## Check for updates

#### **OPEN ACCESS**

EDITED BY Mujeeb Ur Rehman, Livestock and Dairy Development Department, Pakistan

REVIEWED BY Basavaraj S. Mathapati, Indian Council of Medical Research (ICMR), India

\*CORRESPONDENCE Paolo Bonilauri paolo.bonilauri@izsler.it

<sup>†</sup>These authors have contributed equally to this work

#### SPECIALTY SECTION

This article was submitted to Veterinary Infectious Diseases, a section of the journal Frontiers in Veterinary Science

RECEIVED 31 October 2022 ACCEPTED 29 November 2022 PUBLISHED 15 December 2022

#### CITATION

Reggiani A, Rugna G and Bonilauri P (2022) SARS-CoV-2 and animals, a long story that doesn't have to end now: What we need to learn from the emergence of the Omicron variant. *Front. Vet. Sci.* 9:1085613. doi: 10.3389/fvets.2022.1085613

#### COPYRIGHT

© 2022 Reggiani, Rugna and Bonilauri. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or

reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# SARS-CoV-2 and animals, a long story that doesn't have to end now: What we need to learn from the emergence of the Omicron variant

## Alessandro Reggiani<sup>†</sup>, Gianluca Rugna<sup>†</sup> and Paolo Bonilauri<sup>\*†</sup>

Istituto Zooprofilattico Sperimentale della Lombardia e dell'Emilia Romagna, Brescia, Italy

OIE, the world organization for animal health, recently released an update on the state of the art of knowledge regarding SARS-CoV-2 in animals. For farmed animals, ferrets and minks were found to be highly susceptible to the virus and develop symptomatic disease both in natural conditions and in experimental infections. Lagomorphs of the species Oryctolagus cuniculus are indicated as highly susceptible to the virus under experimental conditions, but show no symptoms of the disease and do not transmit the virus between conspecifics, unlike raccoon dogs (Nyctereutes procyonoides), which in addition to being highly susceptible to the virus under experimental conditions, can also transmit the virus between conspecifics. Among felines, the circulation of the virus has reached a level of cases such as sometimes suggests the experimental use of vaccines for human use or treatments with monoclonal antibodies. But even among wild animals, several species (White-tailed deer, Egyptian rousettes, and minks) have now been described as potential natural reservoirs of the virus. This proven circulation of SARS-CoV-2 among animals has not been accompanied by the development of an adequate surveillance system that allows following the evolution of the virus among its natural hosts. This will be all the more relevant as the surveillance system in humans inevitably drops and we move to surveillance by sentinels similar to the human flu virus. The lesson that we can draw from the emergence of Omicron and, more than likely, its animal origin must not be lost, and in this mini-review, we explain why.

### KEYWORDS

SARS-CoV-2, wildlife, surveillance, diagnostic test, evolution, spillover, zoonosis

# 1. Introduction

The *Coronaviridae* is a well-known and studied family of viruses in veterinary medicine, and there are several farmed, wild, or pet animal species that are infected or have diseases from this family of viruses. Bats are the wild animals that host the majority of known coronaviruses, including the virus associated with severe respiratory syndrome (SARS), named SARS-CoV-2 (1, 2). Having known this, when SARS-CoV-2 emerged as a pandemic virus of zoonotic origin, an intensive research for a possible intermediate host of the virus began, research that is still incomplete today (3–5).

We are entering the 4th year of the pandemic and although the SARS-CoV-2 has substantially been sustained by human-to-human transmission, there are many factors, not all of which have been clarified, that determine the extent of transmission of the virus in the human population. Human-to-human transmission is strongly increased when the emergence of highly transmissible variants occurs, the latest being Omicron. The Omicron variant (B.1.1.529), first identified in November 2021, quickly spread around the world, causing another global epidemic, but its origin remains unclear (6). Genetic studies on Omicron BA.1 and BA.2 protein S indicate a high affinity for the mouse ACE2 receptor, while the ancestral SARS-CoV-2 S-trimer binds better to the cat receptor than to the mouse receptor, suggesting a possible human-cat-mousehuman evolutionary path for the emergence of Omicron BA.1 and BA.2 (7, 8).

Nevertheless, it is certain that the SARS-CoV-2 virus is able to infect and transmit among many animal species. There are four documented cases of animal to human transmission: the first well-documented case was in mink, where humanto-mink, mink-to-mink and back mink-to-human transmission of SARS-CoV-2 was observed (9). Another case in which the evidence supports the animal to human transmission is that which took place in Hong Kong between hamsters and humans (10). Although the exact mechanism by which white-tailed deer were infected by SARS-CoV-2 has not yet been clarified, human-to-deer transmission is now well-established, such as deer-to-deer transmission and reverse transmission from deer to human is strongly suspected (11, 12). The latest case is supported by genetic studies and involved a Thai vet and a cat in 2021. The vet is suspected of having been infected after being sneezed on by a cat that belonged to positive owners (dad and son). The genetic variant involved was B.1.167.2, Delta (13).

Current knowledge indicates that wildlife does not play a significant role in the spread of SARS-CoV-2 in humans, but the circulation of the virus in wild animal populations can affect the health of these populations and can facilitate the emergence of new variants of the virus. Furthermore, when we shift our attention away from pets, from those kept in captivity, such as zoos, and from farmed animals, it is much more difficult to track and control infections in animals. This is especially the case for viruses circulating in wild animals, whose role in the current pandemic has not yet been clarified, and it will take a long time and much research to do so.

