Check for updates

#### **OPEN ACCESS**

EDITED BY Elena Velickova, Saints Cyril and Methodius University of Skopje, North Macedonia

REVIEWED BY Ahmed A. Zaky, National Research Centre, Egypt José Armando Ulloa, Autonomous University of Nayarit, Mexico

\*CORRESPONDENCE Seydi Yıkmış 🖂 syikmis@nku.edu.tr Moneera O. Aljobair 🖂 moaljobair@pnu.edu.sa

RECEIVED 27 March 2025 ACCEPTED 16 June 2025 PUBLISHED 02 July 2025

#### CITATION

Yıkmış S, Türkol M, Tokatlı Demirok N, Al B, Mohamed Ahmed IA and Aljobair MO (2025) Advancing sustainable juice processing through thermosonication: functional enrichment of apple juice with pollen.

Front. Sustain. Food Syst. 9:1601419. doi: 10.3389/fsufs.2025.1601419

#### COPYRIGHT

© 2025 Yıkmış, Türkol, Tokatlı Demirok, Al, Mohamed Ahmed and Aljobair. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Advancing sustainable juice processing through thermosonication: functional enrichment of apple juice with pollen

## Seydi Yıkmış<sup>1</sup>\*, Melikenur Türkol<sup>2</sup>, Nazan Tokatlı Demirok<sup>2</sup>, Berna Al<sup>1</sup>, Isam A. Mohamed Ahmed<sup>3</sup> and Moneera O. Aljobair<sup>4</sup>\*

<sup>1</sup>Department of Food Technology, Tekirdağ Namık Kemal University, Tekirdağ, Türkiye, <sup>2</sup>Department of Nutrition and Dietetics, Tekirdağ Namik Kemal University, Tekirdag, Türkiye, <sup>3</sup>Department of Food Science and Nutrition, College of Food and Agricultural Sciences, King Saud University, Riyadh, Saudi Arabia, <sup>4</sup>Department of Sports Health, College of Sports Sciences & Physical Activity, Princess Nourah Bint Abdulrahman University, Riyadh, Saudi Arabia

The development of functional beverages requires the use of sustainable technologies to increase both nutritional value and consumer appreciation. This study aims to investigate the effects of thermosonication, a sustainable food processing approach, on Uruset apple juice to produce a functional beverage enriched with pollen. In the study, four process parameters-temperature, time, amplitude, and pollen concentration—were optimized. As a result of the experimental design using response surface methodology (RSM), significant increases were observed in total phenolic substance (TPC), total flavonoid (TFC), and antioxidant capacity [DPPH (2,2-diphenyl-1-picrylhydrazyl) and CUPRAC (copper reducing antioxidant capacity)] values. At the same time, significant improvements were observed in the content of phenolic compounds, including chlorogenic acid, gallic acid, guercetin, and catechin. Thermosonication process partially suppressed the enzymatic activity (PPO) and ensured the preservation of phenolic compounds and positively affected the shelf life of the product. Sensory analysis results showed that apple pollen juice processed with thermosonication received higher scores in terms of taste, odor, and general liking compared to both the control and pasteurized groups. Optimum process conditions were determined to be 40.9°C temperature, 5.63 min process time, 61.01% amplitude, and 60 mg/100 mL pollen. The findings reveal that a sustainable, functional beverage with enhanced nutritional and sensory quality can be developed through the synergistic effect of thermosonication and a natural pollen additive. This approach offers an innovative and environmentally friendly solution that can be applied on an industrial scale as an alternative to traditional heat treatments. In the future, the production of functional beverages supported by innovative processes such as thermosonication could help to promote healthier and more environmentally friendly products within the food industry.

#### KEYWORDS

sustainable food processing, apple juice, functional beverage, pollen, thermosonication, phenolic compounds

# **1** Introduction

Fruit juices are important sources of bioactive compounds such as vitamin C, carotenoids, and phenolics. The world's most consumed fruit juice, particularly in Europe and North America, is apple (Franz et al., 2009; Lu et al., 2010). Uruset apple juice (red inside and out) contains more bioactive compounds than other apple juices (Yıkmış, 2019; Coşkun and Aşkın, 2016; Abacı and Sevindik, 2014). Ultrasonication is a term defined as "the generation of energy by sound waves that vibrate at 20,000 or more vibrations per second" (Lima et al., 2015; Zhang et al., 2016). Ultrasonic equipment generally operates at frequencies between 20 kHz and 10 MHz. Fruit juices are susceptible to deterioration from heat, micro-organisms, enzymes, oxygen and light during processing and storage (Liang et al., 2006). Compared to heat treatment, ultrasonication inactivates pathogenic microorganisms and enzymes that cause spoilage. In addition, the low temperature applied in the process helps preserve the product's taste, smell, texture, and nutrients. The result is a product with characteristics very close to its fresh form (Tiwari and Mason, 2012; McClements, 1995; Leighton, 2007; Chandrapala et al., 2012). The effect of the ultrasound process can be increased by using temperature, pressure, or both. The combination of ultrasound with moderate heat is referred to as thermosonication; the combination with pressure is termed manosonication; the use of ultrasound in conjunction with ultraviolet light is designated photosonication; and the combination of ultrasound with both temperature and pressure is referred to as manothermosonication (Chemat et al., 2011; Şengül et al., 2011). Pollen, a natural ingredient, was added in this study to increase the nutritional content and functional properties of apple juice. Bee pollen is a mixture of flower pollen from various botanical sources, collected by bees and agglutinated by enzymes and nectar, and is a valuable product of the beehive (Bertoncelj et al., 2023). Bee pollen is a good food for bees and people. It consists of 13-55% carbohydrates, 10-40% protein, 0.2-3.0% mineral substances, 1-12% lipids, 0.3-20% fibre, and other organic compounds (İzol et al., 2025). The flavonoids found in bee pollen include kaempferol, quercetin, delphinidin, luteolin, naringenin, galangin, isorhamnetin, rutin, pinocembrin, catechin and apigenin (Yildiz and Maskan, 2022).

In a study investigating the effect of thermosonication treatment (20 kHz; 30 µm; 5 min; 30°C) on bioactive compounds of watermelon (Citrullus lanatus cv.) juice, it was found that the coefficient of determination (R<sup>2</sup>) for the estimated quality parameters showed good correlation with the experimental data at a confidence level of 95% (Rawson et al., 2011). In another study, the application of thermosonication (15 and 25 min, 80% amplitude) on the shelf life and the antioxidant, microbiological, and physicochemical properties of prickly pear (Opuntia ficusindica) juice was investigated. It was reported that after 14 days of storage, especially for 80% and 25 min treatment, the phenolic content increased, and there was an increase in antioxidant activity [2,2-azinobis (3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) and DPPH] until the end of storage (Del Socorro Cruz-Cansino et al., 2015). Carrot juice was sonicated at 50°C, 54°C, and 58°C for 10 min (24 kHz, 120 µm amplitude) (acoustic power 2204.40, 2155.72, 2181.68 mW/mL, respectively). The researchers evaluated the thermosensation process as a promising technology

to minimize physicochemical changes during storage, delay microbial growth, and improve the quality of carrot juice by preserving bioactive compounds (Martínez-Flores et al., 2015). Thermosonication is a new and applicable technique used instead of traditional heat treatments. It has been reported that ultrasonication is generally more effective when combined with moderate heat. It has been stated that this combined process increases microbial and enzymatic inactivation with heat and cavitation; this is achieved without altering the quality of the juice (Anaya-Esparza et al., 2017).

After the literature review, Uruset apple juice was preferred because of the high bioavailability of Uruset apple juice and the fact that there is no study on thermosonication on Uruset apple juice. This study aims to develop Uruset apple juice as a functional beverage with the addition of pollen. Concurrently, response surface methodology was employed to optimize the beverage's bioactive components and antioxidant properties. Furthermore, a comprehensive comparative analysis was conducted between pollen-fortified thermosonicated apple juice and both thermally pasteurized and untreated control samples. The evaluated parameters included: bioactive characteristics [phenolic compound composition and polyphenol oxidase (PPO) activity], physicochemical properties (pH, titratable acidity and soluble solids content), biochemical composition (sugar content and organic acid profile), as well as sensory attributes.

# 2 Materials and methods

## 2.1 Material

The research used Uruset apple grown in the Ardahan region as material. Analyses were conducted in Tekirdağ Namık Kemal University Çorlu Vocational School Food Technology Laboratories.

