



Results of Dynamic Contrast-Enhanced Ultrasound Correlate With Treatment Outcome in Canine Neoplasia Treated With Electrochemotherapy and Interleukin-12 Plasmid Electrotransfer

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Electrochemotherapy (ECT) and/or gene electrotransfer of plasmid DNA encoding interleukin-12 (GET pIL-12) are effective treatments for canine cutaneous, subcutaneous, and maxillofacial tumors. Despite the clinical efficacy of the combined treatments of ECT and GET, data on parameters that might predict the outcome of the treatments are still lacking. This study aimed to investigate whether dynamic contrast-enhanced ultrasound (DCE-US) results of subcutaneous tumors differ between tumors with complete response (CR) and tumors without complete response (non-CR) in dogs treated with ECT and GET pIL-12. Eight dogs with a total of 12 tumor nodules treated with ECT and GET pIL-12 were included. DCE-US examinations were performed in all animals before and immediately after therapy as well as 8 h and 1, 3, and 7 days later. Clinical follow-up examinations were performed 7 and 14 days, 1 and 6 months, and 1 year after treatment. Numerous significant differences in DCE-US parameters were noted between tumors with CR and non-CR tumors; perfusion and perfusion heterogeneity were lower in CR tumors than in non-CR tumors. Therefore, studies with larger numbers of patients are needed to investigate whether DCE-US results can be used to predict treatment outcomes and to make effective decisions about the need for repeated therapy or different treatment combinations in individual patients.

Keywords: dog, contrast-enhanced ultrasonography, electrochemotherapy, immunotherapy, gene electrotransfer, bleomycin, cisplatin, IL-12 plasmid

INTRODUCTION

Dynamic contrast-enhanced ultrasound (DCE-US) is a simple, readily available, non-invasive, safe, and inexpensive method for assessing tissue perfusion at the capillary level and correlates with histological results of vessel density in preclinical (1–3) and clinical studies (4–7). Moreover, DCE-US has been used to predict the efficacy of antiangiogenic treatments of various tumors in preclinical (3) and human clinical trials (8–17), whereas DCE-US had no predictive value in canine tumors treated with radiotherapy (18). Furthermore, DCE-US results correlated with the results of advanced diagnostic imaging (19). The contrast agents used in DCE-US are gas-filled microbubbles stabilized in a lipoprotein shell that have a diameter of 1–3 μm , which is small enough to migrate freely through the circulation and large enough to remain in the vascular space (20–22). Capillary filling results in diffuse enhancement of perfused tissue. Most of the contrast agent is excreted through the lungs within 20 min after administration (23). The advantages of contrast agents used in DCE-US over those used in dynamic contrast-enhanced computed tomography (DCE-CT) are numerous: the contrast agents allow real-time imaging, there is no ionizing radiation, they are neither nephro- nor hepatotoxic, and they have very few, very mild side effects (22–24). In humans, contraindications to microbubble administration include pulmonary hypertension and impaired cardiopulmonary function (24). In a large number of dogs in which DCE-US examinations were performed, <1% developed an immediate effect, including vomiting and/or syncope, or delayed adverse effects, including vomiting (23).

Electrochemotherapy (ECT) and/or gene electrotransfer of plasmid DNA encoding interleukin-12 (GET pIL-12) are effective treatments for cutaneous, subcutaneous and maxillofacial tumors in dogs (25–41), superficial cell carcinoma in cats (42), cutaneous tumors in ferrets (43) and sarcoid tumors in horses (44, 45). Several preclinical (46–49) and clinical studies in veterinary patients (26–28, 33, 36, 50) have shown that the effect of ECT is potentiated by GET pIL-12, and ECT has become an established standard of care for a variety of human cancers: cutaneous and subcutaneous tumors, including melanoma, squamous cell carcinoma, basal cell carcinoma, and other metastases (51–58); hepatocellular carcinoma and colorectal liver metastases (59–63); pancreatic neoplasia (64–66); and others. A portion of the antitumor efficacy of electroporation (EP)-based therapies arises from the effect of EP on the vasculature of the treated tumor, inducing a local blood flow effect, namely, “vascular lock,” i.e., small blood vessel vasoconstriction and increased wall permeabilization (67–74). The vascular effects of EP are enhanced by the use of chemotherapeutic agents in ECT treatment, and the effect lasts longer in tumors than in healthy tissue, namely, the “vascular disrupting effect” (67, 68, 72).

Despite the clinical efficacy of the combined treatments of ECT and GET pIL-12, there is still a lack of data on parameters that might predict the outcome of the treatments. For EP-based treatments, it could be assumed that the therapy should be repeated if it does not reflect in the expected “vascular lock” immediately after the treatment and/or anti-angiogenic effects in the days after.

This pilot study aimed to investigate whether the DCE-US results from subcutaneous tumors correlate with treatment outcomes in dogs treated with ECT combined with GET pIL-12.

MATERIALS AND METHODS

Design and Setting

Eight dogs (seven females and one male) with a total of 12 superficial tumor nodules (11 mast cell tumors and 1 neurofibrosarcoma) treated with ECT and GET pIL-12 were included (**Table 1**). Their mean age with standard deviation was 8.0 ± 2.3 years. Six dogs had one tumor, and two dogs had three tumors. Each nodule was measured in three perpendicular directions (a, b, c), and tumor volume was calculated using the formula: $V = a \times b \times c \times \pi/6$. Owners of the dogs signed an informed consent form before inclusion.

ECT Combined With GET pIL-12

The procedures were performed under general anesthesia: the dogs received 0.2 mg/kg midazolam (Midazolam Torrex, Torrex Pharma GesmbH, Vienna, Austria) intravenously, and anesthesia was induced by 3–6 mg/kg propofol (Diprivan, Zeneca, Grangemouth, United Kingdom) administered intravenously and maintained with isoflurane (Isoflurin, Vetpharma Animal Health, Barcelona, Spain). All patients received fluid therapy throughout the procedure by administering Hartmann’s solution (B Braun Melsungen AG, Melsungen, Germany) at a rate of 5 mL/kg/h.

An electrical pulse generator, Cliniporator™ (IGEA s.r.l., Carpi, Italy), was used to deliver electrical pulses through either plate, hexagonal, or needle electrodes. The selection of electrode type, voltage, duration, and frequency of the electrical pulses was based on ECT (23, 35, 48–52) and GET studies (25–28, 33) (**Table 2**).

For the ECT procedure, two dogs received bleomycin (Blenoxane, Bristol-Myers, NY, USA) at a concentration of 3 mg/mL intravenously at a dose of 0.3 mg/kg, and six dogs received cisplatin (cis-diammine dichloroplatin II, Cisplatin Accord 1 mg/mL, Accord Health Care, Warsaw, Poland) at a concentration of 1 mg/mL and at a dose of 1 mg/cm³ intratumorally (**Table 2**).

