



Potential Feed Additives as Antibiotic Alternatives in Broiler Production

Habtamu Ayalew^{1,2}, Haijun Zhang^{1*}, Jing Wang¹, Shugeng Wu¹, Kai Qiu¹, Guanghai Qi¹, Ayalew Tekeste², Teketay Wassie³ and Demissie Chanie²

¹ Laboratory of Quality and Safety Risk Assessment for Animal Products on Feed Hazards (Beijing) of the Ministry of Agriculture and Rural Affairs Feed Research Institute, Chinese Academy of Agricultural Sciences, Beijing, China, ² College of Veterinary Medicine and Animal Sciences, University of Gondar, Gondar, Ethiopia, ³ Key Laboratory of Agro-Ecological Processes in Subtropical Region, National Engineering Laboratory for Pollution Control and Waste Utilization in Livestock and Poultry Production, Hunan Provincial Engineering Research Center for Healthy Livestock and Poultry Production, Institute of Subtropical Agriculture, Chinese Academy of Sciences, Changsha, China

This article aimed to describe the current use scenario, alternative feed additives, modes of action and ameliorative effects in broiler production. Alternative feed additives have promising importance in broiler production due to the ban on the use of certain antibiotics. The most used antibiotic alternatives in broiler production are phytochemicals, organic acids, prebiotics, probiotics, enzymes, and their derivatives. Antibiotic alternatives have been reported to increase feed intake, stimulate digestion, improve feed efficiency, increase growth performance, and reduce the incidence of diseases by modulating the intestinal microbiota and immune system, inhibiting pathogens, and improving intestinal integrity. Simply, the gut microbiota is the target to raise the health benefits and growth-promoting effects of feed additives on broilers. Therefore, naturally available feed additives are promising antibiotic alternatives for broilers. Then, summarizing the category, mode of action, and ameliorative effects of potential antibiotic alternatives on broiler production may provide more informed decisions for broiler nutritionists, researchers, feed manufacturers, and producers.

Keywords: antibiotic alternatives, broiler, feed additives, phytochemical, prebiotic, probiotic

OPEN ACCESS

Edited by:

Bing Dong,
China Agricultural University, China

Reviewed by:

Muhammad Saeed,
Northwest A&F University, China
Khalid M. Mahrose,
Zagazig University, Egypt

*Correspondence:

Haijun Zhang
zhanghaijun@caas.cn

Specialty section:

This article was submitted to
Animal Nutrition and Metabolism,
a section of the journal
Frontiers in Veterinary Science

Received: 16 April 2022

Accepted: 09 May 2022

Published: 17 June 2022

Citation:

Ayalew H, Zhang H, Wang J, Wu S,
Qiu K, Qi G, Tekeste A, Wassie T and
Chanie D (2022) Potential Feed
Additives as Antibiotic Alternatives in
Broiler Production.
Front. Vet. Sci. 9:916473.
doi: 10.3389/fvets.2022.916473

INTRODUCTION

The poultry sector is one of the largest food industries in the globe (1). In the near future, by 2050, it would be projected to be 121% of the year 2005 production (2). It has continual growth and industrialization in many parts of the world (3). Particularly, broiler production has shown exponential growth in global meat consumption and business profit, which will be higher in the next century (4–6). This could be because of its comparative advantages including good quality of nutrition, delicious taste, low-fat content, short production period, low production cost, rapid economic progress, and affordable price even for poor levels of society (7, 8). The production has ascended from 9 to 132 million tons in the year range of 1961 to 2019 (3). Seventeen percent of global output is produced in the United States, which is the world's largest poultry meat producer followed by China and Brazil (3).

Per capita, meat consumption has been an increase in the world in which poultry meat accounts 70% of total meat consumed. Over 66 billion broilers are slaughtered in the world each year (9). From these amounts of slaughtered birds, nearly 110 million tons are produced per annum. Per capita broiler meat consumption is higher in developed countries (10). For example, the average broiler meat consumption per capita

in the United States, Brazil, and China is 48, 44.2, and 8.3 kg/head/yr, respectively, in 2017 (11). These exponential broiler meat demands are an alarm to boost production.

Antibiotics have been used for many decades in the poultry industry to enhance production, promote growth performance, and protect birds from pathogenic microbes (12–16). For example, supplementation of broilers' diet with antibiotics could increase body weight gain by 5.8 % (17). This improvement was explained by improved appetite and feed conversion efficiency, stimulation of the immune system, and increased vitality and regulation of the intestinal microflora (18).

Antibiotics are also important for fighting infectious pathologies (16, 19, 20) such as necrotic enteritis and coccidiosis (21). Broadly, antibiotics are used in phytosanitary treatments, feed additives, and prophylactic treatments in animals and humans.

Despite its important role, improper uses of antibiotics in animal farming have been reported to increase antimicrobial resistance bacteria as a public health threat (22–24), residues in animal products, and cause environmental pollution (25, 26). Consequently, the use of antibiotics as growth promoters was banned by the European Union in 2005 (27) and China in 2020 (28). To minimize health risks, consumers have great preferences for conventional broiler meat, resulting in shift to antibiotic-free broiler meat production around the globe (13, 14). The ban on antibiotic use, combined with consumers' preferences, provoked scholars to look for antibiotic alternatives (29). This is important to apply sustainable feeding strategies of potential antibiotic alternatives for increasing antibiotic-free broiler meat production (30, 31). Therefore, this review aimed to explain the current use scenario, mode of action, ameliorative effects, and feeding strategies of different antibiotic alternatives including phyto-genic groups (marine algae, herbs, plant extract, and essential oils), prebiotics, probiotics, and enzymes in broiler production.

