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*CORRESPONDENCE Timothy S. Frey frey.146@osu.edu

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Crop rotation and a rye cover crop have minor impacts on soil health, microbial communities, and soybean yield in Ohio

Timothy S. Frey^{1*}, Denis A. Shah², Laura E. Lindsey³, Christine Sprunger^{4,5,6}, Horacio D. Lopez-Nicora⁷ and M. Soledad Benitez Ponce¹

¹Department of Plant Pathology, College of Food, Agricultural, and Environmental Sciences, The Ohio State University, Wooster, OH, United States, ²Department of Plant Pathology, Kansas State University, Manhattan, KS, United States, ³Department of Horticulture and Crop Science, College of Food, Agricultural, and Environmental Sciences, The Ohio State University, Columbus, OH, United States, ⁴Department of Plant, Soil, and Microbial Sciences, Michigan State University, East Lansing, MI, United States, ⁵W.K. Kellogg Biological Station, Michigan State University, Hickory Corners, MI, United States, ⁶Plant Resilience Institute, Michigan State University, East Lansing, MI, United States, ⁷Department of Plant Pathology, College of Food, Agricultural, and Environmental Sciences, The Ohio State University, Columbus, OH, United States

Crop rotations in the Midwest U.S. have become less diversified over the last twenty years while at the same time cover crop acreage has increased. The impact of these changes on soil health and soil microbial communities is not entirely understood. In rotational plots at two sites in Ohio, we investigated the impact of a corn-soy-wheat crop rotation and a rye cover crop on soil carbon, nitrogen, soil organic matter, and their relationships with soybean yield. Rhizosphere fungal, arbuscular mycorrhizal fungal, and bacterial communities were also assessed, as well as soil health and yield parameters. Microbial communities were measured using long and short-read amplicon sequencing on the PacBio and Illumina platforms. Small changes in soil health parameters were observed, including increased soil protein in the cover crop treatment at one of the sites. We also observed increasing yields in the diversified rotations and cover crops at one site. Soil bacterial and fungal communities showed differences driven by site, and AM fungi showed differences by rotation in one site-year combination. Network analysis revealed several fungal network modules correlated with increases in POXC and bacterial network modules correlated with soil protein and respiration. This research uniquely addresses the interactions between cover crops and diversified rotations and their impact on soil health in no-till production in Ohio.

KEYWORDS

fungi, bacteria, rye, soybean, communities, mycorrhizae, soil

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Introduction

Cover crops are one of the most ancient crop management practices. They are mentioned in Chinese writing as far back as the 5th century BC (1). The Roman agricultural writer Columella advised 1st-century farmers that "Where no kind of manure is to be had, I think the cultivation of lupines will be found the readiest and best substitute. If they are sown about the middle of September in a poor soil and then plowed in, they will answer as well as the best manure." (2). Cover crops are distinguished from cash crops and forage crops in that they are grown for benefits that do not include the sale (cash crop) or direct use (forage crop) of the crop (3). Historically, cover crops have been grown to address many agronomic needs, including reduction of soil erosion (4), prevention of nutrient leaching (5), nitrogen runoff control (6), increasing beneficial insects (7) and microorganisms (8), and assisting in integrated pest management (9). Cover crop adoption in the United States is still low, only on 4% of total cropland in 2022, but has increased from 6.1 million ha to 7.2 million ha from 2017 to 2022 (10, 11). In Ohio, adoption increased from 290,160 to 303,956 ha from 2017 to 2022, and the surrounding states (Indiana, Michigan, Pennsylvania, and Kentucky) recorded increases of 7 to 17% (10). Adoption rates vary but have increased in most cash crops, including soybean, which saw an increase in adoption from 4.9% in 2015 to 7% in 2018 (3).

In contrast to cover cropping, crop rotation is common throughout the United States, including Ohio. In Ohio, economic and policy considerations have driven growers towards a corn-soybean (CS) rotation (12–14). The CS rotation is replacing more diverse rotations, which might include wheat or hay (14, 15). Rotations with more crop diversity lead to positive improvements in soil health (16), greater soil carbon (17), increased microbial biomass and diversity (18), and improved soil water content (19). Our previous research contrasting a corn-soybean-wheat (CSW) rotation with a CS rotation in long-term plots indicated small but positive changes in total nitrogen, soil organic matter (SOM), total carbon (20), bacterial (21), and fungal communities (22).

In the fall of 2020, rye (Secale cereale) cover crop treatments were added to this long-term experiment to test if they could supplement the positive effects of a CSW rotation. Rye is one of the most frequently adopted cover crops in the eastern United States due to its superior cold tolerance and rapid growth in fall and winter after the typical cash crops (corn and soybean) have been planted (3, 10). This advantage of rye - fall planting and growth through the winter- allows flexibility, integrating well with Ohio's most common cash crop rotation of corn and soybean. Additionally, rve is one of the more economical cover crops, with seed costs averaging US\$40.20 per ha (3). Rye cover crops help reduce weed biomass (23), reduce nitrate loading and leaching from drainage (24), help with soil moisture retention (25), and improved soil organic carbon in long-term experiments (26). Previous research specifically focused on the benefits of rye cover crops in Ohio reported little consistent benefit to soil fertility in continuous corn or soybean (27). Hanrahan et al. (28) reported that cover crops, including rye, reduced nitrogen losses but did not affect phosphorous loss in different rotational schemes in Ohio. To increase soybean yield with rye adoption, early cover crop planting and late cover crop termination were found to be important (29). Mukherjee and Lal (30) showed the benefits of other cover crop species, including pea (*Pisum sativum*) and turnip (*Brassica rapa*), showing increased soybean yields in a no-till system.

In the current study, we investigated some important indicators of soil health, including soil protein, soil respiration, SOM, permanganate oxidizable carbon (POXC) and microbial communities, including fungal, arbuscular mycorrhizal (AM) fungal and bacterial communities. Soil protein can indicate available organic nitrogen (31) and nitrogen management (32). Soil respiration, which is an indicator of the outflux of carbon from soil (33), is responsive to climate (34) and management changes (35). SOM consists of plant and animal residues at various stages of decomposition (36). It is an important soil health indicator as a carbon and nitrogen source (37), and for mineral nutrient cycling (38), water retention (39), and soil structure (40). POXC is a soil chemical fraction that reacts to permanganate and may indicate plant-mediated soil processes (41); it is sensitive to recent management changes. Cultural practices such as crop rotation and cover crops influence soil microbial communities as deep as 60 cm (42). These communities are critical for soil health, including nutrient cycling (43), phosphorous leaching (44), SOM formation, transformation, and decomposition (45), decreasing pathogen pressure (46), and contributing to other soil properties (47). Microbial community changes observed with cover cropping included shifts toward symbiotrophs and a decrease in saprotrophs (48), increases in the fungi:bacteria ratio, AM fungal colonization, and spore density (49).

