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# Impact of chronic benzene poisoning on aberrant mitochondrial DNA methylation: A prospective observational study

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Benzene is used as an industrial solvent, which may result in chronic benzene poisoning (CBP). Several studies suggested that CBP was associated with mitochondrial epigenetic regulation. This study aimed to explore the potential relation between CBP and mitochondrial DNA (mtDNA) methylation. This prospective observational study enrolled CBP patients admitted to Shenzhen Prevention and Treatment Center for Occupational Diseases hospital and healthy individuals between 2018 and 2021. The white blood cell (WBC), red blood cell (RBC), hemoglobin (HB), and platelet (PLT) counts and mtDNA methylation levels were measured using blood flow cytometry and targeted bisulfite sequencing, respectively. A total of 90 participants were recruited, including 30 cases of CBP (20 females, mean age 43.0  $\pm$  8.0 years) and 60 healthy individuals (42 females, mean age 43.5  $\pm$  11.5 years). This study detected 168 mitochondrial methylation sites >0 in all study subjects. The mtDNA methylation levels in the CBP cases were lower than the healthy individuals [median  $\pm$  interquartile-range (IQR), 25th percentile, 75th percentile: (1.140  $\pm$ 0.570, 0.965, 1.535)% vs. median  $\pm$  IQR, 25th percentile, 75th percentile: (1.705  $\pm$  0.205,1.240,2.445)%, P < 0.05]. Additionally, the spearman correlation analysis showed that the mtDNA methylation levels were positively correlated with the counts of circulating leukocytes [WBC (r = 0.048, P = 0.036)] and platelets [PLT (r = 0.129, P < 0.01)]. We provided solid evidence of association between CBP and aberrant mtDNA methylation.

#### KEYWORDS

benzene poisoning, mitochondrial DNA, methylation, prospective observational study, white blood cells (WBC), platelet

## Introduction

Exposure to benzene, an industrial solvent, may inflict chronic benzene poisoning (CBP) on the workers, especially in developing countries, as well as certain lung diseases due to severe hazardous occupational exposure through the respiratory tract (1). Benzene was verified to cause several pathological conditions in humans *via* altering DNA, and it has been classified as a human hematological carcinogen by the International Agency for Research on Cancer (IARC) (2). Metabolism of benzene to toxic metabolites is generally thought to be a critical event in benzene-induced toxicity (3). Benzene-induced hematotoxicity and leukemia may be attributed to these metabolites, which could cause cell apoptosis, mitochondrial dysfunction, and oxidative stress. Benzene could induce DNA alterations in exposed workers including DNA methylation, structural damage such as single and double-strand DNA breaks, and accumulation of point mutations (4, 5). In a previous report, a significantly elevated level of mitochondrial DNA (mtDNA) copy number was observed in the benzene-exposed group compared with in the healthy individuals (6). The changing in mtDNA may be an early molecular event in human cells after they are stimulated by endogenous or exogenous oxidative stress (7). Rapid depolarization of mitochondrial inner membrane potential and subsequent damage to oxidative phosphorylation could be induced by oxidative stress (8). Adults exposed to low levels of benzene have impaired hematopoiesis function (9).

DNA methylation plays a very crucial role in the pathogenesis and progression of CBP (10). Previous research has shown that long-term occupational exposure to benzene with concentration below the allowable exposure limit may still alter DNA methylation of anti-oncogene, which may ultimately lead to the development of cancer (11). A number of biochemical changes are induced by benzene exposure, including changes in histone acetylation and DNA methylation (12). Previously, some studies reported that altered global DNA methylation and endoreduplication were potential mechanisms of aneuploidy induction by the benzene metabolite hydroquinone (HQ) in TK6 cells (13). Signal net analysis of differential methylation genes associated with CBP showed that two key hypomethylated STAT3 was identified (14). The methylation level at the third CpG site of p15 promoter region that may permit unlimited proliferation of the cell and is thought to be associated with leukemogenesis, was significantly higher in the CBP group than in the control group (15). In patients who had occupational CBP, mitochondrial cytochrome c oxidase subunit I gene methylation was low and correlated with WBC counts (16). The role of epigenetic regulation of methylation in the pathological regulation of CBP deserves further study. Therefore, based on mitochondrial oxidation, the author hypothesized that aberrant mtDNA methylation might play an important regulatory role in those at risk of exposure to benzene or benzene-related compounds. This study aimed to explore the potential relations between mtDNA methylation and CBP.

