



# Serotype Diversity and Antimicrobial Resistance Profile of *Salmonella enterica* Isolates From Freshwater Turtles Sold for Human Consumption in Wet Markets in Hong Kong

Violaine Albane Colon<sup>1</sup>, Kittitat Lugsomya<sup>1</sup>, Hoi Kiu Lam<sup>1</sup>, Lloyd Christian Wahl<sup>2</sup>, Rebecca Sarah Victoria Parkes<sup>3,4</sup>, Catherine Anne Cormack<sup>5</sup>, Jule Anna Horlbog<sup>6,7</sup>, Marc Stevens<sup>6</sup>, Roger Stephan<sup>6</sup> and Ioannis Magouras<sup>1,2\*</sup>

## OPEN ACCESS

### Edited by:

Min Yue,  
Zhejiang University, China

### Reviewed by:

Yujie Hu,  
China National Center for Food Safety  
Risk Assessment, China  
Xiaowu Jiang,  
Yichun University, China  
Ali Raza Jahejo,  
Shanxi Agricultural University, China

### \*Correspondence:

Ioannis Magouras  
ioannis.magouras@cityu.edu.hk

### Specialty section:

This article was submitted to  
Veterinary Infectious Diseases,  
a section of the journal  
Frontiers in Veterinary Science

Received: 04 April 2022

Accepted: 20 June 2022

Published: 22 July 2022

### Citation:

Colon VA, Lugsomya K, Lam HK, Wahl LC, Parkes RSV, Cormack CA, Horlbog JA, Stevens M, Stephan R and Magouras I (2022) Serotype Diversity and Antimicrobial Resistance Profile of *Salmonella enterica* Isolates From Freshwater Turtles Sold for Human Consumption in Wet Markets in Hong Kong. *Front. Vet. Sci.* 9:912693. doi: 10.3389/fvets.2022.912693

<sup>1</sup> Department of Infectious Diseases and Public Health, Jockey Club College of Veterinary Medicine and Life Sciences, City University of Hong Kong, Kowloon, Hong Kong SAR, China, <sup>2</sup> Centre for Applied One Health Research and Policy Advice, Jockey Club College of Veterinary Medicine and Life Sciences, City University of Hong Kong, Kowloon, Hong Kong SAR, China, <sup>3</sup> Department of Veterinary Clinical Sciences, Jockey Club College of Veterinary Medicine and Life Sciences, City University of Hong Kong, Kowloon, Hong Kong SAR, China, <sup>4</sup> Centre for Animal Health and Welfare, Jockey Club College of Veterinary Medicine and Life Sciences, City University of Hong Kong, Kowloon, Hong Kong SAR, China, <sup>5</sup> Jockey Club College of Veterinary Medicine and Life Sciences, City University of Hong Kong, Kowloon, Hong Kong SAR, China, <sup>6</sup> Institute for Food Safety and Hygiene, Vetsuisse Faculty, University of Zurich, Zürich, Switzerland, <sup>7</sup> Vetsuisse Faculty, National Reference Center for Enteropathogenic Bacteria and Listeria (NENT), Institute for Food Safety and Hygiene, University of Zurich, Zürich, Switzerland

Chelonians are recognized as a source of human salmonellosis through direct contact or consumption of their meat. Freshwater turtles sold for food are widely available in wet markets in Asia. In this pilot study, 50 turtles belonging to three species were randomly sampled from wet markets throughout Hong Kong. The turtles were humanely euthanised and their feces or the colon were sampled for *Salmonella* culture. The *Salmonella* isolates obtained were serotyped and examined for phenotypic antimicrobial resistance and the presence of antimicrobial resistance genes. The study reports a high prevalence (42%, 95% CI: 29.4–55.8) and considerable serotype diversity of *Salmonella* among turtles sold in wet markets. The most common among the 11 serotypes isolated were S. Oranienburg and S. Thompson, which have been reported in turtles previously. The serotype S. Manhattan is reported in chelonians for the first time. Resistance to streptomycin and chloramphenicol was common, despite the latter being banned from aquaculture in mainland China since 2002. Resistance against fluoroquinolones and third-generation cephalosporins which represent first-line treatment options for salmonellosis was also observed. The multidrug-resistance gene *cfr* is identified for the first time in *Salmonella*. This is a worrying finding as it indicates an expansion of the *cfr* reservoir and potential horizontal spread to other bacteria. The results of this study emphasize the need for close surveillance of *Salmonella* from turtles sold as food and better regulation of turtle farming to safeguard public health and improve animal welfare.

**Keywords:** *Salmonella*, turtles, antimicrobial resistance, *cfr* gene, wet markets, zoonoses, Hong Kong

## INTRODUCTION

Freshwater turtles are widely available in wet markets in South East Asia and Hong Kong and are primarily sold for consumption (1). A survey of 950,251 turtles for sale at wholesale and retail outlets in Hong Kong and Guangdong Province between 2000 and 2003 revealed that 77 different species, including endangered and critically endangered species, were sold (2). In Hong Kong, turtles are mostly imported from farms in Asian countries and mainland China, where large-scale turtle farming is estimated to be a multi-billion-dollar industry (1, 3). A study of 684 turtle farms in mainland China showed that ~127 million turtles across 11 different species are sold each year, of which the most common is the Chinese softshell turtle (SS) (*Pelodiscus sinensis*), accounting for over 97.6% of turtles sold (3).

There are numerous biological risks associated with the consumption of reptile products, including infections caused by bacteria, parasites, and exposure to biotoxins (4). *Salmonellae* are part of the normal intestinal flora of turtles, and turtle-associated salmonellosis has been a recognized public health issue for several decades (5, 6). While a high proportion of these infections is attributed to contact of young children with pet turtles (6, 7), salmonellosis has also been linked to the consumption of green turtles (*Chelonia mydas*) in Australia (8, 9) and snapping turtles (*Chelydra serpentina*) in Japan (10). Of the 2,659 *Salmonella* (*S.*) serotypes (11), seven have been implicated in reptile-associated salmonellosis in humans. These include *S. Paratyphi B* var Java, *S. Poona*, *S. Pomona*, *S. Marina*, *S. Stanley*, *S. Litchfield*, and *S. Newport*, and the most commonly reported *S. Typhimurium* and *S. Enteritidis* (6, 7).

*Salmonellae* are estimated to cause 93.8 million cases of gastroenteritis and 155,000 deaths globally each year (12). In Hong Kong, salmonellosis was the second most common bacterial cause of food poisoning from 2003 to 2011, with 3,250 cases (13). Although no cases of salmonellosis associated with turtle meat consumption in Hong Kong are published to date, close surveillance of *Salmonella* from all potential sources is essential to safeguard public health and for the timely detection of emerging serotypes. Furthermore, the ongoing spread of antimicrobial resistance (AMR) and the risk of dissemination of AMR genes (ARGs) in the population represents an additional challenge associated with *Salmonella* infections (14). Infections with resistant *Salmonella* are harder to treat and cause increased morbidity and mortality rates (15, 16).

There is limited information available on turtle farming practices, including antimicrobial usage, despite the large number of turtles farmed and consumed in Asia. Data on AMR in *Salmonella* isolated from turtles destined for human consumption is also sparse, as most studies have focused on wild turtles or captive turtles raised for the pet industry. One study from wet markets in Shanghai reported that most isolates (84%) were resistant to multiple antimicrobials (>3) (17). Other studies have looked at AMR in *Salmonella* from captive populations of freshwater turtles (18, 19). Red-eared sliders (RES) (*Trachemys scripta elegans*) sold in pet shops carried *Salmonella* with resistance against tetracycline, gentamycin, kanamycin, streptomycin, and sulfamethoxazole/trimethoprim (19).

To the author's knowledge, no previous studies have characterized *Salmonella* in turtles sold for human consumption in Hong Kong and research from other regions in Asia is scarce. The objectives of this pilot study were (1) to describe the frequency and serotypes of *Salmonella* in freshwater turtles sold in wet markets for human consumption, and (2) to characterize the AMR profile of the *Salmonella* isolates using phenotypic and molecular approaches. A better understanding of the zoonotic risks from turtle meat will provide a basis for improving consumer's and other stakeholder's awareness. Furthermore, it will stimulate discussions on developing clear guidelines on turtle farming and sale that could benefit animal welfare and safeguard public health.