# 2. SARS-CoV-2 evolution

As soon as the first genomic sequence of SARS-CoV-2 became available, it was analyzed to determine its nature and its putative evolutionary path. Despite its origin has not been completely understood, it has been demonstrated

that this virus descends from bat coronaviruses and could have been transmitted to humans via a still unknown intermediate host (14, 15). The main feature that has enabled SARS-CoV-2 to infect humans and spread around the world is its ability to bind the ACE2 receptor, which has been demonstrated to be an ancestral trait of sarbecoviruses, lost during the evolution of some clades, that is still evolvable and worth further attention (16). In contrast, the furin cleavage site in the spike protein, whose loss impairs viral pathogenicity (17), is a much rarer feature in other SARS-like viruses; its origin is still unclear, possibly obtained *via* recombination, and has raised debates about the natural origin of SARS-CoV-2 (18).

After a first wave of pandemic in early 2020, carried out by the original Wuhan strain, SARS-CoV-2 has begun to accumulate mutations, eventually leading to the generation of different linages, some of which have been declared as Variants Of Concern (VOC) by the WHO. Data from Nextstrain (https:// nextstrain.org/ncov/gisaid/global/6m) show how these VOC have all evolved independently from the original strain, rather than one from another (19). The main source of mutations is the RNA-dependent RNA polymerase (RdRp) encoded in the viral genome, which can incorporate mutations into the genomic template during replication. In the context of a highly transmissible virus that recently jumped to humans, this has led to a growing adaptation to the new host via the selection of more fit clades. Moreover, as the time passed, more people became immune due to previous infections and/or vaccinations, thus adding a selective pressure to viral evolution. Finally, the role of recombination events between different linages of SARS-CoV-2 or with other related CoVs should be taken into account.

The first VOC (PANGO lineage B.1.1.7) was detected in November 2020 in the United Kingdom and later named Alpha by the WHO, showing a high number of mutations gained in a relatively short time, giving rise to the hypothesis of it being generated by the chronic, prolonged infection of immunocompromised hosts (20). The main of these mutations in the spike protein are the N501Y and a deletion of aminoacids 69 and 70. In particular, N501Y could be related to an adaptation of the virus to mice hosts (21). In any case, even though a second wave was due to Alpha, no significant reduction in vaccine efficacy was linked to this variant (22). In October 2020 a new VOC (PANGO lineage B.1.617.2) was identified in India, named Delta, whose increased fitness allowed it to rapidly replace Alpha and other locally circulating variants (like Gamma and Mu) to generate a third global pandemic wave in 2021. Delta bore new mutations that enhanced its transmissibility by 40-60% compared to Alpha (23) and enabled immune escape from therapeutical mAbs, convalescent sera, and vaccination (24).

In November 2021, a fifth VOC (PANGO lineage B.1.1.529) named Omicron was detected in South Africa, then rapidly

spread around the world to give rise to a fourth pandemic wave between the end of 2021 and the beginning of 2022. Moreover, Omicron sublineages (identified, for nomenclature easiness reasons, with aliases such as BA.1 to BA.5) arose during 2022 until BA.5 became the most circulating variant globally in the past 6 months (https://nextstrain.org/ncov/gisaid/global/6m, accessed on 26/10/2022), also due to Omicron's shorter incubation periods and maybe milder clinical symptoms (25). Most of the sublineages known so far, whose description exceeds the purpose of this mini-review, are resumed and well-described in (26).

The main feature of the Omicron VOC is the higher number of mutations that it possesses, compared to the original Wuhan strain and to the previously circulating variants, initially harboring around 50 mutations, of which 32 are located in the spike protein (27). This rapid accumulation of mutations could be owed to two main different scenarios: first, the virus could have achieved a chronic infection in an immune-compromised patient, where it could generate variants without a potent cellular immune response and possibly under a positive selective pressure due to vaccination (28, 29); alternatively, SARS-CoV2 could have jumped back to some animal species and there, due to a different selective pressure, could have accumulated different mutations before getting to humans again. A mouse origin for Omicron has been proposed in a study that shows how pre-outbreak Omicron mutations are more consistent with an evolutionary history in mice rather than in human. This is also confirmed by an increased binding affinity with mouse ACE2 (30, 31). This adaptation could also be due to a different selective pressure exerted by some innate immunity intracellular effectors able to edit the viral genome, like APOBECs (32), ADAR (33), or ROS (34).

The hypothesis of a virus being able to infect and mutate in animal species has been confirmed in minks too. Su and colleagues (35) characterized some mutations in the spike protein of mink-derived viruses, such as Y453F and F486L, increasing the affinity for mink ACE2 while reducing that for human ACE2 (although maintaining infectivity). Interestingly, the circulation of these mink-adapted variants among humans dropped after mink culling policies were adopted (35). However, of all the mutations detected in minkderived viruses in the Netherlands and in Denmark, 18 were found to be prevalent in both countries, and those were similar to human-derived mutations (36). Thus, due to its ability to cross the species barrier and propagate in animals, a One-Health approach should be implemented to identify putative SARS-CoV-2 reservoirs.