## **Methods**

### Study design and population

This prospective observational study enrolled CBP patients admitted to Shenzhen Prevention and Treatment Center for Occupational Diseases and healthy individuals from several major factories between 2018 and 2021. Written informed consent was obtained from all patients and healthy individuals before sample and data collection. The principle of the Helsinki Declaration for using human subjects was employed. The study protocol was approved by the ethics committee of the hospital. CBP patients were diagnosed between 2018 and 2021 by the authorized local Occupational Disease Diagnostic Team, with (a) total white blood cell (WBC) counts  $<4.0 \pm 10^9$ /L or WBC counts between  $4.0 \pm 10^9$ /L and  $4.5 \pm 10^9$ /L and platelet (PLT) counts  $<80 \pm 10^9$ /L, and repeated confirmation of these counts in a peripheral blood sample after a few months; (b) documented benzene exposure as a result of employment in a factory for at least 6 months, (c) exclusion of other known causes of abnormal blood cells counts, such as chloromycetin use and ionizing radiation. The diagnostic criteria for occupational CBP are issued by the Ministry of Health and Family Planning Committee (17). Eligible healthy individuals had normal physical examination results and no history of genetic diseases.

## Data collection

The medical records of these patients were independently reviewed by at least two hematopathologists. Each one of the participants donated 2 ml of venous blood and their demographic data were recorded. All subjects underwent a rigorous physical examination.

#### WBC and PLT and HB and RBC determinations

The WBC, PLT, hemoglobin (HB), and red blood cells (RBC) counts were evaluated with SYSMEX automatic blood analyzer (Sysmex Corporation, Kobe, Japan).

#### **DNA** extraction

Nucleic acids were extracted from the blood samples by extraction DNA Mini kit (Sangon Biotech, Shanghai, China), according to the manufacturer's instruction. DNA quantity was assessed by the spectrometer (Thermo Scientific NanoDrop, NanoDrop Technologies, Wilmington, DE, USA). The qualified DNA was stored at  $-80^{\circ}$ C until analysis.

#### DNA methylation profiling

Genomic DNA was treated for bisulfite sequencing with EpiTect Bisulfite Sequencing kit (Qiagen). Based on the Acegen Targeted Methyl Panel multiple targeted sulfite sequencing system software, and the target sequence (mitochondrial gene ID: NC\_012920.1) multiple sulfite PCR amplification primer pool (upstream primer pool A and downstream primer pool B, primer length 26-40 bases, annealing temperature 55-65; the length of the amplicon is 150-250 bp), the sequencing library was prepared using the Acegen Targeted Methyl Panel Kit amplification system. The primer sequences are listed in Supplementary Table S1. First, 50-200 ng of genomic DNA was recovered by sulfite conversion (ZYMO EZ DNA Methylation-Gold Kit, Zymo Research, Irvine, CA, USA) for multiple sulfite PCR amplification for 25-33 cycles; targeted amplification: the product was purified and recovered with 1X magnetic beads (Agencourt AMPure XP, Beckman Coulter) and used for the preparation of small DNA fragment libraries. Acegen DNA Library Prep Kit (Acegen, Cat. No. AG0810) was used for double-end 8 bp-index library construction. The structure is shown in the figure below. The prepared library was subjected to qualitative detection and concentration determination by Qubit 3.0 and Agilent 2100

Abbreviations: ATP, adenosine triphosphate; ROS, reactive oxygen species; WBC, white blood cell; PLT, platelet; RBC, red blood cells; HB, hemoglobin; mtDNA, mitochondrial DNA; SNP, single nucleotide polymorphism; CBP, chronic benzene poisoning; IARC, International Agency for Research on Cancer; RNR1, 12S ribosomal RNA; RNR2, 16S ribosomal RNA; TRNQ, tRNA glutamine; TRNM, tRNA methionine; TRNW, tRNA tryptophan; TRNS1, tRNA serine 1; ND2, NADH dehydrogenase 2; ND5, NADH dehydrogenase 5; CO1, cytochrome c oxidase subunit 1; CO2, cytochrome c oxidase subunit 2; CO3, cytochrome c oxidase subunit 3; ATP6, ATP synthase membrane subunit 6; CYB, cytochrome-b.

not  $<200\times$ .