## MATERIALS AND METHODS

A list of wet markets in Hong Kong was made using publicly available information from the website of the Food and Environmental Hygiene Department (20). Due to the lack of information on live turtle availability and trade characteristics in wet markets, all 94 wet markets were visited twice weekly over a period of 3 months, and those selling live turtles were recorded. Three turtle species were available, namely SS, RES and Chinese striped neck turtles (CSN, *Mauremys sinensis*). From the final list of 28 wet markets where live turtles were available, 21 wet markets were randomly selected, and 50 freshwater turtles were sampled twice between January and March 2021. The wet markets sampled were distributed across all districts of Kowloon (9 wet markets from 5 districts) and Hong Kong island (6 wet markets from 4 districts) and 6 out of 9 districts in the New Territories (7 wet markets). The country of origin and whether the turtles were wild-caught or farmed was recorded.

A physical examination was performed on each turtle to record the general health condition and any obvious external lesions or abnormalities. Physical examinations were performed by a board-certified reptile veterinarian. Data including sex, weight and age group were collected. The turtles were anesthetized by intravenous injections of alfaxalone (Alfaxan<sup>®</sup>, Jurox Pty Limited, Rutherford, NSW 2320, Australia) at a dose of 10–20 mg/kg. Once anesthesia was confirmed, euthanasia was induced by an intravenous injection of pentobarbital (Dorminal 20%, Alfasan, 3449 JA Worden, The Netherlands) at a dose of 100 mg/kg, following the American Veterinary Medical Association guidelines (21). Fecal samples, if present, were collected during post-mortem examination and placed into sterile tubes with Amies agar gel transport swab (Thermo Fisher Scientific Australia Pty Ltd., Melbourne, Australia). If no feces were present, the colon area was swabbed, and the swabs were stored in a similar manner to the feces. The turtles were collected over 22 days and samples were processed the same day the turtles were bought.

### *Salmonella* Isolation and Identification

Each sample was placed in 10 ml buffered peptone water (BPW; Thermo Fisher Scientific Australia Pty Ltd., Melbourne, Australia) and incubated at 37°C for 18 h. Following incubation,

0.1 ml of cultured BPW was used to inoculate 10 ml Rappaport-Vassiliadis Soya Peptone broth (RVS; bioMérieux, Marcy-l'Étoile, France) at 41.5°C for 24 h. Cultured RVS was streaked on Xylose Lysine Deoxycholate agar (XLD; Thermo Fisher Scientific Australia Pty Ltd., Melbourne, Australia) and incubated at 37°C for 24 h (22). *Salmonella Typhimurium* (ATCC:14028<sup>TM</sup>) was used as positive control. Putative *Salmonella* colonies (black color) were selected for species identification by Matrix-Assisted Laser Desorption/Ionization Time-Of-Flight (MALDI-TOF; Bruker, Massachusetts, US) mass spectrometry and analyzed by MALDI Biotyper<sup>®</sup> (Bruker, Massachusetts, US).

## Salmonella Serotyping

The obtained *Salmonella* strains were initially serotyped at the National Center for Enteropathogenic Bacteria and Listeria (NENT) at the University of Zurich, Switzerland. Typing was performed by slide agglutination with commercially available antisera (Sifin Diagnostics GmbH, Berlin, Germany) according to the Kauffmann-White-Le Minor scheme.

## Antimicrobial Susceptibility Testing

Antimicrobial susceptibility testing against 15 antimicrobials was performed by Kirby-Bauer disk diffusion test. The antimicrobials included ampicillin; AMP (10 µg), cefotaxime; CTX (30 µg), ceftazidime; CAZ (30 µg), meropenem; MEM (10 µg), imipenem; IPM (10 µg), ertapenem; ETP (10 µg), ciprofloxacin; CIP (5 µg), streptomycin; S (10 µg), gentamicin; GEN (10 µg), amikacin; AMK (30 µg), sulfamethoxazole-trimethoprim; SXT (23.75 µg/1.25 µg), doripenem; DOR (10 µg), chloramphenicol; CHL (30 µg), azithromycin; AZM (15 µg) (Thermo Fisher Scientific Australia Pty Ltd., Melbourne, Australia). A colistin (CST) (0.5–32 µg/ml) (MilliporeSigma, Massachusetts, US) susceptibility test was performed using the broth microdilution test. The zone of inhibition and minimal inhibitory concentration were interpreted using the clinical breakpoints published by the Clinical and Laboratory Standards Institute (CLSI), M100 31st edition (23). Extended-Spectrum β-lactamases confirmatory test was performed by Combination Disk Test (23). Linezolid (LZD) and chloramphenicol (CHL) MIC testing for the isolated, *cfr* carrying *S. IIIb* 50:k:z was performed using Etest<sup>®</sup> strips (bioMérieux, Marcy-l'Étoile, France).

## Whole Genome Sequencing

Whole genome sequencing was used for final confirmation and was performed as described previously (24). Briefly, paired-end libraries were produced and sequenced on an Illumina MiniSeq sequencer (Illumina, San Diego, CA, USA). Reads were assembled using Spades 3.13.1 (25) in Shovill 1.0.4 (<https://github.com/tseemann/shovill>). Whole genome-based *Salmonella* Serotyping was performed using SeqSero (26) with standard settings. Antimicrobial resistance gene analysis was done using the NCBI AMRFinderPlus database (27) in Ridom SeqSphere v7.7.5 (Ridom GmbH, Münster, Germany) using standard settings.

**TABLE 1** | The total number and proportion of turtles positive for *Salmonella enterica* by species, origin and sex.

Species	N	% positive samples (n)	95% CI
CSN	8	87.5 (7)	52.9–97.8
RES	11	72.7 (8)	43.4–90.3
SS	31	19.4 (6)	9.2–36.3
<b>Origin</b>			
Farmed	35	51.4 (18)	35.6–67.0
Wild	15	20.0 (3)	7.0–45.2
<b>Sex</b>			
Female	19	78.9 (15)	56.7–91.5
Male	31	9.4 (6)	9.2–36.3
<b>Total</b>	50	42.0 (21)	29.4–55.8

CSN, Chinese stripe-necked turtle (*Mauremys sinensis*); RES, Red-eared slider (*Trachemys scripta elegans*); SS, Chinese softshell turtle (*Pelodiscus sinensis*).