CURRENT SCENARIO OF ANTIBIOTIC USE

The intensity of using antibiotics could vary among nations (32). China is among the world's leading antibiotic producers and consumer, particularly in livestock products (4, 33). This was supported by Ziping (34), who reported that antibiotic use in China is 5 times higher than the international average. Although antimicrobial use in animal production in China increased until 2014, it has fallen in recent years (34). Antimicrobial consumption is projected to be 67% by 2030 and nearly double in Russia, Brazil, China, India, and South Africa (4).

Although antimicrobial consumption in livestock has received little attention, an expert opinion suggests that global consumption of antimicrobials in animals is twice more than in humans (4, 35). In many countries, most commercial broiler producers have reported antibiotic use, i.e., in Ghana (97%) (16), Nepal (90%) (36), Nigeria (89%) (37), Bangladesh (98%) (38), and the United States (40%) (39). Broiler farm intensification could be a driving force for the use of antibiotics as feed additives in developed countries, whereas increasing demand for poultry meat and eggs for food security could be a factor

in the developing world and may lead to the risk of developing antibiotic-resistant microbes (40–43).

Globally, the most commonly applied antibiotics to food animal production include tetracyclines, sulfonamides, and penicillins (44). However, this review finds that there are differences in using antibiotics types in different nations that might be due to antibiotic-producing capability, access, price, and banned antibiotics policy platform. Tetracycline, aminoglycosides, penicillins, and fluoroquinolones in Ghana (45), tetracycline, penicillins, and sulfonamides in South Africa (46), bacitracin, tylosin, tetracycline, salinomycin, virginiamycin, and bambarmycin in North America (29), and erythromycin, penicillins, tylosin, tetracycline, and vancomycin in China (34) are commonly used antibiotics.

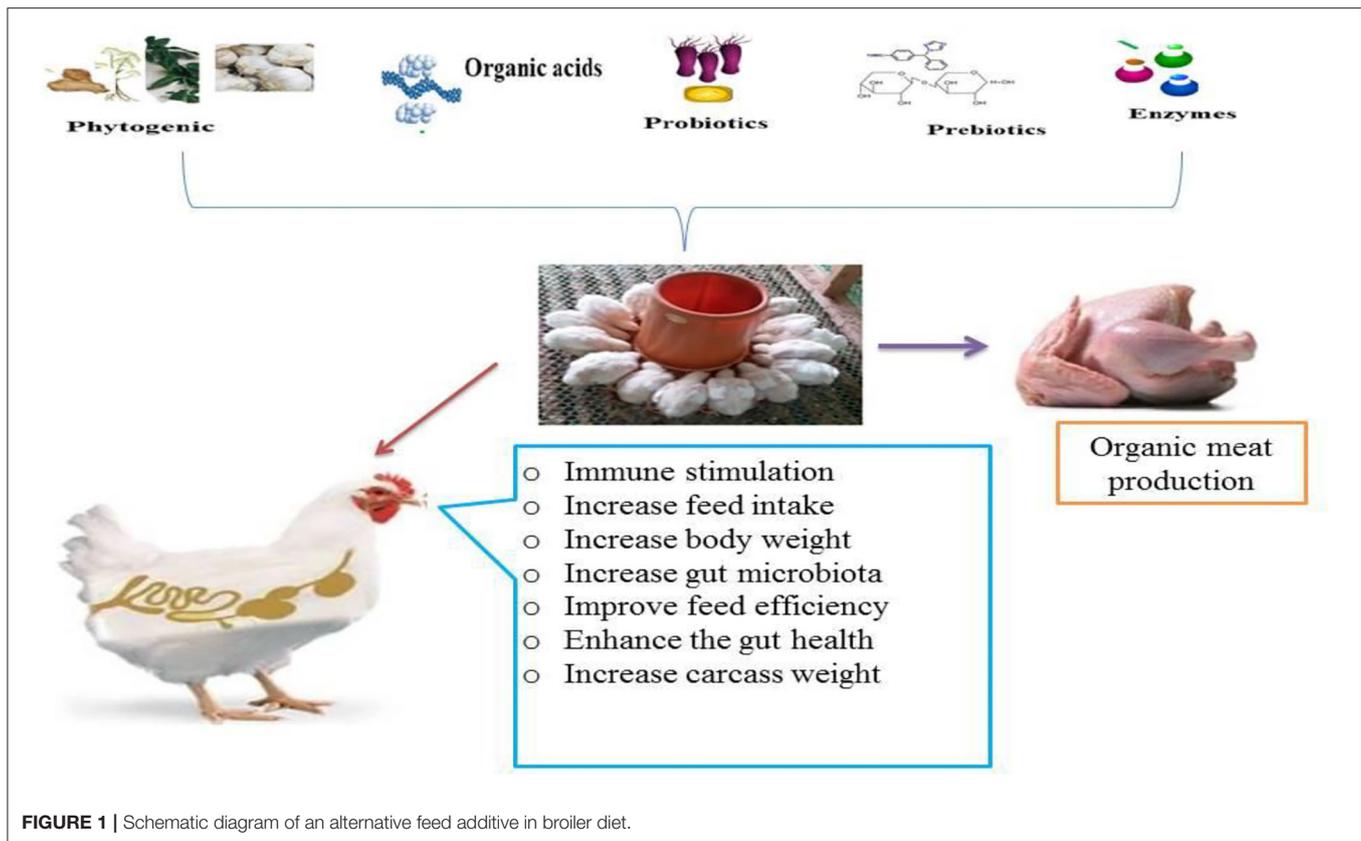
Although the use of antibiotics has ameliorative effects as mentioned above, it has been banned for a decade in different countries because of potential development of antibiotic-resistant human pathogenic bacteria (15, 47, 48). The European Union (EU) has banned non-therapeutic antibiotics used as growth promoters and feed additives in animal production since 2006 (42). Although the ban was applied before a decade and consumers have preferred organic livestock products, antibiotics are still used in livestock as growth promoters. Therefore, feed additives could be familiar as antibiotic alternatives in the poultry production sector, with a great interest in improving growth performance and feed conversion ratio, maintaining healthy intestinal microbial populations, and improving the overall health of birds (20, 49–52).

