



Ups and Downs in 75 Years of Electrocochleography

Jos J. Eggermont^{1,2*}

¹Department of Psychology, University of Calgary, Calgary, AB, Canada, ²Department of Physiology and Pharmacology, University of Calgary, Calgary, AB, Canada

Before 1964, electrocochleography (ECoChG) was a surgical procedure carried out in the operating theatre. Currently, the newest application is also an intra-operative one, often carried out in conjunction with cochlear implant surgery. Starting in 1967, the recording methods became either minimal- or not-invasive, i.e., trans-tympanic (TT) or extra tympanic (ET), and included extensive studies of the arguments pro and con. I will review several valuable applications of ECoChG, from a historical point of view, but covering all 75 years if applicable. The main topics will be: (1) comparing human and animal cochlear electrophysiology; (2) the use in objective audiometry involving tone pip stimulation—currently mostly pre cochlear implantation but otherwise replaced by auditory brainstem response (ABR) recordings; (3) attempts to diagnose Ménière's disease and the role of the summing potential (SP); (4) early use in diagnosing vestibular schwannomas—now taken over by ABR screening and MRI confirmation; (5) relating human electrophysiology to the effects of genes as in auditory neuropathy; and (6) intracochlear recording using the cochlear implant electrodes. The last two applications are the most recently added ones. The “historical aspects” of this review article will highlight the founding years prior to 1980 when relevant. A survey of articles on Pubmed shows several ups and downs in the clinical interest as reflected in the publication counts over the last 75 years.

OPEN ACCESS

Edited by:

Jeffery Lichtenhan,
Washington University in St. Louis,
USA

Reviewed by:

Spencer Smith,
University of Arizona, USA
William Peter Gibson,
University of Sydney, Australia

*Correspondence:

Jos J. Eggermont
eggermont@ucalgary.ca

Received: 14 November 2016

Accepted: 11 January 2017

Published: 24 January 2017

Citation:

Eggermont JJ (2017) Ups and
Downs in 75 Years of
Electrocochleography.
Front. Syst. Neurosci. 11:2.
doi: 10.3389/fnsys.2017.00002

Keywords: auditory nerve, summing potential, compound action potential, cochlear microphonic, Ménière's disease, vestibular schwannoma, auditory neuropathy, cochlear implants

INTRODUCTION

Electrocochleography (ECoChG) is a technique for recording sound-evoked cochlear and auditory nerve population responses from the round window, the cochlear wall (promontory), eardrum and external ear canal. One observes (**Figure 1**) that there are several ups and downs in the number of publications across the years, potentially reflecting the waxing and waning interest for ECoChG as a diagnostic tool. Overall, there is a trend for a slow increase in the output.

Early Surgical Recordings

The first indications of the feasibility of recording cochlear potentials came from Fromm et al. (1935); the responses obtained in two humans with perforated eardrums were small and no cathode ray display could be obtained. Improved recording and amplification techniques gave better cochlear microphonics (CM) recordings (Perlman and Case, 1941; Lempert et al., 1947, 1950). Perlman and Case (1941) placed an electrode on the cochlea, first in monkeys and later in human ears. They found that CM could be obtained regularly in humans with a nearly normal

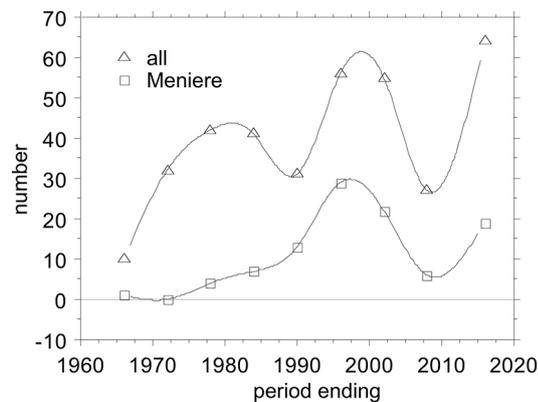


FIGURE 1 | Ups and Downs in the number of publications on Electrocochleography (ECoChG), overall and applied to Ménière's disease. I tabulated all ECoChG articles between 1941 and present that I could find in PubMed and added those from a 1976 conference proceeding (Ruben et al., 1976), amounting to 358 publications covering 75 years. I grouped them in periods of 6 years, because that bracketed the early minimum-intervention recording period from 1967–1972. The first 25 years before that I grouped together. The last period ending July 2016 covers 6.5 years.

audiogram. The potentials could be clearly detected in a loudspeaker or headphones. I consider this the start of ECoChG, albeit that only later Lempert et al. (1947) coined the term “cochleogram”. They carried out recordings in 11 human ears in the course of surgical interventions for otosclerosis, tinnitus or Ménières' disease. They could record responses from the round window in six ears but not from the promontory (no waveforms shown). In a follow-up study, Lempert et al. (1950) could record responses—again no waveforms were shown—in 13 out of 32 ears. They also suggested the placement of the electrode through the eardrum onto the promontory as a feasible non-surgical technique. Then the Ruben era ensued when Ruben et al. (1959, 1960) recorded CMs from the round window with clear waveforms for moderate level sounds produced by tuning forks and human whistles. The feasibility of ECoChG as a diagnostic method was advanced when Ruben et al. (1961) recorded the first compound action potential (CAP) with clear N_1 and N_2 components from the round window. They quantified especially the N_1 latency to a click and found that it was longer at threshold in humans compared to cats. Ruben et al. (1962) extended their recordings to children with serious verbal communication difficulty, seriously impaired speech, and who gave no subjective evidence of hearing. Ruben and Walker (1963) recorded CAPs in Ménières' disease and found them similar to those in other humans when recorded from the round window. Finally, Bordley et al. (1964) reviewed the results obtained by the Ruben group in 63 patients, among those ECoChGs obtained before and after stapes surgery. Clear nearly noise free N_1N_2 waveforms were shown. Ronis (1966) also presented some results pre- and post-stapedectomy, and suggested the use of the N_1 latency as a “valid index of improved sound conduction”. Reviewing his work, Ruben (1967) mentioned three important

topics in ECoChG: (1) the correlation of physiological and psychoacoustic properties; (2) the investigation of certain diseases; and (3) the objective diagnosis of individual cases of deafness. The Ruben era of using ECoChG was characterized by improved CM measurements and clear CAP recordings at moderate-to-high stimulus levels. It was still impossible to measure the CAP near the subjective threshold, and ECoChG as based on round-window recording was still an operating room technique.

Non-Surgical Period

The non-surgical period, started in 1967—time points refer to **Figure 1**—with the first publications by two groups one in Tokyo, Japan, led by Nobuo Yoshie and Toru Ohashi (Yoshie et al., 1967; Yoshie, 1968; Yoshie and Ohashi, 1969), and the other in Bordeaux, France, led by Michel Portmann and Jean-Marie Aran (Portmann et al., 1967; Aran and Le Bert, 1968; Aran, 1971; Portmann and Aran, 1971). The Japanese group started with extra-tympanic (ET) recordings but later on also used trans-tympanic (TT) recording as well. The Bordeaux group only used TT recordings. The period ending 1978 reflects in part the fairly large output from the Leyden group in Netherlands starting with Eggermont et al. (1974) and only using TT recording. The period ending 1984 signaled a starting interest in applying ECoChG to the diagnosis of Ménière's disease. After a reduction in the output in the period ending 1990, things picked up again in the following 12 years with a surge of papers on improving the use of ECoChG in the diagnosis of Ménière's disease. This was followed by a slump, both in the total number of ECoChG articles and in the Ménière articles, potentially by disappointment in the clinical usefulness of the ECoChG (Nguyen et al., 2010). The last six and a half years again show a steep incline in interest for ECoChG fueled by its use in auditory neuropathy, its revival in Ménière's disease following better diagnostic use of all information in the recorded waveforms, as intra-operative tests for cochlear implantation, and using the cochlear implant electrodes to perform multi-channel ECoChG.

BASIC PRINCIPLES

Hair Cell Potentials

Both inner hair cells (IHC) and outer hair cells (OHC) generate receptor potentials in response to sound (Russell and Sellick, 1978; Dallos et al., 1982). It has long been known that compound responses from the cochlea reflecting these hair cell potentials can be recorded at remote sites such as the round window, tympanic membrane or even from the scalp, and can be used clinically. These responses are called the CM and the summing potential (SP). The CM is produced almost exclusively from OHC receptor currents and when recorded from the round window (RW) membrane is dominated by the responses of OHCs in the basal turn. The SP is a direct-current component resulting from the non-symmetric depolarization-hyperpolarization response of the cochlea, which can be of positive or negative polarity, and is likely also generated dominantly by the OHCs (Russell, 2008).

The Summating Potential

Dallos (1972) recorded, using electrodes in the scala vestibule and scala tympani, a differential component (DIF SP) and an overall component (AVE SP) of the cochlea. The DIF SP represents the DC-shift between scala vestibuli and scala tympani, and the AVE SP represents the DC shift of the entire cochlea relative to the neck muscle potential. The AVE SP is positive at the site of maximum stimulation and negative elsewhere in the cochlea. For a round-window recording from the guinea pig, one nearly always measures a positive SP^+ when stimulation occurs with high-frequency, high-level tone bursts. This, therefore, may be compared to the AVE SP recorded from the first turn.

In human ECoChG recordings from the promontory, the SP is most often negative in polarity (SP^-). Sometimes, a change of sign is observed when the frequency of the tone burst is increased while keeping the intensity the same. A sequence of this kind is shown in **Figure 2** taken from Eggermont (1976c). A comparable polarity transition occurring between 4 kHz and 8 kHz was shown in Dauman et al. (1988). For this ear of a patient with Ménière's disease, a distinct SP^- is observed for a stimulus

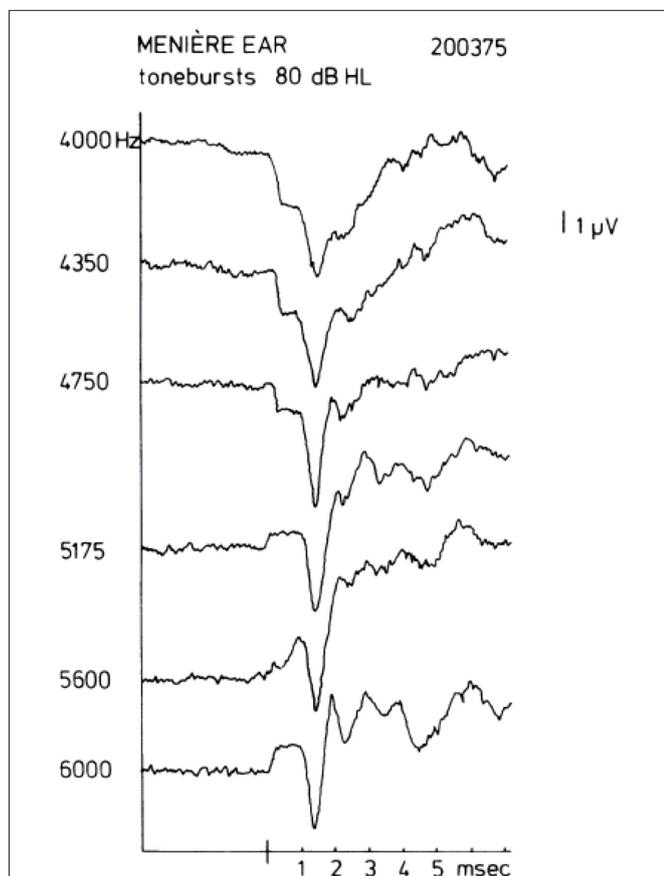


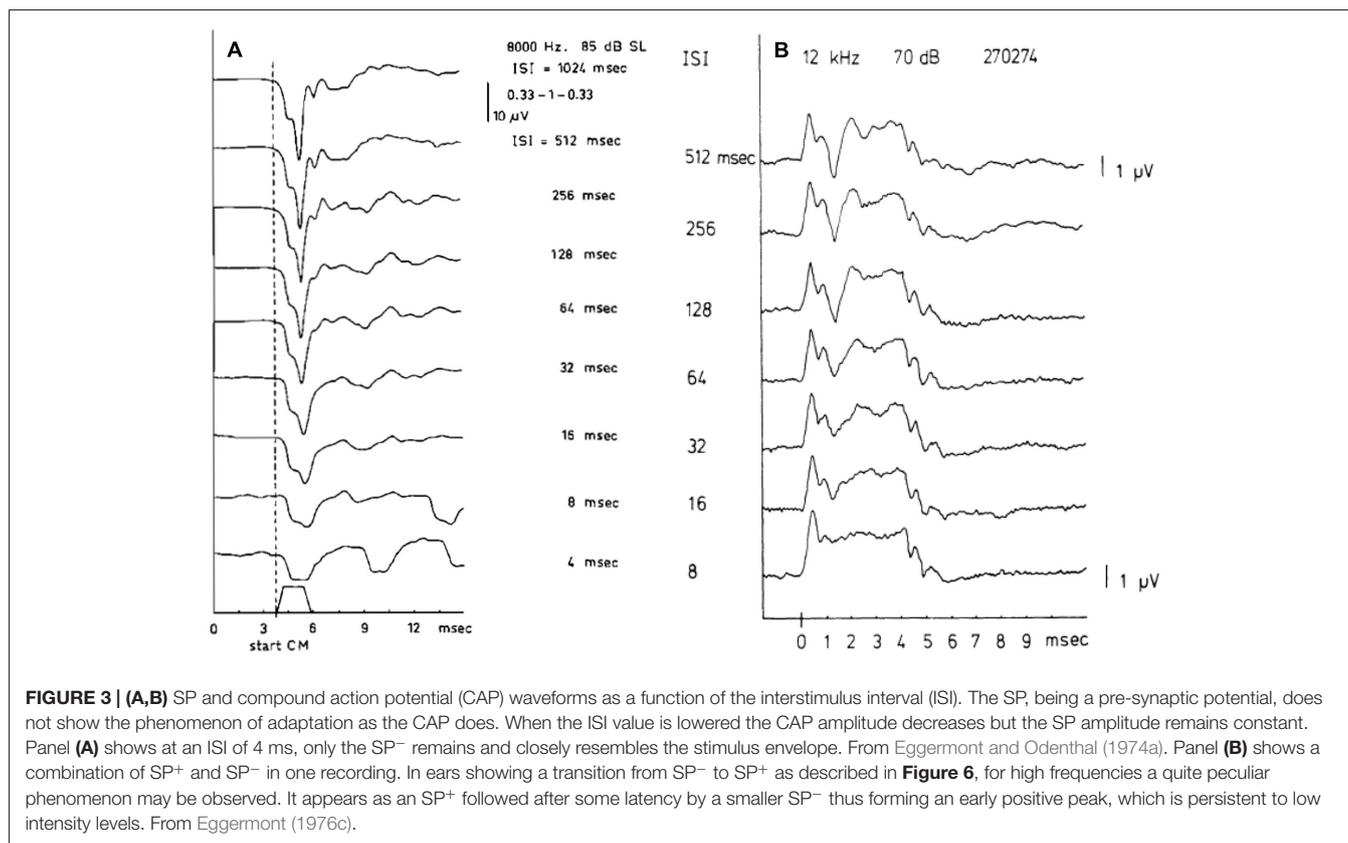
FIGURE 2 | Transition of a negative to a positive summating potential (SP^+) with increase in tone burst frequency. In some ears, a quite sudden change in the sign of the SP may be observed. In this example, a change occurs between 4750 Hz and 5175 Hz. This type of transition from SP^- to SP^+ typically occurs between 4 kHz and 8 kHz. From Eggermont (1976c).

of 4000 Hz at an intensity of 80 dB HL; an increase in the tone burst frequency to 4350 or 4750 Hz leads to a clear drop in the SP^- amplitude. A further increase in frequency to 5175 Hz gives an SP^+ , whose magnitude increases slightly when the frequency is raised further. The same type of change is observed from the AVE SP recorded from the guinea pig's first cochlear turn (Dallos, 1972), where, at an intensity of 60 dB SPL, the AVE SP is typically negative for frequencies up to 3000 Hz, about zero at 6000 Hz, and positive for higher frequencies. This could explain the changes if the promontory recorded SP is considered a mix of positive and negative AVE SPs generated in the basal turn depending on the resistance paths through the promontory into the cochlea and via the round window, i.e., the electroanatomy of the recording site.

As a pre-synaptic potential the SP will not be affected by adaptation. Increasing the repetition rate of the stimuli will isolate the SP^- from the CAP as illustrated in **Figure 3A**. This works for both the SP^+ and SP^- , but shows surprisingly in this example that the SP^+ may consist of a sharp transient and a sustained part (**Figure 3B**). This likely results from a combination of a short latency SP^+ followed by a superimposed slightly longer latency SP^- , both having a duration equal to the tone burst, and the SP^- originating from a slightly more apical region. We noted that this effect persists at least down to 55 dB HL. That this SP^+/SP^- complex is not of neural origin is demonstrated by its persistence at short interstimulus intervals (ISI; **Figure 3B**). The CAP disappears nearly completely at an ISI of 8 ms, but the SP combination remains. The finding that the SP^+ occurs more often in Ménière's disease (Eggermont, 1976c; Dauman et al., 1988) could be caused by a changed electroanatomy, potentially attributable to an endolymphatic hydrops.

