Check for updates

OPEN ACCESS

EDITED BY Francesco Panza, University of Bari Aldo Moro, Italy

REVIEWED BY Bruno Moukette, Purdue University Indianapolis, United States Ann Van Eeckhaut, Vrije Universiteit Brussel, Belgium Chaomeng Liu, Capital Medical University, China

*CORRESPONDENCE Zuoli Sun Zuolisun83@163.com

[†]These authors have contributed equally to this work

RECEIVED 11 January 2025 ACCEPTED 21 April 2025 PUBLISHED 12 May 2025

CITATION

Liu M, Pan W, He J, Ling S, He Y, Yang J, Mao P and Sun Z (2025) Unveiling chiral amino acid alterations and glycine dysregulation in late-life depression through targeted metabolomics. *Front. Psychiatry* 16:1558796. doi: 10.3389/fpsyt.2025.1558796

COPYRIGHT

© 2025 Liu, Pan, He, Ling, He, Yang, Mao and Sun. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Unveiling chiral amino acid alterations and glycine dysregulation in late-life depression through targeted metabolomics

Mingxia Liu^{1†}, Weigang Pan^{1†}, Jing He¹, Sihai Ling¹, Yi He¹, Jian Yang¹, Peixian Mao¹ and Zuoli Sun^{1,2*}

¹Beijing Key Laboratory of Mental Disorders, National Clinical Research Center for Mental Disorders & National Center for Mental Disorders, Beijing Anding Hospital, Capital Medical University, Beijing, China, ²Laboratory for Clinical Medicine, Capital Medical University, Beijing, China

Background: Late-life depression (LLD) is a major depressive disorder that is highly prevalent among older people, and there are currently no validated biomarkers for the diagnosis and treatment of LLD. Although dysregulated amino acid metabolism has been increasingly implicated in neuropsychiatric disorders, including LLD, most existing studies overlook the chiral nature of amino acids, potentially leading to inaccurate or incomplete findings. To address this gap, this study aimed to precisely characterize the serum chiral amino acid profiles in patients with LLD and identify potential biomarkers.

Methods: Using liquid chromatography tandem mass spectrometry combined with a chiral derivatization technique, the serum levels of 34 amino acids were analyzed in 53 LLD patients and 37 healthy controls (HCs).

Results: Significant alterations in both D- and L-enantiomers were observed, including reduced levels of D-methionine, D-glutamic acid, D-threonine, and L-threonine, alongside elevated glycine levels in LLD compared to HCs. The combination of D-methionine and glycine demonstrated moderate discriminatory power for distinguishing LLD from HCs, with an area under the curve of 0.71. Notably, glycine levels were significantly lower in antidepressant treatment responders than in non-responders. Additionally, D- and L-glutamic acid levels were differentially associated with specific cognitive function indicators.

Discussion: These findings underscore the importance of accounting for amino acid chirality in biomarker research and highlight chiral amino acids as promising candidates for the diagnosis of LLD and the prediction of treatment response.

KEYWORDS

late-life depression, chiral amino acid, biomarker, LC-MS/MS, targeted metabolomics

1 Introduction

Late-life depression (LLD) can be defined as major depressive disorder (MDD) occurring in individuals aged 60 years or older (1). The global prevalence of LLD is estimated at 13.3%, which is notably higher than the incidence of depression in younger populations (2). Despite the high prevalence, LLD remains a complex and multifactorial disease with no definitive biomarkers for clinical diagnosis or effective evaluation of treatment responses. Researchers have suggested that LLD may be linked to alterations in neurotransmitter systems (3), inflammatory pathways (4), neuroimaging (5), molecular genetics (6), and environmental stressors (7), but they are not yet sufficiently conclusive. Consequently, identifying objective markers for LLD has always been a significant challenge in both clinical and research settings.

Among the various pathophysiological mechanisms, amino acid neurotransmitter systems dysfunction is thought to play an important role in depression. Recent studies have reported elevated serum levels of glutamic acid (Glu), aspartic acid (Asp), and glycine (Gly) in patients with MDD, with Glu and phenylalanine (Phe) also correlating with the severity of depressive symptoms (8). In contrast, lower levels of methionine (Met), Phe, tryptophan (Trp), tyrosine (Tyr), and non-enzymatic antioxidants have been associated with drug-naive, first-episode MDD patients (9). Furthermore, branched-chain amino acids have been implicated in depression, with negative associations observed between valine (Val) and leucine (Leu) levels and depressive symptoms in young adults (10). Deficiencies in essential amino acids not only increase the risk of depression but may also impair the efficacy of pharmacological treatments. Consequently, some studies have demonstrated that amino acid supplementation, such as Trp, could potentially improve therapeutic outcomes in depressive patients (11).

Common amino acids, except Gly, generally exhibit a chiral center, which theoretically results in the presence of both L- and D-amino acid stereoisomers. It has long been assumed that the majority of amino acids in higher animals are L-enantiomers, with the D-forms being scarcely present. However, recent advancements in analytical techniques have led to the identification of various D-amino acids, such as D-serine (D-Ser), D-Asp, and D-Glu, in mammals, including humans (12). These D-amino acids have been shown to exhibit distinct biological functions and are increasingly recognized as potential clinical targets for brain disorders (13, 14). Despite the growing body of evidence, many studies have treated amino acids as a collective group, which may overlook significant variations between individual enantiomers and lead to imprecise conclusions. This is particularly relevant in the case of multifactorial and complex LLD, where only a limited number of studies have specifically investigated the role of chiral amino acids in the pathophysiology of the disorder. In 2023, our research team developed a novel, efficient liquid chromatography tandem-mass spectrometry (LC-MS/MS) method based on N^{α} -(5fluoro-2,4-dinitrophenyl)-L-leucinamide derivatization for quantifying chiral amino acids in serum. This method was initially applied to the analysis of serum chiral amino acids in LLD patients, and several potential biomarkers for LLD were identified (15).

The current study analyzed and verified the serum profiles of 17 L-amino acids, 14 D-amino acids, asparagine (Asn), Gly, and γ -aminobutyric acid (GABA) in both LLD patients and healthy controls (HCs). We assessed the differences in amino acid levels between these groups, investigated their changes before and after treatment across different response populations, and examined their correlations with symptom scores in LLD.

