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EDITED BY

Ciro Fernando Bustillo LeCompte,
Toronto Metropolitan University, Canada

REVIEWED BY

Nitya Kumar,
Royal College of Surgeons in Ireland, Bahrain
Jaye Marchiandi,
International Agency for Research on Cancer,
France

*CORRESPONDENCE

Jue Wang
✉ wangjue@must.edu.mo
Xin Lai
✉ laix6@mail.sysu.edu.cn
Yu Li
✉ yuli@must.edu.mo

[†]These authors have contributed equally to
this work and share first authorship

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Assessment of ovarian dysfunction induced by environmental toxins: a systematic review

Lu An^{1†}, Yali Huang^{1†}, Yunkai Wang^{1†}, Shuting Shen¹,
Xuexing Luo², Xiaoyan Liang^{3,4}, Liming Lu⁵, Chunzhi Tang⁵,
Jinglin Lin^{4,6}, Ting Su^{4,6}, Meiqi Zhan^{4,6}, Dongying Wang^{4,6},
Jue Wang^{1,7*}, Xin Lai^{4,6*} and Yu Li^{1,7*}

¹Faculty of Chinese Medicine, Macau University of Science and Technology, Macao, Macao SAR, China, ²Faculty of Humanities and Arts, Macau University of Science and Technology, Macao, Macao SAR, China, ³Department of Reproductive Center, The Sixth Affiliated Hospital, Sun Yat-sen University, Guangzhou, China, ⁴Biomedical Innovation Center, The Sixth Affiliated Hospital, Sun Yat-sen University, Guangzhou, China, ⁵Medical College of Acu-Moxi and Rehabilitation, Guangzhou University of Chinese Medicine, Guangzhou, China, ⁶Department of Traditional Chinese Medicine, The Sixth Affiliated Hospital, Sun Yat-sen University, Guangzhou, China, ⁷State Key Laboratory of Quality Research in Chinese Medicines, Macau University of Science and Technology, Macao, Macao SAR, China

Objective: This study systematically assess the potential impact of various environmental pollutants as chemical, airborne, and heavy metal on ovarian function in women, focusing on ovarian reserve such as anti-Müllerian hormone (AMH) and antral follicle count (AFC) as well as hormone levels like follicle-stimulating hormone (FSH) and estradiol (E2). By reviewing epidemiological evidence, this research aims to elucidate the reproductive toxicity of these pollutants and provide scientific support for public health policy to protect reproductive health in women of childbearing age.

Methods: Following the PRISMA-P guidelines, a comprehensive search was conducted in PubMed, EMBASE, Cochrane Library, and Web of Science databases to include all relevant studies up to July 30, 2024. The Newcastle-Ottawa Scale (NOS) and the Grading of Recommendations Assessment, Development and Evaluation (GRADE) approach were used to assess study quality.

Results: This study ultimately included 40 cohort study reports derived from 33 distinct studies that analyzed the effects of 20 pollutant types on ovarian function. Results indicate that pollutants, such as perfluoroalkyl and polyfluoroalkyl substances (PFAS), phthalates (PAEs), triclosan, Polychlorinated Biphenyls (PCBs), PM_{2.5}, and SO_x, have a significantly negative impact on ovarian function, especially among younger women (<35 years). Long-term exposure to particulate matter (PM)_{2.5} and PM₁₀ is associated with a substantial decrease in ovarian reserve, while heavy metals (e.g., lead and cadmium) also demonstrate reproductive toxicity. However, these conclusions require validation due to both methodological limitations in the original studies (e.g., heterogeneous exposure assessments and residual confounding) and challenges in evidence synthesis (e.g., inconsistent outcome measures across cohorts), highlighting the need for further research to address these constraints.

Conclusion: This review underscores that specific pollutants (e.g., PCBs, PFAS, PM) pose substantial risks to reproductive health in women of childbearing age, particularly in highly polluted environments. The findings underscore the

importance of regular ovarian health monitoring, especially for women at higher risk due to occupational or environmental factors.

Systematic review registration: PROSPERO CRD42024567744 (accessible at https://www.crd.york.ac.uk/prospERO/display_record.php?ID=CRD42024567744).

KEYWORDS

ovary, ovarian function, ovarian reserve, environmental pollutants, persistent organic pollutants, air pollutants, heavy metal

Introduction

Environmental pollutants pose significant global public health risks due to their increasing types and concentrations in air, water, and soil driven by industrialization, urbanization, and agricultural modernization (1). Numerous studies have shown that air pollutants, such as particulate matter (PM) (2), ozone (O₃), sulfur dioxide (SO₂), and nitrogen oxides (NO_x) (3), are significantly associated with respiratory diseases (4), cardiovascular diseases (5, 6), and neurological disorders (7, 8). For instance, long-term exposure to air pollution can increase the risk of atherosclerosis by inducing oxidative stress and inflammatory responses (9–11). Additionally, chemical pollutants including phthalates (PAEs) (12, 13) and bisphenol A (BPA) (14), act as endocrine disruptors (15), causing hormonal imbalances that can lead to diseases such as diabetes (16), obesity, and cancer (17–19). Persistent organic pollutants (POPs), which resist degradation (20), and heavy metals like lead (Pb), mercury (Hg), and cadmium (Cd) accumulate through the food chain, and prolonged exposure can cause kidney damage (21, 22), neurological disorders (23), and osteoporosis (24).

With the growing awareness of the harmful effects of environmental pollutants, increasing studies have begun to explore their impact on reproductive health, particularly ovarian function in women (25–27). The ovary, essential for oocyte production (28, 29) and sex hormone secretion (30), regulates reproductive functions and contributes to overall health and metabolism balance (31, 32). Ovarian dysfunction, linked to menstrual irregularities, infertility, premature ovarian failure, and increased risks of cardiovascular disease and osteoporosis (33–35), can be exacerbated by pollutants through mechanisms such as inflammation, oxidative stress, and endocrine disruption (36) (Figure 1). Environmental pollutants are classified as persistent and non-persistent types (37). Persistent pollutants [e.g., Polychlorinated Biphenyls (PCBs), Per- and Polyfluoroalkyl Substances (PFAS), dioxins], have stable chemical structures, enabling them to remain in the environment for years or even decades (38, 39) and potentially disrupt ovarian reserve and oocyte quality through endocrine disruption and DNA damage (40–42). Non-persistent pollutants, such as PAEs (43) and BPA (44), are rapidly metabolized and excreted by the body (45). However, due to their widespread presence in daily life (46, 47), chronic low-level exposure to these compounds may still disrupt hormonal balance and contribute to ovarian dysfunction (48, 49). Air pollutants like PM_{2.5}, PM₁₀, O₃, and NO_x (50–52), can damage ovarian cells by inducing inflammation and oxidative stress (53, 54). As well as long-term exposure to heavy metals like Pb and Cd promotes ovarian cell apoptosis, gene mutations, and DNA damage (55), reducing ovarian reserve (56) and increasing the risk of infertility (57).

In the following sections, several common and significant pollutants are categorized, with a focus on their widespread use,

primary exposure pathways, half-life, and specific impacts on ovarian function. Please refer to Table 1 for all abbreviations and their full names.

Chemical pollutants

PFAS, PAEs, organochlorine pesticides (OCPs), PCBs, dioxin, parabens, and BPA are widespread in industry, agriculture, soil, and daily necessities due to their chemical stability. These substances enter the human body through drinking water, food, air and skin contact, accumulating over time and adversely affect ovarian function (58).

PFAS are a class of synthetic chemicals renowned for their high stability, with half-lives exceeding half a century in the environment (39). They are widely used in waterproof, oil-repellent, and stain-resistant materials, as well as heat-resistant products, such as food packaging, non-stick cookware, and textiles (59). Exposure occurs primarily through drinking water, food, and inhalation, with PFAS persisting in the human body for 3 to 8.5 years (60). Studies indicated that PFAS exposure significantly impacts ovarian health by inducing excessive reactive oxygen species and triggering cell apoptosis through the intrinsic pathway (61). This oxidative stress impairs mitochondrial function and disrupts structural stability of the oocyte complex, ultimately leading to oocyte apoptosis and necrosis (62).

PAEs are common plasticizers widely used in plastic products, cosmetics, and cleaning products. Despite PAEs have a relatively short half-life (12 h) (63), accumulate with frequent exposure via ingestion, inhalation and skin contact (45, 64), can lead to long-term adverse effects on the human body. PAEs can enhance inflammatory gene expression in human ovaries while reducing ovarian cholesterol and steroid synthesis (65). Animal experiments revealed that exposure to di (2-ethylhexyl) phthalate (DEHP) in female lactating mice significant increase in the expression of DNA damage marker γH2AX in ovarian cells, intensified ovarian cell apoptosis, and notably inhibits proliferation of ovarian granulosa cells (66).

OCPs, like dichlorodiphenyltrichloroethane (DDT) and hexachlorobenzene (HCB), were once widely used in agriculture (67). Although they have been banned (68), they remain in the environment due to their stable physicochemical properties, at high and persistent concentrations, with half-lives in soil that can reach 20 to 30 years (69). OCPs can enter the human body through the food chain, especially through animal-derived foods including poultry, meat, eggs, milk, and in particular, animal fat (70). Once absorbed, OCPs can persist for 10 to 20 years, with the dichlorodiphenyldichloroethylene (DDE) component potentially remaining for a lifetime (71). OCPs disrupt hormone signaling pathways, exhibiting anti-estrogenic and anti-androgenic effects. By inhibiting aromatase activity and steroidogenic enzyme systems, they disrupt endocrine function and negatively impact gonadal health (71).

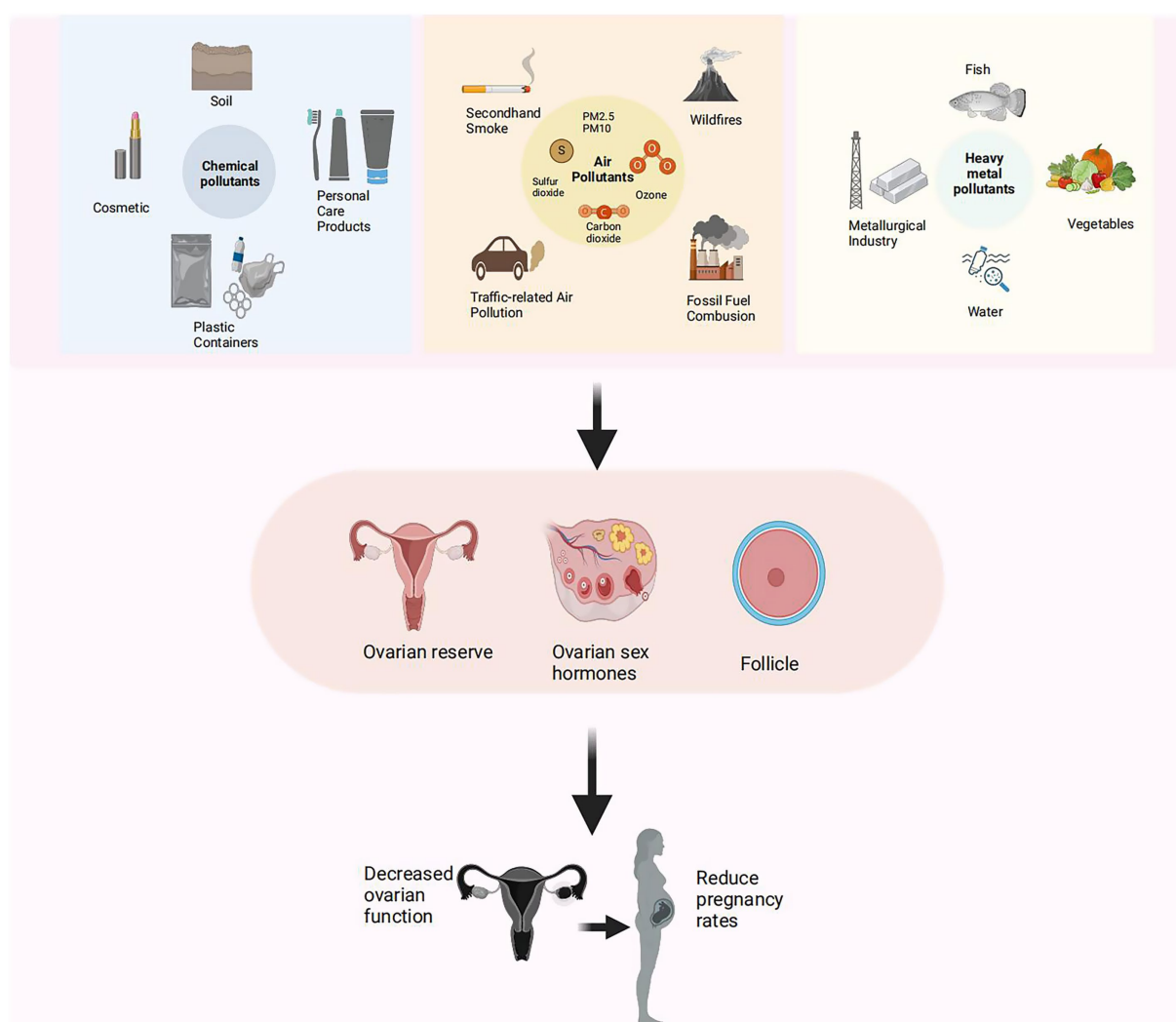


FIGURE 1

Effects of environmental pollutants and heavy metals on ovarian reserve and fertility. Schematic diagram illustrating sources of environmental pollutants (left), including chemical, airborne, and heavy metal contaminants, and their associations with reduced ovarian reserve, hormonal disruption, and lower pregnancy rates (right).

PCBs have been widely used in industrial applications such as printing inks, copying paper, coolants and insulating materials for electrical equipment, paints, and lubricants (72). Although they are now banned internationally, PCBs are still slowly and persistently released from old equipment and landfills into environmental air, soil, water, and sediments (72, 73), eventually contaminating the human food chain (74). Their long half-life in the human body, lasting decades, poses a prolonged health risk (75). PCBs disrupt the expression of crucial enzyme genes, such as CYP17, 3 β -HSD, and CYP19 (76, 77), involved in steroid hormone biosynthesis, thereby negatively impacting the synthesis and secretion of ovarian hormones.

Dioxins are not produced intentionally and have no practical value, they are unintentional by-products of numerous industrial activities and all combustion processes (59, 78). These chemicals are highly persistent in the environment, particularly 2,3,7,8-Tetrachlorodibenzo-p-dioxin (TCDD), which has a half-life of 10 to 12 years in soil (79). Human exposure occurs primarily through dietary intake, especially from the consumption of meat, milk, eggs, fish, and related products (80), with a half-life in the human body

ranging from 5 to 10 years (81). A retrospective cohort study revealed that each 10-fold increase in serum TCDD levels was associated with a 25% prolongation of time-to-pregnancy (TTP) and an approximate doubling of infertility risk (82). Animal studies showed that TCDD exposure increases granulosa cell apoptosis in growing follicles and disrupts follicular maturation (83), leading to a reduction in pre-antral and antral follicle numbers (84).

Parabens, widely used as preservatives in cosmetics, pharmaceuticals, and food products (85), are among the most prevalent ingredients in cosmetics and personal care products after water (86). Although their half-life in the body relatively short (a few hours to days) (87), exposure through skin absorption or ingestion can trigger oxidative stress responses in the body (88), leading to ovarian cell damage or apoptosis (89). These effects may ultimately impair fertility.

BPA is widely used in the production of plastic products, food cans, and thermal paper (90). The primary route of BPA exposure is ingestion of contaminated food (91). Although its half-life in the body is short (4–6 h) (92), continuous exposure from daily

TABLE 1 List of abbreviations.

Abbreviations	Full name	Abbreviations	Full name
POPs	Persistent organic pollutants	PM _{2.5}	Particulate matter 2.5
PFAS	Per- and polyfluoroalkyl substances	PM ₁₀	Particulate matter 10
PFCs	Perfluorochemicals	PAHs	Polycyclic aromatic hydrocarbons
PFOS	Perfluorooctane sulfonate	SO _x	Sulfur oxides
n-PFOA	n-Perfluorooctanoic acid	SO ₂	Sulfur dioxide
Sm-PFOS	Sum of branched-chain perfluorooctane sulfonate	NO _x	Nitrogen oxides
PAEs	Phthalates	NO ₂	Nitrogen dioxide
MEP	Mono-ethyl phthalate	O ₃	Ozone
MBzP	Mono-benzyl phthalate	CO	Carbon monoxide
DEHP	Di (2-ethylhexyl) phthalate	BTEX	Benzene, Toluene, Ethylbenzene, and Xylene
MiNP	Mono-iso-nonyl phthalate	Hg	Mercury
MiDP	Mono-iso-decyl phthalate	Cd	Cadmium
MiBP	Mono-iso-butyl phthalate	Pb	Lead
MBP/MnBP	Mono-n-butyl phthalate	Cr	Chromium
MEHP	Mono-(2-ethyl-hexyl)phthalate	Ba	Barium
MEOHP	Mono(2-ethyl-5-oxohexyl) phthalate	As	Arsenic
DBP	Di-n-butyl phthalate	E2	Estradiol
BPs	Benzophenones	P	Progesterone
OCPs	Organochlorine pesticides	T	Testosterone
HCB	Hexachlorobenzene	AFC	Antral follicle count
DDT	Dichloro-diphenyl-trichloroethane	AMH	Anti-Müllerian hormone
DDE	Dichlorodiphenyldichloroethylene	FSH	Follicle-stimulating hormone
PCBs	Polychlorinated biphenyls	INH B	Inhibin B
BPA	Bisphenol A	LH	Luteinizing hormone
MP	Methylparaben	FF	Follicular fluid
PP	Propylparaben	ER	Estrogen receptors
TCDD	2,3,7,8-tetrachlorodibenzo-p-dioxin	PCOS	Polycystic ovary syndrome
TCS	Triclosan	ART	Assisted reproductive technologies
PM	Particulate matter	IVF	In vitro fertilization
n3PUFA	n-3 polyunsaturated fatty acids	AI	Artificial intelligence
BMI	Body mass index		

Full terminology corresponding to abbreviated forms used throughout the text.

necessities results in long-term endocrine disruption (93). An *in vitro* study demonstrated that exposure to 100 µM BPA significantly inhibited cell proliferation and promoted autophagy in human granulosa cells (94). Studies in mice found that BPA impairs folliculogenesis, leading to cystic expansion of the follicles, a decrease in the number of granulosa cells within the follicles, and a corresponding decline in hormone levels associated with granulosa cells, including estradiol (E2), progesterone (P), and anti-Müllerian hormone (AMH) (95).