More data is needed to help researchers and farmers interpret the relationship between microbial community changes, other soil health indicators, and soil functioning (50, 51). Microbial communities are important as a potential predictor of soil health because they respond quickly to changes to the soil brought about by human interventions (52, 53) and microbial analysis is becoming cheaper and more accessible (50). This study contributes by analyzing the relationships between microbial communities and soil health indicators with rotations and cover crops and their impact on yield.

In the current study, we investigated the impact of adding rye to CS and CSW rotations-including the impact of a first year rye cover crop implementation - on soil health, soil microbial communities, and soybean productivity. We hypothesized that (Supplementary Figure S1): 1. A rye cover crop will not have an impact on soybean yield in CS or CSW rotation (29, 54). 2. The addition of a rye cover crop will improve soil health characteristics, including carbon, as measured from soil respiration and POXC (55, 56), nitrogen in the form of soil protein (57), and SOM (58), regardless of the rotation. 3. Adding a rye cover crop will shift microbial communities in a CSW rotation (59). 4. Changes in microbial communities will be associated with changes in soil health metrics (42, 60). Understanding how cover crops influence a variety of soil health indicators could inform future management decisions of Ohio growers, especially regarding rotation length and cover crop adoption.

Materials and methods

Rotation and cover crop plots

Rotation plots were established at two sites in Ohio in 2013. The first was in northwestern Ohio at the Northwest Agricultural Research Station (NWARS) in Custar, OH (41°21'30.07" N, 83°76'36.55" W); the second was in western Ohio at the Western Agricultural Research Station (WARS) in South Charleston, OH (39°86'04.82" N, 83°67'72.81" W) (Supplementary Figure S2). The treatments were a rotation of corn and soybean (CS) and a rotation of corn, soybean, and winter wheat (CSW) (20). Plots were arranged in a randomized complete block design with four replicates per treatment, with every crop in the rotation represented in at least one plot each year. Plots were managed using notill practices. The rye cover crop treatment was implemented in the fall of 2020 as a split-plot design. Within each rotation plot, one of the subplots was managed with a rye (r) cover crop between corn harvest and soybean planting (CrSW and CrS), and the other subplot left fallow as with the previously established rotations (CSW and CS, Supplementary Figures S3, S4). Farm managers selected commercially available high-yielding soybean cultivars, which varied by year (Supplementary Table S1). Depending on weather and soil conditions, soybeans and corn were planted between April and June and harvested in October or November. Farm managers collected yield and crop moisture data at the end of the growing season and applied herbicides and fertilization as needed, following commercial production practices throughout all trials (61). Herbicides were used to terminate cover crops as well as prior to planting. Fungicides and insecticides were applied as seed treatment to soybean, but no foliar fungicide was applied during the experiment. Corn and soybean cultivars, planting dates, and herbicide applications are summarized in Supplementary Table S1.

Soil sampling

Rhizosphere samples were collected at NWARS and WARS in 2021 and 2022 from soybean seedlings following the first rye cycle in each plot. In each subplot, eight samples were collected when the soybean was between V1 and V3 growth stages (62). Plants were dug up to a depth of 10-15 cm using a small shovel, with minimal disturbance to the roots. The shovels were cleaned with 70% ethanol and wiped with a towel until thoroughly cleaned between each subplot. Bulk soil was gently separated from the roots. The remaining rhizosphere soil was collected and homogenized using soil sieves (size 0.2 cm) to form one composite sample per subplot. These samples were placed in Ziploc plastic bags and transported on ice to the laboratory. The soil was placed in a -20°C freezer in the laboratory until freeze-drying. Samples for microbial community analysis were lyophilized for 24 hours or until completely dry. Lyophilized samples were stored at -80°C until DNA extraction.

Environmental conditions

Air temperature, soil temperature, precipitation, and soil properties were acquired from the Ohio State University CFAES weather station data. Stations are located at the same sites as the plots; the data is publicly accessible at https://weather.cfaes.osu.edu/. Specifically, cover crop growing season temperature included the dates between the sowing and the termination of the cover crop. The average temperature during the soybean growing season was taken for the days between sowing and harvest. Total precipitation, average relative humidity, and growing degree days were estimated for the entire soybean growing season. Soil temperature was measured at 5 cm and 10 cm depths and averaged over the growing season. Sand, silt, and clay percentages were collected at the 10 cm depth from the same sites and reported in an earlier dataset (63).

Soil health measurements

The following measurements were taken from soil samples to assess soil health: permanganate oxidizable carbon (POXC), soil protein, SOM, and soil respiration. These analyses were conducted in the Sprunger Lab, Benitez Ponce Lab, or the Service Testing and Research (STAR) Lab at The Ohio State University. Before analysis, soil samples from each subplot were homogenized, air-dried, and ground to pass through a 2 mm sieve. The POXC fraction was determined using methods described by Culman (64). Briefly, 2.5g of each sample was reacted with 2 ml of KMnO₄ and 18 ml of H₂O for two min and then allowed to settle for 10 min. Afterward, 0.5 ml of the supernatant of this solution was diluted in 49.5 ml of H₂O and read on a spectrophotometer at 550 nm. The mass of POXC was calculated by comparing it to a standard curve using the equation in Culman (64). Soil protein was calculated using the protocol for autoclaved-citrate extractable soil after Hurisso et al. (31). Eight ml of 0.02M sodium citrate (pH 7.0) was added to each tube containing 1 g of soil and mixed well. The tubes were then autoclaved for 30 min. After autoclaving, tubes were cooled and centrifuged for 15 minutes at 3100 g. Protein measurements were then taken using a Bradford assay (Pierce Bradford Protein Assay Kit, ThermoFisher, Waltham, MA) and read on a spectrophotometer at 590nm. Protein concentration was calculated by comparing it to a standard curve. Percent SOM was calculated using the loss on ignition method (65). Each sample was dried at 105 °C. The samples were weighed, heated to 360 °C for two hours, and then weighed again after the temperature dropped below 150 °C. SOM was estimated from the weight loss-on-ignition using regression analysis. Soil respiration was calculated from air-dried soil using the Cornell method described in Moebius-Clune et al. (66). Briefly, 20 g of air-dried, sieved soil was placed into a mason jar with a trap assembly, and the beaker was filled with the trapping solution (9 ml of 0.5M KOH). Then, 7 ml of ddH₂O was pipetted into the jar and wicked into the soil. The jar was incubated for four days, at the end of which the conductivity of the trap solution was measured. We used data collected by Huo et al. (20) and Frey et al. (22) to look at yield and soil health dynamics over the rotation treatment and after the incorporation of the cover crop treatment.