2 Materials and methods

2.1 Subject recruitment and sample collection

A total of 53 patients with LLD and 37 HCs were recruited from Beijing Anding Hospital, Capital Medical University, following a protocol approved by the hospital's Human Research and Ethics Committee (2020-Scientific Research-97) (16). Written informed consent was obtained from all participants after a full explanation of the study. Inclusion criteria for LLD patients were as follows: (1) age \geq 60 years with at least 6 years of education; (2) onset of first depressive episode after age 60; (3) diagnosis of MDD based on the Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition (DSM-5); (4) a Hamilton Depression Rating Scale-17 (HAMD-17) score \geq 17; and (5) no use of antidepressant medication at the time of enrollment. Inclusion criteria for HCs included: (1) age ≥ 60 years; (2) no history of psychiatric disorders and normal cognitive function; and (3) no use of psychotropic medications. Exclusion criteria for all participants were: (1) history of manic or hypomanic episodes; (2) comorbid dementia or other psychiatric/medical disorders; (3) severe physical illnesses (e.g., cardiovascular, hepatic, or renal disease); (4) history of brain injury; (5) substance abuse or dependence; and (6) Mini-Mental State Examination (MMSE) score ≤ 20 for those with primary education, or ≤ 24 for those with education at or above middle school level.

Patients diagnosed with LLD were treated for 8 weeks with either escitalopram or sertraline. The initial dose of escitalopram was 5–10 mg/day, with adjustments allowed up to 20 mg/day based on the patient's clinical response and tolerability. For sertraline, the starting dose was 25–50 mg/day, with titration permitted up to 100 mg/day as needed. Depression severity was assessed using the HAMD-17, while cognitive function was evaluated using the Repeatable Battery for the Assessment of Neuropsychological Status (RBANS) at both baseline and after 8 weeks of treatment. Patients who showed at least a 50% reduction in their HAMD-17 scores post-treatment compared with the initial assessment were classified as responders, while others were categorized as nonresponders (17, 18). RBANS included assessments of five cognitive domains: immediate memory, visuospatial, language, attention, and delayed memory.

For both LLD patients and HCs, the blood sampling was done during 8-12 AM after fasting for more than eight hours and

participants were instructed to maintain a light diet in the days preceding sample collection. Blood samples were collected before the start of treatment (baseline) and after 8 weeks. The serum was separated by centrifugation at 1500 x g for 15 minutes and stored at -80°C until further analysis.

2.2 Amino acids analysis

Serum amino acid levels were measured using liquid chromatography-tandem mass spectrometry (HPLC-MS/MS) as previously described (14, 15). In brief, 50 μ L of serum was mixed with 450 μ L of methanol (MeOH) and vortexed for 5 minutes at 1500 rpm. The mixture was then centrifuged at 16,000 x g for 10 minutes, and the supernatant was evaporated to dryness under nitrogen. The dried residues were reconstituted in 50 μ L of water and derivatized by adding 5 μ L of triethylamine, followed by 50 μ L of N^{α}-(5-fluoro-2,4-dinitrophenyl)-L-leucinamide (1% in acetonitrile). After thorough mixing, the reaction mixture was incubated at 55 °C for 40 minutes, and the reaction was terminated by the addition of 5 μ L of formic acid.

The analysis was performed on an XBridge[®] C18 column (130 Å, 3.5 μ m, 2.1 mm × 150 mm; Waters, USA) using an UltiMate 3000 HPLC system (Dionex, Sunnyvale, CA, USA) coupled with an API 4000 MS/MS system (AB Sciex, Concord, Canada). The mobile phase consisted of solvent A (0.1% formic acid in water) and solvent B (0.1% formic acid in acetonitrile), with a flow rate of 0.2 mL/min. The gradient was programmed as follows: 30% B (0–10 min), 30%-60% B (10–34 min), 60% B (34–39 min), 60%-30% B (39–40 min), followed by a 5 minutes equilibration period.

D- and L-amino acids were identified by comparing the retention times and peak area changes before and after the addition of an enantiomeric standard. Quantification was based on the peak area relative to a calibration curve derived from standards. The HPLC-MS/MS method underwent rigorous validation, as detailed in previous studies (15).

2.3 Statistical analysis

Data analysis was conducted using SPSS (version 26.0, SPSS Inc., Chicago, IL, USA). Descriptive statistics were used to summarize the demographic and clinical characteristics of participants. Continuous variables are presented as mean \pm standard deviation (SD), and the normality of their distribution was assessed using the Shapiro-Wilk test. For group comparisons, appropriate statistical tests were selected based on distribution assumptions. For categorical variables such as gender and family psychiatric history, the chi-squared test was used. For amino acid levels that were not normally distributed, data normalization was carried out using logarithmic, quadratic, or reciprocal transformation. For continuous variables that were normally distributed or achieved normality after transformation, differences in amino acid levels between LLD patients and HCs were analyzed using Student's t-test. For non-normally distributed continuous variables, the Mann-Whitney U test was employed to assess differences in amino acid levels between the two groups. To analyze the changes in amino acid levels from baseline to week 8 of treatment in LLD patients, the paired-samples Wilcoxon signed-rank test was used for non-normally distributed data, and the pairedsamples t-test was used for normally distributed data. To evaluate the associations between amino acid concentrations and continuous variables (e.g., age and LLD symptom scores), Pearson's correlation test was used for data following a Gaussian distribution, while Spearman's correlation test was applied for data with a non-Gaussian distribution. To assess the relationship between amino acid levels and diagnosis, logistic regression was performed with diagnosis as the dependent variable and amino acid levels as explanatory variables, using forward stepwise selection. The receiver operating characteristic (ROC) curve and the area under the ROC curve (AUC) were used to evaluate the diagnosis value of amino acids levels in LLD patients. A repeated measures analysis of variance (RMANOVA) was employed to examine the interaction between time and group (time × group) in relation to treatment effects and amino acid levels. RMANOVA considered both time and treatment as factors. Statistical significance was set at p < 0.05 for all analyses.

3 Results

3.1 Demographic and clinical data of LLD patients and HCs

The demographic and clinical characteristics of 53 LLD patients and 37 HCs are summarized in Table 1. No significant differences were found between the LLD and HCs in terms of age (p = 0.076), gender (p = 0.524), or body mass index (BMI) (p = 0.791). However, significant differences were observed regarding education levels and family history of mental illness. LLD patients tended to have fewer years of education and were more likely to have a family history of mental illness compared to the HCs (p < 0.001). All LLD patients had moderate to severe depression, with HAMD-17 scores ranging from 17 to 34. After 8 weeks of antidepressant treatment, there was a significant reduction in the HAMD-17 scores (p < 0.05).

TABLE 1	Comparison	of	demographic	and	clinical	variables	in	LLD	and
HCs grou	ips.								