# 3. Surveillance in wildlife

The need for monitoring animal species to detect novel SARS-CoV-2 variants has been discussed in the literature

as part of a One-Health approach (37). The likelihood of viral evolution and re-entry into the human population from wild animal reservoirs raises concerns about the infection of wildlife. Indeed, a SARS-CoV-2 sylvatic cycle, which may evade surveillance, could offer numerous chances for recurring spillback into human populations and other susceptible wild species (38). Furthermore, because animals harbor several endemic CoVs (39), recombination events capable of introducing in SARS-CoV-2 genomic traits of increasing fitness could not be ruled out, as was the case with SARS-CoV and MERS-CoV (40), aside from being the hypothetical mechanism of SARS-CoV-2 emergence (41).

Our understanding of the virus, as well as its epidemiology and transmission, is still evolving. Natural and experimental infections in wildlife species may drive implementation of risk-based surveillance systems for SARS-CoV-2 in wildlife, by restricting the range of animal species to be monitored (42).

## 3.1. Natural infections in wildlife

Although the contribution of wildlife to the emergence of SARS-CoV-2 is still unknown exactly (43), great concerns regarding the virus spilling-back and becoming endemic in animals have been expressed in light of several human-to-animal transmission events (Figure 1). To date, 735 SARS-CoV-2 animal events in 31 species were reported from 39 Countries in the SARS-ANI global dataset (44). SARS-CoV-2 has been demonstrated to have a wide host range, infecting a number of captive, farmed, and free-ranging wildlife species in addition to humans and domestic animals (45). Pigs and cattle have been found to be extremely insensitive to SARS-CoV-2 and poultry species are not at all susceptible to the virus (39). However, natural infections have been so far reported in a number of wild felids, in mustelids, e.g., Eurasian river otter (Lutra lutra), white-tailed deer (Odocoileus virginianus), and gorillas (46–52).

SARS-CoV-2 strains in wildlife generally mirrored the epidemiologic situation in humans; in early phase of pandemics animals were infected by ancestral strains, whereas in the latter stages diverse VOCs was detected, with Delta variant being the predominant (53).

In certain cases, human-to-animal transmission led to onward epizootic circulation of SARS-CoV-2, as in the case of farmed American minks reported from many European and American Countries (39, 54–56), and of white-tailed deer (*Odocoileus virginianus*) in North America (11, 12) (Figure 1). Transmission within animal populations may promote the origin of new variants as a consequence of evolutionary changes in the virus. Strain tracking by whole genome sequencing evidenced a mink-selected SARS-CoV-2 lineage, named Cluster 5, which has further spilled-back to humans



(46, 57). Additionally, the transmission of a highly divergent SARS-CoV-2 strain (B.1.641) from a white-tailed deer to humans was recently suspected in Canada (58).

## 3.2. Experimental infections in wildlife

Since the beginning of the pandemics, researchers have been investigating the susceptibility of various animal species to SARS-CoV-2 (38). Many studies focused on peridomestic wild species, such as carnivores, mustelids, rodents, and chiropters, which live in close proximity to humans and are thus at increased risk of virus exposure (59).

Among peridomestic wildlife, the striped skunk (*Mephitis* mephitis) and the raccoon (*Procyon lotor*), were experimentally infected with the SARS-CoV-2 strain USA-WA1/2020; all inoculated animals seroconverted, suggesting that both species are susceptible to SARS-CoV-2 infection. However, it appeared unlikely for SARS-CoV-2 to establish itself in wild raccoon and skunk populations due to the lack of virus isolation from raccoons and the low amount of virus shed by skunks (38). Rabbits of the species *Oryctolagus cuniculus* have been also demonstrated susceptible to experimental infection with SARS-CoV-2, developing an asymptomatic infection with an acute phase of viral shedding. Even though there is no evidence of a natural infection in wild lagomorphs, these findings call for more research (60). In contrast, experimentally infected raccoon dogs (*Nyctereus*  *procyonoides*) efficiently transmitted the virus to in-contact animals (61).

To date, two bat species, i.e., big brown bats (*Eptesicus fuscus*) and Egyptian fruit bats (*Rousettus aegyptiacus*), have been tested in the laboratory for virus susceptibility and shed capacity (62, 63). While one of the in-contact Egyptian fruit bats became infected after being exposed to inoculated animals, none of the experimentally infected big brown bats shed virus or transmitted it, nor did either species of bat develop clinical signs of disease. However, there are over 1,400 bat species worldwide and each species may respond differently to infection (64).

Research has shown a broad susceptibility of North American rodents to SARS-CoV-2 (65). Under experimental condition, deer mice of the species Peromyscus maniculatus nebrascensis and P. m. rufinus, could transmit the virus to naive conspecifics, implying that they could act as a wildlife reservoir for the virus (66). Rodent susceptibility to SARS-CoV-2 varies by species, which is not surprising given that rodents are the largest and most diverse order of mammals. Importantly, under experimental conditions, mice are not susceptible to early pandemic variants of SARS-CoV-2, but they are susceptible to certain VOCs such as B.1.1.7/Alpha, BA.1.1/Omicron, and especially B.1.351/Beta (67, 68). Recently, Omicron variant has been shown to infect minks, who can then effectively spread the virus to other conspecifics (69). Therefore, a continuous monitoring of newly emerged variants for their cross-species transmission is strongly recommended.

## 3.3. Diagnostic tests

Detecting viral RNA using a Real Time reverse transcription polymerase chain reaction (Real Time RT-PCR) test is currently the gold standard to detect an active SARS-CoV-2 infection (70). Several Real Time RT-PCR kits have been developed and approved, globally (71, 72). Reduction of false-negative as a result of SARS-CoV-2 rapid mutation, could be overcome by applying highly conserved region targets and multiple targets (73, 74). However, sensitivity of molecular tests is also influenced by pre-analytical factors, such as low viral loads, viral shedding time, and type of collected specimens. In addition, it should be considered the time of collection in passive surveillance, since sample degradation could occur between the death of the wild animal and sample collection.