FEED ADDITIVES AS ANTIBIOTIC ALTERNATIVES

Feed additives are non-nutritive natural products added to basal diet as minor components of the diet to improve feed quality and food from animal origins and improve animal performance and health. They also promote ingestion, absorption, nutrient assimilation, and growth of animals by affecting physiological processes such as immune function and stress resistance (53). It has been reported that feed additives could be used as antibiotic alternatives for broilers to reduce mortality rates and enhance performance without jeopardizing the environment and consumer health (20). The common feed additives tested in poultry are phyto-genic feed additive groups including essential oils (20, 51, 52, 54), herbal extracts (55–57), organic acids (58, 59), and others like prebiotics (15, 60), probiotics (15, 61), and enzymes (62–64) (Figure 1).

Phyto-genic Feed Additives

Phyto-genic feed additives are plant-origin extracted compounds that include a wide range of substances such as herbs, spices, botanicals, oleoresins, and essential oils that are used in poultry production (65–73) (Table 1). According to Madhupriya et al.' (68) explanation, PFAs are natural, less toxic, residue-free, and ideal feed additives for poultry compared with synthetic antibiotics.



Herbs are flowering plants whose stem does not become woody and persistent, and are valued for their medical properties, flavor, and scent (51), whereas spices are pungent or aromatic substances of vegetable origin that are used as seasonings and preservatives (51). Botanicals or photobiotics are parts of a plant like roots, leaves, and barks, which are used to make drugs for medical use. Essential oils are any class of volatile oils obtained from plants; they possess the odor and other characteristic properties of plants and are used chiefly in the manufacture of perfumes, flavors, and pharmaceuticals (51). The most widely used herbs and spices for Phyto feed additives in poultry production are oregano, thyme, garlic, horseradish, chili, cayenne, pepper, peppermint, cinnamon, anise, clove, rosemary derivatives, citrus, and sage (68, 82).

A growing body of evidence has shown that supplementation of phytogetic feed additives in broilers' diet improve intestinal functions (83, 84), increase nitrogen retention and fiber digestibility, enhance growth performance (85), reduce inflammation (86), and improve anti-oxidative (51, 87) and antimicrobial activities (88) (Table 1). Altogether, the above findings suggest that PFAs have beneficial effects to improve performance and broiler health (54, 73, 75–77, 89, 90).

Phytogetic Mode of Action

Studies have shown that the growth and health-promoting effects of PFAs are associated with their biological activities including antimicrobial, antioxidant, immunomodulatory, and

anti-inflammatory (54, 68, 91–93). For instance, Superliv concentrate premix (SCP), AV/HGP/16 premix (AVHGP), and bacteriostatic herbal growth promoter (BHGP) have been increasing the feed efficiency of broilers by modulation of the muscle mTOR pathway and hepatic lipolytic programs; thus, they are promising for muscle protein synthesis and hepatic lipogenesis reduction (94). This is aligned with (95, 96) that have shown that PFAs modulate the expression of feeding-related hypothalamic neuropeptides and result in feed efficiency (FE) improvement. FE is also controlled by peripheral intermediary metabolism like lipid metabolism and protein synthesis-associated signaling pathways, which are modulated by bioactivities of PFAs.

PFAs also improve the palatability, digestibility, absorption of the feed nutrients, control animal intestinal microbiome structure, improve performance and feed quality through positively reflected of biological activities of plant secondary compounds with the action of antioxidative properties and slow microbial growth in poultry (97–99). In addition, they have been shown to enhance gut health by reducing bacterial colony populations, lessening fermentation products including ammonia and biogenic amines, decreasing the activity of the gut-associated lymphatic system, and increasing prececal nutrient digestion. Beneficial phytogetic compounds derived from their bioactive molecules are carvacrol, thymol, cineole, linalool, anethole, eugenol, capsaicin, allicin, allyl isothiocyanate, and piperine (65, 68). Most of these active secondary plant

TABLE 1 | Phytogetic feed additives and ameliorative effects on broiler production.

Feed additives	Level of supplementation	Findings	Sources
Essential oils (Origanum genus)	300–600 g/kg	Increase in the average daily gain	(74)
Cinnamon	2 g/kg	Improve growth performance	(75)
Lippia Javanica leaf meal	5 g/kg	Improve daily gain and slaughter weight	(76)
The mixture of garlic and black pepper powder	5 g/kg) and 1 g/kg)	Increase in weight gain	(77)
Pennyroyal (Mentha pulegium L.)	2%	Increase in average daily gain	(78)
Neem (Azadirachta indica)	7 g/kg	Favorable influences on the immune	(79)
B. subtilis with enramycin	UBT-MO ₂ /kg	Increase in body weight and relative weight of the thymus	(80)
Milk kefir	2%	Improvement on body mass and chicken consumption index	(81)

metabolites belong to the classes of isoprene derivatives, flavonoids, and glucosinolates, which act as antibiotics or antioxidants (100, 101).

Organic Acids as Feed Additives

Organic acids are weak acids that have a carboxylic acid group (R-COOH) and nutritional values and antimicrobial effects in animal feeds (102–104). Organic acids have been used in animal feeds for many years because of the ban on the use of antibiotics (59). In line with these findings (15, 105, 106) reported that organic acids are considered as effective antibiotic alternatives in animal feeds. The most commonly used organic acids in the broilers' diet are acetic, butyric, citric, formic, propionic, malic, tartaric, and lactic acids (15, 28, 107).