DNA extraction

DNA was extracted from homogenized rhizosphere soil samples taken from each subplot. DNA was extracted using a

TaKaRa 159 NucleoSpin[®]96 Soil kit (#740787), following the manufacturer's instructions. Two-hundred and fifty mg of the homogenized and lyophilized soil sample from each subplot was loaded into each of the provided bead tubes and agitated using an MP Biomedicals FastPrep-24 5G sample preparation system. For the extraction, both optional buffers, SL1 and Enhancer SX, were used to increase efficiency. A DNA extraction with a buffer and no soil was included as a negative control for the extraction kit. Before library preparation, DNA concentration was assessed by gel electrophoresis, with band intensities compared using the average of two ThermoScientific GeneRuler 1Kb ladders using ImageJ (67). This was compared to nanodrop measurement. After extraction, DNA was stored at -20 °C until amplicon sequencing library preparation.

Amplicon library preparation and sequencing

DNA libraries for fungal community analysis through amplicon sequencing were prepared using a 2-step PCR process. First, sequences for the target gene of interest were amplified. This was done using two different primer sets ITS1F (5' CTT-GGT-CAT-TTA-GAG-GAA-GTAA-3', 68) and ITS4 (5'-TCC-TCC-GCT-TAT-TGA-TAT-GC-3', 69) for general fungi and AML1 (5'-ATC AAC TTT CGA TGG TAG GAT AGA-3') and AML2 (5'-GAA CCC AAA CAC TTT GGT TTC C-3') for AM fungi (70) tailed with universal sequences (5' blocked, F -/5AmMC6/GCA-GTC-GAA-CAT-GTA-GCT-GAC-TCA-GGT-CAC-primer-3', R-5'/5AmMC6/TGG-ATC-ACT-TGT-GCA-AGC-ATC-ACA-TCG-TAG-primer-3') necessary for the addition of the barcode primers in the next step. PCR conditions recommended by PacBio were followed with annealing temperatures at 61 °C for ITS1F/4 and AML1/2 (71). The product from these reactions was then used to attach PACBIO barcode primers (PacBio Barcoded Universal F/R Primers Plate-96v2) using a PCR with conditions as recommended by the manufacturer with 20 cycles and 64 °C annealing temperature. KAPA HiFi HotStart polymerase (Roche, Basel, Switzerland) and filtered pipette tips were used for all reactions. Samples were then pooled into two pools per year (96 samples in each pool) using the best practices for equimolar pooling as recommended by PacBio. Libraries were then concentrated and cleaned using an AMPure (Beckman Coulter, Brea, CA, USA) magnetic bead cleanup. Cleaned fungi and AM fungi libraries were then submitted to the University of Louisville for PacBio sequencing (PacBio, Menlo Park, CA, U.S.A). The remaining library preparation, including the attachment of SMRTbell adaptors, was carried out at the University of Louisville Sequencing Technology Center. Bacterial communities were sequenced at the Molecular Cellular Imaging Center (MCIC) at Ohio State University (College of Food, Agriculture and Environmental Sciences, Wooster Campus). The initial amplification was done using the 515F (5'-GTG CCA GCM GCC GCG GTA A-3', 72) and 806R (5'-GGA CTA CHV GGG TWT CTA AT-3', 73) primers which target the V3-V4 region of the 16s rDNA. A second double-indexing PCR was then conducted. Samples were then pooled into a single sequencing library in equimolar concentrations. Bacteria DNA was sequenced using an Illumina MiSeq (Illumina, San Diego, CA. U.S.A.) and a 2x300 kit at the MCIC. The Commercial ZymoBIOMICS Microbial Community DNA (D6300; Zymo Research Irvine, CA) and the Mycobiome Genomic DNA Mix (ATCC MSA-1010tm) were included as positive controls. A water sample was included as a DNA extraction control for bacterial and fungal libraries. Fastq sequences are available at the NCBI SRA, Bioproject PRJNA1190478.

Bioinformatics

Primers and adapters were removed from demultiplexed fastq files using cutadapt (74) in the Unix shell environment at the Ohio Supercomputing Center (OSC, The Ohio State University, Columbus, OH, U.S.A). The dada2 package (Ver. 1.26, 75) in R (76) was used for error rate calculation, primer trimming, chimera detection and removal, Amplicon Sequencing Variant (ASV) calling, and taxonomic assignment. R packages are available via Bioconductor (www.bioconductor.org) or the Comprehensive R Archive Network (CRAN, cran.r-project.org), unless otherwise noted. Decontam (Ver. 1.12, 77), was used for contamination removal. The UNITE database (Ver. 29.11.2022) was used for taxonomic assignment in fungi (78), and the Maarjam database was used for taxonomic assignment of AM fungi (79). Illumina bacterial reads were processed using the nf-core/ampliseq pipeline version 2.5.0 (80, 81), using the default settings. This pipeline runs adapter removal, primer trimming, read quality filtering, pair-end merging, dereplication, and chimera removal. ASV inference and taxonomic assignments were conducted using dada2 (75) within nfcore/ampliseq, and the SILVA database (Ver. 138.1) was used for taxonomic assignment (82). Singletons were removed from all datasets.

Soil health analysis

Differences in soil health measurements were analyzed by siteyear using the split-plot ANOVA function 'sp. plot' in the agricolae package (Ver. 1.3-7, 83). Multi-year trends in yield, SOM, and POXC, starting from 2018, were evaluated for changes after the addition of the cover crop treatments (Figure 1).

Microbial community analysis

Fungal, AM fungal, and bacterial diversity were investigated using alpha diversity measures; specifically, observed richness and Shannon index (84), using the estimate_richness function in the phyloseq package (Ver. 1.22.3, 85). Differences in bacterial, fungal, and AM fungal communities were analyzed using a PERMANOVA multivariate analysis using the adonis2 function with 999 permutations and the distance metrics from the Bray-Curtis dissimilarity in the vegan package (Ver. 2.6-8, 86). Non-metric multidimensional (NMDS) plots were generated using the Bray-Curtis distance (87) in the vegan package.