# 2.2 Preparation of Uruset apple juice concentrate and pollen addition

Uruset apple juice was first made by breaking fresh Uruset apple fruits in a blender (ISO LAB Blender, 602.21.001) at low speed for 30 s and obtaining fruit pulp. The mixture was filtered. The prepared concentrates were stored in 50 mL sterile sample containers with lids at  $-18^{\circ}$ C. Pollen samples were obtained from different beekeepers operating in Tekirdağ and were used in the analyses after being mixed and homogenized.

# 2.3 Experimental design of response surface methodology

Uruset apple juice was analyzed using the response surface method Minitab Statistical Analysis Software (Minitab 18.1.1) to understand the effect of thermosonication application on quality parameters. Response surface method (RSM) was used. A fivelevel, three-factor central composite design was chosen as the experimental design. There are 31 trial points for optimization (Tables 1, 2). The adequacy of the model was assessed by

Independent variables			Level		
	-1.68	-1	0	1	1.68
X <sub>1</sub> : Temperature (°C)	30	35	40	45	50
X <sub>2</sub> : Time (min.)	2	4	6	8	10
X3: Amplitude (%)	40	50	60	70	80
X <sub>4</sub> : Pollen (mg/100 mL)	40	50	60	70	80

TABLE 1 Independent variable values and their corresponding proportions used in response surface method.

considering the  $R^2$  and the adjusted  $-R^2$  coefficients, the lack of fit tests, and the ANOVA results. Independent variables were determined as temperature (X<sub>1</sub>), time (X<sub>2</sub>), amplitude (X<sub>3</sub>), and pollen (X<sub>4</sub>). Dependent variables were selected, such as phenolic substance, flavonoid substance, and antioxidant values. The second-degree-polynomial equation shown in the following equation was used to create the model Equation 1:

$$y = \beta_0 + \sum_{i=1}^{3} \beta_i X_i + \sum_{i=1}^{3} \beta_{ii} X_i^2 + \sum_{\substack{i=1\\i < j}}^{3} \sum_{i=1}^{3} \beta_{ij} X_i X_j \qquad (1)$$

In the equation, Y is the dependent variable,  $\beta o$  is the intercept term,  $\beta i$  is the first-degree (linear) equation coefficient,  $\beta ii$  is the second-degree equation coefficient,  $\beta_{ij}$  is the two-factor cross-interaction coefficient, and X<sub>i</sub> and X<sub>j</sub> are the independent variables.

### 2.4 Determination of bioactive compounds

The total phenolic content amount was determined spectrophotometrically using the Folin–Ciocalteu method with modifications. The absorbance was measured with a UV–VIS spectrophotometer (SP-UV/VIS-300SRB, Spectrum Instruments, Victoria, Australia) at a wavelength of 765 nm. Subsequently, the findings were expressed as milligrams of gallic acid equivalents per liter (mg GAE/L) (Zhishen et al., 1999). Total flavonoid content was determined by the aluminum chloride colorimetric analysis method. The total content of flavonoids is expressed in mg of quercetin equivalents (CE/L) (Singleton et al., 1999). The CUPRAC (Cu (II) ion-reducing antioxidant capacity) method has been described in a previous study by Apak et al. (2006), and the DPPH scavenging activity method, as described by Singh et al. (2002), was used to determine the antioxidant capacity of Uruset apple juice. Analyses were performed in triplicate.

### 2.5 Physicochemical analysis

The temperature of the samples to be measured for pH was brought to +20°C, and the pH analysis of the samples was performed using the Hanna Instruments HI 2002 pH/ORP device (Cemeroğlu, 2010). Analyses were performed in triplicate. The refractometric method determined the amount of dissolved dry matter (Cemeroğlu, 2010). Measurements were made at 20°C using a PCE-032 brand model refractometer, and the results were expressed as °Brix. Analyses were performed in triplicate. Titratable acidity was determined potentiometrically by titrating the samples with 0.1 N NaOH solution to pH 8.1 (Sadler and Murphy, 2010). Analyses were performed in triplicate.

### 2.6 Analysis of organic acid and sugar

The amount of organic acids in the samples was determined according to the method specified by Castellari et al. (2000). The method was established by preparing standards in the expected concentration ranges for organic acids (citric, malic, ascorbic) and sugars (sucrose, glucose, fructose). Citric and malic acids from organic acids were determined at 210 nm and ascorbic acid at 243 nm with the diode array detector (DAD detector). Results are given as mg/100 mL for organic acid. The amount of sugar in the samples was determined according to the method specified by Sturm et al. (2003). Sugars were determined in the refractive index detector (RID detector). Sugar results are given in g/100 mL.

## 2.7 Analysis of phenolic compounds

Phenolic compounds were analyzed using an Agilent 1,260 Infinity chromatograph with a diode array detector (DAD). As outlined in the study by Portu et al. (2017), the chromatography process was carried out utilizing a C-18 Agilent column ( $250 \times 4.6 \text{ mm}$ ; 5 µm packing). The column temperature was fixed at 30°C with a flow rate of 0.80 mL/min. Detection was carried out at 280, 320, and 360 nm. The concentrations of these compounds are expressed as µg/mL. The results for phenolic compounds are given as the average of the analyses of three samples.

# 2.8 Analysis of polyphenol oxidase enzyme (PPO) activity

0.1 mL of Uruset apple juice and 2.8 mL (0.1 M, pH = 6.8) Na-phosphate buffer solution was added to the cuvette in the spectrophotometer cuvette holder. The solution was left for 3 min to extract PPO. At the end of the incubation period, 0.1 mL (0.1 M) catechol solution was added and mixed, and absorbance measurement was started immediately at 410 nm. Absorbance measurements were recorded at 15-s intervals, and a linear absorbance-time graph was drawn using the data obtained for 5 min. Using the slope of the graph, PPO enzyme activity was calculated as Unit/mL (U/mL) as specified in Cemeroğlu (2010).

#### TABLE 2 Measured responses used in the experimental design for RSM.

Run no.		Independe	nt variables		Dependent variables							
Temperature		Time (X <sub>2</sub> )	Amplitude	Pollen	TPC (mg GAE/L)		TFC (mg CE/L)		DPPH (inhibition %)		CUPRAC (inhibition %)	
	(X <sub>1</sub> ) (°C)	(min.)	(X <sub>3</sub> ) (%)	(mg/100 mL) (X₄)	Experimental data	RSM predicted	Experimental data	RSM predicted	Experimental data	RSM predicted	Experimental data	RSM predicted
1	35	4	70	70	861.44	860.77	344.58	344.63	55.40	55.65	59.72	59.98
2	35	4	70	50	856.58	857.16	342.63	342.98	58.07	58.14	60.11	60.03
3	45	4	70	70	874.72	875.42	349.89	349.97	59.30	59.44	62.37	62.15
4	40	6	60	60	870.56	870.42	355.62	355.51	60.27	60.27	64.02	64.24
5	40	6	60	60	870.56	870.42	355.62	355.51	60.27	60.27	64.14	64.24
6	40	6	60	60	870.48	870.42	355.34	355.51	60.23	60.27	64.37	64.24
7	30	6	60	60	862.16	862.34	346.75	346.08	59.27	59.08	61.81	61.55
8	40	6	40	60	864.07	863.66	346.65	346.03	59.38	59.19	61.44	61.07
9	40	6	60	60	870.56	870.42	355.62	355.51	60.27	60.27	64.38	64.24
10	35	4	50	50	867.57	867.40	347.03	347.45	59.61	59.91	60.87	60.98
11	45	8	50	70	853.78	853.95	341.51	341.55	57.88	57.95	59.91	59.80
12	40	10	60	60	859.27	859.70	343.71	343.60	57.27	56.99	60.73	60.98
13	40	6	80	60	865.86	866.35	346.34	346.60	58.7	58.42	60.18	60.51
14	45	8	50	50	860.12	860.30	344.05	344.13	54.31	54.52	60.36	60.23
15	50	6	60	60	868.84	868.73	347.54	347.84	58.90	58.62	60.4	60.63
16	45	8	70	50	867.54	867.53	347.02	347.15	56.33	56.59	60.88	60.52
17	40	6	60	60	870.12	870,42	355.24	355.51	60.21	60.27	64.32	64.24
18	40	6	60	40	865.13	865.22	346.05	345.80	57.66	57.36	60.71	60.84
19	45	4	50	50	866.72	867.26	346.69	346.87	58.76	58.85	59.73	59.88
20	45	8	70	70	867.21	866.88	346.88	346.60	58.79	58.95	60.16	60.19
21	40	2	60	60	867.43	867.07	346.97	346.72	58.81	58.61	60.87	60.60
22	40	6	60	60	870.12	870.42	355.62	355.51	60.27	60.27	64.39	64.24
23	35	8	70	50	863.66	863.22	345.46	345.44	58.55	58.78	61.61	61.61
24	35	8	50	70	860.12	860.00	344.05	344.49	59.10	59.41	61.36	61.62
25	35	4	50	70	864.55	865.31	346.82	347.07	58.61	58.49	60.67	60.83
26	35	8	70	70	860.14	860.35	344.06	344.26	58.31	58.36	60.35	60.01
27	45	4	50	70	867.43	867.38	346.97	347.13	59.98	60.21	60.87	61.00
28	35	8	50	50	868.51	868.57	347.40	347.70	58.77	58.77	63.31	63.33
29	40	6	60	80	862.49	862.48	345.00	344.88	58.47	58.30	60.53	60.36