Variables	LLD	HCs	Р	
Ν	53	37	-	
Age (year)	66.68 ± 4.08	65.16 ± 3.74	0.076 ^a	
Gender (Male/Female)	18/35	15/22	0.524 ^b	
BMI (score)	23.95 ± 3.01	24.14 ± 2.43	0.791 ^a	
Education (years)	10.04 ± 3.11	13.24 ± 3.22	< 0.001 ^a	
Family psychiatric history	43 (81%)	37 (100%)	< 0.001 ^b	
Baseline HAMD-17	21.66 ± 3.64	-	_	
Follow-up HAMD-17	9.24 ± 5.87	-	-	

LLD, late-life depression; HCs, healthy controls; BMI, body mass index; HAMD-17, Hamilton Depression Rating Scale-17. All values are expressed as mean \pm standard deviation. ^a Statistical *p* values derive from the t-test (t). ^b Statistical *p* values derive from the chi-squared test (χ^2).

3.2 Comparison of amino acid levels between LLD patients and HCs

There were significant differences (p < 0.05) between the LLD and HCs in 5/34 amino acids, that was, D-Met, D-Glu, D-threonine (D-Thr), L-Thr, and Gly (Supplementary Table S1 and Figure 1). Specifically, D-Met (log10-transformed: 0.05 ± 0.03 versus $0.06 \pm$ 0.03μ mol/L, p = 0.010), D-Glu (log10-transformed: 0.2228 ± 0.09 versus 0.25 ± 0.07 µmol/L, p = 0.025), D-Thr (log10-transformed: 0.38 ± 0.13 versus 0.43 ± 0.12 µmol/L, p = 0.031), and L-Thr (200.49 ± 50.15 versus 225.84 ± 46.52 µmol/L, p = 0.017) were significantly lower in LLD patients compared to HCs, while Gly levels (435.57 ± 94.09 versus 403.76 ± 128.85 µmol/L, p = 0.013) were significantly higher in LLD patients. However, when applying the Bonferroni correction with a critical p value of 0.00147 (0.05/34), none of the amino acids retained statistical significance.



FIGURE 1

Comparison of amino acid levels between LLD patients and HCs. (A) Volcano plot for differential amino acids. Significantly regulated amino acids between groups determined by p value (p < 0.05). Green dots represent decreased amino acids in LDD, red dots represent increased amino acids in LDD, and gray dots represent non-significant amino acids between groups. (B) Serum levels of D-Met, D-Glu, D-Thr, L-Thr, and Gly in LLD patients and HCs.

3.3 Age-dependent changes in amino acid levels

Age-related variations of the above significantly different amino acids (D-Met, D-Glu, D-Thr, L-Thr, and Gly) were assessed using Pearson or Spearman correlation analysis (Figure 2). In the LLD group, significant age-related decreases were observed in the levels of Gly and L-Thr (r = -0.387, p = 0.004, and r = -0.282, p = 0.041, respectively), whereas no such correlations were found in the HCs group (r = 0.195, p = 0.247, and r = -0.187, p = 0.267, respectively). In contrast to Gly and L-Thr, the levels of D-Met, D-Glu, and D-Thr did not show significant age-related changes in either the LLD or HCs.

3.4 Differentiating LLD patients from HCs using amino acids

To determine the association between amino acids and diagnosis, a forward stepwise logistic regression analysis using D-Met, D-Glu, D-Thr, L-Thr, and Gly as independent variables was performed. The Nagelkerke's \mathbb{R}^2 was 0.159, and the model's coefficient was significant in the omnibus goodness-of-fit test ($\chi 2 = 11.298$, df = 2, p = 0.004). At a significance level of p = 0.05, Gly (p = 0.049) and D-Met (log10-transformed: p = 0.005) were retained in the model and showed a significant association with the LLD diagnosis (Supplementary Table S2).

Following these results, a receiver operating characteristic (ROC) curve analysis was performed to evaluate Gly and D-Met's ability to distinguish LLD patients from HCs. The optimal cut-off values for Gly and D-Met (log10-transformed) were determined to be 390.500 and -1.217, with a sensitivity of 0.755 and 0.514, specificity of 0.649 and 0.755, and area under the curve (AUC) values of 0.655 and 0.649, respectively. Combining Gly and D-Met (log10-transformed) improved the AUC to 0.701 (Supplementary Table S3, Figure 3), indicating a fair discriminatory power.

3.5 Changes in amino acid levels with different treatment outcomes

Significant differences (p < 0.05) were observed in the levels of 19 out of 34 amino acids before and after antidepressant treatment (Supplementary Table S1). After applying the Bonferroni correction with a critical p value of 0.00147 (0.05/34), D-Ser, D-Thr, L-Asp, D-Asp, L-Glu, D-Glu, D-Trp, L-histidine (L-His), and D-His remained significantly altered, with all of these amino acids showing a notable increase following treatment.

To explore the relationship between amino acid levels and treatment effectiveness, LLD patients were divided into responders (n = 36 [73.47%]) and non-responders (n = 13 [26.53%]). The RMANOVA was conducted, with amino acid levels as the dependent variable and time (before and after treatment) as a factor. As shown in Table 2, significant time effects were observed for L-Ser, D-Ser, D-Thr, L-Asp, D-Asp, L-Glu, D-Glu, D-Trp, L-His, D-His, D-arginine (D-Arg), and L-lysine (L-Lys) (p < 0.05), all of which significantly increased after treatment, except for L-Lys. After applying the Bonferroni correction, only L-Asp, D-Asp, and D-His remained significant. Additionally, a significant group effect was found for Gly (p = 0.028), with responders exhibiting significantly lower Gly levels than non-responders. However, no statistically significant interactions between group and time were observed in this study.

3.6 Associations of amino acid levels and LLD symptom scores

To assess the relationship between changes in key amino acid levels and improvements in LLD symptoms as measured by the HAMD-17 and RBANS scales after 8 weeks of antidepressant treatment, Pearson or Spearman correlation analysis was performed. As shown in Figure 4, changes in D-Glu levels were significantly correlated with reductions in the RBANS immediate



FIGURE 2

Age-dependent changes in the serum levels of Gly and L-Thr in LLD and HCs groups. Correlation analysis of age with (A) Gly levels and (B) L-Thr levels in the serum samples of HCs (black) and LLD (red).



memory scores (r = -0.284, p = 0.048), and changes in L-Glu levels were significantly associated with reduced scores for RBANS attention scores (r = -0.288, p = 0.047). No significant correlations were found between other amino acids of interest and LLD symptom scores.