Pairwise differential abundance was determined using the DESeq2 package (Ver. 1.40.2, 88); here, we compared the CS



Soybean yield and soil health measurements over a five-year period in long-term rotation plots at two sites. (A, B) yield, (C, D) POXC. (E, F) soil organic matter. Trends in measurements are plotted before (2018-19) the addition of the cover crop treatment and after (2021-22), dashed vertical line indicates the timing of the addition of the cover crop treatment. Error bars represent +/- one standard deviation. NWARS (Northwestern Agricultural Research Station), WARS (Western Agricultural Research Station). CS= corn-soybean Rotation (blue lines), CSW= corn-soybean-wheat Rotation (orange lines).

rotation to the CSW rotation and the rye cover crop to the no cover crop controls. Differences were deemed significant if they met the following cutoffs: (i) a log2 fold threshold of 1.5 and (ii) a p value threshold of 0.01, limiting the differential taxa to those that differed by +/- 150% with a 1% false discovery rate.

Co-occurrence networks were created using the NetCoMI package (Ver. 1.1.0, 89) which is available at https://github.com/stefpeschel/ NetCoMi. To reduce noise, the 250 ASVs with the highest abundance were selected for analysis. The cluster_fast_greedy method and the Sparcc correlation estimator (89) were used for network construction. Associations were calculated using the Pearson correlation approach with a threshold of 0.4, including the default normalization and zero handing options. Networks were constructed using the netConstruct function and compared between rotation and cover crop for both years of the study. For correlation analysis, subsets of reads were selected to reduce noise; these included combining reads for indicator species for each organism type, differential abundant species for each organism type, and network modules generated in NetCoMI. A supplemental file (Network Module ASV table.csv) contains the information on which ASVs make up individual modules. Correlation analysis used the corrplot package (Ver. 0.95, 90) and base R stats functions. Figures were generated using BioRender, ggplot2 (Ver. 3.5.1, 91), ggpubr (Ver. 0.6.0, 92), and vegan.

The core bacterial, fungal, and AM fungal group members for the various treatments were determined using the R package microbiome (Ver. 1.23.1, 93). ASVs were filtered into genera, and the total abundancies and frequencies of the samples were ranked. Only genera present in the 95th percentiles of total relative abundance and frequency were considered as core taxa, as described by Cantoran et al. (94). The relative abundance of ASVs was calculated as the count of each ASV divided by the average number of ASVs within each sample. Bacterial and fungal core genera identity and relative abundances by sample were visualized using bubble plots in ggplot2. The core analysis was run by site-year; one combination (WARS – 2021) did not have enough samples after filtering to complete the analysis.

A set of highly abundant, potentially biologically relevant genera (i.e., known to contain pathogenic or beneficial species) representing a

range of frequencies of occurrence were selected for linear mixed model and Bayesian analysis. For fungi these included common soil fungi, potential pathogens or beneficial organisms, Fusarium (95), Trichoderma (96), Mortierella (97), Fusicolla (98), Neoascochyta (99), and Arthobotrys (100). As well as a set of fungi that were highly abundant in the analysis, including Saitozyma, Tetracladium, Solicoccozyma, Penicillium, Clonostachys, Alternaria, Ustilago, and Exophiala. Bacteria genera selected for these analyses included biologically relevant genera in soils or agriculture, based on the literature, Pseudomonas (101), Bacillus (102), Bradyrhizobium (103), Paenibacillus (104), Streptomyces (105), Deinococcus (106). As well as genera that were highly abundant in our samples including Granulicella, Luteolibacter, Nitrospira, Rhodanobacter, Gemmatimonas, Acidibacter, and Acidothermus. AM fungal genera selected included the only genera represented across both years of the dataset, these are biologically relevant as well: Archeaospora, Claroideoglomus, Diversispora, Glomus, and Paraglomus (107). These sets of genera were also used for variance partitioning using the VariancePartition R package (Ver, 1.36.2, 108) to ascertain genera whose variance may be best explained by the soil health factors. This package uses linear mixed models for partitioning the variation in a feature (in our case, fungal and bacterial species counts) based on measured covariates (108). The covariates included fixed categorical variables (year, location, rotation, and cover crop treatment) and random variables (yield, soil protein, soil respiration, POXC, and SOM).

For the Bayesian analysis, genera counts were first fit using a Dirichlet-multinomial distribution with and without correction for zero inflation using the R packages MicroBVS (109), and ZIDM (110). These packages are available at https://github.com/mkoslovsky/ MicroBVS and https://github.com/mkoslovsky/ZIDM, respectively. As the distribution fitting indicated zero inflation was a feature of the data, a Bayesian zero-inflated Dirichlet-multinomial (ZIDM) model was then used to analyze the multivariate compositional counts (assuming excess zeros in the counts). The ZIDM model simultaneously estimates both individual- and population-level genera abundances and covariates associated with microbial abundances and zero-inflation by incorporating a sparsity-inducing spike-and-slab prior (110). All parameter and hyperparameter priors were kept at their defaults. Models were run until convergences were reached, usually between 40,000 and 50,000 iterations. The inclusion of genera-covariate associations in the model was determined using a marginal posterior probability of inclusion (MPPI) of 0.5 for bacteria and 0.45 for fungi. Results were visualized using ggplot2 pheatmap (Ver. 1.0.12, 111), and matsbyname (Ver. 0.6.10, 112).

The code for all analyses in this study is available on GitHub at https://github.com/calconey/freyetal2025.

Results

Impacts of diversified rotations and cover crops on soil health parameters

Soybean yield, POXC, soil protein, percent SOM, and soil respiration were measured to ascertain the impact of diversified rotations and cover crops on soybean productivity and soil health. In 2021 and 2022, split-plot ANOVAs indicated no statistically significant impacts on soybean yield (Table 1). There was a trend of increases in yield at 3 out of 4 site-year combinations for the CSW rotation, but none reached statistical significance. The trends for plots with the cover crop treatment were split (and non-significant), with half trending to higher yields without the cover crop and the other half trending higher with the rye cover crop. Rotational treatments did not impact POXC. POXC was significantly higher without the cover crop in one site-year combination (2022-NWARS, p = 0.001). POXC was generally higher in the no cover crop control at NWARS and trended higher in the rye treatment at WARS. At both sites, all POXC levels were significantly lower in 2021 than in 2022. Soil protein (g_ACE protein/kg soil) levels were similar across rotation and cover crop treatments at NWARS. At WARS, soil protein was significantly higher in the rye cover crop in 2022 and trended higher after the rye cover crop in 2021 (p = 0.03). Percent SOM was significantly increased at NWARS in the no cover crop treatment in 2021 (p = 0.03) and the CS treatment at WARS in 2022 (p = 0.04). Soil respiration was not significantly impacted in site-year combinations but increased significantly at both sites in 2022 over 2021 by about 20-30 µg CO2-C g-1 day-1 each year.