The Cochlear Microphonics

I had never much faith in the clinical use of the CM (Eggermont, 1976c), amplified by the fact that in Ménière's disease the CM amplitude for 85 dB HL tone bursts was up to CAP thresholds of 70 dB HL independent of hearing loss (Eggermont, 1979a). However, as a consequence of the decisive use of CM in the diagnosis of auditory neuropathy (see "Auditory Neuropathy" Section) it is time to take a new look. The CM is an electric response that can be recorded from almost anywhere in the cochlea and from the cochlear surface (e.g., the round window), as first demonstrated by Wever and Bray (1930). Early on, Tasaki et al. (1954) showed that CM to all frequencies might be recorded with differential electrodes from the first turn in the guinea pig cochlea. Experiments in kanamycin-intoxicated guinea pigs, which destroys the OHC, showed that the CM produced by the IHCs was about 30–40 dB less sensitive than that generated by the OHCs (Dallos and Wang, 1974). However, absent CM is not an absolute indicator of non-functional OHCs, as Liberman et al. (2002) have shown in mice lacking prestin, the distortion-product oto-acoustic emissions (DPOAEs) are elevated to correspond to the hearing loss, whereas the CM is not significantly reduced compared to normal controls. The CM recorded at the promontory or in the ear canal thus arises primarily from OHCs in the more basal portions of the



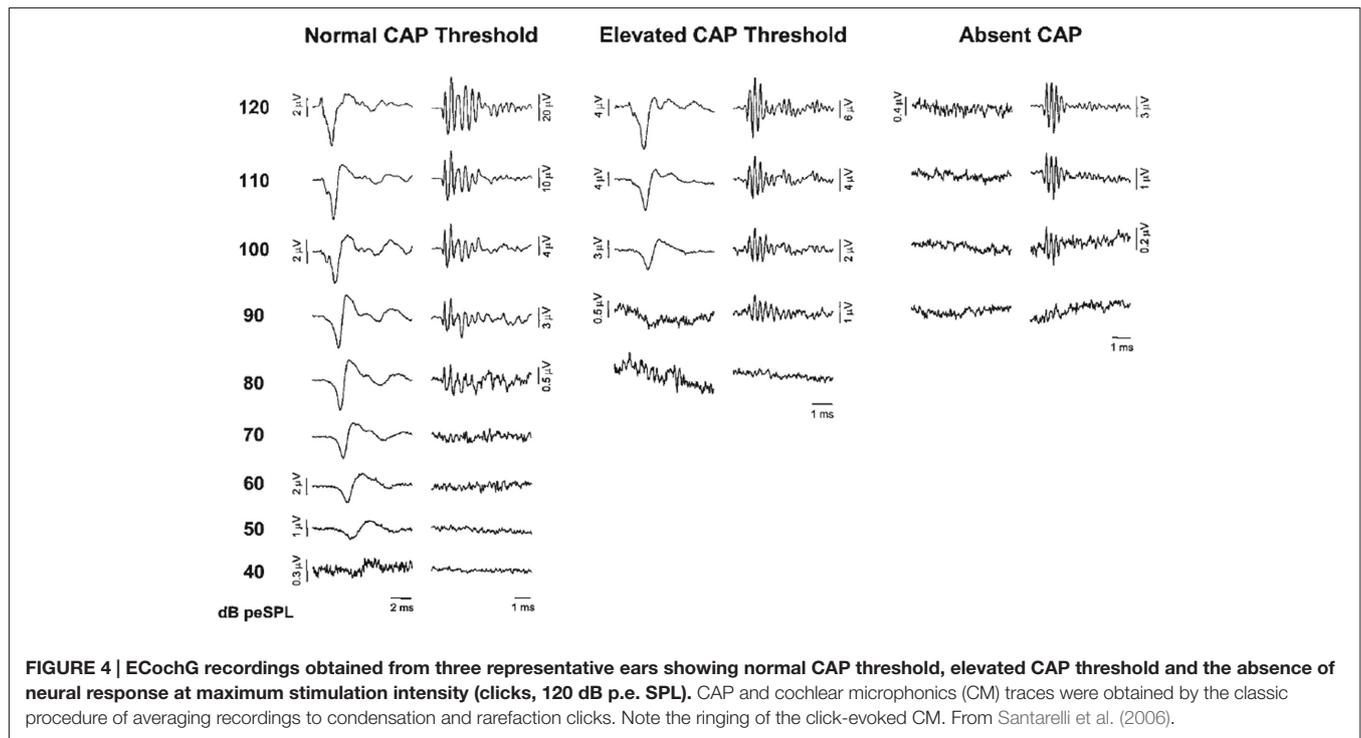
cochlea, while the apical regions make a negligible contribution to its generation (Johnstone and Johnstone, 1966; Patuzzi et al., 1989; Withnell, 2001). However, CM as recordable from the promontory may not only be generated in the basal turn and may for low frequencies also include neural contributions (Chertoff et al., 2012, 2014; Kamerer et al., 2016).

Santarelli et al. (2006) recorded CM, SP and CAPs using TT ECoChG in 502 normal hearing subjects and with varying degrees of sensorineural hearing impairment, and in 20 auditory neuropathy patients. They distinguished three categories (**Figure 4**), those with a normal CAP threshold in which case the CM to clicks is detectable to about 80 dB peak equivalent sound pressure level (p.e. SPL) (~50 dB HL), those with an elevated CAP threshold often accompanied by a CM with similar threshold, and those without CAP, where the CM might indicate functioning OHCs, as in auditory neuropathy. Santarelli et al. (2006) found that CM was almost always detected when recording TT ECoChG in ears with varying degrees of hearing impairment or even with profound hearing loss, and thus, in the presence of extensive OHCs loss (Eggermont, 1979a; Arslan et al., 1997; Schoonhoven et al., 1999). Even in the 202 ears of children (mean age 2.6 ± 4.2 years) with no CAPs recorded at 120 dB p.eSPL, the CM was always detected albeit with elevated threshold (99.1 ± 7.9 dB p.e.SPL, compared to 41.1 ± 9.5 dB in normal controls) and reduced amplitude (7.5 ± 9.7 μ V, compared to 29.1 ± 33.1 μ V in normal controls). According to Santarelli et al. (2006) “this finding challenges the widely accepted view

that the CM is strictly related to OHC electrical activity with only a minor contribution from IHCs”. An important finding was that the presence of central nervous system pathology and normal hearing thresholds seemed to enhance CM amplitude compared to normal hearing ears. This amplitude enhancement was often accompanied by prolonged CM duration, albeit that this duration enhancement was also observed in about half of completely normal ears (Gibbin et al., 1983; Liu et al., 1992; Santarelli et al., 2006). The amplitude enhancement was attributed to a dysfunction of the medial efferent system through a reduced inhibitory influence on OHCs, leading, in turn, to enhanced cochlear amplification. Santarelli et al. (2006) also compared DPOAEs with CM in the same ears with a wide range of CAP thresholds and found the presence of DPOAEs “a more sensitive indicator of hearing threshold preservation than CM amplitude”.

Interlude: Transtympanic vs. Extratympanic Recording

Of interest for addressing this choice I present four prospective studies that compared TT and ET ECoChG in the same ears, and in two cases by simultaneous recording. Mori et al. (1982) concluded that TT showed higher amplitudes but the same latency as ET. Noguchi et al. (1999) confirmed this and also found that TT and ET had same threshold detection levels, and same slopes for the CAP amplitude-intensity functions. Of relevance for diagnostics of Ménière’s disease, to be reviewed later, is



that TT recordings often show positive summing potentials (SP^+) for high-frequency tone bursts and negative summing potentials (SP^-) for lower frequencies, whereas in ET only SP^- were recorded (Mori et al., 1982). No significant difference in the SP/CAP ratio was found between TT and ET recordings (Roland et al., 1995). Schoonhoven et al. (1995) made simultaneous ET and TT recordings in 30 patients with various types and degrees of cochlear hearing loss. They found that ET responses were reduced in amplitude with respect to TT responses by a factor of 0.43 on average. ET and TT latencies were identical. This suggests that when the hearing loss is not too large both recording methods are equally applicable. Modifying the ET technique by using two identical high-impedance electrodes on the tympanic membrane (active) and as a reference in the ear canal, resulted in a signal to noise increase by >2.6 dB (Kumaragama et al., 2015).

The Compound Action Potential Phenomenology

I will introduce TT tone-burst ECochG with a typical intensity series of the CAP, obtained in a normal hearing subject for 2000 Hz tone burst stimulation (Figure 5). In this series of responses, an interesting transition takes place around 65 dB HL. If the intensity of 65 dB is taken as a starting point for the analysis, it is evident that an increase in intensity leads to a relative increase of the N_1 peak of the CAP with respect to the N_2 , whereas lowering the intensity favors the second peak over the first (Eggermont and Odenthal, 1974a,b; Eggermont et al., 1974; Eggermont, 1976c). A similar bifurcation around 55–60 dB HL for click responses was reported by Yoshie (1976). A detailed

analysis of the same phenomenon obtained in response to a 2 kHz half-sine wave stimulus from the external ear canal was carried out by Elberling (1973). He presented stimuli in consecutive 2.5 dB steps over the intensity range of 72.5–95 dB p.e. SPL. The two peaks were of about the same magnitude at 85–90 dB p.e. SPL, which corresponds to about 65–70 dB HL. It is tempting to attribute these two peaks to contributions from populations of auditory nerve fibers (ANFs) with low and medium thresholds respectively (Bourien et al., 2014).

A contrasting series of CAP waveforms for two types of sensorineural hearing loss with recruitment, resulting from: (a) Ménière's disease; and (b) neonatal asphyxia, is shown in Figure 6, again for stimulation with a 2000 Hz tone burst. It is noted that in the neonatal asphyxia waveforms only the early N_1 is present (see Figure 5), whereas in the Ménière ear the CAP is much broader and dominated by the relatively large and long lasting SP (see "The Cochlear Microphonics" Section).

In case of loudness recruitment one often (but not always; Eggermont, 1976c) observes a steep increase in the amplitude of the CAP with stimulus level as illustrated in Figure 7. This shows a series of typical input-output curves for Ménière ears with the median curve obtained in 20 normal ears. All ears show the increase in steepness compared to the median control amplitude-level function (for which the threshold at the $0.1 \mu V$ level was at 0 dB HL). This mimics the steeper increase of loudness with increasing sound level.

Early on it was noted that the adaptation and post-masking recovery in human CAPs was clearly different from animal data (Coats and Dickey, 1972; Eggermont and Spoor, 1973; Eggermont and Odenthal, 1974a,b). This is illustrated in

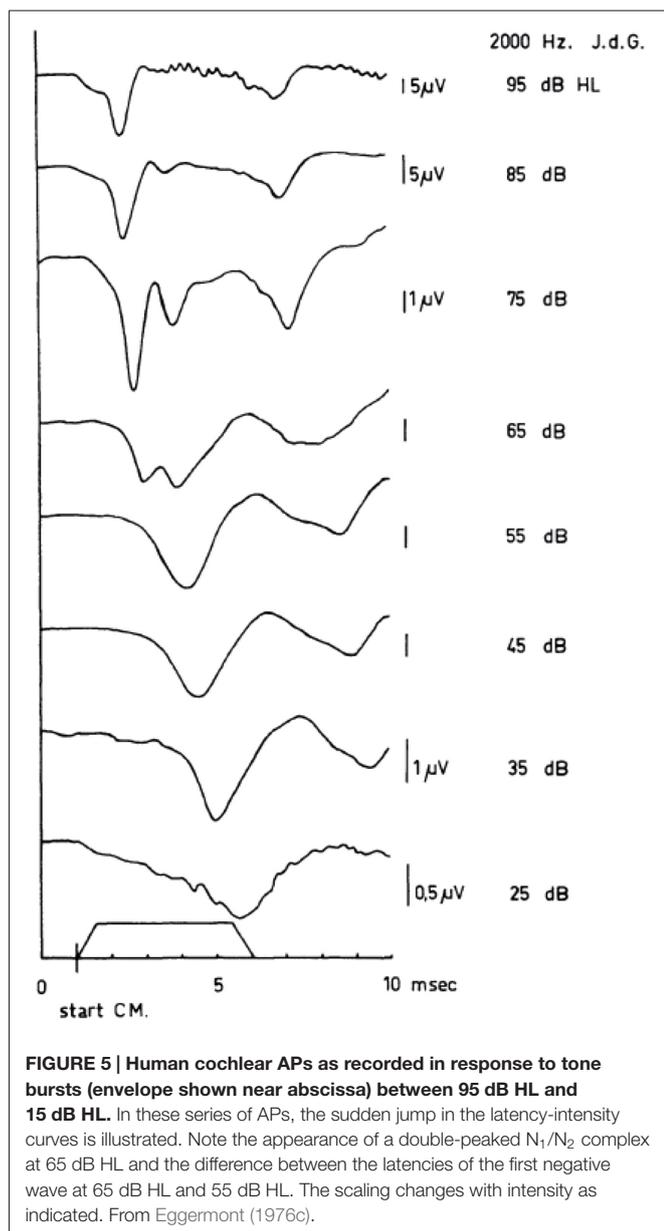


Figure 8. Here we compare the adaptation of the CAP amplitude for stimulation with tone-burst trains of various ISI and in the recovery from forward masking as a function of post-masker delay (Δt) in guinea pigs and humans. Coats and Dickey (1972) found that the post masking recovery of click loudness in their ECochG participants was nearly complete at $\Delta t = 100$ ms, which compared well with the animal electrophysiological results, but not with the human ECochG. This suggests that CAPs, which depend on neural firing synchrony, do not reflect loudness measures.

The Composition of the Compound Action Potential

This topic got a lot of attention in the 1970s, where investigators aimed at understanding how to interpret human ECochG recordings. Early on, Gasser and Grundfest (1939) had used

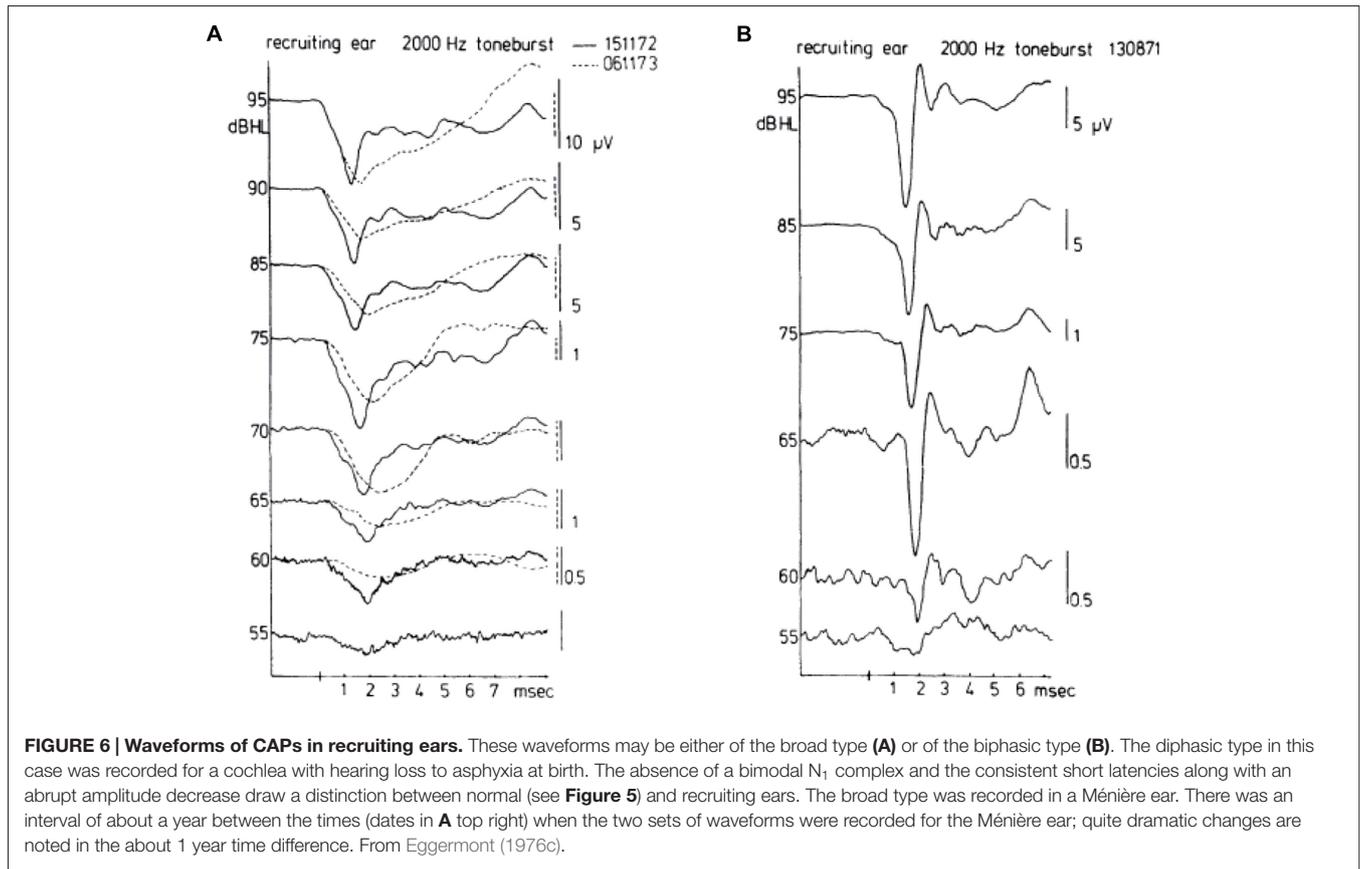
convolution to predict the waveform of the CAP evoked by electrical stimulation of the saphenous nerve of the cat from the distribution of nerve fiber diameters (resulting in a latency distribution) and a hypothetical individual fiber unit response. Twenty years later, Goldstein and Kiang (1958) pointed out that, under the assumption that unit responses add with equal weight to the recording electrode, the CAP-waveform could indeed be expressed as a convolution integral:

$$CAP(t) = N \int_0^t s(\tau) a(t - \tau) d\tau$$

where N is the number of nerve fibers, $s(t)$ the latency distribution function and $a(t)$ the unit response. A unit response, recorded from a nerve end, will be normally diphasic in shape and this has been postulated for the auditory nerve by Teas et al. (1962), de Boer (1975) and Elberling (1976a,b) and first demonstrated by Kiang et al. (1976). The convolution is allowed under the conditions of statistical independency of the individual contributions. When using a click as a stimulus, the latency distribution function may be considered reflecting the envelope of the impulse response function of the peripheral hearing organ. For individual fibers such impulse response functions may be obtained from the cross correlation between the nerve fiber response and a white noise stimulus evoking them (de Boer, 1969).