For the GET procedure, the pCMVcaIL-12 plasmid encoding canine IL-12 was used, isolated using the Qiagen Endo-Free kit (Qiagen, Hilden, Germany), and diluted to a concentration of 1 mg/mL in endotoxin-free water (Qiagen). Quality control and quantification were performed (28). The plasmid was injected at a dose of 2 mg per patient (27, 28, 33) peritumorally in two dogs and intratumorally in six dogs (**Table 2**). When more than one tumor was present in a patient, the dose of pIL-12 was divided among the tumors proportional to the tumor volume (**Table 2**).

DCE-US

DCE-US examinations were performed in all animals before and immediately after therapy as well as 8 h and 1, 3, and 7 days later (**Figure 1**). For the first two measurements, the dogs were under general anesthesia (described above for ECT combined with GET) but were awake for the DCE-US measurements at 8 h and 1, 3, and 7 days after therapy. The contrast agent Sonovue

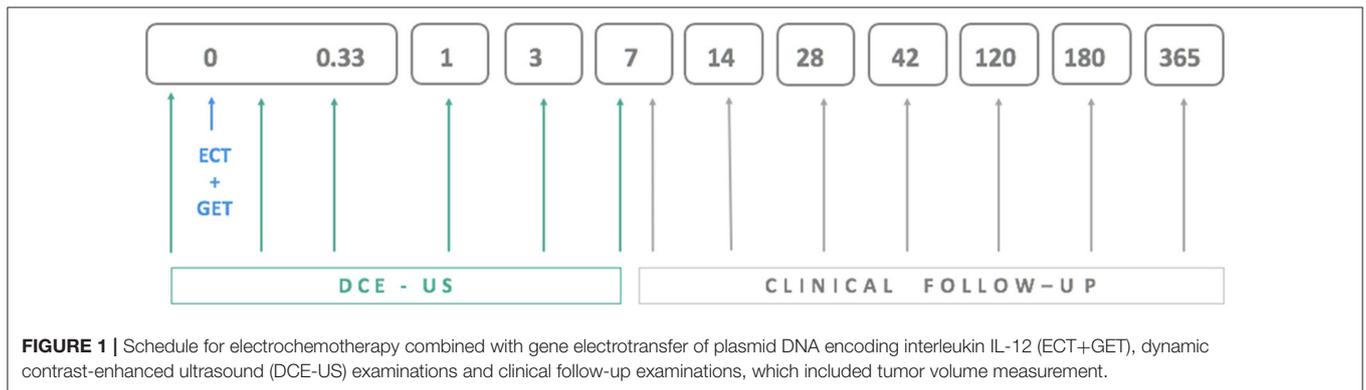
TABLE 1 | Characteristics of the eight dogs treated with electrochemotherapy (ECT) combined with gene electrotransfer of plasmid DNA encoding canine interleukin-12 (GET pIL-12).

| Patient no. | Age | Sex | Breed | Weight (kg) | Tumor | No of tumors | Tumor volume (cm ³) |
|-------------|----------|--------|------------------------|-------------|-------------------|--------------|---------------------------------|
| 1 | 9 y 11 m | Female | Mixed breed | 25.0 | Mast cell tumor | 1 | 0.38 |
| 2 | 9 y 7 m | Female | German hunting terrier | 11.3 | Mast cell tumor | 3 | 0.04; 0.004; 0.01 |
| 3 | 9 y 10 m | Female | Beagle | 17.3 | Neurofibrosarcoma | 1 | 23.00 |
| 4 | 10 y 8 m | Female | Mixed breed | 34.4 | Mast cell tumor | 3 | 1.10; 0.10; 0.004 |
| 5 | 6 y 5 m | Female | Basset hound | 29.0 | Mast cell tumor | 1 | 5.00 |
| 6 | 4 y 10 m | Female | Golden retriever | 34.2 | Mast cell tumor | 1 | 0.92 |
| 7 | 5 y 11 m | Male | Boston terrier | 10.7 | Mast cell tumor | 1 | 0.37 |
| 8 | 6 y 7 m | Female | Bernese mountain dog | 40.3 | Mast cell tumor | 1 | 0.27 |

TABLE 2 | Electrochemotherapy (ECT) combined with gene electrotransfer of plasmid DNA encoding canine interleukin-12 (GET pIL-12) treatment regimens (dosages, route of administration, type of electrodes and pulse parameters) and outcome per patient.

| Patient no. | ECT | | | | | GET pIL12 | | | Outcome |
|-------------|------|----------------------------------|----------------|--------------------|------------------|------------------------------|--------------------|------------------|----------------------------------|
| | Drug | Dose (mg) | Administration | Type of electrodes | Pulse parameters | pIL-12 administration (2 mg) | Type of electrodes | Pulse parameters | |
| 1 | BLM | 6.90 | i.v. | Plate | ECT pulses | p.t. | MEA | GET pulses | CR |
| 2 | CDDP | T1: 0.60 T2: 0.20 T3: 0.20 | i.t. | Plate | ECT pulses | p.t. | MEA | GET pulses | Non-CR (PD) Non-CR (PD) CR |
| 3 | CDDP | 12.00 | i.t. | / | / | i.t. | Hexagonal | ECT pulses | Non-CR (PR) |
| 4 | BLM | 10.47 | i.v. | / | / | i.t. | Hexagonal | ECT pulses | Non-CR (PD) CR CR |
| 5 | CDDP | 5.00 | i.t. | / | / | i.t. | Hexagonal | ECT pulses | CR |
| 6 | CDDP | 0.90 | i.t. | / | / | i.t. | Hexagonal | ECT pulses | CR |
| 7 | CDDP | 0.37 | i.t. | / | / | i.t. | Needle | ECT pulses | Non-CR (PR) |
| 8 | CDDP | 0.27 | i.t. | / | / | i.t. | Plate | ECT pulses | CR |

BLM, bleomycin; CDDP, cisplatin; i.v., intravenously; i.t., intratumorally; p.t., peritumorally; T, tumor nodule; ECT pulses, 8 pulses of 100 μs duration with an amplitude to electrode distance ratio of 1,300 V/cm and a frequency of repetition of 5 kHz; GET pulses = 150 ms pulse of amplitude 170 V/cm; / = electroporation was performed simultaneously for ECT and GET; CR, tumor with complete response; MEA, multielectrode array; non-CR, tumor without complete response; PD, progressive disease; PR, partial response.