10.3389/fsufs.2025.1601419

CUPRAC (inhibition %)

DPPH (inhibition %)

Dependent variables

RSM

Experimental

RSM

erimental

Exp

RSM

ental

БХр

RSM

Experimental

X<sup>4</sup>)

PC (mg GAE/L)

FC (mg CE/L)

data

64.24 60.93

64.14 61.05

60.27 59.13

60.23 58.98

355.51 347.68

355.34 347.99

870.42 869.60

870.12 869.97

70 60

4 6

355.58 441.87 %0.24

871.04 682.63 27%

50.00

61.01

3.63

40 45 40.90

TS-UJ

Experimental values % Difference

60 50 64.14

67.51 %4.9

60.27 61.80 %0.02

## 2.9 Sensory analysis

Ultrasound-treated pollen-fortified Uruset apple juice was compared with thermal pasteurized apple juice and fresh Uruset apple juice. The study was conducted in two replicates. Panelists were asked to evaluate the samples' taste, odor, color, texture, and general acceptability. Fifty trained panelists participated in the sensory evaluation of the juice samples. All samples were coded using a random three-digit alphabet. Sensory properties were determined using a 9-point hedonic scale (0–9). Scale scores: excellent, 9; very good, 8; good, 7; acceptable, 6; poor (initial odorless, tasteless development) < 6; lower score accepted 6. The product was considered unacceptable after the first odor or tastelessness (Portu et al., 2017).

## 2.10 Statistical analysis

Statistical analyses of the comparative analyses of the study were performed using the SPSS 20.0 (SPSS Inc., Chicago, United States) program. Samples were compared using the One-way ANOVA multiple comparison-Tukey test. The statistical significance level was determined as p < 0.05. The response surface method was analyzed using Minitab Statistical Analysis Software (Minitab 18.1.1).

## **3** Results and discussion

### 3.1 Optimization of bioactive compounds

This study evaluated the effects of temperature  $(X_1)$ , time  $(X_2)$ , amplitude  $(X_3)$  and pollen concentration  $(X_4)$  on the bioactive properties of Uruset apple juice using RSM. Dependent and independent variables are shown in Table 2. The optimization process was carried out to increase total phenolic substance (TPC), total flavonoid substance (TFC) and antioxidant activities (DPPH and CUPRAC methods). The effects of temperature  $(X_1)$ , time  $(X_2)$ , amplitude  $(X_3)$  and pollen  $(X_4)$  independent variables on TPC (Equation 2), TFC (Equation 3), DPPH (Equation 4) and CUPRAC (Equation 5) properties of Uruset apple juice are shown in the following equations. ANOVA results are given in Table 3.

$$\operatorname{TPC}\left(\operatorname{mg}\frac{\operatorname{GAE}}{\operatorname{L}}\right) = 834.6 + 1.001 X_{1} + 13.651 X_{2} - 2.047 X_{3} \\ + 1.089 X_{4} - 0.04878 X_{1}X_{1} - 0.4393 X_{2}X_{2} \\ - 0.013533 X_{3}X_{3} - 0.016421 X_{4}X_{4} - 0.2030 X_{1}X_{2} \\ + 0.06290 X_{1}X_{3} + 0.01108 X_{1}X_{4} + 0.06119 X_{2}X_{3} \\ - 0.08087 X_{2}X_{4} + 0.01425 X_{3}X_{4}$$
(2)

$$\operatorname{TFC}\left(\operatorname{mg}\frac{\operatorname{CE}}{\operatorname{L}}\right) = 88.9 + 5.603 X_{1} + 10.846 X_{2} + 1.247 X_{3} \\ + 2.809 X_{4} - 0.08551 X_{1}X_{1} - 0.6471 X_{2}X_{2} \\ - 0.022992 X_{3}X_{3} - 0.025424 X_{4}X_{4} - 0.07495 X_{1}X_{2} \\ + 0.02641 X_{1}X_{3} + 0.00318 X_{1}X_{4} + 0.02760 X_{2}X_{3} \\ - 0.03547 X_{2}X_{4} + 0.005075 X_{3}X_{4}$$
(3)

TABLE 2 (Continued)

Run no

<u>ndep</u>endent vari<u>ables</u>

ime  $(X_2)$ 

emperature (X1) (°C)

31

X; temperature; X; stime; X; amplitude; X; poller; T'S-U]; Thermosonicated optimized pollen Uruset apple juice; TPC: total phenolic content; TFC: total flavonoid content; DPPH (2,2-diphenyl-1-picryhhydrazyl) antioxidant capacity; CUPRAC: copper reducing antioxidant capacity; GAE:

quercetin equivalents

gallic acid equivalent; CE:

05

		TPC (mg GAE/L)		TFC (mg CE/L)		DPPH (inhibition %)		CUPRAC (inhibition %)		
Source	DF	F-value	P-value	F-value	P-value	F-value	P-value	F-value	P-value	
Model	14.00	210.82	0.000	290.18	0.000	66.17	0.000	72.64	0.000	
Linear	4.00	175.77	0.000	37.00	0.000	25.15	0.000	7.39	0.001	
$X_1$	1.00	260.67	0.000	32.66	0.000	5.14	0.038	16.26	0.001	
X <sub>2</sub>	1.00	347.84	0.000	102.68	0.000	61.24	0.000	2.78	0.115	
X <sub>3</sub>	1.00	45.77	0.000	3.34	0.086	13.85	0.002	6.07	0.025	
$X_4$	1.00	48.78	0.000	9.30	0.008	20.36	0.000	4.44	0.051	
Square	4.00	212.66	0.000	882.76	0.000	87.62	0.000	205.95	0.000	
$X_1X_1$	1.00	181.59	0.000	922.09	0.000	55.91	0.000	229.37	0.000	
$X_2X_2$	1.00	376.91	0.000	1351.95	0.000	168.72	0.000	275.81	0.000	
X <sub>3</sub> X <sub>3</sub>	1.00	223.60	0.000	1066.68	0.000	59.59	0.000	274.39	0.000	
$X_4X_4$	1.00	329.19	0.000	1304.33	0.000	164.64	0.000	305.83	0.000	
2-way interaction	6.00	232.97	0.000	63.91	0.000	79.22	0.000	27.26	0.000	
$X_1X_2$	1.00	281.49	0.000	63.42	0.000	157.11	0.000	51.47	0.000	
$X_1X_3$	1.00	675.64	0.000	196.87	0.000	65.39	0.000	51.86	0.000	
$X_1X_4$	1.00	20.95	0.000	2.85	0.111	120.32	0.000	21.00	0.000	
X <sub>2</sub> X <sub>3</sub>	1.00	102.30	0.000	34.40	0.000	49.18	0.000	7.53	0.014	
$X_2X_4$	1.00	178.72	0.000	56.83	0.000	65.87	0.000	31.54	0.000	
$X_3X_4$	1.00	138.71	0.000	29.08	0.000	17.45	0.001	0.14	0.718	
Error	16.00									
Lack-of-fit	10.00	6.72	0.015	7.18	0.013	122.76	0.000	4.98	0.031	
Pure error	6									
Total	30									
R <sup>2</sup>		99.4	6%	99.61%		98.30%		98.45%		
Adjusted R <sup>2</sup>		98.9	9%	99.26%		96.82%		97.10%		
Predicted R <sup>2</sup>		97.0	9%	97.82	97.87%		90.26%		91.81%	

#### TABLE 3 ANOVA in the regression model of the combination test.

 $X_1$ : temperature;  $X_2$ : time;  $X_3$ : amplitude;  $X_4$ : Pollen; DF—degrees of freedom;  $R^2$ —coefficient of determination. TPC: total phenolic substance; TFC: total flavonoid, antioxidant capacity; DPPH: 2,2-diphenyl-1-picrylhydrazyl; CUPRAC: copper reducing antioxidant capacity. p < 0.05, statistically significant; p < 0.01, statistically highly significant.

