



Assessing the Present and Future Habitat Suitability of *Caligus rogercresseyi* (Boxshall and Bravo, 2000) for Salmon Farming in Southern Chile

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The sea louse (Caligus rogercresseyi) is the most relevant parasite for the farmed salmon industry in Chile, the second largest producer worldwide. Although spatial patterns of C. rogercressevi have been addressed from data obtained from established monitoring and surveillance programs, studies on its spatial ecology are limited. A wide geographic distribution of C. rogercresseyi is presumed in Chile; however, how this species could potentially be distributed in space is unknown. Our study presents an analysis of the habitat suitability for C. rogercresseyi in the entire area occupied by marine sites of salmon farms in Chile. Habitat suitability modeling was used to explore the likelihood of species spatial occurrence based on environmental characteristics. Due to the expanding salmon industry in southern Chile, we studied C. rogercresseyi habitat suitability models for present (average of 2005-2010) and two future projections (2050 and 2100) under different climate change scenarios. Models were constructed with the maxent algorithm using a large database of spatial C. rogercresseyi occurrences from the Chilean fisheries health authority and included 23 environmental variables obtained from the Ocean Rasters for Analysis of Climate and Environment (Bio-ORACLE). Habitat suitability models indicated that water temperature, water salinity, and current velocity of waters were the most important characteristics limiting C. rogercresseyi distribution in southern Chile. Habitat suitability models for current climate indicated a heterogeneous pattern with C. rogercresseyi being present in waters with temperature range 12.12-7.08°C (sd = 0.65), salinity range 33.7-25.5 pss (sd = 1.73), and current water velocity range 0.23–0.01 m^{-1} (sd = 0.02). Predictions for future projections in year 2050 and year 2100 suggest new clumped dispersion of the environmental conditions for C. rogercresseyi establishment. Our results suggest complexity and a wide dispersion

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of the biogeographic distribution of the *C. rogercresseyi* habitat suitability with potential implications for control strategies and environmental issues for salmon farming in Chile. Further investigations are required into *C. rogercresseyi* distribution in southern Chile considering the possible effect of climate change.

Keywords: sea lice, copepod, ecological modeling, habitat suitability model, maximum entropy, climate change, aquaculture, salmon farming

INTRODUCTION

The sea louse (Caligus rogercresseyi) is the most relevant parasite of salmon farming in Chile, the second-largest salmon producer worldwide. This marine copepod feeds on the mucus, skin, and blood of parasitized fish, altering the osmotic barrier, reducing appetite, decreasing food conversion, damaging the carcass, and as a consequence producing substantial economic losses (1). The three species of salmonids intensively farmed in Chile: Atlantic salmon (Salmon salar), rainbow trout (Oncorhynchus mykiss), and coho salmon (O. kisutch), are all susceptible to C. rogercresseyi infection (2). The life cycle of this copepod species includes eight different development stages: two nauplius and one copepodid, four chalimus, and the adult (3). The first three stages are planktonic, and the others are parasitic. The cost of the salmon production unit in Chile increases USD\$1.4/kg for the treatment against C. rogercresseyi (4). In addition, the economic impact of sea lice globally is estimated at USD\$650 million per year, considering the loss in weight gain and the predisposition to secondary diseases (5).

A wide geographic distribution of C. rogercresseyi is presumed according to the reports of salmon farming in southern Chile. Since the beginning of salmon farming in the 1980s on region X [Los Lagos], sea lice infestation has been reported with farms producing rainbow trout (O. mykiss) particularly on the island of Chiloé (2). With the advance of the salmon industry in the 1990s toward the region XI [Aysén], sea lice infestations were also confirmed in farmed Atlantic salmon (S. salar). These two regions are considered as northern Patagonian fjords, a complex and heterogeneous topographic and oceanographic habitat composed of fjords, gulfs, channels, and semi-enclosed water bodies (6). However, for the decade of 2000s, a parasitosis prevalence of 53.4% was estimated in salmon farms in both regions with a decreasing gradient for the farms located further south (7). Besides, with the health crisis due to Infectious Salmon Anemia virus (ISAV) (8), some farming companies expanded their salmon production to more isolated, pristine, and cold sites in the southernmost part of the country. Recently in 2017, the presence of C. rogercresseyi was reported for the first time in region XII [Magallanes], the southernmost sites with salmon farming in Chile, even though these isolated farms represent <10% of the industry (5).

Usually, the distribution patterns of *C. rogercresseyi* have been approached with an epidemiological focus or considering the effectiveness of control measures interests (7, 9–15), yet studies of spatial ecological patterns of this parasite are rare (16). However, how this species is distributed in space is unknown in Chile (17). Furthermore, this ambitious question can be partially answered by addressing *C. rogercresseyi* presence as a function of environmental conditions in the area occupied by the salmon industry. Predicting the spatial distribution of the environmental conditions relevant to *C. rogercresseyi* is necessary to know the potential geographic range where the health and economic impacts of this parasite to salmon farming in Chile are expected.

According to the ecological niche theory, a species only thrives within definite ranges of environmental conditions (18, 19). The habitat suitability of a species is the predictions of presence/absence of the likelihood of occurrence based on environmental characteristics in a statistical modeling process (20). However, the concept of suitable habitat models (HSMs) should be differentiated from ecological niche modeling, considering that the latter in a function that links the fitness of the species to their environment (21). Therefore, ecological niche modeling is a quantitative framework that includes evolutionary processes, interspecific relationships, migration (movement), and other biotic and abiotic aspects for individuals (22). The HSMs framework estimates a qualitative output (usually the presence) considering the relationship between environmental variables of biological relevance for the individuals.