Benzophenones (BPs) are commonly found in the formulations of sunscreens, nail polishes, enamel paints, bath products, and skincare products, absorbed through skin and have a half-life of a few days (96). Hydroxylated forms such as benzophenone-3 (BP-3), benzophenone-1 (BP-1), and benzophenone-2 (BP-2) exhibit

estrogenic and anti-androgenic activities in both *in vitro* and *in vivo* experiments (97, 98), thereby interfering with the reproductive system specifically by inhibiting the development of oocytes (98).

Triclosan (TCS) is an antimicrobial agent in personal care products such as soaps, toothpaste, and deodorants, as well as medical devices (99). TCS primarily absorbed through skin absorption and oral ingestion (100). It is metabolized within 15–29 h (101), but its continuous release and environmental persistence can disrupt the endocrine system, leading to reproductive disorders (102). *In vitro* studies showed TCS (0–10 µM) stimulated E2 and P secretion and upregulated steroidogenic genes in human granulosa KGN cells, indicating its endocrine-disrupting potential (103). Research by Wang et al. indicated that exposure to 0.16 mg/L TCS disrupts the hypothalamic–pituitary–ovarian axis in female Yellow River carp by

increasing the synthesis and secretion of E2 and upregulating the hypothalamic mRNA expression levels (104).

Air pollutants

The increasingly severe issue of air pollution, particularly its impact on health, has become a globally recognized concern, especially in developing countries (105, 106). Alongside national development, accelerated urbanization, expanded industrial production, and the prevalence of motorized transportation have significantly increased human exposure to harmful air pollutants (107). These pollutants, mainly including waste gases from industrial emissions (108), road dust, vehicle exhaust (109), and secondary pollutants (110) formed through complex reactions in the atmosphere such as PM (PM10 and PM2.5), Sulfur Oxides (SO_x), and NO_x, can penetrate the alveolar-capillary barrier, enter the bloodstream, and reach other tissues and organs (111).

Evidence from related mechanisms indicates that long-term inhalation of multiple air pollutants can lead to systemic inflammation (112), endothelial dysfunction (113), DNA methylation (114), and granulocyte apoptosis, thereby affecting the normal function of the reproductive system (114). The adverse health effects of PM may stem from its particulate characteristics (size, mass, and even shape) or the chemical components it adsorbs (115, 116). Its impacts on the human body primarily include cytotoxicity, oxidative stress, and inflammatory effects (117). By observing the cytotoxic effects of PM2.5 on Chinese hamster ovary cells over 24 h (118), it was found that PM can continuously activate the nuclear factor-kappa B signaling pathway, promoting the expression of apoptotic genes and proteins, and accelerating the apoptosis of ovarian granulosa cells and oocytes (119). Additionally, SO₂, nitrogen dioxide (NO₂), O₃, and CO are all classified as “criteria” air pollutants by the U. S. Environmental Protection Agency (<https://www.epa.gov/criteria-air-pollutants>). People are often exposed to these pollutants simultaneously, making it difficult to determine the individual effects of specific pollutants. Current mechanistic research suggests that these pollutants may collectively exert negative impacts on the human reproductive system through various pathways such as hormonal disruption (120), oxidative stress induction, cellular DNA alterations (121), and epigenetic changes (122, 123).

Heavy metal pollutants

Heavy metals, as a class of pollutants with persistent toxicity, originate from both human activities such as mining, coal burning, metal processing, and transportation, as well as natural phenomena like rock weathering and volcanic eruptions (124–126). In the environment, common heavy metal contaminants like Cd, Hg, Pb and chromium (Cr) can cause damage to human organs even at trace exposure levels (127). Heavy metals primarily enter the human body through contaminated drinking water, food intake (especially seafood) (128, 129), and smoking (130). Due to their resistance to biodegradation and their tendency to accumulate in organisms while being difficult to metabolize and excrete, heavy metals pose a long-term and continuous threat to human health (131).

Selenia Miglietta and her colleagues found that the levels of Pb and Cd within follicles might be associated with morphological changes in the ultrastructures (such as the endoplasmic reticulum, mitochondria, and nucleus) of oocytes and cumulus cells (132). These morphological changes could potentially lead to the arrest of oocyte maturation, thereby impairing their fertilization capabilities, weakening the steroidogenic activity of cumulus cells, and inducing cumulus cell apoptosis. Furthermore, exposure to multiple toxic metals [such as Cd, Hg, barium (Ba), and arsenic (As)] may also increase the risk of developing polycystic ovary syndrome (PCOS) by disrupting the endocrine system (133, 134).

Despite existing research revealing the association between environmental pollutants and ovarian function, there remain gaps in the depth and breadth of these studies. Current research often focuses on the effects of a single pollutant, lacking exploration of the synergistic effects of multiple pollutants and the cumulative effects of long-term exposure. Furthermore, there is a lack of systematic and comprehensive analysis regarding the effects of multiple pollutants on ovarian function. Therefore, this systematic review aims to address these research gaps by comprehensively evaluating the impact of overall and multiple environmental pollutants (including chemical, gaseous, and heavy metals) on female ovarian function. This review provides an in-depth assessment of pollutant-related reproductive toxicity, helping reduce the incidence of female infertility, support public health policies to protect women's reproductive health, and providing reliable scientific evidence for clinical practice and policy-making.

Materials and methods

Search strategy and selection criteria

This systematic review was designed based on the Preferred Reporting Items for Systematic Review and Meta-Analysis Protocols (PRISMA-P) checklist (135) and the updated version of the Cochrane Handbook for Systematic Reviews (136). The review protocol was registered in the PROSPERO database (Registration number: CRD42024567744).

To comprehensively review the literature on the effects of environmental pollutants on ovarian function, we searched the following electronic databases: PubMed, EMBASE, Cochrane Library, and Web of Science. The search strategy is provided in [Supplementary Table 1](#). The search covered all relevant studies from the inception of each database to July 30, 2024. Additionally, through this comprehensive search strategy, we aimed to expand the scope of the research and ensure the comprehensiveness and accuracy of the findings. During the literature screening process, the inclusion and exclusion criteria were as follows:

Inclusion Criteria: Women of reproductive age (menarche to premenopause, typically 15–49 years) (137) exposed to environmental pollutants (e.g., POPs, industrial waste, heavy metals); women with ovarian dysfunction, primary ovarian insufficiency, poor ovarian response, premature ovarian failure, or reduced ovarian reserve; studies reporting at least one primary outcome measure (e.g., antral follicle count [AFC], AMH, follicle-stimulating hormone [FSH], E2); cohort studies exploring the relationship between environmental pollutant exposure and ovarian function.

Exclusion Criteria: Women with ovarian dysfunction due to iatrogenic factors (including radiotherapy, chemotherapy, and medication), or lifestyle factors. Lifestyle-related ovarian dysfunction was determined based on predefined clinical criteria (138), including obesity [body mass index (BMI) ≥ 30] (139), excessive alcohol intake (>14 units/week) (140), or chronic sleep deprivation (141) with circadian dysregulation (142). Women of reproductive age with ovarian tumors, polycystic ovary syndrome, or other organic ovarian diseases; pregnant women; non-cohort studies, such as cross-sectional studies or case-control studies; studies not reporting relevant outcome measures; non-English publications (143); reviews, editorials, conference abstracts, and unpublished studies.

Data extraction and quality appraisal

Data extraction

Three independent reviewers (LA, YH and YW) participated in the screening, eligibility assessment, and selection review process. Reviewer L. A. was responsible for downloading and conducting an initial screening to systematically review the literature and exclude studies irrelevant to the research question or not meeting the predefined criteria. Reviewers MZ, DW and SS and S. T. S. independently assessed the eligibility of the studies that passed the initial screening based on the predefined inclusion criteria. For studies with disagreements, reviewers YL and XL conducted further independent assessment and made the final decision to ensure scientific rigor and the validity of the selected studies.

After determining the eligibility of cohort studies, reviewers YH and YW independently completed data extraction. To ensure data consistency and accuracy, LA subsequently cross-checked a portion of the extracted data. All data were collected and organized using a pre-designed standardized Excel spreadsheet. Specific data extracted included: first author's name, publication date, country of study, study design type, study objective, data source, sample size, type of exposure pollutants, mode of exposure measurement, characteristics of the study population (e.g., age, ethnicity), primary outcome measures, and relevant statistical indicators. All these details aimed to construct a comprehensive and accurate database to support further analysis of the impact of environmental pollutants on ovarian function.

Quality appraisal

Due to the significant differences in design and implementation between observational studies and randomized controlled trials, we used the NOS (144–146) and the Grading of Recommendations Assessment, Development and Evaluation (GRADE) (147–150) approach to systematically assess the quality and certainty of the evidence from cohort studies.

The NOS primarily evaluates studies in three domains: selection/exposure of study subjects, comparability between groups, and outcome measurement. High-quality studies were those with NOS scores of 8 or higher, medium-quality studies scored between 5 and 7, and low-quality studies scored below 5.

The three authors (LA, YH and YW) independently assessed the certainty of evidence for each type of compound's impact on ovarian function using the GRADE framework. This assessment included evaluating the risk of bias at the outcome level, inconsistency, indirectness, imprecision, and other biases (such as publication bias

or observational study design). GRADE evidence certainty levels: high (research unlikely to change confidence), moderate (research may importantly impact confidence), low (research very likely to impact confidence), and very low (estimates highly uncertain) (147). According to Murad and colleagues (151), the GRADE guidelines were appropriately applied to the narrative synthesis process. In doing so, we referred to Dr. Nafiso Ahmed's team's refined version of the GRADE scoring criteria (152), which was more carefully adapted and optimized to address the differences in research methods involved in each research question. The adaptations were discussed and agreed upon with the working group. For more detailed information, please consult [Supplementary Table 2](#).

Outcome criteria

The effects of environmental pollutants on ovarian function are complex and diverse (153). AMH, AFC, FSH, and E2 are important indicators of ovarian function and are highly sensitive to environmental pollutants, making them effective measures for assessing the potential damage of these pollutants on ovarian function (29, 154).

AMH is a hormone produced by small follicles in the ovary, mainly secreted by antral and pre-antral follicles, and serves as a stable indicator of ovarian reserve. A decrease in AMH levels indicates a decline in ovarian function (155–157). AFC refers to the number of antral follicles detected in the ovary via ultrasound, directly reflecting ovarian reserve. Therefore, higher AFC values indicate better ovarian reserve (158, 159). FSH is a hormone secreted by the pituitary gland that stimulates follicular development in the ovary and reflects the ovary's response to stimulation. FSH is an important hormone for assessing ovarian function, and elevated FSH levels represent ovarian failure or decreased ovarian function due to negative feedback regulation (160). E2 is the primary estrogen secreted by the ovary, and its levels reflect follicular development and ovarian hormone secretion function, with abnormal changes indicating ovarian dysfunction (161, 162). Since AMH and AFC are relatively stable and highly correlated, the expression of AMH is barely affected by the cyclic fluctuations of gonadotropin, remaining relatively stable throughout the menstrual cycle (155, 163, 164). The most accepted predictor for female fertility is ovarian reserve (155). Therefore, in the GRADE framework, we evaluate them as direct evidence of ovarian function.

Data analysis

Due to the complex and diverse effects of environmental pollutants, particularly the variation in chemical derivatives and study characteristics (e.g., study design, exposure dose, exposure duration, and participant demographics), conducting a meta-analysis presented certain challenges. Therefore, this study used a narrative synthesis approach, combining qualitative and quantitative data from existing literature to comprehensively understand the impact of these pollutants on ovarian function. The narrative analysis summarized the main findings of each study, analyze the sources of heterogeneity, explore the potential mechanisms of different environmental pollutants, and discussed the consistency and differences in study results. For studies with significant heterogeneity, specific factors

influencing the outcomes were further examined to enhance the reliability of the analysis.

This study will focus on analyzing the specific effects of environmental pollutant exposure on ovarian function, summarizing and comparing reproductive indicators reported in different studies, including AMH, AFC, FSH, and E2, to evaluate differences or common trends in these indicators across studies. Quantitative analysis will also be conducted to assess the extent to which different types of environmental pollutants, such as BPA, PAEs, PCBs, and heavy metals (e.g., Pb, Cd), impact AMH, AFC, FSH, and E2.

Results

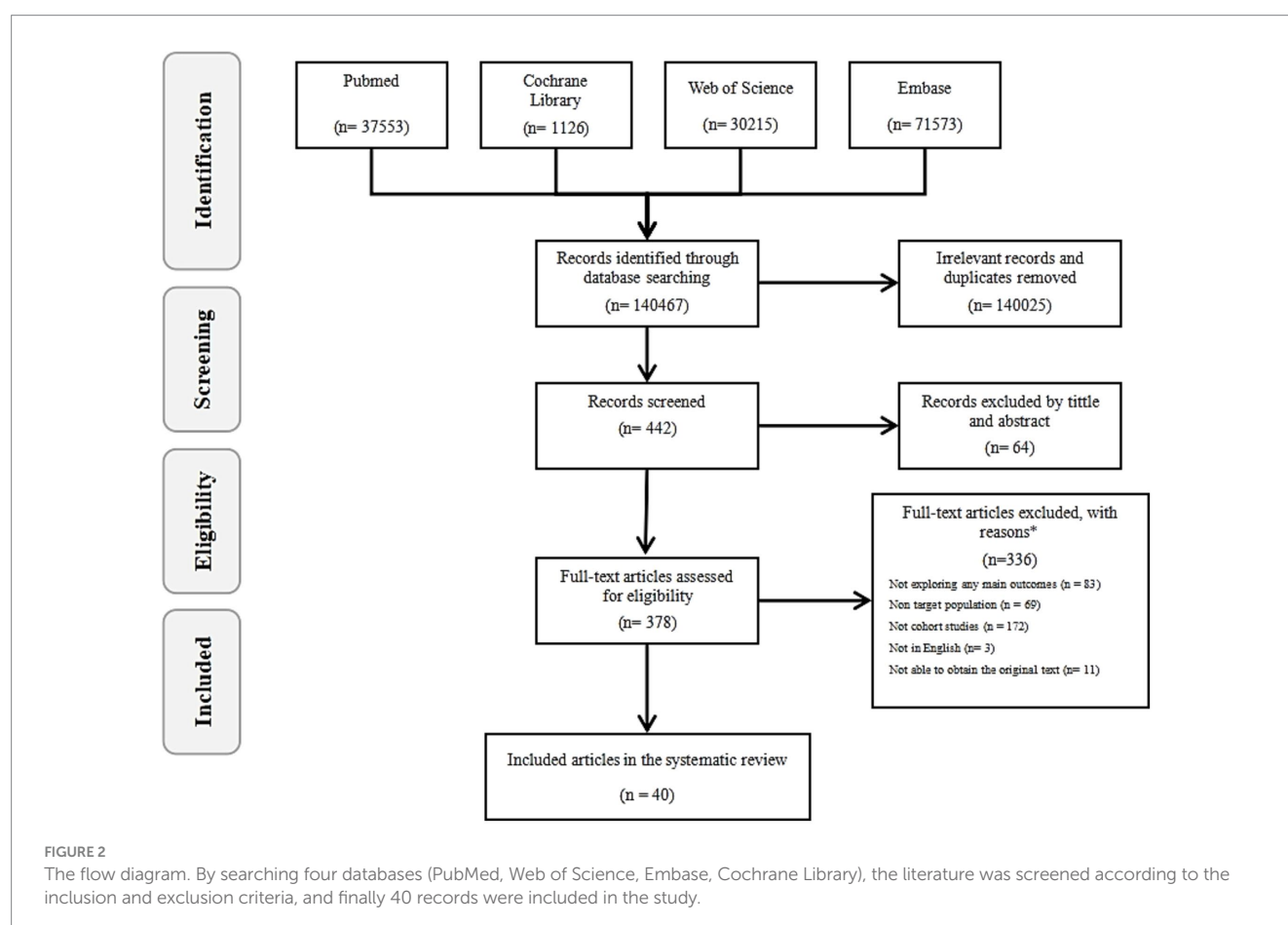
A total of 140,467 results were initially obtained from four databases (PubMed, Web of Science, Embase, Cochrane Library). After removing duplicates and irrelevant entries (140,025), 442 articles remained for further screening (Figure 2). Of these, 64 were excluded at the title and abstract screening stage. Following a full-text review of remaining 378, 338 were excluded based on exclusion criteria, leaving 40 cohort study reports from 33 different studies that analyzed the effects of 20 types of pollutants on ovarian function. Of these 40 reports, 53% (21 studies) reported the effects of chemical industrial pollutants on ovarian reserve, including PFAS, PAEs, BPs, OCPs, PCBs, and other chemical pollutants. Additionally, 37% (15 studies)

reported the effects of air pollutants on ovarian reserve, including gaseous pollutants (e.g., SO_x, NO_x, O₃, Benzene, Toluene, Ethylbenzene, and Xylene [BTEX]) and PM (e.g., PM_{2.5}, PM₁₀). The remaining 10% (4 studies) reported the effects of heavy metals, including Hg, Cd, and Pb, on ovarian function.