Investigating the single nerve fiber firing pattern for non-click stimuli will result in a modified weighting function $s^*(t)$ which may be found by convolution of the true impulse response and the stimulus envelope. A second convolution of the new $s^*(t)$ with the unit response $a(t)$ will then give the CAP to this new stimulus (de Boer, 1975) after summation over all contributing units. In a practical situation, either in modeling or analyzing, the number of contributing units has to be restricted. This may be done by forming groups of nearly equivalent units. It might thus be useful to divide the cochlear partition into small regions about 3 mm long (corresponding to about half-an-octave in frequency) and study the narrow-band CAPs (NAPs) evoked on these small segments. Since the human cochlea is innervated by about 25,000 (Hall, 1967) to 31,000 (Rasmussen, 1940) afferent nerve fibers, such a 3 mm segment is assumed to comprise about 2500–3100 individual nerve fibers. The thresholds of the fibers in each segment are supposed to be approximately distributed in the same way across low-, medium- and high-threshold ones (Kiang et al., 1965; Rutherford and Roberts, 2008; Bourien et al., 2014).

Teas et al. (1962) introduced an experimental technique for such a separation of the CAP recorded from the guinea pig cochlea into about 10 NAPs. A high-pass noise-masking stimulus with a number of discrete high-pass cut-off frequencies was used. Subtracting CAP responses obtained in the presence of high-pass noise with cutoff frequencies being $\frac{1}{2}$ octave apart, results in NAPs, which can be assigned to particular narrow-band segments each characterized by a central frequency (CF). This technique has first been used in human ET ECochG by Elberling (1974) for the analysis of click-evoked CAPs. Later on this method was applied by Eggermont (1976d, 1979b,c) using TT

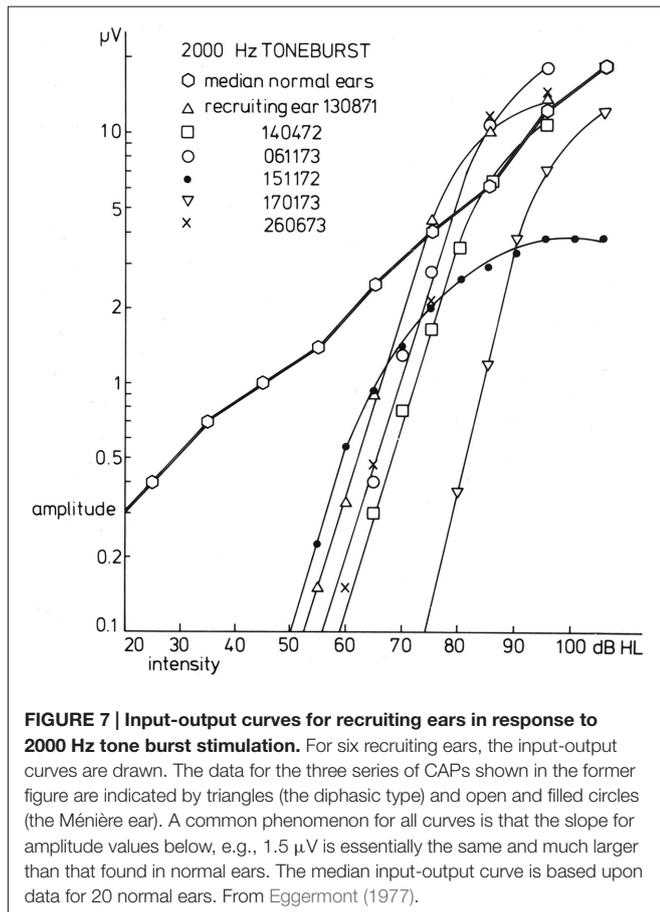


recording of responses to click and tone burst stimuli to elucidate the frequency specific character of these types of stimulation. An example of such a separation of the CAP into NAPs for the human cochlea upon click stimulation is shown in Figure 9. The click intensity is 90 dB p.e. SPL, and the NAPs are essentially diphasic in shape and their latencies range from 1.4 ms to 5.8 ms. The CAP latency is 1.4 ms and is therefore mainly dominated by the most basal contributions, due to the diphasic waveforms the contributions from segments with lower CFs tend to cancel each other and are therefore not seen in the CAP. It seems appropriate to use narrow-band waveform for the highest CF, with the shortest duration, as an estimate of the unit response. It is noted that double peaked CAP responses as shown for 2 kHz tone burst in Figure 5, and observed here for 4 kHz high-pass noise masking of the click evoked CAP, are not the result of changes in the NAP waveforms but result from changes in the cancellation of responses from different CF regions.

A plot of the NAP-amplitude (negative deflection only) as a function of the CF, which may be related to distance from the stapes (Greenwood, 1961; von Békésy, 1963), shows for a click level of 90 dB p.e. SPL (Figure 10) a gradual increase in amplitude for higher central frequencies. For lower click intensities, the contributions from both the high- and the low-frequency side rapidly decrease, while the central region (about 3 kHz) still contributes the same. For relatively

low intensities, the activation area seems to be reduced to a more narrow frequency-selective region likely related to the external ear canal and middle ear resonances, which favor the parts in the spectrum around 2–3 kHz, where the human ear has its greatest sensitivity. In normal ears, and ears with high-frequency hearing loss, click evoked CAP thresholds will reflect the patency of this 2–4 kHz region.

More recently, in an elaborate and detailed study, Lichtenhan and Chertoff (2008) were able to estimate the number of ANFs, N , contributing to the CAP, as well as the post-stimulus time histogram summed across nerve fibers, $s(t)$, and unit response, $a(t)$, before and after TTS. They found that TTS resulted in: a broadening and decreased latency of $s(t)$, and decreased N . Their model unit response, $a(t)$, based on the whole nerve click CAP showed a lower oscillation frequency and more rapid decay. This could have been improved by using a high CF NAP. These results suggested that TTS causes fewer ANFs to contribute to the CAP and those that do are more basally located with lower response synchrony and more quickly decaying and lower frequency oscillations. Lichtenhan and Chertoff (2008) suggested that this type of analysis might be useful in quantifying the number and location of surviving ANFs in patients with hearing loss. Similarly, Earl and Chertoff (2010) fit the analytic CAP to gerbils with partial lesions of the auditory nerve. The model parameter N at high-stimulus levels

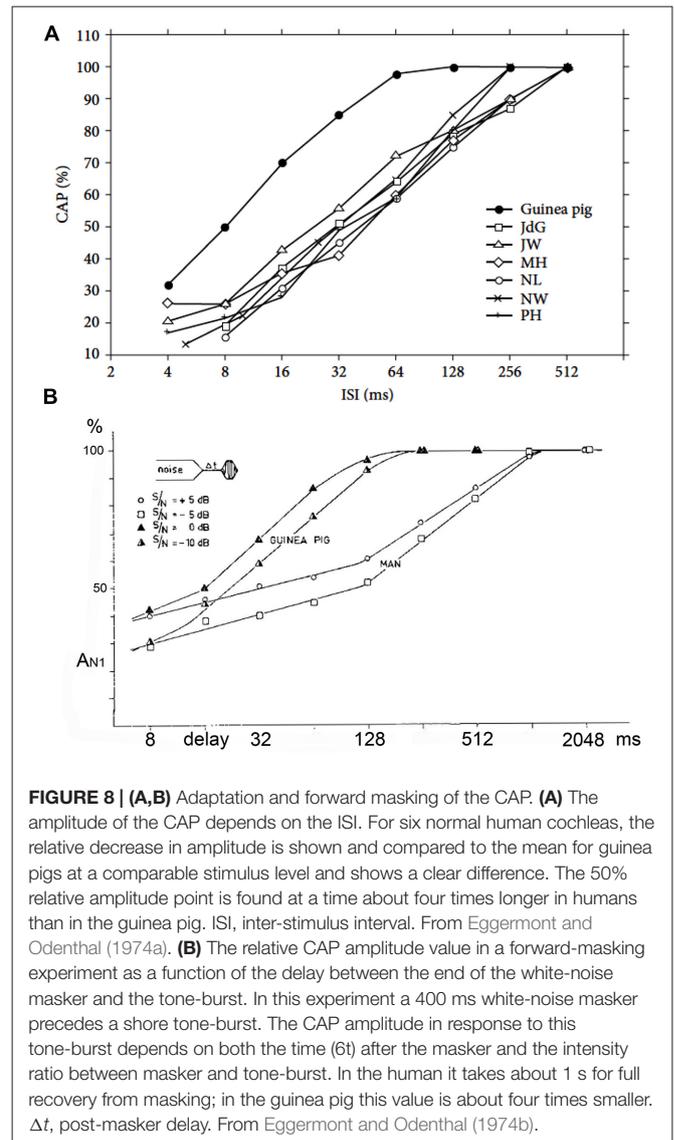


was strongly correlated with normal nerve area suggesting, that it is a good predictor of auditory nerve survival. The model parameter N also seemed to be a better predictor of the condition of the auditory nerve than the conventional measure of CAP amplitude.

Validation of the Use of NAPs

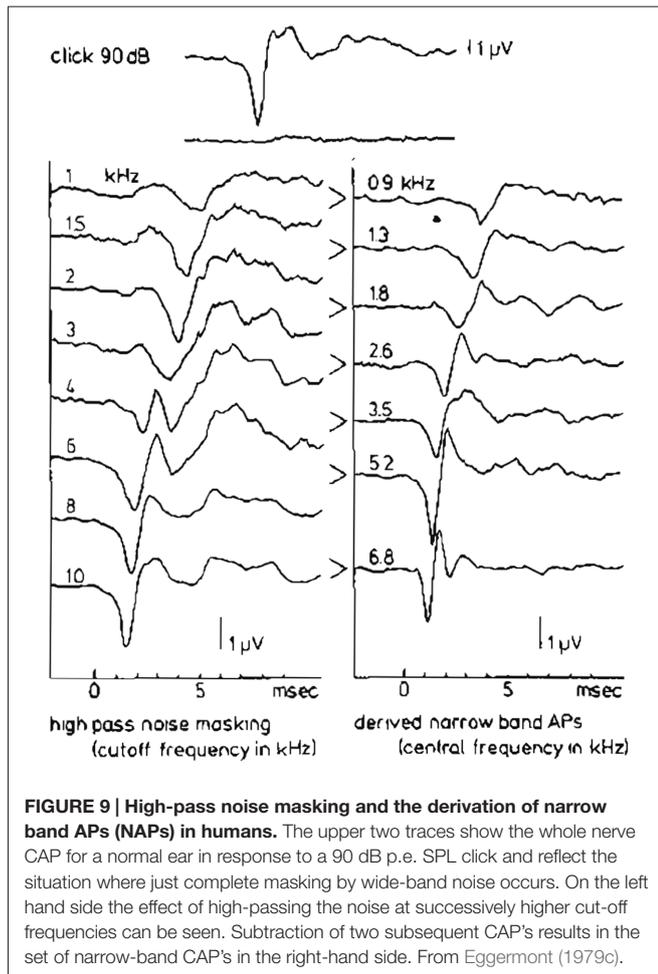
Evans and Elberling (1982) validated the use of the high-pass noise masking technique by comparing single-unit recordings and CAP measurements in the cat under conditions of high-pass masking. They computed the NAPs in a cat and compared them with the CFs of single cochlear fiber responses contributing to these NAP regions. With one main exception, the conclusions drawn on the origin of the frequency components of the NAPs were found to be valid in the normal cat. The exceptions were fibers with characteristic frequencies below 1–2 kHz, where the high-pass masking derived location was less specific. Taking into account the low-frequency hearing range of the cat, which is shifted upwards with about 1 octave compared to humans, Evans and Elberling (1982) predicted that the high-pass masking technique would be valid in normal humans for frequencies down to 0.5–1 kHz. This in effect validated the use of the latency distribution function.

Further experimental evidence for the applicability of the NAP technique in pathological cochleas came from recordings



in normal and noise-exposed guinea pigs (Versnel et al., 1992), which looked at the validity of using the same unit response along the CF range and in normal vs. hearing loss ears. They used a technique pioneered by Kiang et al. (1976) involving spike-triggered averaging of round window “noise”. In that way one can estimate the unit response for units with CFs corresponding to locations along the cochlear partition. Their findings in normal cochleas confirmed the earlier data from Prijs (1986), namely that the unit response was diphasic and had a fairly constant amplitude of about 0.1 μV . In noise-exposed cochleas, waveform, latency and amplitude of the negative component of the unit response remained unchanged.

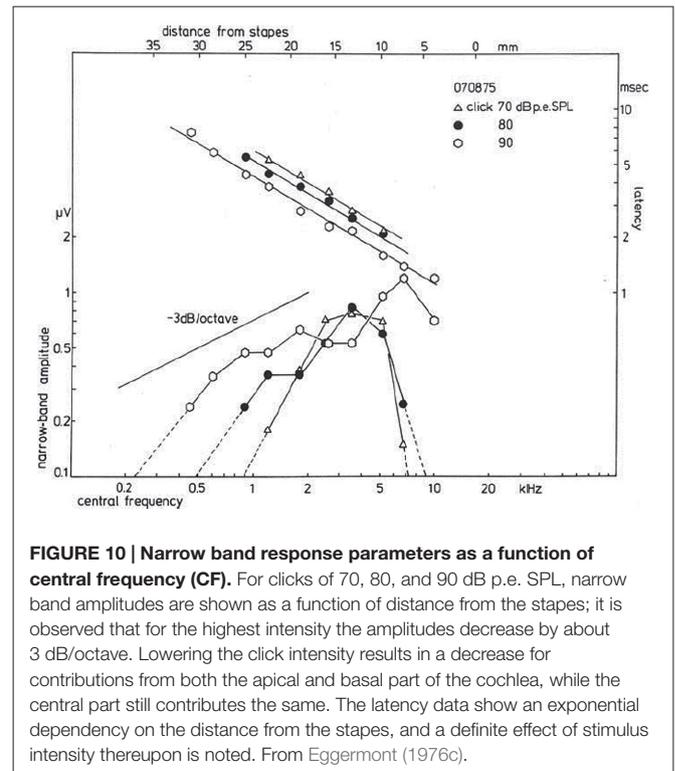
Delays estimated from NAPs have recently been used to generate chirps, which synchronize auditory nerve discharges along the length of the cochlea and yield larger amplitude CAP responses than clicks, presumably due to greater ANF synchrony along the cochlear partition (Chertoff et al., 2010).



Diagnosis Based on the Waveform of the Compound Action Potential

Portmann and Aran (1971) were the first to point to a potential diagnostic use of the click-evoked CAP waveform. They distinguished four typical response patterns: the normal response, the recruiting response (not unlike that in **Figure 6B**), the broad or prolonged response often seen in Ménière ears (see **Figure 6A**), and the abnormal response, which showed an initially positive SP (**Figure 3B**). Yoshie (1976) also paid attention to the abnormal waveforms found in Ménière ears and resulting from SP⁺ and/or SP⁻ interaction with the CAP. Much attention was paid on the so-called low- and high-amplitude and latency functions with a cross-over point at the bifurcating CAP waveform (see **Figure 5**).