This paper presents an analysis of the habitat suitability for C. rogercresseyi in southern Chile. The ongoing expansion of the salmon industry in Chile is an important characteristic of this parasitic dynamic. Although salmon production fell from 700,000 to 500,000 tons due to the ISAV health crisis in 2008, the industry has shown a growing recovery to 900,000 tons in 2014 (23). A central issue is a transnational and capitalistic salmon industry supported by the idea of economic growth from the national state. The foregoing poses health and environmental challenges for the development of this activity in the future. For this reason, we studied the predictions for HSMs using a presence-only method (Maxent) for the present (average of 2005-2010) and future projections (2050 and 2100) under different climate change scenarios. The importance of our mapping of HSMs is to know the spatial heterogeneity that C. rogercresseyi could exhibit under different scenarios and predict the possible impacts of this important health issue to the salmon industry.

METHODS

Study Area

The study area covers a latitudinal extension of \sim 1,500 km of coastline waters in southern Chile (41 to 54°S), across three geographical regions (region X [Los Lagos], region XI [Aysén], and region XII [Magallanes]) where salmon production





is established (**Figure 1**). This area covers 100% of the country's salmon farming activity. The average annual water temperature is 9.1° C (minimum of 6.6, maximum of 12.3), and the average annual water salinity is 33.32 pps (minimum of 25.38, maximum of 34.01).

Caligus rogercresseyi Data

The estimation of HSMs is based on a large database of spatial occurrences of C. rogercresseyi. We obtained the data from the monitoring program for C. rogercresseyi [Specific Sanitary Program for the Surveillance and Control of Caligidosis—SSPSC] maintained by The Chilean National Fisheries and Aquaculture Service (Sernapesca) (24). The objective of the SSPSC program is to monitor C. rogercressevi abundance in all active salmon farms throughout time and assess sanitary measures to control this parasitic infestation. The trained staff at each farm takes a random sample of 40 fish for the count total of individuals by parasitic phases of three categories: juveniles phases (includes chalimus I, II, III, and IV), mobile adults (males and no gravid females), and gravid female (egg sacs strings) (25). The data obtained of SSPSC consists of C. rogercresseyi abundance and the geographic location (decimal latitude and longitude coordinates) of all farms in operation from January 2012 to December 2018 on region X, XI, and XII in southern Chile. We consider C. rogercresseyi occurrences in each report of the presence of ovigerous female in the study area. The rationale behind this is that the gravid female indicates that the life cycle of the copepods is complete in a geographical site as a biological occurrence. Besides, the gravid female phase can be identified with the naked eye in the field as chains of eggs are visible and it is \sim 5 mm long (3). A C. rogercresseyi occurrence database was generated in MS Excel, including geographic coordinates. Duplicate occurrence records and absolute zeros of farm abundance during uptime were removed since it is assumed that the susceptible fish host is present but there is no evidence of interaction with the parasite and the design of the SSPSC program is to estimate the infection (the presumed false absences are very uncertain) (26). In addition, duplicate occurrences were removed because it is advisable to avoid multiple records for the same species in the same grid cell for the modeling process as salmon farms can report the C. rogercresseyi more than once. A parasite prevalence was estimated without distinction between old and new farms infected (number of positive farms by study period/number of active farms by study period). To determine optimal prevalence thresholds for inclusion as an occurrence in the modeling dataset, we created subsets based on the 100, 80, 60, 50, and 20 percentile prevalence distribution. In this case, the original data included records from 790 active farms for the 100 percentile (corresponding active farms by prevalence thresholds 80, 60, 50, and 20 percentile were 632, 474, 395, and 158 farms).

Accessible Location

The HSMs require a geographic location that is accessible to the species for a certain period (20). Thus, we defined a boundary area given the dispersal abilities of *C. rogercresseyi* based on occurrences in southern Chile. According to the available knowledge, the planktonic and infective phases can be dispersed 30 km around (27). Therefore, we imported the MS Excel occurrence database into a Geographic Information System (Quantum GIS, version 3.4) to estimate a buffer area. A layer polygon in shapefile was generated covering an area around 30 km from each occurrence of the database as an accessible location for *C. rogercresseyi* in the area occupied by the salmon industry.

Environmental Variables

Another important assumption of the HSMs is the identification of the environmental conditions necessary for the subsistence of the species. We used 23 environmental variables of the Bio-ORACLE (ocean rasters for analysis of climate and environment) marine surface dataset (http://www.bio-oracle.org) provided in grids with cells equally spaced in latitude and longitude with a spatial resolution of 5 arcmins (c. 9.2 km) (28). Bio-ORACLE is a global dataset consisting of geophysical, biotic, and climate variables for a present (average 2005-2010) and two future projections (2050 and 2100) based on Representative Concentration Pathways (RCP). The RCP projections describe different climate futures scenarios considered possible depending on the amount of greenhouse gases emitted in the years to come at the following temperature thresholds: RCP 2.6 (global temperature rise <1°C by 2100), RCP 4.5 (>1°C), RCP 6.0 $(>2^{\circ}C)$, and RCP 8.5 $(>3^{\circ}C)$. In other words, RCP 8.5 represents the worst-case climate change scenario (29). The Bio-ORACLE raster layers were imported into QGIS for clipping the "accessible location" proposed for the C. rogercresseyi using the shapefile as a mask.

To reduce the dimensionality of the 23 environmental variables, we examined with a Principal Component Analysis (PCA) using the PCA4cd plugin (QGIS, version 3.4). This analysis summarizes uncorrelated variables giving significant environmental information for HSMs (20). After reducing the number of variables by a statistical test, biological criteria were applied to the *C. rogercresseyi* life cycle, limiting the number of variables to conditions of water temperature °C, water salinity pss (practical salinity scale), and currents water velocity m⁻¹ (3, 16, 30–32).