4 Discussion

The primary findings of this study were summarized as follows: 1) Statistical analysis identified significant differences (p < 0.05) in the concentrations of 5 out of 34 amino acids between LDD patients and HCs. Serum levels of D-Met, D-Glu, D-Thr, and L-Thr were notably lower, whereas Gly levels were elevated in the LDD groups compared to HCs (Supplementary Table S1, Figure 1). 2) Gly and L-Thr exhibited significant age-dependent declines in LDD patients (Figure 2). 3) The combination of D-Met and Gly exhibited moderate discriminatory ability to distinguish LDD patients from HCs, with an AUC of 0.701 (Figure 3). 4) Following antidepressant treatment, levels of L-Asp, D-Asp, and D-His significantly increased. Moreover, Gly levels were significantly lower in responders compared to non-responders (Table 2). 5) Significant correlations were observed between reductions in RBANS scores and the levels of D-Glu and L-Glu (Figure 4).

This study revealed a significant elevation in serum Gly levels among LLD patients compared to HCs. These findings aligned with prior research, which also reported increased plasma and serum Gly levels in individuals with depression (8, 19). Additionally, a notable group effect was identified for Gly, with responders displaying significantly lower Gly levels than non-responders. Supporting this, Hebbring et al. conducted a metabolomic analysis of plasma

samples categorized by escitalopram response or remission status, demonstrating a negative association between Gly levels and treatment outcomes (20). Further investigation by Hebbring et al. included genotyping tag single-nucleotide polymorphisms (SNPs) related to genes involved in Gly synthesis and degradation. This analysis identified the rs10975641 SNP in the Gly dehydrogenase gene as significantly associated with treatment outcome phenotypes. Elevated Gly levels may contribute to the development of depression through their role as co-agonists of Nmethyl-D-aspartate (NMDA) receptors (8). NMDA receptors, a subtype of ionotropic glutamate receptors, exhibit high calcium permeability and are primarily activated by Glu and Asp, with Gly acting as a co-agonist (21). Impaired NMDA receptor function is linked to cognitive deficits, while excessive activation can result in excitotoxicity and neuronal damage. This may correspond to our finding that the levels of Gly and its precursor, L-Thr, decrease with age in the LLD group, paralleling the decline in cognitive function. Overactivation of NMDA receptors leads to excessive calcium influx, which disrupts mitochondrial function, promotes oxidative stress through increased reactive oxygen species, and activates proinflammatory pathways, ultimately causing tissue and organ damage (22). Although physiological activation of the NMDA receptor is necessary for cell survival, overactivation is a signal for cell death (23). As a result, dysregulated expression and dysfunction of NMDA receptors have been associated with a range of neuropsychiatric disorders, such as autism spectrum disorders, epilepsy, schizophrenia, and related conditions (24).

Our findings indicated that D-Met levels were significantly reduced in the serum of LLD patients, with notable increases observed after antidepressant treatment. Interestingly, previous studies have detected a significant decrease in Met levels in depression patients (25, 26). Met, an essential sulfur-containing amino acid, has antioxidative activity. Wang et al. demonstrated that the anti-inflammatory mechanism of L-Met involves inhibiting nuclear factor-kB activation and upregulating glutathione Stransferase (27). In LLD patients, the reduction in Met might be linked to the abnormal pro-inflammatory state. Notably, Met is also associated with NMDA receptor regulation. Bilen et al. found that Met promotes stress resilience and alleviates social avoidance behavior by decreasing cortical NMDA receptor expression and activity through an epigenetic mechanism involving histone methylation (28). Furthermore, it is worth highlighting that L-Met is the immediate precursor of S-adenosyl-methionine (SAMe), a crucial methyl donor involved in DNA methylation and brain monoamine synthesis, both of which are key processes in mood regulation (29). SAMe has been shown to have significant antidepressant effects, and lower concentrations of SAMe have been reported in the cerebrospinal fluid (CSF) of severely depressed individuals. SAMe administration leads to increased CSF SAMe levels and improvements in depressive symptoms (30). The recent study by Sarkisova et al. revealed that L-Met treatment produced effects comparable to those of established