The inclusion of organic acids in the broilers' diet has been shown to improve protein and carbohydrate digestibility (108), fight against pathogenic bacteria (105), and (106) enhance the feed conversion rate, nutrient utilization, and growth rate of broilers (109, 110).

Organic Acid Mode of Action

Diets with poor protein quality have more indigestible proteins reaching the GIT, which end up with high protein fermentation (111). This high protein fermentation causes discomfort in the animal body and negatively affects its growth rate because of high volatile fatty acids and ammonia and production of other gases (112). Organic acids are good supplement alternatives in such types of feed to acidify the GIT environment (113) and improve

nutrient utilization, which results in activeness of the protease enzyme. For example, Suiryranrayna and Ramana (114) reported stimulation of protein digestion by converting pepsinogen to pepsin by supplementation of organic acids. Moreover, organic acids reduce pH in the GIT, which enhances pepsin activity, and increases the digestibility of nitrogen, phosphorus, and other minerals (15, 115). These acid anions react with calcium, phosphorus, magnesium, and zinc, thus enhancing their digestibility. Peptides produced by pepsin proteolysis stimulate the release of gastrin and cholecystokinin hormones, which regulate protein digestion and absorption (116, 117).

Organic acids have been used as feed preservatives for protecting feed from microbial and fungal deterioration with the mechanism of acidification (118). These are a powerful tool in maintaining the health of the gastrointestinal tract of poultry, resulting in improvement in birds' production performance. For example, sanguinarine suppresses the growth of some harmful acid intolerance bacteria such as *E-coli*, *Salmonella spp.*, and *Clostridium perfringens* that cause gastrointestinal distress (119), resulting in enhanced appetite and feed intake and improving growth (120). Reduction of competition for microbial nutrients in the host thereby increases the availability of nutrients (121), consequently increases BWG, and improves FCR (122, 123). Organic acids also affect the histological structure of the gastrointestinal tract; Consequently, improve nutrient absorption, maximized nutrient utilization efficiency, and improved growth performance (54). As a conclusion remark from different studies; organic acids and their salts are used to reduce a load of pathogenic microorganisms in the intestine, activate digestive enzymes, improve digestibility, and increase the absorption of nutrients, gut microflora function, and performance of chickens (Table 2).

Prebiotics Feed Additives

Prebiotics are indigestible carbohydrates by the host animal but can be utilized by useful GIT microorganisms (54, 141–143). Prebiotics are found in different food sources such as oats, barley, dandelion greens, chicory, chia seeds, flax seeds, onion, garlic, almonds, and artichoke (144). Green algae (Chlorophyta) are also considered prebiotic because of the presence of water-soluble sulfated polysaccharides; the perform gut microbiota modulation and immunomodulation, and they have antioxidant, antibacterial, anti-hyperlipidemia, and anti-diabetic properties (145).

Potential prebiotics that have been fed to broilers include fructan, oligofructose, inulin, fructooligosaccharides, galactan, galactooligosaccharides, xylooligosaccharides (XOS), pectin, fiber components, and milk oligosaccharides (146–149). Refined functional carbohydrates (RFCs) including mannan-oligosaccharides (MOSSs), β -glucan, and D-mannose, which are derived from the cell wall of *Saccharomyces cerevisiae*, are a readily available source of prebiotics for animal use (150). From these, mannan-oligosaccharides and fructooligosaccharides are the most common commercial feed nutrients in poultry feed production (151). In connection with their economic importance for producers, prebiotics also have no residual effect and do not develop any resistance for broiler product consumers (141).

TABLE 2 | Organic acids, their derivatives, and ameliorative effects on broiler production.

Organic acids	Level of supplementation	Finding	Sources
CA	2%	Improve epithelial cell proliferation and villi height of gastrointestinal tract	(124)
CA, avilamycin	0.5 and 0.001%, respectively	Significantly increase growth performances at 35 days	(125)
BA	0.2%	Increase CW, breast meat yield, FCR, dressing % and reduce abdominal fat	(126, 127)
SB	0.6 and 1.2 g/kg	Increase ADG and FCR during 1–21 days period	(123)
N-butyric acid and 50% MB	250–7,000 mg/kg	Reduce <i>Salmonella Typhimurium</i> or <i>Clostridium perfringens</i>	(128)
MESB	800 mg/kg	Higher total body weight, daily gain and FCR at 35 days	(129)
PCB	0.3 g/kg	Increase weight gain	(130)
FA	5 g/kg	Increase BWG, dressing percentage and reduce FCR	(131)
KDF	5 g/kg	Increase BWG, dressing percentage and reduce FCR	(131)

FCR, feed conversion ratio; CW, carcass weight; SB, sodium butyrate; MESB, microencapsulated sodium butyrate; ADG, average daily gain; PCB, protected calcium butyrate; CA, citric acid; BA, butyric acid; FA, formic acid; KDF, potassium di-formate; BWG, body weight gain.

Supplementation of prebiotics can improve growth performance and antibody titer against infectious bursal disease in broilers (Table 3) (133). Prebiotics are also useful for changing the microbial population of the intestine (31, 149, 152, 153); for example, dietary MOS (1g/kg) increase *Lactobacillus* and *Bifidobacterium* contents (154), increase the length of the villain (155), prevention of colon cancer, minimize disease-causing bacteria and increases daily weight gain (156, 157), and medical therapy of broiler (158). Generally, the beneficial effects of prebiotics are alteration of gut microorganisms that enable to increase their numbers, increase digestibility, reduce pathogenic bacteria, increase mineral and vitamin absorbability, maintain optimal intestinal pH, and maximize nutrients utilization (142, 143, 159).