The Modeling

The HSMs analysis is limited to occurrence data (presence) because the SSPCS program has a sanitary approach to establish farms that need chemical control for the parasitic infestation. Therefore, there are no directed efforts to estimate the strict absence of C. rogercresseyi. We use Maxent (maximum entropy modeling) as an algorithm that simulates pseudo-absences to complement the dependent variable (33). In this case, Maxent works as a Generalized Linear Model with Poisson distribution where the number of occurrence counts is a function of environmental variables representing a prediction of indices as habitat suitability for C. rogercresseyi on the study area (34). However, like any generalized prediction of environmental conditions, these models cannot capture ecological interactions, limiting themselves to an estimate where the species is most likely to occur. The foregoing should be considered with caution because these estimates do not represent quantitative estimates that suggest a greater or lesser occurrence of the species in the study area.

To fit HSMs Maxent models, we used specific settings: raw output (this feature estimates the ratio of the relative suitability of one pixel vs. another's pixel within the accessible location), 20% of random test (a quantity of the data is withheld every time and used for testing), with 100 replicates (amount of k-folds models to train), bootstrap (training data is selected by sampling with replacement from the presence points), and deactivating extrapolate and do clamping options (no projections outside the limits of the training data). Then, we made a range of models for the present using different regularization multiplier to avoid overfitting (0.5, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0). The output directory for these models was the environmental variables of the present (2005-2010). For these models, we estimated the Akaike information criterion (AIC) using the toolbox of Niche Analyst software (Niche A, version 3.0) (35). According to the reference framework of Multimodel Inference, under similar conditions, the models with the lowest AIC presents better-estimated values for a suitable habitat (36). To fit HDM Maxent models for two future projections (2050 and 2100), we used the same specific settings applying them to four climate change scenarios (RCP 2.6, RCP 4.5, RCP 6.0, and RCP 8.5), and AIC values were estimated for the range of the different regularization multiplier (0.05-5.0) selecting the AIC smallest model. The modeling process was developed using Maxent 3.4.1 (33).

To evaluate the models based on each prevalence threshold, we use the Area Under the Curve (AUC) by summing the estimates values for the Receiver Operating Characteristic (ROC). The ROC is a graphical estimate of sensitivity against 1specificity for all possible thresholds of the model. The summary statistic of AUC calculates values between 0 and 1 from ROC, where values >0.5 suggest that our model is better than random (37). In this case, the AUC values >0.5 and near 1 are useful to compare the predictive strength of multiple only presence data models using the same data (38). We selected as the best model the prevalence threshold for which models showed the highest AUC value. To evaluate HSM uncertainty, we detected areas of strict extrapolation as places where environmental conditions are non-analogous to those in areas across which the models were calibrated (39). The mobility-oriented parity metric (MOP nearest 5% of reference cloud) was used to identify such extrapolative areas resulting from calibration areas, compared with conditions under present and future climate scenarios using "ntbox" R package (40). Then, we excluded areas with strict extrapolation to represent only potential suitable areas with higher levels of certainty. After establishing the best model by AUC close to 1, we used the mean of the 100 replicates to generate a binary map (QGIS version 2.18) using as threshold the highest value that included 95% of occurrences (omission error 5%).

RESULTS

The original data included 68,114 records from 790 active farms during the study period with a 92% parasitic prevalence. A total

of 728 C. rogercresseyi occurrences from salmon farms were used to select the HSMs (Supplementary Material). Of these occurrences, 308 were established in Region X [Los Lagos], 409 in Region XI [Aysén], and 11 in Region XII [Magallanes] during the years 2012-2018. According to PCA analyses, only three environmental variables are necessary to explore the variability of the environmental conditions in the study area: mean water temperature, mean water salinity, and mean current water velocity. In addition, as low temperatures limit the development of C. rogercresseyi according to the biological cycle, we added a temperature minimum to the environmental variables set (this characteristic present correlation <0.7). The most parsimonious HSMs based on AIC scores included a regularization multiplier value of 0.5 in Maxent software (default value is 1). Besides, Maxent models performed better than random for each time prediction and showed a good fit (AUC > 0.95). Predicted habitat suitability for C. rogercresseyi using maximum entropy models is shown in Figure 1. The Maxent thresholds for climate change scenarios are shows in Figure 2.

The majority of predicted suitable habitat areas were near the coastline of the study area, where most salmon farms are located. However, a heterogeneous pattern is observed with a greater C. rogercresseyi presence in the northern sites. Therefore, the suitable habitat for C. rogercresseyi at current climate is widely distributed in waters with temperature range 12.12- $7.08^{\circ}C$ (sd = 0.65), salinity range 33.7-25.5 pss (sd = 1.73), and current water velocity range 0.23–0.01 m⁻¹ (sd = 0.02) across the study area. Besides, in region XII a clumped area (54° S, 71° W) is observed with suitable environmental conditions for C. rogercresseyi. In addition, the suitable habitat for C. rogercressevi is limited by water temperature minimum <5.5°C across the study area. Future predictions for 2050, under RCP 2.6 estimate a slight advance of the suitable habitat near Guamblin Island National Park (44° S, 74° W) toward the south in region XI but without uniform distribution toward region XII (Figure 1). Future predictions for 2050 under RCP 4.5 estimate a loss of the suitable habitat (44° S, 74° W) and a slight advance of new areas in River Island (45° S, 74° W) toward the south in region XI but without uniform distribution toward region XII (Figure 1). In the case of the 2050 predictions under RCP 6.0 and 85, a loss of the RCP 2.6 and RCP 4.5 new areas is perceived in region XI but with an increase in suitable areas in region XII on Mornington Island (49° S, 75° W), in sites not occupied by salmon farms under current conditions. Future predictions for 2100 predictions suggest greater coverage areas compared to models for 2050, mainly in Region XII. Model predictions for 2100 under RCP 26 indicated a focus in Region XI in the north part of Campana Island (48° S, 75° W). In addition, under RCP 45 projections the new area extends continuously to Region XII from Merino Jarpa Island (47° S, 74° W) to the northern part of Duke of York Island (50° S, 75° W) and an isolated area on Rennell Island (51° S, 74° W). Future predictions 2100 under RCP 60 indicated the contraction of the suitable habitat observed in the RCP45 projections and a new continuous area in region XII from the Bernardo O'Higgins national park (50° S, 74° W) to the south of the Alacalufes Natural Reserve (52 $^{\circ}$ S, 73 $^{\circ}$ W). Future predictions for 2100 under RCP 85 suggest a partial reduction