Variable	Responders ± 1	(n=36) Mean SD ^a	Non-responde ± S	P ^b			
	Baseline	Week 8	Baseline	Week 8	Time	Group	Interaction
L-Ser	268.93 ± 54.86	308.07 ± 78.04	286.28 ± 58.33	307.75 ± 57.69	0.043	0.462	0.601
D-Ser	3.21 ± 0.89	3.99 ± 1.39	2.87 ± 0.75	3.31 ± 0.69	0.003	0.096	0.688
L-Ala	600.72 ± 131.66	663.54 ± 186.54	696.04 ± 168.44	705.82 ± 206.59	0.376	0.121	0.451
D-Ala	2.55 ± 0.94	3.00 ± 1.22	2.64 ± 1.29	2.85 ± 1.06	0.090	0.948	0.841
L-Pro	321.79 ± 87.33	348.32 ± 138.43	354.65 ± 132.54	340.06 ± 111.49	0.831	0.752	0.572
D-Pro	0.84 ± 0.28	0.87 ± 0.37	0.80 ± 0.32	0.98 ± 0.45	0.198	0.896	0.284
L-Val	424.68 ± 104.66	448.08 ± 150.34	454.74 ± 143.33	455.46 ± 132.84	0.706	0.609	0.763
D-Val	0.39 ± 0.15	0.43 ± 0.12	0.37 ± 0.12	0.42 ± 0.12	0.081	0.685	0.932
L-Thr	196.62 ± 52.22	218.08 ± 70.26	205.45 ± 45.94	222.99 ± 56.07	0.134	0.634	0.879
D-Thr	0.39 ± 0.14	0.47 ± 0.15	0.37 ± 0.10	0.45 ± 0.15	0.004	0.577	0.747
L-Ile	143.13 ± 52.96	138.83 ± 69.25	154.29 ± 60.21	128.03 ± 53.84	0.144	0.957	0.462
D-Ile	0.09 ± 0.07	0.10 ± 0.05	0.08 ± 0.05	0.08 ± 0.04	0.815	0.305	0.325
L-Leu	213.37 ± 72.91	221.43 ± 96.41	244.98 ± 94.88	223.01 ± 81.83	0.674	0.466	0.366
D-Leu	0.20 ± 0.08	0.23 ± 0.05	0.21 ± 0.07	0.20 ± 0.06	0.293	0.551	0.149
L-Asp	27.21 ± 10.14	36.81 ± 14.92	31.36 ± 17.49	44.69 ± 17.53	<0.001	0.184	0.462
D-Asp	0.37 ± 0.12	0.46 ± 0.14	0.38 ± 0.14	0.51 ± 0.17	<0.001	0.596	0.456
L-Glu	96.87 ± 35.60	126.68 ± 56.20	121.18 ± 39.97	147.49 ± 59.38	0.011	0.052	0.683
D-Glu	0.21 ± 0.09	0.26 ± 0.09	0.24 ± 0.09	0.31 ± 0.12	0.004	0.176	0.893
L-Trp	59.51 ± 24.92	59.66 ± 34.57	62.52 ± 29.05	56.62 ± 33.71	0.636	0.999	0.618
D-Trp	0.05 ± 0.02	0.06 ± 0.02	0.05 ± 0.02	0.05 ± 0.02	0.012	0.537	0.321
L-Met	38.77 ± 15.12	41.65 ± 18.09	44.88 ± 16.15	41.23 ± 16.92	0.829	0.469	0.405
D-Met	0.05 ± 0.03	0.06 ± 0.03	0.05 ± 0.02	0.05 ± 0.02	0.105	0.53	0.121
L-His	109.43 ± 20.02	133.56 ± 33.75	119.41 ± 28.22	133.13 ± 24.42	0.002	0.402	0.467
D-His	0.12 ± 0.05	0.16 ± 0.04	0.13 ± 0.05	0.15 ± 0.04	<0.001	0.955	0.369
L-Phe	117.16 ± 41.52	120.97 ± 64.59	135.22 ± 55.99	132.55 ± 77.81	0.964	0.318	0.796
D-Phe	0.13 ± 0.05	0.15 ± 0.05	0.13 ± 0.06	0.14 ± 0.04	0.194	0.890	0.629
L-Arg	123.40 ± 34.97	131.73 ± 40.46	126.77 ± 39.80	125.56 ± 35.51	0.598	0.867	0.595
D-Arg	0.30 ± 0.12	0.33 ± 0.10	0.29 ± 0.10	0.33 ± 0.08	0.015	0.974	0.93
L-Tyr	49.62 ± 27.27	56.55 ± 27.41	57.37 ± 27.24	78.66 ± 27.41	0.854	0.306	0.901
L-Lys	55.00 ± 32.25	45.90 ± 31.41	61.40 ± 30.70	42.61 ± 28.92	0.007	0.861	0.330
L-Gln	998.97 ± 181.06	1195.99 ± 267.57	1078.13 ± 252.83	1078.27 ± 221.94	0.077	0.707	0.087
Asn	98.26 ± 22.70	110.24 ± 27.64	105.76 ± 21.84	109.94 ± 27.24	0.119	0.530	0.375
Gly	433.03 ± 102.69	436.57 ± 77.57	475.40 ± 122.12	469.02 ± 99.70	0.086	0.028	0.769
GABA	0.65 ± 0.40	0.77 ± 0.54	0.65 ± 0.35	0.72 ± 0.41	0.054	0.863	0.544

TABLE 2 Results of repeated measure analysis of variance (RMANOVA) with amino acid levels as the dependent variable across time.

LLD, late-life depression; Ser, serine; Ala, alanine; Pro, proline; Val, valine; Thr, threonine; Ile, isoleucine; Leu, leucine; Asp, aspartic acid; Glu, glutamic acid; Trp, tryptophan; Met, methionine; His, histidine; Phe, phenylalanine; Arg, arginine; Tyr, tyrosine; Lys, lysine; Gln, glutamine; Asn, asparagine; Gly, glycine; GABA, γ -aminobutyric acid. ^a Values are expressed as mean ± standard deviation (SD). The concentration unit of all amino acids is presented as μ mol/L. ^b Significant *p* values are in bold type.



antidepressants like imipramine and fluoxetine in WAG/Rij rats (31). However, research on the biomechanism and therapeutic potential of D-Met in depression remains underexplored. Given the diverse roles of Met isomers in various biological processes-ranging from the growth depression of broilers (32) to enzymatic activities in human erythrocytes (33)-further investigation into the specific biological functions of D-Met in the context of depression is warranted.

In addition to Gly and Met, Glu has also exhibited significant changes with depression. Notably, a substantial decrease in D-Glu levels was observed in patients with LLD, while both L- and D-1Glu levels showed significant increases following treatment. Furthermore, these changes in Glu concentrations were significantly correlated with cognitive function. These findings are consistent with a recent study by Bian et al., which reported a significant reduction in serum Glu levels in depression patients. This result was further corroborated in chronic mild unpredictable stress (CUMS) rat models (34). L-Glu has garnered increased attention due to its crucial role in the citric acid cycle (35) and its function as a neurotransmitter in the vertebrate nervous system, which controls cognitive processes in the brain (36). In contrast, the physiological role of D-Glu remains less understood. While D-Glu has been identified in bacterial cell walls (37), rat tissues (38), and human blood (14), its functions are not well-characterized. Recent research has shown that both D-Glu and L-Glu can induce transporter-mediated presynaptic autoinhibition of neurotransmitter release (39). Additionally, D-Glu has been explored as a potential modulator of hormonal secretion (40). Our study found that D-Glu and L-Glu were differentially associated with specific cognitive function indicators on the RBANS, with D-Glu correlating with immediate memory scores and L-Glu with attention scores. This novel finding underscores the potential role of chiral amino acid metabolism in cognitive dysfunction in LLD and highlights the importance of examining amino acids from the perspective of their chiral enantiomers. Although direct evidence linking RBANS scores to glutamic acid remains limited, prior studies support the involvement of glutamatergic signaling in cognitive processes. Glutamatergic and GABAergic systems contribute to memory formation and learning processes via long-term potentiation (LTP). In vitro studies have demonstrated that manipulating GABAergic and glutamatergic transmission through receptor-specific ligands affects LTP. For instance, blocking glutamatergic transmission reduces LTP and impairs learning, whereas blocking GABAergic transmission facilitates LTP and learning (41). Animal studies have shown that forebrain concentrations of glutamate and glutamine were found to increase after passive avoidance learning, suggesting glutamate/ glutamine synthesis during the learning process (42). Furthermore, glutamate levels positively correlate with learning success, while GABA levels negatively correlate with learning performance in the anterior cingulate cortex during reward-based learning paradigms (43). Recent research also indicates that disruptions in both GABAergic and glutamatergic systems impair memory and relearning ability (41, 44). These findings are consistent with our observation that Glu levels are decreased in LLD, and the change in Glu concentration before and after treatment is negatively correlated with the reduction rate of RBANS scores. Together, these results suggest that glutamatergic signaling may play a critical role in the cognitive dysfunction associated with LLD.