Description of studies

Supplementary Table 3 provides an overview of the characteristics of included studies. The 33 studies included women of reproductive age (18–54), with sample sizes ranging from 44 to 19,861 participants. These studies were conducted in the United States (*n* = 10), China (*n* = 8), Poland (*n* = 3), Italy (*n* = 2), South Korea (*n* = 2), Norway (*n* = 1), Denmark (*n* = 1), Sweden (*n* = 1), Israel (*n* = 1), India (*n* = 1), Iran (*n* = 1), Vietnam (*n* = 1), and South Africa (*n* = 1).

The quality of the studies was assessed using the NOS for cohort studies, with 27 studies (70%) rated as high quality and 13 studies (30%) rated as medium quality. Figure 3 presents the bias assessment results of these studies. The quality of evidence for each pollutant's effect on ovarian function was independently assessed using the GRADE assessment method. Among the 20 pollutants studied, GRADE evidence quality was rated as high for 1 pollutant (5%), moderate for 3 pollutants (15%), and 16 pollutants (80%) as low to very low. Detailed information regarding the GRADE evidence profile, characteristics of the study population, NOS assessment of studies



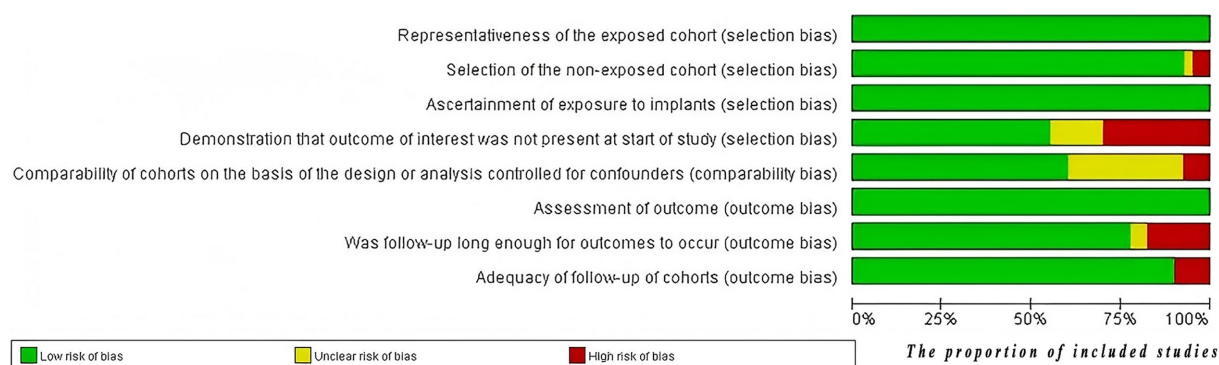


FIGURE 3

The risk of bias graph. The evaluations were conducted using the NOS, and the results figure was exported using RevMan software.

quality, and results of the subgroup analysis can be found in [Supplementary Tables 2, 4–6](#).

Association between chemical pollutants and ovarian function

The GRADE rating table is shown in [Table 2](#).

Polyfluoroalkyl substances

Four prospective studies explored the relationship between PFAS exposure and ovarian function, involving a total of 1,611 women. Of these studies, three were rated as “High” and one as “Medium” according to the NOS, with a GRADE certainty rating of moderate.

Björvang et al. (165) investigated a cohort of women undergoing *in vitro* fertilization (IVF) and found that the total PFAS levels in blood and follicular fluid (FF) were associated with higher baseline AFC, but a lower chance of obtaining high-quality embryos. The study also suggested that PFAS might be associated with lower AMH levels, although not significantly. This result was consistent with findings from Crawford et al. (166), who found no association between serum PFAS and AMH. Barrett et al. (167) discovered that PFAS exposure had different effects in nulliparous and parous women. Among nulliparous women, serum perfluorooctane sulfonate (PFOS) concentrations were significantly negatively correlated with salivary E2 ($\beta = -0.025$, 95% CI: -0.043, -0.007), and these women showed higher sensitivity in ovarian hormone secretion to PFAS exposure compared to parous women. A longitudinal cohort study (168) also found that participants with higher serum PFAS concentrations had a shorter time to natural menopause, with FSH acting as a key mediator in the association between linear-chain perfluorooctanoate (n-PFOA), sum of branched-chain perfluorooctane sulfonate (Sm-PFOS) exposure, and natural menopause incidence, with total effect sizes of 26.9% (95% CI: 15.6, 38.4) and 13.2% (95% CI: 0.0, 24.5), respectively.

Summary: Although the specific strength of the associations varied across studies, the overall evidence consistently indicates that PFAS exposure may adversely affect ovarian function in women. Considering the prospective study designs, the moderate quality ratings from NOS and GRADE, and consistent findings across the studies, we can conclude that the weight of evidence is sufficient to

support the conclusion that PFAS exposure poses a threat to female reproductive health.

Phthalates

Seven studies investigated the relationship between PAEs exposure and ovarian function in women, including six prospective studies and one retrospective study with a total of 2,334 participants. According to the NOS assessment, six studies were rated as “High” quality, and one as “Medium” quality, with GRADE evidence quality rated as low.

Evidence unrelated to ovarian parameters

The only retrospective study reported no association between PAEs and ovarian parameters. No significant dose–response relationship was observed between urinary PAEs metabolites and serum AMH levels (169).

Evidence of decreased ovarian parameters

Five prospective reports indicated that PAEs exposure was associated with a decline in ovarian parameters. Data from the Midlife Women’s Health Study (MWHs) (170) showed that higher cumulative exposure to urinary PAEs metabolites over the four-year study period was associated with lower E2 levels in year four. Mono-ethyl phthalate (MEP) and mono-benzyl phthalate (MBzP) contributed the most, with a doubling in MBzP concentration resulting in a 3.17% decrease in E2 levels (95% CI: -5.72, -0.55%). In the Environment and Reproductive Health (EARTH) study (171), a prospective association was observed between 11 urinary PAEs metabolites and AFC among women undergoing infertility treatment. Across the entire study cohort, higher quartiles of urinary DEHP metabolites were associated with lower AFC, particularly in women under 37 years.

An Indian study (172) found that women in the highest quartile of mono-iso-nonyl phthalate (MiNP) and mono-iso-decyl phthalate (MiDP) exposure had significantly lower AFC and serum AMH levels compared to those in the lowest quartile. Women in the highest quartile of mono-n-butyl phthalate (MBP) also exhibited a declining trend in E2 levels ($p = 0.016$). Higher levels of FF MEP were negatively correlated with average AFC and serum AMH levels, although these trends were not statistically significant. Beck et al. (173) reported that women in the highest tertiles of mono-iso-butyl phthalate (MiBP) and

TABLE 2 GRADE (grading of recommendations assessment, development and evaluation) evidence profile table.

No. of studies	Study quality	Inconsistency	Indirectness	Imprecision	Publication bias	Certainty
Polyfluoroalkyl Substances						
4	No concerns	Serious concerns	Borderline	Borderline	No concerns	Moderate certainty ⊕ ⊕ ⊕ ⊖
Phthalates						
7	No concerns	Borderline	No concerns	Serious concerns	No concerns	Very low certainty ⊕ ⊖ ⊖ ⊖
Benzophenones						
1	No concerns	No concerns	No concerns	Serious concerns	Serious concerns	Very low certainty ⊕ ⊖ ⊖ ⊖
Organochlorine Pesticides						
3	Serious concerns	Borderline	No concerns	Borderline	No concerns	Low certainty ⊕ ⊕ ⊖ ⊖
Polychlorinated Biphenyls						
2	Serious concerns	Serious concerns	No concerns	Serious concerns	No concerns	Very low certainty ⊕ ⊖ ⊖ ⊖
Bisphenol A						
3	Borderline	Borderline	Serious concerns	Serious concerns	No concerns	Very low certainty ⊕ ⊖ ⊖ ⊖
Parabens						
1	No concerns	No concerns	No concerns	Serious concerns	Serious concerns	Very low certainty ⊕ ⊖ ⊖ ⊖
Dioxin						
2	Borderline	Borderline	Serious concerns	Borderline	No concerns	Very low certainty ⊕ ⊖ ⊖ ⊖
Triclosan						
2	Borderline	Borderline	No concerns	Borderline	No concerns	Very low certainty ⊕ ⊖ ⊖ ⊖
PM2.5						
12	No concerns	Borderline	No concerns	No concerns	No concerns	High ⊕ ⊕ ⊕ ⊕
PM10						
9	No concerns	Serious concerns	No concerns	No concerns	No concerns	Low certainty ⊕ ⊕ ⊖ ⊖
SO_x						
6	Borderline	Borderline	No concerns	No concerns	No concerns	Moderate certainty ⊕ ⊕ ⊕ ⊖
NO_x						
10	Borderline	Serious concerns	No concerns	No concerns	No concerns	Low certainty ⊕ ⊕ ⊖ ⊖
O₃						
7	Borderline	Serious concerns	No concerns	No concerns	No concerns	Very low certainty ⊕ ⊖ ⊖ ⊖
CO						
5	Borderline	Borderline	No concerns	No concerns	No concerns	Very low certainty ⊕ ⊖ ⊖ ⊖

(Continued)

TABLE 2 (Continued)

No. of studies	Study quality	Inconsistency	Indirectness	Imprecision	Publication bias	Certainty
BTEX						
1	No concerns	No concerns	No concerns	Serious concerns	Serious concerns	Very low certainty ⊕ ⊕ ⊕ ⊕
Tobacco smoke						
1	No concerns	No concerns	Serious concerns	Serious concerns	Serious concerns	Very low certainty ⊕ ⊕ ⊕ ⊕
Hg						
3	No concerns	Borderline	Borderline	No concerns	No concerns	Moderate certainty ⊕ ⊕ ⊕ ⊕
Cd						
1	No concerns	Borderline	Borderline	Serious concerns	Serious concerns	Very low certainty ⊕ ⊕ ⊕ ⊕
Pb						
1	No concerns	No concerns	Serious concerns	Serious concerns	Serious concerns	Very low certainty ⊕ ⊕ ⊕ ⊕

mono-n-butyl phthalate (MnBP) had an average of five fewer antral follicles compared to the reference group ($p < 0.01$).

Yao et al. (174) conducted an age-stratified analysis and found that, with similar levels of PAEs exposure, the relationship between PAEs metabolite concentration and AFC was positively correlated in women aged ≥ 35 years, whereas it was negatively correlated in women under 35 years. Among younger women, the second and third quartiles of MEP and \sum DEHP (sum of DEHP metabolites) were associated with a 6.50% (95% CI: $-12.8, -0.18\%$) and 7.37% (95% CI: $-13.8, -0.89\%$) reduction in AFC, respectively.

Evidence of increased ovarian parameters

Three studies mentioned that PAEs exposure could lead to increased ovarian parameters. A study from Denmark (173) found that women in the highest tertile of urinary mono-(2-ethyl-hexyl) phthalate (MEHP) had 26% higher E2 levels compared to those in the lowest tertile (95% CI: $5-51\%$, $p = 0.02$). Women in the highest tertiles of MiNP and MiDP showed significant increases in AMH levels by 44% (95% CI: $11-87\%$, $p < 0.01$) and an average increase of six antral follicles (95% CI: $2.14-9.00$, $p < 0.01$), respectively. Hoffmann-Dishon et al. (175) observed a positive correlation between E2 levels in FF and MnBP/MiBP levels ($p < 0.05$). Additionally, Yao et al. (174) analyzed 525 women undergoing IVF treatment and found that MBP, mono(2-ethyl-5-oxohexyl) phthalate (MEOHP), and \sum PAEs (total PAEs) concentrations were positively correlated with AFC in women aged ≥ 35 , whereas a negative correlation was observed in those younger than 35.

Summary: Across seven studies with varied geographic locations, time periods, and exposure ranges, the majority of high-quality prospective studies consistently demonstrated a decline in ovarian parameters, particularly in younger women (age < 35), suggesting a negative impact of PAEs on ovarian function. Although a definitive conclusion on the specific effects of PAEs on the ovaries has not yet been reached, existing evidence indicates that they have a clear disruptive effect on ovarian function. Given the widespread

presence of PAEs in daily life and the potential risks of exposure, we should attach great importance to the potential health issues they may pose.

Benzophenones

Only one study examined the effect of BPs exposure on ovarian function in a cohort of 111 women. This study was rated as “High” quality by the NOS, with a GRADE evidence quality rating of very low.

This Danish study (173) investigated the association between the concentrations of six BPs in the 155 FF samples of 111 women undergoing IVF and fertility outcomes. It was found that women in the highest tertile of BP-3 had 30% higher E2 levels compared to the reference group (95% CI: $2\%; 65\%$, $p = 0.02$). Notably, higher BP-3 levels were also significantly associated with a lower likelihood of live birth.

Summary: Currently, only one high-quality study from Denmark exists in this field. The positive correlation between BP-3 concentration and E2 levels observed in this study raises interest; however, this finding contrasts with the observed reduction in live birth rates among women with higher BP-3 levels, adding complexity to the interpretation. Despite the study’s high quality, the GRADE evidence quality rating was “low,” potentially reflecting limitations such as sample size. More high-quality, large-scale studies are needed to further verify and refine these findings.

Organochlorine pesticides

Three prospective studies investigated the effect of OCPs exposure on ovarian function, involving a total of 637 women. All three studies were rated as “Medium” quality by the NOS, with a GRADE evidence quality rating of low.

A Swedish study (165) analyzed nine OCPs in blood and FF and their association with assisted reproductive technologies (ART) outcomes. HCB in the blood was found to be significantly associated with lower AMH levels, negatively correlated with clinical pregnancy and live birth rates. However, the Study of Metals and Assisted

Reproductive Technologies (SMART) (176), found that women with higher FF DDT concentrations had higher average E2 peak levels compared to women with lower DDT concentrations (1.45; 95% CI 0.92, 1.97; $p < 0.0001$). A study investigating the impact of serum OCPs on AMH levels in women from a South African village reported no evidence of an association between AMH levels and DDT exposure (177).

Summary: The three studies showed inconsistent results regarding the effect of OCP exposure on ovarian function. Due to the inconsistencies in the results, small sample sizes, regional limitations, and the overall low quality of evidence, no definitive conclusions can be drawn at this time.

Polychlorinated biphenyls

Two prospective studies investigated the effect of PCB exposure on ovarian function, involving a total of 217 women. Both studies were rated as “Medium” quality by the NOS, with a GRADE evidence quality rating of very low.

The SMART study (176) also analyzed the impact of PCBs on IVF outcomes in women, finding that women with high concentrations of PCB-151, PCB-170 and PCB-180 in FF had 47, 32 and 32% fewer baseline antral follicles, respectively, compared to women with low levels ($p < 0.05$). Additionally, women with high FF levels of PCB-138 and PCB-153 had E2 values 35 and 41% lower than those with low levels ($p < 0.05$). The study further revealed a significant association between PCB exposure and reduced embryo implantation rates. Conversely, Björvang et al. (165) found that PCBs exposure increased the number of antral follicles and enhanced ovarian responsiveness, but resulted in a decline in embryo quality.

Summary: The evidence suggests that PCBs negatively impact ovarian function, but the evidence quality from existing prospective studies is very low. Although high concentrations of PCB-151, PCB-170, and PCB-180 were associated with fewer antral follicles, methodological limitations and small sample sizes necessitate cautious interpretation of these findings.

Bisphenol A

Three prospective studies investigated the effect of BPA exposure on ovarian function, involving a total of 274 women. Of these, two were rated as “High” quality and one as “Medium” by the NOS, with a GRADE evidence quality rating of very low.

Mok-Lin et al. (178) found a positive correlation between serum E2 levels and total oocyte count ($p < 0.001$). However, each log unit increase in specific gravity-BPA was associated with a 12% decrease in oocyte count ($p = 0.007$) and a 213 pg/mL reduction in E2 peak ($p = 0.03$). These findings are consistent with the SMART study (179), which found a negative correlation between BPA and peak E2 levels ($p = 0.06$), although BPA was not associated with AFC or FSH levels. In contrast, a study from South Korea (180) did not find any significant effect of BPA on E2 levels or embryo quality.