Trends in yield, soil health, and environmental conditions

Soybean yields were steady, on average 4.1 metric tons at NWARS and 4.6 metric tons at WARS, over the five years measured. At NWARS, soybean yields were higher in the CSW rotation compared to the CS rotation in all years but 2022. At NWARS, soybean yields were more variable with respect to rotations, with the CS rotation having higher yields in 2018 and 2021 and the CSW rotation having higher yields in 2019, 2020, and 2022. Within both sites, soybean yields from 2022 were similar to 2018.

POXC measurements decreased over time at both sites, particularly after adding the cover crop treatments at the end of the 2020 growing season. At NWARS, POXC peaked in 2020 at 726.96 mg/kg of soil in the CSW rotation but declined by 207% to 350 mg/kg soil in the CS rotation in 2022. At WARS, POXC followed similar numbers, peaked in 2020 in the CS rotation at 617.1 mg/kg soil and declined by 221% to 278 mg/kg soil by 2022 after adding the cover crop treatments in 2020. SOM remained steady at NWARS, starting at around 5% in 2018 and ending at 5.2% in 2022. SOM changes were similar at WARS, except in 2020, when it dropped to 2.5%, rebounding to 5.5% in 2021, a deviation best explained by a measurement error. At both sites, SOM numbers were similar in 2022 to the time of the first measurements in 2018.

Environmental conditions from 2018 to 2022 are presented in Supplementary Figure S4. Annual mean temperature at WARS ranged from 10 to 12 °C and 11.1 to 12 °C at NWARS. Total annual precipitation at WARS ranged from 72.6 to 113.5 cm. At NWARS, total precipitation ranged from 69.9 to 118.7 cm. Average daily relative humidity ranged from 77.1 to 80.5% at WARS and 77.9 to 82.0 at NWARS. Mean 5 cm soil temperature ranged from 11.1 to 12.6 °C at NWARS and 13.0 to 14.3 °C at WARS.

		2021				2022			
		CS	CSW	No Cover	Rye	CS	CSW	No Cover	Rye
NWARS	Soybean Yield (metric tons/ha)	4.2	4.5	4.4	4.3	4.9	4.8	4.9	4.7
	POXC (mg/kg)	547.3	552.6	557.5	542.5	350.0	425.5	483.3*	292.2
	Soil Protein (g_ACE protein/kg soil)	3.4	3.3	3.3	3.3	2.5	2.7	2.6	2.6
	Soil Organic Matter (%)	5.3	5.4	5.9*	5.1	5.2	5.1	5.2	5.1
	Soil Respiration (µg CO ₂ -C g-1 day-1)	52.6	58.0	56.8	53.7	74.2	73.1	71.2	76.1
WARS	Soybean Yield (metric tons/ha)	5.1	5.1	5.5	5.6	4.5	4.8	4.7	4.7
	POXC (mg/kg)	464.4	464.6	454.8	474.2	326.4	278.2	327.8	276.7
	Soil Protein (g_ACE protein/kg soil)	2.8	3.0	2.8	2.9	3.4	3.8	3.4	3.8*
	Soil Organic Matter (%)	5.4	5.5	5.4	5.5	4.9*	4.5	4.7	4.7
	Soil Respiration (µg CO ₂ -C g-1 day-1)	56.9	52.0	55.1	53.8	89.2	90.4	90.7	88.9

TABLE 1 Soybean yield and soil health impacts under rotation and cover crop treatments.

Asterisk (*), bolding and dark shading indicate a significant difference (*p* <= 0.05) in a split-plot ANOVA. Each year and site were analyzed separately. CS, corn-soybean Rotation; CSW, corn-soybean-wheat Rotation; WARS, Western Agricultural Research Station; NWARS, Northwestern Agricultural Research Station.

Sequencing and bioinformatics summary

For the 2021 fungal sequencing, 136,720 long reads (median of 4,615 per sample) were assembled into 1,237 ASVs with an average read length of 608bp. For the 2022 fungal sequencing, 292,377 long reads (a median of 7,897.5 per sample) were assembled into 2,638 ASVs with an average read length of 610bp. For the 2021 AM fungal sequencing, 320,272 long reads (median of 7,442 reads per sample) were assembled into 377 ASVs with an average read length of 7,442 reads per sample) were assembled into 377 ASVs with an average read length of 798bp. For the 2022 AM fungal sequencing, 322,967 long reads (a median of 7,911 reads per sample) were assembled into 2,163 ASVs with an average read length of 795bp. For the 2021 bacterial sequencing, 1,869,800 reads (median of 54,881 reads per sample) were assembled into 7625 ASVs with an average read length of 253bp. For the 2022 bacterial sequencing, 2,094,082 reads (a median of 60,051 bacterial reads per sample) were assembled into 8,088 ASVs with an average read length of 253bp.

Soil microbial communities – alpha and beta diversity

Location, diversified rotations, and cover crops did not impact soil fungal communities in 2021. In 2022, changes in soil fungal communities were driven by location (Table 2); statistically significant differences between the two sites were detected in observed diversity (p = 0.005) or Shannon diversity index (p =0.01) for fungal populations, but not between rotation or cover crop treatments. (Supplementary Figure S5). According to PERMANOVA the location by rotation interaction was a driver of fungal communities in 2021, the year the first cover crop intervention was made. However, that pattern was not repeated in 2022 (Table 2). Beta diversity patterns were consistent with the PERMANOVA analysis, which showed that location was a primary driver of community structure (Supplementary Figures S8, S9). Rotation was a driver of arbuscular mycorrhizal fungal community structure, in 2021 (Table 2), but not in 2022 (Table 2). There was no difference in AM fungi alpha diversity metrics in location or rotation treatments in 2021 or 2022 (Supplementary Figure S6). The 2021 cover crop treatment drove a significant increase in the Shannon diversity of AM fungi (Supplementary Figure S6E, p = 0.03) which was not maintained in 2022. Beta diversity analysis did not detect an effect of cover crops (Supplementary Figures S8C, D) on AM fungal communities.

Location strongly impacted bacterial communities in both years (Table 2, p = 0.001; Supplementary Figures S8, S9). In 2021, observed diversity and Shannon index were not significantly different between rotation and cover crop treatments (Supplementary Figure S7). In 2022, the observed alpha diversity (p = 0.01) and Shannon diversity index (p = 0.007) were significantly higher at the WARS site (Supplementary Figure S7B). There were no differences in any alpha diversity metrics for rotation or cover crop treatments in 2022.