$$\begin{aligned} \text{DPPH} \left(\text{inhibition}\%\right) &= 45.45 + 0.139 \, X_1 + 1.951 \, X_2 \\ &\quad + 0.0355 \, X_3 + 0.2022 \, X_4 - 0.01422 \, X_1 X_1 \\ &\quad - 0.1543 \, X_2 X_2 - 0.003669 \, X_3 X_3 - 0.006098 \, X_4 X_4 \\ &\quad - 0.07964 \, X_1 X_2 + 0.01028 \, X_1 X_3 + 0.01394 \, X_1 X_4 \\ &\quad + 0.02228 \, X_2 X_3 + 0.02578 \, X_2 X_4 - 0.002654 \, X_3 X_4 \end{aligned} \tag{4}$$

CUPRAC (inhibition%) = 
$$-36.63 + 1.788 X_1 + 6.369 X_2$$
  
+  $0.6602 X_3 + 0.9251 X_4 - 0.03146 X_1 X_1$   
-  $0.2156 X_2 X_2 - 0.008603 X_3 X_3 - 0.009083 X_4 X_4$   
-  $0.04981 X_1 X_2 + 0.01000 X_1 X_3 + 0.00636 X_1 X_4$   
-  $0.00953 X_2 X_3 - 0.01950 X_2 X_4 + 0.000256 X_3 X_4$  (5)

ANOVA results showed that all independent variables ( $X_1$ - $X_4$ ) had a significant effect on the measured responses (p < 0.05). Temperature and time were found to have the most significant effects.

## 3.1.1 Total phenolic compound (TPC)

All factors showed high significance (p < 0.0001) in second-order effects. The highest F-values were recorded for X<sub>2</sub> (F = 347.84) and X<sub>4</sub> (F = 329.19), indicating that the duration and pollen amount were important in preserving phenolic compounds (Figure 1). Especially the increase in duration had a positive effect on total phenolic compounds, and this result is consistent with previous studies on the enrichment of phenolic compounds (Gadioli Tarone et al., 2021; Kaur et al., 2022).

Total flavonoid substance (TFC), especially  $X_2$  (F = 1351.95) and  $X_4$  (F = 1304.33) variables, have the highest effect, revealing that time and pollen concentration play a critical role in the preservation of these compounds. An increase in the amount of TFC was observed in thermosonication treatment (Figure 2). Similar results were observed in samples of orange juice, whey (Oliveira et al., 2022), black, red, and white currant juices (Kidoń and Narasimhan, 2022).

Figures 3, 4 show response surface plots of DPPH and CUPRAC, respectively, RSM as a function of the significant interaction factors.





Significant improvements were observed with increasing temperature and amplitude in both methods, confirming the potential of ultrasound-assisted processing to enhance the extraction of bioactive compounds.

Increasing thermosonication parameters, as in our study, was reported to increase antioxidant activity in black carrot juice (Hasheminya and Dehghannya, 2022). Similar results were observed in other studies where thermosonication application to cashew apple juice and amora (*Spondius pinnata*) juice increased antioxidant activity values (Deli et al., 2022; Nayak et al., 2022). Pairwise interactions revealed complex relationships between variables and were statistically significant (p < 0.01). X<sub>1</sub> and X<sub>2</sub>





were found to significantly affect all responses, particularly a strong synergy with the CUPRAC value (F = 51.47). X<sub>1</sub> and X<sub>3</sub> were found to be significant, especially for TPC (F = 675.64) and TFC (F = 196.87), indicating the effect of ultrasound treatment in enhancing the extraction of phenolic compounds. X<sub>2</sub> × X<sub>4</sub> had high significance on DPPH (F = 65.87) and CUPRAC (F = 31.54), indicating that longer treatment time could increase antioxidant capacity by increasing pollen content.

The coefficient of determination ( $R^2$ ) values are above 98%, indicating excellent agreement between experimental and predicted data. The adjusted  $R^2$  and predicted  $R^2$  values are also exceptionally high, proving the reliability of the developed models. These findings show that the predictive power of RSM models is relatively high and can be used safely in optimization processes. The optimum process conditions were determined as 40.9°C temperature, 5.63 min process time, 61.01% amplitude, and 60 mg/100 mL pollen. The difference



between experimental and predicted values is less than 5%, which supports the reliability of the optimization process. This study provides a scientific basis for the industrial-scale development of Uruset apple juice and offers important findings for the production of functional beverages. In particular, the potential of ultrasoundassisted processing to increase the bioavailability of bioactive compounds is of great importance for developing functional products for consumer health.

## 3.2 Bioactive compounds

Fruits are the preferred source of bioactive compounds with health benefits for preventing and/or treating disease (De La Fuente-Carmelino et al., 2025). The main bioactive compounds are secondary metabolites with antioxidant, antimicrobial, anti-inflammatory, and other properties (Meena et al., 2024). Figure 5 shows significant differences (p < 0.05) in total phenolic compounds, flavonoid compounds, DPPH, and CUPRAC content. TPC and TFC were significantly affected by thermosonication, with the highest values observed in these samples. The results are parallel to previous studies that found that the thermosonication process increased TPC and TFC values (Yıkmış et al., 2022; Giovagnoli-Vicuña et al., 2022). The present study observed a significant difference in DPPH values between P-UJ and TS-UJ samples. Similarly, Mukhtar et al. (2024) found that the applied thermosonication process (25 kHz at 40 and 50°C) provided a significant increase in DPPH value compared to heat treatment (100°C, 4 min) of watermelon-beetroot juice. Additionally, Li et al. (2025) reported that thermosonication process (60°C, 15 min) provided significantly higher antioxidant activity compared to the pasteurization process (85°C for 10 min).

Pearson correlation plot shows the effect of thermosonication and pollen addition on physicochemical, sensory and bioactive properties in Uruset apple juice. A very strong positive correlation (r > 0.99) was observed between sugar components (fructose, glucose, sucrose and sorbitol), indicating that the components increased or decreased together. Also, a high level of positive correlation (r > 0.95) was found between total phenolic content (TPC), total flavonoid content (TFC) and antioxidant capacity (DPPH and CUPRAC), indicating that thermosonication and pollen addition increased the preservation and extraction of bioactive compounds (Figure 6).

## 3.3 Physicochemical parameters

In terms of physicochemical properties, pH levels remain relatively constant between treatments and are slightly higher than TS-UJ (3.57), C-UJ (3.54), and P-UJ (3.53) (Figure 7). Similar to these results, no significant change in pH value was observed after thermosonication treatment in the hog plum juice study conducted by Oladunjoye et al. in 2021 (Oladunjoye et al., 2021). The finding that pH value from physicochemical parameters did not show any significant difference compared to the control group is consistent with the result of this study. Total soluble solids (TSS, °Brix) content was the highest in TS-UJ (14.14), followed by C-UJ (13.80) and P-UJ (13.77), indicating that thermosonication may contribute to the release of soluble compounds and increase the sweetness and flavor of the juice. Titratable acidity (TA) remained relatively unchanged, with



Pearson correlates the coefficients between the physicochemical parameters, organic acid, sugar, phenolic compounds, polyphenol oxidase enzyme activity, sensory properties of the optimized thermosonicated Uruset apple juice samples.



values ranging from 5.72 to 5.78 g/L, indicating that the process has a minimal effect on acidity, which is very important for flavor stability. Pearson's positive correlation coefficients among the physicochemical parameters (pH, TSS, and TA) were significantly correlated with myricetin (1), cinnamic acid (1), and ferulic acid (0.99). In parallel with our study, it was stated that the thermosonication process of anthocyanin-enriched tomato juice did not cause a significant change

in pH value (Lafarga et al., 2019). However, the significant increase in TSS and TA values after thermosyncation may be due to the difference in the sample and the applied temperature–time combination. The findings that thermal pasteurization process did not affect the pH, TSS (% Brix), and TA (g/L) values of Uruset apple juice are similar to the results of studies conducted on physalis juice for the same purposes (Rabie et al., 2015).

## 3.4 Organic acid profile

The organic acid composition analysis of Uruset apple juice shows minor differences among control (C-UJ), pasteurized (P-UJ), and thermosonicated (TS-UJ) samples. Table 4 presents the results of organic acid analysis of C-UJ, P-UJ and TS-UJ Uruset apple juice. Total organic acid content is slightly higher in TS-UJ  $(1.33 \pm 0.01 \text{ g}/100 \text{ mL})$  compared to C-UJ  $(1.30 \pm 0.02 \text{ g}/100 \text{ mL})$ and P-UJ ( $1.25 \pm 0.01 \text{ g}/100 \text{ mL}$ ), but the differences are not statistically significant (p > 0.05). Among the individual organic acids, citric acid is the dominant component, followed by malic acid and tartaric acid, all showing minimal differences among the treatments. The citric acid content in TS-UJ ( $0.97 \pm 0.013 \text{ g}/100 \text{ mL}$ ) higher is significantly (p < 0.05)than in P-UI  $(0.91 \pm 0.01 \text{ g/100 mL})$ , indicating that thermosonication can better preserve this organic acid. At the same time, pasteurization appears to cause slight deterioration. However, malic acid and tartaric acid levels remain unchanged in all treatments (p > 0.05), indicating that these acids are more stable under different processing conditions. As demonstrated in the study by Yıkmış et al. (2022), the implementation of thermosonication treatment on freshly squeezed pomegranate juice did not lead to a substantial change in the amount of malic acid, with a (p < 0.05) level of significance, similar to our study.