(Bracco, Milan, Italy) was administered into the cephalic vein at dose 0.5–1.5 mL per dog, depending on the weight of the animal (<10 kg: 0.06 mL/kg, 10–20 kg: 0.05 mL/kg, 20–30 kg: 0.04 mL/kg, and >30 kg: 0.03 mL/kg).

Ultrasound examinations were performed with a Resona 7 ultrasound scanner and a linear probe L11-3u with a frequency of 3–10 MHz (Mindray, Shenzhen, China). Low mechanical index harmonic non-linear ultraband contrast imaging was used. From

the time of contrast application, a 90-s recording was made. The Dicom files of the examinations were imported into the free software Vuebox™ (Bracco, Milan, Italy) to quantify tissue perfusion with DCE-US. Each tumor was carefully delineated, and two additional regions of interest (ROIs), each representing half of the tumor and the reference region representing the tissue below the tumor, were drawn. The arrival time of the contrast agent was manually selected. For each of the perfusion or time-intensity curves representing signal intensity over time, the following was recorded: basic intensity (BI) when no contrast agent was present, peak intensity (PI), and time to peak (TTP) in ms, the time at which the contrast intensity reached its peak. PE (peak enhancement) was calculated as the difference between PI and BI.

The dogs were closely monitored for adverse effects of contrast administration: vomiting, respiratory distress, syncope, nausea, and other effects. The dogs were hospitalized for the first three measurements. For the last three measurements, the dogs were monitored as outpatients for immediate effects (<1 h) and by the owner for delayed effects.

Clinical Follow-Up Examinations

Clinical follow-up examinations that included measurement of the three perpendicular tumor dimensions were performed at 7 and 14 days, 1 and 6 months and 1 year after treatment (Figure 1). Tumors were classified as having a complete response (CR) or not having a complete response (non-CR) with the latter including partial response (at least 30% decrease in tumor size), progressive disease (>20% increase in tumor size), and stable disease (neither sufficient shrinkage to qualify for partial response nor sufficient increase to qualify for progressive disease) according to RECIST (75) and iRECIST (76) criteria.

Statistical Analysis

Statistical software R, version 3.6.2, was used for the statistical analysis (77). The parameters of interest are defined in Table 3.

The normality of the data was tested using the Shapiro Wilk test. Data were not normally distributed; therefore, the comparison between CR and non-CR tumors for each variable was calculated using the Wilcoxon rank-sum test. Statistical significance was set at 5%.

RESULTS

Response to Therapy

The size of the tumors varied from 0.004 to 23.0 cm³ (median 0.32 cm³). The median baseline tumor volume did not differ between CR and non-CR tumors. Furthermore, none of the DCE-US parameters correlated with pretherapy tumor volume.

There were seven tumors with CR in six dogs: dogs 1, 2, 4, 5, 6, and 8. Clinical follow-up examination of the tumor of dog 1 is presented in Figure 2. There were five tumors without CR in four dogs (dogs 2, 3, 4, and 7; Table 2). Three of the non-CR tumors were classified as progressive disease, and two showed a partial response (Table 2).

No adverse effects of contrast administration were noted in our study.

DCE-US Results

The PE values were significantly lower in tumors with CR than in tumors with non-CR at all time points after therapy, except on day 3 (Figure 3). The difference in PE between CR and non-CR tumors was highest 8 h after therapy and gradually decreased in the following days but remained statistically significant on days 3 and 7. Note that the CR tumors showed no contrast enhancement immediately and 8 h after therapy; this finding is in contrast to non-CR tumors, which were still filled with microbubbles (Figure 4).

The PE ratio (Table 3) was significantly reduced in CR tumors compared to non-CR tumors immediately, 8 h and 7 days after therapy. The difference steadily increased over time, reaching a 45-fold decrease by day 7 (Figure 5).

The PE change (Table 3) was significantly reduced in CR tumors at four time points: immediately, 8 h, and 1 and 7 days after therapy. The highest difference between the two tumor groups (20-fold decrease) was observed immediately after therapy. With time, the difference decreased but remained statistically significant (Figure 6).

The percentage change in TTP from baseline TTP (TTP_{ch}) was more than three times greater 7 days after therapy in tumors with CR than in tumors without CR (Figure 7) because TTP increased from baseline in CR tumors and decreased from baseline in non-CR tumors.

The percentage difference in the change in PE between the two parts of the tumor (PE_{ROI dif_{ch}}), which describes tumor heterogeneity in PE, was more than 10–13 times lower immediately, 8 h and 3 days after therapy in tumors with CR compared to tumors with non-CR (Figure 8).

The parameters TTP, TTP_{ROI dif}, PE_{ROI dif}, and TTP ratio (Table 3) were not significantly different between CR and non-CR tumors.

When results for both groups (CR vs. non-CR tumors) regardless of the time were compared, PE, PE ratio, PE_{ch}, TTP_{ROI dif}, and TTP_{ROI dif_{ch}} were significantly lower, and TTP_{ch} was significantly higher in tumors with CR than in tumors with non-CR (Tables 3, 4).