Summary: Mok-Lin et al. and the SMART study observed negative associations between BPA levels and both oocyte count and E2 peak levels, although these findings were not consistent across all markers, as BPA was not linked to AFC or FSH levels. Additionally, the South Korean study found no impact of BPA on E2 levels or embryo quality. Therefore, while some evidence suggests that BPA may negatively impact ovarian function, the overall evidence is not entirely consistent and may be influenced by various factors.

Comprehensive analysis indicates that BPA may have adverse effects on ovarian function; however, the quality of current evidence is low. This conclusion is primarily constrained by the overall insufficient sample size in the studies, which subsequently leads to a lower rating in the GRADE evidence quality evaluation system.

Parabens

Only one study reported on the impact of parabens exposure on ovarian function in a cohort of 192 women, which was rated as “High” quality by the NOS, with a GRADE evidence quality rating of very low.

A report from the EARTH study (181) analyzed the association between urinary parabens concentrations and ovarian reserve markers. Methylparaben (MP) and propylparaben (PP) were detected in over 99% of urine samples. Researchers observed a decreasing trend in AFC in women with higher PP concentrations ($p = 0.07$), and an increasing trend in day-3 FSH levels, consistent with the negative correlation between FSH and AFC ($p = 0.002$).

Summary: This study indicated that high urinary PP concentrations were associated with a decreasing trend in AFC and an increasing trend in FSH levels, although the association did not reach conventional significance levels. This finding suggests the potential adverse impact of PP on ovarian reserve function.

Dioxins

Two studies analyzed the effect of dioxins exposure on ovarian function in women, one prospective and one retrospective with a total of 472 participants. The prospective study was rated as “High” quality and the retrospective study as “Medium” by the NOS, with both studies receiving a GRADE evidence quality rating of very low.

One study analyzed the relationship between ovarian function and serum levels of TCDD in women who had been exposed to high concentrations of TCDD 20 years earlier (182). The other study investigated the impact of TCDD by analyzing steroid hormone levels in saliva and serum from women in high-exposure and low-exposure regions (183). However, neither study found conclusive evidence of a significant effect of TCDD on ovarian function.

Summary: The first study suggested a potential impact of TCDD on female reproductive health, while the second study analyzed the relationship between steroid hormone levels and TCDD exposure, but could not conclusively demonstrate a direct effect on ovarian function. Although no definitive conclusions were drawn, these studies provide important clues for understanding dioxins’ potential risks to the human endocrine system and reproductive health.

Triclosan

Two prospective studies investigated the effect of TCS exposure on ovarian function, involving a total of 620 women. One study was rated as “High” quality by the NOS, and the other as “Medium,” with a GRADE evidence quality rating of very low.

Jurewicz et al. (184) explored that urinary TCS concentrations significantly reduced AFC ($p = 0.03$), but no associations were observed between TCS exposure and other parameters such as E2, FSH, and AMH levels. Additionally, the EARTH study (185) found a negative correlation between specific gravity-adjusted urinary TCS concentration and AFC ($p = 0.04$) with younger (<35 years) and leaner ($\text{BMI} < 25 \text{ kg/m}^2$) women experienced a greater decline in AFC.

Summary: Both prospective studies indicated that TCS exposure may have adverse effects on ovarian function, as shown by the

negatively correlation between urinary TCS concentrations and ovarian function parameters. These findings provide valuable insights into this area of research.

Air pollutants and ovarian function

Particulate matter

Fourteen articles analyzed the impact of PM (PM_{2.5} and PM₁₀) exposure on ovarian function in women, derived from thirteen different studies. These studies included 54,348 participants for PM_{2.5} analyses and 45,410 participants for PM₁₀ evaluations. Ten of the studies were rated as “High” quality by the NOS, while three were rated as “Medium.” The GRADE evidence quality for PM_{2.5} was rated as high, whereas for PM₁₀ it was rated as low.

Seven studies (186–193) consistently found a significant association between PM_{2.5} or PM₁₀ exposure and decreased AFC and AMH levels. The negative impact of PM_{2.5} on ovarian reserve was equivalent to aging by approximately two years (186). In women diagnosed with infertility, the impact of PM₁₀ on AFC was particularly pronounced (188). Research teams led by Wang (192) and Pang (190) highlighted that the impact of PM_{2.5} on AMH was especially prominent during the early transition from primary follicles to preantral follicles. However, there were divergent findings regarding the age-dependent impact of PM_{2.5}: La Marca et al. (193) observed that the negative effect of PM_{2.5} on AMH diminished with age; Xinyan Wang et al. (192) demonstrated that stronger inverse associations were observed in women <35 years; Lanlan Fang et al. found that after age stratification, PM_{2.5}/PM₁₀ exposure had significantly stronger negative effects on E2 and FSH levels in older women (>30 years) ($p < 0.05$) (194); whereas Wiecek (188) study suggested that older women (>35 years) were more affected by PM_{2.5}, with more pronounced declines in AFC and AMH.

One study found a monotonic decrease in FSH levels with PM_{2.5} and PM₁₀ exposure, along with a “U”-shaped exposure-response curve for E2 levels (194). Another study reported that women with higher PM₁₀ exposure levels in the early luteal phase had higher E2 levels ($p = 0.02$) (195). Nevertheless, four studies (196–199) found no significant association between PM_{2.5} or PM₁₀ exposure and ovarian reserve markers. For example, LaPointe et al. (197) did not observe any correlation between PM_{2.5} and AFC, but noted that high PM_{2.5} exposure led to reduced total oocyte counts, mature/metaphase II oocyte numbers, and ovarian sensitivity index.

Summary: PM_{2.5} exposure appeared to have a broader and more profound impact on ovarian reserve function, affecting not only AMH and AFC but also oocyte quantity and quality, while PM₁₀ effects seemed to be more specific, notably impacting women with infertility and affecting E2 levels.

Gaseous pollutants

SO_x, NO_x, O₃, carbon monoxide (CO), BTEX, and tobacco smoke are common gaseous pollutants, with SO_x and NO_x most commonly appearing in the form of SO₂ and NO₂.

Thirteen studies analyzed the effect of gaseous pollutants exposure on ovarian function in women, each contributing one article. The analyses included 29,243 participants for SO_x, 50,939 for NO_x, 48,882 for O₃, 23,720 for CO, 806 for BTEX, and 132 for tobacco smoke exposure. The NOS quality ratings showed that eight studies were

rated as “High” and five as “Medium.” The GRADE quality ratings and specific outcomes for the impact of various gaseous pollutants on ovarian function are as follows:

Moderate-quality evidence (188, 191, 194, 199) suggests that SO_x has a predominantly negative correlation with ovarian markers such as AFC, AMH, and FSH, with stronger negative associations observed in older women (>30 years) (194), although some studies (189, 198) did not find a significant association between SO₂ and AMH. Very low-quality evidence (188–191, 193, 196–199) shows that NO_x have little effect on AMH and AFC, with only one study (194) indicating a positive correlation between NO₂ and E2, FSH. Very low-quality evidence (189, 190, 194, 199) indicates that O₃ generally shows no association with ovarian reserve markers, although a few studies reported a negative correlation between O₃ and AMH (192, 200), and a positive correlation with FSH and E2 (194). Very low quality evidence (189, 197, 199) did not find a significant association between CO and ovarian reserve, except for one study suggesting a negative correlation between CO and FSH, E2 (194). Very low-quality evidence (198) for BTEX suggests no significant association with AMH levels, while very low quality evidence (195) for tobacco smoke indicates a significant association with increased E2 levels.

Summary: Findings reveal considerable variability in the impact of different gaseous pollutants on ovarian function in women. SO_x generally shows a negative correlation with ovarian function markers, while the effects of NO_x, O₃, and CO are inconclusive. Tobacco smoke may be associated with increased E2 levels.

Heavy metals and ovarian function

Due to the limited availability of cohort studies on the impact of heavy metals on ovarian function, this section provides an overview without detailed subdivision. Three independent studies (reported in four articles) were identified examining Cd, Hg, and Pb exposure. These analyses included 810 participants for Hg exposure, 525 for Cd exposure, and 525 for Pb exposure. The NOS quality ratings showed that two studies were rated as “High” and one as “Medium.” The GRADE quality ratings and specific outcomes for their impact on ovarian function are as follows:

Moderate-quality evidence suggests mixed findings regarding the impact of Hg on ovarian reserve, with one out of three studies showed a positive correlation between Hg and AFC (201), while the other two did not find a significant association (202, 203). Very low-quality evidence (203, 204) suggested a positive correlation between Cd and AMH and E2 levels. Another study with very low-quality evidence (203) found no significant correlation between Pb and ovarian markers. The study by Mínguez-Alarcón et al. (201) a positive correlation was observed between hair Hg and AFC, specifically among women consumed more than 0.125% of their total caloric intake from n-3 polyunsaturated fatty acids (n3PUFA) weekly. In this study higher Hg tertiles were associated with an increasing trend in AFC ($p = 0.004$), it is possible that the higher exposure group consumed more fish, which may reflect the positive effect of n3PUFA in fish rather than the impact of the heavy metal itself (201–203). In addition, the research by Kim (204) found that higher blood Cd concentrations led to increased testosterone and AMH levels, with each 0.1 µg/L rise in blood Cd increasing the probability of a mild

polycystic ovarian syndrome (PCOS) phenotype by 18% (RR 1.18; 95% CI 1.06, 1.31).

Summary: The impact of heavy metals on ovarian function is a complex and relatively understudied area. Existing evidence on the direct association between heavy metals and ovarian function remains contentious and may be influenced by other factors, such as n3PUFA intake, making it difficult to clearly delineate the effect of heavy metals on ovarian function. Therefore, more high-quality studies are needed to explore this area in depth.

Discussion

This review synthesizes findings from 33 cohort studies, covering twenty different categories of environmental pollutants. These pollutants, widely present in various consumer products, were explored their associations with ovarian function, with GRADE assessments indicating that PFAS, PAEs, TCS, PM, and SO_x have the clearest evidence of impact on ovarian reserve. Specific manifestations include: PFAS significantly reduces AMH levels and suppresses E2 secretion; PAEs correlate with diminished AMH and AFC; elevated TCS concentrations are linked to reduced AFC; PM2.5/PM10 exposure leads to declines in AFC and AMH; while SO_x broadly disrupts ovarian biomarkers (AFC, AMH, FSH, [Supplementary Table 7](#) summarizes and correlates the effects of these pollutants on ovarian function with their potential mechanisms of action). However, due to the limited number of studies and the heterogeneity in study designs, the impact of other pollutants remains inconclusive.

Chemical pollutants are particularly relevant due to their diverse forms and frequent contact in daily life, especially for women, who may be more exposed to personal care products than men. The negative impact of these chemical pollutants on the ovaries is substantial. They not only disrupt normal ovarian hormone secretion but also impede proper oocyte development. Multiple studies have indicated that various pollutants like PAEs (such as DEHP, MiBP, and MnBP) ([171](#), [173](#)), PCBs (specifically PCB-151, PCB-170, PCB-180) ([176](#)), parabens (PP) ([181](#)), and TCS ([184](#), [185](#)) adversely affect ovarian reserve by inhibiting antral follicle growth, causing follicular cell cycle arrest and increasing follicle atresia. This impact is especially pronounced in younger women (<35 years) ([185](#), [188](#)), aligning with findings ([181](#)) that elevated urinary PP levels are associated with increased day-3 FSH levels and reduced AFC, which closely resembles clinical features of premature ovarian insufficiency ([205](#)). Although some PAEs metabolites (such as MEOHP and Σ PAEs) ([174](#)), PFAS ([165](#)), and certain PCBs (PCB-118) ([165](#)) may increase AFC. This likely reflects accelerated recruitment of primordial follicles, potentially promoting early follicle depletion. Animal studies ([174](#), [206](#)) support this view as well: exposure to di-n-butyl phthalate (DBP), the precursor of MBP, promotes the depletion of follicular follicles by accelerating primordial follicle recruitment in rats. These observations suggest that a higher baseline AFC in exposed individuals does not correlate with improved fertility outcomes, as pollutants also lower embryo quality during IVF ([165](#)), revealing the complex and far-reaching impact of these pollutants on ovarian health. Moreover, these pollutants exhibit estrogenic effects, can bind to estrogen receptors (ER) alpha and ER beta, thereby influencing the expression of endogenous estrogen to some extent ([207](#)).

Among air pollutants, PM2.5 is the most clearly harmful to ovarian function, followed by PM10, both of which directly affect

ovarian reserve. Due to it is smaller in diameter, PM2.5 is more easily to penetrate the alveolar-capillary barrier and reaches the ovaries. The organic matter, soluble metals, and other substances contained within PM2.5 can interfere with ovarian function ([208](#), [209](#)), with polycyclic aromatic hydrocarbons (PAHs) being particularly representative ([195](#)). PAHs exposure can lead to accelerated loss of primordial follicles, resulting in premature ovarian reserve depletion ([210](#)). The effects of PAHs exposure may also extend to subsequent generations, leading to reduced ovarian reserve.

Research on heavy metals and ovarian function is limited, but numerous animal studies have emphasized the reproductive toxicity of heavy metals ([211](#)) like Cd, Hg and Pb, which do not metabolize and are toxic at any concentration ([212](#)). Humans are primarily exposed through contaminated water, seafood, and smoking ([212](#)). However, individuals with high exposure tend to consume more fish ([201–203](#)), and the positive effects of n3PUFA in fish on ovarian reserve may mask the adverse effects of heavy metals. Therefore, future research needs to employ more rigorous experimental designs to investigate the impact of heavy metals on ovarian function.

Limitations and strengths of the study

Study design limitations

We systematically integrated comprehensive reports from 33 cohort studies, most of which ($N = 20$) that remained after screening were infertile patients, potentially limiting generalizability to the broader population. On the other hand, this highlights the potential correlation between environmental pollutants and infertility, warranting further research ([213](#), [214](#)). Additionally, the observed contradictions in the association between environmental pollutant exposure and ovarian function across studies may be attributed to several factors. First, the magnitude of exposure (low vs. high) and duration of exposure may influence outcomes, as populations exposed to high levels of environmental pollutants (e.g., those living near industrial areas) may experience different health effects than those with background exposure ([215](#), [216](#)). Second, methodological variations among studies may lead to inconsistent results. These include differences in study design (e.g., cross-sectional vs. longitudinal), sample size, methods for measuring environmental pollutants (e.g., blood serum, follicular fluid, urine), and statistical approaches (e.g., adjusting for different confounders). Future studies would benefit from standardized exposure assessment protocols, such as harmonized biomonitoring matrices (e.g., urine vs. serum) and consistent quantification methods. Moreover, the timing of sample collection (e.g., follicular phase vs. luteal phase of the menstrual cycle) can significantly impact hormone measurements ([195](#)), potentially masking or exaggerating associations ([217](#), [218](#)). Third, exposure variations may contribute to these inconsistencies. Different types of environmental pollutants have distinct chemical properties, metabolic pathways, and biological effects, which may result in varying impacts on ovarian function. Furthermore, differences in study populations could play a critical role. The age, BMI, lifestyle factors (e.g., smoking, alcohol consumption), and reproductive history (nulliparous vs. parous) may modify the effects of environmental pollutants on ovarian health. For example, age differences further influenced outcomes ([200](#), [219](#)), as ovarian decline in women aged ≥ 35 years may obscure pollutant effects, particularly with chemical pollutants and PM ([190](#), [191](#), [220](#)). Notably, for certain compounds (e.g., BPs and

PCBs), the limited number of related studies resulted in a weak evidence base, making it challenging to comprehensively assess their effects on ovarian function. To address these limitations, we recommend prioritizing longitudinal cohort studies with repeated exposure measurements and multi-country collaborative designs to account for geographic variability in pollutant profiles. By recognizing and analyzing these factors, we can better understand the complexity of the relationship between environmental pollutants and ovarian function and identify areas for future research.

Strengths of the study

Compared with previous research, this review has several significant strengths. Firstly, it included pollutants from various exposure pathways such as air, water, and soil, enhancing the overall understanding of the impact of environmental pollutants on ovarian function. Secondly, this study covered all relevant articles from the establishment of the database to the present day, and multiple geographical regions, reflecting differences in exposure across locations. The multidimensional analysis of ovarian function evaluation indicators (e.g., AMH, AFC, FSH, E2) further enhanced understanding of the impact of pollutants on ovarian reserve and function. Lastly, this review systematically assessed the quality of the included studies used the NOS and the GRADE framework, increasing the robustness and reliability of the conclusions.

Policy and practice recommendations

Based on the study findings, it is recommended that public health regulations be strengthened concerning certain chemical pollutants (e.g., PCBs, PFAS), particularly to protect pregnant and reproductive-age women (221–223). Existing frameworks such as the EU REACH regulations (224, 225) and US EPA PFAS Action Plan (226) have demonstrated effectiveness in reducing exposure to legacy pollutants through restrictions on production and environmental discharge. However, we identifies emerging contaminants (e.g., BPA) that currently fall outside these regulatory frameworks despite evidence of ovarian toxicity (227). Policies should prioritize not only reducing exposure to individual pollutants but also investigating the combined and cumulative effects of multiple contaminants (228), as mandated by the 2021 EU Chemicals Strategy for Sustainability (229). Public education initiatives must be strengthened to encourage lifestyle and dietary habits that minimize pollutant exposure, modeled after successful programs like the US National Biomonitoring Program's community outreach (230). Enhancing dynamic biomarker monitoring can further improve the accuracy of long-term exposure assessments.