Evaluating virus-specific antibodies is a rapid and affordable method to determine the status of the disease in individuals as well as carry out epidemiologic surveys in animal populations. The Spike (S)-protein and Nucleocapside (N)-protein are widely used in serological tests for SARS-CoV-2 infection. Due to the functional activity of the S-protein's receptor-binding domain (RBD), the level of anti-RBD antibody is generally regarded as a surrogate marker of the viral neutralizing potential of a serum. As a result, RBD-based serology assays are commonly used in human and veterinary clinical practice. Nevertheless, these tests are prone to decreased sensitivity because of the high rate of mutation for RBD epitopes, indicating the need of a timely updating based on the antigenic properties of the newly emerging SARS-CoV-2 variants (75). In contrast, N-based tests should be less affected because the N gene is the most conserved SARS-CoV-2 gene (76). However, comparison of two commercial N-based ELISAs and an RBD-based indirect ELISA, against virus neutralizing test (VNT), which is considered the gold standard for SARS-CoV-2 antibody detection, showed a lower sensitivity of both N-based assays (77).

Ideally, both serology and RT-PCR would be integrated into a surveillance program for SARS-CoV-2 in wildlife. Given the short duration of viral shedding, serologic investigations provide a more realistic picture of actual SARS-CoV-2 exposure in animals. However, detection of the virus allows for genome sequencing, which improves understanding of how SARS-CoV-2 evolves in animals (78, 79). Indeed, inter-species transmission and evolution in new hosts can also result in viral diversity, as seen during the human-mink-human transmission of SARS-CoV-2 (35) and the sustained transmission of the virus in the white-tailed deer populations (58).

# 4. Conclusion and proposals

Surveillance of zoonotic diseases in wildlife is challenging since it requires a lot of labor and is somewhat expensive, and there are often limited resources available. When adopting "One-Health" surveillance, the emergence of new diseases present opportunities for the development of novel surveillance techniques (80). To reduce costs or improve animal population coverage, it is crucial to evaluate novel sampling procedures, implement hazard-specific surveillance, and evaluate the efficacy of diagnostic tests, with the aim of avoiding loss of sensitivity even as new strains emerge, as it could come with SARS-CoV-2 (81–83).

A priority list of species groups for initial consideration can be created using the current evidence of natural and experimental infections. Additionally, the design of a wildlife surveillance system needs to be driven by a thorough understanding of animal demographics, social network structure, disease transmission paths, and a deep knowledge of the human-animal interface (84).

A large passive surveillance study on the circulation of the virus in wild animals is underway in northeastern Italy (Ministero della Salute, grant IZSLER PRC0062021), in which organs and fecal samples of numerous species will be analyzed. We expect to have enough coverage of animals and species analyzed by 2023 to provide the results to the scientific community.

However, we can already say that detecting the virus's circulation in these situations is particularly difficult, and as a result, efforts should be multiplied in various European regions and Countries in order to maximize the probability of detecting the virus's presence among wild animals and maximize our knowledge of a virus that will undoubtedly continue to circulate among human and animal populations indefinitely.

# Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

# Funding

This research was funded by the Ministry of Health, Italy (grant E55F21002740006 – IZSLER PRC2021006).

# Acknowledgments

The authors would like to thank all the One Health professionals who are working to fight this pandemic together.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

# References

1. Cui J, Li F, Shi ZL. Origin and evolution of pathogenic coronaviruses. *Nat Rev Microbiol.* (2019) 17:181–92. doi: 10.1038/s41579-018-0118-9

2. Fan Y, Zhao K, Shi ZL, Zhou P. Bat coronaviruses in China. Viruses. (2019) 11:210. doi: 10.3390/v11030210

3. Spyros L, Hughes J, Martin D, Swanepoel P, de Klerk A, Lourens R, et al. Exploring the natural origins of SARS-CoV-2 in the light of recombination. *Genome Biol Evol.* (2022) 14:evac018. doi: 10.1093/gbe/evac018

4. Cattaneo AM. Reviewing findings on the polypeptide sequence of the SARS-CoV-2 S-protein to discuss the origins of the virus. *Fut Virol.* (2022) 17:393–402. doi: 10.2217/fvl-2021-0233

5. Andersen KG, Rambaut A, Lipkin WI, Holmes EC, Garry RF. The proximal origin of SARS-CoV-2. *Nat Med.* (2020) 26:450–2. doi: 10.1038/s41591-020-0820-9

6. Xia S, Wang L, Zhu Y, Lu L, Jiang S. Origin, virological features, immune evasion and intervention of SARS-CoV-2 Omicron sublineages. *Signal Transduct Target Ther.* (2022) 7:1–7. doi: 10.1038/s41392-022-01105-9

7. Xu Y, Wu C, Cao X, Gu C, Liu H, Jiang M, et al. Structural and biochemical mechanism for increased infectivity and immune evasion of Omicron BA.2 variant compared to BA.1 and their possible mouse origins. *Cell Res.* (2022) 32:609–20. doi: 10.1038/s41422-022-00672-4