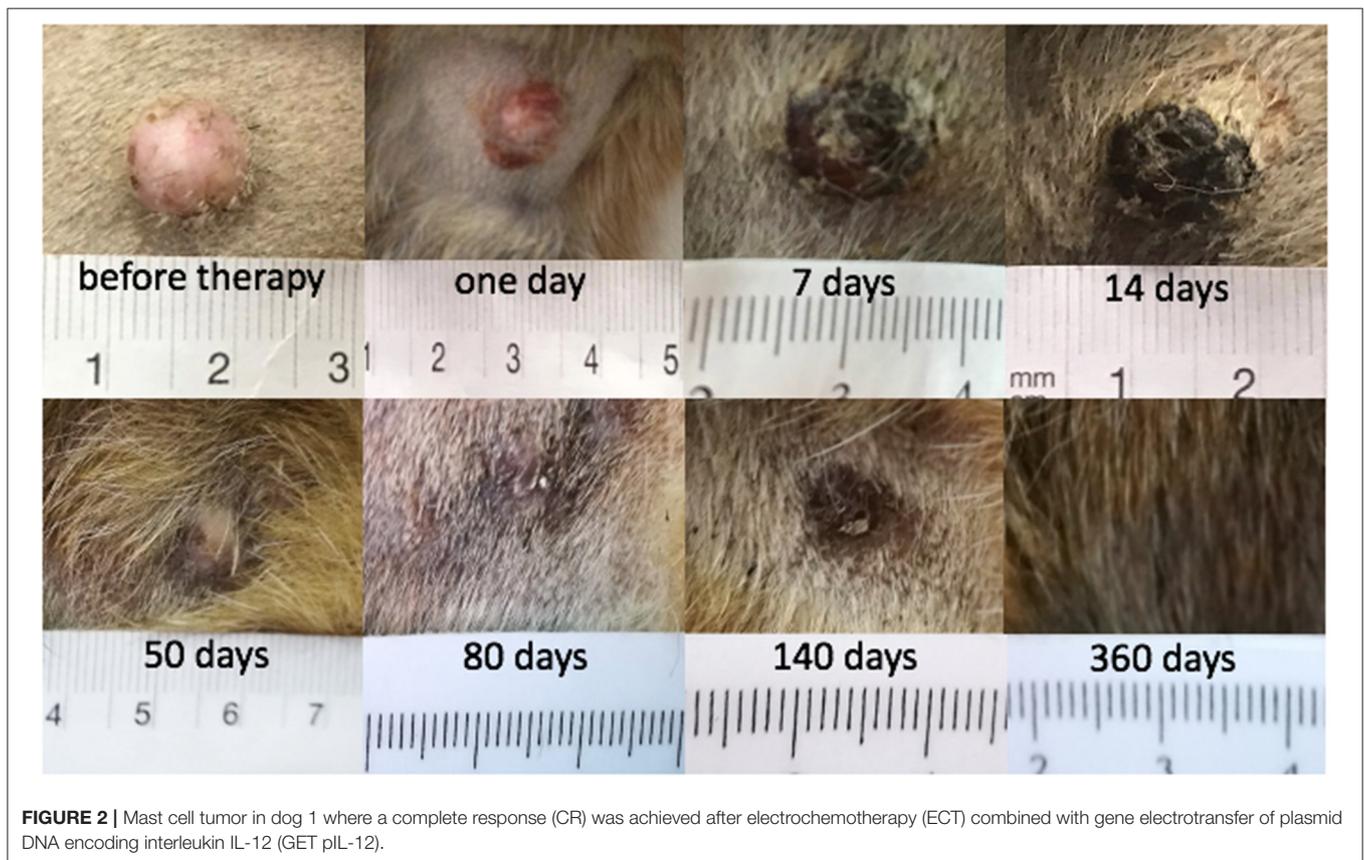
DISCUSSION

This study shows a significant difference in DCE-US results between canine tumors that achieved a CR to ECT combined with GET pIL-12 and non-CR tumors. After therapy, perfusion of tumors was lower in the CR group, and perfusion heterogeneity was greater in non-CR tumors.

To the best of our knowledge, this is the first study to compare DCE-US results with treatment outcomes in canine tumors treated with ECT combined with GET pIL-12. Our results are consistent with human studies that have shown DCE-US results to be a useful tool for predicting the efficacy of other antiangiogenic treatments in metastatic renal cell carcinoma, advanced hepatocellular carcinoma, colorectal carcinoma, metastatic breast cancer, gastrointestinal stromal

TABLE 3 | Definition of tumor volume and dynamic contrast-enhanced ultrasound parameters (DCE-US) and their calculation.

| Parameter | Symbol | Calculation |
|--|--------------------------------------|--|
| Volume of tumor before the treatment (cm ³) | V | $a \times b \times c \times \pi/6$ (a,b,c = perpendicular tumor dimensions) |
| Peak enhancement | PE | = Peak Intensity (PI) – Basic Intensity (BI) |
| Time to peak | TTP | |
| Ratio between PE of the tumor and PE of the reference (PE _{ref}) | PE ratio | $= \frac{PE}{PE_{ref}}$ |
| Ratio between TTP of the tumor and TTP of the reference | TTP ratio | $= \frac{TTP}{TTP_{ref}}$ |
| Percentage change in PE | PE ch | $= \frac{PE_L}{PE_0} \times 100 - 100$ |
| Percentage change in TTP | TTP ch | $= \frac{TTP_L}{TTP_0} \times 100 - 100$ |
| Percentage difference in PE between ROI1 and ROI2 | PE _{ROI} dif | $= \frac{PE_{ROI1} - PE_{ROI2}}{(PE_{ROI1} + PE_{ROI2})/2} \times 100$ |
| Percentage difference in TTP between ROI1 and ROI2 | TTP _{ROI} dif | $= \frac{TTP_{ROI1} - TTP_{ROI2}}{(TTP_{ROI1} + TTP_{ROI2})/2} \times 100$ |
| Percentage difference in change of PE between ROI1 and ROI2 | PE _{ROI} dif _{ch} | $= \frac{PE_{chROI1} - PE_{chROI2}}{(PE_{chROI1} + PE_{chROI2})/2} \times 100$ |
| Percentage difference in change of TTP between ROI1 and ROI2 | TTP _{ROI} dif _{ch} | $= \frac{TTP_{chROI1} - TTP_{chROI2}}{(TTP_{chROI1} + TTP_{chROI2})/2} \times 100$ |



tumors, and metastatic melanoma (9–11, 14–16). Tumors included in this study were compared between the two groups regarding tumor perfusion and perfusion heterogeneity. Before therapy, no differences in perfusion parameters and perfusion heterogeneity were noted between CR and non-CR tumors. Perfusion decreased after therapy in all CR tumors, which is consistent with previously described vascular lock and vascular disrupting action of EP-based therapies (63–74). The decrease in tumor perfusion was greater in CR compared with non-CR tumors, supporting the assumption that therapy is more likely

to be effective if it is reflected in immediate “vascular lock.” Furthermore, the difference remained statistically significant until day 7 with the exception of day 3. This finding is consistent with the expectation that the outcome of therapy is less likely to be favorable when therapy does not show antiangiogenic effects that result in decreased perfusion in the days after treatment. A similar trend was observed on day 3. However, due to owner non-compliance and thus missing data from one dog with three tumors, the difference was not statistically significant.