Clinically, these findings emphasize the need for proactive measures to protect reproductive-age women from environmental pollutants. Regular ovarian health assessments, tailored guidance, and advanced technologies such as AI-driven exposure tools (231) are crucial for identifying high-risk individuals and implementing targeted prevention strategies. Practical steps, including avoiding plastic products, reducing air pollution exposure, and selecting uncontaminated food, can significantly lower risks (232). Physicians should educate patients, particularly women in reproductive age, planning pregnancy or women at higher risk due to occupational

and environmental factors, on minimizing pollutant exposure and leveraging early screening methods for ovarian dysfunction (233).

For researchers, future research should expand sample sizes to include women of different ages and reproductive states to clarify the long-term impact of these pollutants on ovarian reserve, incorporating longer follow-up periods and more precise exposure assessments. More attention should also be given to research on male exposure to gain a comprehensive understanding of the impact of environmental pollutants on couple fertility (234). Rigorous study designs, especially for heavy metals, should control for confounders like fish consumption. This review also highlights gaps in the literature regarding the effects of some pollutants on ovarian function, including the long-term effects of pollutants with long half-lives and the mechanisms of pollutant impact on ovarian function, which urgently need more high-quality research to provide policy-relevant evidence, particularly for regulatory agencies evaluating chemical prioritization under programs like the US EPA's Toxic Substances Control Act (235).

Conclusion

This systematic review highlights the substantial impact of environmental pollutants on ovarian function, emphasizing the role of chemical pollutants (e.g., PFAS, PAEs, PCBs), air pollutants (e.g., PM2.5, PM10), and heavy metals (e.g., Pb, Cd) in reducing ovarian reserve and impairing hormonal function. Women of reproductive age, particularly those exposed to high levels of these pollutants, face increased risks of infertility.

The review reveals significant gaps in current research, particularly in understanding the cumulative and synergistic effects of multiple pollutants and their long-term health outcomes. These findings advocate for stricter public health policies, enhanced education on minimizing exposure, and advanced monitoring technologies to protect reproductive health. For clinical practice, regular ovarian health assessments and personalized advice on reducing pollutant exposure are essential to mitigate risks. Future research should prioritize rigorous, large-scale studies exploring the molecular mechanisms of pollutant-induced ovarian dysfunction to develop effective preventive and therapeutic strategies.

Data availability statement

The original contributions presented in the study are included in the article/[Supplementary material](#), further inquiries can be directed to the corresponding authors.

Author contributions

LA: Data curation, Formal analysis, Investigation, Resources, Software, Validation, Visualization, Writing – original draft. YH: Data curation, Formal analysis, Investigation, Resources, Software, Validation, Writing – original draft. YW: Data curation, Formal analysis, Investigation, Resources, Software, Validation, Writing – original draft. SS: Data curation, Formal analysis, Investigation, Resources, Validation, Writing – review & editing. XXL: Formal analysis, Methodology,

Software, Supervision, Visualization, Writing – review & editing. XYL: Methodology, Project administration, Supervision, Validation, Writing – review & editing. LL: Methodology, Project administration, Supervision, Validation, Writing – review & editing. CT: Methodology, Project administration, Supervision, Validation, Writing – review & editing. JL: Data curation, Formal analysis, Methodology, Writing – review & editing. TS: Data curation, Formal analysis, Methodology, Writing – review & editing. MZ: Data curation, Formal analysis, Writing – review & editing. DW: Data curation, Investigation, Project administration, Writing – review & editing. JW: Software, Visualization, Writing – review & editing, Methodology, Data curation, Project administration, Conceptualization. XL: Supervision, Writing – review & editing, Conceptualization, Data curation, Funding acquisition, Formal analysis, Writing – original draft. YL: Writing – review & editing, Conceptualization, Funding acquisition, Supervision, Methodology, Resources, Project administration.

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References

- Landrigan PJ, Fuller R, Acosta NJR, Adeyi O, Arnold R, Basu NN, et al. The lancet commission on pollution and health. *Lancet*. (2018) 391:462–512.
- Lelieveld J, Evans JS, Fnais M, Giannadaki D, Pozzer A. The contribution of outdoor air pollution sources to premature mortality on a global scale. *Nature*. (2015) 525:367–71. doi: 10.1038/nature15371
- Guxens M, Garcia-Esteban R, Giorgis-Allemand L, Fornis J, Badaloni C, Ballester F, et al. Air pollution during pregnancy and childhood cognitive and psychomotor development: six European birth cohorts. *Epidemiology*. (2014) 25:636–47. doi: 10.1097/EDE.0000000000000133
- Wang M, Aaron CP, Madrigano J, Hoffman EA, Angelini E, Yang J, et al. Association between long-term exposure to ambient air pollution and change in quantitatively assessed emphysema and lung function. *JAMA*. (2019) 322:546–56. doi: 10.1001/jama.2019.10255
- Maitre A, Bonnetterre V, Huillard L, Sabatier P, de Gaudemaris R. Impact of urban atmospheric pollution on coronary disease. *Eur Heart J*. (2006) 27:2275–84. doi: 10.1093/eurheartj/ehl162
- Boogaard H, Patton AP, Atkinson RW, Brook JR, Chang HH, Crouse DL, et al. Long-term exposure to traffic-related air pollution and selected health outcomes: A systematic review and meta-analysis. *Environ Int*. (2022) 164:107262. doi: 10.1016/j.envint.2022.107262
- Cohen AJ, Brauer M, Burnett R, Anderson HR, Frostad J, Estep K, et al. Estimates and 25-year trends of the global burden of disease attributable to ambient air pollution: an analysis of data from the global burden of diseases study 2015. *Lancet*. (2017) 389:1907–18. doi: 10.1016/S0140-6736(17)30505-6
- Weuve J, Bennett EE, Ranker L, Gianattasio KZ, Pedde M, Adar SD, et al. Exposure to air pollution in relation to risk of dementia and related outcomes: An updated systematic review of the epidemiological literature. *Environ Health Perspect*. (2021) 129:96001. doi: 10.1289/EHP8716
- Brook RD, Rajagopalan S, Pope CA, Brook JR, Bhatnagar A, Diez-Roux AV, et al. Particulate matter air pollution and cardiovascular disease: an update to the scientific statement from the American Heart Association. *Circulation*. (2010) 121:2331–78. doi: 10.1161/CIR.0b013e3181dbec1
- Holme SAN, Sigsgaard T, Holme JA, Holst GJ. Effects of particulate matter on atherosclerosis: a link via high-density lipoprotein (HDL) functionality? *Part Fibre Toxicol*. (2020) 17:36. doi: 10.1186/s12989-020-00367-x
- Kelly FJ, Fussell JC. Linking ambient particulate matter pollution effects with oxidative biology and immune responses. *Ann N Y Acad Sci*. (2015) 1340:84–94. doi: 10.1111/nyas.12720
- Fruh V, Claus Henn B, Weuve J, Wesselink AK, Orta OR, Heeren T, et al. Incidence of uterine leiomyoma in relation to urinary concentrations of phthalate and phthalate alternative biomarkers: A prospective ultrasound study. *Environ Int*. (2021) 147:106218. doi: 10.1016/j.envint.2020.106218
- Meeker JD, Sathyanarayana S, Swan SH. Phthalates and other additives in plastics: human exposure and associated health outcomes. *Philos Trans R Soc Lond Ser B Biol Sci*. (2009) 364:2097–113. doi: 10.1098/rstb.2008.0268
- vom Saal FS, Hughes C. An extensive new literature concerning low-dose effects of bisphenol A shows the need for a new risk assessment. *Environ Health Perspect*. (2005) 113:926–33. doi: 10.1289/ehp.7713
- Diamanti-Kandaraki E, Bourguignon J-P, Giudice LC, Hauser R, Prins GS, Soto AM, et al. Endocrine-disrupting chemicals: an Endocrine Society scientific statement. *Endocr Rev*. (2009) 30:293–342. doi: 10.1210/er.2009-0002
- Young SS, Yu M. Association of bisphenol A with diabetes and other abnormalities. *JAMA*. (2009) 301:721–722.
- Ahern TP, Broe A, Lash TL, Cronin-Fenton DP, Ulrichsen SP, Christiansen PM, et al. Phthalate exposure and breast Cancer incidence: A Danish Nationwide cohort study. *J Clin Oncol*. (2019) 37:1800–9. doi: 10.1200/JCO.18.02202
- Zhang X, Guo N, Jin H, Liu R, Zhang Z, Cheng C, et al. Bisphenol A drives di (2-ethylhexyl) phthalate promoting thyroid tumorigenesis via regulating HDAC6/PTEN and c-MYC signaling. *J Hazard Mater*. (2022) 425:127911. doi: 10.1016/j.jhazmat.2021.127911
- Carpenter DO. Health effects of persistent organic pollutants: the challenge for the Pacific Basin and for the world. *Rev Environ Health*. (2011) 26:61–9. doi: 10.1515/reveh.2011.009
- Joffin N, Noirez P, Antignac JP, Kim MJ, Marchand P, Falabregue M, et al. Release and toxicity of adipose tissue-stored TCDD: direct evidence from a xenografted fat model. *Environ Int*. (2018) 121:1113–20. doi: 10.1016/j.envint.2018.10.027
- Nicholson JK, Kendall MD, Osborn D. Cadmium and mercury nephrotoxicity. *Nature*. (1983) 304:633–5. doi: 10.1038/304633a0
- Su F, Zeeshan M, Xiong LH, Lv JY, Wu Y, Tang XJ, et al. Co-exposure to perfluoroalkyl acids and heavy metals mixtures associated with impaired kidney

Conflict of interest

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Supplementary material

The Supplementary material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpubh.2025.1575418/full#supplementary-material>

function in adults: A community-based population study in China. *Sci Total Environ.* (2022) 839:156299. doi: 10.1016/j.scitotenv.2022.156299

23. Paithankar JG, Saini S, Dwivedi S, Sharma A, Chowdhuri DK. 103 heavy metal associated health hazards: An interplay of oxidative stress and signal transduction. *Chemosphere.* (2021) 262:128350. doi: 10.1016/j.chemosphere.2020.128350

24. Beier EE, Maher JR, Sheu TJ, Cory-Slechta DA, Berger AJ, Zuscik MJ, et al. Heavy metal lead exposure, osteoporotic-like phenotype in an animal model, and depression of Wnt signaling. *Environ Health Perspect.* (2013) 121:97–104. doi: 10.1289/ehp.1205374

25. Zhou S, Xi Y, Chen Y, Zhang Z, Wu C, Yan W, et al. Ovarian dysfunction induced by chronic whole-body PM2.5 exposure. *Small.* (2020) 16:e2000845. doi: 10.1002/sml.202000845

26. Pan W, Ye X, Yin S, Ma X, Li C, Zhou J, et al. Selected persistent organic pollutants associated with the risk of primary ovarian insufficiency in women. *Environ Int.* (2019) 129:51–8. doi: 10.1016/j.envint.2019.05.023

27. Hannon PR, Niermann S, Flaws JA. Acute exposure to Di (2-Ethylhexyl) phthalate in adulthood causes adverse reproductive outcomes later in life and accelerates reproductive aging in female mice. *Toxicological sciences: an official journal of the society of Toxicology.* (2016) 150:97–108.

28. Jones ASK, Hannum DF, Machlin JH, Tan A, Ma Q, Ulrich ND, et al. Cellular atlas of the human ovary using morphologically guided spatial transcriptomics and single-cell sequencing. *Sci Adv.* (2024) 10:eadm7506. doi: 10.1126/sciadv.adm7506

29. Testing and interpreting measures of ovarian reserve: a committee opinion. *Fertil Steril.* (2020) 114:1151–7.

30. Svechnikov K, Söder O. Ontogeny of gonadal sex steroids. *Best Pract Res Clin Endocrinol Metab.* (2008) 22:95–106. doi: 10.1016/j.beem.2007.09.002

31. Zlotorynski E. Long-lived proteomes in healthy ovaries. *Nat Rev Mol Cell Biol.* (2024) 25:596. doi: 10.1038/s41580-024-00764-6

32. Cedars MI. Evaluation of female fertility-AMH and ovarian reserve testing. *J Clin Endocrinol Metab.* (2022) 107:1510–9. doi: 10.1210/clinem/dgac039

33. Metka M, Holzer G, Heytmanek G, Huber J. Hypergonadotropic hypogonadic amenorrhea (World Health Organization III) and osteoporosis. *Fertil Steril.* (1992) 57:37–41. doi: 10.1016/s0015-0282(16)54773-6

34. Kalantaridou SN, Naka KK, Bechlioulis A, Makrigiannakis A, Michalis L, Chrousos GP. Premature ovarian failure, endothelial dysfunction and estrogen-progesterone replacement. *Trends Endocrinol Metab.* (2006) 17:101–9. doi: 10.1016/j.tem.2006.02.003

35. Schover LR. Premature ovarian failure and its consequences: vasomotor symptoms, sexuality, and fertility. *J Clin Oncol.* (2008) 26:753–8. doi: 10.1200/JCO.2007.14.1655

36. Ding T, Yan W, Zhou T, Shen W, Wang T, Li M, et al. Endocrine disrupting chemicals impact on ovarian aging: evidence from epidemiological and experimental evidence. *Environ Pollut.* (2022) 305:119269. doi: 10.1016/j.envpol.2022.119269

37. Ruiz-Castell M, Le Coroller G, Pexaras A, Ciprián GM, Fagherazzi G, Bohn T, et al. Characterizing the adult exposure in men and women from the general population: results from the EHES-LUX study. *Environ Int.* (2023) 173:107780. doi: 10.1016/j.envint.2023.107780

38. Dien NT, Hirai Y, Koshida J, Sakai SI. Factors affecting multiple persistent organic pollutant concentrations in the air above Japan: A panel data analysis. *Chemosphere.* (2021) 277:130356. doi: 10.1016/j.chemosphere.2021.130356

39. Ruyle BJ, Thackray CP, Butt CM, LeBlanc DR, Tokranov AK, Vecitis CD, et al. Centennial persistence of forever chemicals at military fire training sites. *Environ Sci Technol.* (2023) 57:8096–106. doi: 10.1021/acs.est.3c00675

40. Yu K, Zhang X, Tan X, Ji M, Chen Y, Wan Z, et al. Multigenerational and transgenerational effects of 2, 3, 7, 8-tetrachlorodibenzo-p-dioxin exposure on ovarian reserve and follicular development through AMH/AMHR2 pathway in adult female rats. *Food Chem Toxicol.* (2020) 140:111309. doi: 10.1016/j.fct.2020.111309

41. Levine L, Hall JE. Does the environment affect menopause? A review of the effects of endocrine disrupting chemicals on menopause. *Climacteric.* (2023) 26:206–15. doi: 10.1080/13697137.2023.2173570

42. Bjerregaard-Olesen C, Ghisari M, Bonefeld-Jørgensen EC. Activation of the estrogen receptor by human serum extracts containing mixtures of perfluorinated alkyl acids from pregnant women. *Environ Res.* (2016) 151:71–9. doi: 10.1016/j.envres.2016.07.001

43. Li Y, Xiao N, Liu M, Liu Y, He A, Wang L, et al. Dysregulation of steroid metabolome in follicular fluid links phthalate exposure to diminished ovarian reserve of childbearing-age women. *Environmental Pollution.* (2023) 330:121730. doi: 10.1016/j.envpol.2023.121730

44. Artacho-Córdón F, Ríos-Arrabal S, León J, Frederiksen H, Sáenz JM, Martín-Olmedo P, et al. Adipose tissue concentrations of non-persistent environmental phenols and local redox balance in adults from southern Spain. *Environ Int.* (2019) 133:105118. doi: 10.1016/j.envint.2019.105118

45. Johns LE, Cooper GS, Galizia A, Meeker JD. Exposure assessment issues in epidemiology studies of phthalates. *Environ Int.* (2015) 85:27–39. doi: 10.1016/j.envint.2015.08.005

46. Wong KH, Durrani TS. Exposures to endocrine disrupting Chemicals in Consumer Products-A Guide for pediatricians. *Curr Probl Pediatr Adolesc Health Care.* (2017) 47:107–18. doi: 10.1016/j.cped.2017.04.002

47. Tagne-Fotso R, Riou M, Saoudi A, Zeghnoun A, Frederiksen H, Berman T, et al. Exposure to bisphenol A in European women from 2007 to 2014 using human biomonitoring data - the European joint Programme HBM4EU. *Environ Int.* (2024) 190:108912. doi: 10.1016/j.envint.2024.108912

48. Weng X, Zhu Q, Liao C, Jiang G. Cumulative exposure to phthalates and their alternatives and associated female reproductive health: body burdens, adverse outcomes, and underlying mechanisms. *Environ Sci Technol.* (2023) 57:8189–212. doi: 10.1021/acs.est.3c00823

49. Zhang N, Zhao Y, Zhai L, Bai Y, Wei W, Sun Q, et al. Urinary concentrations of bisphenol A and its alternatives: potential predictors of and associations with antral follicle count among women from an infertility clinic in northern China. *Environ Res.* (2024) 249:118433. doi: 10.1016/j.envres.2024.118433