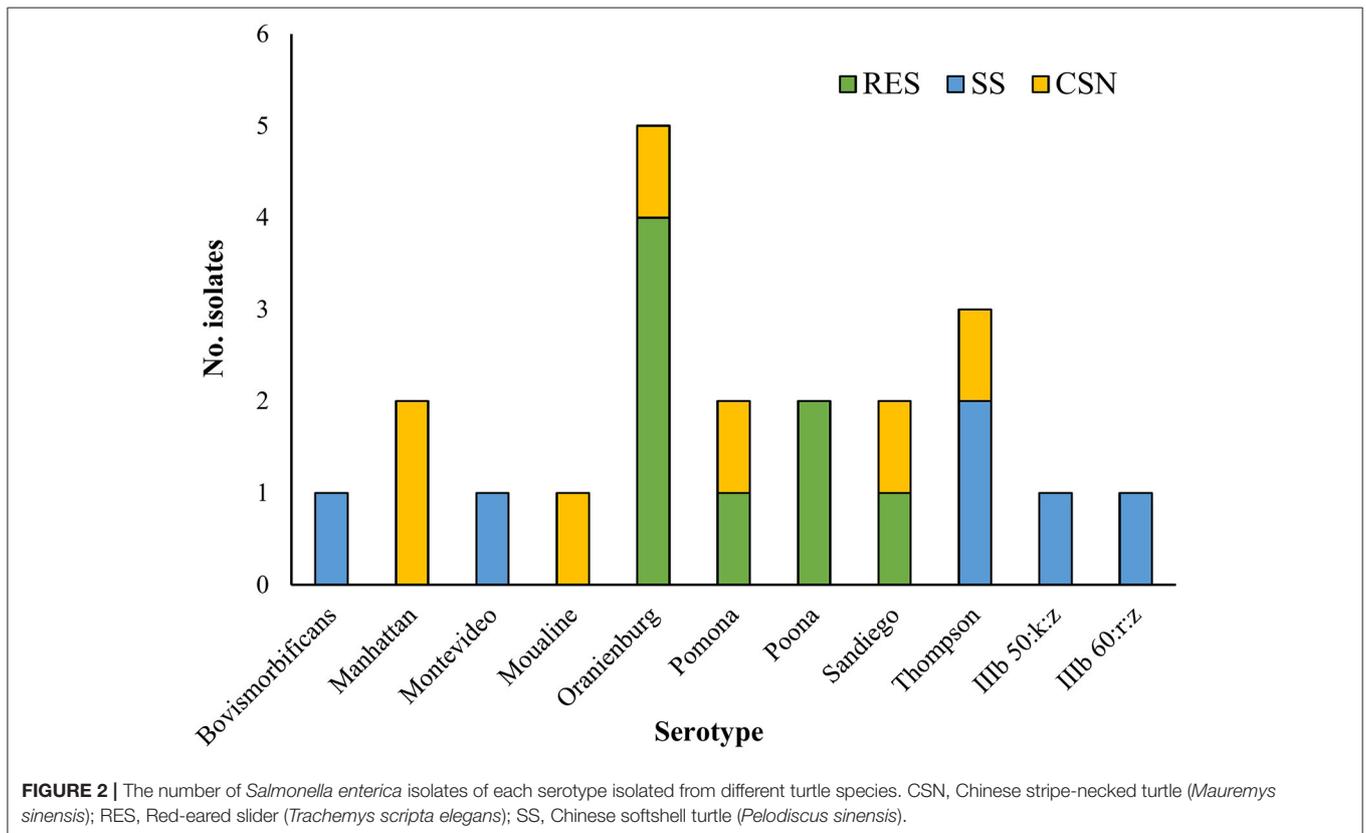
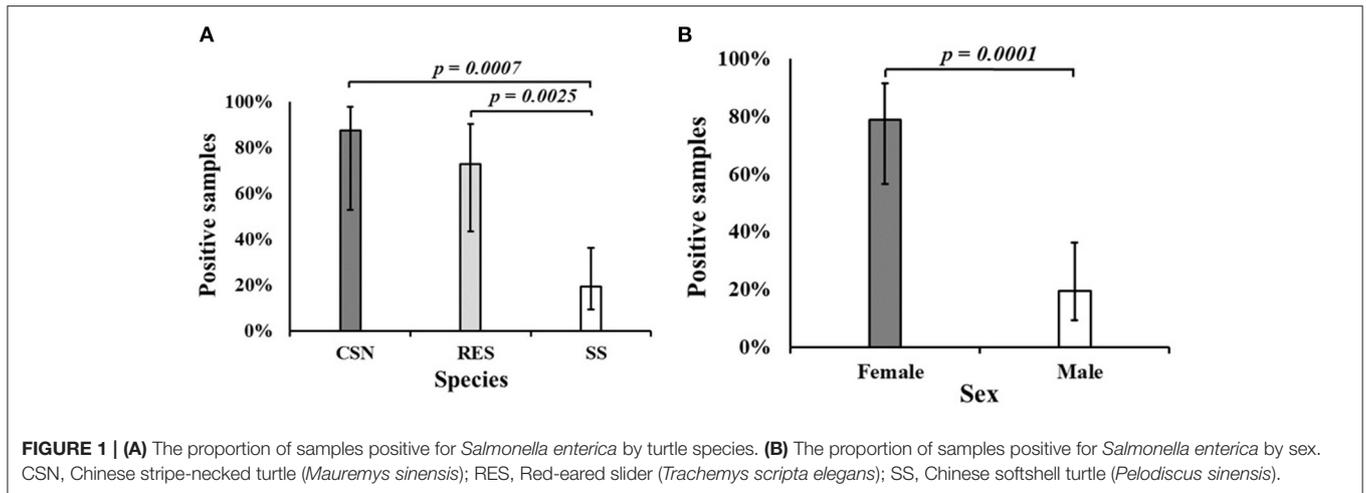
## Statistical Analysis

Collection of datapoints and descriptive statistics were performed on an electronic spreadsheet (Excel, Microsoft Corp, Redmond, Wash). Association between pairs of variables was assessed by Fisher's exact test of independence using the online statistical tool Stangroom (28). Confidence intervals were calculated using the Wilson score method using the online calculator Eptools (29).

## RESULTS

A total of 50 turtles were randomly sampled from 21 wet markets. The turtle species distribution was 16% (8/50) CSN, 22% (11/50) RES and 62% (31/50) SS (**Table 1**). Thirty-one turtles were male (62%) and all belonged to the species SS, and 19 turtles were female (38%) and were either CSN or RES. According to the information provided by the vendor, 30% (15/50) of turtles were wild-caught and 70% (35/50) were farmed. Thirteen of the wild turtles were SS and 2 were CSN. All farmed turtles originated from mainland China except for two turtles imported from Thailand. Four turtles were classified as juvenile (one CSN and three RES), while the rest were adult individuals. The mean weight of the turtles was 0.94 kg (median 0.92 kg, range 0.61–1.64 kg). The mean weight for RES was 0.84 kg, for SS it was 0.99 kg and for CSN was 0.81 kg. Feces was present in 68% (34/50) of the turtles (four CSN, 10 RES and 20 SS). The remaining animals (16/50) were sampled using swabs.

*Salmonella enterica* was isolated from 21 turtles (42%), including 87.5% of CSN (7/8), 72.7% of RES (8/11) and 19.4% of SS (6/31). A statistically significant difference ( $p < 0.001$ ) was found between the proportion of positive SS and both CSN and RES (**Figure 1A**). A significant difference ( $p < 0.001$ ) was found between the proportion of positive male (19.4%) and positive female turtles (78.9%) (**Figure 1B**). Feces were present in 34 turtles and 50.0% of these were positive for *S. enterica* (17/34) while 25.0% of turtles without feces were positive for *S. enterica* (4/16) although no significant difference was found ( $p = 0.129$ ). A greater proportion of farmed turtles (50.4%) were positive for



*S. enterica* compared to wild-caught (20%) samples although no significant difference was found ( $p = 0.061$ ).

Two isolates belonged to *S. enterica* subsp. *diarizonae* and the remaining nineteen isolates belonged to *S. enterica* subsp. *enterica*. The serotypes *S. Oranienburg* and *S. Thompson* were isolated 5 and 3 times respectively. There was no overlap between the serotype profile of isolates from RES, which included *S. Oranienburg*, *S. Poona*, *S. Pomona* and *S. Sandiego*, and the serotype profile of isolates from SS which included *S. Bovismorbificans*, *S. Montevideo*, *S. Thompson*, *S. IIIb 50:k:z* and

*S. IIIb 60:r:z* (Figure 2). A list of the *S. enterica* isolates, including GenBank accession Numbers can be found in Table 2. All but one *S. Oranienburg* isolates were either sensitive or intermediate resistant to the antimicrobials tested. One isolate showed phenotypic resistance to azithromycin (Table 3). All three *S. Thompson* isolates were multidrug-resistant (i.e., resistant to at least one antimicrobial agent in three or more antimicrobial classes). In total, eight isolates (38.1%) were resistant to at least one antimicrobial, and four isolates (19%) were resistant to seven or more antimicrobials. Resistance to chloramphenicol (33.3%,

**TABLE 2** | Serotypes, turtle species of origin and GenBank accession numbers of *Salmonella enterica* isolates.

ID	<i>Salmonella enterica</i> subspecies	Serotype	Turtle species	Accession Number
1	<i>S. enterica</i> subsp. <i>enterica</i>	Bovismorbificans	SS	JAKMZU000000000
2	<i>S. enterica</i> subsp. <i>diarizonae</i>	IIIb 50:k:z	SS	JAKMZY000000000
3	<i>S. enterica</i> subsp. <i>diarizonae</i>	IIIb 60:r:z	SS	JAKMZM000000000
4	<i>S. enterica</i> subsp. <i>enterica</i>	Manhattan	CSN	JAKMZR000000000
5	<i>S. enterica</i> subsp. <i>enterica</i>	Manhattan	CSN	JAKMZQ000000000
6	<i>S. enterica</i> subsp. <i>enterica</i>	Montevideo	SS	JAKMZJ000000000
7	<i>S. enterica</i> subsp. <i>enterica</i>	Mouline	CSN	JAKMZL000000000
8	<i>S. enterica</i> subsp. <i>enterica</i>	Oranienburg	CSN	JAKNAC000000000
9	<i>S. enterica</i> subsp. <i>enterica</i>	Oranienburg	RES	JAKNAB000000000
10	<i>S. enterica</i> subsp. <i>enterica</i>	Oranienburg	RES	JAKMZX000000000
11	<i>S. enterica</i> subsp. <i>enterica</i>	Oranienburg	RES	JAKMZV000000000
12	<i>S. enterica</i> subsp. <i>enterica</i>	Oranienburg	RES	JAKMZS000000000
13	<i>S. enterica</i> subsp. <i>enterica</i>	Pomona	CSN	JAKMZT000000000
14	<i>S. enterica</i> subsp. <i>enterica</i>	Pomona	RES	JAKMZP000000000
15	<i>S. enterica</i> subsp. <i>enterica</i>	Poona	RES	JAKMZW000000000
16	<i>S. enterica</i> subsp. <i>enterica</i>	Poona	RES	JAKMZN000000000
17	<i>S. enterica</i> subsp. <i>enterica</i>	Sandiego	CSN	JAKMZO00000000
18	<i>S. enterica</i> subsp. <i>enterica</i>	Sandiego	RES	JAKMKZK000000000
19	<i>S. enterica</i> subsp. <i>enterica</i>	Thompson	CSN	JAKNAA000000000
20	<i>S. enterica</i> subsp. <i>enterica</i>	Thompson	SS	JAKMZZ000000000
21	<i>S. enterica</i> subsp. <i>enterica</i>	Thompson	SS	JAKMZI000000000

CSN, Chinese stripe-necked turtle (*Mauremys sinensis*); RES, Red-eared slider (*Trachemys scriptaelegans*); SS, Chinese softshell turtle (*Pelodiscus sinensis*).