However, these typical waveforms and their presumed reflection of the underlying disturbances in the peripheral hearing organ can be studied more insightfully by using the narrow-band response derivation. **Figure 11** shows such a (CF-restricted in these illustrations) narrow-band analysis for a normal ear, a Ménière ear and for an ear affected by an acoustic neuroma (vestibular Schwannoma). For the normal ear the narrow band responses at the three central frequencies shown are essentially biphasic in shape (see **Figure 9**). Since in this

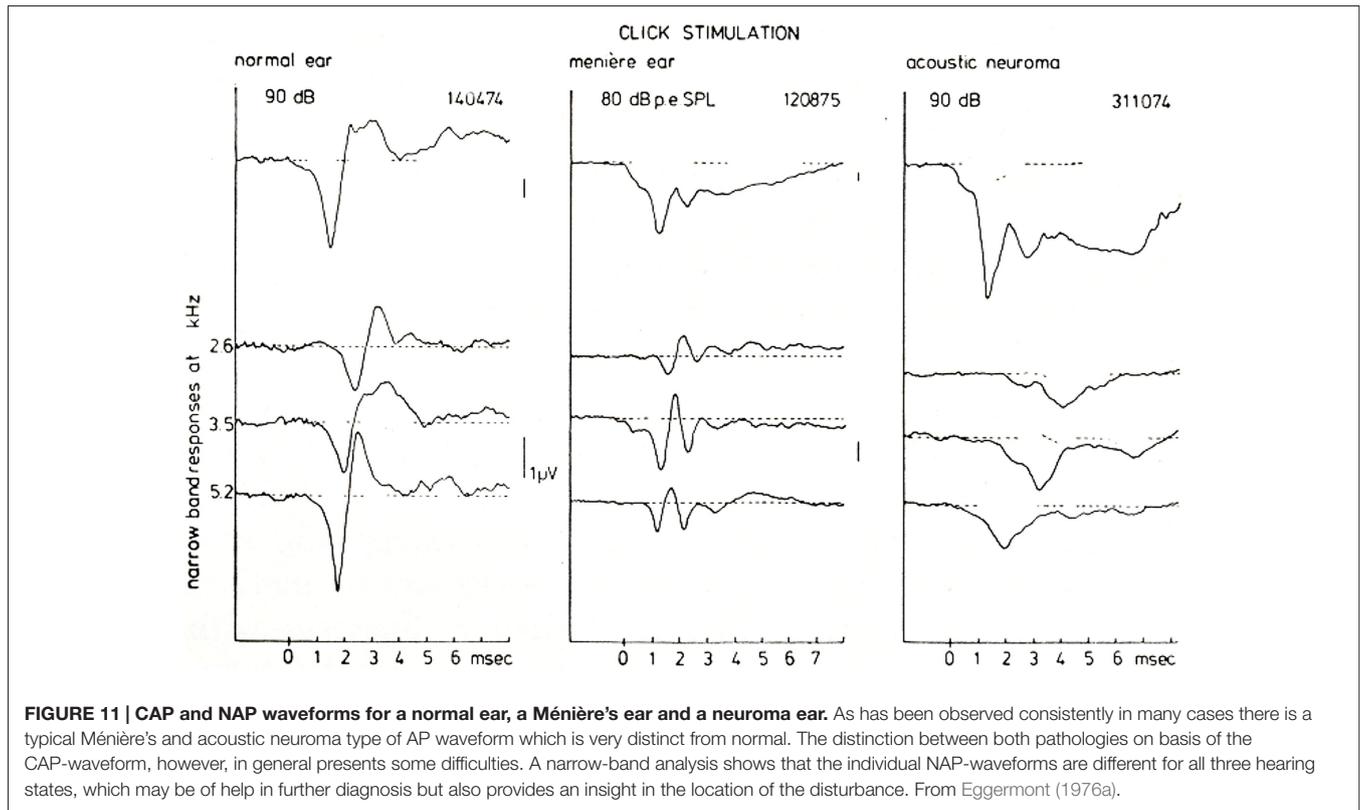


basal part of the cochlea the traveling wave velocity is around 20 m/s (Eggermont, 1976d), these 3 mm wide narrow bands are traversed by the traveling wave in about 0.15 ms. One may say therefore that these single nerve fibers will fire in nearly perfect synchrony. This implies that for the most basal part of the cochlea the NAP reflects the unit-response waveform contribution to the CAP.

For the Ménière ear the recorded CAP is dominated to a large extent by the relatively large negative SP. The narrow band analysis, however, shows an additional feature, namely that the unit contribution is composed out of two biphasic waveforms with a delay of about 1 ms. This may point to repeated firing by the fibers in the indicated narrow-bands in response to the same click. This fact may contribute to the over-recruitment often observed in Ménière ears but this will need a more detailed study.

The acoustic neuroma ear shows essentially the same type of broad CAP waveform as found in the Ménière ear (see also Aran, 1971). However, the SP is relatively smaller than the CAP and seems not to account for the broadening of the CAP in the same way as in the Ménière ear. High-pass masking shows that the NAPs are monophasic in this situation. The addition of the NAPs therefore does not produce cancellation of activity after the onset of the CAP as found in normal and Ménière ears, but instead produces a broad CAP. In this situation the NAP waveform may reflect a change in the unit contribution as a result of nerve conduction block due to the presence of the tumor (Beagley et al., 1977).

The mechanisms that produce these striking differences in NAPs seem very useful in diagnosis. Especially the close



similarity of the CAP waveforms for the Ménière ear and the neuroma ear is completely removed when looking at the narrow band responses.

CLINICAL APPLICATIONS

Objective Audiograms

Sample objective audiograms for frequencies of 500 Hz to 8 kHz obtained with TT tone burst ECoChG were first shown in Eggermont et al. (1974). For a more restricted frequency range, Yoshie (1973) performed a regression analysis in 56 patients between TT CAP thresholds for tone pips with the audiometric thresholds for the frequencies 2, 4, and 8 kHz. The regression lines showed slopes that ranged from 0.75 (2000 Hz) to 0.83 (4000 Hz) with correlation coefficients very near 0.90. Almost all of the points in his scattergram were within ± 15 dB from the regression line, suggesting good clinical application.

In a group of 96 patients in which behavioral audiometry was available, Spoor and Eggermont (1976) compared the audiogram with TT ECoChG tone burst evoked CAP thresholds. Given an ECoChG threshold, the practical question concerns the prediction of the subjective threshold. Regression analysis showed that the slope of the regression line was close to unity for each frequency. For 1, 2 and 4 kHz, the mean difference between ECoChG and subjective measures was 0 dB. At 500 Hz, the mean difference between the ECoChG and the subjective thresholds was

about -10 dB, i.e., the subjective threshold is 10 dB higher than the ECoChG threshold. At 8000 Hz this was the same, and the spread of ECoChG thresholds at 500 Hz and 8 kHz was higher than at 1, 2 and 4 kHz. Standard deviations for the different frequencies varied from 7.5 dB to 11 dB, resulting in a 95% confidence level of 15–22 dB around the mean.

Schoonhoven et al. (1996) further investigated the relation between TT response thresholds for tone bursts with octave frequencies from 500 Hz to 8000 Hz and audiometric thresholds in 148 ears. Similar analyses of ET thresholds were reported for a subset of 30 ears in which TT and ET physiological responses were simultaneously recorded. They found that TT ECoChG thresholds were highly correlated with audiometric thresholds. Linear regression analysis showed that audiometric thresholds might be predicted from physiological thresholds with an error in the estimate of 11 dB. ET ECoChG permitted similar predictions but with a larger uncertainty of 16 dB. It appeared that ECoChG thresholds increase slightly less with increasing cochlear dysfunction than do pure tone thresholds. They considered this a result of the different stimulus durations on which the two threshold measurements are based and the difference in temporal integration between normal and pathological ears.

Recent animal studies have provided an interesting alternative. Lichtenhan et al. (2013) described a novel technique to estimate low-frequency cochlear thresholds that uses the auditory nerve overlapped waveform (ANOW) response in the guinea pig. They showed that for frequencies of 700 Hz and

below, ANOW thresholds were mostly 10–20 dB more sensitive than onset-CAP thresholds and 10–20 dB less sensitive than the most sensitive single-AN-fiber thresholds. The results show that ANOW can be used to objectively estimate thresholds at very low frequencies in a high frequency-specific manner. A subsequent study (Lichtenhan et al., 2014) demonstrated that in guinea pigs this ANOW response originates in the apex of the cochlea. This technique could potentially be used to assess very low frequency information more accurately than current ECoChG procedures allow.

Ménière's Disease

The Importance of the Summating Potential for Diagnosis

The first report using non-surgical recording (TT) in Ménière patients ($N = 22$) was by Schmidt et al. (1974). They reported that the SP^- value, although often pronounced compared to the CAP amplitude, was almost the same as found in normal hearing ears, however distinctly larger than the SP^- amplitude observed in non-Ménière ears with high-frequency hearing loss. The first abnormal waveforms in a Ménière patient were shown in Schmidt et al. (1975) and later in Eggermont (1976a) and Odenthal and Eggermont (1976), all using TT ECoChG. The use of the SP/AP amplitude ratio was first reported at an ECoChG conference in 1974 organized by Bob Ruben in New York City and later published in Eggermont (1976b). However, I found this technique not useful for diagnosing individual patients. The mean SP/AP ratio in normal ears was level dependent and decreasing from about 0.3 at 95 dB HL to 0.07 at 55 dB HL, the upper limit was barely level dependent and about 0.45. In Ménière ears, the mean SP/AP value was nearly level (55–95 dB HL) independent at 0.35, with upper limits up to 0.6. In “hair cell loss” ears, the mean SP/AP ratio was strongly level dependent, from 0.25 at 95 dB HL decreasing to 0.06 at 75 dB HL. The upper boundary was around 0.6 at 95 dB HL and decreasing to 0.35 at 75 dB HL. Example waveforms contrasting a Ménière ear and a non-Ménière hearing loss ear, were shown in **Figure 6**. Note that the CAP amplitude in this study was taken from the level of the SP^- and not from the baseline, which would include the SP^- in case of tone burst stimulation, and thus reduces the calculated SP/AP ratio. For tone burst evoked responses measuring CAP amplitude from the SP level (either SP^+ or SP^-) seems to be the best procedure. For click evoked responses it is more difficult to assess the decaying SP level and here the least ambiguous way would be calculating amplitudes with respect to the pre stimulus baseline. Separate norms have to be established for ET and TT recordings.

Gibson et al. (1977) were more optimistic for SP use in diagnostic procedures by the observation of an “apparent widening of the SP/AP waveform”. They considered this as caused by an enlarged SP^- , enhanced relative to the CAP, and “believed to be related directly to the presence of endolymphatic hydrops”. This was followed up Gibson et al. (1983) by a comparison of 32 normal, 40 sensory-loss ears, and 44 Ménière ears. They concluded that the diagnostic value would be increased if the SP amplitude was expressed as a percentage of the

CAP amplitude, i.e., as an SP/AP ratio. In normal ears, the mean SP/AP ratio was 25% (range 10%–63%). In sensory damage, the SP/AP ratio was on average 13% (range 0%–29%), and in Ménière's ears, the mean SP/AP ratio was 51% (range 29%–89%). In this series, an SP/AP ratio of 29% suggested a useful diagnostic dividing mark between the sensory damage and ears affected with Ménière's disease. Note the large overlap in SP/AP range between normal and Ménière ears.

An extensive study of the SP^- in 112 patients with Ménière's disease compared to 22 normal ears was carried out by Eggermont (1979a). He divided the Ménière ears in a low-threshold (≤ 50 dB HL) and a high-threshold (> 50 dB HL) group. The SP^- values at a range of intensity levels (55–85 dB HL) were not significantly different from normal for the low-threshold group, whereas the high-threshold group showed significantly smaller SP^- amplitudes for 2, 4 and 8 kHz tone bursts. For 2 kHz the median amplitude value was independent of the hearing threshold up to 45 dB HL, and for larger losses there was a sharp decrease in the SP^- amplitude. The same phenomenon was found for 4000 Hz: up to 55 dB HL there was a slow decrease in the SP^- amplitude, and for higher threshold values a sharp loss. Thus the pattern at both frequencies showed a boundary value around 50 dB HL. The changes at 8000 Hz, however, seem more gradual relative to the amount of hearing loss, making separation artificial at this frequency. Eggermont (1979a) concluded that “in Ménière ears hearing losses up to about 50 dB are not related to changes in the hair cells, since the SP^- does not change, whereas the increase in the amount of hearing loss above 50 dB HL is paralleled by a loss in sensitivity of the SP and is therefore related to a functional loss of hair cells”.

Coats (1981) used clicks and ET recording, and measured both the SP^- and the N_1 from baseline, which tend to make the SP/AP, in fact an $SP/(SP + AP)$, ratio smaller. However, this may have a small effect when using clicks as the SP then is of small duration. Despite that Coats found that the SP/AP ratio for detection was 64%. I would not consider this a value useful for diagnosing individual cases. Goin et al. (1982) reported that the SP/AP was the most efficient diagnostic measure, with 62% of the Ménière's group demonstrating abnormal ratios compared to 4% of the normal control group and 17% of the cochlear group. However, they did not report the “abnormal value” used. Kanzaki et al. (1982) using TT and ET ECoChG found that “It was not possible to differentiate Ménière's disease from sudden deafness on the basis of large SP/AP ratios alone. Such ratios were found frequently in both diseases”. Ferraro et al. (1985) used ET ECoChG in 55 suspected Ménière patients and found that “the presence of hearing loss combined with aural fullness or pressure was the strongest predictor of an enlarged SP/AP ratio”. The Bordeaux group (Dauman et al., 1988) investigated the SP to 1, 2, 4 and 8 kHz tone bursts in 50 Ménière patients, 10 sensory loss patients and five normal hearing controls. They found that the mean SP amplitude was larger in the Ménière's disease group for 1, 2 and 8 kHz compared to controls. However, the ears with larger negative SPs at low frequencies also had larger CAPs, measured from the level of the SP.

In a large series of studies Mori et al. (1987a) investigated differences between TT and ET ECoChGs in the use of the SP/AP ratio for click and tone burst stimuli. The N_1 amplitude included the tail of the SP. They found that the SP⁻/AP ratio at 80 dB nHL was higher for a click with the ET than with the TT method. The SP⁻ elicited by tone bursts of mid to low frequencies was found more stable in Ménière's disease than SP⁻ elicited by a click (Mori et al., 1987b). An important observation was that there was no relationship between the ratio of the SP⁻ amplitude between both ears and the hearing threshold level at any frequency. In contrast, CAP amplitude ratio between both ears was significantly correlated ($r = -0.419$, $p < 0.01$) to the average hearing threshold level at 2–8 kHz, but not at 0.25–1 kHz ($p > 0.05$). This suggested that the increase in the SP⁻/AP ratio with the deterioration of the hearing at higher frequencies (Mori et al., 1987b) resulted from a decrease in CAP amplitude rather than an increase in SP⁻ amplitude (Mori et al., 1988; Asai and Mori, 1989). When the SP⁻/AP ratio threshold for abnormality was set at 0.43, they found that “ears with abnormal SP⁻ had a significantly worse hearing loss at high frequencies (2–8 kHz) than ears with normal SP⁻, whereas there was no significant difference in hearing loss at low frequencies (0.25–1 kHz) between both ears” (Mori et al., 1993).

The value of the SP/AP ratio that is considered indicative for Ménière's disease varies between studies. We have seen that Gibson et al. (1983) favored a value of 0.29, whereas Mori et al. (1993) used 0.43. Koyuncu et al. (1994) used 0.33, Aso and Watanabe (1994) suggested 0.42, Pou et al. (1996) used a definite positive result for a ratio >0.5 , and definite negative below 0.35. In a meta analysis of various studies, Wuyts et al. (1997) proposed an SP/AP ratio for click stimulation >0.35 using TT-ECoChG, or >0.42 using ET-ECoChG, as indicative of hydrops.

Specificity and sensitivity is important for any diagnostic test. Sass (1998) used TT ECoChG in a group of 61 patients (61 ears) with the clinical diagnosis of Ménière's disease and 15 patients (21 ears) with cochlear hearing loss of other etiologies, and 13 normal hearing subjects to assess the ability of the SP/AP ratio method to separate different cochlear disorders. Sass (1998) found a sensitivity of the click SP/AP ratio of 62% and a specificity of 95%. Inclusion of the 1-kHz burst-evoked SP amplitudes increased sensitivity to 82%, without changing specificity. Inclusion of the 2 kHz tone burst had no further effect on sensitivity or specificity. Sass et al. (1998) added the latency difference for condensation and rarefaction clicks, which was significantly larger in Ménière's disease compared to normal and non-Ménière hearing loss (as was also found by Orchik et al., 1998; Ge and Shea, 2002), and found that “the sensitivity of TT ECoChG, obtained by using measurements of SP/AP ratios and the SP amplitude at 1 kHz burst stimulation, increased from 83% to 87% by addition of the condensation-rarefaction shift measurement”. The specificity of TT ECoChG obtained by this combination of variables was 100%.