Although the observed changes were small, the slightly higher organic acid content in TS-UJ suggests that thermosonication may help preserve acidity, which may be beneficial for flavor, microbial stability, and antioxidant properties. The lower total organic acid content in pasteurized samples suggests that thermal treatment may lead to slight degradation of some acid components, potentially affecting sensory and preservation qualities of the juice. These findings suggest that thermosonication may be a preferable alternative to pasteurization in terms of preserving the natural acid composition of Uruset apple juice and preserving its freshness and stability.

# 3.5 Sugar profile

Sugar composition analysis of Uruset apple juice revealed significant differences among control (C-UJ), pasteurized (P-UJ), and thermosonicated (TS-UJ) samples. Sugar content is given in Table 4. Total sugar content was highest in TS-UJ ( $10.01 \pm 0.00 \text{ g}/100 \text{ mL}$ ), followed by C-UJ ( $9.90 \pm 0.00 \text{ g}/100 \text{ mL}$ ) and lowest in P-UJ ( $9.32 \pm 0.04 \text{ g}/100 \text{ mL}$ ) with statistically significant differences (p < 0.05) among all treatments. These results indicate that thermosonication may contribute to maintaining or even increasing sugar concentrations, while pasteurization decreases slightly. Unlike our study, Liao et al. (2020) found that ultrasonication treatment did not significantly affect the total sugar

TABLE 4 Organic acid, sugar, and phenolic compounds analysis results of C-UJ, P-UJ and TS-UJ.

Studied compound		Samples					
		C-UJ (µg/mL)	P-UJ (μg/mL)	TS-UJ (μg/mL)			
Organic acid (g/100 mL)	Citric Acid	$0.95 \pm 0.013^{a}$	$0.91\pm0.01^{a}$	$0.97\pm0.013^{\rm b}$			
	Malic Acid	$0.29\pm0.00^{a}$	$0.28\pm0.00^{\rm a}$	$0.29 \pm 0.00^{a}$ $0.07 \pm 0.00^{a}$			
	Tartaric Acid	$0.07\pm0.00^{a}$	$0.06\pm0.00^{\mathrm{a}}$				
	Total	8.77 ± 0.65	$8.12\pm0.47$	8.69 ± 0.36			
Sugars (µg/mL)	Fructose	$5.69\pm0.07^{\rm b}$	$5.31\pm0.03^{\rm a}$	$5.74\pm0.08^{\rm b}$			
	Glucose	$2.20\pm0.03^{ab}$	$2.11\pm0.02^{\rm a}$	$2.24\pm0.03^{\rm b}$			
	Sucrose	$1.90\pm0.027^{\rm b}$	$1.79\pm0.02^{\rm a}$	$1.92\pm0.02^{\rm b}$			
	Sorbitol	$0.20\pm0.00^{\rm b}$	$0.19\pm0.00^{\rm a}$	$0.20\pm0.00^{\rm b}$			
	Total	$9.90\pm0.00^{\rm b}$	$9.32\pm0.04^{\rm a}$	$10.01 \pm 0.00^{\circ}$			
Phenolic compounds (μg/ mL)	Klorogenic acid	$61.43 \pm 1.56^{ab}$	$55.83 \pm 1.52^{a}$	$64.39 \pm 0.91^{\rm b}$			
	Gallic Acid	$4.10\pm0.03^{\text{b}}$	$3.73\pm0.03^{\rm a}$	$4.39 \pm 0.07^{\circ}$			
	Caffeic acid	$3.10\pm0.05^{\rm b}$	$2.81\pm0.06^{a}$	$3.22\pm0.05^{\rm b}$			
	p-Coumaric acid	$1.42\pm0.04^{ab}$	$1.29\pm0.04^{\rm a}$	$1.57\pm0.04^{\rm b}$			
	Quercetin	$13.26\pm0.19^{\rm b}$	$12.04 \pm 0.20^{a}$	$13.79 \pm 0.20^{\rm b}$			
	Phloridzin	9.23 ± 0.13°	$8.40 \pm 0.11^{a}$	$8.77\pm0.12^{ab}$			
	Catechin	$13.33 \pm 0.29^{ab}$	$12.03 \pm 0.41^{a}$	$13.86 \pm 0.30^{\rm b}$			
	Epicatechin	$15.39 \pm 0.33^{a}$	$14.07 \pm 0.20^{a}$	16.03 ± 1.15ª			
	Cinnamic acid	$0.00 \pm 0.00^{a}$	$0.00 \pm 0.00^{a}$	$0.01\pm0.00^{\rm b}$			
	Ferulic acid	$0.23\pm0.00^{a}$	$0.21\pm0.00^{a}$	$0.47\pm0.02^{\rm b}$			
	Rutin	$0.05\pm0.00^{\rm b}$	$0.00\pm0.00^{a}$	$0.08 \pm 0.00^{\circ}$			
	Myricetin	$0.00 \pm 0.00^{a}$	$0.00\pm0.00^{a}$	$0.19\pm0.00^{\rm b}$			
	Total	$121.28 \pm 2.38^{\mathrm{b}}$	$110.22 \pm 2.37^{a}$	$126.59 \pm 2.68^{\rm b}$			

The results are the mean  $\pm$  standard deviation (n = 3). The values marked with different letters within the line are significantly different from each other (p < 0.05). C-UJ: Control Uruset juice; P-UJ: Pasteurized Uruset apple juice; TS-UJ: Thermosonicated optimized pollen Uruset apple juice.

content of clear red pitaya juice (p > 0.05). Among individual sugars, fructose, glucose, and sucrose are the main components, and fructose is the dominant sugar in all treatments. TS-UJ exhibits the highest fructose content (5.74  $\pm$  0.08 g/100 mL), which is significantly higher than P-UJ  $(5.31 \pm 0.03 \text{ g}/100 \text{ mL})$  (p < 0.05), indicating that thermal and ultrasonic effects can increase sugar release or reduce degradation. Similarly, glucose and sucrose levels are slightly lower in pasteurized samples than control and thermosonic juices, indicating a small degradation effect during conventional heat treatment. Sorbitol content remains unchanged throughout the treatments, indicating that this sugar alcohol is more stable under different processing conditions. Statistical analysis showed that TS-UJ maintained significantly higher total sugar levels than P-UJ (p < 0.05). These findings highlight that thermosonication may be an effective method to preserve or improve the natural sugar profile of Uruset apple juice, making it a promising alternative to conventional pasteurization techniques. Ultrasound treatment provides mechanical effects through shear forces, allowing greater solvent penetration into the sample matrix (Mala et al., 2021).