(dashed line) of different Representative Concentration Pathways (RCP 2.6, 4.5, 6.0, and 8.5) in two different projection future scenarios (2,050 and 2,100) compared to the present scenario 2000–2014 threshold (continuous line). The suitable conditions predicted for future scenarios (threshold) are similar to the present conditions, but they differ because different feature types allow different possible shapes of response curves.

of the areas, maintaining favorable conditions in Puerto Natales (47° S, 74° W) and new areas appearing to the south between the Recalada island (53° S, 74° W) and the Carlos island (54° S, 73° W) in region XII. The MOP results showed areas of strict extrapolation as uncertainty predictions when looking at results for calibration areas agreement among them, mainly in Region XI (46° S, 74° W) near Laguna San Rafael National Park, and in Region XII (52° S, 69° W) near San Gregorio Village (**Figure 1**).

DISCUSSION

We used presence-only maximum entropy modeling to represent the habitat suitability of C. rogercresseyi in southern Chile at present and projected for 2050 and 2100. Mapping outputs of HSMs suggest that habitat suitability almost entirely covers the area occupied by the salmon industry overpass Los Lagos to Aysén regions. Besides, future predictions for the years 2050 and 2100 suggest growing suitable habitat contiguous to the Aysén region with an isolated and delimited new area of favorable environmental conditions for C. rogercresseyi in the Magallanes region. We cautiously suggest avoiding farming use of these newly suitable areas shown in future predictions (Figure 1). The parasite dynamics would be similar in these new areas to that observed in northern areas, which would be added to the costs of maintaining salmon production and control measures in more remote sites (41). This heterogeneous pattern and the environmental variables detected involved have potential implications in C. rogercresseyi biogeography, environmental issues, and control strategies.

Using the maxent algorithm to project probable future scenarios based on current data entails uncertainty. Other statistical methods as generalized linear models or generalized additive models consider strict species absence reducing uncertainty (42). A fundamental limitation of presence-only data using maxent is that some sites in the study area are sampled more intensively than others (38). For our data, it is relevant to consider that *C. rogercresseyi* infection requires a susceptible host density at existing farms (43, 44). Therefore, we obtained models that combine the *C. rogercresseyi* habitat suitability distribution with the distribution of salmon farms. On the other hand, MOP results identify high uncertainty areas with strict extrapolation risks as a key approach in this regard, suggesting prediction variation depending on the environmental data used to create models.

Temperature is the most important environmental characteristic to predict the *C. rogercresseyi* habitat suitability, with less cold waters within the study area favoring its occurrence. Development rates of parasitic copepods are dependent on temperature, altering their geographic distribution and increasing infestations in farms at higher temperatures (45). Besides, it is known that the development time of the different phases of *C. rogercresseyi* is affected by temperature (3). The complete life cycle can take 45 days at a mean temperature of 10.3° C. However, the life cycle can be completed in just

18 days at a temperature of 16.7° C. This is in agreement with our results in which we found more suitable habitats for *C. rogercresseyi* in Los Lagos and Aysén regions due to warmer waters compared to Magallanes region. On the other hand, predictions based on future climate change scenarios suggest that habitat suitability can be expanded in a specific area of Magallanes region. Moreover, this should be taken with caution because increasing the mean temperature could potentially reduce the development time of the parasitic phases or could also cause distress in the copepod dynamic (e.g., patterns prediction for 2100 in scenarios RCP 4.5, 6.0, and 8.5; **Figure 1**) (46, 47).

Minimum temperature defines the environmental limit of the geographic range for *C. rogercresseyi* in our models. The temperature threshold of $<4.2^{\circ}$ C not only stops the development of all phases but also causes the death of this species of copepod (3). This threshold is only observed for the southernmost sites in our study area where there are no occurrences of the species. However, the increase in the mean temperature expected with climate change projections would modify the minimum temperature threshold, expanding the habitat suitability for *C. rogercresseyi* in Chile (29). This implies the need to understand how the salmon farming in Chile interacts with the environment, mainly the conflicts over marine space with other aquaculture activities and whether a more sustainable industry is possible (23).

Salinity and current velocity are the variables with the least importance in our models. In other species of sea lice in the northern hemisphere (Lepeophtheirus salmonis and C. elongatus), low salinities <30 pss have been observed to decline reproductive and survival rates (48). For the species C. rogercresseyi, a variable tolerance and sensitivity to salinities <20 pss have been described (31). Experimental studies show variations between 0, 15-100, 45-100, and 100% C. rogercresseyi survival rate at salinity concentrations of 0, 10, 20, and 30 pss, respectively. However, the slight salinity effect observed in our models may be because the low concentration is associated with specific geographical sites, such as estuaries and brackish waters. Concerning velocity current, it is understood that current can flush the planktonic phases and the infective phase toward neighboring farms (27). Nonetheless, this effect could be limited on a large scale, since it has been reported that for L. salmonis the current can transport the phases only between around 7.3 and 10 km (49). In addition, information is still lacking on the role of current velocity in the transport of planktonic phases of C. rogercresseyi in the complex fjord systems of southern Chile.