8. Liu Z, Lu L, Jiang S. Up or down: where comes Omincron? Cell Res. (2022) 22:1. doi: 10.1038/s41422-022-00675-1

9. Oude Munnink BB, Sikkema RS, Nieuwenhuijse DF, Molenaar RJ, Munger E, Molenkamp R, et al. Transmission of SARS-CoV-2 on mink farms between humans and mink and back to humans. *Science*. (2021) 371:172–7. doi: 10.1126/science.abe5901

10. Yen HL, Sit THC, Brackman CJ, Chuk SSY, Gu H, Tam KWS, et al. Transmission of SARS-CoV-2 delta variant (AY.127) from pet hamsters to humans, leading to onward human-to-human transmission: a case study. *Lancet.* (2022) 399:1070–8. doi: 10.1016/S0140-6736(22)00326-9

11. Hale VL, Dennis PM, McBride DS, Nolting JM, Madden C, Huey D, et al. SARS-CoV-2 infection in free-ranging white-tailed deer. *Nature*. (2022) 602:481–6. doi: 10.1038/s41586-021-04353-x

12. Kuchipudi SV, Surendran-Nair M, Ruden RM, Yon M, Nissly RH, Vandegrift KJ, et al. Multiple spillovers from humans and onward transmission of SARS-CoV-2 in white-tailed deer. *Proc Nat Acad Sci USA*. (2022) 119:e2121644119. doi: 10.1073/pnas.2121644119

13. Sila T, Sunghan J, Laochareonsuk W, Surasombatpattana S, Kongkamol C, Ingviya T, et al. Suspected cat-to-human transmission of SARS-CoV-2, Thailand, July-September 2021. *Emerg Infect Dis.* (2022) 28:1485–8. doi: 10.3201/eid2807.212605

14. Boni MF, Lemey P, Jiang X, Lam TT-Y, Perry BW, Castoe TA, et al. Evolutionary origins of the SARS-CoV-2 sarbecovirus lineage responsible for the COVID-19 pandemic. *Nature Microbiology.* (2020) 5:1408–17. doi: 10.1038/s41564-020-0771-4

15. Zhou P, Yang X-L, Wang X-G, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. (2020) 579:270–3. doi: 10.1038/s41586-020-2951-z

16. Starr TN, Zepeda SK, Walls AC, Greaney AJ, Alkhovsky S, Veesler D, et al. ACE2 binding is an ancestral and evolvable trait of sarbecoviruses. *Nature*. (2022) 603:913–8. doi: 10.1038/s41586-022-04464-z

17. Johnson B, Xie X, Bailey A, Kalveram B, Lokugamage K, Muruato A, et al. Loss of furin cleavage site attenuates SARS-CoV-2 pathogenesis. *Nature*. (2021) 591:4. doi: 10.1038/s41586-021-03237-4

18. Chan YA, Zhan SH. The emergence of the spike furin cleavage site in SARS-CoV-2. *Mol Biol Evol.* (2022) 39:msab327. doi: 10.1093/molbev/m sab327

19. Telenti A, Hodcroft EB, Robertson DL. The evolution and biology of SARS-CoV-2 variants. *Cold Spring Harb Perspect Med.* (2022) 12:a041390. doi: 10.1101/cshperspect.a041390

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

20. Corey L, Beyrer C, Cohen M, Michael N, Bedford T, Rolland M. SARS-CoV-2 variants in patients with immunosuppression. *N Engl J Med.* (2021) 385:NEJMsb2104756. doi: 10.1056/NEJMsb2104756

21. Gu H, Chen Q, Yang G, He L, Fan H, Deng Y-Q, et al. Adaptation of SARS-CoV-2 in BALB/c mice for testing vaccine efficacy. *Science*. (2020) 369:1603–7. doi: 10.1101/2020.05.02.073411

22. Muik A, Wallisch AK, Sänger B, Swanson KA, Mühl J, Chen W, et al. Neutralization of SARS-CoV-2 lineage B.1.1.7 pseudovirus by BNT162b2 vaccine-elicited human sera. *Science*. (2021) 371:1152–3. doi: 10.1126/science.abg6105

23. Kumar S, Thambiraja TS, Karuppanan K, Subramaniam G. Omicron and Delta variant of SARSARS-CoV-2: a comparative computational study of spike protein. *J Med Virol.* (2022) 94:1641–9. doi: 10.1002/jmv.27526

24. Planas D, Veyer D, Baidaliuk A, Staropoli I, Guivel-Benhassine F, Rajah MM, et al. Reduced sensitivity of SARS-CoV-2 variant Delta to antibody neutralization. *Nature.* (2021) 596:276–80. doi: 10.1038/s41586-021-03777-9

Jansen L, Tegomoh B, Lange K, Showalter K, Figliomeni J, Abdalhamid B, et al. Investigation of a SARS-CoV-2 B.1.1.529 (Omicron) Variant Cluster — Nebraska, November–December 2021. Morbid Mortal Weekly Rep. (2021) 70:1782–4. doi: 10.15585/mmwr.mm705152e3

26. Focosi D, McConnell S, Casadevall A. The Omicron variant of concern: diversification and convergent evolution in spike protein, and escape from anti-Spike monoclonal antibodies. *Drug Resist Updat.* (2022) 65:100882. doi: 10.1016/j.drup.2022.100882

27. Khandia R, Singhal S, Alqahtani T, Kamal MA, El-Shall NA, Nainu F, et al. Emergence of SARS-CoV-2 Omicron (B11529) variant, salient features, high global health concerns and strategies to counter it amid ongoing COVID-19 pandemic. *Environ Res.* (2022) 209:112816. doi: 10.1016/j.envres.2022.112816