Bioanalyzer. The qualified library was used for NGS sequencing using the Illumina sequencing system. The sequencing strategy was pairedend index sequencing, and the sequencing read length was PE150; (a

#### **Bioinformatics analysis**

Qualified data is the basis of information analysis, so quality control of obtained raw data is an essential aspect of data analysis, which mainly includes: (1) Output of raw reads after disembarkation. (2) A clean read is obtained by removing sequencing adapters and low-quality bases and reads using Trimmomatic-0.36. Subsequently, the clean reads and target sequences were aligned and analyzed using methylation sequencing data alignment software (bsmap, version v2.73). The overall statistics are as follows: Raw Q20 (%) average efficiency is 96.45%, clean Q20 (%) average efficiency is 97.62%, mapping\_rate (%) average efficiency is 96.45%. All data are >95%.

The methylation level of CG bases on each target sequence of each sample was calculated. The methylation level was calculated according to the following formula: methylation level of C site = number of reads supporting methylation/(supporting methylation number of reads basement + number of reads supporting unmethylation). Detectable site methylation ratio levels for each sample were counted; if the depth was  $<30\times$ , it would be expressed as NA. And the methylation level of each site in each sample was counted; if the depth was  $<30\times$ , it would be expressed as NA. Enrichment of functions and signaling pathways of genetic loci with differential methylation expression was performed based on KOBAS v3.0 (http://bioinfo.org/kobas/) using the KEGG database with corrected *P*-value < 0.05.

### Statistical analysis

All analyses were performed with SPSS software version 16.0 (SPSS Inc., Chicago, III., USA). Demographic data, such as age at sampling, and sex, were compared between groups using Student's *t*-test or Fisher's exact test. Mann Whitney's *U*-test were utilized to identify a discrepancy of methylated mtDNA levels between the groups. The correlation between mtDNA methylation levels and blood cells counts was determined using Spearmans correlation coefficient. A two-side *P*-value <0.05 was considered significant.

## Results

A total of 90 participants were recruited including 30 cases of CBP (20 females, aged 43.0  $\pm$  8.0 years) and 60 healthy individuals (42

females, aged 43.5  $\pm$  11.5 years). There was no difference in age and gender distribution between the CBP patients and healthy individuals (all P > 0.05), while the CBP patients had significantly lower blood indices, including WBC count, PLT count, HB concentration, and RBC count (all P < 0.001), than the healthy individuals (Table 1).

The average read depth of the sequencing data was  $\geq$ 2,000. The sequencing results showed a total of 168 CpG mtDNA methylation sites >0. The raw sequence can be accessed at https://www.ncbi.nlm.nih.gov/sra/PRJNA877783. The total mtDNA methylation level in the case group was lower than that in the control group, and the difference was statistically significant. In total, 168 CpG were identified, of which 47 showed up-regulated methylation



#### FIGURE 1

Line chart for visualization of methylation levels. Based on the absolute coordinates of the detected gene region, the methylation level of each locus in each sample is plotted, and marked with different colors according to biological groups, and the average methylation of each locus within the group is calculated. This form more intuitively displays the position of differential methylation on the gene, the average methylation level and the significance of the difference. There are significantly fewer mtDNA methylations in patients with CBP than in healthy individuals in the following regions: RNR1, RNR2, TRNQ, TRNM, TRNW, TRNS1, ND2, NDT, COL, CO2, CO3, ATP6, and CYB.

TABLE 1	Basic profile	of the patients	and control group.
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	Age (year)	Female (%)	WBC (10 <sup>9</sup> /L)	PLT (10 <sup>9</sup> /L)	HB (g/L)	RBC (10 <sup>12</sup> /L)
Cases	$43.0\pm8.0$	68.3	$3.78 \pm 1.387$	$153.80\pm58.31$	$113.33\pm16.34$	$3.86\pm0.65$
Controls	$43.5\pm11.5$	70	$5.74 \pm 1.413$	$244.92\pm51.99$	$138.22\pm13.22$	$4.89\pm0.65$
Т	1.565	0.026▲	6.248	7.524	7.768	7.101
SIG	0.121	0.872	$P < 0.001^{*}$	$P < 0.001^{*}$	$P < 0.001^{*}$	P < 0.001*

\*P < 0.05.