Prebiotics Mode of Action

Prebiotics can affect host health in different ways, such as production of metabolites like lactic acid, microbial metabolism modification, and increase in epithelium cell integrity (160, 161). Prebiotics are used to modulate the ecosystem of gut elements including alteration of the intestinal microbiota, stimulation of the immune system, improvement of the epithelium, and

TABLE 3 | Prebiotics and their ameliorative effects on broiler production.

Prebiotics	Level of supplementation	Finding	Sources
FOS	0.25%	Improve productivity of broiler Increase lactobacillus in the ileum	(132)
MOS	0.05%	Improve productivity of broiler Increase lactobacillus in the ileum	(132)
MOS	1.5 g/kg	Improve WG and FCR Improve the antibody titer against IBD	(133)
IMO	5–10 g/kg	Improve WG Increase feed conversion rate Increase the caecal populations of lactobacilli and bifidobacteria Decrease the caecal <i>Escherichia coli</i> Increase the caecal VFA	(134)
RFC	50–100 g/t	Improve ADG Decrease cecal <i>Campylobacter</i> counts The high dose also increases FBW	(135)
Autolyzed WY and YCW	1.5–2 g/kg	Improve BWG, FCR, and Meat yield Positive effect on ileal protein digestibility as well as trypsin and chymotrypsin activities	(136)

ADG, average daily gain; FBW, final body weight; FOS, fructo-oligosaccharide; IMO, isomalto-oligosaccharide; IBD, infectious bursal disease; MOS, mannan-oligosaccharide; RFC, refined functional carbohydrate; VFA, volatile fatty acid; WY, whole yeast; YCW, yeast cell wall product.

regulation of interaction between the host and the intestinal microbiota (162).

Prebiotics could be a selective substrate for a limited number of beneficial bacteria to alter the colon microflora in favor of a healthier gastrointestinal environment (149, 152). For example, they serve as a substrate for endogenous beneficial bacteria, thus promoting competitive exclusion of pathogenic microbes and selective colonization by beneficial microbes (60). Mazanko et al. (159) also reported that a prebiotic feed supplement creates an unfavorable condition for pathogenic organisms by altering the pH of the intestine. It establishes a healthy microbial community in the intestine of broilers by enhancing the abundance of *Lactobacilli* and *Bifidobacteria* and reducing the titers of Coliform (163, 164). *Bifidobacterium* and *Lactobacillus* have manase enzymes; they selectively bind mannan oligosaccharides only for harmful bacteria, which normally do not have this enzyme (157). The effect of mannan oligosaccharides on broilers is increase in the daily weight gain of broilers by 4–8% (156, 157).

The sustainable ability of prebiotics in acidic environments and to remain resistant to distinct digestive enzymes in the

small intestine make them an extraordinary tool to boost the growth of beneficial gut microbes that ferment them, leading to production of short-chain fatty acids, vitamins, and other fragmented molecules or some antibacterial substances such as bacteriocin against pathogenic microorganisms (165, 166). These fermented products of beneficial microbes due to prebiotic feed additives also improve the integrity of intestinal epithelial cells, which further increase the absorption of nutrients and enhance the growth performance of animals (115, 162).

The modulation of the intestinal microbiota with prebiotic feed additives is associated with immune responses (162) (Figure 2). Oligosaccharides have been reported to present immunomodulatory beneficial effects on the gut, such as modifying clearance efficiency of pathogenic bacteria, activating T cell-dependent immune responses, and repression of pro-inflammatory cytokines (167, 168). Inhibiting pathogen colonization with prebiotics can decrease pathogen-associated molecular patterns, which are produced by pathogenic microorganisms (169). The produced molecule can be recognized by pattern recognition receptors (PRRs), including toll-like receptors (TLRs) and NOD-like receptors (NLRs), which are expressed on the surface of sentinel cells (170, 171). Once pattern recognition receptors (PRRs) recognize pathogen-associated molecular patterns (PAMPs), sentinel cells such as macrophages, epithelial cells, dendritic cells, and mast cells are activated and produce cytokines for regulation of further innate immune

responses (171). They can be recognized by receptors of immune cells, consequently modulating host immunity systems.

Probiotics Feed Additives

Probiotics are “live strains of strictly selected microorganisms that, when used in adequate amounts, confer a health benefit on the host” (172). Similarly, probiotics are beneficial bacteria that can fight pathogens in the gastrointestinal tract of chickens like subclinical necrotic enteritis (173), stimulate growth (174–176), and improve the immunity of the host (143, 177–179). Probiotics strains have been also providing feeding efficiency improvement, intestinal protection, antioxidant capacity and apoptosis (180), use of nutrients (181), energy digestibility, disappearance of non-starch polysaccharides (182), and microbial profile of cecum and litter (Table 4) (183).

Selection and use of microorganisms as feed additives are not an easy task; their risks, handling procedures, and adaptability to the environment should be considered. Some microbes will participate in the spread of antibiotic resistance (*enterococcus*) and produce toxin substances (*Bacillus cereus* strains) (184). The recommended dose for most probiotic strains is 10×9 colony-forming units of feed (CFU/KG). Care should also be taken when mixing probiotics. The water should be free from any disinfectant or chlorine. Administration or offering of a probiotic feed additive solution should be within 6–12 h after mixing with water. If animals are on antibiotic treatment, it is