28. Kupferschmidt K, Vogel G. How bad is Omicron? Some clues are emerging. *Science.* (2021) 374:1304–5. doi: 10.1126/science.acx9782

29. Li X. Omicron: call for updated vaccines. J Med Virol. (2022) 94:1261-3. doi: 10.1002/jmv.27530

30. Cameroni E, Bowen JE, Rosen LE, Saliba C, Zepeda SK, Culap K, et al. Broadly neutralizing antibodies overcome SARS-CoV-2 Omicron antigenic shift. *Nature*. (2022) 602:664–70. doi: 10.1038/s41586-021-04386-2

31. Wei C, Shan KJ, Wang W, Zhang S, Huan Q, Qian W. Evidence for a mouse origin of the SARS-CoV-2 Omicron variant. *J Genet Genomics*. (2021) 48:1111–21. doi: 10.1016/j.jgg.2021.12.003

32. Willems L, Gillet NA. APOBEC3 interference during replication of viral genomes. Viruses. (2015) 7:2999–3018. doi: 10.3390/v7062757

33. Pfaller CK, George CX, Samuel CE. Adenosine deaminases acting on RNA (ADARs) and viral infections. *Annu Rev Virol.* (2021) 8:239-64. doi: 10.1146/annurev-virology-091919-065320

34. Kong Q, Lin CL. Oxidative damage to RNA: mechanisms, consequences, and diseases. *Cell Mol Life Sci.* (2010) 67:1817–29. doi: 10.1007/s00018-010-0277-y

35. Su C, He J, Han P, Bai B, Li D, Cao J, et al. Molecular basis of mink ACE2 binding to SARS-CoV-2 and its mink-derived variants. *J Virol.* (2022) 96:e0081422. doi: 10.1128/jvi.00814-22

36. Khalid M, Alshishani A, Al-Ebini Y. Genome similarities between humanderived and mink-derived SARS-CoV-2 make mink a potential reservoir of the virus. *Vaccines*. (2022) 10:1352. doi: 10.3390/vaccines10081352

37. McDowell A. Pharmaceutics for free-ranging wildlife: case studies to illustrate considerations and future prospects. *Int J Pharm.* (2022) 628:122284. doi: 10.1016/j.ijpharm.2022.122284

38. Francisco R, Hernandez SM, Mead DG, Adcock KG, Burke SC, Nemeth NM, et al. Experimental susceptibility of North American Raccoons (*Procyon lotor*) and striped skunks (*Mephitis mephitis*) to SARS-CoV-2. *Front Vet Sci.* (2021) 8:715307. doi: 10.3389/fvets.2021.715307

39. Bonilauri P, Rugna G. Animal coronaviruses and SARS-CoV-2 in animals, what do we actually know? *Life*. (2021) 11:20123. doi: 10.3390/life11020123

40. Kozlakidis Z. Evidence for recombination as an evolutionary mechanism in coronaviruses: is SARS-CoV-2 an exception? *Front Public Health.* (2022) 2022:859900. doi: 10.3389/fpubh.2022.859900

41. Makarenkov V, Mazoure B, Rabusseau G, Legendre P. Horizontal gene transfer and recombination analysis of SARS-CoV-2 genes helps discover its close relatives and shed light on its origin. *BMC Ecol Evol.* (2021) 21:5. doi: 10.1186/s12862-020-01732-2

42. WHO. Joint Statement on the Prioritization of Monitoring SARS-CoV-2 Infection in Wildlife and Preventing the Formation of Animal Reservoirs. (2022). Available online at: https://www.woah.org/en/joint-statement-onthe-prioritization-of-monitoring-SARS-CoV-2-infection-in-wildlife-andpreventing-the-formation-of-animal-reservoirs/ (accessed October 30, 2022).

43. Jiang X, Wang R. Wildlife trade is likely the source of SARS-CoV-2. *Science*. (2022) 2022:add8384. doi: 10.1126/science.add8384

44. SARS-ANI VIS. A Global Open Access Dataset of Reported SARS-CoV-2 Events in Animals. (2022). Available online at: https://vis.csh.ac.at/sars-ani/ (accessed October 30, 2022).

45. Meekins DA, Gaudreault NN, Richt JA. Natural and experimental SARS-CoV-2 infection in domestic and wild animals. *Viruses.* (2021) 13:101993. doi: 10.3390/v13101993

46. Munnink BBO, Sikkema RS, Nieuwenhuijse DF, Molenaar RJ, Munger E, Molenkamp R, et al. Transmission of SARS-CoV-2 on mink farms between humans and mink and back to humans. *Science*. (2021) 371:172–7. doi: 10.1126/science.abe590

47. McAloose D, Laverack M, Wang L, Killian ML, Caserta LC, Yuan F, et al. From people to panthera: natural SARS-CoV-2 infection in tigers and lions at the Bronx Zoo. *MBio.* (2020) 22:231959. doi: 10.1101/2020.07.22.213959

48. Nagy A, Stará M, Vodička R, Černíková L, Jiřincová H, Křivda V, et al. Reverse-zoonotic transmission of SARS-CoV-2 lineage alpha (B117) to great apes and exotic felids in a zoo in the Czech Republic. *Arch Virol.* (2022) 167:1681–5. doi: 10.1007/s00705-022-05469-9