A Pearson Chi-square value.

level and 121 down-regulated. The differences of methylation level between CBP cases and the healthy individuals in the up-regulated gene loci were not statistically significant. The differences of methylation level in some down-regulated gene loci between CBP and the healthy individuals were statistically significant. Among the 168 loci, methylation level in the case group was lower than in the control group in 79 loci, and the differences were statistically significant (all P < 0.05). There were significantly fewer mtDNA methylations in patients with CBP than in healthy individuals in the following regions: RNR1, RNR2, TRNQ, TRNM, TRNW, TRNS1, ND2, ND5, CO1, CO2, CO3, ATP6 and CYB (all *P* < 0.05; Figure 1). KEGG pathway analyses identified significantly enriched pathways in the differentially methylated CpG, including aminoacyltRNA biosynthesis, oxidative phosphorylation, Parkinson's disease, ribosome biogenesis in eukaryotes, Staphylococcus aureus infection, retrograde endocannabinoid signaling, pertussis, ribosome, thermogenesis, complement and coagulation cascades, systemic lupus erythematosus, Alzheimer's disease, Huntington disease, metabolic pathways signaling pathway (Figure 2). The mtDNA methylation levels from the CBP patients [median  $\pm$  IQR, 25th percentile, 75th percentile: (1.140 ± 0.570, 0.965, 1.535)%] were significantly lower than those from the healthy individuals [median  $\pm$  IQR, 25th percentile, 75th percentile: (1.705  $\pm$  0.205, 1.240, 2.445)%; U = 525, P = 0.018] (Figure 3).

Additionally, the Spearmans correlation coefficient showed that the mtDNA methylation levels were positively correlated with the counts of circulating leukocytes [WBC (r = 0.220, P = 0.036)] and of platelets [PLT (r = 0.359, P < 0.01); Figure 4]. The Spearman correlation analysis among all patients. (a) Correlation between age and methylation levels in CBP (r = 0.144, P = 0.436). (b) Correlation between work year and methylation levels in CBP (r = 0.228, P = 0.223; Figure 5).

## Discussion

The results of this study revealed that CBP patients had significantly aberrant/lower mtDNA methylation level compared to the healthy individuals. This finding highlights the effect of CBP on mtDNA methylation and peripheral blood cells which may provide clues for further research on the mechanisms of peripheral blood cytopenia during the development of CBP.

Benzene exposure, both occupational and non-occupational, has the potential to cause cancer in humans (18). Early diagnosis and management of CBP have always been a challenge to clinicians. The diagnosis standard of CBP is based on a history of exposure, clinical presentation, and change in blood indices. However, these



Scatter plot plus bee colony plot showing differences between cases and controls. Statistical analysis by MannWhitney's *U*-test found that there is a significant difference between the cases [(median  $\pm$  IQR, 25th percentile, 75th percentile: (l.140  $\pm$  0.570, 0.965, 1.535)%] and controls [(median  $\pm$  IQR, 25th percentile, 75th percentile: (1.705  $\pm$  0.205, l.240, 2.445)] (*P* < 0.05). The methylation site detection results are shown in the figure.



FIGURE 2

Pathway enrichment of the identified mitochondrion genes. Fold enrichment is calculated by dividing gene ratio (the number of genes enriched in the pathway divided by the total number of analyzed genes) with background ratio in KEGG database. *P*-values were adjusted.



The Spearman correlations among the indicators in all participants. The line in red represented a positive correlation. (A) WBC and methylation (r = 0.220, P = 0.036). (B) PLT and methylation (r = 0.359, P < 0.05).



diagnostic features are difficult to determine in the early stage of the disease. Although molecular diagnostic methods have been explored for use in auxiliary diagnosis and toxicological mechanism studies of CBP, there is still much for improvement. This study performed a comprehensive mtDNA methylation analysis to better identify biomarkers of aberrant DNA methylation in CBP. DNA methylation is a stable epigenetic mark involved in the regulation of gene expression and disease pathophysiology (19). Some studies have suggested that the methylation of specific loci in DNA from body fluid samples has been found useful in cancer diagnosis and prognosis (20). DNA damage and subsequent dysfunction in the mitochondrion are important factors in a range of human diseases due to their roles in cell metabolism (21). Thus, there is increasing interest in the role of mitochondrial epigenetics in common diseases associated with mitochondrial dysfunction (22). In this regard, several studies have searched for relationships between mtDNA methylation levels and diseases. For example, reduced mtDNA copy number and increased methylation levels of specific loci at mtDNA have been shown to increase the risk of macrosomia (23). Additionally, mtDNA hypermethylation has been shown to precede diabetic retinopathy, with mtDNA methylation decreasing after the onset of diabetic retinopathy (24). Therefore, mitochondrial epigenetic markers are crucial in early disease detection and disease prevention.