The pattern predicted by our models is consistent with previous reports of this species of copepod in Chile. The first spatial and temporal description of *C. rogercresseyi* proposes a cluster pattern according to the number of farms in operation during 1999–2000 (50). Subsequent analyses with data from 2007 identified a decreasing pattern from north to south on the abundance of *C. rogercresseyi* in salmon farms (7, 51). However, our study shows a clumped advance of the suitability conditions under the assumptions of climate change, aggravating

sea lice infestations, and their economic and environmental consequences in the medium-term future for Chile. Besides, according to our results, a latitudinal gradient is observed in the suitable habitat of *C. rogercresseyi*, where its likelihood of occurrence increases toward the north in our study area. It is necessary to expand the biogeographic knowledge of the *C. rogercresseyi* due to the trend of expansion of aquaculture in the face of the reduction of the global capture fisheries. Besides, there are hypotheses about a wider distribution range of the *C. rogercresseyi* from Peru to Argentina (passing through Chile), complicating the control strategies by the natural sources of the parasite (17, 52).

The habitat suitability distribution proposed by our models reflects a challenge for sea lice control strategies in Chile. The environmental conditions of less cold temperature $>10^{\circ}$ C, salinities > 25 pss, and low velocity of currents will guarantee the thrive of the C. rogercresseyi in Los Lagos and Aysén regions for the next 80 years. Consequently, the loss of sensitivity of sea lice to chemical control can be repeated in future scenarios in southern Chile. Before 2007, emamectin benzoate was exclusively used to control C. rogercresseyi infestations, causing loss of sensitivity to this antiparasitic drug (53). After 2007, the health authority approved the use of deltamethrin, followed by diflubenzuron in 2009, cypermethrin in 2010, and azamethiphos in 2013 (14). The implementation of azamethiphos as a new alternative of chemical control in 2013 coincides with an increase of C. rogercresseyi abundance at salmon farms in region X [Los Lagos] (54). Although up to now the use of azamethiphos maintains limited parasitic loads, it is necessary to improve the proper management of chemicals and propose non-chemical options due to the expansion and emergence of antiparasitic-resistant C. rogercresseyi.

In addition, the impacts of C. rogercresseyi chemical control on non-target aquatic organisms is an environmental concern of relevance under the projections of geographic expansion for this copepod in future scenarios. Emamectin benzoate, diflubenzuron, deltamethrin, cypermethrin, and hydrogen peroxide at the recommended dose reduce the feeding and mobility, producing paralysis and dead in copepods and causing >95% mortality in another zooplankton and phytoplankton organisms under laboratory conditions (55, 56). Otherwise, cypermethrin and deltamethrin in high doses cause 100% mortality in the marine amphipods Monocorophium insidiosum (57). Furthermore, the synergistic effect of cypermethrin and deltamethrin combination on crustacean Daphnia magna has been described to cause superior toxicity to the half-maximum effective concentration [EC50; (58)]. In larger crustaceans, specifically the stage IV lobsters (Homarus americanus), a high concentration of azamethiphos produces agitation, relaxing erratically, and aggressiveness and causes 40% mortality in 4 h of experimental continuous exposure (59). Emamectin benzoate lacks these toxic effects in H. americanus (60). However, it is alarming that the pelagic marine ecosystem of Chile is characterized by high endemism of copepods (61-63), marine organisms very sensitive to toxicity effects of chemical control used against C. rogercresseyi.

Other impacts of C. rogercresseyi include their interaction with native fish, where low abundances are observed for native fish Rock cod (Eleginnops maclovinus) and Chilean silverside (Odontesthes regia) (2). However, C. rogercressevi parasitism on salmonid production species shows high abundances and infestations in Atlantic salmon (S. salar) and rainbow trout (Oncorhynchus mykiss), while low abundances are shown in coho salmon (O. kisutch) due to a better immune response mediated by macrophages (64). Besides, stocking density $(>22 \text{ kg m}^{-3})$, the season of the year (autumn), depth of water at the farm and cage location (>50 m), and chemical control (>1 week later) are all factors associated with higher C. rogercresseyi infestation rates (16, 51). Furthermore, the growing trend of salmon farming worldwide expects a growing host density, causing negative feedback for this host-parasitic dynamics (44).

The maps from HSMs provide a useful guide to hypothesize the occurrence of C. rogercresseyi in different climate change scenarios, mainly for the Magallanes region. However, these models are limited by only-presence data of salmon farms in Chile. We lack interpretation to establish the limited dispersal and potential distribution for using pseudo-absences. In addition, our study area could be small for the spatial distribution of this species by ignoring biotic interactions (competition, commensalism, multi-species studies) and the migration process (20). In this respect, we limit our predictions to the area of calibration of the models, with utility only for the area occupied by the salmon industry in Chile. Interpolation is acceptable and necessary because we used the largest database of C. rogercresseyi occurrence that is available. Our results suggest complexity and a wide dispersion of the biogeographic behavior of the C. rogercresseyi in the south of Chile; further investigations are required in these areas.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

ML-L wrote the manuscript and analyzed and interpreted the data. JE-D and NZ-V collected the data and prepared the figures. CA review and wrote the manuscript. FM wrote the manuscript and conceived and planned the study. All authors contributed to the article and approved the submitted version.