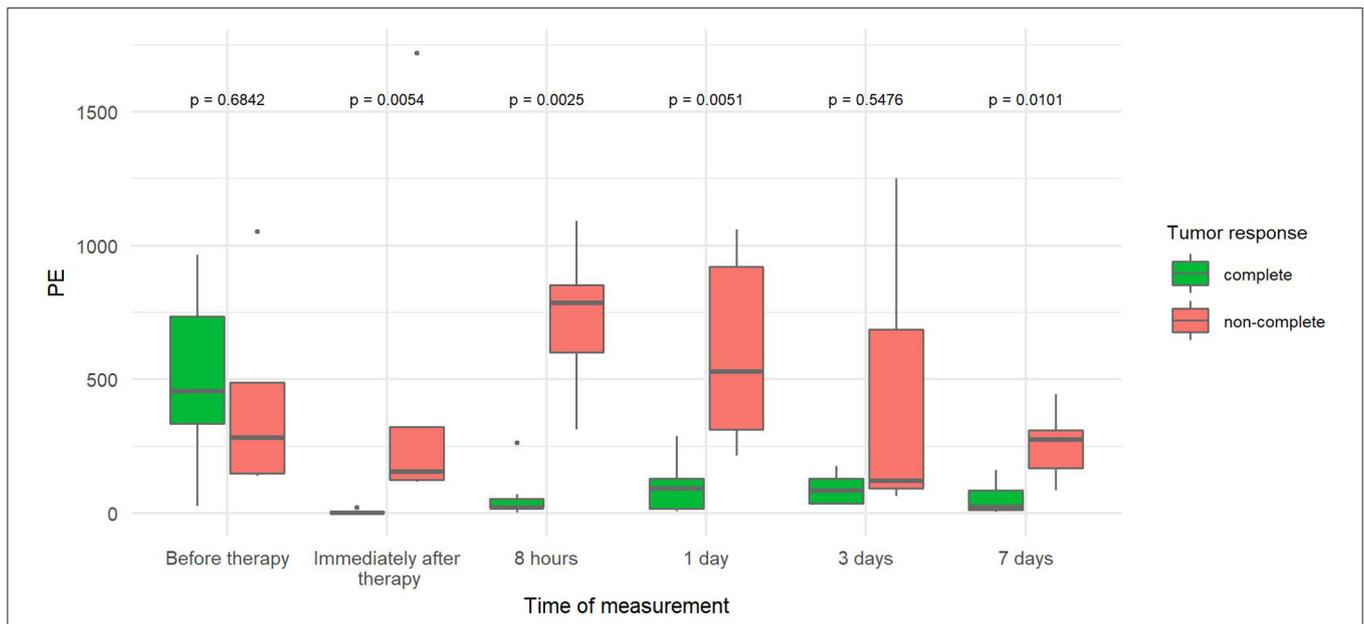


FIGURE 3 | Comparison of peak enhancement (PE) at different time points between canine tumors with complete (CR) and non-complete (non-CR) responses to combined electrochemotherapy treatment with gene electrotransfer of plasmid DNA encoding interleukin IL-12 (ECT GET pIL-12). Note the *p*-values presented at the top of the image. Note that PE is significantly lower for complete response tumors than for non-complete response tumors at all time points after therapy except at 3 days.

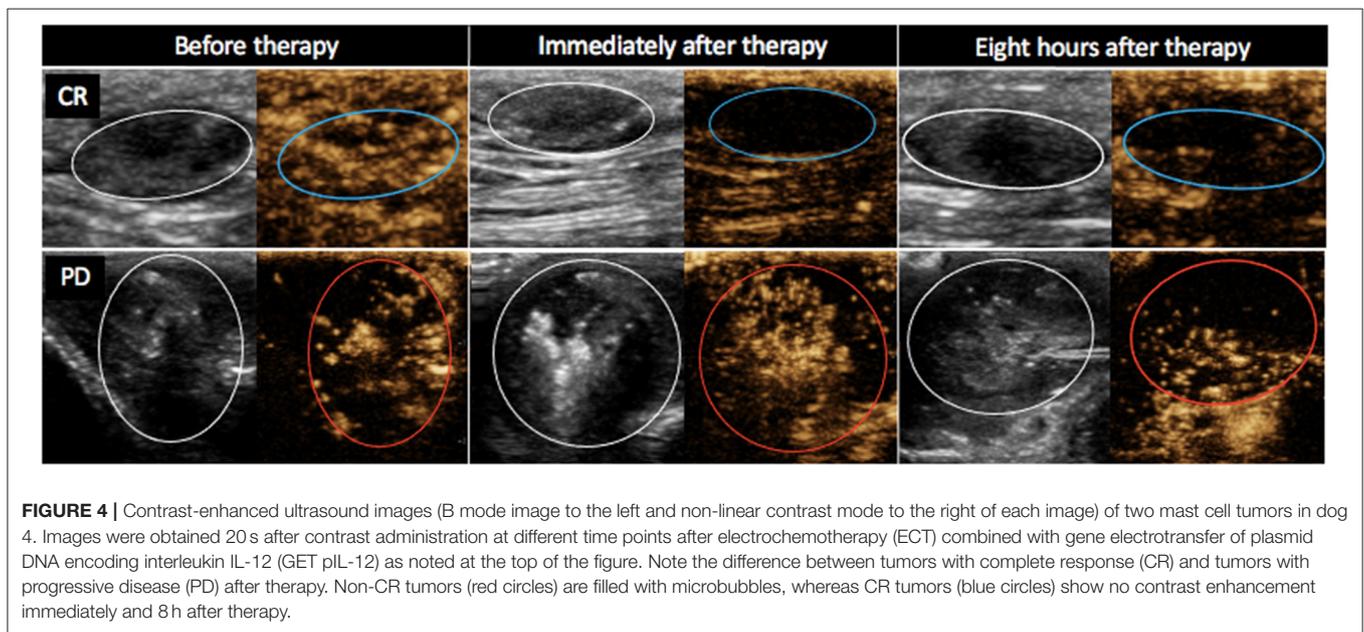


FIGURE 4 | Contrast-enhanced ultrasound images (B mode image to the left and non-linear contrast mode to the right of each image) of two mast cell tumors in dog 4. Images were obtained 20 s after contrast administration at different time points after electrochemotherapy (ECT) combined with gene electrotransfer of plasmid DNA encoding interleukin IL-12 (GET pIL-12) as noted at the top of the figure. Note the difference between tumors with complete response (CR) and tumors with progressive disease (PD) after therapy. Non-CR tumors (red circles) are filled with microbubbles, whereas CR tumors (blue circles) show no contrast enhancement immediately and 8 h after therapy.