Negative outlooks on the use of ECoChG parameters in the diagnosis of Ménière's disease started to emerge in the

late 1990s. Levine et al. (1998) using CAP amplitude, SP amplitude, and CAP latency concluded that: “ECOG has limited value in the diagnosis of Ménière's disease. It appears to correlate with the length of time patients experience symptoms and their audiometric findings. It was not correlated with the number of symptoms that the patient experienced at the time that the study was conducted”. This was echoed by Kim et al. (2005), who reported that abnormally elevated SP/AP ratios (>0.4) in definite Ménière's disease were found in 66.7%. In less than definite Ménière's disease this was only slightly lower by 52.7%, which was not significantly different. Consequently, based on the SP/AP ratio approximately 30% of those with definite Ménière's disease would not be classified as having Ménière's disease. Because of its lack of sensitivity, ECoChG was considered not to play a decisive role in determining the presence or absence of Ménière's disease. Gibson (2009) also found that click SP/AP measurements did not significantly differentiate between Ménière's ears and non-Ménière's ears. However, tone burst SP-amplitude measurements were found significantly different between the two groups, particularly for frequencies at 500 Hz, 1 kHz, and 2 kHz. Recently, Oh et al. (2014) reported that: “Statistically significant differences were not demonstrated in the SP/AP amplitude ratio or SP_{area}/AP_{area} ratio between the definite Ménière's, probable Ménière's, overall Ménière's, or control groups”. These less than positive findings were echoed by a questionnaire on the clinical utility of ECoChG in the diagnosis of Ménière's disease among members of the American Otological Society (AOS) and American Neurotology Society (ANS). It was found that “For approximately half of respondents, ECoChG has no role in their clinical practice. ECoChG was used routinely by only 1 in 6 respondents” (Nguyen et al., 2010). However, introducing more extensive measures such as SP/AP area ratio (Ferraro and Tibbils, 1999) in some studies appeared to increase the diagnostic sensitivity (Devaiah et al., 2003). By combining SP amplitude, SP area, SP/AP area ratio and total SP-AP area, sensitivity and specificity values increased to 92% and 84%, respectively (Al-momani et al., 2009). In contrast, Baba et al. (2009) found that the combination of these parameters as well as using SP/AP area alone did not have greater sensitivity than SP/AP amplitude ratios.

Evaluating Mechanisms of Ménière's Disease

Dehydrating agents such as glycerol have been routinely administered since the report of Klockhoff and Lindblom (1966) to reduce the presumed endolymphatic hydrops in Ménière's disease and improve hearing thresholds. Here are some of the pioneering ECoChG studies. Moffat et al. (1978) tested 13 patients diagnosed with Ménière's disease using TT ECoChG during glycerol administration. Decrease of the SP⁻ was a common finding and occurred more often than threshold changes. Coats and Alford (1981) administered glycerol to 11 Ménière and 20 non-Ménière ears. ET-recorded SP amplitudes decreased, and 250–1000 Hz thresholds improved, and CAP amplitudes from the ears with Ménière's disease also decreased after glycerol ingestion, but to a lesser degree. None of these changes

were found in non-Ménière ears. Gibson and Morrison (1983) presented a single case study showing a large SP compared to the CAP, which after dehydration with glycerol showed a decrease in the SP and no change in the CAP so that the SP/CAP ratio became almost normal. Dauman et al. (1986) evaluated the “effect of orally administered glycerol on the SP and CAP amplitudes by means of automated recordings repeated every 5 min. SP values were remarkably constant in the control group. A decrease in SP absolute amplitude was observed in most patients with Ménière’s disease and some subjects with uncertain diagnoses, specifically at low frequencies.” Takeda and Kakigi (2010) evaluated 632 patients (727 ears) with vertigo/dizziness, of which 334 patients had a definite Ménière’s diagnosis. They found an enhanced SP in 56.3% of patients with Ménière’s disease, mostly where the disease duration was ≥ 2 years and/or the frequency of attacks was several times a year. Hearing improvement induced by the glycerol test did not produce a change in the SP/AP ratio—likely because both SP and AP increased or decreased together—and there was no significant difference between the glycerol test results and the incidence of an enhanced SP. Takeda and Kakigi (2010) suggested that the ECoChG seems to indicate that the enhanced SP in Ménière’s disease might be caused by the malfunction of the hair cells, not by the displacement of the basilar membrane toward the scala tympani, i.e., not by an endolymphatic hydrops. Fukuoka et al. (2012) evaluated 20 patients with a 3T MRI scanner and ECoChG after glycerol application. They found that ECoChG was positive (SP/AP > 0.3) for hydrops in 15/20 patients and with MRI hydrops was detected in all but one of the patients.

The alternative to dehydration is the effect of salt loading, which was supposed to produce endolymphatic hydrops symptoms. After baseline ECoChG studies, Gamble et al. (1999) administered 4 g of sodium chloride daily for 3 days to controls and Ménière’s disease patients. The control group of 13 healthy volunteers with normal baseline ECoChG and pure tone audiometry was tested under similar conditions. Gamble et al. (1999) performed ET ECoChG using alternating polarity clicks presented at a rate of 9.7/s at 95 dB nHL. A SP/AP ratio of 0.37 was considered the upper limit of normal. One or both ears in 38% of the patients in the study group with normal baseline SP/AP ratios and symptoms of inner ear fluid imbalance converted to abnormal. The mean SP/AP ratio of the control group for the conditions before and after salt-load was not statistically different ($p = 0.48$), whereas the difference in the mean SP/AP ratio in the study group after salt loading was statistically significant.

An animal experiment on the effects of endolymphatic hydrops, which is assumed to displace the basilar membrane towards the scala tympani and thereby increase the SP was carried out by Klis and Smoorenburg (1994). They used perfusion of the perilymphatic space with a hypotonic solution, which increased the SP and decreased the CAP amplitude, and corroborated the idea that static displacements of the basilar membrane indeed may underlie the enlarged SP and in particular the enlarged SP/AP ratio.

Vestibular Schwannoma

One of the first studies using ECoChG in the diagnosis of vestibular Schwannoma was by Morrison et al. (1976) who evaluated the findings in 56 surgically confirmed ears. They proposed that there are at least three separate criteria to be considered in reaching or strongly suspecting a diagnosis of such pathology. These are broadening of the CAP waveform (loss of the positive peak separating the N_1 and N_2), observation of a clear CM response, and presence of the CAP even when using stimulus intensities which are not audible in the patients’ affected ears. Beagley et al. (1977) explored in an animal study why the normally biphasic CAP changed into a monophasic one and attributed it to a neural block. This fits well with the monophasic NAPs often obtained in these tumors (see **Figure 11**).

In a large study Eggermont et al. (1980) compared the use of ECoChG and auditory brainstem response (ABR) in the diagnosis of surgically confirmed vestibular Schwannoma in 45 patients. ECoChG results provided evidence that, for hearing losses up to at least 60 dB HL, the origin is cochlear (**Figure 12**). We concluded that ECoChG as the sole test for detection of vestibular Schwannoma appeared to be of limited diagnostic value. In combination with ABR, ECoChG generally provided a clear N_1 in cases where ABR wave I could not be detected, and so raised its diagnostic value.

CAP phenomenology in vestibular Schwannoma ears is distinctly different from normal ears and often also from ears with sensorineural hearing loss (**Figure 13A**). In 30% of the studied vestibular Schwannoma cases, Eggermont et al. (1980) found that the N_1 latencies were longer than those of Ménière’s disease. Whereas long CAP duration is found with use of tone burst stimulation, especially for 2 kHz, it does not occur in the NAP derivation (**Figure 13B**). Most cases with abnormally long

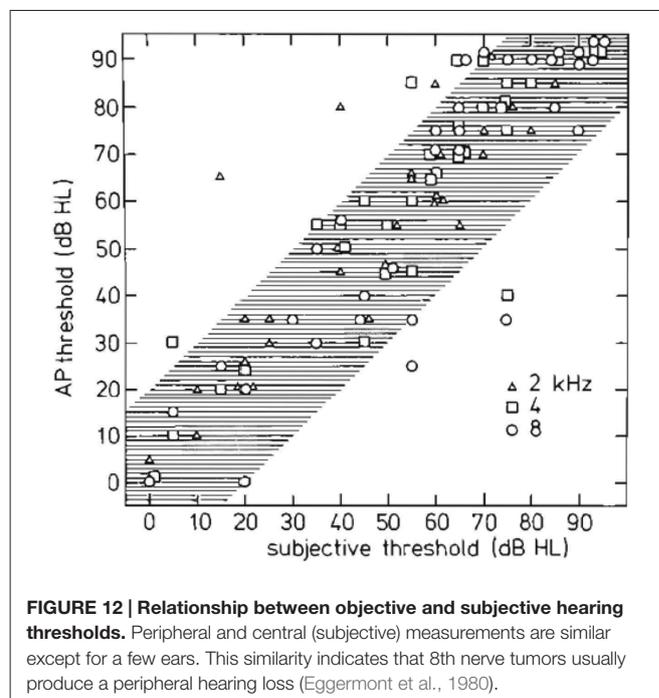


FIGURE 12 | Relationship between objective and subjective hearing thresholds. Peripheral and central (subjective) measurements are similar except for a few ears. This similarity indicates that 8th nerve tumors usually produce a peripheral hearing loss (Eggermont et al., 1980).

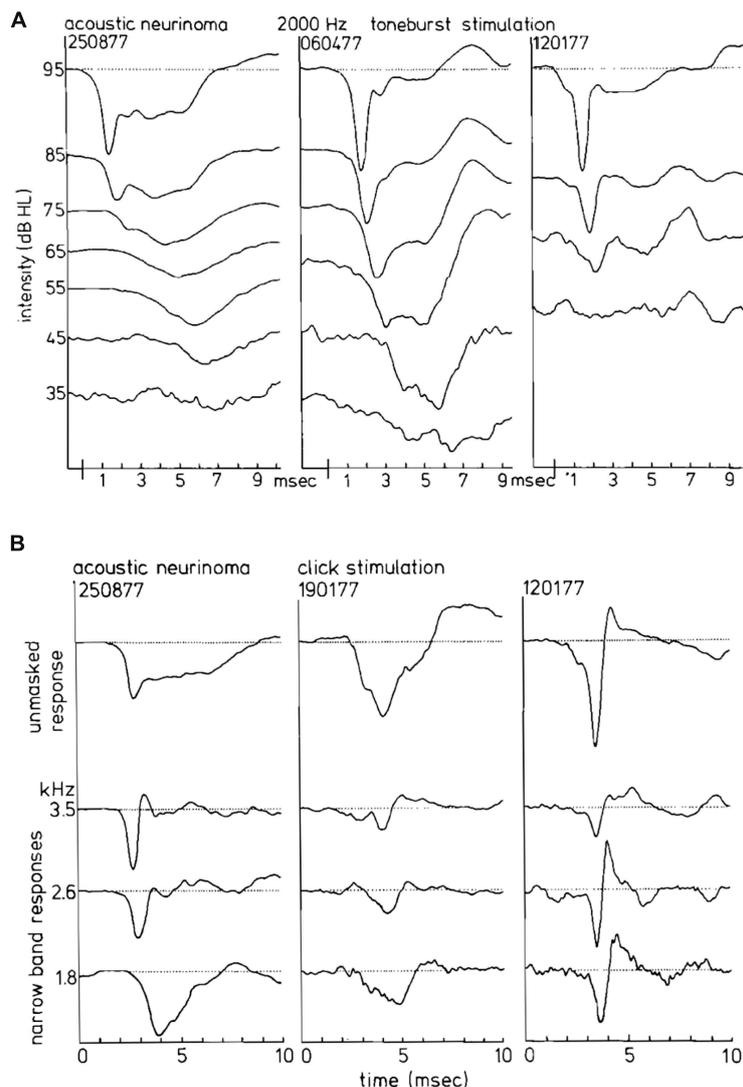


FIGURE 13 | (A) CAP waveforms in response to 2 kHz tone burst stimulation in three ears with acoustic neuroma. Depending on the individual case as well as on stimulus intensity, broad characteristic waveforms or nearly normal CAPs can be found. It appears that the CAP waveform is not consistently abnormal in acoustic neuroma ears. **(B)** Narrow band AP waveforms in acoustic neuroma ears. From dominantly monophasic NAPs in the left series to strictly biphasic narrow band responses in the right series, reminding us of a sensorineural hearing loss, the relationship to the CAP waveform is clear. From Eggermont et al. (1980).

N_1 latencies also had monophasic narrow band contributions. In this situation, the usual canceling of positive and negative deflections leading to sharply peaked CAPs is lacking. The result is broad CAPs and abnormally long CAP latencies in the middle intensity range.

Correspondingly, the width of the CAP, resulting from the monophasic NAP contributions can be distinctly larger than in normal ears (**Figure 14A**), whereas the amplitude of the SP^- is clearly lower than in Ménière's disease (**Figure 14B**). Thus the abnormally broad CAPs, especially those with short latencies (**Figure 13A**), are due to this NAP effect and not to a pronounced SP^- , as in Ménière's disease.

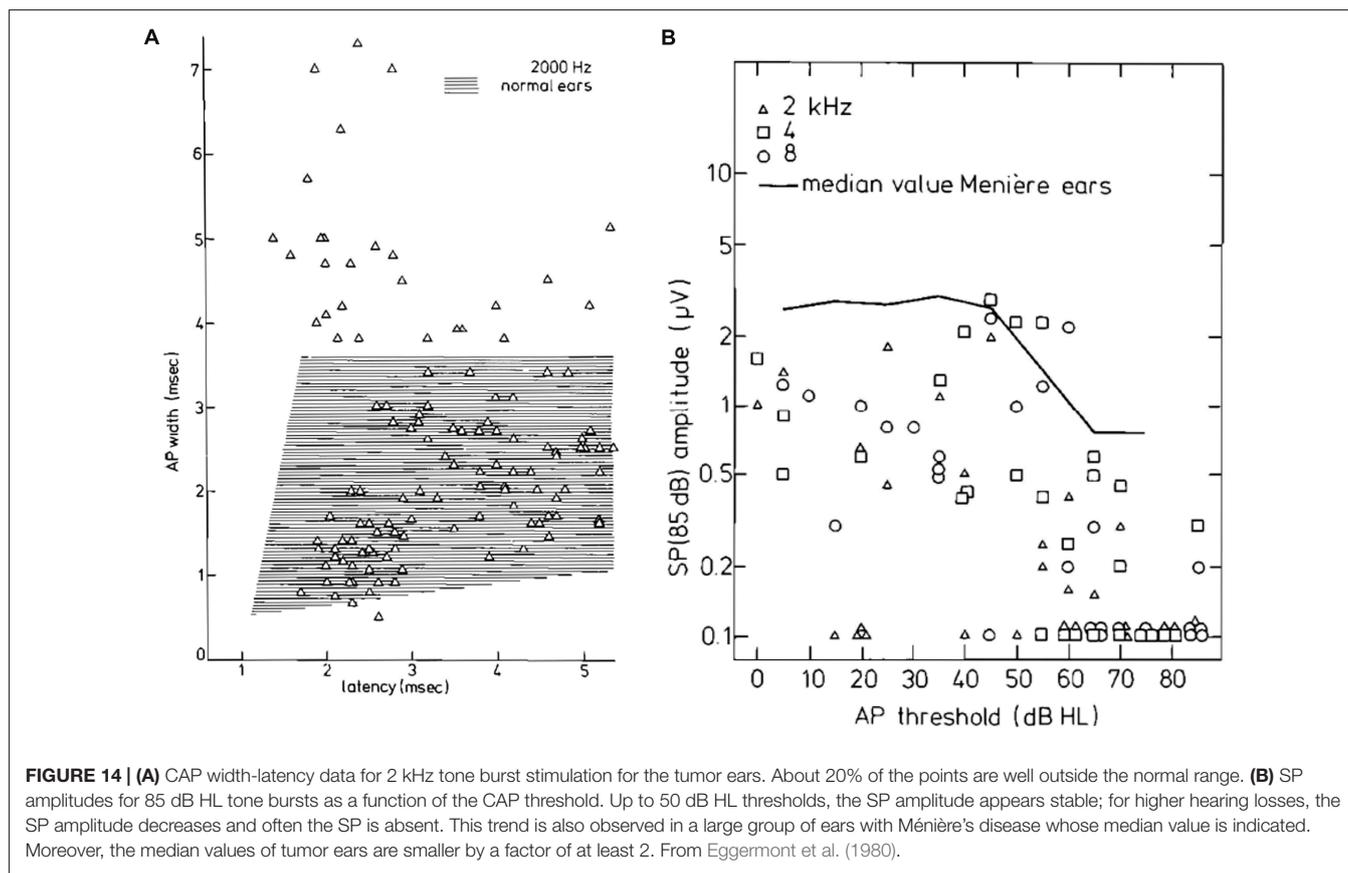
Finally, Eggermont et al. (1980) found that the dominant effect of vestibular Schwannomas, causing a hearing loss, is

on the cochlea probably resulting from interference with the blood supply. Because most ECoChG parameters indicate a pure cochlear hearing loss without neural involvement, assessing the state of hearing at the peripheral site of the internal auditory meatus therefore has limited value in the differential diagnosis. An exception is when the CAP thresholds are much lower than the behavioral ones. This was later independently confirmed by Prasher and Gibson (1983).

CURRENT INTEREST

Cochlear Implants

Telemetry capabilities became commercially available in 1998 (e.g., Shallop et al., 1999) for the measurement of the electrically



evoked CAP (eCAP) from the auditory nerve in cochlear implant recipients. The eCAP is recorded via the intracochlear electrodes of the implant. Because the eCAP is a short-latency evoked potential, it overlaps with the stimulus artifact. All newer CI systems are equipped with two-way telemetry capabilities and artifact rejection that allow for measurement of electrode impedance and the eCAP. The eCAP is recorded as a negative peak (N_1) at about 0.2–0.4 ms following stimulus onset, followed by a much smaller positive peak (P_1) or plateau occurring at about 0.6–0.8 ms. The amplitude of the eCAP can be as large as 1 mV, which is much larger in magnitude than the CAP (up to 30 μV) recorded by TT ECoChG in normal ears (Eggermont et al., 1974).