## 3.6 Phenolic compounds profile

The phenolic compound result of Uruset apple juice is given in Table 4. The phenolic compound analysis of Uruset apple juice reveals significant differences among control (C-UJ), pasteurized (P-UJ) and thermosonicated (TS-UJ) samples. Total phenolic content was highest in TS-UJ (126.59  $\pm$  2.68  $\mu$ g/mL), followed by C-UJ (121.28  $\pm$  2.38  $\mu$ g/ mL) and lowest in P-UJ (110.22  $\pm$  2.37 µg/mL). There were statistically significant differences (p < 0.05) between pasteurized and thermosonicated samples. This result suggests that thermosonication may better preserve phenolic compounds than pasteurization, which has been shown to degrade certain bioactive compounds. Among the phenolic compounds, chlorogenic acid, quercetin, catechin, and epicatechin are found in relatively high concentrations and show significant differences depending on the processing method. For example, chlorogenic acid levels in TS-UJ ( $64.39 \pm 0.91 \,\mu\text{g/mL}$ ) are significantly higher (p < 0.05) than in P-UJ (55.83 ± 1.52 µg/mL), indicating that pasteurization leads to more significant loss of this compound. Similarly, quercetin and catechin levels are statistically higher in TS-UJ than P-UJ, supporting the hypothesis that thermosonication preserves flavonoids more effectively than conventional heat treatment. In a recent study, Erdal et al. (2022) discovered that p-coumaric acid and quercetin levels increased after ultrasound treatment. This outcome is consistent with the findings of this study. The level of caffeic acid was measured in the thermal pasteurised group (P-UJ) at 2.81 µg/mL, in the TS-UJ group at  $3.22 \,\mu\text{g/mL}$ , and in the C-UJ group at  $3.10 \,\mu\text{g/mL}$ . These differences are also statistically significant. Following the findings of the present study, the levels of caffeic and coumaric acids in black carrots were found to be comparatively low chlorogenic acid levels (Anandhi et al., 2024; Blando et al., 2021). Furthermore, pasteurization seems to have a reducing effect on most phenolic compounds, evidenced by significantly lower levels of gallate (p < 0.05), caffeic acid (p < 0.01), and phloridzin (p < 0.05) in P-UJ compared to TS-UJ and C-UJ. Interestingly, sinapic acid and myricetin were detected only in trace amounts in TS-UJ but not in the other samples, which may be due to structural changes induced by thermosonication. The presence of these compounds in TS-UJ but not in C-UJ suggests that ultrasound-induced cavitation and heat may facilitate the release or transformation of bound phenolic compounds. Statistical analysis highlights that TS-UJ showed significant differences (p < 0.05 or lower) compared to both C-UJ and P-UJ for most phenolic compounds. This suggests that thermosonication may preserve and enhance the extractability of certain phenolics. The moderate positive correlation (r  $\approx$  0.6–0.8) between phenolic compounds and sensory properties (taste, odor and overall acceptability) indicates that thermosonication positively affects product quality and consumer appreciation. Particularly, compounds such as chlorogenic acid, quercetin and caffeic acid showed high positive correlation, indicating that thermosonication and pollen addition supported the extraction of these components. These findings highlight the strong influence of processing conditions on phenolic compound stability and suggest that thermosonication is a promising technology to preserve and potentially enrich the bioactive profile of Uruset apple juice.

## 3.7 Polyphenol oxidase enzyme activity

Polyphenol oxidase (PPO) activity in Uruset apple juice showed significant differences among control (C-UJ), pasteurized (P-UJ) and thermosonicated (TS-UJ) samples. PPO results are given in Figure 8. The highest PPO activity was observed in C-UJ (70.81%), while P-UJ showed the lowest activity (42.49%), indicating that pasteurization effectively inactivated PPO enzymes. TS-UJ showed a moderate value (54.52%), indicating that thermosensation led to a partial decrease in enzyme activity but was not as effective as pasteurization. The moderate inactivation in TS-UJ can be attributed to the combined thermal and cavitation effects of thermosonication, which disrupted enzyme structures while preserving more bioactive compounds than conventional heat treatments. The strong negative correlation  $(r \approx -0.8)$  between polyphenol oxidase (PPO) activity and phenolic compounds and antioxidant capacity indicates that inactivation of PPO contributes to preserving phenolic compounds and increasing antioxidant capacity. Similar to our study, Aadil et al. reported that thermosonication application caused a decrease in the PPO activity of grapefruit juice (Aadil et al., 2015). Considering that PPO is responsible for enzymatic browning, the decrease in its activity in P-UJ





and TS-UJ implies that both treatments can improve juice's visual appeal and shelf-life stability. The synergistic effect of temperature and ultrasound was more effective in denaturing PPO (Zhang et al., 2017).

## 3.8 Sensory properties

The sensory analysis results of Uruset apple juice are given in Figure 9. The sensory analysis results of Uruset apple juice revealed significant differences among control (C-UJ), pasteurized (P-UJ), and thermosonicated (TS-UJ) samples, highlighting the effect of processing methods on consumer perception. Color, odor, taste, turbidity, and overall acceptability scores were significantly lower in pasteurized samples (P-UJ), while thermosonicated samples (TS-UJ) generally maintained higher sensory attributes than the control group. Shen et al. (2021) observed that the overall liking of ultrasound-treated apple juice increased, which is similar to our study. In particular, the color and odor intensity of TS-UJ was rated significantly higher than P-UJ (p < 0.05), indicating that thermosonication better preserves the fresh and natural characteristics of the juice. In addition, the taste and overall acceptability of TS-UJ were statistically superior to P-UJ (p < 0.05), indicating that thermosonication may increase flavor perception and consumer preference compared to conventional heat treatment. The lower acceptability of P-UJ may be attributed to the thermal degradation of volatile compounds and bioactive components, leading to reduced sensory appeal. Moreover, turbidity, a critical quality parameter affecting consumer perception, was significantly higher in TS-UJ than P-UJ (p < 0.05), indicating that thermosonication preserves the natural turbidity of apple juice, which is generally associated with higher bioactive content. Tukey's HSD test revealed significant differences between the treatments, indicating that thermosonication maintains or even improves the sensory profile of Uruset apple juice compared to pasteurisation. These findings align with previous studies showing that nonthermal technologies can improve sensory properties while minimizing nutrient loss, making them a promising alternative for high-quality juice processing. The positive correlation observed between sensory properties and antioxidant capacity indicates that thermosonication process improves both nutritional and sensory quality. Taste showed a significant negative correlation with phloridzin (-0.82) and polyphenol oxidase (-0.83). These results show that thermosonication is an effective method for developing a functional beverage by optimizing the bioactive and sensory properties of Uruset apple juice. Given that sensory properties play an important role in consumer acceptance, the results support the potential application of thermosonication in the juice industry to improve product quality while maintaining the desired sensory attributes.

# 3.9 Principal component analysis (PCA)

PCA (Principal Component Analysis) plot visually summarizes the changes in sensory and chemical properties of Uruset apple juice samples due to different processing methods (control, pasteurization, and thermosonication) and the relationships between these properties. Two main components explain almost all of the total variance (100%) (first component 73.6%; second component 26.4%). The first principal component (PC1) shows a strong relationship with phenolic compounds (especially chlorogenic acid, gallic acid, catechin, quercetin), sugars (fructose, glucose, sorbitol), organic acids (malic acid, tartaric acid), total soluble solids (TSS), turbidity, polyphenol oxidase (PPO) and pH (Figure 10). This shows that PC1 mainly reflects chemical quality indicators and thermosonication treated samples (TS-UJ) are richer in these parameters. The control group (C-UJ) shows a higher enzymatic



activity, especially with a negative separation of polyphenol oxidase activity. The second component (PC2) in the graph explains 26.4% of the total variance and clearly represents sensory parameters (taste and general acceptability). Sensory parameters show a relatively independent distribution from other chemical properties. It is seen that thermosonicated samples (TS-UJ) are more sensory acceptable and preferable by the consumer. Thus, PCA results confirm that thermosonication is effective both in preserving and increasing bioactive components and in developing a product with high general acceptability. This situation reveals that thermosonication offers significant advantages compared to traditional methods for Uruset apple juice.

# 4 Conclusion

This study was conducted to optimize the bioactive and sensory properties of thermosonicated fortified-pollen Uruset apple juice. The thermosonication process and pollen addition increased the apple juice's total phenolic and flavonoid content and improved its antioxidant capacity. Especially phenolic compounds were preserved at higher levels and increased in quantity after thermosonication. The addition of pollen increased the stability of phenolic compounds and had a positive effect on antioxidant activity. The sensory analysis showed that the thermosonication process and pollen addition improved the apple juice's taste, odour, and general acceptability. The decrease in polyphenol oxidase activity contributed to preserving phenolic compounds. The synergistic effect of the thermosonication process and pollen addition was demonstrated by the positive correlation between sugar components and phenolic compounds. These results show that the thermosonication process and pollen content effectively produce functional beverages by improving the bioactive and sensory properties of Uruset apple juice. The study demonstrated the potential of valuable bioactive ingredients such as pollen in the beverage industry and supported the applicability of thermosonication in the production of functional beverages. These findings provide a scientific basis for the industrial processing of Uruset apple juice and the development of functional products.

# Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

# Author contributions

SY: Supervision, Investigation, Methodology, Conceptualization, Data curation, Validation, Writing – review & editing,

Writing – original draft, Project administration. MT: Writing – review & editing, Formal analysis, Validation, Writing – original draft, Conceptualization. NTD: Software, Writing – review & editing, Validation, Writing – original draft, Visualization. BA: Writing – original draft, Writing – review & editing, Resources, Formal analysis. IMA: Writing – review & editing, Writing – original draft. MOA: Writing – original draft, Funding acquisition, Writing – review & editing.

## Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was supported by Princess Nourah Bint Abdulrahman University Researchers Supporting Project Number (PNURSP2025R251), Princess Nourah Bint Abdulrahman University, Riyadh, Saudi Arabia.