In a study of nine spontaneous canine tumors treated with GET pIL-12 (25), a significant decrease in the DCE-US parameters wash-in area under the curve describing relative blood volume and wash-in rate describing blood flow velocity was observed at days 8 and 35 compared to baseline, consistent with the results of our study. Similar parameters reflecting relative blood volume and blood flow velocity, namely PE and TTP, respectively, were investigated by the same authors in a

study of five dogs treated with GET pIL-12 in combination with metronomic cyclophosphamide (30). They observed a significant decrease in PE and prolonged TTP 35 days after treatment, which is consistent with our results, while in contrast to our results, an increase in PE was observed 8 days after treatment. The difference in the dynamics of PE can most likely be attributed to the fact that the study by Cicchelerio et al. (30) investigated GET pIL-12 (without ECT), which does not

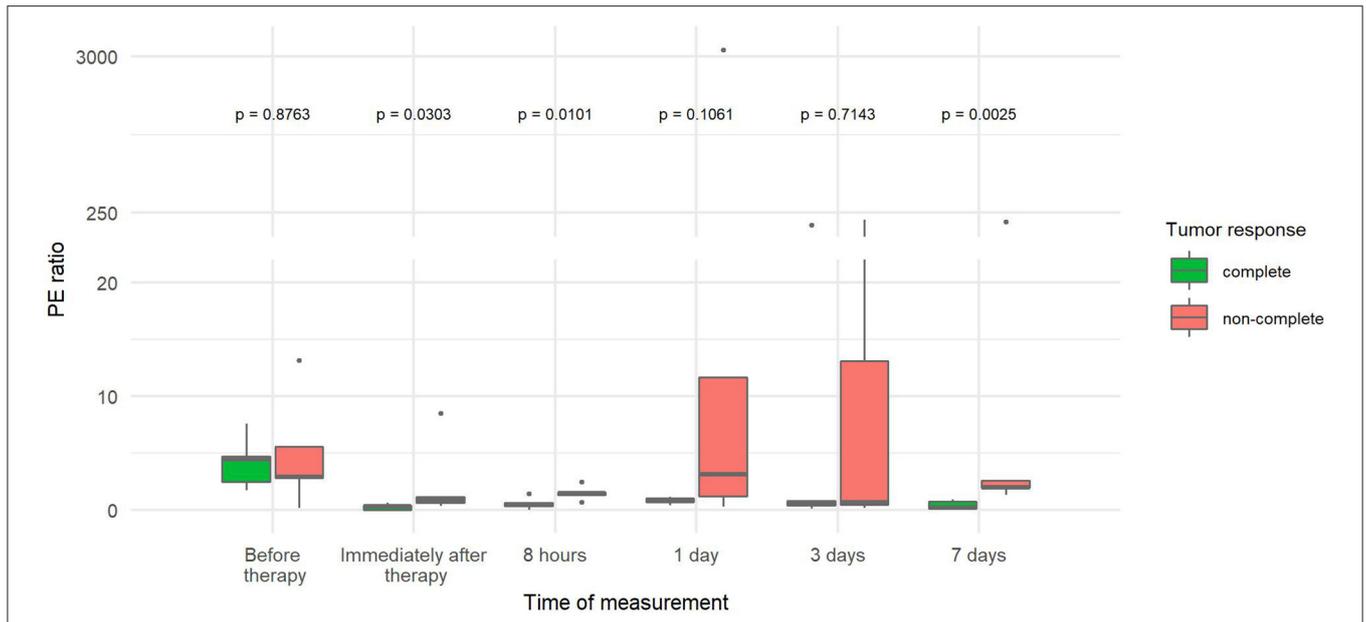


FIGURE 5 | Comparison of ratios between tumor peak enhancement and reference peak enhancement (PE ratio) at different time points between canine tumors with a complete and non-complete response to electrochemotherapy (ECT) with gene electrotransfer of plasmid DNA encoding interleukin IL-12 (GET pIL-12). Note the *p*-values presented at the top of the image. PE ratio is significantly lower for complete response tumors compared with non-complete response tumors immediately after therapy and at 8 h and 7 days.