7/21) and streptomycin (28.6%, 6/21) were the most common phenotypes. Phenotypic resistance to the macrolide azithromycin was observed in five isolates. Phenotypic ciprofloxacin resistance was detected in four isolates and intermediate resistance in further four isolates (three *S. Thompson* and one *S. Montevideo*). No isolates were found to be carbapenem-resistant although three isolates (14.3%), two *S. Thompson* and one *S. IIIb 50:k:z*, were deemed to be ESBL-producing. A summary of the AMR phenotypes observed using disk diffusion test is shown in **Table 3**. The MIC values for chloramphenicol and linezolid in *S. IIIb 50:k:z* were  $\geq 256$   $\mu\text{g/ml}$  for both antimicrobials.

Whole genome sequencing of the isolates revealed 33 unique ARGs spanning 11 antimicrobial groups (**Tables 4a,b**). The chloramphenicol and florfenicol resistance gene *floR*, and the sulphonamide resistance gene *sul2* were the most common ARGs, both present in 33.3% (7/21) of isolates. The aminoglycoside-inactivating phosphotransferase genes *aph(6)-Id* and *aph(3'')-Ib* were the second most common, co-occurring in the same 28.6% (6/21) of isolates. Furthermore,  $\beta$ -lactamase (BLAs) genes were found in 23.8% (5/21) of isolates. The *bla*<sub>TEM-1</sub> and *bla*<sub>CMY-2</sub> ARGs were isolated in all *S. Thompson* ( $n = 3$ ) isolates. No ARGs were found in isolates belonging to *S. Oranienburg* ( $n = 5$ ), *S. Pomona* ( $n = 2$ ), *S. Poona* ( $n = 2$ ) and *S. IIIb 60:r:z* ( $n = 1$ ) serotypes. Antimicrobial resistance genes specific to aminoglycosides, amphenicols, sulphonamides and tetracyclines were identified in seven isolates, consisting of all *S. Thompson* ( $n = 3$ ), *S. Manhattan* ( $n = 2$ ), *S. Montevideo* ( $n = 1$ ) and *S. IIIb 50:k:z* ( $n = 1$ ) serotypes. The azithromycin resistance genes *mph(A)* and *erm(B)* were detected in one (*S. Thompson*)

and three (one *S. IIIb 50:k:z* and two *S. Thompson*) isolates, respectively. Five quinolone ARGs were detected in seven different isolates. These included plasmid-mediated resistance genes *qnrS1*, *qnrA1*, *aac(6)-Ib-cr* and *qepA8S*. *Mouline* ( $n = 1$ ), *S. Sandiego* ( $n = 2$ ) and *S. Bovismorbificans* ( $n = 1$ ) isolates which each only had ARGs for one single antimicrobial group, while the single *S. IIIb 50:k:z* isolate had resistance genes specific for ten different antimicrobial groups.

## DISCUSSION

This study found a high proportion of *Salmonella* carriage with a variety of AMR phenotypes and genotypes in turtles sold for food in wet markets throughout Hong Kong. A higher proportion of turtles sampled randomly from wet markets in Hong Kong (42%, 21/50) were positive for *S. enterica* than in a previous study from Shanghai, China (29.7%, 51/172), in which only SS were sampled (17). In the current study, 19.4% of SS were positive for *Salmonella*, significantly ( $p < 0.001$ ) fewer than CSN (87.5%) and RES (72.7%). Published data for *Salmonella* in chelonians varies across different studies and different turtle populations, therefore findings are difficult to compare between studies. In Japan, the prevalence of *S. enterica* among pet shop RES was 53.7% (130/227) (19). In Shanghai, prevalence among pet turtles (species not specified) was found to be 18.9% (31/164) (17) and in pet turtles (several species) from Korea it was 50% (17/34) (18). In a pet shop in Spain, the proportion of turtles positive for *S. enterica* was 75.0% (18/24) (30).

**TABLE 3 |** Antimicrobial sensitivity of *Salmonella enterica* isolates against individual antimicrobials using disk diffusion test.

ID	<i>Salmonella</i> serotype	CTX	CAZ	MEM	IPM	ETP	DOR	AMP	CIP	GEN	S	AMK	SXT	AZM	CHL	CST	ESBL-PE	CRE	Antibiogram
1	Bovismorbificans	S	S	S	S	S	S	S	I	S	S	S	S	S	S	S	-	-	-
2	IIIb 50:k:z	R	R	S	S	S	S	R	I	S	R	S	R	R	R	S	ESBL-PE	-	CTX-CAZ-AMP-S-SXT-AZM-CHL
3	IIIb 60:r:z	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
4	Manhattan	S	S	S	S	S	S	S	I	S	R	S	I	S	R	S	-	-	S-CHL
5	Manhattan	S	S	S	S	S	S	S	I	S	R	S	S	S	R	S	-	-	S-CHL
6	Montevideo	S	S	S	S	S	S	R	R	S	S	S	R	S	R	S	-	-	AMP-CIP-SXT-CHL
7	Mouline	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
8	Oranienburg	S	S	I	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
9	Oranienburg	I	S	I	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
10	Oranienburg	S	S	S	S	S	S	S	S	S	S	S	S	R	S	S	-	-	AZM
11	Oranienburg	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
12	Oranienburg	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
13	Pomona	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
14	Pomona	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
15	Poona	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
16	Poona	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
17	Sandiego	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
18	Sandiego	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	-	-	-
19	Thompson	R	R	S	S	S	S	R	R	S	R	S	R	R	R	S	ESBL-PE	-	CTX-CAZ-AMP-CIP-S-SXT-AZM-CHL
20	Thompson	R	R	S	S	S	S	R	R	S	R	S	R	R	R	S	ESBL-PE	-	CTX-CAZ-AMP-CIP-S-SXT-AZM-CHL
21	Thompson	I	I	S	S	S	S	R	R	R	R	S	R	R	R	S	-	-	AMP-CIP-GEN-S-SXT-AZM-CHL
	<i>n</i> (%) resistant	3	3	0	0	0	0	5	4	1	6	0	5	5	7	0			
		(14.3)	(14.3)	(0)	(0)	(0)	(0)	(23.8)	(19)	(4.8)	(28.6)	(0)	(23.8)	(23.8)	(33.3)	(0)			

S, sensitive; I, intermediate; R, resistant; CTX, cefotaxime; CAZ, ceftazidime; MEM, meropenem; IPM, imipenem; ETP, ertapenem; DOR, doripenem; AMP, ampicillin; CIP, ciprofloxacin; GEN, gentamicin; S, streptomycin; AMK, amikacin; SXT, trimethoprim-sulfamethoxazole; AZM, azithromycin; CHL, chloramphenicol; CST, colistin; ESBL-PE, extended-spectrum beta-lactamase-producing Enterobacteriaceae.