Differential abundance analysis

In 2021, differential abundance analysis using DEseq2 for fungal communities revealed that 24 ASVs were significantly more abundant in the CS rotation, while 7 ASVs were more abundant in the CSW rotation (Figure 2A, $p \le 0.01$). In the cover crop treatments, 6 ASVs (including *Fusarium*) were significantly more abundant in the no cover crop controls (Figure 2C, $p \le 0.01$). In comparison, 23 ASVs were significantly more abundant under the rye cover crop. In 2022, 18 ASVs were more abundant in the CS rotation, (Figure 2B, $p \le 0.01$), while 25 ASVs were significantly more abundant in the CSW rotation. In the cover crop treatments, 15 ASVs were more abundant in the no cover crop controls, (Figure 2D, $p \le 0.01$), and 8 ASVs were more abundant under the rye cover crop.

Microbial Community	Location	Rotation	Cover	Location: Rotation	Location: Cover	Rotation: Cover	Location: Rotation: Cover
Fungi 2021	0.07	0.751	0.859	0.057	0.026**	0.288	0.588
Fungi 2022	0.001***	0.968	0.309	0.974	0.195	0.871	0.724
AM Fungi 2021	0.068*	0.04**	0.134	0.176	0.284	0.275	0.121
AM Fungi 2022	0.757	0.795	0.308	0.87	0.951	0.918	0.803
Bacteria 2021	0.001***	0.116	0.501	0.312	0.747	0.365	0.423
Bacteria 2022	0.001***	0.314	0.909	0.378	0.636	0.759	0.779

TABLE 2 Permutational Multivariate Analysis of Variance (PERMANOVA) for fungal, arbuscular mycorrhizal (AM) fungal, and bacterial community composition.

Numbers indicate p values from PERMANOVA results for fungal, AM fungi, and bacterial communities at NWARS (Northwestern Agricultural Research Station) and WARS (Western Agricultural Research Station). (*= p < 0.1 (italics), ** = p < 0.05, *** = p < 0.001 (bold)).

In 2021, differential abundance analysis for AM fungal communities revealed 7 ASVs more abundant in the CS rotation and 7 ASVs more abundant in the CSW rotation (Figure 3A, $p \le 0.01$). Comparing the cover crop treatments, 10 ASVs were more abundant in the no cover crop control, and 14 ASVs were more abundant under the rye cover crop treatment (Figure 3C, $p \le 0.01$). In 2022, 7 ASVs were more abundant in the CSW rotation (Figure 3B, p <= 0.01). In the cover crop treatments, 18 AM fungi fungal ASVs were more abundant in the no cover crop control (Figure 3D, p <= 0.01), while 19 ASVs were more abundant in the rye cover crop treatment.

For bacterial communities, differential abundance analysis in 2021 revealed one ASV that was significantly more abundant in the CS rotation and one ASV that was more abundant in the CSW rotation (Supplementary Figure S10A, $p \leq 0.01$). In the cover crop treatment, only two differentially abundant ASVs were identified, both of which were more abundant in the no cover crop control (Supplementary Figure S10C). In 2022, one bacterial ASVs was differentially abundant in the CSW rotation and one ASV was differentially abundant in the CSW rotation (Supplementary Figure S10C). No differentially abundant ASVs were identified in the cover crop treatments in 2022.

Network analysis and module correlations

Association networks between fungal, AM fungal, and bacterial ASVs recovered from CS and CSW rotations (Figure 4) and cover crop treatments (Figure 5) were estimated for each year of study. Network metrics (Supplementary Tables S2–S4) varied for rotation and cover crop treatments, especially between years and the type of organism sampled. Fungal and AM fungal communities had more modules in 2021 than in 2022, containing several smaller groups of interconnected ASVs, probably due to differences in sequencing runs (Supplementary Table S2). Bacteria communities generally had fewer network modules than fungi or AM fungi. Fungal communities had higher modularity values than AM fungi and bacteria, indicating a higher possibility of community connectedness. AM fungi communities had high variation in modularity and edge density compared to fungi and bacteria,

with values that were consistently in the same range. Bacterial communities generally had the highest edge density, although some groups of AM fungi also had high edge density. More closely associated modules tended to contain groups associated with WARS or NWARS, such as the two large groupings in bacterial communities (Figure 4).

To determine if any trends could be detected using groups of associated organisms, ASVs from individual fungal, AM fungal, and bacterial network modules were averaged, and a correlation analysis was conducted with yield and soil health parameters (Supplementary Figures S11, S12). Three fungal modules and one bacterial module in 2021 suggested a positive correlation with POXC (Supplementary Figure S11B) (Figures 6A-D). These three modules contain similar taxa, including Ustilago, Humicola, and Exophiala. In fungal communities, in 2022, there was a trend of stronger negative associations with soil protein in modules containing Lacrymaria, Mortierella, and Ustilago ASVs. Bacterial communities in 2021 had some large modules that showed a positive trend: four modules with yield, four with POXC, and six with soil protein (Supplementary Figure S11E). In 2022, the predominant pattern for bacterial modules was that eleven out of sixteen modules were positively associated with soil protein and nine out of sixteen modules were positively associated with respiration. In contrast, most of the bacterial modules from 2022 were weakly and negatively associated with yield, POXC, and SOM. ASVs identified as Sphingomonas, Bacillus, Gaiella, and Acidothermus were some of the more abundant bacteria in these modules. Some of the strongest positive (Supplementary Figures S13A, B) and negative (Supplementary Figures S13C, D) associations were extracted to illustrate these trends.

Core taxa

Nine fungal taxa were identified as core taxa in the rotation and cover crop treatments in 2021, and ten were identified as core taxa in 2022. From this *Ustilago*, *Mortierella*, and *Fusarium* were most abundant in the rotational and cover crop treatments in 2021 (Supplementary Figure S14). and *Papulaspora*, *Lacrymaria*, and one unidentified fungal ASV in 2022. In both years there were no



significant differences in the largest groups of core fungal taxa between rotational or cover crop treatments at either site. Because of some dropout samples, we could not conduct this analysis on AM Fungi.

Twenty-five bacterial genera were identified as core taxa for rotation treatments and cover crop treatments in 2021, and twentysix were identified in 2022. Core bacterial taxa in the rotational and cover crop treatments in 2021 included *Gemmatimonas*, *Sphingomonas*, and *Bryobacter* at NWARS in 2021 and *Candidatus* Uedobacter, *Gemmatimonas*, and *Sphingomonas* at WARS in 2021. Core bacterial taxa in the rotational and cover crops treatments in 2022 included *Gemmatimonas*, *Candidatus* Uedobacter, *and Sphingomonas* at NWARS *and Gemmatimonas*, *Bacillus*, and *Acidothermus* at WARS. In both years there were no significant differences in the largest groups of core bacterial taxa between rotational or cover crop treatments at either site.