# Acknowledgments

The authors gratefully acknowledge the financial support provided by TÜBİTAK (Scientific and Technological Research Council of

## References

Aadil, R. M., Zeng, X. A., Zhang, Z. H., Wang, M. S., Han, Z., Jing, H., et al. (2015). Thermosonication: a potential technique that influences the quality of grapefruit juice. *Int. J. Food Sci. Technol.* 50, 1275–1282. doi: 10.1111/IJFS.12766

Abacı, Z., and Sevindik, E. (2014). Determination of Bioactive Compounds and Total Antioxidant Capacity in Apple Varieties Grown in Ardahan Region. Süleyman Demirel University Faculty of Agriculture Journal and Yüzüncü Yıl University Journal of Agricultural Sciences.

Anandhi, E., Shams, R., Dash, K. K., Bhasin, J. K., Pandey, V. K., and Tripathi, A. (2024). Extraction and food enrichment applications of black carrot phytocompounds: a review. *Appl. Food Res.* 4:100420. doi: 10.1016/J.AFRES.2024.100420

Anaya-Esparza, L. M., Velázquez-Estrada, R. M., Roig, A. X., García-Galindo, H. S., Sayago-Ayerdi, S. G., and Montalvo-González, E. (2017). Thermosonication: An Alternative Processing for Fruit and Vegetable Juices. *Trends Food Sci. Technol.* 61, 26–37. doi: 10.1016/J.TIFS.2016.11.020

Apak, R., Güçlü, K., Özyürek, M., Esin Karademir, S., and Erçğ, E. (2006). The Cupric Ion Reducing Antioxidant Capacity and Polyphenolic Content of Some Herbal Teas. *Int. J. Food Sci. Nutr.* 57, 292–304. doi: 10.1080/09637480600798132

Bertoncelj, J., Lilek, N., and Korošec, M. (2023). Bee Pollen Carbohydrates Composition and Functionality. *Pollen Chem. Biotechnol.*, 51–69. doi: 10.1007/978-3-031-47563-4\_3

Blando, F., Marchello, S., Maiorano, G., Durante, M., Signore, A., Laus, M. N., et al. (2021). Bioactive Compounds and Antioxidant Capacity in Anthocyanin-Rich Carrots: A Comparison between the Black Carrot and the Apulian Landrace "Polignano". *Carrot. Plants* 10:564. doi: 10.3390/PLANTS10030564

Castellari, M., Versari, A., Spinabelli, U., Galassi, S., and Amati, A. (2000). An improved HPLC method for the analysis of organic acids, carbohydrates, and alcohols in grape musts and wines. *J. Liquid Chromatog. Relat. Technol.* 23, 2047–2056. doi: 10.1081/JLC-100100472

Cemeroğlu, B. (2010). Gıda Analizleri. 2nd Edn. Nobel Yayıncılık: Ankara.

Chandrapala, J., Oliver, C., Kentish, S., and Ashokkumar, M. (2012). Ultrasonics in Food Processing – Food Quality Assurance and Food Safety. *Trends Food Sci. Technol.* 26, 88–98. doi: 10.1016/J.TIFS.2012.01.010

Chemat, F., Zill-e-Huma, and Khan, M. (2011). Applications of ultrasound in food technology: processing, preservation and extraction. *Ultrason. Sonochem.* 18, 813–835. doi: 10.1016/J.ULTSONCH.2010.11.023

Coşkun, S., and Aşkın, M. (2016). Determination of Pomological and Biochemical Characteristics of Some Local Apple Varieties. Süleyman Demirel University Faculty of Agriculture Journal and Yüzüncü Yil University Journal of Agricultural Sciences. Turkey) under the 2209-A Student Research Project Program (project number 1919B012318028).

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## **Generative AI statement**

The author(s) declare that no Gen AI was used in the creation of this manuscript.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

De La Fuente-Carmelino, L., Anticona, M., Ramos-Escudero, F., Casimiro-Gonzales, S., and Muñoz, A. M. (2025). Commercial Plant-Based Functional Beverages: A Comparative Study of Nutritional Composition and Bioactive Compounds. *Beverages* 11:26. doi: 10.3390/BEVERAGES11010026

Del Socorro Cruz-Cansino, N., Ramírez-Moreno, E., León-Rivera, J. E., Delgado-Olivares, L., Alanís-García, E., Ariza-Ortega, J. A., et al. (2015). Shelf Life, Physicochemical, Microbiological and Antioxidant Properties of Purple Cactus Pear (Opuntia Ficus Indica) Juice after Thermoultrasound Treatment. *Ultrason. Sonochem.* 27, 277–286. doi: 10.1016/J.ULTSONCH.2015.05.040

Deli, M. G. E. P., Kirit, B. D., Ağçam, E., and Akyıldız, A. (2022). The Effects of Thermosonication on Quality Parameters of Cashew Apple Nectar: An Optimization Study for Processing Conditions. *Appl. Food Res.* 2:100217. doi: 10.1016/J.AFRES.2022.100217

Erdal, B., Yıkmış, S., Demirok, N. T., Bozgeyik, E., and Levent, O. (2022). Effects of Non-Thermal Treatment on Gilaburu Vinegar (*Viburnum Opulus* L.): Polyphenols, Amino Acid, Antimicrobial, and Anticancer Properties. *Biology* 11:926. doi: 10.3390/BIOLOGY11060926

Franz, C. M. A. P., Specht, I., Cho, G. S., Graef, V., and Stahl, M. R. (2009). UV-C-Inactivation of Microorganisms in Naturally Cloudy Apple Juice Using Novel Inactivation Equipment Based on Dean Vortex Technology. *Food Control* 20, 1103–1107. doi: 10.1016/J.FOODCONT.2009.02.010

Gadioli Tarone, A., Keven Silva, E., de Freitas, D., Queiroz Barros, H., Baú Betim Cazarin, C., and Roberto Marostica Junior, M. (2021). High-Intensity Ultrasound-Assisted Recovery of Anthocyanins from Jabuticaba by-Products Using Green Solvents: Effects of Ultrasound Intensity and Solvent Composition on the Extraction of Phenolic Compounds. *Food Res. Int.* 140:110048. doi: 10.1016/J.FOODRES.2020.110048

Giovagnoli-Vicuña, C., Briones-Labarca, V., Romero, M. S., Giordano, A., and Pizarro, S. (2022). Effect of Extraction Methods and In Vitro Bio-Accessibility of Microencapsulated Lemon Extract. *Molecules* 27:4166. doi: 10.3390/MOLECULES27134166

Hasheminya, S. M., and Dehghannya, J. (2022). Non-Thermal Processing of Black Carrot Juice Using Ultrasound: Intensification of Bioactive Compounds and Microbiological Quality. Int. J. Food Sci. Technol. 57, 5848–5858. doi: 10.1111/IJFS.15901

İzol, E., Turhan, M., Yılmaz, M. A., Çağlayan, C., and Gülçin, İ. (2025). Determination of Antioxidant, Antidiabetic, Anticholinergic, Antiglaucoma Properties and Comprehensive Phytochemical Content by LC-MS/MS of Bingöl Honeybee Pollen. *Food Sci. Nutr.* 13:e4531. doi: 10.1002/FSN3.4531

Kaur, B., Panesar, P. S., and Anal, A. K. (2022). Standardization of Ultrasound Assisted Extraction for the Recovery of Phenolic Compounds from Mango Peels. *J. Food Sci. Technol.* 59, 2813–2820. doi: 10.1007/S13197-021-05304-0 Kidoń, M., and Narasimhan, G. (2022). Effect of Ultrasound and Enzymatic Mash Treatment on Bioactive Compounds and Antioxidant Capacity of Black, Red and White Currant Juices. *Molecules* 27:318. doi: 10.3390/MOLECULES27010318

Lafarga, T., Ruiz-Aguirre, I., Abadias, M., Viñas, I., Bobo, G., and Aguiló-Aguayo, I. (2019). Effect of Thermosonication on the Bioaccessibility of Antioxidant Compounds and the Microbiological, Physicochemical, and Nutritional Quality of an Anthocyanin-Enriched Tomato Juice. *Food Bioprocess Technol.* 12, 147–157. doi: 10.1007/S11947-018-2191-5

Leighton, T. G. (2007). What Is Ultrasound? Prog. Biophys. Mol. Biol. 93, 3-83. doi: 10.1016/J.PBIOMOLBIO.2006.07.026

Li, L., Su, H., Pang, L., Pan, Y., Li, X., Xu, Q., et al. (2025). Thermosonication enhanced the bioactive, antioxidant, and flavor attributes of freshly squeezed tomato juice. *Ultrason. Sonochem.* 115:107299. doi: 10.1016/J.ULTSONCH.2025.107299