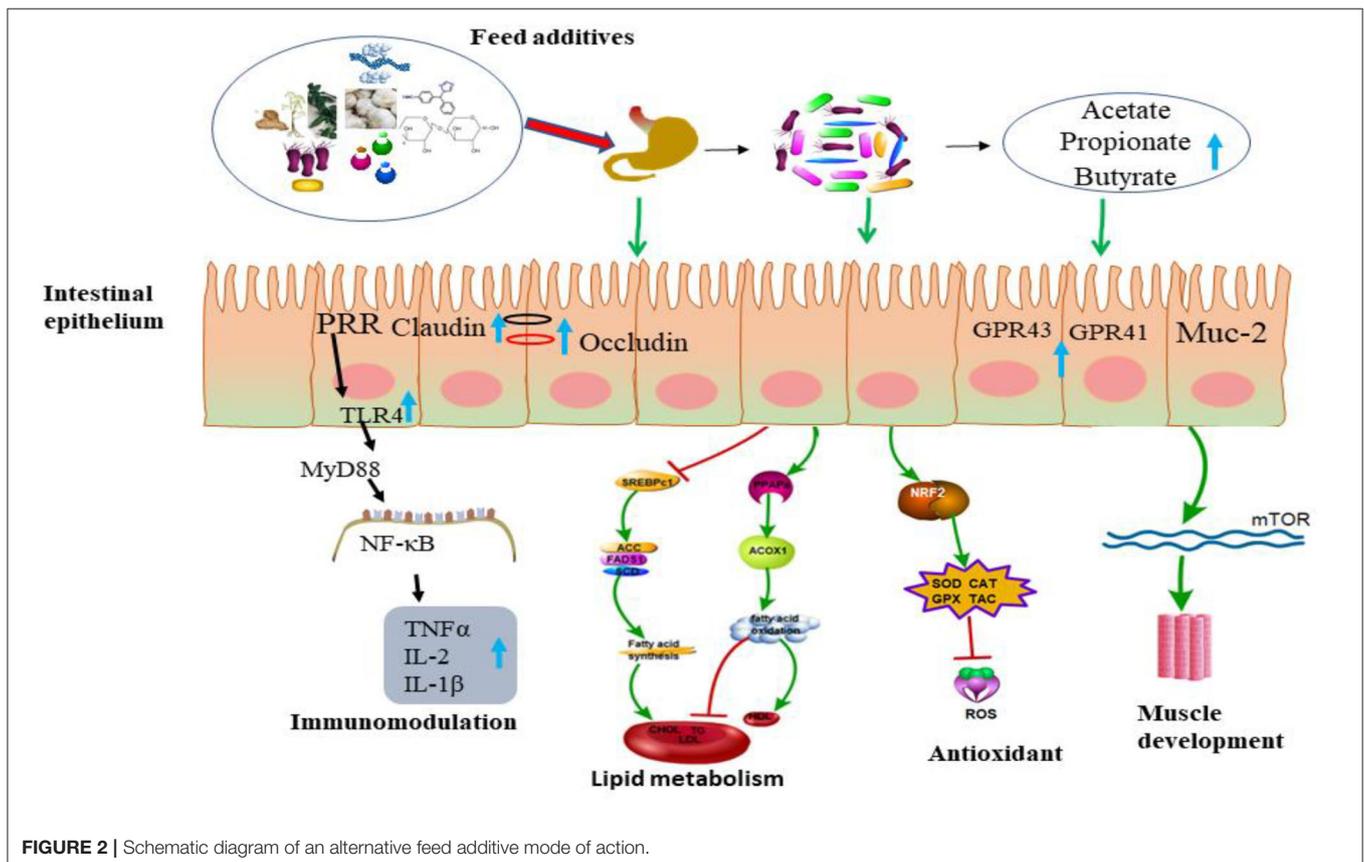


TABLE 4 | Probiotics and their ameliorative effects on broiler production.

Probiotics	Level of supplementation	Finding	Sources
Mixture of <i>Bacillus licheniformis</i> and <i>Bacillus subtilis</i> spores	0.05%	Significantly improve the FCR	(137)
Multi-strain probiotic (11 <i>Lactobacillus</i> strains)	1 g/kg	Increase FCR Improve BWG Increase the caecal populations of lactobacilli and bifidobacteria Increase the caecal VFA Decrease the caecal <i>Escherichia coli</i>	(134)
Protexin	2 g/kg	Improve the growth performance	(133)
Promax	1 g/L	Improve BW and the hemato-biochemical profile	(138)
Normosil	1 mL/kg	Increase the average daily gain Increase the level of blood erythrocytes Improve carcass quality	(139)
<i>Lact. lactis</i> and <i>L. plantarum</i>	10 ⁹ cfu/mL and 10 ¹² cfu/mL, respectively	Lower the serum cholesterol, triglyceride, and total lipid contents Increase contents of blood glucose and total protein	(140)

BWG, body weight gain; FCR, feed conversion rate; VFA, volatile fatty acid.

highly recommended that the treatment be withdrawn 24–48 h before administering probiotics (185).

The most used microorganisms as feed additives in poultry production are bacterial strains, mostly Gram-positive *Bifidobacterium*, and lactic acid bacteria groups such as *Bacillus*, *Enterococcus*, *Lactobacillus*, *Pediococcus*, *Streptococcus*, *Aspergillus*, *Candida*, and *Saccharomyces*. However, fungi and yeast strains are also used, mainly from the species *Saccharomyces cerevisiae* and *Kluyveromyces* (184, 186–188).

Probiotics Mode of Action

The main modes of action of probiotics include antagonistic action toward pathogenic bacteria by secreting products that inhibit their development such as bacteriocins, organic acids, and hydrogen peroxide, and competitive exclusion by competing with bacteria for locations in the intestinal mucous membrane to adhere to nutrients (47). Lowering the gut pH through the volatile fatty acids and organic acids produced during probiotic product breakdown is the most common probiotic mode of action (189, 190). The low pH in the intestine suppresses the colonization of pathogens in the digestive tract, thereby competitively inhibiting the effects of pathogens (191). Probiotics are also used to modulate the intestinal microbiota, for immunomodulation, and to improve intestinal

integrity (192, 193). Other principal mechanisms of probiotics are competition for binding sites where probiotics adhere to the intestinal epithelium wall, hindering competition and joining of pathogenic microorganisms; this higher concentration of the beneficial microbiota is also the driving force to have an advantage in the competition for nutrients (20). Findings showed that probiotics have nutritional effects, increasing fiber digestion and enzymatic activity in birds to be efficient in feed nutrient utilization (133). The finding of Wang et al. (194) stated that supplementation of broilers with *Bacillus subtilis* in the diet was more effective in performance in heat stress conditions through the immunity modulated by the microbiota.