Variance partitioning

For the selected subset of fungal genera, year was the most significant contributor to the variance (19%) in taxa counts (Supplementary Figure S15A). Of the soil health characteristics, soil respiration was the most significant contributor to the variance of the selected fungi. *Mortierella* and *Fusicolla* had significant variation explained by soil respiration differences (Figure 7A). *Tetracladium* had a larger variance explained by soil protein. For

AM fungal genera (Figure 7B), year was the most significant contributor to the variance (9%) (Supplementary Figure S15B). Significant contributions to *Paraglomus* variance were explained by soil respiration and POXC. For the bacterial genera in the variance partitioning analysis, year was the largest contributor to variance (43%, Supplementary Figure S15C). Location explained the largest percentage of variance for some genera, including *Bacillus* and *Paenibacillus* (Figure 7C). Yield contributed 12% of variation to both *Pseudomonas* and *Bacillus*. Soil respiration contributed 21% of the variation to *Nitrospira*. Other measured covariates accounted for only small portions of the variation for each genus we measured. Overall, the variation contributed by our covariates was lower for AM fungi than for the fungi or bacteria. Soil respiration contributed to variation within all the microbial groups, whereas soil organic matter only for variation within fungi.

Bayesian analysis

A zero-inflated Dirichlet-multinomial linear regression model correcting with Bayesian variable selection (110), identified subsets of fungal, AM fungal, and bacterial genera associated with the measured environmental and soil covariates. Model convergence and covariate inclusion using a marginal posterior probability of inclusion (MPPI) are shown in Supplementary Figures S16 and S17. The following associations were identified for fungal genera at a cutoff of 0.5 MPPI (Figure 8A): *Fusarium* was positively associated



with clay soils; *Saitozyma* was positively associated with higher air temperatures; and *Tetracladium* was associated with higher soil temperatures. AM fungal genera *Claroideoglomus* and *Paraglomus* were associated with higher air temperatures, and *Glomus* was associated with 2022 (Figure 8B). Several associations were identified for bacterial genera at a cutoff of 0.45 MPPI (Figure 8C). *Acidothermus* was positively associated with the no cover crop control and the rye cover crop treatment, location, and weakly positively associated with soil protein. *Bacillus* was positively associated with air temperature and rain. *Gemmatimonas* were positively associated with clay content and 2021 and negatively associated with sand content, WARS, and air temperature. *Luteobacter* was positively associated with air temperature.

Discussion

The current paper builds upon previous studies of a long-term experiment on crop rotation by expanding the rotation's diversity due to adding a rye cover crop before soybean (20–22). There were no significant changes in soybean yield in the two years after adding the cover crop. Soybean yield was stable between 2018-2022. This yield stability is consistent with our hypothesis that adding a rye cover crop to a CSW rotation before soybean will not decrease soybean productivity. Previous studies have reported mixed effects of the impact of rye cover crops on soybean crops. Overmyer et al. (29)

found that a rye/oat cover crop positively affected soybean yield in Ohio when combined with late cover crop termination. Early sowing of soybeans after rye termination significantly negatively affected soybean yield, while later planting dates did not impact yield (113). Other studies have shown no impact (25) or a negative effect (58, 114) on soybean yield. Rye cover crops may not impact overall soybean productivity but do have other positive benefits that may warrant their addition to rotations, including helping to deal with excess nitrogen (115), nitrogen runoff (116, retention of soil moisture (25), and weed suppression (117). Another possibility is that any positive changes that may be achieved with the cover crop may not have had time to establish after one cycle, and any impact on yield may lag beyond that even further (118). Many studies showing the positive benefits of cover crops looked at the situation in the timeframe of a decade after the implementation of the cover crop, while our study has undergone just one cycle (25).

The current study did not detect any further improvements in soil health indicators since the 2019 measurements; any subsequent effects of the rotations were not detected with the measured parameters, with SOM significantly decreasing at WARS in 2022. Some changes were observed with the addition of the cover crop treatment, including a significant decrease in POXC at NWARS in 2022, an increase in soil protein at WARS in 2022, and a decrease in SOM at NWARS in 2021. POXC and SOM were measured at each site over the five years the plots have been monitored. There was a consistent trend of decline in POXC levels after the addition of the cover crops. POXC measurements declined by over 200% at both



sites between 2020 and 2022 and although declines were seen in treatments with and without cover crops, the decline in the cover crops was higher. This contrasts with our hypothesis that adding the cover crop would positively affect soil health qualities, including POXC, soil protein, soil respiration, and SOM. The CSW rotation had previously been observed to increase SOM at one of our sites (20), and this has also been observed in other studies utilizing a rye cover crop in a shorter rotation (119). It also contrasts with other studies that have reported increased (120) or no impact of cover crop on POXC (121). The impacts on POXC were unlikely due to changes in weather, as temperature and precipitation trends at both sites have been stable throughout the last five years and are within the climate norms for the region. There may be changes in how carbon is partitioned after adding the cover crops, with carbon cycling out of the POXC fraction and into another (122). Five-year trends in SOM have been relatively consistent at the studied sites, starting at around 5% at both sites in 2018 and still around 5% in 2022. Tyler (58) reported increased SOM by adding cover crops. We did not observe any changes with rotation or cover crops in the present study. Previous studies looking at diversified rotations (123) and cover crops (56) have reported increases in soil carbon. However, we did not observe any consistent changes in soil respiration in these plots. Positive changes in soil health metrics due to cover crops may take more years to stabilize or may be buffered by the soil at our locations (124). We may not have seen these changes after the initial year of cover crop introduction due to the buffering effects of soil, the large influence of the cash crop, or our application of no-till. Alternatively it is possible that these are not the right indicators for detecting the impact of the cover crop. Other indicators for soil health that have been proposed for measuring impacts of management changes, including cover cropping, include soil organic carbon (125), bulk density, soil structure (55), and microbial biomass carbon (126).