The ability to record high quality eCAP data was early on shown by Frijns et al. (2002). Their recordings showed clear N_1 and P_1 peaks with amplitude up to 400 μV , under the condition that there was at least one contact space between the stimulating and recording electrodes. They also found that responses were larger and tended to peak at recording sites around apical and basal stimulating electrodes. This suggested a limited spread of excitation. Campbell et al. (2015) recorded from CI patients who retained audiometric thresholds between 75 and 90 dB HL at 500 Hz in their implanted ear. In response to acoustical stimulation they obtained eCAPs including CM and SP responses. The eCAP thresholds were similar to the audiometric thresholds. Dalbert

et al. (2015b) used eCAPs to follow the post-surgery changes in hearing in CI patients, which were largely due to middle ear effusion, resulting from the surgery and disappeared over time.

From their modeling studies, Briare and Frijns (2005) noted that the calculated eCAPs based on the theoretical unit response did not match the measured human eCAP obtained using neural response telemetry (Frijns et al., 2002). Briare and Frijns (2005) found the potential solution to the discrepancy from a study by Miller et al. (2004) that indicated that two APs are present, and that the initial positive peak, when present, in the eCAP originates from antidromic APs originating from a relatively central site on the nerve fiber, likely close the ganglion cell body, of AP initiation. Thus, the dendrite may be responsible for the generation of the P_0 peak. Note that in acoustic stimulation the site of initial spike excitation is likely the proximal dendrite (Hossain et al., 2005).

The study by Miller et al. (2004) indicated that the state of neural degeneration of the fibers has a big influence on the presence of the P_0 peak in the unit response, as also implied by Rattay et al. (2001). Briare and Frijns (2006) used this to show that a large P_0 peak in the eCAP occurs before the N_1P_1 complex when the fibers are not degenerated. They suggested that the absence of this peak might be used as an indicator for degeneration of the proximal dendrite.

Westen et al. (2011) evaluated the use of the unit response as a unitary response in a convolution integral to predict the eCAP and found evidence for changes in the unit response with stimulus level. This suggested that the unit responses for different electrodes are not independent, likely caused by strong synchronization across fibers at high stimulus levels. Therefore the eCAP cannot be predicted from the unit responses, and consequently, the inverse problem assessing the patency of the ANFs on basis of the eCAP is not unambiguous.

Recently, Strahl et al. (2016) used a deconvolution model to estimate the nerve firing probability based on a biphasic unit response and the eCAP, both in guinea pigs and human implantees. They found that the estimated nerve firing probability was bimodal and could be parameterized by two Gaussian distributions with an average latency difference of 0.4 ms. The ratio of the scaling factors of the late and early component increased with neural degeneration in the guinea pig. The two-component firing probability was attributed to either latency differences in the population of nerve fibers resulting from late firing due to excitation of the proximal dendrite, compared to direct, central to the cell body, activation of the ANFs. They suggested that the deconvolution of the eCAP could be used to reveal these two separate firing components in the auditory nerve, which may elucidate degeneration of the proximal dendrite.

Intraoperative recording from the round window or from the promontory during cochlear implant surgery has also been reported about in a recent series of articles (Mandalà et al., 2012; Calloway et al., 2014; McClellan et al., 2014; Dalbert et al., 2015a; Formeister et al., 2015; Adunka et al., 2016). I will not dwell on this ECochG use as it will be part of another set of articles in this Special Topic.

Auditory Neuropathy

The diagnosis of “auditory neuropathy” usually does not require more than the presence of a superficial phenomenology consisting of recordable OAEs and absent or very poorly defined ABRs. Patients with auditory neuropathy also may have mild to moderate hearing loss and more severe speech perception deficits than expected based on the audiogram. However, there is quite a bit more differentiation with respect to underlying genetic and peripheral hearing mechanisms. This has led among others to use of a new term “synaptopathy”, which puts one of the mechanism in the IHC ribbon synapses (Khimich et al., 2005; Kujawa and Liberman, 2009; Moser et al., 2013). It should be noted that acquired synaptopathy (Kujawa and Liberman, 2009) is completely different from that resulting from the *OTOF* mutation. Acquired synaptopathy resulting from a TTS following noise exposure, shows normal otoacoustic emission, normal ABR thresholds and waveforms and putatively a reduction in wave I amplitude at high stimulus levels. It is obvious that in such cases the CM, SP and CAP will all be normal, with a putative reduction in CAP amplitude at high stimulus levels, although this has been disputed (Bourien et al., 2014).

Another umbrella term is “dys-synchrony”, which can describe anything from the non-synchronous transmitter

substance release from the ribbon synapses, resulting in onset desynchrony in the ANF firings, to changes in the peripheral dendrite of the spiral ganglion slowing down of APs along the ANFs (Rance and Starr, 2015) which also results in a large spread of spike latencies and hence poorly shaped ABRs. Ears affected by auditory neuropathy show a large CM riding on a large positive potential, presumably and SP⁺ (Gibson and Sanli, 2007).

Harrison (1998) found that scattered IHC loss, resulting from carboplatin administration in chinchillas resulted in normal oto-acoustic emissions, and CM whereas ABR thresholds were significantly elevated. He suggested that this type of damage could also result from longterm cochlear hypoxia and be a likely candidate for certain types of auditory neuropathy in humans.

Genes underlying two common forms of auditory neuropathy are *OTOF* resulting in synaptopathy and *OPA1* resulting in neuropathy of the spiral ganglion dendrites. Because IHC exocytosis was almost completely abolished in an *otoferlin* knock-out mouse model, *otoferlin* should have a role in a late step of exocytosis from the ribbon synapses. *Otoferlin* appears to mediate the replenishment of the ready releasable vesicle pool, and plays a role in the vesicle recruitment to the active zone membrane (Wichmann, 2015).

Huang et al. (2012) recorded the cochlear potentials CM, SP and CAPs by ECochG before cochlear implantation in patients diagnosed with familial optic atrophy which suggested an auditory neuropathy. Genetic analysis identified a R445H mutation in the *OPA1* gene. Audiological studies showed preserved DPOAEs and absent or abnormally delayed ABRs. TT ECochG showed prolonged low amplitude negative potentials without auditory nerve CAPs. After cochlear implantation, hearing thresholds, speech perception and synchronous activity in auditory brainstem pathways were restored. This suggests that deafness accompanying this *OPA1* mutation is due to altered function of the dendritic portions of the spiral ganglion.

Santarelli et al. (2009) recorded abnormal click-evoked cochlear potentials with TT ECochG from four children with *OTOF* mutations to evaluate the physiological effects resulting from abnormal neurotransmitter release by IHCs. The children were profoundly deaf with absent ABRs and preserved otoacoustic emissions consistent with auditory neuropathy. Cochlear potentials evoked by clicks from 60 dB p.e. SPL to 120 dB p.e. SPL were compared to recordings obtained from 16 normally hearing children. The CM showed normal amplitudes from all but one ear, consistent with the preserved DPOAEs. After canceling the CM, the remaining cochlear potentials were of negative polarity with reduced amplitude and prolonged duration compared to controls. These cochlear potentials were recorded as low as 50–90 dB below behavioral thresholds in contrast to the close correlation in normal hearing controls between cochlear potentials and behavioral threshold (see **Figure 4**). SPs were identified in five out of eight ears with normal latency whereas CAPs were either absent or of low amplitude. Stimulation at high rates reduced amplitude and duration

of the prolonged potentials, consistent with their neural generation site and not comprising SP^- s. The remaining low-amplitude prolonged negative potentials are consistent with sustained exocytosis and decreased phasic neurotransmitter release (Khimich et al., 2005) resulting in abnormal dendritic activation and impairment of auditory nerve firing. This study suggests that mechano-electrical transduction and cochlear amplification are normal in patients with *OTOF* mutations.

Santarelli et al. (2013) then compared acoustically- and electrically-evoked potentials of the auditory nerve in patients with postsynaptic or presynaptic auditory neuropathy with underlying mutations in the *OPAI* or *OTOF* gene, respectively. Among non-isolated auditory neuropathy disorders, mutations in the *OPAI* gene are believed to cause disruption of auditory nerve discharge by affecting the unmyelinated portions of human ANFs. TT ECoChG was used to record click-evoked responses from two adult patients carrying the R445H *OPAI* mutation, and from five children with mutations in the *OTOF* gene. The CM amplitude was normal in all subjects. Prolonged negative responses were recorded as low as 50–90 dB below behavioral threshold in subjects with *OTOF* mutations (Figure 15A) whereas in the *OPAI* disorder the prolonged potentials were correlated with hearing threshold (Figure 15B). A CAP was superimposed on the prolonged activity at high stimulation intensity in two children with mutations in the *OTOF* gene while CAPs were absent in the *OPAI* disorder. Electrically-evoked eCAPs (see “Cochlear Implants” Section) could be recorded from subjects with *OTOF* mutations but not from

OPAI mutations following cochlear implantation (Santarelli et al., 2015).

Santarelli et al. (2015) further characterized the hearing dysfunction in *OPAI*-linked disorders. Nine of 11 patients carrying *OPAI* mutations inducing haplo-insufficiency had normal hearing function. Eight patients carrying *OPAI* missense variants underwent cochlear implantation. The use of cochlear implant improved speech perception in all but one patient. ABRs were recorded in response to electrical stimulation in five of six subjects, whereas no eCAP was evoked from the auditory nerve through the cochlear implant. These findings corroborate that the impaired mechanism in patients carrying *OPAI* missense variants is desynchronized ANF firings resulting from neural degeneration affecting the terminal dendrites (Santarelli et al., 2015).

SUMMARY

Ruben (1967)’s three important topics in ECoChG were: (1) The correlation of physiological and psychoacoustic properties. (2) The investigation of certain diseases. (3) The objective diagnosis of individual cases of deafness. After 50 years we can make up the balance of the outcome of these three points.

Point one includes objective audiometry, which is quite accurate but is largely superseded by the non-invasive ABR. ECoChG may remain the method of choice when objective hearing test have to be done under anesthesia. One may also say that intra-operative monitoring falls in this category. This likely becomes an important topic in relation to cochlear implantation. Several important differences between human and animal electrophysiology were found in some temporal response properties, such as adaptation and forward masking. Here the human data showed much larger time constants than those in common experimental animals. However, in these cases the human psychoacoustic data did not show any difference from the animal electrophysiological data. This requires further investigation. In addition the purported relation between oto-acoustic emission and CM needs more detailed study. Correlating the recorded eCAPs with a CI with applicable psychoacoustics needs to be further explored.

Point two, the investigation of certain diseases has been largely focused on Ménière’s disease, and has shown that for hearing losses up to 50 dB the OHC are not affected—normal SP and CM—and do not cause the fluctuating hearing loss. More promise hold the recent investigations of various genetic forms of auditory neuropathy, where ECoChG powerfully illustrates the effects of the pre- and post-synaptic mechanisms on the temporal aspects of auditory nerve activity.

Point three, the objective diagnosis of individual cause of deafness, has focused primarily on vestibular schwannoma and Ménière’s disease, which show comparable broad and long lasting SP–CAP waveforms. ECoChG highlighted the different underlying causes as relatively—compared to the CAP—large

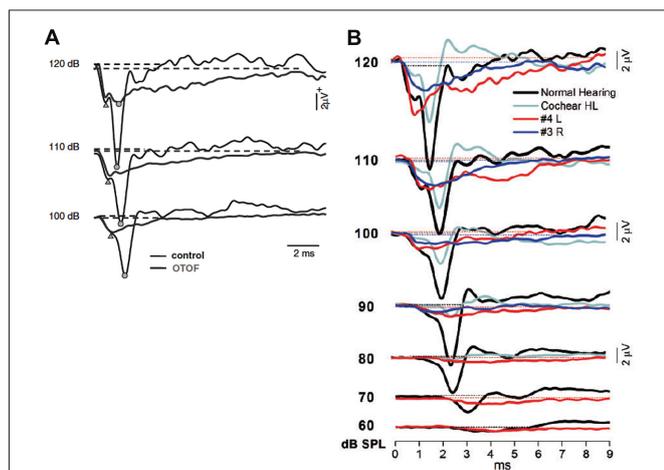


FIGURE 15 | (A) Comparison between the SP–CAP potentials recorded from one subject with *OTOF* mutations and one control. The curves for the *OTOF* subject are superimposed on the recordings obtained from one control at intensities up to 120 dB p.e. SPL to highlight the similarities of the SP component between controls and patients with *OTOF* mutations. Open circles and triangles refer to the CAP and SP peaks, respectively. From Santarelli et al. (2009). **(B)** ECoChG waveforms obtained after CM cancellation from two representative *OPAI* patients are superimposed on the corresponding responses recorded from one normal hearing control and from one hearing-impaired child with cochlear hearing loss (Cochlear HL) at decreasing stimulus intensity. From Santarelli et al. (2015).

SP (Ménière's disease) and monophasic unit contributions (vestibular schwannomas), respectively. However, the specificity and sensitivity of ECochG in these disorders has so far precluded reliable diagnosis in individual cases.

Point four, given the ambiguities of distinguishing SP⁻ from a desynchronized CAP in auditory neuropathy, and the interpretation of CM as a purely presynaptic potential, it is obvious that further basic research is needed into the limits of applicability of these traditionally considered "isolatable responses" in ECochG.

REFERENCES

- Adunka, O. F., Giardina, C. K., Formeister, E. J., Choudhury, B., Buchman, C. A., and Fitzpatrick, D. C. (2016). Round window electrocochleography before and after cochlear implant electrode insertion. *Laryngoscope* 126, 1193–1200. doi: 10.1002/lary.25602
- Al-momani, M. O., Ferraro, J. A., Gajewski, B. J., and Ator, G. (2009). Improved sensitivity of electrocochleography in the diagnosis of Ménière's disease. *Int. J. Audiol.* 48, 811–819. doi: 10.3109/14992020903019338
- Aran, J. M. (1971). The electrocochleogram. Recent results in children and in some pathological cases. *Arch. Klin. Exp. Ohr.-, Nas.- U. Kehlk. Heilk.* 198, 128–141. doi: 10.1007/BF00419529
- Aran, J. M., and Le Bert, G. (1968). Les reponses nerveuses cochleaires chez l'homme: image du fonctionnement de l'oreille et nouveau test d'audiometrie objective. *Rev. Laryngol. Otol. Rhinol.* 89, 361–378.
- Arslan, E., Turrini, M., Lupi, G., Genovese, E., and Orzan, E. (1997). Hearing threshold assessment with auditory brainstem response (ABR) and Electrocochleography (ECochG) in uncooperative children. *Scand. Audiol. Suppl.* 46, 32–37.
- Asai, H., and Mori, N. (1989). Change in the summing potential and action potential during the fluctuation of hearing in Ménière's disease. *Scand. Audiol.* 18, 13–17. doi: 10.3109/01050398909070716
- Aso, S., and Watanabe, Y. (1994). Electrocochleography in the diagnosis of delayed endolymphatic hydrops. *Acta Otolaryngol. Suppl.* 511, 87–90. doi: 10.3109/00016489409128307
- Baba, A., Takasaki, K., Tanaka, F., Tsukasaki, N., Kumagami, H., and Takahashi, H. (2009). Amplitude and area ratios of summing potential/action potential (SP/AP) in Ménière's disease. *Acta Otolaryngol.* 129, 25–29. doi: 10.1080/00016480701724888
- Beagley, H. A., Legoux, J. P., Teas, D. C., and Remond, M. C. (1977). Electrocochleographic changes in acoustic neuroma: some experimental findings. *Clin. Otolaryngol. Allied Sci.* 2, 213–219. doi: 10.1111/j.1365-2273.1977.tb01360.x
- von Békésy, G. (1963). Hearing theories and complex sounds. *J. Acoust. Soc. Am.* 35, 588–601. doi: 10.1121/1.1918543
- de Boer, E. (1969). Encoding of frequency information in the discharge pattern of auditory nerve fibres. *Int. Audiol.* 8, 547–556. doi: 10.3109/05384916909070224
- de Boer, E. (1975). Synthetic whole-nerve action potentials for the cat. *J. Acoust. Soc. Am.* 58, 1030–1045. doi: 10.1121/1.380762
- Bordley, J. E., Ruben, R. J., and Lieberman, A. T. (1964). Human cochlear potentials. *Laryngoscope* 74, 463–479. doi: 10.1288/00005537-196404000-00001
- Bourien, J., Tang, Y., Batrel, C., Huet, A., Lenoir, M., Ladrech, S., et al. (2014). Contribution of auditory nerve fibers to compound action potential of the auditory nerve. *J. Neurophysiol.* 112, 1025–1039. doi: 10.1152/jn.00738.2013
- Briaire, J. J., and Frijns, J. H. M. (2005). Unraveling the electrically evoked compound action potential. *Hear. Res.* 205, 143–156. doi: 10.1016/j.heares.2005.03.020
- Briaire, J. J., and Frijns, J. H. M. (2006). The consequences of neural degeneration regarding optimal cochlear implant position in scala tympani: a model approach. *Hear. Res.* 214, 17–27. doi: 10.1016/j.heares.2006.01.015

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and approved it for publication.