Enzymes as Feed Additives

Enzymes are catalysts of biochemical processes that are composed of proteins, amino acids with minerals, and vitamins (195). Enzymes are the most important and useful additives in the animal feed industry (196). They can be obtained from plants, animals, and microorganisms (197). Enzymes, as feed additives in broiler production, are produced by fermentation of fungi and bacteria and are used for maximization of feed conversion efficiency (FCE) (15). Although animals produce endogenous enzymes that are involved in digestion, they do not efficiently degrade feedstuff and take advantage of all their nutritional components; therefore, exogenous enzymes are supplemented to increase animal performance (195, 196, 198). Pectinases, amylases, cellulase, galactosidases, β glucanases, xylanases, associated enzyme phytases, proteases, and lipases are commonly used exogenous enzymes in the animal feed industry (Table 5) (196, 197, 207). These exogenous enzymes are mainly used in monogastric animals like poultry and swine (208).

The supplementation of enzymes for broilers has nutritionally, economically, and environmentally justifiable advantages (209). The use of enzymes in the chicken diet resulted in high feed utilization efficiency, reduction of digesta viscosity, enhanced digestion and absorption of nutrients, and increased feed intake and weight gain (18, 196, 210, 211). Xylanase has increased crude protein digestibility, feed intake, nitrogen and fiber absorption, and weight gain in broilers (211, 212). Phytases increase the utilization of phytate phosphorus in feeds (210). A multi-enzyme complex (Avizyme) composed of xylanases, proteases, and amylases is used to improve nutritional quality, reduce the viscosity of diets, increase body weight, decrease mortality, and increase the amount of net energy (213). It also improves the intestinal health of animals (214). Generally, different studies have reported that the use of exogenous feed enzymes in poultry diets is becoming familiar to overcome the adverse effects of anti-nutritional factors, and improve the digestion of dietary components and bird performance.

Enzymes must be active under physiological conditions prevailing in the animal's digestive tract and must complement the characteristics of dietary ingredients and additives to realize their functions (209, 215).

TABLE 5 | Enzymes, target substrates, and their benefits in broiler production.

Broad classes of enzymes	Specific example	Substrate	Target feedstuff	Level of supplementation	Ameliorative Effect	Sources
Carbohydrases	Xylanases	Arabinoxylans	Wheat, rye, triticale, barley, fibrous plant materials	3,200–24,000 IU/kg	Increase starch and nitrogen digestibility and improve AIDE	(199)
	α -Galactosidases	Oligosaccharides	Soybean meal, grain, legumes	50 mg/kg of diet	Improves intestinal histology and morphology	(200)
	α -amylase	Starch	Cereal grains, grain legumes	300–2,250 IU/kg	Improve the apparent ileal digestibility of energy	(201)
	β -Glucanases	β -Glucan	Barley, oats, and rye	20 IU/	Reduce viscosity, increases dry matter of digesta, and available energy	(202) (203) (204)
	β -Mannanase	Cell wall matrix (fiber components)	Plant-derived ingredients, fibrous plant materials	200–400 mg/kg		
	Cellulases Hemicellulases Pectinases			20 IU/kg 20 IU/kg 53 IU/ kg		
Proteases	Proteases	Proteins	All plant protein sources	30,000 IU/kg	Increase FI and FCR, increase N retention, reduce abdominal fat	(205)
Phytases	Phytates	Phytic acid	All plant-derived ingredients	500 – 1,500 FTU/kg	Increase FI, BW, FCR, CW, and GIT organs length	(206)

AIDE, apparent ileal digestible energy; BW, body weight; CW, carcass weight; FCR, feed conversion ratio; FI, feed intake.

Enzyme Mode of Action

Each enzyme has a different and interdependent mode of action; its use in combination with feed formulations must be carried out carefully to achieve maximum ameliorative effects (197). Broiler diets containing a large amount of NSP lead to increased digesta viscosity, thus depression in growth performance (216). Carbohydrase enzymes are added to broiler diets to overcome this type of difficulty, consequently improving nutrient utilization and increasing the productivity of birds. For example, hydrolysis of non-starch polysaccharides (NSPs) into smaller oligosaccharides with carbohydrase results in decrease in digesta viscosity and release of encapsulated nutrients (217). Produced small oligosaccharides during NSP hydrolysis could also have a prebiotic advantage (218). The hemicellulose in agro-industrial byproducts, particularly Palm kernel expeller (PKE), is partially hydrolyzed with enzyme treatment, thus obtaining oligosaccharides (DP<6) that have prebiotic-like effects. Based on Chen et al.'s (219) results, the untreated PKE contained 20.93 g/kg oligosaccharides, but after treatment, the oligosaccharide content increased to 28.91 and 59.71 g/kg for PKEENZ and SPKEENZ, respectively. Zhang et al. (220) also reported that smaller oligosaccharides such as xylooligosaccharide (XOS) come from hydrolysis of NSPs, which have been shown to have prebiotic-like effects.

Enzymes act on nutrients having main effects on substrates to which they are directed as well as having side effects. They initiate

and control the rate of biological reactions by which substrates are changed into useful products (195). NSP hydrolysis products are fermented by beneficial bacteria such as *Bifidobacter* and *Lactobacilli* spp., thus, producing short-chain fatty acids (221). Increased SCFA concentration is often associated with increase in the population of beneficial bacteria and decrease in pathogenic bacteria (19). Some SCFAs are also used as an available energy source to the host for growth (222).