Location was the dominant driver of any differences observed in our microbial communities. Although there were changes in the abundance of individual bacterial, fungal, and AM fungal ASVs observed using DESeq2 analysis, few of these patterns were consistent year-over-year. *Fusarium* ASVs were more abundant in the CS rotation in 2021, although this pattern was not repeated in 2022. We had previously identified *Fusarium* as an indicator species



of the CS rotation in 2018 and 2019 (22) along with other potential plant pathogens (Neoascochyta and Aphanomyces). However, these differences could be due to differences in the indicator species and DeSeq analysis. In the first year of the rye treatment, rye did not influence overall Fusarium reads. However, several individual Fusarium ASVs were higher in either the no cover crop control or the rye treatment; perhaps the impact on Fusarium will only be observed after several years of the cover crop treatment. Differences in individual AM fungal genera were challenging to parse, as many ASVs are identified to the same genus, and most of these genera have ASVs that are differentially abundant in both rotation and cover crop treatments. The resolution of our analysis may not be sufficient to track these changes, or a more detailed analysis of individual ASVs may be required to identify oligotypes that could be grouped together. A few ASVs were differentially abundant, including multiple Clairoideoglomus ASVs in the CS and CSW rotation in 2021 and 2022, a single Diversispora ASV in the CSW rotation, and the no cover crop control in 2021. AM fungi are important to promote in soil as they are known to assist crop species in resisting environmental stress. This has been shown in *Clairoideoglomus* (127) and other species (128). They are also known to help plants acquire phosphorous; this has been shown in *Diversispora* (129) and other species (130). Bacterial communities contained very few differentially abundant species, indicating little impact of the rotation or cover crop treatment on bacterial communities. Although there was some variation in core taxa by site, there was minimal variation in core taxa over rotational or cover crop treatments. Overall, the microbial community results are consistent with our hypothesis that adding the cover crop would impact fungal, AM fungal, and bacterial communities. Microbiome variation is often driven by regional differences in climate and soil (131). Cover crops have been reported to influence soil fungal communities (132), AM fungal communities (133), and soil bacterial communities (134).

Location and year were consistently the highest contributors to the variance of different genera of fungi and bacteria. Soil respiration explained 20% of variance in *Nitrospira* sp. bacteria. *Nitrospira* is involved in nitrogen transformation in soil (135) and so plays a role in N availability in soil, which can, in turn, affect respiration (136). Rotation was a source of variance in *Bacillus* and



FIGURE 6

Fungal (A, B, D) and bacterial (C) reads from amplicon sequencing variants (ASVs), which comprise associated network modules plotted against permanganate oxidizable carbon (POXC) levels in the soil. Points represent average reads in each sample; a dashed black line shows the smoothed trend, the gray shaded area is a 95% percent confidence interval for the trend curve, and the blue line shows a linear regression. The linear correlation coefficient (*R*) is shown in the upper left of each panel.



FIGURE 7

Sources of variation in amplicon sequencing variant (ASV) counts for selected fungal (A), arbuscular mycorrhizal fungal (B), and bacterial (C) genera. CS, corn-soybean rotation; CSW, corn-soybean-wheat rotation; WARS, Western Agricultural Research Station; NWARS, Northwestern Agricultural Research Station.



Luteolibacter. Management choices that increase these species should be considered as both are potential plant symbionts (137), and Bacillus are notably well characterized for their plant growth promotion (138) and disease suppression activity (139). In fungi, soil respiration contributed about 5% of the variation to Mortierella and Penicillium. Mortierella and Penicillium are common soil fungi involved in ecosystem services (140) and plant growth promotion (141). Cover crops were a small but significant (4%) contributor to variance in Tetracladium. Tetracladium are most well known as root endophytes and are known to assist with phosphate solubilization (142). Bayesian association analysis also identified possible covariates that are important for these genera. These results are consistent with our hypothesis that changes in microbial communities would be associated with soil health changes we observed. Microbial communities are associated with changes in many soil health indicators (60).

We combined and averaged ASVs from modules identified by network analysis for an alternative attempt understanding whether groups of associated fungi or bacteria impact soybean yield or soil health properties While many of these modules showed slight or insignificant correlations with yield and soil health parameters, some showed potential trends. Several modules from the 2021 fungal network analysis showed a consistent positive trend with increasing POXC. These modules were dominated by a few ASVs, particularly an ASV identified as *Ustilago*. *Ustilago* is a common plant pathogen that is the causal organism of smut disease on aerial portions of maize. There is evidence that *Ustilago* can also infect the roots of some plants (143), and at least its teliospores are soil-borne. Although *Ustilago* is known to manipulate plant carbon metabolism in above-ground portions of the plant (144), there is no known connection to soil carbon. In 2022, another consistent trend was an increase in almost all bacterial network modules as soil protein and respiration increased. Bacteria contribute to carbon and nitrogen dynamics in soil (145).

Any changes or lack thereof in soil health and microbial communities must be discussed in the context that the current study looked at the first year after implementing the cover crop plots. It may take up to 10 years before significant changes are observed after implementing a change in agricultural practices (25, 146). The diversified rotation reached 10 years in the field in 2024, while the cover crop will have completed two repetitions only in each plot at the time of publication. The decision to include no-till as a part of this rotation and cover crop scheme likely contributed to the lack of impact the treatments had on yield, as no-till is known to widely impact yield and soil health (147). Another pitfall in this experiment is related to the differences in the structure of microbial networks generated across the years. Although the library preperation and sequencing were done by the same facility, and using the same primers, coverage from each experiment was different. This likely contributes to some variation between years in our experiments.

In conclusion, the initial impact on soil health and soil microbial communities of a rye cover crop interjected into an established no-till rotation was minimal. Location, encompassing site-specific weather and soil qualities, seems to be a more critical factor in determining microbial communities (148). Any potential benefits of adding a cover crop to a CSW rotation may not be observed soon after the initial intervention. Thus, multiple years of implementing cover cropping may be needed before observing any benefits in soil health. This will have to be considered when calculating the economic cost of such a change in management strategy. However, there are other benefits that we did not measure here, such as improvements in N runoff and weed suppression, which may increase the economic viability of cover crops in CSW production.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: https://www.ncbi.nlm. nih.gov/, PRJNA1190478.

Author contributions

TF: Data curation, Formal analysis, Investigation, Methodology, Software, Visualization, Writing – original draft, Writing – review & editing, Conceptualization. DS: Formal analysis, Methodology, Software, Visualization, Writing – original draft, Writing – review & editing, Conceptualization. LL: Conceptualization, Project administration, Writing – review & editing, Methodology, Investigation. CS: Conceptualization, Investigation, Methodology, Resources, Writing – review & editing. HL-N: Formal analysis, Writing – review & editing. MB: Conceptualization, Funding acquisition, Investigation, Methodology, Project administration, Resources, Supervision, Writing – original draft, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fsoil.2025.1535734/ full#supplementary-material

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