49. Gortázar C, Barroso-Arévalo S, Ferreras-Colino E, Isla J, de la Fuente G, Rivera B, et al. Natural SARS-CoV-2 infection in Kept Ferrets, Spain. *Emerg Infect Dis.* (2021) 27:1994–6. doi: 10.3201/eid2707.210096

50. Shriner SA, Ellis JW, Root JJ, Roug A, Stopak SR, Wiscomb GW, et al. SARS-CoV-2 exposure in Escaped Mink, Utah, USA. *Emerg Infect Dis.* (2021) 27:988–90. doi: 10.3201/eid2703.204444

51. Aguiló-Gisbert J, Padilla-Blanco M, Lizana V, Maiques E, Muñoz-Baquero M, Chillida-Martínez E, et al. First description of SARS-CoV-2 infection in two feral American Mink (Neovison vison) caught in the wild. *Animals.* (2021) 11:51422. doi: 10.3390/ani11051422

52. Padilla-Blanco M, Aguiló-Gisbert J, Rubio V, Lizana V, Chillida-Martínez EV, Cardells J, et al. The finding of the severe acute respiratory syndrome coronavirus (SARS-CoV-2) in a Wild Eurasian River Otter (*Lutra lutra*) highlights the need for viral surveillance in wild mustelids. *Front Vet Sci.* (2022) 9:826991. doi: 10.3389/fvets.2022.826991

53. Cui S, Liu Y, Zhao J, Peng X, Lu G, Shi W, et al. An updated review on SARS-CoV-2 infection in animals. *Viruses.* (2022) 14:71527. doi: 10.3390/v14071527

54. Moreno A, Lelli D, Trogu T, Lavazza A, Barbieri I, Boniotti M, et al. SARS-CoV-2 in a mink farm in Italy: case description, molecular and serological diagnosis by comparing different tests. *Viruses*. (2022) 14:81738. doi: 10.3390/v14081738

55. Badiola JJ, Otero A, Sevilla E, Marín B, García Martínez M, Betancor M, et al. SARS-CoV-2 outbreak on a Spanish mink farm: epidemiological, molecular, and pathological studies. *Front Vet Sci.* (2022) 8:5004. doi: 10.3389/fvets.2021.805004

56. Chaintoutis SC, Thomou Z, Mouchtaropoulou E, Tsiolas G, Chassalevris T, Stylianaki I, et al. Outbreaks of SARS-CoV-2 in naturally infected mink farms: impact, transmission dynamics, genetic patterns, and environmental contamination. *PLoS Pathog.* (2021) 17:e1009883. doi: 10.1371/journal.ppat.1009883

57. Burkholz S, Pokhrel S, Kraemer BR, Mochly-Rosen D, Carback RT, Hodge T, et al. Paired SARS-CoV-2 spike protein mutations observed during ongoing SARS-CoV-2 viral transfer from humans to minks and back to humans. *Infect Genet Evol.* (2021) 93:104897. doi: 10.1016/j.meegid.2021.104897

58. Pickering B, Lung O, Maguire F, Kruczkiewicz P, Kotwa JD, Buchanan T, et al. Divergent SARS-CoV-2 variant emerges in white-tailed deer with deerto-human transmission. *Nat Microbiol.* (2022) 2022:9. doi: 10.1038/s41564-022-01268-9

59. Bosco-Lauth AM, Root JJ, Porter SM, Walker AE, Guilbert L, Hawvermale D, et al. Peridomestic mammal susceptibility to severe acute respiratory syndrome coronavirus 2 infection. *Emerg Infect Dis.* (2021) 27:2073–80. doi: 10.3201/eid2708.210180

60. Freuling CM, Breithaupt A, Müller T, Sehl J, Balkema-Buschmann A, Rissmann M, et al. Susceptibility of raccoon dogs for experimental SARS-CoV-2 infection. *Emerg Infect Dis J.* (2020) 26:203733. doi: 10.3201/eid2612. 203733

61. Mykytyn AZ, Lamers MM, Okba NMA, Breugem TI, Schipper D, van den Doel PB, et al. Susceptibility of rabbits to SARS-CoV-2. *Emerg Microbes Infect.* (2021) 10:1–7. doi: 10.1080/22221751.2020.1868951

62. Hall JS, Knowles S, Nashold SW, Ip HS, Leon AE, Rocke T, et al. Experimental challenge of a North American bat species, big brown bat (*Eptesicus fuscus*), with SARS-CoV-2. *Transbound Emerg Dis.* (2021) 68:3443–52. doi: 10.1111/tbed.13949

63. Schlottau K, Rissmann M, Graaf A, Schön J, Sehl J, Wylezich C, et al. SARS-CoV-2 in fruit bats, ferrets, pigs, and chickens: an experimental transmission study. *Lancet Microbe.* (2020) 1:e218–25. doi: 10.1016/S2666-5247(20)30089-6

64. Bosco-Lauth AM, Porter SM, Fox KA, Wood ME, Neubaum D, Quilici M. Experimental infection of Brazilian free-tailed bats (*Tadarida brasiliensis*) with two strains of SARS-CoV-2. *Viruses.* (2022) 14:81809. doi: 10.3390/v14081809

65. Lewis J, Zhan S, Vilander AC, Fagre AC, Kiaris H, Schountz T. SARS-CoV-2 infects multiple species of North American deer mice and causes clinical disease in the California mouse. *BioRxiv*. (2022) 2022:504888. doi: 10.1101/2022.08.22.504888