ACKNOWLEDGMENTS

This study was supported by the Natural Sciences and Engineering Research Council of Canada (NSERC; award number: 1206-2010 RGPIN).

- Calloway, N. H., Fitzpatrick, D. C., Campbell, A. P., Iseli, C., Pulver, S., Buchman, C. A., et al. (2014). Intracochlear electrocochleography during cochlear implantation. *Otol. Neurotol.* 35, 1451–1457. doi: 10.1097/MAO.0000000000000451
- Campbell, L., Kaicer, A., Briggs, R., and O'Leary, S. (2015). Cochlear response telemetry: intracochlear electrocochleography via cochlear implant neural response telemetry pilot study results. *Otol. Neurotol.* 36, 399–405. doi: 10.1097/MAO.0000000000000678
- Chertoff, M. E., Earl, B. R., Diaz, F. J., and Sorensen, J. L. (2012). Analysis of the cochlear microphonic to a low-frequency tone embedded in filtered noise. *J. Acoust. Soc. Am.* 132, 3351–3362. doi: 10.1121/1.4757746
- Chertoff, M. E., Earl, B. R., Diaz, F. J., Sorensen, J. L., Thomas, M. L., Kameron, A. M., et al. (2014). Predicting the location of missing outer hair cells using the electrical signal recorded at the round window. *J. Acoust. Soc. Am.* 136, 1212–1224. doi: 10.1121/1.4890641
- Chertoff, M., Lichtenhan, J., and Willis, M. (2010). Click- and chirp-evoked human compound action potentials. *J. Acoust. Soc. Am.* 127, 2992–2996. doi: 10.1121/1.3372756
- Coats, A. C. (1981). The summing potential and Meniere's disease. I. Summing potential amplitude in meniere and non-Meniere ears. *Arch. Otolaryngol.* 107, 199–208. doi: 10.1001/archotol.1981.007904000101001
- Coats, A. C., and Alford, B. R. (1981). Ménière's disease and the summing potential. III. Effect of glycerol administration. *Arch. Otolaryngol.* 107, 469–473.
- Coats, A. C., and Dickey, J. R. (1972). Postmasking recovery of human click action potentials and click loudness. *J. Acoust. Soc. Am.* 52, 1607–1612. doi: 10.1121/1.1913293
- Dalbert, A., Pffnner, F., Rüüsl, C., Thoele, K., Sim, J. H., Gerig, R., et al. (2015a). Extra- and intracochlear electrocochleography in cochlear implant recipients. *Audiol. Neurootol.* 20, 339–348. doi: 10.1159/000438742
- Dalbert, A., Sim, J. H., Gerig, R., Pffnner, F., Roosli, C., and Huber, A. (2015b). Correlation of electrophysiological properties and hearing preservation in cochlear implant patients. *Otol. Neurotol.* 36, 1172–1180. doi: 10.1097/MAO.0000000000000768
- Dallos, P. (1972). Cochlear potentials. A status report. *Audiology* 11, 29–41. doi: 10.3109/00206097209072579
- Dallos, P., Santos-Sacchi, J., and Flock, A. (1982). Intracellular recordings from cochlear outer hair cells. *Science* 218, 582–584. doi: 10.1126/science.7123260
- Dallos, P., and Wang, C. Y. (1974). Bioelectric correlates of kanamycin intoxication. *Audiology* 13, 277–289. doi: 10.3109/00206097409071685
- Dauman, R., Aran, J. M., Charlet de Sauvage, R., and Portmann, M. (1988). Clinical significance of the summing potential in Ménière's disease. *Am. J. Otol.* 9, 31–38.
- Dauman, R., Aran, J. M., and Portmann, M. (1986). Summing potential and water balance in Ménière's disease. *Ann. Otol. Rhinol. Laryngol.* 95, 389–395. doi: 10.1177/000348948609500413
- Devaiah, A. K., Dawson, K. L., Ferraro, J. A., and Ator, G. A. (2003). Utility of area curve ratio electrocochleography in early Ménière's disease. *Arch. Otolaryngol. Head Neck Surg.* 129, 547–551. doi: 10.1001/archotol.129.5.547
- Earl, B. R., and Chertoff, M. E. (2010). Predicting auditory nerve survival using the compound action potential. *Ear Hear.* 31, 7–21. doi: 10.1097/AUD.0b013e3181ba748c

- Eggermont, J. J. (1976a). Electrophysiological study of the normal and pathological human cochlea. II. Neural responses. *Rev. Laryngol. Otol. Rhinol.* 97, 497–506.
- Eggermont, J. J. (1976b). “Summating potentials in electrocochleography. Relation to hearing pathology,” in *Electrocochleography*, eds R. J. Ruben, C. Elberling, and G. Salomon (Baltimore: University Park Press), 67–87.
- Eggermont, J. J. (1976c). “Electrocochleography,” in *Handbook of Sensory Physiology*, (Vol. 5, part 3) eds W. D. Keidel and W. D. Neff (New York, NY: Springer-Verlag), 626–705.
- Eggermont, J. J. (1976d). Analysis of compound action potential responses to tone bursts in the human and guinea pig cochlea. *J. Acoust. Soc. Am.* 60, 1132–1139. doi: 10.1121/1.381214
- Eggermont, J. J. (1977). Electrocochleography and recruitment. *Ann. Otol. Rhinol. Laryngol.* 86, 138–149. doi: 10.1177/000348947708600202
- Eggermont, J. J. (1979a). Summating potentials in Ménière’s disease. *Arch. Otorhinolaryngol.* 222, 63–74. doi: 10.1007/bf00456340
- Eggermont, J. J. (1979b). Narrow-band AP latencies in normal and recruiting human ears. *J. Acoust. Soc. Am.* 65, 463–470. doi: 10.1121/1.382345
- Eggermont, J. J. (1979c). Compound action potentials: tuning curves and delay times. *Scand. Audiol.* 9, (Suppl), 129–139.
- Eggermont, J. J., Don, M., and Brackmann, D. E. (1980). Electrocochleography and auditory brainstem electric response in patients with pontine angle tumors. *Ann. Otol. Rhinol. Laryngol. Suppl.* 89, 1–19. doi: 10.1177/000348948008905601
- Eggermont, J. J., and Odenthal, D. W. (1974a). Electrophysiological investigation of the human cochlea: recruitment, masking and adaptation. *Audiology* 13, 1–22. doi: 10.3109/00206097409089333
- Eggermont, J. J., and Odenthal, D. W. (1974b). Frequency selective masking in electrocochleography. *Rev. Laryngol. Otol. Rhinol.* 95, 489–496.
- Eggermont, J. J., Odenthal, D. W., Schmidt, P. R., and Spoor, A. (1974). Electrocochleography Basic principles and clinical application. *Acta Otolaryngol. Suppl.* 316, 1–84.
- Eggermont, J. J., and Spoor, A. (1973). Cochlear adaptation in guinea pigs. A quantitative description. *Audiology* 12, 193–220. doi: 10.1080/00206097309089320
- Elberling, C. (1973). Transitions in cochlear action potentials recorded from the ear canal in man. *Scand. Audiol.* 2, 151–159. doi: 10.3109/01050397309044948
- Elberling, C. (1974). Action potentials along the cochlear partition recorded from the ear canal in man. *Scand. Audiol.* 3, 13–19. doi: 10.3109/01050397409044959
- Elberling, C. (1976a). “Simulation of cochlear action potentials recorded from the ear canal in man,” in *Electrocochleography*, eds R. J. Ruben, C. Elberling, and G. Salomon (Baltimore: University Park Press), 151–168.
- Elberling, C. (1976b). Modeling action potentials. *Rev. Laryngol. Otol. Rhinol.* 97, 527–537.
- Evans, E. F., and Elberling, C. (1982). Location-specific components of the gross cochlear action potential: an assessment of the validity of the high-pass masking technique by cochlear nerve fibre recording in the cat. *Audiology* 21, 204–227. doi: 10.3109/00206098209072740
- Ferraro, J. A., Arenberg, I. K., and Hassanein, R. S. (1985). Electrocochleography and symptoms of inner ear dysfunction. *Arch. Otolaryngol.* 111, 71–74. doi: 10.1001/archotol.1985.00800040035001
- Ferraro, J. A., and Tibbils, R. P. (1999). SP/AP area ratio in the diagnosis of Ménière’s disease. *Am. J. Audiol.* 8, 21–28. doi: 10.1044/1059-0889(1999/001)
- Formeister, E. J., McClellan, J. H., Merwin, W. H. III, Iseli, C. E., Calloway, N. H., Teagle, H. F., et al. (2015). Intraoperative round window electrocochleography and speech perception outcomes in pediatric cochlear implant recipients. *Ear Hear.* 36, 249–260. doi: 10.1097/AUD.0000000000000106
- Frijns, J. H., Briaire, J. J., de Laat, J. A., and Grote, J. J. (2002). Initial evaluation of the clarion cii cochlear implant: speech perception and neural response imaging. *Ear Hear.* 23, 184–197. doi: 10.1097/00003446-200206000-00003
- Fromm, B., Nysten, C. O., and Zotterman, Y. (1935). Studies in the mechanism of the Wever and Bray effect. *Acta Otolaryngol. (Stockh)* 22, 477–486. doi: 10.3109/00016483509118125
- Fukuoka, H., Takumi, Y., Tsukada, K., Miyagawa, M., Oguchi, T., Ueda, H., et al. (2012). Comparison of the diagnostic value of 3 T MRI after intratympanic injection of GBCA, electrocochleography and the glycerol test in patients with Ménière’s disease. *Acta Otolaryngol.* 132, 141–145. doi: 10.3109/00016489.2011.635383
- Gamble, B. A., Meyerhoff, W. L., Shoup, A. G., and Schwade, N. D. (1999). Salt-load electrocochleography. *Am. J. Otol.* 20, 325–330.
- Gasser, H. S., and Grundfest, H. (1939). Axon diameters in relation to the spike dimensions and the conduction velocity in mammalian a fibers. *Am. J. Physiol.* 127, 393–414.
- Ge, X., and Shea, J. J. Jr. (2002). Transtympanic electrocochleography: a 10-year experience. *Otol. Neurotol.* 23, 799–805. doi: 10.1097/00129492-200209000-00032
- Gibbin, K. P., Mason, S. M., and Kent, S. E. (1983). Prolongation of the cochlear microphonic in man. Cochlear microphonic ringing. *Acta Otolaryngol.* 95, 13–18. doi: 10.3109/00016488309130910
- Gibson, W. P. (2009). A comparison of two methods of using transtympanic electrocochleography for the diagnosis of Ménière’s disease: click summating potential/action potential ratio measurements and tone burst summating potential measurements. *Acta Otolaryngol. Suppl.* 560, 38–42. doi: 10.1080/00016480902729843
- Gibson, W. P., and Morrison, A. W. (1983). Electrocochleography and the glycerol dehydration test: A case study. *Br. J. Audiol.* 17, 95–99. doi: 10.3109/03005368309078915
- Gibson, W. P., Prasher, D. K., and Kilkenny, G. P. (1983). Diagnostic significance of transtympanic electrocochleography in Ménière’s disease. *Ann. Otol. Rhinol. Laryngol.* 92, 155–159. doi: 10.1177/000348948309200212
- Gibson, W. P. R., Ramsden, R. T., and Moffat, D. A. (1977). Clinical electrocochleography in the diagnosis and management of Ménière’s disease. *Audiology* 16, 389–401. doi: 10.3109/00206097709071852
- Gibson, W. P. R., and Sanli, H. (2007). Auditory neuropathy: an update. *Ear Hear.* 28, 102S–106S. doi: 10.1097/aud.0b013e3180315392
- Goin, D. W., Staller, S. J., Asher, D. L., and Mischke, R. E. (1982). Summating potential in Ménière’s disease. *Laryngoscope* 92, 1383–1389. doi: 10.1288/00005537-198212000-00008
- Goldstein, M., and Kiang, N. Y. S. (1958). Synchrony of neural activity in electric responses evoked by transient acoustic stimuli. *J. Acoust. Soc. Am.* 30, 107–114. doi: 10.1121/1.1909497
- Greenwood, D. D. (1961). Critical bandwidth and the frequency coordinates of the basilar membrane. *J. Acoust. Soc. Am.* 33, 1344–1356. doi: 10.1121/1.1908437
- Hall, J. G. (1967). Hearing and primary auditory centres of whales. *Acta Otolaryngol.* 63, 244–250. doi: 10.3109/00016486709123587
- Harrison, R. V. (1998). An animal model of auditory neuropathy. *Ear Hear.* 19, 355–361. doi: 10.1097/00003446-199810000-00002
- Hossain, W. A., Antic, S. D., Yang, Y., Rasband, M. N., and Morest, D. K. (2005). Where is the spike generator of the cochlear nerve? Voltage-gated sodium channels in the mouse cochlea. *J. Neurosci.* 25, 6857–6868. doi: 10.1523/JNEUROSCI.0123-05.2005
- Huang, T., Santarelli, R., and Starr, A. (2012). Mutation of *OPA1* gene causes deafness by affecting function of auditory nerve terminals. *Brain Res.* 1300, 97–104. doi: 10.1016/j.brainres.2009.08.083
- Johnstone, J. R., and Johnstone, B. M. (1966). Origin of the summating potential. *J. Acoust. Soc. Am.* 40, 1405–1413. doi: 10.1121/1.1910240
- Kamerer, A. M., Diaz, F. J., Peppi, M., and Chertoff, M. E. (2016). The potential use of low-frequency tones to locate regions of outer hair cell loss. *Hear. Res.* 342, 39–47. doi: 10.1016/j.heares.2016.09.006
- Kanzaki, J., Ouchi, T., Yokobori, H., and Inoh, T. (1982). Electrocochleographic study of summating potentials in Ménière’s disease. *Audiology* 21, 409–424. doi: 10.3109/00206098209072755
- Khimich, D., Nouvian, R., Pujol, R., Tom Dieck, S., Egner, A., Gundelfinger, E. D., et al. (2005). Hair cell synaptic ribbons are essential for synchronous auditory signalling. *Nature* 434, 889–894. doi: 10.1038/nature03418
- Kiang, N. Y. S., Moxon, E. C., and Kahn, A. R. (1976). “The relationship of gross potentials recorded from the cochlea to single unit activity in the auditory