50. Luderer U, Lim J, Ortiz L, Nguyen JD, Shin JH, Allen BD, et al. Exposure to environmentally relevant concentrations of ambient fine particulate matter (PM2.5) depletes the ovarian follicle reserve and causes sex-dependent cardiovascular changes in apolipoprotein E null mice. *Part Fibre Toxicol.* (2022) 19:5. doi: 10.1186/s12989-021-00445-8

51. Gai H-F, An J-X, Qian X-Y, Wei Y-J, Williams JP, Gao G-L. Ovarian damages produced by aerosolized fine particulate matter (PM2.5) pollution in mice: possible protective medications and mechanisms. *Chin Med J.* (2017) 130:1400–10. doi: 10.4103/0366-6999.207472

52. Conforti A, Mascia M, Cioffi G, De Angelis C, Coppola G, De Rosa P, et al. Air pollution and female fertility: a systematic review of literature. *Reprod Biol Endocrinol.* (2018) 16:117. doi: 10.1186/s12958-018-0433-z

53. Hwang S, Hood RB, Hauser R, Schwartz J, Laden F, Jones D, et al. Using follicular fluid metabolomics to investigate the association between air pollution and oocyte quality. *Environ Int.* (2022) 169:107552. doi: 10.1016/j.envint.2022.107552

54. Zhang X, Wang X, Li H, Wang H, Du D, Huang H. ATF3 mediates PM2.5-induced apoptosis and inflammation in ovarian granulosa cells. *J Ovarian Res.* (2024) 17:215. doi: 10.1186/s13048-024-01539-5

55. Qu J, Niu H, Wang J, Wang Q, Li Y. Potential mechanism of lead poisoning to the growth and development of ovarian follicle. *Toxicology.* (2021) 457:152810. doi: 10.1016/j.tox.2021.152810

56. Qu J, Wang Q, Sun X, Li Y. The environment and female reproduction: potential mechanism of cadmium poisoning to the growth and development of ovarian follicle. *Ecotoxicol Environ Saf.* (2022) 244:114029. doi: 10.1016/j.ecoenv.2022.114029

57. Lv B, Peng Y, Peng Y-d, Wang Z, Song Q-S. Integrated transcriptomics and proteomics provide new insights into the cadmium-induced ovarian toxicity on *Pardosa pseudoannulata*. *Chemosphere.* (2022) 297:134255. doi: 10.1016/j.chemosphere.2022.134255

58. Johansson HKL, Svingen T, Fowler PA, Vinggaard AM, Boberg J. Environmental influences on ovarian dysgenesis - developmental windows sensitive to chemical exposures. *Nat Rev Endocrinol.* (2017) 13:400–14. doi: 10.1038/nrendo.2017.36

59. George AJ, Birnbaum LS. Dioxins vs. PFAS: science and policy challenges. *Environ Health Perspect.* (2024) 132:85003. doi: 10.1289/EHP14449

60. Rosato I, Bonato T, Fletcher T, Batzella E, Canova C. Estimation of per- and polyfluoroalkyl substances (PFAS) half-lives in human studies: a systematic review and meta-analysis. *Environ Res.* (2024) 242:117743. doi: 10.1016/j.envres.2023.117743

61. Liu Z, Cui Z, Li C, Lu K, Chen K, Cui W, et al. Exposure to perfluorodecanoic acid impairs follicular development via inducing granulosa cell necroptosis. *Ecotoxicol Environ Saf.* (2024) 287:117268. doi: 10.1016/j.ecoenv.2024.117268

62. Yi Y, Feng Y, Shi Y, Xiao J, Liu M, Wang K. Per- and polyfluoroalkyl substances (PFASs) and their potential effects on female reproductive diseases. *Toxics.* (2024) 12:539–54. doi: 10.3390/toxics12080539

63. Hoppin JA, Brock JW, Davis BJ, Baird DD. Reproducibility of urinary phthalate metabolites in first morning urine samples. *Environ Health Perspect.* (2002) 110:515–8. doi: 10.1289/ehp.02110515

64. Duty SM, Ackerman RM, Calafat AM, Hauser R. Personal care product use predicts urinary concentrations of some phthalate monoesters. *Environ Health Perspect.* (2005) 113:1530–5. doi: 10.1289/ehp.8083

65. Varik I, Zou R, Bellavia A, Rosenberg K, Sjunnesson Y, Hallberg I, et al. Reduced ovarian cholesterol and steroid biosynthesis along with increased inflammation are associated with high DEHP metabolite levels in human ovarian follicular fluids. *Environ Int.* (2024) 191:108960. doi: 10.1016/j.envint.2024.108960

66. Liu JC, Xing CH, Xu Y, Pan ZN, Zhang HL, Zhang Y, et al. DEHP exposure to lactating mice affects ovarian hormone production and antral follicle development of offspring. *J Hazard Mater.* (2021) 416:125862. doi: 10.1016/j.jhazmat.2021.125862

67. Kaushik P, Kaushik G. An assessment of structure and toxicity correlation in organochlorine pesticides. *J Hazard Mater.* (2007) 143:102–11.

68. Seo SH, Choi SD, Batterman S, Chang YS. Health risk assessment of exposure to organochlorine pesticides in the general population in Seoul, Korea over 12 years: A cross-sectional epidemiological study. *J Hazard Mater.* (2022) 424:127381. doi: 10.1016/j.jhazmat.2021.127381

69. Zhang C, Liu L, Ma Y, Li F. Using isomeric and metabolic ratios of DDT to identify the sources and fate of DDT in Chinese agricultural topsoil. *Environ Sci Technol.* (2018) 52:1990–6. doi: 10.1021/acs.est.7b05877

70. Lee SA, Dai Q, Zheng W, Gao YT, Blair A, Tessari JD, et al. Association of serum concentration of organochlorine pesticides with dietary intake and other lifestyle factors among urban Chinese women. *Environ Int.* (2007) 33:157–63. doi: 10.1016/j.envint.2006.08.010
71. Turusov V, Rakitsky V, Tomatis L. Dichlorodiphenyltrichloroethane (DDT): ubiquity, persistence, and risks. *Environ Health Perspect.* (2002) 110:125–8. doi: 10.1289/ehp.02110125
72. Zhu M, Yuan Y, Yin H, Guo Z, Wei X, Qi X, et al. Environmental contamination and human exposure of polychlorinated biphenyls (PCBs) in China: a review. *Sci Total Environ.* (2022) 805:150270. doi: 10.1016/j.scitotenv.2021.150270
73. Peng Y, Wu J, Luo X, Zhang X, Giesy JP, Mai B. Spatial distribution and hazard of halogenated flame retardants and polychlorinated biphenyls to common kingfisher (*Alcedo atthis*) from a region of South China affected by electronic waste recycling. *Environ Int.* (2019) 130:104952. doi: 10.1016/j.envint.2019.104952
74. Tang X, Shen C, Shi D, Cheema SA, Khan MI, Zhang C, et al. Heavy metal and persistent organic compound contamination in soil from Wenling: an emerging e-waste recycling city in Taizhou area. *China J Hazard Mater.* (2010) 173:653–60. doi: 10.1016/j.jhazmat.2009.08.134
75. Milbrath MO, Wenger Y, Chang CW, Emond C, Garabrant D, Gillespie BW, et al. Apparent half-lives of dioxins, furans, and polychlorinated biphenyls as a function of age, body fat, smoking status, and breast-feeding. *Environ Health Perspect.* (2009) 117:417–25. doi: 10.1289/ehp.11781
76. Warner J, Osuch JR, Karmaus W, Landgraf JR, Taffe B, O'Keefe M, et al. Common classification schemes for PCB congeners and the gene expression of CYP17, CYP19, ESR1 and ESR2. *Sci Total Environ.* (2012) 414:81–9. doi: 10.1016/j.scitotenv.2011.10.044
77. Murugesan P, Balaganesh M, Balasubramanian K, Arunakaran J. Effects of polychlorinated biphenyl (Aroclor 1254) on steroidogenesis and antioxidant system in cultured adult rat Leydig cells. *J Endocrinol.* (2007) 192:325–38. doi: 10.1677/joe.1.06874
78. Shibamoto T, Yasuhara A, Katami T. Dioxin formation from waste incineration. *Rev Environ Contam Toxicol.* (2007) 190:1–41. doi: 10.1007/978-0-387-36903-7_1
79. Zheng GJ, Leung AO, Jiao LP, Wong MH. Polychlorinated dibenzo-p-dioxins and dibenzofurans pollution in China: sources, environmental levels and potential human health impacts. *Environ Int.* (2008) 34:1050–61. doi: 10.1016/j.envint.2008.02.011
80. Fiedler H. (editor). Dioxins and furans (PCDD/PCDF) In: Persistent organic pollutants (2003). 123–201.
81. Sorg O, Zennegg M, Schmid P, Fedosyuk R, Valikhnovskiy R, Gaide O, et al. 2, 3, 7, 8-tetrachlorodibenzo-p-dioxin (TCDD) poisoning in victor Yushchenko: identification and measurement of TCDD metabolites. *Lancet.* (2009) 374:1179–85. doi: 10.1016/S0140-6736(09)60912-0
82. Eskenazi B, Warner M, Marks AR, Samuels S, Needham L, Brambilla P, et al. Serum dioxin concentrations and time to pregnancy. *Epidemiology.* (2010) 21:224–31. doi: 10.1097/EDE.0b013e3181cb8b95
83. Yu K, Zhang X, Tan X, Ji M, Chen Y, Tao Y, et al. Transgenerational impairment of ovarian induced by 2, 3, 7, 8-tetrachlorodibenzo-p-dioxin (TCDD) associated with Igf 2 and H19 in adult female rat. *Toxicology.* (2019) 428:152311. doi: 10.1016/j.tox.2019.152311
84. Salisbury TB, Marcinkiewicz JL. In utero and lactational exposure to 2, 3, 7, 8-tetrachlorodibenzo-p-dioxin and 2, 3, 4, 7, 8-pentachlorodibenzofuran reduces growth and disrupts reproductive parameters in female rats. *Biol Reprod.* (2002) 66:1621–6. doi: 10.1095/biolreprod66.6.1621
85. Calafat AM, Ye X, Wong LY, Bishop AM, Needham LL. Urinary concentrations of four parabens in the U.S. population: NHANES 2005–2006. *Environ Health Perspect.* (2010) 118:679–85. doi: 10.1289/ehp.0901560
86. Błędzka D, Gromadzińska J, Wąsowicz W. Parabens. From environmental studies to human health. *Environ Int.* (2014) 67:27–42. doi: 10.1016/j.envint.2014.02.007
87. Fisher M, Mac Pherson S, Braun JM, Hauser R, Walker M, Feeley M, et al. Paraben concentrations in maternal urine and breast milk and its association with personal care product use. *Environ Sci Technol.* (2017) 51:4009–17. doi: 10.1021/acs.est.6b04302
88. Zhao Y, Liu Y, Chen Y, Wu X, Xiao Q, Li C, et al. Exposure to parabens and associations with oxidative stress in adults from South China. *Sci Total Environ.* (2021) 774:144917. doi: 10.1016/j.scitotenv.2020.144917
89. Jurewicz J, Radwan M, Wielgomas B, Karwacka A, Klimowska A, Kałużny P, et al. Parameters of ovarian reserve in relation to urinary concentrations of parabens. *Environ Health.* (2020) 19:26. doi: 10.1186/s12940-020-00580-3
90. von Goetz N, Wormuth M, Scheringer M, Hungerbühler K. Bisphenol A: how the most relevant exposure sources contribute to total consumer exposure. *Risk Anal.* (2010) 30:473–87. doi: 10.1111/j.1539-6924.2009.01345.x
91. Loganathan SN, Kannan K. Occurrence of bisphenol A in indoor dust from two locations in the eastern United States and implications for human exposures. *Arch Environ Contam Toxicol.* (2011) 61:68–73. doi: 10.1007/s00244-010-9634-y
92. Thayer KA, Doerge DR, Hunt D, Schurman SH, Twaddle NC, Churchwell MI, et al. Pharmacokinetics of bisphenol A in humans following a single oral administration. *Environ Int.* (2015) 83:107–15. doi: 10.1016/j.envint.2015.06.008
93. Tarafdar A, Sirohi R, Balakumaran PA, Reshmy R, Madhavan A, Sindhu R, et al. The hazardous threat of Bisphenol A: toxicity, detection and remediation. *J Hazard Mater.* (2022) 423:127097. doi: 10.1016/j.jhazmat.2021.127097
94. Li C, Cui Z, Liu Z, Fan H, Lan Y, Luo J, et al. MiR-204 regulates autophagy and cell viability by targeting BDNF and inhibiting the NTRK2-dependent PI3K/Akt/mTOR pathway in a human granulosa cell line exposed to bisphenol A. *Ecotoxicol Environ Saf.* (2024) 287:117304. doi: 10.1016/j.ecoenv.2024.117304
95. Lin M, Hua R, Ma J, Zhou Y, Li P, Xu X, et al. Bisphenol A promotes autophagy in ovarian granulosa cells by inducing AMPK/mTOR/ULK1 signalling pathway. *Environ Int.* (2021) 147:106298. doi: 10.1016/j.envint.2020.106298
96. Kim S, Choi K. Occurrences, toxicities, and ecological risks of benzophenone-3, a common component of organic sunscreen products: a mini-review. *Environ Int.* (2014) 70:143–57. doi: 10.1016/j.envint.2014.05.015
97. Park MA, Hwang KA, Lee HR, Yi BR, Jeung EB, Choi KC. Benzophenone-1 stimulated the growth of BG-1 ovarian cancer cells by cell cycle regulation via an estrogen receptor alpha-mediated signaling pathway in cellular and xenograft mouse models. *Toxicology.* (2013) 305:41–8. doi: 10.1016/j.tox.2012.12.021
98. Kinnberg KL, Petersen GI, Albrechtsen M, Minghlani M, Awad SM, Holbech BE, et al. Endocrine-disrupting effect of the ultraviolet filter benzophenone-3 in zebrafish, *Danio rerio*. *Environ Toxicol Chem.* (2015) 34:2833–40. doi: 10.1002/etc.3129
99. Hao Z, Wu Q, Li Z, Li Y, Li Q, Lai X, et al. Maternal exposure to triclosan constitutes a yet unrecognized risk factor for autism spectrum disorders. *Cell Res.* (2019) 29:866–9. doi: 10.1038/s41422-019-0220-1
100. Yueh MF, Tukey RH. Triclosan: A widespread environmental toxicant with many biological effects. *Annu Rev Pharmacol Toxicol.* (2016) 56:251–72. doi: 10.1146/annurev-pharmtox-010715-103417
101. Luo N, Chen J, Chen X, Wang M, Niu X, Chen G, et al. Toxicity evolution of triclosan during environmental transformation and human metabolism: misgivings in the post-pandemic era. *Environ Int.* (2024) 190:108927. doi: 10.1016/j.envint.2024.108927
102. Weatherly LM, Gosse JA. Triclosan exposure, transformation, and human health effects. *J Toxicol Environ Health B Crit Rev.* (2017) 20:447–69. doi: 10.1080/10937404.2017.1399306
103. Du Y, Wang B, Cai Z, Zhang H, Wang B, Liang W, et al. The triclosan-induced shift from aerobic to anaerobic metabolism link to increased steroidogenesis in human ovarian granulosa cells. *Ecotoxicol Environ Saf.* (2021) 220:112389. doi: 10.1016/j.ecoenv.2021.112389
104. Wang F, Guo X, Chen W, Sun Y, Fan C. Effects of triclosan on hormones and reproductive axis in female Yellow River carp (*Cyprinus carpio*): potential mechanisms underlying estrogen effect. *Toxicol Appl Pharmacol.* (2017) 336:49–54. doi: 10.1016/j.taap.2017.10.005
105. Lu F, Xu D, Cheng Y, Dong S, Guo C, Jiang X, et al. Systematic review and meta-analysis of the adverse health effects of ambient PM2.5 and PM10 pollution in the Chinese population. *Environ Res.* (2015) 136:196–204. doi: 10.1016/j.envres.2014.06.029
106. Odo DB, Yang IA, Dey S, Hammer MS, van Donkelaar A, Martin RV, et al. Ambient air pollution and acute respiratory infection in children aged under 5 years living in 35 developing countries. *Environ Int.* (2022) 159:107019. doi: 10.1016/j.envint.2021.107019
107. Shang Y, Sun Z, Cao J, Wang X, Zhong L, Bi X, et al. Systematic review of Chinese studies of short-term exposure to air pollution and daily mortality. *Environ Int.* (2013) 54:100–11. doi: 10.1016/j.envint.2013.01.010
108. Mishra M, Chen PH, Bisquera W Jr, Lin GY, Le TC, Dejchanchaiwong R, et al. Source-apportionment and spatial distribution analysis of VOCs and their role in ozone formation using machine learning in central-West Taiwan. *Environ Res.* (2023) 232:116329. doi: 10.1016/j.envres.2023.116329
109. Fan MY, Zhang YL, Lin YC, Li L, Xie F, Hu J, et al. Source apportionments of atmospheric volatile organic compounds in Nanjing, China during high ozone pollution season. *Chemosphere.* (2021) 263:128025. doi: 10.1016/j.chemosphere.2020.128025
110. Huang Y, Wang L, Zhang S, Zhang M, Wang J, Cheng X, et al. Source apportionment and health risk assessment of air pollution particles in eastern district of Chengdu. *Environ Geochem Health.* (2020) 42:2251–63. doi: 10.1007/s10653-019-00495-0
111. Brain JD. The respiratory tract and the environment. *Environ Health Perspect.* (1977) 20:113–26. doi: 10.1289/ehp.7720113
112. Seaton A, Mac Nee W, Donaldson K, Godden D. Particulate air pollution and acute health effects. *Lancet.* (1995) 345:176–8. doi: 10.1016/s0140-6736(95)90173-6
113. Olesiejuk K, Chalubiński M. How does particulate air pollution affect barrier functions and inflammatory activity of lung vascular endothelium? *Allergy.* (2023) 78:629–38. doi: 10.1111/all.15630
114. Dang S, Ding D, Lu Y, Su Q, Lin T, Zhang X, et al. PM (2.5) exposure during pregnancy induces hypermethylation of estrogen receptor promoter region in rat uterus and declines offspring birth weights. *Environ Pollut.* (2018) 243:851–61. doi: 10.1016/j.envpol.2018.09.065
115. Han Y, Zhu T. Health effects of fine particles (PM2.5) in ambient air. *Sci China. Life Sci.* (2015) 58:624–6. doi: 10.1007/s11427-015-4878-4
116. Lyu Y, Su S, Wang B, Zhu X, Wang X, Zeng EY, et al. Seasonal and spatial variations in the chemical components and the cellular effects of particulate matter collected in northern China. *Sci Total Environ.* (2018) 627:1627–37. doi: 10.1016/j.scitotenv.2018.01.224
117. Muñoz-Planillo R, Kuffa P, Martínez-Colón G, Smith BL, Rajendiran TM, Núñez G. K⁺ efflux is the common trigger of NLRP3 inflammasome activation by bacterial

toxins and particulate matter. *Immunity*. (2013) 38:1142–53. doi: 10.1016/j.immuni.2013.05.016