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REFERENCES

- Johnson SC, Treasurer JW, Bravo S, Nagasawa K, Kabata Z. A review of the impact of parasitic copepods on marine aquaculture. *Zool Stud.* (2004) 43:229–43. Available online at: http://zoolstud.sinica.edu.tw/Journals/ 43.2/229.pdf
- Carvajal J, González L, George-Nascimento M. Native sea lice (Copepoda: Caligidae) infestation of salmonids reared in netpen systems in southern Chile. Aquaculture. (1998) 166:241–6. doi: 10.1016/S0044-8486(98)00301-9
- González L, Carvajal J. Life cycle of *Caligus rogercresseyi*, (Copepoda: Caligidae) parasite of Chilean reared salmonids. *Aquaculture*. (2003) 220:101– 17. doi: 10.1016/S0044-8486(02)00512-4
- Dresdner J, Chávez C, Quiroga M, Jiménez D, Artacho P, Tello A. Impact of Caligus treatments on unit costs of heterogeneous salmon farms in Chile. *Aquac Econ Manag.* (2019) 23:1–27. doi: 10.1080/13657305.2018.1449271
- Arriagada G, Valenzuela-Muñoz V, Arriagada AM, Núñez-Acuña P, Brossard M, Montecino K, et al. First report of the sea louse *Caligus rogercresseyi* found in farmed Atlantic salmon in the Magallanes region, Chile. *Aquaculture*. (2019) 512:734386. doi: 10.1016/j.aquaculture.2019.734386
- Niklitschek EJ, Soto D, Lafon A, Molinet C, Toledo P. Southward expansion of the Chilean salmon industry in the Patagonian Fjords: main environmental challenges. *Rev Aquac.* (2013) 5:172–95. doi: 10.1111/raq.12012
- Hamilton-West C, Arriagada G, Yatabe T, Valdés P, Hervé-Claude LP, Urcelay S. Epidemiological description of the sea lice (*Caligus rogercresseyi*) situation in southern Chile in August 2007. *Prev Vet Med.* (2012) 104:341– 5. doi: 10.1016/j.prevetmed.2011.12.002
- Asche F, Hansen H, Tveteras R, Tveteras S. The salmon disease crisis in Chile. J Agric Resour Econ. (2010) 24:405–11. doi: 10.1086/mre.24.4.426 29664
- Arriagada GA, Stryhn H, Campistó JL, Rees EE, Sanchez J, Ibarra R, et al. Evaluation of the performance of pyrethroids over different life stages of *C. rogercresseyi* in southern Chile. *Aquaculture*. (2014) 426–7:231– 7. doi: 10.1016/j.aquaculture.2014.02.007
- Arriagada G, Hamilton-West C, Nekouei O, Foerster C, Müller A, Lara M, et al. *Caligus rogercresseyi* infestation is associated with Piscirickettsia salmonis-attributed mortalities in farmed salmonids in Chile. *Prev Vet Med.* (2019) 171:104771. doi: 10.1016/j.prevetmed.2019.104771
- Arriagada G, Sanchez J, Stryhn H, Vanderstichel R, Campistó JL, Ibarra R, et al. A multivariable assessment of the spatio-temporal distribution of pyrethroids performance on the sea lice *Caligus rogercresseyi* in Chile. *Spat Spatiotemporal Epidemiol.* (2018) 26:1–13. doi: 10.1016/j.sste.2018.04.004
- Bravo S, Nuñez M, Silva MT. Efficacy of the treatments used for the control of *Caligus rogercresseyi* infecting Atlantic salmon, *Salmo salar* L., in a new fish-farming location in Region XI, Chile. J Fish Dis. (2013) 36:221– 8. doi: 10.1111/jfd.12023
- Bravo S, Treasurer J, Sepulveda M, Lagos C. Effectiveness of hydrogen peroxide in the control of *Caligus rogercresseyi* in Chile and implications for sea louse management. *Aquaculture*. (2010) 303:22–7. doi: 10.1016/j.aquaculture.2010.03.007
- Marín SL, Ibarra R, Medina MH, Jansen PA. Sensitivity of *Caligus rogercresseyi* (Boxshall and Bravo 2000) to pyrethroids and azamethiphos measured using bioassay tests-A large scale spatial study. *Prev Vet Med.* (2015) 122:33– 41. doi: 10.1016/j.prevetmed.2015.09.017
- Rozas M, Asencio G. Assessment of epidemiologic situation of caligiasis in Chile: toward to effective control strategy. *Salmociencia*. (2007) 2:43–59. Available online at: https://www.researchgate.net/publication/285708305_

Sanitary Program for the Surveillance and Control of Caligidosis (SSPSC Caligidosis).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fvets. 2020.615039/full#supplementary-material

Evaluacion_de_la_Situacion_Epidemiologica_de_la_Caligiasis_en_Chile_ Hacia_una_estrategia_de_control_efectiva

