulate neural encoding and expression of depression-related behaviour. *Nature.* (2013) 493:537. doi: 10.1038/nature11740
49. Chang CH, Grace AA. Amygdala-ventral pallidum pathway decreases dopamine activity after chronic mild stress in rats. *Biol Psychiatry.* (2014) 76:223–30. doi: 10.1016/j.biopsych.2013.09.020
50. Chaudhury D, Walsh JJ, Friedman AK, Juarez B, Ku SM, Koo JW, et al. Rapid regulation of depression-related behaviours by control of midbrain dopamine neurons. *Nature.* (2013) 493:532. doi: 10.1038/nature11713
51. Authement ME, Kodangattil JN, Gouty S, Rusnak M, Symes AJ, Cox BM, et al. Histone deacetylase inhibition rescues maternal deprivation-induced gabaergic metaplasticity through restoration of AKAP signaling. *Neuron.* (2015) 86:1240–52. doi: 10.1016/j.neuron.2015.05.024
52. Vaiserman AM, Koliada AK. Early-life adversity and long-term neurobehavioral outcomes: epigenome as a bridge? *Hum Genomics.* (2017) 11. doi: 10.1186/s40246-017-0129-z
53. Torres-Berrio A, Issler O, Parise EM, Nestler EJ. Unraveling the epigenetic landscape of depression: focus on early life stress. *Dialogues Clin Neurosci.* (2019) 21:341–57. doi: 10.31887/DCNS.2019.21.4/enestler
54. Juruena MF, Gadelrab R, Cleare AJ, Young AH. Epigenetics: A missing link between early life stress and depression. *Prog Neuropsychopharmacol Biol Psychiatry.* (2021) 109. doi: 10.1016/j.pnpbp.2020.110231
55. Godfrey KM, Costello PM, Lillycrop KA. The developmental environment, epigenetic biomarkers and long-term health. *J Dev Orig Health Dis.* (2015) 6:399–406. doi: 10.1017/s204017441500121x
56. Jakobsson J, Cordero MI, Bisaz R, Groner AC, Buskamp V, Bensadoun JC, et al. KAP1-mediated epigenetic repression in the forebrain modulates behavioral vulnerability to stress. *Neuron.* (2008) 60:818–31. doi: 10.1016/j.neuron.2008.09.036
57. Uchida S, Hara K, Kobayashi A, Otsuki K, Yamagata H, Hobara T, et al. Epigenetic status of *Gdnf* in the ventral striatum determines susceptibility and adaptation to daily stressful events. *Neuron.* (2011) 69:359–72. doi: 10.1016/j.neuron.2010.12.023
58. Morales M, Margolis EB. Ventral tegmental area: cellular heterogeneity, connectivity and behaviour. *Nat Rev Neurosci.* (2017) 18:73–85. doi: 10.1038/nrn.2016.165
59. Sesack SR, Aoki C, Pickel VM. Ultrastructural localization of D2 receptor-like immunoreactivity in midbrain dopamine neurons and their striatal targets. *J Neurosci.* (1994) 14:88–106.
60. Steffensen SC, Taylor SR, Horton ML, Barber EN, Lyle LT, Stobbs SH, et al. Cocaine disinhibits dopamine neurons in the ventral tegmental area via use-dependent blockade of GABA neuron voltage-sensitive sodium channels. *Eur J Neurosci.* (2008) 28:2028–40. doi: 10.1111/j.1460-9568.2008.06479.x
61. van Zessen R, Phillips JL, Budygin EA, Stuber GD. Activation of VTA GABA neurons disrupts reward consumption. *Neuron.* (2012) 73:1184–94. doi: 10.1016/j.neuron.2012.02.016
62. Martin TA, Smith HR, Luessen DJ, Chen R, Porrino LJ. Functional brain activity is globally elevated by dopamine D2 receptor knockdown in the ventral tegmental area. *Brain Res.* (2020) 1727. doi: 10.1016/j.brainres.2019.146552
63. Kim HJ, Kim M, Kang B, Yun S, Ryeo SE, Hwang D, et al. Systematic analysis of expression signatures of neuronal subpopulations in the VTA. *Molecular Brain.* (2019) 12. doi: 10.1186/s13041-019-0530-8
64. Ignacio ZM, Reus GZ, Abelaira HM, Maciel AL, de Moura AB, Matos D, et al. Quetiapine treatment reverses depressive-like behavior and reduces DNA methyltransferase activity induced by maternal deprivation. *Behav Brain Res.* (2017) 320:225–32. doi: 10.1016/j.bbr.2016.11.044
65. Wei J, Cheng J, Waddell NJ, Wang ZJ, Pang XD, Cao Q, et al. DNA Methyltransferase 3A is involved in the sustained effects of chronic stress on synaptic functions and behaviors. *Cereb Cortex.* (2021) 31:1998–2012. doi: 10.1093/cercor/bhaa337
66. Mehta D, Klengel T, Conneely KN, Smith AK, Altmann A, Pace TW, et al. Childhood maltreatment is associated with distinct genomic and epigenetic profiles in posttraumatic stress disorder. *Proc Natl Acad Sci U S A.* (2013) 110:8302–7. doi: 10.1073/pnas.1217750110
67. Teh AL, Pan H, Chen L, Ong ML, Dogra S, Wong J, et al. The effect of genotype and in utero environment on interindividual variation in neonate DNA methylomes. *Genome Res.* (2014) 24:1064–74. doi: 10.1101/gr.171439.113

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