118. Peng H, Zhao XH, Bi TT, Yuan XY, Guo JB, Peng SQ. PM (2.5) obtained from urban areas in Beijing induces apoptosis by activating nuclear factor-kappa B. *Mil Med Res*. (2017) 4:27. doi: 10.1186/s40779-017-0136-3

119. Ogliari KS, Lichtenfels AJ, de Marchi MR, Ferreira AT, Dolnikoff M, Saldiva PH. Intrauterine exposure to diesel exhaust diminishes adult ovarian reserve. *Fertil Steril*. (2013) 99:1681–8. doi: 10.1016/j.fertnstert.2013.01.103

120. Radwan M, Jurewicz J, Polańska K, Sobala W, Radwan P, Bochenek M, et al. Exposure to ambient air pollution—does it affect semen quality and the level of reproductive hormones? *Ann Hum Biol*. (2016) 43:50–6. doi: 10.3109/03014460.2015.1013986

121. Ding R, Jin Y, Liu X, Zhu Z, Zhang Y, Wang T, et al. Characteristics of DNA methylation changes induced by traffic-related air pollution. *Mutat Res Genet Toxicol Environ Mutagen*. (2016) 796:46–53. doi: 10.1016/j.mrgentox.2015.12.002

122. Carré J, Gatimel N, Moreau J, Parinaud J, Léandri R. Does air pollution play a role in infertility?: a systematic review. *Environ Health*. (2017) 16:82. doi: 10.1186/s12940-017-0291-8

123. Walton RT, Mudway IS, Dundas I, Marlin N, Koh LC, Aithadji L, et al. Air pollution, ethnicity and telomere length in East London schoolchildren: an observational study. *Environ Int*. (2016) 96:41–7. doi: 10.1016/j.envint.2016.08.021

124. Luo H, Wang Q, Guan Q, Ma Y, Ni F, Yang E, et al. Heavy metal pollution levels, source apportionment and risk assessment in dust storms in key cities in Northwest China. *J Hazard Mater*. (2022) 422:126878. doi: 10.1016/j.jhazmat.2021.126878

125. Pan H, Lu X, Lei K. A comprehensive analysis of heavy metals in urban road dust of Xi'an, China: contamination, source apportionment and spatial distribution. *Sci Total Environ*. (2017) 609:1361–9. doi: 10.1016/j.scitotenv.2017.08.004

126. Schiavo B, Meza-Figueroa D, Morton-Bermea O, Angulo-Molina A, González-Grijalva B, Armienta-Hernández MA, et al. Metal (loid) bioaccessibility and risk assessment of ashfall deposit from Popocatepetl volcano, Mexico. *Environ Geochem Health*. (2024) 46:354. doi: 10.1007/s10653-024-02135-8

127. Okubo H, Nakayama SF. Periconceptional maternal diet quality influences blood heavy metal concentrations and their effect on low birth weight: the Japan environment and children's study. *Environ Int*. (2023) 173:107808. doi: 10.1016/j.envint.2023.107808

128. Liang G, Gong W, Li B, Zuo J, Pan L, Liu X. Analysis of heavy metals in foodstuffs and an assessment of the health risks to the general public via consumption in Beijing, China. *Int J Environ Res Public Health*. (2019) 16:909–18. doi: 10.3390/ijerph16060909

129. Papadopoulou E, Haug LS, Sakhi AK, Andrusaityte S, Basagaña X, Brantsaeter AL, et al. Diet as a source of exposure to environmental contaminants for pregnant women and children from six European countries. *Environ Health Perspect*. (2019) 127:107005. doi: 10.1289/EHP5324

130. Pinto E, Cruz M, Ramos P, Santos A, Almeida A. Metals transfer from tobacco to cigarette smoke: evidences in smokers' lung tissue. *J Hazard Mater*. (2017) 325:31–5. doi: 10.1016/j.jhazmat.2016.11.069

131. Salim SA, Sarraf Ov N, Dana Z, Hashami Z, Afrah A, Sadeghi E, et al. A comprehensive image of environmental toxic heavy metals in red meat: A global systematic review and meta-analysis and risk assessment study. *Sci Total Environ*. (2023) 889:164100. doi: 10.1016/j.scitotenv.2023.164100

132. Miglietta S, Cristiano L, Battaglione E, Macchiarelli G, Nottola SA, De Marco MP, et al. Heavy metals in follicular fluid affect the ultrastructure of the human mature cumulus-oocyte complex. *Cells*. (2023) 12:2577–98. doi: 10.3390/cells12212577

133. Liang C, Zhang Z, Cao Y, Wang J, Shen L, Jiang T, et al. Exposure to multiple toxic metals and polycystic ovary syndrome risk: endocrine disrupting effect from as, Pb and Ba. *Sci Total Environ*. (2022) 849:157780. doi: 10.1016/j.scitotenv.2022.157780

134. Wang X, Zhang Y, Peng J, Zhang H, Jiang T, Zhang Z, et al. Association between exposure to multiple toxic metals in follicular fluid and the risk of PCOS among infertile women: the mediating effect of metabolic markers. *Biol Trace Elem Res*. (2024) 203:775–89. doi: 10.1007/s12011-024-04236-y

135. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ*. (2021) 372:n71

136. Cumpston M, Li T, Page MJ, Chandler J, Welch VA, Higgins JP, et al. Updated guidance for trusted systematic reviews: a new edition of the Cochrane handbook for systematic reviews of interventions. *Cochrane Database Syst Rev*. (2019) 10:Ed000142

137. Safarinezhad A, Lamyian M, Ahmadi F, Montazeri A. Mental health literacy of reproductive age women: a qualitative study. *BMC Public Health*. (2025) 25:204. doi: 10.1186/s12889-025-21488-y

138. Teede HJ, Tay CT, Laven J, Dokras A, Moran LJ, Piltonen TT, et al. Recommendations from the 2023 international evidence-based guideline for the assessment and Management of Polycystic Ovary Syndrome. *Fertil Steril*. (2023) 120:767–93. doi: 10.1016/j.fertnstert.2023.07.025

139. Silvestris E, de Pergola G, Rosania R, Loverro G. Obesity as disruptor of the female fertility. *Reprod Biol Endocrinol*. (2018) 16:22. doi: 10.1186/s12958-018-0336-z

140. Hawkins Bressler L, Bernardi LA, De Chavez PJ, Baird DD, Carnethon MR, Marsh EE. Alcohol, cigarette smoking, and ovarian reserve in reproductive-age African-American women. *Am J Obstet Gynecol*. (2016) 215:758.e1–9.

141. Kloss JD, Perlis ML, Zamzow JA, Culnan EJ, Gracia CR. Sleep, sleep disturbance, and fertility in women. *Sleep Med Rev*. (2015) 22:78–87. doi: 10.1016/j.smrv.2014.10.005

142. Shelling AN, Ahmed Nasef N. The role of lifestyle and dietary factors in the development of premature ovarian insufficiency. *Antioxidants*. (2023) 12:1601–21. doi: 10.3390/antiox12081601

143. Nussbaumer-Streit B, Klerings I, Dobrescu AI, Persad E, Stevens A, Garrity C, et al. Excluding non-English publications from evidence-syntheses did not change conclusions: a meta-epidemiological study. *J Clin Epidemiol*. (2020) 118:42–54. doi: 10.1016/j.jclinepi.2019.10.011

144. Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. *Eur J Epidemiol*. (2010) 25:603–5. doi: 10.1007/s10654-010-9491-z

145. Wells G, Shea B, O'Connell D, Peterson J, Welch V, Losos M, et al. The Newcastle-Ottawa scale (NOS) for assessing the quality of nonrandomized studies in Meta-analysis. Editor: GA Wells. Ottawa, Ontario: The Ottawa Health Research Institute (2011).

146. Cook DA, Reed DA. Appraising the quality of medical education research methods: the medical education research study quality instrument and the Newcastle-Ottawa scale-education. *Acad Med*. (2015) 90:1067–76. doi: 10.1097/ACM.0000000000000786

147. Guyatt GH, Oxman AD, Vist GE, Kunz R, Falck-Ytter Y, Alonso-Coello P, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ*. (2008) 336:924–6. doi: 10.1136/bmj.39489.470347.AD

148. Guyatt G, Oxman AD, Akl EA, Kunz R, Vist G, Brozek J, et al. GRADE guidelines: 1. Introduction-GRADE evidence profiles and summary of findings tables. *J Clin Epidemiol*. (2011) 64:383–94. doi: 10.1016/j.jclinepi.2010.04.026

149. Neumann I, Santesso N, Akl EA, Rind DM, Vandvik PO, Alonso-Coello P, et al. A guide for health professionals to interpret and use recommendations in guidelines developed with the GRADE approach. *J Clin Epidemiol*. (2016) 72:45–55. doi: 10.1016/j.jclinepi.2015.11.017

150. Xie CX, Machado GC. Clinimetrics: grading of recommendations, assessment, development and evaluation (GRADE). *J Physiother*. (2021) 67:66. doi: 10.1016/j.jphys.2020.07.003

151. Murad MH, Mustafa RA, Schünemann HJ, Sultan S, Santesso N. Rating the certainty in evidence in the absence of a single estimate of effect. *Evid Based Med*. (2017) 22:85–7. doi: 10.1136/ebmed-2017-110668

152. Ahmed N, Barnett P, Greenburgh A, Pemovska T, Stefanidou T, Lyons N, et al. Mental health in Europe during the COVID-19 pandemic: a systematic review. *Lancet Psychiatry*. (2023) 10:537–56. doi: 10.1016/S2215-0366(23)00113-X

153. Vabre P, Gatimel N, Moreau J, Gayraud V, Picard-Hagen N, Parinaud J, et al. Environmental pollutants, a possible etiology for premature ovarian insufficiency: a narrative review of animal and human data. *Environ Health*. (2017) 16:37. doi: 10.1186/s12940-017-0242-4

154. Barad DH, Weghofer A, Gleicher N. Comparing anti-Müllerian hormone (AMH) and follicle-stimulating hormone (FSH) as predictors of ovarian function. *Fertil Steril*. (2009) 91:1553–5. doi: 10.1016/j.fertnstert.2008.09.069

155. Xu H, Zhang M, Zhang H, Alpadi K, Wang L, Li R, et al. Clinical applications of serum anti-Müllerian hormone measurements in both males and females: An update. *Innovation*. (2021) 2:100091. doi: 10.1016/j.xinn.2021.100091

156. Buratini J, Dellaqua TT, Dal Canto M, La Marca A, Carone D, Mignini Renzini M, et al. The putative roles of FSH and AMH in the regulation of oocyte developmental competence: from fertility prognosis to mechanisms underlying age-related subfertility. *Hum Reprod Update*. (2022) 28:232–54. doi: 10.1093/humupd/dmab044

157. Broer SL, Dölleman M, Opmeer BC, Fauser BC, Mol BW, Broekmans FJ. AMH and AFC as predictors of excessive response in controlled ovarian hyperstimulation: a meta-analysis. *Hum Reprod Update*. (2011) 17:46–54. doi: 10.1093/humupd/dmq034

158. Iliodromiti S, Anderson RA, Nelson SM. Technical and performance characteristics of anti-Müllerian hormone and antral follicle count as biomarkers of ovarian response. *Hum Reprod Update*. (2015) 21:698–710. doi: 10.1093/humupd/dmu062

159. Broer SL, Mol BW, Hendriks D, Broekmans FJ. The role of antimüllerian hormone in prediction of outcome after IVF: comparison with the antral follicle count. *Fertil Steril*. (2009) 91:705–14. doi: 10.1016/j.fertnstert.2007.12.013

160. Weghofer A, Margreiter M, Fauser Y, Schaetz T, Brandstetter A, Boehm D, et al. Age-specific FSH levels as a tool for appropriate patient counselling in assisted reproduction. *Hum Reprod*. (2005) 20:2448–52. doi: 10.1093/humrep/dei076

161. Chauvin S, Cohen-Tannoudji J, Guigon CJ. Estradiol signaling at the heart of Folliculogenesis: its potential deregulation in human ovarian pathologies. *Int J Mol Sci*. (2022) 23:512–31. doi: 10.3390/ijms23010512

162. Segawa T, Teramoto S, Omi K, Miyauchi O, Watanabe Y, Osada H. Changes in estrone and estradiol levels during follicle development: a retrospective large-scale study. *Reprod Biol Endocrinol*. (2015) 13:54. doi: 10.1186/s12958-015-0051-y