**TABLE 4a** | Antimicrobial resistance genes identified in *Salmonella enterica* isolates from turtles.

ID	Serotype	Aminoglycosides					Amphenicols	Beta-lactams	Fosfomycin
		AMK	GEN	HYG	KAN	S	FLR/CHL	Cephalosporins	FOF
1	Bovismorbificans								
2	IIIb 50:k:z	<i>aac(6')-Ib-cr5</i>			<i>aph(3')-Ia</i>	<i>aadA16 / aph(3'')-Ib / aph(6)-Id</i>	<i>floR</i>	<i>bla<sub>CMY-2</sub></i>	
3	IIIb 60:r:z								
4	Manhattan					<i>aph(3'')-Ib / aph(6)-Id</i>	<i>floR</i>		
5	Manhattan					<i>aph(3'')-Ib / aph(6)-Id</i>	<i>floR</i>		
6	Montevideo	<i>aac(6')-Ib-cr5</i>	<i>aac(3)-IV</i>	<i>aph(4)-Ia</i>		<i>aadA2</i>	<i>catB3 / floR</i>	<i>bla<sub>OXA-1</sub></i>	
7	Moualine								<i>fosA7</i>
8	Oranienburg								
9	Oranienburg								
10	Oranienburg								
11	Oranienburg								
12	Oranienburg								
13	Pomona								
14	Pomona								
15	Poona								
16	Poona								
17	Sandiego								<i>fosA7</i>
18	Sandiego								<i>fosA7</i>
19	Thompson				<i>aph(3')-Ia</i>	<i>aph(3'')-Ib / aph(6)-Id</i>	<i>floR</i>	<i>bla<sub>TEM-1</sub></i>	<i>bla<sub>CMY-2</sub></i>
20	Thompson					<i>aph(3'')-Ib / aph(6)-Id</i>	<i>floR</i>	<i>bla<sub>TEM-1</sub></i>	<i>bla<sub>CMY-2</sub></i>
21	Thompson				<i>aph(3')-Ia</i>	<i>aph(3'')-Ib / aph(6)-Id</i>	<i>floR</i>	<i>bla<sub>TEM-1</sub></i>	<i>bla<sub>CMY-2</sub></i>

AMK, Amikacin; GEN, Gentamicin; HYG, Hygromycin B; KAN, Kanamycin; S, Streptomycin; FLR, Florfenicol; CHL, Chloramphenicol; FOF, Fosfomycin.

**TABLE 4b |** Antimicrobial resistance genes identified in *Salmonella enterica* isolates from turtles.

ID	Serotype	Lincosamides/Streptogramins	Macrolides AZM/ERY/TEL/TYL	Quinolones	Rifamycins	Sulphonamides		Tetracyclines
						TMP	TET	
1	Bovismorbificans			<i>gyrA_S83F</i>				
2	IIIb 50:k:z	<i>cfr</i>	<i>cfr / erm(B) / mph(E) / msr(E)</i>	<i>aac(6')-Ib-cr5</i>	<i>arr-3</i>	<i>sul1 / sul2</i>	<i>dfrA12 / dfrA27</i>	<i>tet(A)</i>
3	IIIb 60:r:z							
4	Manhattan			<i>qnrS1</i>		<i>sul2</i>		<i>tet(D)</i>
5	Manhattan			<i>qnrS1</i>		<i>sul2</i>		<i>tet(D)</i>
6	Montevideo			<i>aac(6')-Ib-cr5 / qnrA1</i>	<i>arr-3</i>	<i>sul1 / sul2</i>	<i>dfrA12</i>	<i>tet(B)</i>
7	Moualine							
8	Oranienburg							
9	Oranienburg							
10	Oranienburg							
11	Oranienburg							
12	Oranienburg							
13	Pomona							
14	Pomona							
15	Poona							
16	Poona							
17	Sandiego							
18	Sandiego							
19	Thompson		<i>erm(B) / mph(E) / msr(E)</i>	<i>qepA8 / qnrS1</i>	<i>arr-2</i>	<i>sul1 / sul2</i>	<i>dfrA12 / dfrA14</i>	<i>tet(A)</i>
20	Thompson		<i>mph(A)</i>			<i>sul2</i>		<i>tet(A)</i>
21	Thompson		<i>erm(B) / mph(E) / msr(E)</i>	<i>qepA8 / qnrS1</i>	<i>arr-2</i>	<i>sul1 / sul2</i>	<i>dfrA12 / dfrA14</i>	<i>tet(A)</i>

AZM, Azithromycin; ERY, Erythromycin; TEL, Telithromycin; TYL, Tylosin; TMP, Trimethoprim; TET, Tetracycline.

The most common *Salmonella* serotypes isolated from turtles in this study were Oranienburg and Thompson. The latter was—together with *S. Typhimurium*—one of the two predominant serotypes identified in native European pond turtle (*Emys orbicularis*) and introduced RES in natural ponds in Spain (31). In addition, *S. Thompson* was also the most common serotype (17%, 14/85) isolated from SS and pet turtles in wet markets in Shanghai, China (17). *Salmonella* Oranienburg and *S. Sandiego* were isolated from RES sold in pet shops in Japan (19). *Salmonella* Manhattan is the only serotype isolated from turtles in the current study that—to the best of the author's knowledge—has never been reported in chelonians before. *Salmonella* Manhattan has been isolated from other reptiles, including Iguanas (*Conolophus subcristatus*) from the Galápagos Islands (32), captive Andros and Bahamian rock iguanas (*Cyclura cychlura* and *Cyclura rileyi*) (33), and northern water snakes (*Nerodia sipedon sipedon*) (34). Some of the serotypes identified here have also been linked to human salmonellosis. *Salmonella* Pomona, which is considered to be particularly pathogenic (35), caused ~18% of human salmonellosis cases due to turtle exposure in the USA between 2006 and 2014 (5). This serotype is particularly prevalent in turtles and other reptiles, as it has been found in 39% of free-living introduced RES caught in mainland China (35), and 12% of reptiles sampled from pet shops in Spain (36). *Salmonella* Sandiego was identified in three and *S. Pomona* and *S. Poona* each in two out of eight outbreaks of turtle-associated salmonellosis in young children during 2011–2013 across 41 states of the USA (37). *Salmonella* Thompson, along with *S. Typhimurium*, was among the four most-frequently recovered serotypes from human patients in Shanghai (38). In Hong Kong, the five main serotypes reported in human salmonellosis (regardless of origin of the infection) are *S. Enteritidis* (31.8%), *S. Typhimurium* (16.1%), *S. Stanley* (6.4%), *S. Derby* (6.0%) and *S. Agona* (2.5%). None of these serotypes were isolated from turtles in this study.

Phenotypic AMR was identified in eight (38.1%) and genotypic AMR in 11 (52.4%) of all 21 *Salmonella* isolates. The proportion of samples with AMR was relatively low compared to a similar study performed in wet markets in Shanghai where 100% ( $n = 82$ ) of isolates showed resistance to at least one antimicrobial and 84.1% to at least three antimicrobials (17). A high level of AMR was also observed among pet reptiles in Spain, where 100% ( $n = 75$ ) of *Salmonella* isolates were resistant to at least one of the 12 antimicrobials tested, and 72% were multidrug-resistant (36). Antimicrobial resistance genes for cephalosporin were detected in 19% (4/21) of isolates in this study. Furthermore, five quinolone ARGs were detected in seven different isolates. These findings are significant as cephalosporins and fluoroquinolones are the antimicrobials of choice to treat salmonellosis in humans and any *Salmonella* showing resistance to these drugs is a major concern for public health. Resistance to extended-spectrum cephalosporins and fluoroquinolones is particularly worrying since they represent the first-line antimicrobials to treat invasive salmonellosis in children and in adults respectively (39). Furthermore, the quinolone ARGs are significant in their ability to confer resistance by horizontal gene exchange (40).

In Hong Kong, 21% of *Salmonella* isolates collected between 2002 and 2004 from human cases were multidrug-resistant (41). The results from this and the current study are in stark contrast to findings from 1986 to 1996, where 99% of *S. enterica* serotype enteritidis strains isolated in Hong Kong were susceptible to 17 of the 19 antimicrobial agents tested (42), emphasizing the rapid emergence of AMR in *Salmonella enterica*. In the current study, resistance to chloramphenicol (33.3%, 7/21) and streptomycin (28.6%, 6/21) was the most common phenotypic AMR among the *Salmonella* isolates. The chloramphenicol and florfenicol resistance gene *floR* was one of the most common ARGs detected. Despite chloramphenicol being banned from aquaculture in mainland China since 2002 and streptomycin being a non-authorized antimicrobial drug, these two antimicrobials seem to be commonly used in aquaculture in mainland China (43). Furthermore, studies conducted in SS turtles for human consumption from mainland China in 2012 and 2016 showed that chloramphenicol residues could be detected in turtles' tissues. Thirteen antimicrobials have been authorized for use in aquaculture in mainland China: doxycycline, enrofloxacin, florfenicol, flumequine, neomycin, norfloxacin, oxolinic acid, sulfadiazine, sulfamethazine, sulfamethoxazole, sulfamonomethoxine, thiamphenicol and trimethoprim (43). Resistance against chloramphenicol and streptomycin in *Salmonella* isolates was also described in a study performed on pet turtles in South Korea, where 82.9% of *Salmonella* isolates were resistant against streptomycin (18). Similarly, these two antimicrobials are banned from use in aquaculture in South Korea. The authors hypothesized that these AMR patterns were due to the unregistered use of these drugs in pet turtles (18). However, this hypothesis could not be verified as the pet turtles were purchased from pet stores or online shops with no available information about their origin and breeding conditions. A similar hypothesis about the use of unauthorized antimicrobial drugs could be plausible for the current study.