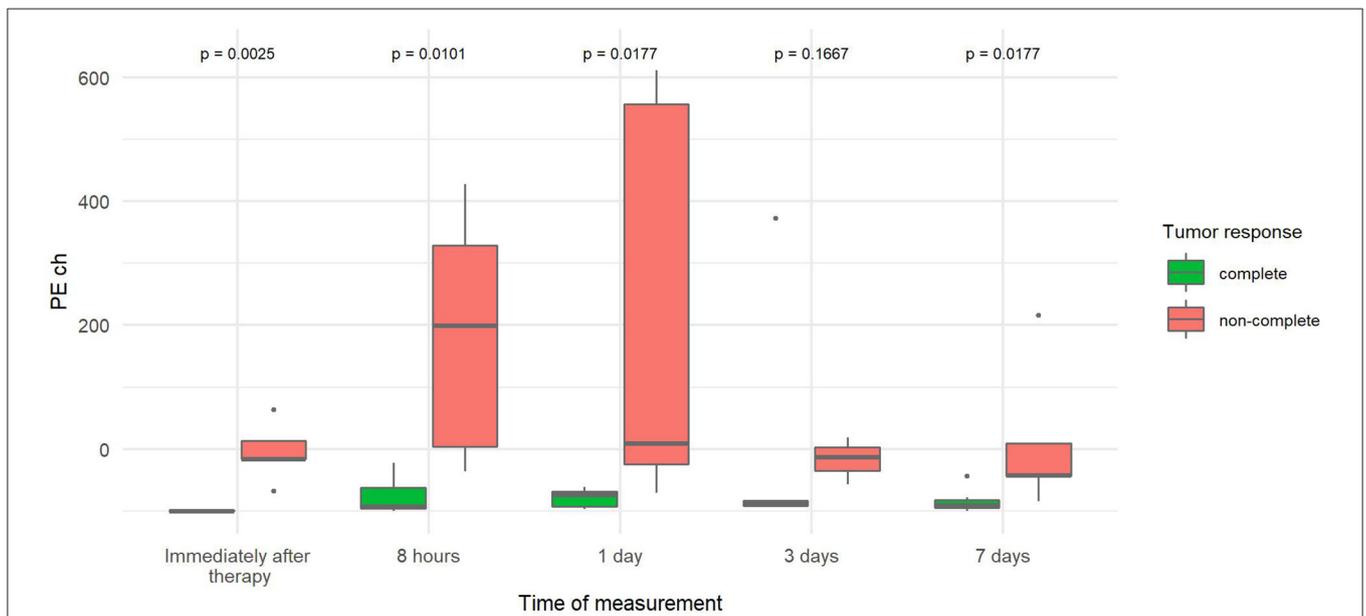
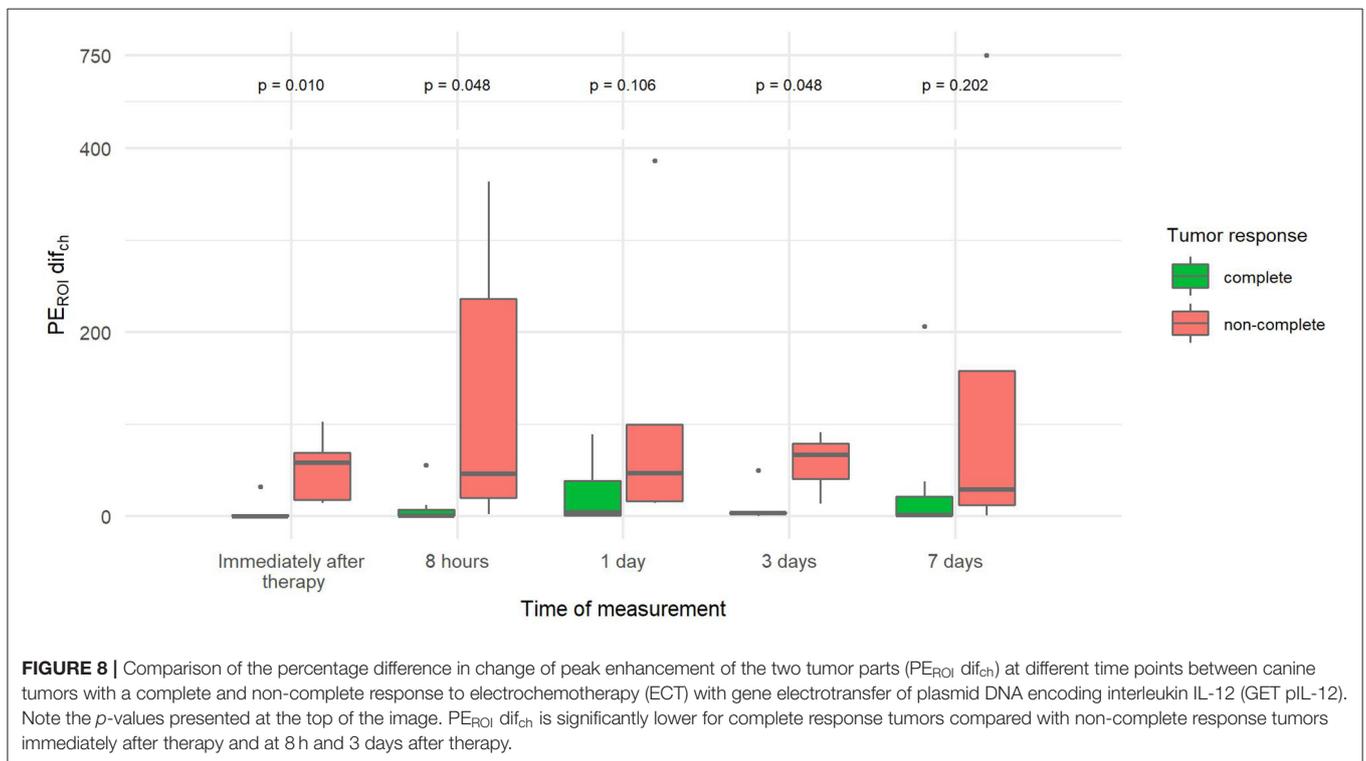
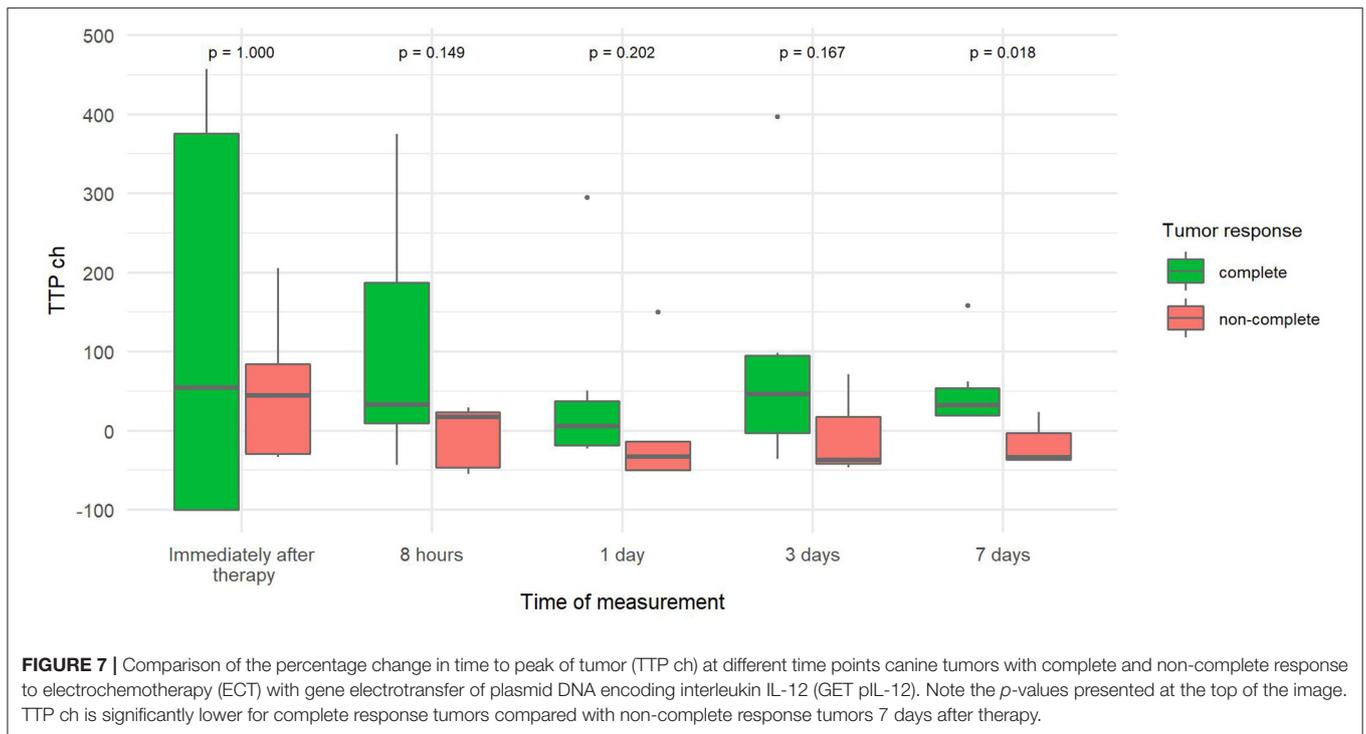


FIGURE 6 | Comparison of the percentage change in peak enhancement of tumor (PE ch) at different time points between canine tumors with a complete and non-complete response to electrochemotherapy (ECT) with gene electrotransfer of plasmid DNA encoding interleukin IL-12 (GET pIL-12). Note the *p*-values presented at the top of the image. PE ch is significantly lower for complete response tumors compared with non-complete response tumors immediately after therapy and at 8 h and 7 days.

induce a “vascular disrupting effect,” i.e., cytotoxic effect of chemotherapeutic drugs on vascular endothelial cells (67, 68, 72) that can be observed after treatment with ECT. Treatment

with GET pIL-12 results in a much less pronounced antitumor effect (27), also observed in this study, as no clinically relevant outcome was observed (30). However, it is likely that the decrease



in PE and prolongation of TTP 35 days after treatment is a consequence of the antiangiogenic effect of gene therapy. An additional difference between our studies is also the fact that the latter study (30) included five different tumor types and DCE-US

was performed only at three time points, i.e., before therapy and at days 8 and 35; our data cannot be directly compared because they were collected at different time points and after different treatments.