Thr levels were significantly reduced in the serum of patients with LLD, with a notable increase in D-Thr observed following antidepressant treatment. Reductions in Thr levels have also been reported in the serum of patients with treatment-resistant depression (TRD) (45), as well as in the medial prefrontal cortex, plasma, and urine of animal models of depression and TRD (46– 48). However, these findings are not entirely consistent, as increased Thr levels have been observed in the serum of patients with MDD and postpartum depression (49, 50). Thr is an essential amino acid crucial for the nervous system, capable of crossing the blood-brain barrier and being converted into Gly (50). Collectively, these studies suggest that Thr may play an important role in the pathophysiology and treatment of depression, warranting further investigation.

In addition to the aforementioned changes, significant differences were also found in amino acid levels before and after antidepressant treatment. Specifically, significant differences were observed in the levels of 19 out of 34 amino acids, and D-Ser, D-Thr, L-Asp, D-Asp, L-Glu, D-Glu, D-Trp, L-His, and D-His remained significantly altered after applying the Bonferroni correction. Notably, all of these amino acids showed a marked increase following treatment, which is consistent with our previous findings (51). These results suggest that antidepressant therapyspecifically treatment with Selective Serotonin Reuptake Inhibitors (SSRIs) such as escitalopram and sertraline-may have measurable regulatory effects on amino acid metabolism. Several of the significantly altered amino acids, such as Glu, Asp, and D-Ser, are well known to participate in neurotransmission and neuroplasticity, potentially linking their changes to therapeutic response (52, 53). It is worth noting that the baseline levels of these significantly increased amino acids were generally lower in LLD patients compared to HCs, and these findings imply that SSRI treatment may ameliorate the dysregulation of serum metabolites in LLD by restoring or enhancing amino acid metabolism. This highlights the potential of these amino acids as biomarkers reflecting neurochemical changes associated with clinical improvement.

Several limitations of this study should be noted. First, the sample size for each group, particularly the responder and nonresponder groups, was relatively small. Second, the participants were only assessed at two time points-baseline and after 8 weeks of treatment-limiting the ability to explore the dynamics of group differences over time. Third, while several amino acid biomarkers, particularly D-amino acids, were identified, the existing reports on these biomarkers are limited, and their mechanisms remain poorly understood. Therefore, further validation through additional methodologies, such as animal studies, is required. Future research should aim to recruit larger sample sizes, incorporate more frequent follow-up visits, and evaluate the clinical usefulness of these biomarkers.

5 Conclusions

In conclusion, this pilot study demonstrates reduced serum levels of D-Met, D-Glu, D-Thr, and L-Thr, along with an elevated level of Gly in patients with LLD. These findings suggest that the combination of D-Met and Gly may serve as a potential diagnostic indicator for LLD, while Gly levels could be used as a biomarker for evaluating treatment efficacy in this population.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

Ethics statement

The studies involving humans were approved by the Ethics Committee of Beijing Anding Hospital, Capital Medical University (2020-Scientific Research-97). The trial was registered in the Chinese Clinical Trial Registry (ChiCTR2100042370). The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

ML: Conceptualization, Funding acquisition, Methodology, Visualization, Writing – original draft, Writing – review & editing. WP: Data curation, Resources, Software, Writing – original draft, Writing – review & editing. JH: Data curation, Methodology, Writing – original draft. SL: Investigation, Validation, Writing – review & editing. YH: Conceptualization, Formal Analysis, Writing – review & editing. JY: Conceptualization, Funding acquisition, Investigation, Writing – review & editing. PM: Project administration, Resources, Writing – review & editing. ZS: Conceptualization, Funding acquisition, Investigation, Supervision, Validation, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was supported by R&D Program of Beijing Municipal Education Commission (KM202310025007), Beijing Hospitals Authority Innovation Studio of Young Staff Funding Support (202336), and the National Natural Science Foundation of China (82171525), Beijing Hospitals Authority's Ascent Plan (DFL20241901).

Acknowledgments

The authors would like to thank all study participants.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer CL declared a shared affiliation with the authors to the handling editor at the time of review.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpsyt.2025.1558796/full#supplementary-material

References

1. Taylor WD. Clinical practice. Depression in the elderly. N Engl J Med. (2014) 371:1228–36. doi: 10.1056/NEJMcp1402180

2. Shorey S, Ng ED, Wong CHJ. Global prevalence of depression and elevated depressive symptoms among adolescents: A systematic review and meta-analysis. Br J Clin Psychol. (2022) 61:287–305. doi: 10.1111/bjc.12333

3. Frisardi V, Panza F, Farooqui AA. Late-life depression and Alzheimer's disease: The glutamatergic system inside of this mirror relationship. *Brain Res Rev.* (2011) 67:344–55. doi: 10.1016/j.brainresrev.2011.04.003

4. Xia J, Chen M, Shao H, Chen H, Zhu S, Zhao M, et al. Inflammation and late-life depression: Unraveling the complex relationship and potential therapeutic strategies. *Front Biosci.* (2023) 28:356. doi: 10.31083/j.fbl2812356

5. Khundakar AA, Thomas AJ. Cellular morphometry in late-life depression: A review of postmortem studies. *Am J Geriatr Psychiatry*. (2014) 22:122–32. doi: 10.1016/j.jagp.2013.06.003

6. Ryan J, Ancelin ML. Genetic and epigenetic regulation of the serotonin transporter gene in late-life depression. *J Geriatr Psychiatry Neurol.* (2019) 32:175–7. doi: 10.1177/0891988719841725

7. Kamiya Y, Doyle M, Henretta JC, Timonen V. Depressive symptoms among older adults: The impact of early and later life circumstances and marital status. *Aging Ment Health*. (2013) 17:349–57. doi: 10.1080/13607863.2012.747078

8. Ho CSH, Tay GWN, Wee HN, Ching J. The utility of amino acid metabolites in the diagnosis of major depressive disorder and correlations with depression severity. *Int J Mol Sci.* (2023) 24:2232. doi: 10.3390/ijms24032231

9. Islam MR, Ali S, Karmoker JR, Kadir MF, Ahmed MU, Nahar Z, et al. Evaluation of serum amino acids and non-enzymatic antioxidants in drug-naive first-episode major depressive disorder. *BMC Psychiatry*. (2020) 20:333. doi: 10.1186/s12888-020-02738-2