Both *S. Manhattan* isolates were resistant against several antimicrobials. *Salmonella* Manhattan isolated from terrestrial wild iguanas (*Conolophus subcristatus*) from the Galapagos islands did not exhibit any resistance to antimicrobials (32). In the other two studies that identified *S. Manhattan* in captive iguanas (*Cyclura cychlura* and *Cyclura rileyi*) (33) and in northern water snakes (*Nerodia sipedon sipedon*) from Pennsylvania (34), no AMR testing was performed.

Similarly to the findings in turtles from wet markets in Shanghai (17), all *S. Thompson* isolates in this study were multidrug-resistant. While a phenotypic resistance against ciprofloxacin was detected, no ARGs against ciprofloxacin were found for this serotype. Resistance against ciprofloxacin is very problematic as it is the antimicrobial of choice to treat salmonellosis in humans. Ciprofloxacin-resistant *Salmonella* have increased in mainland China from 2.3% in 2006 to 5.9% in 2013 (44).

The *bla*<sub>CMY-2</sub>, which codes for the plasmid-mediated AmpC  $\beta$ -lactamase CMY-2 that hydrolyses third-generation cephalosporins (45) was detected in four isolates. Three of these isolates were phenotypically resistant to both third-generation cephalosporins tested by disk diffusion and the fourth had an intermediate phenotype. Because *bla*<sub>CMY-2</sub> is encoded within a

plasmid, it can be transmitted horizontally and spread among bacterial populations in animals and humans (46). The *bla*<sub>TEM-1</sub> gene, which was found in the three *S. Thomson* isolates, codes for the TEM-1  $\beta$ -lactamase. Mutation of this gene by only two single nucleotide polymorphisms (SNPs) can produce an ESBL capable of degrading third generation cephalosporins (47). The *bla*<sub>TEM-1</sub> gene is amongst the most common ESBL genes found in *Salmonella* isolates in other studies (48, 49).

The *cfr* gene was detected in one *S. IIIb* 50:k:z isolate from a SS imported from Thailand. To the best of the author's knowledge, this is the first report of the *cfr* gene in *Salmonella*. This is an important finding as *cfr* confers resistance to five classes of antimicrobials, namely phenicols, lincosamides, oxazolidinones, pleuromutilins, and streptogramin A (50). Although oxazolidinones are not or only partially effective against Gram-negative bacteria due to their intrinsic resistance (51) the presence of the *cfr* gene in *Salmonella* could represent an additional reservoir for its potential horizontal spread to Gram-positive bacteria. The detection of *cfr* from *Salmonella* in turtles sold for consumption is particularly worrying as food is one of the main matrices responsible for transferring AMR determinants to humans. The *cfr* gene has also been identified in several species of staphylococcal bacteria (52), and in species within the genera *Bacillus*, *Enterococcus*, *Streptococcus*, *Macrocooccus*, *Jeotgalicoccus*, *Proteus*, and *Escherichia* (53). It has also been detected in Methicillin-resistant *Staphylococcus aureus* (MRSA) strains from animals, humans (54) and food items (frozen dumpling) in mainland China (55). It was also identified in *Pasteurella multocida* isolated from poultry in mainland China (56).

In cases of salmonellosis that require antimicrobial treatment, the first-line of therapy is typically ciprofloxacin, azithromycin or the third-generation cephalosporin, ceftriaxone, although treatment options also include ampicillin and trimethoprim-sulfamethoxazole depending on the resistance profile (57). Worryingly, a phenotypic resistance and ARGs to all three commonly used first-line antimicrobial groups were detected in isolates from this study. *Salmonella* has been placed on the WHO high-priority list for the development of new antimicrobials because of the emergence of fluoroquinolone resistance (58).

An obvious sex bias was observed in our samples, as all SS were male and all RES and CSN were females. Sex determination in most turtles, including RES and CSN, is considered to be temperature-dependent, giving rise to males at lower temperatures and females at higher temperatures (i.e., over 30°C) (59, 60). In SS however, recent studies indicate that sex determination might have a genetic basis (61). Given the general lack of knowledge on turtle farming, it is difficult to make a hypothesis on the drivers of sex bias in the turtle market. Since sex and species are potential confounding variables, these results should be interpreted with care. The prevalence of *Salmonella* in several wild turtle species in the US was significantly higher in females than in males (62). This could potentially explain the differences observed in the current study, but it would require equal representation of both sexes in all three species.

The representativeness of the turtle population sampled needs to be interpreted with caution given the relatively small sample size and short sampling period. There is a lack of

detailed information on turtle trade in Hong Kong SAR (i.e., turtle availability and volume in wet markets, species and sex distribution etc.) and data on *Salmonella* prevalence. Deciding on the right sampling strategy was challenging and the total number of samples taken from each retail location could not be adjusted accordingly. This project therefore used a pilot study approach. The results demonstrate a complex demographic structure (i.e., sex and species distribution) that might potentially persist even if a larger number of turtles is sampled.

Finally, the diversity of *Salmonella* serovars described here might be underestimated as only one isolation method and incubation temperature was used. However, the aim of the study was to generate some baseline data and to further characterize the isolated *Salmonella* strains using whole genome sequencing.

## CONCLUSION

This pilot study reports a high prevalence and serotype diversity of *Salmonella* among chelonians sold as food in Hong Kong wet markets, with the serotype *S. Manhattan* being—to the best of the author's knowledge—reported in chelonians for the first time. Resistance was detected against antimicrobials banned from aquaculture in mainland China and those recommended as first-line treatment for salmonellosis. The multidrug-resistance gene *cfr* is—to the best of the author's knowledge—reported for the first time in *Salmonella*. This is a worrying finding as it indicates an expansion of the *cfr* reservoir and the potential for horizontal spread to other bacteria. A systematic surveillance of *Salmonella* ideally from a representative sample of farmed turtles is essential to safeguard public health and for the timely detection of emerging threats.

## DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/supplementary material.

## ETHICS STATEMENT

The animal study was reviewed and approved by Animal Research Ethics Sub-Committee of City University of Hong Kong (Internal Ref: A-0592).

## AUTHOR CONTRIBUTIONS

VC performed the necropsies and wrote the body of the manuscript. KL and HL performed culture and sensitivity testing. LW did the data analysis and assisted with manuscript writing. RP and CC assisted with study design and discussion writing. JH and MS carried out molecular analysis. RS assisted with laboratory data interpretation and discussion. IM developed the research idea and supervised the project. All authors contributed to the discussion and comments on the manuscript.

## FUNDING

This work was supported by the City University of Hong Kong start-up grant for new Faculty (project no. 9610449).

## ACKNOWLEDGMENTS

We would like to thank Nicole Cernela (NENT) for her technical support with Illumina sequencing.