163. La Marca A, Stabile G, Arsenio AC, Volpe A. Serum anti-Müllerian hormone throughout the human menstrual cycle. *Hum Reprod.* (2006) 21:3103–7. doi: 10.1093/humrep/del291
164. Rombauts L, Onwude JL, Chew HW, Vollenhoven BJ. The predictive value of antral follicle count remains unchanged across the menstrual cycle. *Fertil Steril.* (2011) 96:1514–8. doi: 10.1016/j.fertnstert.2011.09.005
165. Björvang RD, Hallberg I, Pikki A, Berglund L, Pedrelli M, Kiviranta H, et al. Follicular fluid and blood levels of persistent organic pollutants and reproductive outcomes among women undergoing assisted reproductive technologies. *Environ Res.* (2022) 208:112626. doi: 10.1016/j.envres.2021.112626
166. Crawford NM, Fenton SE, Strynar M, Hines EP, Pritchard DA, Steiner AZ. Effects of perfluorinated chemicals on thyroid function, markers of ovarian reserve, and natural fertility. *Reprod Toxicol.* (2017) 69:53–9. doi: 10.1016/j.reprotox.2017.01.006
167. Barrett ES, Chen C, Thurston SW, Haug LS, Sabaredzovic A, Fjeldheim FN, et al. Perfluoroalkyl substances and ovarian hormone concentrations in naturally cycling women. *Fertil Steril.* (2015) 103:1261–70.e3. doi: 10.1016/j.fertnstert.2015.02.001
168. Ding N, Harlow SD, Randolph JF, Mukherjee B, Batterman S, Gold EB, et al. Perfluoroalkyl substances and incident natural menopause in midlife women: the mediating role of sex hormones. *Am J Epidemiol.* (2022) 191:1212–23. doi: 10.1093/aje/kwac052
169. Du YY, Guo N, Wang YX, Hua X, Deng TR, Teng XM, et al. Urinary phthalate metabolites in relation to serum anti-Müllerian hormone and inhibin B levels among women from a fertility center: a retrospective analysis. *Reprod Health.* (2018) 15:33. doi: 10.1186/s12978-018-0469-8
170. Babadi RS, Williams PL, Preston EV, Li Z, Smith RL, Strakovsky RS, et al. Phthalates and sex steroid hormones across the perimenopausal period: A longitudinal analysis of the midlife women's health study. *Environ Int.* (2024) 188:108770. doi: 10.1016/j.envint.2024.108770
171. Messerlian C, Souter I, Gaskins AJ, Williams PL, Ford JB, Chiu YH, et al. Urinary phthalate metabolites and ovarian reserve among women seeking infertility care. *Hum Reprod.* (2016) 31:75–83. doi: 10.1093/humrep/dev292
172. Parikh FR, Uttamchandani S, Sawkar S, Panpalia M, Naik N, Sinkar P, et al. The impact of follicular fluid phthalate metabolites on the ovarian reserve and ovarian function in Indian women undergoing intracytoplasmic sperm injection. *F&S Science.* (2024) 5:107–20. doi: 10.1016/j.xfss.2023.11.001
173. Beck AL, Rehfeld A, Mortensen LJ, Lorenzen M, Andersson AM, Juul A, et al. Ovarian follicular fluid levels of phthalates and benzophenones in relation to fertility outcomes. *Environ Int.* (2024) 183:108383. doi: 10.1016/j.envint.2023.108383
174. Yao YC, Du YY, Guo N, Liu FH, Deng TR, Li YF. Associations between urinary phthalate concentrations and antral follicle count among women undergoing in vitro fertilization. *Front Endocrinol.* (2024) 14:1286391–9. doi: 10.3389/fendo.2023.1286391
175. Hoffmann-Dishon N, Barnett-Izhaki Z, Zalko D, Hemi R, Farzam N, Hauser R, et al. Endocrine-disrupting chemicals in follicular fluid and follicular reproductive hormone levels. *J Assist Reprod Genet.* (2024) 41:1637–42. doi: 10.1007/s10815-024-03101-0
176. Bloom MS, Fujimoto VY, Storm R, Zhang L, Butts CD, Sollohub D, et al. Persistent organic pollutants (POPs) in human follicular fluid and in vitro fertilization outcomes, a pilot study. *Reprod Toxicol.* (2017) 67:165–73. doi: 10.1016/j.reprotox.2017.01.004
177. Whitworth KW, Baird DD, Steiner AZ, Bornman RMS, Travlos GS, Wilson RE, et al. Anti-Müllerian hormone and lifestyle, reproductive, and environmental factors among women in rural South Africa. *Epidemiology.* (2015) 26:429–35. doi: 10.1097/EDE.0000000000000265
178. Mok-Lin E, Ehrlich S, Williams PL, Petrozza J, Wright DL, Calafat AM, et al. Urinary bisphenol A concentrations and ovarian aging among women undergoing IVF. *Int J Androl.* (2010) 33:385–93. doi: 10.1111/j.1365-2605.2009.01014.x
179. Bloom MS, Kim D, Vom Saal FS, Taylor JA, Cheng G, Lamb JD, et al. Bisphenol A exposure reduces the estradiol response to gonadotropin stimulation during in vitro fertilization. *Fertil Steril.* (2011) 96:672–7.e2. doi: 10.1016/j.fertnstert.2011.06.063
180. Kim HK, Ko DH, Lee W, Kim KR, Chun S, Song J, et al. Body fluid concentrations of bisphenol A and their association with in vitro fertilization outcomes. *Hum Fertil (Camb).* (2021) 24:199–207. doi: 10.1080/14647273.2019.1612104
181. Smith KW, Souter I, Dimitriadis I, Ehrlich S, Williams PL, Calafat AM, et al. Urinary paraben concentrations and ovarian aging among women from a fertility center. *Environ Health Perspect.* (2013) 121:1299–305. doi: 10.1289/ehp.1205350
182. Warner M, Eskenazi B, Olive DL, Samuels S, Quick-Miles S, Vercellini P, et al. Serum dioxin concentrations and quality of ovarian function in women of Seveso. *Environ Health Perspect.* (2007) 115:336–40. doi: 10.1289/ehp.9667
183. Kido T, Van Dao T, Ho MD, Dang ND, Pham NT, Okamoto R, et al. High cortisol and cortisone levels are associated with breast milk dioxin concentrations in Vietnamese women. *Eur J Endocrinol.* (2014) 170:131–9. doi: 10.1530/EJE-13-0410
184. Jurewicz J, Wielgomas B, Radwan M, Karwacka A, Klimowska A, Dziewirska E, et al. Triclosan exposure and ovarian reserve. *Reprod Toxicol.* (2019) 89:168–72. doi: 10.1016/j.reprotox.2019.07.086
185. Mínguez-Alarcón L, Christou G, Messerlian C, Williams PL, Carignan CC, Souter I, et al. Urinary triclosan concentrations and diminished ovarian reserve among women undergoing treatment in a fertility clinic. *Fertil Steril.* (2017) 108:312–9. doi: 10.1016/j.fertnstert.2017.05.020
186. Gaskins AJ, Mínguez-Alarcón L, Fong KC, Abdelmessih S, Coull BA, Chavarro JE, et al. Exposure to fine particulate matter and ovarian reserve among women from a fertility clinic. *Epidemiology.* (2019) 30:486–91. doi: 10.1097/EDE.0000000000001029
187. Hood RB, James P, Fong KC, Mínguez-Alarcón L, Coull BA, Schwartz J, et al. The influence of fine particulate matter on the association between residential greenness and ovarian reserve. *Environ Res.* (2021) 197:111162. doi: 10.1016/j.envres.2021.111162
188. Wiczeorek K, Szczesna D, Radwan M, Radwan P, Polańska K, Kilanowicz A, et al. Exposure to air pollution and ovarian reserve parameters. *Sci Rep.* (2024) 14:461. doi: 10.1038/s41598-023-50753-6
189. Kim H, Choe SA, Kim OJ, Kim SY, Kim S, Im C, et al. Outdoor air pollution and diminished ovarian reserve among infertile Korean women. *Environ Health Prev Med.* (2021) 26:20. doi: 10.1186/s12199-021-00942-4
190. Pang L, Yu W, Lv J, Dou Y, Zhao H, Li S, et al. Air pollution exposure and ovarian reserve impairment in Shandong province, China: the effects of particulate matter size and exposure window. *Environ Res.* (2023) 218:115056. doi: 10.1016/j.envres.2022.115056
191. Liu S, Zhao J, Ye X, Fu M, Zhang K, Wang H, et al. Fine particulate matter and its constituent on ovarian reserve: identifying susceptible windows of exposure. *Sci Total Environ.* (2023) 904:166744. doi: 10.1016/j.scitotenv.2023.166744
192. Wang XY, Zhang S, Yan HH, Ma Z, Zhang YS, Luo HN, et al. Association of exposure to ozone and fine particulate matter with ovarian reserve among women with infertility. *Environ Pollut.* (2024) 340:122845. doi: 10.1016/j.envpol.2023.122845
193. La Marca A, Spaggiari G, Domenici D, Grassi R, Casonati A, Baraldi E, et al. Elevated levels of nitrous dioxide are associated with lower AMH levels: a real-world analysis. *Hum Reprod.* (2020) 35:2589–97. doi: 10.1093/humrep/deaa214
194. Fang L, Ma C, Ma Y, Zhao H, Peng Y, Wang G, et al. Associations of long-term exposure to air pollution and green space with reproductive hormones among women undergoing assisted reproductive technology: A longitudinal study. *Sci Total Environ.* (2023) 905:166941. doi: 10.1016/j.scitotenv.2023.166941
195. Merklinger-Gruchala A, Jasienska G, Thune I, Kapiszewska M. Joint effect of particulate matter and cigarette smoke on women's sex hormones. *BMC Womens Health.* (2022) 22:3–17. doi: 10.1186/s12905-021-01586-w
196. Gregoire AM, Upson K, Niehoff NM, Chin HB, Kaufman JD, Weinberg CR, et al. Outdoor air pollution and anti-Müllerian hormone concentrations in the sister study. *Environ Epidemiol.* (2021) 5:e163–e168.
197. LaPointe S, Lee JC, Nagy ZP, Shapiro DB, Chang HH, Wang Y, et al. Ambient traffic related air pollution in relation to ovarian reserve and oocyte quality in young, healthy oocyte donors. *Environ Int.* (2024) 183:108382. doi: 10.1016/j.envint.2023.108382
198. Namvar Z, Mohseni-Bandpei A, Shahsavani A, Amini H, Mousavi M, Hopke PK, et al. Long-term exposure to air pollution and anti-müllerian hormone rate of decline: a population-based cohort study in Tehran, Iran. *Environ Sci Pollut Res Int.* (2023) 30:86987–97. doi: 10.1007/s11356-023-28394-z
199. Feng X, Luo J, Wang X, Xie W, Jiao J, Wu X, et al. Association of exposure to ambient air pollution with ovarian reserve among women in Shanxi province of North China. *Environ Pollut.* (2021) 278:116868. doi: 10.1016/j.envpol.2021.116868
200. Liu SY, Liu L, Ye X, Fu MJ, Wang W, Zi YH, et al. Ambient ozone and ovarian reserve in Chinese women of reproductive age: identifying susceptible exposure windows. *J Hazard Mater.* (2024) 461:132579. doi: 10.1016/j.jhazmat.2023.132579
201. Mínguez-Alarcón L, Williams PL, Souter I, Sacha C, Amarasiwardena CJ, Ford JB, et al. Hair mercury levels, intake of omega-3 fatty acids and ovarian reserve among women attending a fertility center. *Int J Hyg Environ Health.* (2021) 237:113825. doi: 10.1016/j.ijheh.2021.113825
202. Wright DL, Afeiche MC, Ehrlich S, Smith K, Williams PL, Chavarro JE, et al. Hair mercury concentrations and in vitro fertilization (IVF) outcomes among women from a fertility clinic. *Reprod Toxicol.* (2015) 51:125–32. doi: 10.1016/j.reprotox.2015.01.003
203. Jackson LW, Howards PP, Wactawski-Wende J, Schisterman EF. The association between cadmium, lead and mercury blood levels and reproductive hormones among healthy, premenopausal women. *Hum Reprod.* (2011) 26:2887–95. doi: 10.1093/humrep/der250
204. Kim K, Pollack AZ, Nobles CJ, Sjaarda LA, Zolton JR, Radoc JG, et al. Associations between blood cadmium and endocrine features related to PCOS-phenotypes in healthy women of reproductive age: a prospective cohort study. *Environ Health.* (2021) 20:64. doi: 10.1186/s12940-021-00749-4
205. Jiao X, Meng T, Zhai Y, Zhao L, Luo W, Liu P, et al. Ovarian reserve markers in premature ovarian insufficiency: within different clinical stages and different etiologies. *Front Endocrinol.* (2021) 12:601752. doi: 10.3389/fendo.2021.601752
206. Tran DN, Jung EM, Yoo YM, Ahn C, Kang HY, Choi KC, et al. Depletion of follicles accelerated by combined exposure to phthalates and 4-vinylcyclohexene diepoxide, leading to premature ovarian failure in rats. *Reprod Toxicol.* (2018) 80:60–7. doi: 10.1016/j.reprotox.2018.06.071
207. Gomez E, Pillon A, Fenet H, Rosain D, Duchesne MJ, Nicolas JC, et al. Estrogenic activity of cosmetic components in reporter cell lines: parabens, UV screens, and musks. *J Toxicol Environ Health A.* (2005) 68:239–51. doi: 10.1080/15287390590895054

208. Wang L, Luo D, Liu X, Zhu J, Wang F, Li B, et al. Effects of PM (2.5) exposure on reproductive system and its mechanisms. *Chemosphere*. (2021) 264:128436.
209. Qiu X, Duan L, Gao J, Wang S, Chai F, Hu J, et al. Chemical composition and source apportionment of PM₁₀ and PM_{2.5} in different functional areas of Lanzhou, China. *J Environ Sci*. (2016) 40:75–83. doi: 10.1016/j.jes.2015.10.021
210. Jurisicova A, Taniuchi A, Li H, Shang Y, Antenos M, Detmar J, et al. Maternal exposure to polycyclic aromatic hydrocarbons diminishes murine ovarian reserve via induction of Harakiri. *J Clin Invest*. (2007) 117:3971–8.
211. Xiao C, Lai D. Impact of oxidative stress induced by heavy metals on ovarian function. *J Appl Toxicol*. (2024) 45:107–16. doi: 10.1002/jat.4664
212. Canipari R, De Santis L, Cecconi S. Female fertility and environmental pollution. *Int J Environ Res Public Health*. (2020) 17:8802–19. doi: 10.3390/ijerph17238802
213. Joffe M. Infertility and environmental pollutants. *Br Med Bull*. (2003) 68:47–70. doi: 10.1093/bmb/ldg025
214. Green MP, Harvey AJ, Finger BJ, Tarulli GA. Endocrine disrupting chemicals: impacts on human fertility and fecundity during the peri-conception period. *Environ Res*. (2021) 194:110694. doi: 10.1016/j.envres.2020.110694
215. Shupler M, Hystad P, Birch A, Miller-Lionberg D, Jeronimo M, Arku RE, et al. Household and personal AIR pollution exposure measurements from 120 communities in eight countries: results from the PURE-AIR study. *Lancet Planet Health*. (2020) 4:e451–62. doi: 10.1016/S2542-5196(20)30197-2
216. Cucurachi S, Sala S, Laurent A, Heijungs R. Building and characterizing regional and global emission inventories of toxic pollutants. *Environ Sci Technol*. (2014) 48:5674–82. doi: 10.1021/es405798x
217. Polichetti G, Cocco S, Spinali A, Trimarco V, Nunziata A. Effects of particulate matter (PM (10), PM (2.5) and PM (1)) on the cardiovascular system. *Toxicology*. (2009) 261:1–8. doi: 10.1016/j.tox.2009.04.035
218. Alsberg B, Kesmodel US, Humaidan P, Bungum L. The natural menstrual cycle revisited - can natural cycle be trusted. *J Ovarian Res*. (2024) 17:153. doi: 10.1186/s13048-024-01469-2
219. Gari M, Grimalt JO. Inverse age-dependent accumulation of decabromodiphenyl ether and other PBDEs in serum from a general adult population. *Environ Int*. (2013) 54:119–27. doi: 10.1016/j.envint.2013.01.012
220. Wu S, Hao G, Zhang Y, Chen X, Ren H, Fan Y, et al. Poor ovarian response is associated with air pollutants: A multicentre study in China. *EBioMedicine*. (2022) 81:104084. doi: 10.1016/j.ebiom.2022.104084
221. Ryva BA, Pacyga DC, Anderson KY, Calafat AM, Whalen J, Aung MT, et al. Associations of urinary non-persistent endocrine disrupting chemical biomarkers with early-to-mid pregnancy plasma sex-steroid and thyroid hormones. *Environ Int*. (2024) 183:108433. doi: 10.1016/j.envint.2024.108433
222. Kalloo G, Wellenius GA, McCandless L, Calafat AM, Sjodin A, Karagas M, et al. Profiles and predictors of environmental chemical mixture exposure among pregnant women: the health outcomes and measures of the environment study. *Environ Sci Technol*. (2018) 52:10104–13. doi: 10.1021/acs.est.8b02946
223. Li J, Liu W, Xia W, Zhao H, Zhou Y, Li Y, et al. Variations, determinants, and Coexposure patterns of personal care product chemicals among Chinese pregnant women: A longitudinal study. *Environ Sci Technol*. (2019) 53:6546–55. doi: 10.1021/acs.est.9b01562
224. EU. Chemicals strategy (2020). Available online at: https://environment.ec.europa.eu/strategy/chemicals-strategy_en (Accessed May 16, 2025).
225. EU. Zero Pollution Action Plan (2021). [May 16, 2025]. Available online at: https://environment.ec.europa.eu/strategy/zero-pollution-action-plan_en (Accessed May 16, 2025).
226. Agency USEP. Per-and Polyfluoroalkyl Substances (PFAS) (2024). Available online at: <https://www.epa.gov/sdwa/and-polyfluoroalkyl-substances-pfas> (Accessed May 16, 2025).
227. Agency USEP. Regulatory and Guidance Information by Topic: Toxic Substances (2025). Available online at: <https://www.epa.gov/regulatory-information-topic/regulatory-and-guidance-information-topic-toxic-substances> (Accessed May 16, 2025).
228. Zare Jeddi M, Hopf NB, Louro H, Viegas S, Galea KS, Pasanen-Kase R, et al. Developing human biomonitoring as a 21st century toolbox within the European exposure science strategy 2020–2030. *Environ Int*. (2022) 168:107476. doi: 10.1016/j.envint.2022.107476
229. EU. EU Action Plan: 'Towards Zero Pollution for Air, Water and Soil' (2021). Available online at: <https://eur-lex.europa.eu/legal-content/EN/ALL/?uri=SWD:2021:141:FIN> (Accessed May 16, 2025).
230. Create a Local Network of “Learn the Signs. Act Early”. Delegates (2016). Available online at: <https://blogs.cdc.gov/actearlypromisingpractices/2016/06/17/create-a-local-network-of-learn-the-signs-act-early-delegates/> (Accessed June 28, 2025).
231. AlSaad R, Abd-Alrazaq A, Choucair F, Ahmed A, Aziz S, Sheikh J. Harnessing artificial intelligence to predict ovarian stimulation outcomes in in vitro fertilization: scoping review. *J Med Internet Res*. (2024) 26:e53396. doi: 10.2196/53396
232. Hao Y, Wang Y, Yan L, Xu X, Chen D, Zhao Y, et al. Synthetic phenolic antioxidants and their metabolites in follicular fluid and association with diminished ovarian reserve: A case-control study. *Environ Health Perspect*. (2023) 131:67005. doi: 10.1289/EHP11309
233. Tan T, Junghans C, Varaden D. Empowering community health professionals for effective air pollution information communication. *BMC Public Health*. (2023) 23:2547. doi: 10.1186/s12889-023-17462-1
234. Marchiandi J, Dagnino S, Zander-Fox D, Green MP, Clarke BO. Characterization of chemical Exposome in A paired human preconception pilot study. *Environ Sci Technol*. (2024) 58:20352–65. doi: 10.1021/acs.est.4c04356
235. Agency USEP. TSCA Chemical Substance Inventory (1975). Available online at: <https://www.epa.gov/tsc-a-inventory> (Accessed March 28, 2025).