66. Griffin BD, Chan M, Tailor N, Mendoza EJ, Leung A, Warner BM, et al. SARS-CoV-2 infection and transmission in the North American deer mouse. *Nat Commun.* (2021) 12:3612. doi: 10.1038/s41467-021-23848-9

67. Chen Q, Huang X, Liu YY, Sun M, Ji B, Zhou C, et al. Comparative characterization of SARS-CoV-2 variants of concern and mouse-adapted strains in mice. *J Med Virol.* (2022) 94:3223. doi: 10.1002/jmv.27735

68. Montagutelli X, Prot M, Levillayer L, Salazar EB, Jouvion G, Conquet L, et al. Variants with the N501Y mutation extend SARS-CoV-2 host range to mice, with contact transmission. *BioRxiv.* (2021) 2021:436013. doi: 10.1101/2021.03.18.436013

69. Virtanen J, Aaltonen K, Kegler K, Venkat V, Niamsap T, Kareinen L, et al. Experimental infection of mink with SARS-CoV-2 Omicron variant and subsequent clinical disease. *Emerg Infect Dis J.* (2022) 28:220328. doi: 10.3201/eid2806.220328

70. Bentley E, Mee ET, Routley S, Mate R, Fritzsche M, Hurley M, et al. Collaborative study for the establishment of a WHO international standard for SARS-CoV-2 RNA. Geneva: WHO/BS20202402 (2020).

71. European Commission. COVID-19 in vitro Diagnostic Devices and Test Methods Database. Available online at: https://covid-19-diagnostics.jrc.ec.europa. eu/ (accessed October 30, 2022).

72. U.S. Food and Drug Administration. In vitro Diagnostics EUAs - Antigen Diagnostic Tests for SARS-CoV-2. (2022). Available online at: http://www.fda.gov/medical-devices/coronavirus-disease-2019-covid-19-emergency-use-authorizations-medical-devices/in-vitro-diagnostics-euas-antigen-diagnostic-tests-SARS-CoV-2 (accessed October 30, 2022).

73. Asghar R, Rasheed M, Ul Hassan J, Rafique M, Khan M, Deng Y. Advancements in testing strategies for COVID-19. *Biosensors*. (2022) 12:60410. doi: 10.3390/bios12060410

74. Tahamtan A, Ardebili A. Real-time RT-PCR in COVID-19 detection: issues affecting the results. *Expert Rev Mol Diagn.* (2020) 20:453-4. doi:10.1080/14737159.2020.1757437

75. Rak A, Donina S, Zabrodskaya Y, Rudenko L, Isakova-Sivak I. Crossreactivity of SARS-CoV-2 nucleocapsid-binding antibodies and its implication for COVID-19 serology tests. *Viruses.* (2022) 14:92041. doi: 10.3390/v140 92041

76. Dutta NK, Mazumdar K, Gordy JT. The nucleocapsid protein of SARS-CoV-2: a target for vaccine development. *J Virol.* (2020) 94:20. doi: 10.1128/JVI.00647-20

77. Ratti G, Lelli D, Moreno A, Stranieri A, Trogu T, Giordano A, et al. Comparison of diagnostic performances of different serological tests for SARS-CoV-2 antibody detection in cats and dogs. *Transbound Emerg Dis.* (2022) 2022;14716. doi: 10.1111/tbed.14716

78. Tan CCS, Lam SD, Richard D, Owen CJ, Berchtold D, Orengo C, et al. Transmission of SARS-CoV-2 from humans to animals and potential host adaptation. *Nat Commun.* (2022) 13:2988. doi: 10.1038/s41467-022-30698-6

79. Magateshvaren Saras MA, Patro LPP, Uttamrao PP, Rathinavelan T. Geographical distribution of SARS-CoV-2 amino acids mutations and the concomitant evolution of seven distinct clades in non-human hosts. *Zoonoses Public Health.* (2022) 69:816–25. doi: 10.1111/zph.12971

80. dos S, Ribeiro C, van de Burgwal, Linda HM, Regeer BJ. Overcoming challenges for designing and implementing the One Health approach: a systematic review of the literature. One Health. (2019) 7:100085. doi: 10.1016/j.onehlt.2019.100085

81. Rodríguez-Prieto V, Vicente-Rubiano M, Sánchez-Matamoros A, Rubio-Guerri C, Melero M, Martínez-López B, et al. Systematic review of surveillance systems and methods for early detection of exotic, new and re-emerging diseases in animal populations. *Epidemiol Infect*. (2015) 143:2018–42. doi: 10.1017/S095026881400212X

82. Del Vecchio C, Cracknell Daniels B, Brancaccio G, Brazzale AR, Lavezzo E, Ciavarella C, et al. Impact of antigen test target failure and

testing strategies on the transmission of SARS-CoV-2 variants. Nat Commun. (2022) 13:1–16. doi: 10.1038/s41467-022-33460-0

83. WOAH. Considerations for Sampling, Testing, and Reporting of SARS-CoV-2 in Animals. (2020). Available online at: http://rr-asia.woah.org/wp-content/ uploads/2020/05/sampling\_testing\_and\_reporting\_of\_SARS-CoV-2\_in\_animals\_ 7may\_2020.pdf (accessed October 30, 2022).

84. Delahay RJ, De La Fuente J, Smith GC, Sharun K, Snary EL, Flores Girón L, et al. Assessing the risks of SARS-CoV-2 in wildlife. *One Health Outlook.* (2021) 3:6. doi: 10.1186/s42522-021-00039-6