## REFERENCES

- Ades G, Banks CB, Buhlmann KA, Chan B, Chang HC, Chen TH, et al. Turtle trade in northeast Asia: regional summary (China, Hong Kong, and Taiwan). In: *Proceedings of a Workshop on Conservation and Trade of Freshwater Turtles and Tortoises in Asia, Phnom Penh, Cambodia 1999*. Chelonian Research Foundation (2000). p. 52–54.
- Cheung SM, Dudgeon D. Quantifying the Asian turtle crisis: market surveys in southern China, 2000–2003. *AQUAT Conserv.* (2006) 16:751–70. doi: 10.1002/aqc.803
- Haitao S, Parham JF, Zhiyong F, Meiling H, Feng Y. Evidence for the massive scale of turtle farming in China. *Oryx.* (2008) 42:147–50. doi: 10.1017/S0030605308000562
- Magnino S, Colin P, Dei-Cas E, Madsen M, McLaughlin J, Nöckler K, et al. Biological risks associated with consumption of reptile products. *Int J Food Microbiol.* (2009) 134:163–75. doi: 10.1016/j.ijfoodmicro.2009.07.001
- Bosch S, Tauxe R V, Behravesh CB. Turtle-associated salmonellosis, United States, 2006–2014. *Emerg Infect Dis.* (2016) 22:1149. doi: 10.3201/eid2207.150685
- Sodagari HR, Habib I, Shahabi MP, Dybing NA, Wang P, Bruce M. A review of the public health challenges of Salmonella and turtles. *Vet Sci.* (2020) 7:56. doi: 10.3390/vetsci7020056
- Harris JR, Neil KP, Behravesh CB, Sotir MJ, Angulo FJ. Recent multistate outbreaks of human Salmonella infections acquired from turtles: a continuing public health challenge. *Clin Infect Dis.* (2010) 50:554–9. doi: 10.1086/649932
- O'Grady KA, Krause V. An outbreak of salmonellosis linked to a marine turtle. *Southeast Asian J Trop Med Public Health.* (1999) 30:324–7. doi: 10.1038/sj/ph/1900680
- Draper A, James CL, Pascall JE, Shield KJ, Langrell J, Hogg A. An outbreak of Salmonella Muenchen after consuming sea turtle, Northern Territory, Australia, 2017. *Commun Dis Intell Q Rep.* (2017) 41:E290–4.
- Fukushima H, Okuno J, Fujiwara Y, Hosoda T, Kurazono T, Ohtsuka K, et al. An outbreak of Salmonella food poisoning at a snapping turtle restaurant. *J Japanese Assoc Infect Dis.* (2008) 61:328.
- Issenhuth-Jeanjean S, Roggentin P, Mikoleit M, Guibourdenche M, de Pinna E, Nair S, et al. Supplement 2008–2010 (no. 48) to the White–Kauffmann–Le Minor scheme. *Res Microbiol.* (2014) 165:526–30. doi: 10.1016/j.resmic.2014.07.004
- Majowicz SE, Musto J, Scallan E, Angulo FJ, Kirk M, O'Brien SJ, et al. The global burden of *Salmonella gastroenteritis*. *Clin Infect Dis.* (2010) 50:882–9. doi: 10.1086/650733
- Hong Kong SAR, Department of Health. *Centre for Health Protection*. Review of nontyphoidal Salmonella food poisoning in Hong Kong: CHP (2011). Available online at: [https://www.chp.gov.hk/files/pdf/review\\_of\\_nontyphoidal\\_salmonella\\_food\\_poisoning\\_in\\_hong\\_kong\\_r.pdf](https://www.chp.gov.hk/files/pdf/review_of_nontyphoidal_salmonella_food_poisoning_in_hong_kong_r.pdf) (accessed November 4, 2021).
- Michael GB, Schwarz S. Antimicrobial resistance in zoonotic nontyphoidal Salmonella: alarming trend? *Clin Microbiol Infect.* (2016) 22:968–74. doi: 10.1016/j.cmi.2016.07.033
- Parisi A, Crump JA, Glass K, Howden BP, Furuya-Kanamori L, Vilkins S, et al. Health Outcomes from multidrug-resistant salmonella infections in high-income countries: a systematic review and meta-analysis. *Foodborne Pathog Dis.* (2018) 15:428–36. doi: 10.1089/fpd.2017.2403
- Marchello CS, Birkhold M, Crump JA, Martin LB, Ansah MO, Breggi G, et al. Complications and mortality of non-typhoidal salmonella invasive disease: a global systematic review and meta-analysis. *Lancet Infect Dis.* (2022) 22:692–705. doi: 10.1016/S1473-3099(21)00615-0
- Zhang J, Kuang D, Wang F, Meng J, Jin H, Yang X, et al. Turtles as a possible reservoir of nontyphoidal Salmonella in Shanghai, China. *Foodborne Pathog Dis.* (2016) 13:428–33. doi: 10.1089/fpd.2015.2107
- Hossain S, De Silva BCJ, Dahanayake PS, Shin G-W, Heo G-J. Molecular characterisation of virulence, antimicrobial resistance genes, and class one integron cassettes in salmonella enterica subsp. enterica isolated from pet turtles in Seoul, Korea. *J Exot Pet Med.* (2019) 28:209–17. doi: 10.1053/j.jepm.2018.11.002
- Kuroki T, Ishihara T, Nakajima N, Furukawa I, Une Y. Prevalence of Salmonella enterica subspecies enterica in red-eared sliders *Trachemys scripta elegans* retailed in pet shops in Japan. *Jpn J Infect Dis.* (2019) 72:38–43. doi: 10.7883/yoken.JJID.2018.14
- Food and Environmental Hygiene Department. *List of FEHD Public Markets / Cooked Food Markets*. Food and Environmental Hygiene Department; Available online at: [https://www.fehd.gov.hk/english/pleasant\\_environment/tidy\\_market/Markets\\_CFC\\_list.html](https://www.fehd.gov.hk/english/pleasant_environment/tidy_market/Markets_CFC_list.html) (accessed December 17, 2020).
- Underwood W, Anthony R. *AVMA guidelines for the euthanasia of animals: 2020 edition*. Retrieved on March 2013:2020–2021 (2020).
- International Organization for Standardization. *Microbiology of the food chain—horizontal method for the detection, enumeration and serotyping of Salmonella—part 1: detection of Salmonella spp.* International Organization for Standardization Geneva, Switzerland (2017).
- Clinical and Laboratory Standards Institute. *Performance Standards for Antimicrobial Susceptibility Testing, CLSI supplement M100*, 31st ed. USA (2021).
- Baschera M, Cernela N, Stevens MJA, Liljander A, Jores J, Corman VM, et al. Shiga toxin-producing Escherichia coli (STEC) isolated from fecal samples of African dromedary camels. *One Heal.* (2019) 7:100087. doi: 10.1016/j.onehlt.2019.100087
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, et al. SPAdes: a new genome assembly algorithm and its applications to single-cell Sequencing. *J Comput Biol.* (2012) 19:455–77. doi: 10.1089/cmb.2012.0021
- Zhang S, Yin Y, Jones MB, Zhang Z, Kaiser BLD, Dinsmore BA, et al. Salmonella serotype determination utilizing high throughput genome sequencing data. *J Clin Microbiol.* (2015) 53:1685–92. doi: 10.1128/JCM.00323-15
- Feldgarden M, Brover V, Gonzalez-Escalona N, Frye JG, Haendiges J, Haft DH, et al. AMRFinderPlus and the Reference Gene Catalog facilitate examination of the genomic links among antimicrobial resistance, stress response, and virulence. *Sci Rep.* (2021) 11:12728. doi: 10.1038/s41598-021-91456-0
- Stangroom J. *Fisher Exact Test Calculator*. *Social Science Statistics*. Available online at: <https://www.socscistatistics.com/tests/fisher/default2.aspx> (accessed February 10, 2022).
- Sergeant ESG. *Epitools Epidemiological Calculators*. Ausvet. Available online at: <http://epitools.ausvet.com.au> (accessed February 11, 2022).
- Marin C, Vega S, Marco-Jiménez F. Tiny turtles purchased at pet stores are a potential high risk for Salmonella human infection in the Valencian region, eastern Spain. *Vector Borne Zoonotic Dis.* (2016) 16:455–60. doi: 10.1089/vbz.2016.1950
- Marin C, Ingesa-Capaccioni S, González-Bodi S, Marco-Jiménez F, Vega S. Free-living turtles are a reservoir for Salmonella but not for Campylobacter. *PLoS ONE.* (2013) 8:14–7. doi: 10.1371/journal.pone.0072350
- Franco A, Hendriksen RS, Lorenzetti S, Onorati R, Gentile G, Dell'Omo G, et al. Characterization of Salmonella occurring at high prevalence in a population of the land iguana *Conolophus subcristatus* in Galápagos Islands, Ecuador. *PLoS ONE.* (2011) 6:e23147. doi: 10.1371/journal.pone.0023147
- Geue L, Löschner U. Salmonella enterica in reptiles of German and Austrian origin. *Vet Microbiol.* (2002) 84:79–91. doi: 10.1016/S0378-1135(01)00437-0
- Chambers DL, Hulse AC. Salmonella serovars in the herpetofauna of Indiana County, Pennsylvania. *Appl Environ Microbiol.* (2006) 72:3771–3. doi: 10.1128/AEM.72.5.3771-3773.2006
- Gong S, Wang F, Shi H, Zhou P, Ge Y, Hua L, et al. Highly pathogenic Salmonella Pomona was first isolated from the