Our research found that mtDNA methylation levels globally decreased in CBP patients. We also analyzed the differences of each methylation site in mitochondria between the two groups. The difference of methylation level between CBP and the healthy individuals in any of the up-regulated gene loci was not statistically significant, but the differences in some of the down-regulated gene loci between the two groups were statistically significant. Therefore, the down-regulated genes were the focus of this study. The article published by Wang et al. (16) suggested that the methylation levels of two sites in the mitochondrial CO1 gene region were significantly reduced in benzene poisoned patients. This study also found that the methylation levels of multiple sites in the CO1 gene region were significantly reduced in benzene poisoning patients. However, the specific methylation sites in this study are different from those in Wang's previous study. The region reported in the previous study of Wang et al. was examined using pyrosequencing to detect the short sequence of the gene region, while the current study using a different detection method did not include that methylated gene locus. Previous studies suggest that mtDNA methylation levels may have a regulatory effect on the transcription of target proteins (25). This corresponding regulatory process promotes changes in the expression of downstream target proteins, which lead to the generation of mitochondrial reactive oxygen species (ROS) and apoptosis. In addition, during occupational benzene exposure, oxidative stressors such as ROS may elevate because of benzene and benzene metabolites, which may affect the proliferation of cells at the molecular level in the bone marrow by stimulating various epigenetic modifications, resulting in changes in peripheral blood cells counts (26). This study found that decreased mtDNA methylation levels correlated with lower leukocytes and platelets counts, demonstrating that mtDNA methylation may play a critical role in the pathological process of benzene-induced cytopenia. Pathway analysis identified 14 significant pathways involving AminoacyltRNA biosynthesis, Oxidative phosphorylation. Aminoacyl-tRNA catalyzes the tRNA aminoacylation reaction to generate aminoacyltRNA, which ensures the speed and accuracy of protein synthesis. Oxidative phosphorylation is the main step in ATP synthesis. Protein and ATP biosynthesis is a major step in leukocyte and platelet production. This also explains the reduced methylation levels of associated mitochondrial regulatory genes in the benzene poisoning population when leukocytes and platelets are reduced to regulate the elevated levels of their target proteins used to replenish essential substances for cell synthesis. In addition, we performed a correlation analysis of age and work year with mean mitochondrial methylation levels in the CBP patients and found no significant correlation. This also indicates that age and work year do not have an effect on the results of this study. The results of this study suggest that changes in mitochondrial methylation levels occur after benzene exposure acts to produce toxicity in the organism, independent of the benzene exposure time-of-action effect. As aforementioned, DNA methylation is crucial for multiple cellular processes, thus it is understandable that its deregulation has been linked to serve diseases. In blood DNA, aberrant methylation is detected as a molecular early screening tool for cancer and other serve diseases. As a result, methylation signatures have the potential to serve as response markers in CBP. Future studies should explore whether the changes in mtDNA methylation level occur earlier than in peripheral blood cells counts.

The study also has several limitations. First, the sample size of the case group is relatively limited, thus the findings need to be further validated. Second, this study did not investigate the temporal relationship between the duration of benzene exposure and mtDNA methylation; such a relationship would aid in determining the earliest time when mtDNA modifications occur in those who are at risk. Third, the blood sample we used contained different types of cells, which might lead to confounding, and mtDNA alterations in underrepresented cells might be underestimated.

In conclusion, this study found that CBP might be associated with aberrant/lower mtDNA methylation level which was positively correlated with the counts of circulating leukocytes and platelets. The findings may provide clues for further research on the mechanisms of peripheral blood cytopenia during the development of CBP.

# Data availability statement

The datasets presented in this study be can found online The of the in repositories. names repository/repositories and accession number(s) can be found below: [http://www.ncbi.nlm.nih.gov/sra/PRJNA877783].

# **Ethics statement**

The studies involving human participants were reviewed and approved by the Ethical Committee of Shenzhen Prevention and Treatment Center for Occupational Diseases. The patients/participants provided their written informed consent to participate in this study.

# Author contributions

DW and DL: conceptualization. GF and XY: investigation and data curation. GF: formal analysis and methodology. WZ and NZ: supervision. PL, ZZ, and YW: writing-original draft. DW, DL, and YG: writing-review and editing. All authors contributed to the article and approved the submitted version.

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# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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# Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpubh.2023. 990051/full#supplementary-material

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