- Bravo S, Silva MT, Treasurer J. Factors affecting the abundance of *Caligus rogercresseyi* (Boxshall and Bravo) on farmed salmonids in Chile in the period 2006-2007. *Aquaculture.* (2014) 434:456– 61. doi: 10.1016/j.aquaculture.2014.09.009
- Hemmingsen W, MacKenzie K, Sagerup K, Remen M, Bloch-Hansen K, Dagbjartarson Imsland AK. *Caligus elongatus* and other sea lice of the genus Caligus as parasites of farmed salmonids: a review. *Aquaculture*. (2020) 522:735160. doi: 10.1016/j.aquaculture.2020.735160
- Guisan A, Thuiller W. Predicting species distribution: offering more than simple habitat models. *Ecol Lett.* (2005) 8:993– 1009. doi: 10.1111/j.1461-0248.2005.00792.x
- Austin MP. Spatial prediction of species distribution: an interface between ecological theory and statistical modelling. *Ecol Modell.* (2002) 157:101– 18. doi: 10.1016/S0304-3800(02)00205-3
- Hirzel AH, Le Lay G. Habitat suitability modelling and niche theory. J Appl Ecol. (2008) 45:1372–81. doi: 10.1111/j.1365-2664.2008.01524.x
- Peterson AT. Uses and requirements of ecological niche models and related distributional models. *Biodivers Informatics*. (2006) 3:59–72. doi: 10.17161/bi.v3i0.29
- Peterson AT. Predicting the geography of species' invasions via ecological niche modeling. Q Rev Biol. (2003) 78:419–33. doi: 10.1086/378926
- Chávez C, Dresdner J, Figueroa Y, Quiroga M. Main issues and challenges for sustainable development of salmon farming in Chile: a socio-economic perspective. *Rev Aquac.* (2019) 11:403–21. doi: 10.1111/raq.12338
- Sernapesca. Specific Sanitary Program for the Surveillance and Control of Caligidosis - SSPSC. (2015). Available online at: http://www.sernapesca. cl/programas/programa-sanitario-especifico-de-vigilancia-y-control-decaligidosis
- Sernapesca. Caligus Surveillance Guide for Chile. (2015). Available online at: http://www.sernapesca.cl/index.php?option=com_content&view=article& id=976&Itemid=903
- Escobar LE, Craft ME. Advances and limitations of disease biogeography using ecological niche modeling. *Front Microbiol.* (2016) 7:1–21. doi: 10.3389/fmicb.2016.01174
- Kristoffersen AB, Rees EE, Stryhn H, Ibarra R, Campisto JL, Revie CW, et al. Understanding sources of sea lice for salmon farms in Chile. *Prev Vet Med.* (2013) 111:165–75. doi: 10.1016/j.prevetmed.2013.03.015
- Tyberghein L, Verbruggen H, Pauly K, Troupin C, Mineur F, De Clerck O. Bio-ORACLE: a global environmental dataset for marine species distribution modelling. *Glob Ecol Biogeogr.* (2012) 21:272–81. doi: 10.1111/j.1466-8238.2011.00656.x
- Riahi K, Rao S, Krey V, Cho C, Chirkov V, Fischer G, et al. RCP 8.5-A scenario of comparatively high greenhouse gas emissions. *Clim Change*. (2011) 109:33–57. doi: 10.1007/s10584-011-0149-y
- Bravo S, Erranz F, Lagos C. A comparison of sea lice, *Caligus rogercresseyi*, fecundity in four areas in southern Chile. J Fish Dis. (2009) 32:107– 13. doi: 10.1111/j.1365-2761.2008.01012.x
- Bravo S, Pozo V, Silva MT. The tolerance of *Caligus rogercresseyi* to salinity reduced in southern Chile. *Bull Eur Assoc Fish Pathol.* (2008) 28:198–206. Available online at: https://eafp.org/download/2008-Volume28/Issue%205/ 28_198.pdf
- Bravo S. The reproductive output of sea lice *Caligus rogercresseyi* under controlled conditions. *Exp Parasitol.* (2010) 125:51– 4. doi: 10.1016/j.exppara.2009.12.001

- Phillips SB, Aneja VP, Kang D, Arya SP. Maximum entropy modeling of species geographic distributions. *Ecol Modell.* (2006) 190:231–59. doi: 10.1016/j.ecolmodel.2005.03.026
- Merow C, Smith MJ, Silander JA. A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and settings matter. *Ecography*. (2013) 36:1058–69. doi: 10.1111/j.1600-0587.2013.07872.x
- Qiao H, Peterson AT, Campbell LP, Soberón J, Ji L, Escobar LE. NicheA: creating virtual species and ecological niches in multivariate environmental scenarios. *Ecography*. (2016) 39:805–13. doi: 10.1111/ecog.01961
- Burnham KP, Anderson DR. Multimodel inference: understanding AIC and BIC in model selection. Sociol Methods Res. (2004) 33:261-304. doi: 10.1177/0049124104268644
- Radosavljevic A, Anderson RP. Making better Maxent models of species distributions: complexity, overfitting and evaluation. J Biogeogr. (2014) 41:629–43. doi: 10.1111/jbi.12227
- Elith J, Phillips SJ, Hastie T, Dudík M, Chee YE, Yates CJ. A statistical explanation of MaxEnt for ecologists. *Divers Distrib.* (2011) 17:43– 57. doi: 10.1111/j.1472-4642.2010.00725.x
- Owens HL, Campbell LP, Dornak LL, Saupe EE, Barve N, Soberón J, et al. Constraints on interpretation of ecological niche models by limited environmental ranges on calibration areas. *Ecol Modell*. (2013) 263:10– 8. doi: 10.1016/j.ecolmodel.2013.04.011
- Osorio-Olvera L, Lira-Noriega A, Soberón J, Peterson AT, Falconi M, Contreras-Díaz RG, et al. ntbox: An r package with graphical user interface for modelling and evaluating multidimensional ecological niches. *Methods Ecol Evol.* (2020) 11:1199–206. doi: 10.1111/2041-210X.13452
- Abolofia J, Wilen JE, Asche F. The cost of lice: quantifying the impacts of parasitic sea lice on farmed salmon. *Mar Resour Econ.* (2017) 32:329– 49. doi: 10.1086/691981
- 42. Wiens JA, Stralberg D, Jongsomjit D, Howell CA, Snyder MA. Niches, models, and climate change: assessing the assumptions and uncertainties. *Proc Natl Acad Sci U.S.A.* (2009) 106:19729–36. doi: 10.1073/pnas.0901639106
- Krkošek M. Host density thresholds and disease control for fisheries and aquaculture. Aquac Environ Interact. (2010) 1:21–32. doi: 10.3354/aei00004
- 44. Jansen PA, Kristoffersen AB, Viljugrein H, Jimenez D, Aldrin M, Stien A. Sea lice as a density-dependent constraint to salmonid farming. *Proc R Soc B Biol Sci.* (2012) 279:2330–8. doi: 10.1098/rspb.2012.0084
- Costello MJ. Ecology of sea lice parasitic on farmed and wild fish. Trends Parasitol. (2006) 22:475–83. doi: 10.1016/j.pt.2006.08.006
- Garzke J, Hansen T, Ismar SMH, Sommer U. Combined effects of ocean warming and acidification on copepod abundance, body size and fatty acid content. *PLoS ONE*. (2016) 11:e155952. doi: 10.1371/journal.pone.0155952
- Kurihara H, Shimode S, Shirayama Y. Effects of raised CO2 concentration on the egg production rate and early development of two marine copepods (*Acartia steueri* and *Acartia erythraea*). Mar Pollut Bull. (2004) 49:721– 7. doi: 10.1016/j.marpolbul.2004.05.005
- Boxaspen K. A review of the biology and genetics of sea lice. *ICES J Mar Sci.* (2006) 63:1304–16. doi: 10.1016/j.icesjms.2006.04.017
- 49. Brooks KM. The effects of water temperature, salinity, and currents on the survival and distribution of the infective copepodid stage of sea lice (*Lepeophtheirus salmonis*) originating on Atlantic salmon farms in the Broughton Archipelago of British Columbia, Canada. *Rev Fish Sci.* (2005) 13:177–204. doi: 10.1080/10641260500207109
- Zagmutt-Vergara FJ, Carpenter TE, Farver TB, Hedrick RP. Spatial and temporal variations in sea lice (Copepoda: Caligidae) infestations of three salmonid species famed in net pens in southern Chile. *Dis Aquat Organ.* (2005) 64:163–73. doi: 10.3354/dao064163
- Yatabe T, Arriagada G, Hamilton-West C, Urcelay S. Risk factor analysis for sea lice, *Caligus rogercresseyi*, levels in farmed salmonids in southern Chile. J *Fish Dis*. (2011) 34:345–54. doi: 10.1111/j.1365-2761.2011.01238.x
- 52. Bravo S, Boxshall GA, Conroy G. New cultured host and a significant expansion in the known geographical range of the sea louse *Caligus*