- nerve," in *Electrocochleography*, eds R. J. Ruben, C. Elberling, and G. Salomon (Baltimore: University Park Press), 95–115.
- Kiang, N. Y. S., Watanabe, T., Thomas, E. C., and Clark, L. F. (1965). *Discharge Patterns of Single Fibres in The Cats Auditory Nerve*. Cambridge, MA: MIT Press.
- Kim, H. H., Kumar, A., Battista, R. A., and Wiet, R. J. (2005). Electrocochleography in patients with Ménière's disease. *Am. J. Otolaryngol.* 26, 128–131. doi: 10.1016/j.amjoto.2004.11.005
- Klis, S. F., and Smoorenburg, G. F. (1994). Osmotically induced pressure difference in the cochlea and its effect on cochlear potentials. *Hear. Res.* 75, 114–120. doi: 10.1016/0378-5955(94)90062-0
- Klockhoff, I., and Lindblom, U. (1966). Endolymphatic hydrops revealed by glycerol test. Preliminary report. *Acta Otolaryngol.* 61, 459–462. doi: 10.3109/00016486609127084
- Koyuncu, M., Mason, S. M., and Saunders, M. W. (1994). Electrocochleography in endolymphatic hydrops using tone-pip and click stimuli. *Clin. Otolaryngol. Allied Sci.* 19, 73–78. doi: 10.1111/j.1365-2273.1994.tb01152.x
- Kujawa, S. G., and Liberman, M. C. (2009). Adding insult to injury: cochlear nerve degeneration after "temporary" noise-induced hearing loss. *J. Neurosci.* 29, 14077–14085. doi: 10.1523/JNEUROSCI.2845-09.2009
- Kumaragamage, C., Lithgow, B., and Moussavi, Z. (2015). A new low-noise signal acquisition protocol and electrode placement for electrocochleography (ECOG) recordings. *Med. Biol. Eng. Comput.* 53, 499–509. doi: 10.1007/s11517-015-1251-5
- Lempert, J., Meltzer, P. E., Wever, E. G., and Lawrence, M. (1950). The cochleogram and its clinical application. *Arch. Otolaryngol.* 51, 307–311. doi: 10.1001/archotol.1950.00700020329001
- Lempert, J., Wever, E. G., and Lawrence, M. (1947). The cochleogram and its clinical application. *Arch. Otolaryngol.* 45, 61–67. doi: 10.1001/archotol.1947.00690010068005
- Levine, S., Margolis, R. H., and Daly, K. A. (1998). Use of electrocochleography in the diagnosis of Ménière's disease. *Laryngoscope* 108, 993–1000. doi: 10.1097/00005537-199807000-00008
- Liberman, M. C., Gao, J., He, D. Z., Wu, X., Jia, S., and Zuo, J. (2002). Prestin is required for electromotility of the outer hair cell and for the cochlear amplifier. *Nature* 419, 300–304. doi: 10.1038/nature01059
- Lichtenhan, J. T., and Chertoff, M. E. (2008). Temporary hearing loss influences post-stimulus time histogram and single neuron action potential estimates from human compound action potentials. *J. Acoust. Soc. Am.* 123, 2200–2212. doi: 10.1121/1.2885748
- Lichtenhan, J. T., Cooper, N. P., and Guinan, J. J. Jr. (2013). A new auditory threshold estimation technique for low frequencies: proof of concept. *Ear Hear.* 34, 42–51. doi: 10.1097/AUD.0b013e31825f9bd3
- Lichtenhan, J. T., Hartsock, J. J., Gill, R. M., Guinan, J. J. Jr., and Salt, A. N. (2014). The auditory nerve overlapped waveform (ANOW) originates in the cochlear apex. *J. Assoc. Res. Otolaryngol.* 15, 395–411. doi: 10.1007/s10162-014-0447-y
- Liu, C., Chen, X., and Xu, L. (1992). Cochlear microphonics and recruitment. *Acta Otolaryngol.* 112, 215–220. doi: 10.1080/00016489.1992.11665407
- Mandalà, M., Colletti, L., Tonoli, G., and Colletti, V. (2012). Electrocochleography during cochlear implantation for hearing preservation. *Otolaryngol. Head Neck Surg.* 146, 774–781. doi: 10.1177/0194599811435895
- McClellan, J. H., Formeister, E. J., Merwin, W. H. III, Dillon, M. T., Calloway, N., Iseli, C., et al. (2014). Round window electrocochleography and speech perception outcomes in adult cochlear implant subjects: comparison with audiometric and biographical information. *Otol. Neurotol.* 35, e245–e252. doi: 10.1097/MAO.0000000000000557
- Miller, C. A., Abbas, P. J., Hay-McCutcheon, M. J., Robinson, B. K., Nourski, K. V., and Jeng, F. C. (2004). Intracochlear and extracochlear eCAPs suggest antidromic action potentials. *Hear. Res.* 198, 75–86. doi: 10.1016/j.heares.2004.07.005
- Moffat, D. A., Gibson, W. P., Ramsden, R. T., Morrison, A. W., and Booth, J. B. (1978). Transtympanic electrocochleography during glycerol dehydration. *Acta Otolaryngol.* 85, 158–166. doi: 10.3109/00016487809111922
- Mori, N., Asai, H., Doi, K., and Matsunaga, T. (1987a). Diagnostic value of extratympanic electrocochleography in Ménière's disease. *Audiology* 26, 103–110. doi: 10.3109/00206098709078411
- Mori, N., Asai, H., Sakagami, M., and Matsunaga, T. (1987b). Comparison of summing potential in Ménière's disease between trans- and extratympanic electrocochleography. *Audiology* 26, 348–355. doi: 10.3109/00206098709081562
- Mori, N., Asai, H., and Sakagami, M. (1988). The relationship of SP and AP findings to hearing level in Ménière's disease. *Scand. Audiol.* 17, 237–240. doi: 10.3109/01050398809070711
- Mori, N., Asai, H., and Sakagami, M. (1993). The role of summing potential in the diagnosis and management of Ménière's disease. *Acta Otolaryngol. Suppl.* 501, 51–53. doi: 10.3109/00016489309126214
- Mori, N., Saeki, K., Matsunaga, T., and Asai, H. (1982). Comparison between AP and SP parameters in trans- and extratympanic electrocochleography. *Audiology* 21, 228–241. doi: 10.3109/00206098209072741
- Morrison, A. W., Gibson, W. P., and Beagley, H. A. (1976). Transtympanic electrocochleography in the diagnosis of retrocochlear tumours. *Clin. Otolaryngol. Allied Sci.* 1, 153–167. doi: 10.1111/j.1365-2273.1976.tb00868.x
- Moser, T., Predoehl, F., and Starr, A. (2013). Review of hair cell synapse defects in sensorineural hearing impairment. *Otol. Neurotol.* 34, 995–1004. doi: 10.1097/MAO.0b013e3182814d4a
- Nguyen, L. T., Harris, J. P., and Nguyen, Q. T. (2010). Clinical utility of electrocochleography in the diagnosis and management of Ménière's disease: AOS and ANS membership survey data. *Otol. Neurotol.* 31, 455–459. doi: 10.1097/MAO.0b013e3181d2779c
- Noguchi, Y., Komatsuzaki, A., and Nishida, H. (1999). Cochlear microphonics for hearing preservation in vestibular schwannoma surgery. *Laryngoscope* 109, 1982–1987. doi: 10.1097/00005537-199912000-00016
- Odenthal, D. W., and Eggermont, J. J. (1976). "Electrocochleographic study of Ménière's disease and pontine angle neurinoma," in *Electrocochleography*, eds R. J. Ruben, C. Elberling, and G. Salomon (Baltimore: University Park Press), 331–352.
- Oh, K. H., Kim, K. W., Chang, J., Jun, H. S., Kwon, E. H., Choi, J. Y., et al. (2014). Can we use electrocochleography as a clinical tool in the diagnosis of Ménière's disease during the early symptomatic period? *Acta Otolaryngol.* 134, 771–775. doi: 10.3109/00016489.2014.907500
- Orchik, D. J., Ge, N. N., and Shea, J. J. Jr. (1998). Action potential latency shift by rarefaction and condensation clicks in Ménière's disease. *J. Am. Acad. Audiol.* 9, 121–126.
- Patuzzi, R. B., Yates, G. K., and Johnstone, B. M. (1989). Changes in cochlear microphonic and neural sensitivity produced by acoustic trauma. *Hear. Res.* 39, 189–202. doi: 10.1016/0378-5955(89)90090-7
- Perlman, M. B., and Case, T. J. (1941). Electrical phenomena of the cochlea in man. *Arch. Otolaryngol.* 34, 710–718. doi: 10.1001/archotol.1941.00660040766003
- Portmann, M., and Aran, J. M. (1971). Electro-cochleography. *Laryngoscope* 81, 899–910. doi: 10.1288/00005537-197106000-00010
- Portmann, M., Le Bert, G., and Aran, J. M. (1967). Potentiels cochléaires obtenues chez l'homme en dehors de toute intervention chirurgicale. *Rev. Laryngol. (Bordeaux)* 88, 157–164.
- Pou, A. M., Hirsch, B. E., Durrant, J. D., Gold, S. R., and Kameron, D. B. (1996). The efficacy of tympanic electrocochleography in the diagnosis of endolymphatic hydrops. *Am. J. Otol.* 17, 607–611. (Erratum in 1996)
- Prasher, D. K., and Gibson, W. P. (1983). Brainstem auditory-evoked potentials and electrocochleography: comparison of different criteria for the detection of acoustic neuroma and other cerebello-pontine angle tumours. *Br. J. Audiol.* 17, 163–174. doi: 10.3109/03005368309107881
- Prijs, V. F. (1986). Single-unit response at the round window of the guinea pig. *Hear. Res.* 21, 127–133. doi: 10.1016/0378-5955(86)90034-1
- Rance, G., and Starr, A. (2015). Pathophysiological mechanisms and functional hearing consequences of auditory neuropathy. *Brain* 138, 3141–3158. doi: 10.1093/brain/awv270
- Rasmussen, A. T. (1940). Studies on the eight cranial nerve of man. *Laryngoscope* 50, 67–83. doi: 10.1288/00005537-194001000-00008
- Rattay, F., Lutter, P., and Felix, H. (2001). A model of the electrically excited human cochlear neuron I. Contribution of neural substructures to the generation and propagation of spikes. *Hear. Res.* 153, 43–63. doi: 10.1016/S0378-5955(00)00256-2

- Roland, P. S., Yellin, M. W., Meyerhoff, W. L., and Frank, T. (1995). Simultaneous comparison between transtympanic and extratympanic electrocochleography. *Am. J. Otol.* 16, 444–450.
- Ronis, B. J. (1966). Cochlear potentials in otosclerosis. *Laryngoscope* 76, 212–231. doi: 10.1288/00005537-196602000-00003
- Ruben, R. J. (1967). “Cochlear potentials as a diagnostic test in deafness,” in *Sensorineural Hearing Processes and Disorders*, (Boston: Little, Brown and Cie), 313–338.
- Ruben, R. J., Bordley, J. E., and Lieberman, A. T. (1961). Cochlear potentials in man. *Laryngoscope* 71, 1141–1164. doi: 10.1288/00005537-196110000-00001
- Ruben, R. J., Bordley, J. E., Nager, G. T., Sekula, J., Knickerbocker, G. G., and Fisch, U. (1960). Human cochlear responses to sound stimuli. *Ann. Otol. Rhinol. Laryngol.* 69, 459–479. doi: 10.1177/000348946006900214
- Ruben, R. J., Elberling, C., and Salomon, G. (Eds.). (1976). *Electrocochleography*. Baltimore: University Park Press.
- Ruben, R. J., Knickerbocker, G. G., Sekula, J., Nager, G. T., and Bordley, J. E. (1959). Cochlear microphonics in man. *Laryngoscope* 69, 665–671. doi: 10.1288/00005537-195906000-00004
- Ruben, R. J., Lieberman, A. T., and Bordley, J. E. (1962). Some observations on cochlear potentials and nerve action potentials in children. *Laryngoscope* 72, 545–554. doi: 10.1288/00005537-196205000-00001
- Ruben, R. J., and Walker, A. E. (1963). The eighth nerve action potential in Ménière’s disease. *Laryngoscope* 73, 1456–1464. doi: 10.1288/00005537-196311000-00002
- Russell, I. J. (2008). “Chapter 3.20-Cochlear receptor potentials,” in *The Senses: A Comprehensive Reference*, ed. P. Dallos (Amsterdam: Elsevier), 320–358.
- Russell, I. J., and Sellick, P. M. (1978). Intracellular studies of hair cells in the mammalian cochlea. *J. Physiol.* 284, 261–290. doi: 10.1113/jphysiol.1978.sp012540
- Rutherford, M. A., and Roberts, W. M. (2008). “Chapter 3.22-Afferent synaptic mechanisms,” in *The Senses. A Comprehensive Reference*, ed. P. Dallos (Amsterdam: Elsevier), 366–395.
- Santarelli, R., Del Castillo, I., Rodríguez-Ballesteros, M., Scimemi, P., Cama, E., Arslan, E., et al. (2009). Abnormal cochlear potentials from deaf patients with mutations in the otoferlin gene. *J. Assoc. Res. Otolaryngol.* 10, 545–556. doi: 10.1007/s10162-009-0181-z
- Santarelli, R., del Castillo, I., and Starr, A. (2013). Auditory neuropathies and electrocochleography. *Hearing Balance Commun.* 11, 130–137. doi: 10.3109/21695717.2013.815446
- Santarelli, R., Rossi, R., Scimemi, P., Cama, E., Valentino, M. L., La Morgia, C., et al. (2015). OPA1-related auditory neuropathy: site of lesion and outcome of cochlear implantation. *Brain* 138, 563–576. doi: 10.1093/brain/awu378
- Santarelli, R., Scimemi, P., Dal Monte, E., and Arslan, E. (2006). Cochlear microphonic potential recorded by transtympanic electrocochleography in normally-hearing and hearing-impaired ears. *Acta Otorhinolaryngol. Ital.* 26, 78–95.
- Sass, K. (1998). Sensitivity and specificity of transtympanic electrocochleography in Ménière’s disease. *Acta Otolaryngol.* 118, 150–156. doi: 10.1080/00016489850154838
- Sass, K., Densert, B., and Arlinger, S. (1998). Recording techniques for transtympanic electrocochleography in clinical practice. *Acta Otolaryngol.* 118, 17–25. doi: 10.1080/00016489850155071
- Schmidt, P. H., Eggermont, J. J., and Odenthal, D. W. (1974). Study of Ménière’s disease by electrocochleography. *Acta Otolaryngol. Suppl.* 77, 75–84. doi: 10.1080/16512251.1974.11675748
- Schmidt, P. H., Odenthal, D. W., Eggermont, J. J., and Spoor, A. (1975). Electrocochleographic study of a case of lermoyez’s syndrome. *Acta Otolaryngol.* 79, 287–291. doi: 10.3109/00016487509124687
- Schoonhoven, R., Fabius, M. A., and Grote, J. J. (1995). Input/output curves to tone bursts and clicks in extratympanic and transtympanic electrocochleography. *Ear Hear.* 16, 619–630. doi: 10.1097/00003446-199512000-00008
- Schoonhoven, R., Lamoré, P. J., de Laat, J. A., and Grote, J. J. (1999). The prognostic value of electrocochleography in severely hearing-impaired infants. *Audiology* 38, 141–154. doi: 10.3109/00206099909073016
- Schoonhoven, R., Pijls, V. F., and Grote, J. J. (1996). Response thresholds in electrocochleography and their relation to the pure tone audiogram. *Ear Hear.* 17, 266–275. doi: 10.1097/00003446-199606000-00009
- Shalloo, J. K., Facer, G. W., and Peterson, A. (1999). Neural response telemetry with the nucleus CI24M cochlear implant. *Laryngoscope* 109, 1755–1759. doi: 10.1097/00005537-199911000-00006
- Spoor, A., and Eggermont, J. J. (1976). “Electrocochleography as a method for objective audiogram determination,” in *Davis and Hearing: Essays Honouring Hallowell Davis*, eds S. K. Hirsh, D. H. Eldredge, I. J. Hirsh, and S. R. Silverman (St. Louis, MO: Washington University Press), 411–418.
- Strahl, S. B., Ramekers, D., Nagelkerke, M. M. B., Schwarz, K. E., Spitzer, P., Klis, S. F. L., et al. (2016). “Assessing the firing properties of the electrically stimulated auditory nerve using a convolution model,” in *Physiology, Psychoacoustics and Cognition in Normal and Impaired Hearing. Advances in Experimental Medicine and Biology*, (Vol. 894) eds P. van Dijk, D. Baskent, E. Gaudrain, E. de Kleine, A. Wagner, and C. Lanting (Berlin: Springer), 143–153.
- Takeda, T., and Kakigi, A. (2010). The clinical value of extratympanic electrocochleography in the diagnosis of Ménière’s disease. *ORL J. Otorhinolaryngol. Relat. Spec.* 72, 196–204. doi: 10.1159/000315552
- Tasaki, I., Davis, H., and Eldredge, D. H. (1954). Exploration of cochlear potentials in guinea pig with a microelectrode. *J. Acoust. Soc. Am.* 26, 765–773. doi: 10.1121/1.1907415
- Teas, D. C., Eldredge, D. H., and Davis, H. (1962). Cochlear responses to acoustic transients: an interpretation of whole nerve action potentials. *J. Acoust. Soc. Am.* 34, 1438–1459. doi: 10.1121/1.1918366
- Versnel, H., Pijls, V. F., and Schoonhoven, R. (1992). Round-window recorded potential of single-fibre discharge (unit response) in normal and noise-damaged cochleas. *Hear. Res.* 59, 157–170. doi: 10.1016/0378-5955(92)90112-z
- Westen, A. A., Dekker, D. M., Briare, J. J., and Frijns, J. H. (2011). Stimulus level effects on neural excitation and eCAP amplitude. *Hear. Res.* 280, 166–176. doi: 10.1016/j.heares.2011.05.014
- Wever, E. G., and Bray, C. W. (1930). Auditory nerve impulses. *Science* 71:215. doi: 10.1126/science.71.1834.215
- Wichmann, C. (2015). Molecularly and structurally distinct synapses mediate reliable encoding and processing of auditory information. *Hear. Res.* 330, 178–190. doi: 10.1016/j.heares.2015.07.008
- Withnell, R. H. (2001). Brief report: the cochlear microphonic as an indication of outer hair cell function. *Ear Hear.* 22, 75–77. doi: 10.1097/00003446-200102000-00008
- Wuyts, F. L., Van de Heyning, P. H., Van Spaendonck, M. P., and Molenberghs, G. (1997). A review of electrocochleography: instrumentation settings and meta-analysis of criteria for diagnosis of endolymphatic hydrops. *Acta Otolaryngol. Suppl.* 526, 14–20. doi: 10.3109/00016489709124014
- Yoshie, N. (1968). Auditory nerve action potential responses to clicks in man. *Laryngoscope* 78, 198–215. doi: 10.1288/00005537-196802000-00002
- Yoshie, N. (1973). Diagnostic significance of the electrocochleogram in clinical audiometry. *Audiology* 12, 504–539. doi: 10.3109/00206097309071666
- Yoshie, N. (1976). “Electrocochleographic classification of sensorineural defects: pathological pattern of the cochlear nerve compound action potential in man,” in *Electrocochleography*, eds R. J. Ruben, C. Elberling and G. Salomon (Baltimore, MD: University Park Press), 353–386.
- Yoshie, N., and Ohashi, T. (1969). Clinical use of cochlear nerve action potential responses in man for differential diagnosis of hearing loss. *Acta Otolaryngol. Suppl.* 252, 71–87. doi: 10.3109/00016486909120514
- Yoshie, N., Ohashi, T., and Suzuki, T. (1967). Non-surgical recording of auditory nerve action potentials in man. *Laryngoscope* 77, 76–85. doi: 10.1288/00005537-196701000-00006

Conflict of Interest Statement: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Eggermont. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.