- exotic red-eared slider (*Trachemys scripta elegans*) in the wild in China: implications for public health. *Sci Total Environ.* (2014) 468:28–30. doi: 10.1016/j.scitotenv.2013.08.025
36. Marin C, Lorenzo-Rebenaque L, Laso O, Villora-Gonzalez J, Vega S. Pet reptiles: a potential source of transmission of multidrug-resistant *Salmonella*. *Front Vet Sci.* (2021) 7:1157. doi: 10.3389/fvets.2020.613718
  37. Walters MS, Simmons L, Anderson TC, DeMent J, Van Zile K, Matthias LP, et al. Outbreaks of salmonellosis from small turtles. *Pediatrics.* (2016) 137:e20151735. doi: 10.1542/peds.2015-1735
  38. Zhang J, Jin H, Hu J, Yuan Z, Shi W, Ran L, et al. Serovars and antimicrobial resistance of non-typhoidal *Salmonella* from human patients in Shanghai, China, 2006–2010. *Epidemiol Infect.* (2014) 142:826–32. doi: 10.1017/S0950268813001659
  39. Nair DVT, Venkitanarayanan K, Kollanoor Johny A. Antibiotic-resistant *Salmonella* in the food supply and the potential role of antibiotic alternatives for control. *Foods.* (2018) 7:167. doi: 10.3390/foods7100167
  40. Ruiz J. Transferable mechanisms of quinolone resistance from 1998 onward. *Clin Microbiol Rev.* (2019) 32:e00007-19. doi: 10.1128/CMR.00007-19
  41. Jin Y, Ling JM. Prevalence of integrons in antibiotic-resistant *Salmonella* spp. in Hong Kong. *Jpn J Infect Dis.* (2009) 62:432–9.
  42. Ling JM, Koo IC, Kam KM, Cheng AF. Antimicrobial susceptibilities and molecular epidemiology of *Salmonella enterica* serotype Enteritidis strains isolated in Hong Kong from 1986 to 1996. *J Clin Microbiol.* (1998) 36:1693–9. doi: 10.1128/JCM.36.6.1693-1699.1998
  43. Liu X, Steele JC, Meng X-Z. Usage, residue, and human health risk of antibiotics in Chinese aquaculture: a review. *Environ Pollut.* (2017) 223:161–9. doi: 10.1016/j.envpol.2017.01.003
  44. Kuang D, Zhang J, Xu X, Shi W, Chen S, Yang X, et al. Emerging high-level ciprofloxacin resistance and molecular basis of resistance in *Salmonella enterica* from humans, food and animals. *Int J Food Microbiol.* (2018) 280:1–9. doi: 10.1016/j.ijfoodmicro.2018.05.001
  45. Koga VL, Maluta RP, da Silveira WD, Ribeiro RA, Hungria M, Vespero EC, et al. Characterization of CMY-2-type beta-lactamase-producing *Escherichia coli* isolated from chicken carcasses and human infection in a city of South Brazil. *BMC Microbiol.* (2019) 19:174. doi: 10.1186/s12866-019-1550-3
  46. Palmeira JD, Cunha MV, Carvalho J, Ferreira H, Fonseca C, Torres RT. Emergence and spread of cephalosporinases in wildlife: a review. *Animals.* (2021) 11:1765. doi: 10.3390/ani11061765
  47. Muhammad I, Golparian D, Dillon J-AR, Johansson Å, Ohnishi M, Sethi S, et al. Characterisation of bla TEM genes and types of beta-lactamase plasmids in *Neisseria gonorrhoeae* – the prevalent and conserved bla TEM-135 has not recently evolved and existed in the Toronto plasmid from the origin. *BMC Infect Dis.* (2014) 14:454. doi: 10.1186/1471-2334-14-454
  48. Adel WA, Ahmed AM, Hegazy Y, Torky HA, Shimamoto T. High prevalence of ESBL and plasmid-mediated quinolone resistance genes in *Salmonella enterica* isolated from retail meats and slaughterhouses in Egypt. *Antibiotics.* (2021) 10:881. doi: 10.3390/antibiotics10070881
  49. Qiao J, Alali WQ, Liu J, Wang Y, Chen S, Cui S, et al. Prevalence of virulence genes in extended-spectrum beta-lactamases (ESBLs)-producing salmonella in retail raw chicken in China. *J Food Sci.* (2018) 83:1048–52. doi: 10.1111/1750-3841.14111
  50. Long KS, Poehlsgaard J, Kehrenberg C, Schwarz S, Vester B. The Cfr rRNA methyltransferase confers resistance to phenicols, lincosamides, oxazolidinones, pleuromutilins, and streptogramin A antibiotics. *Antimicrob Agents Chemother.* (2006) 50:2500–5. doi: 10.1128/AAC.00131-06
  51. Zahedi Bialvaei A, Rahbar M, Yousefi M, Asgharzadeh M, Samadi Kafil H. Linezolid: a promising option in the treatment of Gram-positives. *J Antimicrob Chemother.* (2017) 72:354–64. doi: 10.1093/jac/dkw450
  52. Liu X-Q, Wang J, Li W, Zhao L-Q, Lu Y, Liu J-H, et al. Distribution of cfr in *Staphylococcus* spp. and *Escherichia coli* strains from pig farms in China and characterization of a novel cfr-carrying F43:A-:B- plasmid. *Front Microbiol.* (2017) 8:329. doi: 10.3389/fmicb.2017.00329
  53. Vester B. The cfr and cfr-like multiple resistance genes. *Res Microbiol.* (2018) 169:61–6. doi: 10.1016/j.resmic.2017.12.003
  54. Li S-M, Zhou Y-F, Li L, Fang L-X, Duan J-H, Liu F-R, et al. Characterization of the multi-drug resistance gene cfr in methicillin-resistant *Staphylococcus aureus* (MRSA) strains isolated from animals and humans in China. *Front Microbiol.* (2018) 9:2925. doi: 10.3389/fmicb.2018.02925
  55. Zhang F, Wu S, Lei T, Wu Q, Zhang J, Huang J, et al. Presence and characterization of methicillin-resistant *Staphylococcus aureus* co-carrying the multidrug resistance genes cfr and lsa(E) in retail food in China. *Int J Food Microbiol.* (2022) 363:109512. doi: 10.1016/j.ijfoodmicro.2021.109512
  56. Chen H, Deng H, Cheng L, Liu R, Fu G, Shi S, et al. First report of the multiresistance gene cfr in *Pasteurella multocida* strains of avian origin from China. *J Glob Antimicrob Resist.* (2020) 23:251–5. doi: 10.1016/j.jgar.2020.09.018
  57. Chen H-M, Wang Y, Su L-H, Chiu C-H. Nontyphoid salmonella infection: microbiology, clinical features, and antimicrobial therapy. *Pediatr Neonatol.* (2013) 54:147–52. doi: 10.1016/j.pedneo.2013.01.010
  58. Tacconelli E, Carrara E, Savoldi A, Harbarth S, Mendelson M, Monnet DL, et al. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. *Lancet Infect Dis.* (2018) 18:318–27. doi: 10.1016/S1473-3099(17)30753-3
  59. Rhen T, Willingham E, Sakata JT, Crews D. Incubation temperature influences sex-steroid levels in juvenile red-eared slider turtles, *Trachemys scripta*, a species with temperature-dependent sex determination I. *Biol Reprod.* (1999) 61:1275–80. doi: 10.1095/biolreprod61.5.1275
  60. Du W-G, Wang L, Shen J-W. Optimal temperatures for egg incubation in two Geomydrid turtles: *Ocadia sinensis* and *Mauremys mutica*. *Aquaculture.* (2010) 305:138–42. doi: 10.1016/j.aquaculture.2010.03.032
  61. Mu Y, Zhao B, Tang WQ, Sun BJ, Zeng ZG, Valenzuela N, et al. Temperature-dependent sex determination ruled out in the Chinese soft-shelled turtle (*Pelodiscus sinensis*) via molecular cytogenetics and incubation experiments across populations. *Sex Dev.* (2015) 9:111–7. doi: 10.1159/000373903
  62. Hernandez SM, Maurer JJ, Yabsley MJ, Peters VE, Presotto A, Murray MH, et al. Free-living aquatic turtles as sentinels of *Salmonella* spp. for water bodies. *Front Vet Sci.* (2021) 8:674973. doi: 10.3389/fvets.2021.674973

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Colon, Lugsomya, Lam, Wahl, Parkes, Cormack, Horlbog, Stevens, Stephan and Magouras. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.