Liang, Z., Cheng, Z., and Mittal, G. S. (2006). Inactivation of Spoilage Microorganisms in Apple Cider Using a Continuous Flow Pulsed Electric Field System. *LWT Food Sci. Technol.* 39, 351–357. doi: 10.1016/J.LWT.2005.02.019

Liao, H., Zhu, W., Zhong, K., and Liu, Y. (2020). Evaluation of colour stability of clear red pitaya juice treated by Thermosonication. *LWT* 121:108997. doi: 10.1016/J.LWT.2019.108997

Lima, M. D. S., da, M., Toaldo, I. M., Corrêa, L. C., Pereira, G. E., de, D., et al. (2015). Phenolic compounds, organic acids and antioxidant activity of grape juices produced in industrial scale by different processes of maceration. *Food Chem.* 188, 384–392. doi: 10.1016/J.FOODCHEM.2015.04.014

Lu, G., Li, C., Liu, P., Cui, H., Xia, Y., and Wang, J. (2010). Inactivation of Microorganisms in Apple Juice Using an Ultraviolet Silica-Fiber Optical Device. *J. Photochem. Photobiol. B* 100, 167–172. doi: 10.1016/J.JPHOTOBIOL.2010.06.003

Mala, T., Sadiq, M. B., and Anal, A. K. (2021). Optimization of Thermosonication Processing of Pineapple Juice to Improve the Quality Attributes during Storage. *J. Food Meas. Charact.* 15, 4325–4335. doi: 10.1007/S11694-021-01011-8

Martínez-Flores, H. E., Garnica-Romo, M. G., Bermúdez-Aguirre, D., Pokhrel, P. R., and Barbosa-Cánovas, G. V. (2015). Physico-Chemical Parameters, Bioactive Compounds and Microbial Quality of Thermo-Sonicated Carrot Juice during Storage. *Food Chem.* 172, 650–656. doi: 10.1016/J.FOODCHEM.2014.09.072

McClements, D. J. (1995). Advances in the Application of Ultrasound in Food Analysis and Processing, Trends Food Sci. Technol. 6, 293-299. doi: 10.1016/S0924-2244(00)89139-6

Meena, L., Gowda, N. N., Sunil, C. K., Rawson, A., and Janghu, S. (2024). Effect of Ultrasonication on Food Bioactive Compounds and Their Bio-Accessibility: A Review. *J. Food Compos. Anal.* 126:105899. doi: 10.1016/J.JFCA.2023.105899

Mukhtar, K., Nabi, B. G., Manzoor, M. F., Zia, S., Bhat, Z. F., Hussain, S., et al. (2024). Impact of thermal, ultrasonication, and thermosonication processes on the quality profile of watermelon-beetroot juice blend: a comparative study. *J. Food Process. Preserv.* 2024, 1–8. doi: 10.1155/2024/5518914

Nayak, P. K., Chandrasekar, C. M., Gogoi, S., and Kesavan, R. (2022). krishnan Impact of Thermal and Thermosonication Treatments of Amora (Spondius Pinnata) Juice and Prediction of Quality Changes Using Artificial Neural Networks. *Biosyst. Eng.* 223, 169–181. doi: 10.1016/J.BIOSYSTEMSENG.2022.02.012

Oladunjoye, A. O., Adeboyejo, F. O., Okekunbi, T. A., and Aderibigbe, O. R. (2021). Effect of Thermosonication on Quality Attributes of Hog Plum (Spondias Mombin L.) Juice. Ultrason. Sonochem. 70:105316. doi: 10.1016/J.ULTSONCH.2020.105316

Oliveira, G. A. R., Guimarães, J. T., Ramos, G. L. P. A., Esmerino, E. A., Pimentel, T. C., Neto, R. P. C., et al. (2022). Benefits of Thermosonication in Orange Juice Whey Drink Processing. *Innovative Food Sci. Emerg. Technol.* 75:102876. doi: 10.1016/J.IFSET.2021.102876 Portu, J., López, R., Santamaría, P., and Garde-Cerdán, T. (2017). Elicitation with Methyl Jasmonate Supported by Precursor Feeding with Phenylalanine: Effect on Garnacha Grape Phenolic Content. *Food Chem.* 237, 416–422. doi: 10.1016/J.FOODCHEM.2017.05.126

Rabie, M. A., Soliman, A. Z., Diaconeasa, Z. S., and Constantin, B. (2015). Effect of Pasteurization and Shelf Life on the Physicochemical Properties of Physalis (*Physalis Peruviana* L.). *Juice. J Food Proc. Pres.* 39, 1051–1060. doi: 10.1111/JFPP.12320

Rawson, A., Tiwari, B. K., Patras, A., Brunton, N., Brennan, C., Cullen, P. J., et al. (2011). Effect of Thermosonication on Bioactive Compounds in Watermelon Juice. *Food Res. Int.* 44, 1168–1173. doi: 10.1016/J.FOODRES.2010.07.005

Sadler, G. D., and Murphy, P. A. (2010). PH and Titratable Acidity., 219-238. doi: 10.1007/978-1-4419-1478-1\_13

Şengül, M., Erkaya, T., Başlar, M., and Ertugay, M. F. (2011). Effect of Photosonication Treatment on Inactivation of Total and Coliform Bacteria in Milk. *Food Control* 22, 1803–1806. doi: 10.1016/J.FOODCONT.2011.04.015

Shen, Y., Zhu, D., Xi, P., Cai, T., Cao, X., Liu, H., et al. (2021). Effects of temperaturecontrolled ultrasound treatment on sensory properties, physical characteristics and antioxidant activity of cloudy apple juice. *LWT* 142:111030. doi: 10.1016/J.LWT.2021.111030

Singh, R. P., Chidambara Murthy, K. N., and Jayaprakasha, G. K. (2002). Studies on the Antioxidant Activity of Pomegranate (Punica Granatum) Peel and Seed Extracts Using in Vitro Models. *J. Agric. Food Chem.* 50, 81–86. doi: 10.1021/JF010865B

Singleton, V. L., Orthofer, R., and Lamuela-Raventós, R. M. (1999). Analysis of Total Phenols and Other Oxidation Substrates and Antioxidants by Means of Folin-Ciocalteu Reagent. *Methods Enzymol.* 299, 152–178. doi: 10.1016/S0076-6879(99)99017-1

Sturm, K., Koron, D., and Stampar, F. (2003). The Composition of Fruit of Different Strawberry Varieties Depending on Maturity Stage. *Food Chem.* 83, 417–422. doi: 10.1016/S0308-8146(03)00124-9

Tiwari, B. K., and Mason, T. J. (2012). Ultrasound Processing of Fluid Foods. Novel Therm. Non-Thermal Technol. Fluid Foods, 135–165. doi: 10.1016/B978-0-12-381470-8.00006-2

Yıkmış, S. (2019). Optimization of Uruset Apple Vinegar Production Using Response Surface Methodology for the Enhanced Extraction of Bioactive Substances. *Food Secur.* 8:107. doi: 10.3390/FOODS8030107

Yıkmış, S., Ozer, H., Levent, O., Çöl, B. G., and Erdal, B. (2022). Effect of Thermosonication and Thermal Treatments on Antidiabetic, Antihypertensive, Mineral Elements and in Vitro Bioaccessibility of Bioactive Compounds in Freshly Squeezed Pomegranate Juice. J. Food Meas. Charact. 16, 3023–3041. doi: 10.1007/S11694-022-01402-5

Yildiz, R., and Maskan, M. (2022). Optimization of a Green Tea Beverage Enriched with Honey and Bee Pollen. *Int. J. Gastron. Food Sci.* 30:100597. doi: 10.1016/J.IJGFS.2022.100597

Zhang, Y., Liu, X. C., Wang, Y., Zhao, F., Sun, Z., and Liao, X. (2016). Quality Comparison of Carrot Juices Processed by High-Pressure Processing and High-Temperature Short-Time Processing. *Innovative Food Sci. Emerg. Technol.* 33, 135–144. doi: 10.1016/J.IFSET.2015.10.012

Zhang, Z., Niu, L., Li, D., Liu, C., Ma, R., Song, J., et al. (2017). Low intensity ultrasound as a pretreatment to drying of daylilies: impact on enzyme inactivation, color changes and nutrition quality parameters. *Ultrason. Sonochem.* 36, 50–58. doi: 10.1016/J.ULTSONCH.2016.11.007

Zhishen, J., Mengcheng, T., and Jianming, W. (1999). The Determination of Flavonoid Contents in Mulberry and Their Scavenging Effects on Superoxide Radicals. *Food Chem.* 64, 555–559. doi: 10.1016/S0308-8146(98)00102-2