10. Whipp AM, Heinonen-Guzejev M, Pietilainen KH, van Kamp I, Kaprio J. Branched-chain amino acids linked to depression in young adults. *Front Neurosci.* (2022) 16:935858. doi: 10.3389/fnins.2022.935858

11. Haleem D. Nutritional importance of tryptophan for improving treatment in depression and diabetes. *Nutr Rev.* (2022) 81:133–49. doi: 10.3390/genes1030413

12. Lella C, Nestor L, De Bundel D, Vander Heyden Y, Van Eeckhaut A. Targeted chiral metabolomics of d-amino acids: Their emerging role as potential biomarkers in neurological diseases with a focus on their liquid chromatography-mass spectrometry analysis upon chiral derivatization. *Int J Mol Sci.* (2024) 25:12410. doi: 10.3390/ ijms252212410

13. Souza INO, Roychaudhuri R, de Belleroche J, Mothet JP. D-amino acids: New clinical pathways for brain diseases. *Trends Mol Med.* (2023) 29:1014–28. doi: 10.1016/j.molmed.2023.09.001

14. Liu M, Li M, He J, He Y, Yang J, Sun Z, et al. Chiral Amino acid profiling in serum reveals potential biomarkers for Alzheimer's disease. *J Alzheimer's Dis.* (2023) 94:291–301. doi: 10.3233/jad-230142

15. Liu M, He J, Ruan C, Pan W, Mao P, Sun Z, et al. Simultaneous measurement of amino acid enantiomers in the serum of late-life depression patients using convenient LC-MS/MS method with N α -(5-fluoro-2,4-dinitrophenyl)-l-leucinamide derivatization. *J Pharm BioMed Anal.* (2023) 230:115387. doi: 10.1016/j.jpba.2023.115387

16. Pan W, Liu C, Zhu D, Liu Y, Mao P, Ren Y, et al. Prediction of antidepressant efficacy by cognitive function in first-episode late-life depression: A pilot study. *Front Psychiatry.* (2022) 13:916041. doi: 10.3389/fpsyt.2022.916041

17. Davis AK, Barrett FS, May DG, Cosimano MP, Sepeda ND, Johnson MW, et al. Effects of psilocybin-assisted therapy on major depressive disorder. *JAMA Psychiatry*. (2021) 78:481. doi: 10.1001/jamapsychiatry.2020.3285

18. Trivedi MH, Rush AJ, Wisniewski SR, Nierenberg AA, Warden D, Ritz L, et al. Evaluation of outcomes with citalopram for depression using measurement-based care in STARD implications for clinical practice. *Am J Psychiatry*. (2006) 163:28–40. doi: 10.1176/appi.ajp.163.1.28

19. Mitani H, Shirayama Y, Yamada T, Maeda K, Ashby CR Jr., Kawahara R. Correlation between plasma levels of glutamate, alanine and serine with severity of depression. *Prog Neuropsychopharmacol Biol Psychiatry*. (2006) 30:1155–8. doi: 10.1016/j.pnpbp.2006.03.036

20. Ji Y, Hebbring S, Zhu H, Jenkins GD, Biernacka J, Snyder K, et al. Glycine and a glycine dehydrogenase (GLDC) SNP as citalopram/escitalopram response biomarkers in depression: Pharmacometabolomics-informed pharmacogenomics. *Clin Pharmacol Ther.* (2011) 89:97–104. doi: 10.1038/clpt.2010.250

21. Smith Q. Transport of glutamate and other amino acids at the blood-brain barrier. J Nutr. (2000) 130:1016S-22S. doi: 10.1093/jn/130.4.1016S

22. Ma T, Cheng Q, Chen C, Luo Z, Feng D. Excessive activation of NMDA receptors in the pathogenesis of multiple peripheral organs via mitochondrial dysfunction, oxidative stress, and inflammation. *SN Compr Clin Med.* (2020) 2:916041. doi: 10.3389/fpsyt.2022.916041

23. Waxman EA, Lynch DR. N-methyl-D-aspartate receptor subtypes: Multiple roles in excitotoxicity and neurological disease. *Neuroscientist.* (2005) 11:37–49. doi: 10.1177/1073858404269012

24. Mony L, Paoletti P. Mechanisms of NMDA receptor regulation. Curr Opin Neurobiol. (2023) 83:102815. doi: 10.1016/j.conb.2023.102815

25. Ogawa S, Koga N, Hattori K, Matsuo J, Ota M, Hori H, et al. Plasma amino acid profile in major depressive disorder: Analyses in two independent case-control sample sets. *J Psychiatr Res.* (2018) 96:23–32. doi: 10.1016/j.jpsychires.2017.09.014

 Umehara H, Numata S, Watanabe S-y, Hatakeyama Y, Kinoshita M, Tomioka Y, et al. Altered KYN/TRP, Gln/Glu, and Met/methionine sulfoxide ratios in the blood plasma of medication-free patients with major depressive disorder. *Sci Rep.* (2017) 7:4855. doi: 10.1038/s41598-017-05121-6

27. Wang Z, Liang M, Li H, Liu B, Yang L. L-Methionine inhibits 4-hydroxy-2nonenal accumulation and suppresses inflammation in growing rats. *Nutr Res Pract.* (2022) 16:729. doi: 10.4162/nrp.2022.16.6.729

28. Bilen M, Ibrahim P, Barmo N, Abou Haidar E, Karnib N, El Hayek L, et al. Methionine mediates resilience to chronic social defeat stress by epigenetic regulation of NMDA receptor subunit expression. *Psychopharmacol.* (2020) 237:3007–20. doi: 10.1007/s00213-020-05588-8

29. Peng TR, Cheng HY, Wu TW. S-Adenosylmethionine (SAMe) as an adjuvant therapy for patients with depression: An updated systematic review and meta-analysis. *Gen Hosp Psychiatry.* (2024) 86:118–26. doi: 10.1016/j.genhosppsych.2024.01.001

30. Nelson JC. S-adenosyl methionine (SAMe) augmentation in major depressive disorder. Am J Psychiat. (2010) 167:889–91. doi: 10.1176/appi.ajp.2010.10040627

31. Sarkisova KY, Gabova AV, Fedosova EA, Shatskova AB, Narkevich VB, Kudrin VS. Antidepressant and anxiolytic effects of L-methionine in the WAG/Rij rat model of depression comorbid with absence epilepsy. *Int J Mol Sci.* (2023) 24:12425. doi: 10.3390/genes1030413