TABLE 4 | Comparison of overall results of dynamic contrast-enhanced ultrasound parameters (DCE-US) between tumors with complete response (CR) and tumors without complete response (non-CR) treated with electrochemotherapy (ECT) combined with gene electrotransfer (GET pIL-12).

| Parameter | Tumor response | | Wilcoxon rank sum test | |
|--------------------------------------|--|---|-------------------------|---------|
| | CR | Non-CR | | |
| | Median (1 st and 3 rd quartile) | Median (1 st and 3 rd quartile) | <i>p</i> -value | |
| V | Volume of tumor before the treatment (cm ³) | 0.27 (0.06, 0.65) | 0.37 (0.04, 1.10) | 0.8072 |
| PE | Peak enhancement | 32.50 (14.87, 150.90) | 312.56 (155.21, 803.50) | <0.0001 |
| TTP | Time to peak | 9.41 (6.70, 17.07) | 8.90 (6.36, 13.42) | 0.1242 |
| PE ratio | Ratio between PE of the tumor and PE of the reference | 0.64 (0.34, 1.10) | 1.77 (0.84, 3.77) | <0.0001 |
| TTP ratio | Ratio between TTP of the tumor and TTP of the reference | 0.96 (0.57, 1.24) | 0.79 (0.60, 1.29) | 0.6926 |
| PE ch | Percentage change in PE | -92.36 (-97.21, -79.13) | 3.69 (-39.22, 131.43) | <0.0001 |
| TTP ch | Percentage change in TTP | 27.91 (-6.49, 94.75) | 29.06 (-37.56, -26.51) | <0.0001 |
| PE _{ROI} dif | Percentage difference in PE between ROI1 and ROI2 | 17.09 (5.81, 33.75) | 30.20 (9.35, 38.07) | 0.0892 |
| TTP _{ROI} dif | Percentage difference in TTP between ROI1 and ROI2 | 5.71 (0.92, 15.69) | 12.34 (7.02, 20.08) | 0.0673 |
| PE _{ROI} dif _{ch} | Percentage difference in change of PE between ROI1 and ROI2 | 1.59 (0.29, 10.15) | 47.01 (15.34, 100.99) | <0.0001 |
| TTP _{ROI} dif _{ch} | Percentage difference in change of TTP between ROI1 and ROI2 | 16.17 (11.06, 59.41) | 50.00 (21.41, 130.35) | 0.0577 |

A common feature of malignant tumors compared with non-malignant tumors is rapid wash-in (78, 79), which may be associated with shorter TTP. In our study, 7 days after therapy, the percentage change in TTP was greater in CR tumors, i.e., the time to peak increased significantly compared with the measurement before therapy, which may indicate that the tumors became “less malignant” with the treatment performed. These results are similar to those of human studies examining various chemotherapeutic antiangiogenic treatments, in which TTP and wash-in rate decreased after therapy (10, 17, 80). In a study of the efficacy of sorafenib treatment of metastatic renal cancer in humans, the correlation between treatment outcome and the percentage of perfused tissue decreased 3 weeks after treatment (15). In contrast, in studies evaluating DCE-US after radiotherapy for spontaneous tumors in dogs (18, 81), DCE-US results were not predictive of disease outcome. This difference can be explained by an important difference in the mechanism of action of radiotherapy compared to ECT combined with GET pIL-12. Radiotherapy is less efficient in regions of lower perfusion due to the resistance of hypoxic cells to treatment, whereas the presented EP-based treatment exerts antiangiogenic and cytotoxic effects that do not depend on cell oxygenation.

Perfusion heterogeneity is a hallmark of malignant tumors and provides valuable information for discriminating between malignant and benign lesions (82). We demonstrated a significant difference in perfusion heterogeneity between the two different groups based on the clinical response: tumors reaching CR were less heterogeneously perfused than non-CR tumors. This is an important finding as it indicates a possible predictive value of perfusion heterogeneity in therapies based on antiangiogenic effects. Similar results were obtained by DCE-CT in human hepatic neoplasia treated with antiangiogenic therapy; reduced perfusion heterogeneity correlated with better local tumor control and longer survival (83). In contrast, in human cervical cancer treated with radiotherapy and chemotherapy and evaluated with dynamic contrast-enhanced

magnetic resonance imaging (DCE-MRI), decreased perfusion heterogeneity correlated with poorer outcomes due to a lower response to radiotherapy in those parts of the tumor that are hypoxic due to decreased perfusion (82). Therefore, perfusion and perfusion heterogeneity evaluated with DCE-US appear to be useful for predicting the results of antiangiogenic treatments but cannot be used for all types of anticancer therapy.

Investigation of perfusion is an appealing method to predict outcome in tumors treated with antiangiogenic therapies. Our results show that DCE-US is safe for the patient; no adverse effects of contrast administration were noted despite repeated administration. Furthermore, it is a simple method to assess tumor perfusion that can be easily repeated during and after treatment and, based on our study, is associated with treatment outcome in canine tumors treated with a combination of ECT and GET pIL-12. The next clinically applicable step would be to investigate the predictive ability of DCE-US to distinguish between tumors that were successfully treated and those in which the therapy is unlikely to lead to a complete response after EP-based therapies. This information would allow a clinician to perform such a therapeutic procedure on a patient to evaluate if and when the therapy should be repeated, preferably in the early stages of the treatment when targeted therapy adjustments are generally more effective. This was not possible in our study given that a much larger number of patients are needed for logistic regression models. If a predictive value is to be confirmed, appropriate cutoff values for DCE-US parameters should be ascertained to make effective decisions about prognosis and the need for repeated or additional therapy in individual patients.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

ETHICS STATEMENT

The animal study was reviewed and approved by National Ethics Committee at the Administration of the Republic of Slovenia for Food Safety, Veterinary, and Plant Protection (U34401-24/2014/4). Written informed consent was obtained from the owners for the participation of their animals in this study.

AUTHOR CONTRIBUTIONS

DP, MC, GS, NT, and MB: conception and design. MB, NB, NM, and DP: acquisition of data. MB, NB, NM, TK, DP, and SKB: analysis and interpretation of data. MB, NB, DP, and SKB: drafting the article. GS, MC, NT, DP, and SKB: revising the article for intellectual content. All authors: final approval of the completed article.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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