rogercresseyi. Bull Eur Assoc Fish Pathol. (2011) 31:156-60. Available online at: https://eafp.org/download/2011-Volume31/issue-4/710%20Bravo.pdf

- Bravo S, Sevatdal S, Horsberg TE. Sensitivity assessment in the progeny of *Caligus rogercresseyi* to emamectin benzoate. *Bull Eur Assoc Fish Pathol.* (2010) 30:99–105. Available online at: https://eafp.org/download/ 2010-Volume30/Issue%203/Bravo.pdf
- Mancilla-Schulz J, Marín SL, Molinet C. Dynamics of *Caligus rogercresseyi* (Boxshall & Bravo, 2000) in farmed Atlantic salmon (*Salmo salar*) in southern Chile: are we controlling sea lice? *J Fish Dis.* (2019) 42:357– 69. doi: 10.1111/jfd.12931
- Buschmann AH, Riquelme VA, Hernández-González MC, Varela D, Jiménez JE, Henríquez LA, et al. A review of the impacts of salmonid farming on marine coastal ecosystems in the southeast Pacific. *ICES J Mar Sci.* (2006) 63:1338–45. doi: 10.1016/j.icesjms.2006.04.021
- Buschmann AH, Cabello F, Young K, Carvajal J, Varela DA, Henríquez L. Salmon aquaculture and coastal ecosystem health in Chile: analysis of regulations, environmental impacts and bioremediation systems. *Ocean Coastal Manage*. (2009) 52:243–9. doi: 10.1016/j.ocecoaman.2009.03.002
- 57. Tucca F, Díaz-Jaramillo M, Cruz G, Silva J, Bay-Schmith E, Chiang G, et al. Toxic effects of antiparasitic pesticides used by the salmon industry in the marine amphipod Monocorophium insidiosum. *Arch Environ Contam Toxicol.* (2014) 67:139–48. doi: 10.1007/s00244-014-0008-8
- Rose S, Altenburger R, Sturm A. Mixture toxicity effects of sea louse control agents in Daphnia magna. *Chemosphere*. (2016) 144:599–606. doi: 10.1016/j.chemosphere.2015.08.053
- 59. Burridge LE, Haya K, Waddy SL, Wade J. The lethality of anti-sea lice formulations Salmosan (azamethiphos) and Excis (Cypermethrin) to stage IV and adult lobsters (Homarus americanus) during repeated short-term exposures. *Aquaculture*. (2000) 182:27–35. doi: 10.1016/s0044-8486(99)00251-3
- Burridge LE, Hamilton N, Waddy SL, Haya K, Mercer SM, Greenhalgh R, et al. Acute toxicity of emamectin benzoate (SLICETM) in fish feed to *American lobster, Homarus americanus. Aquacult Res.* (2004) 35:713–22. doi: 10.1111/j.1365-2109.2004.01093.x
- Hidalgo P, Escribano P, Vergara O, Jorquera E, Donoso K, Mendoza P. Patterns of copepod diversity in the Chilean coastal upwelling system. *Deep Sea Res Part II*. (2010) 57:2089–97. doi: 10.1016/j.dsr2.2010.09.012
- Morales CM, Torreblanca L, Hormazabal S, Ramírez MC, Nuñez S, Hidalgo P. Mesoscale structure of copepod assemblages in the coastal transition zone and oceanic waters off central-southern Chile. *Prog Oceanogr.* (2010) 84:158–73. doi: 10.1016/j.pocean.2009.12.001
- Pino-Marambio J, Mordue AJ, Birkett M, Carvajal J, Asencio G, Mellado A, Quiroz A. Behavioural studies of host, non-host and mate location by the Sea Louse, Caligus rogercresseyi Boxshall & Bravo, 2000 (Copepoda: Caligidae). *Aquaculture*. (2007) 271:70–6. doi: 10.1016/j.aquaculture.2007.05.025
- 64. Valenzuela-Muñoz V, Boltaña S, Gallardo-Escárate C. Comparative immunity of *Salmo salar* and *Oncorhynchus kisutch* during infestation with the sea louse *Caligus rogercresseyi*: an enrichment transcriptome analysis. *Fish Shellfish Immunol.* (2016) 59:276–87. doi: 10.1016/j.fsi.2016. 10.046

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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