32. Wickramasuriya SK,E, Cho HM, Shin TK, Kim B, Lee M, Seo S, et al. Differential effects of dietary methionine isomers on broilers challenged with acute heat stress. *J Poult Sci.* (2019) 56:195–203. doi: 10.2141/jpsa.0180072

33. Uramatsu M, Liu G, Uramatsu S, Zhang M, Wang W, Nakayama K, et al. Different effects of sulfur amino acids on prolidase and prolinase activity in normal and prolidase-deficient human erythrocytes. *Clin Chim Acta.* (2007) 375:129–35. doi: 10.1016/j.cca.2006.06.027

34. Bian X, Zhou N, Zhao Y, Fang Y, Li N, Zhang X, et al. Identification of proline, 1pyrroline-5-carboxylate and glutamic acid as biomarkers of depression reflecting brain metabolism using carboxylomics, a new metabolomics method. *Psychiatry Clin Neurosci.* (2023) 77:196–204. doi: 10.1111/pcn.13517

35. Lanoue K, Walajtys E, Williamson J. Regulation of glutamate metabolism and interactions with citric-acid cycle in rat-heart mitochondria. *J Biol Chem.* (1973) 248:7171–83. doi: 10.1016/S0021-9258(19)43375-9

36. Mcentee W, Crook T. Glutamate-its role in learning, memory, and the aging brain. *Psychopharmacol.* (1993) 111:391–401. doi: 10.1007/BF02253527

37. Jones W, Soper T, Ueno H, Manning J. D-glutamate-D-amino acid transaminase from bacteria. *Methods Enzymol.* (1985) 113:108–13. doi: 10.1016/S0076-6879(85) 13024-7

38. Jia S, Wang R, Zhang D, Guan Z, Ding T, Zhang J, et al. Quercetin modulates the liver metabolic profile in a chronic unpredictable mild stress rat model based on metabolomics technology. *Food Funct.* (2023) 14:1726–39. doi: 10.1039/d2f003277e

39. Dudel J. Both d- and l-glutamate induce transporter-mediated presynaptic autoinhibition of transmitter release. *Eur J Neurosci.* (2004) 20:161–6. doi: 10.1111/j.1460-9568.2004.03471.x

40. Han H, Miyoshi Y, Koga R, Mita M, Konno R, Hamase K. Changes in d-aspartic acid and d-glutamic acid levels in the tissues and physiological fluids of mice with various d-aspartate oxidase activities. *J Pharm BioMed Anal.* (2015) 116:47–52. doi: 10.1016/j.jpba.2015.05.013

41. Thielen JW, Hong D, Rohani Rankouhi S, Wiltfang J, Fernández G, Norris DG, et al. The increase in medial prefrontal glutamate/glutamine concentration during memory encoding is associated with better memory performance and stronger functional connectivity in the human medial prefrontal-thalamus-hippocampus network. *Hum Brain Mapp.* (2018) 39:2381–90. doi: 10.1002/hbm.24008

42. Hertz L. Reciprocal changes in forebrain contents of glycogen and of glutamate/ glutamine during early memory consolidation in the day-old chick. *Brain Res.* (2003) 994:226–33. doi: 10.1016/j.brainres.2003.09.043

43. Scholl J, Kolling N, Nelissen N, Stagg CJ, Harmer CJ, Rushworth MFS. Excitation and inhibition in anterior cingulate predict use of past experiences. *eLife.* (2017) 6: e20365. doi: 10.7554/eLife.20365

44. Spurny B, Seiger R, Moser P, Vanicek T, Reed MB, Heckova E, et al. Hippocampal GABA levels correlate with retrieval performance in an associative learning paradigm. *NeuroImage*. (2020) 204:116244. doi: 10.1016/j.neuroimage.2019.116244

45. Maes M, Verkerk R, Vandoolaeghe E, Lin A, Scharpe S. Serum levels of excitatory amino acids, serine, glycine, histidine, threonine, taurine, alanine and arginine in treatment-resistant depression: modulation by treatment with antidepressants and prediction of clinical responsivity. *Acta Psychiatr Scand.* (1998) 97:302–8. doi: 10.1111/j.1600-0447.1998.tb10004.x

46. Muneoka K, Shirayama Y, Horio M, Iyo M, Hashimoto K. Differential levels of brain amino acids in rat models presenting learned helplessness or non-learned helplessness. *Psychopharmacol.* (2013) 229:63–71. doi: 10.1007/s00213-013-3080-2

47. Aghajani R, Naderi N, Sadeghi N, Ani M, Ani S, Nasr-Esfahani MH. Distribution of plasma one-carbon metabolism factors and amino acids profile in depression state treated with paroxetine: A model study. *Cell J.* (2023) 25:165–75. doi: 10.22074/ cellj.2023.557498.1056

48. Song J, Ma W, Gu X, Zhao L, Jiang J, Xu Y, et al. Metabolomic signatures and microbial community profiling of depressive rat model induced by adrenocorticotrophic hormone. *J Transl Med.* (2019) 17:224. doi: 10.1186/s12967-019-1970-8

49. Chen H, Xie H, Huang S, Xiao T, Wang Z, Ni X, et al. Development of mass spectrometry-based relatively quantitative targeted method for amino acids and neurotransmitters: Applications in the diagnosis of major depression. *J Pharm BioMed Anal.* (2021) 194:113773. doi: 10.1016/j.jpba.2020.113773

50. Brann E, Malavaki C, Fransson E, Ioannidi MK, Henriksson HE, Papadopoulos FC, et al. Metabolic profiling indicates diversity in the metabolic physiologies associated with maternal postpartum depressive symptoms. *Front Psychiatry.* (2021) 12:685656. doi: 10.3389/fpsyt.2021.685656

51. Wang Y, Zhou J, Ye J, Sun Z, He Y, Zhao Y, et al. Multi-omics reveal microbial determinants impacting the treatment outcome of antidepressants in major depressive disorder. *Microbiome*. (2023) 11:195. doi: 10.1186/s40168-023-01635-6

52. Kakegawa W, Miyoshi Y, Hamase K, Matsuda S, Matsuda K, Kohda K, et al. Dserine regulates cerebellar LTD and motor coordination through the delta2 glutamate receptor. *Nat Neurosci.* (2011) 14:603–11. doi: 10.1038/nn.2791

53. Rampino A, Garofalo M, Nuzzo T, Favia M, Saltarelli S, Masellis R, et al. Variations of blood D-serine and D-aspartate homeostasis track psychosis stages. *Schizophr (Heidelb)*. (2024) 10:115. doi: 10.1038/s41537-024-00537-2