Supplementing glucose oxidase (GOD) in broilers has been reported to increase daily body weight gain, improve meat quality, and enhance digestive ability that is indicated by the nutrients' apparent digestibility and digestive enzymes (223). Different studies also confirm that the increase in body weight gain and FCR of broilers with commercial enzymes is due to the ileal digestibility of crude proteins (224), starch and fat (225), and improvement in ileal non-starch polysaccharide (NSP) digestibility (226). The content of secreted immunoglobulin A and transepithelial electrical resistance are also increased with the GOD supplement, which indicated an enhanced gut barrier. In the general context, dietary GOD supplement could improve the growth performance of broilers in two main mechanisms: 1) by enhancing the digestive function of the gut, which concluded from improved nutrients' apparent digestibility and digestive enzyme, and 2) by increasing the abundance of beneficial bacteria such as *F. prausnitzii*, *Ruminococcaceae*, and *Firmicutes* (223).

CONCLUSION AND FUTURE RESEARCH DIRECTIONS

The ban on certain antibiotics has promoted phytochemicals, organic acids, prebiotics, probiotics, and enzymes as alternatives in broiler production. Antibiotic alternatives have comparable advantages to antibiotics to enhance the production performance and well-being of broilers without human health challenges. Moreover, using antibiotic alternatives can increase body weight, average daily gain, carcass weight, feed conversion ratio, and the nutritive value of feed ingredients, and enhance the gut health of broilers. The main provided effects of alternative feed additives includes immune-modulating, enhance digestion, improving nutrient availability, increase absorbability of nutrients, antimicrobial, antioxidant activity, enhancement of gut integrity, intestinal barrier function or improve intestinal health, nutrient for the host, and modulating the host gut microflora. These different modes of action suggest that there could be symbiotic, antagonistic, and synergistic or combative effects between alternatives or other feed nutrients. Therefore, use of alternative feed additives in broiler production should

highly promoted and further investigations on interaction effects of combined additives, sub-additive, and with diet nutrient, efficiency of utilization, and level of inclusion could be mandatory.

AUTHOR CONTRIBUTIONS

HA carried out the organization and drafting of the manuscript. HZ and TW were involved more in technical editorial support of the drafted manuscript. All authors participated in the evaluation, editing, and approval of the final version of the manuscript.

FUNDING

This study was funded by the Shandong Key Science and Technology Innovation Program (2019JZZY010704), the China Agriculture Research System-Beijing Team for Poultry Industry, and the Agricultural Science and Technology Innovation Program (ASTIP).

REFERENCES

- Chowdhury EU, Morey A. Intelligent packaging for poultry industry. *J App Poul Res.* (2019) 28:791–800. doi: 10.3382/japr/pfz098
- Alexandratos N, Bruinsma J. World Agriculture Towards 2030/2050: The 2012 Revision Agricultural Development Economics Division, Food and Agriculture Organization of the United Nations. Rome, Italy (2012). 1–147. Available online at: <https://www.fao.org/3/ap106e/ap106e.pdf>
- FAO (Food and Agriculture Organization). *Gateway to Poultry Production and Products*. Paris: 26th World's Poultry Congress (2022) (accessed December 22, 2022).
- Van Boeckel TP, Charles B, Marius G, Bryan TG, Simon AL, Timothy PR, et al. Global trends in antimicrobial use in food animals. *Proc Natl Acad Sci USA.* (2015) 112:5649–54. doi: 10.1073/pnas.1503141112
- Mekonnen MM, Neale CM, Ray C, Erickson GE, Hoekstra AY. Water productivity in meat and milk production in the US from 1960 to 2016. *Environ Int.* (2019) 132:1–12. doi: 10.1016/j.envint.2019.105084
- Newsire Globe. *Global Poultry (Broiler) Market Analysis & Forecast 2019–2023: Production, Consumption, Import, and Export*. France: The World's Largest Market and Research Store (2019).
- Najeeb AP, Mandal PK, Pal UK. Efficacy of fruits (red grapes, gooseberry and tomato) powder as natural preservatives in restructured chicken slices. *Int Food Res J.* (2014) 21:2431–6. Available online at: <http://www.ifrj.upm.edu.my>
- Petracci M, Mudalal S, Soglia F, Cavani C. Meat quality in fast-growing broiler chickens. *World's Poul Sci J.* (2015) 71:363–74.
- Faostat (2019). Food and Agriculture Organization of the United Nations, 2019. Production: Crops. Available online at: <http://faostat.fao.org>
- FAO. Where is per capita poultry meat consumption highest? *Poultry Int.* (2018).
- AVEC annual report. *EU28 Poultry Meat Export Trade, Main Tariff Lines*. EU28 Poultry Meat Export Association of Poultry Processors and Poultry Trade in the EU Countries ASBL (2018). p. 1–37.
- United States Department of Agriculture. *Restrictions on Antibiotic Use for Production Purposes in U.S. Livestock Industries Likely To Have Small Effects on Prices and Quantities*. Economic Research Service (2019).
- Dianna VB, Kim MW. Antibiotic-Free Production and Broiler Chicken Meat Safety. Available online at: <https://www.food-safety.com/articles/5971-antibiotic-free-production-and-broiler-chicken-meat-safety>. (2018) (accessed December 22, 2021).
- Hakimul H, Subir S, Shariful I, Aminul I, Rezaul K, Mohammad EHK, et al. Sustainable antibiotic-free broiler meat production: current trends, challenges, and possibilities in a developing country perspective. *Biology.* (2020) 9:1–24. doi: 10.3390/biology9110411
- Lethogonolo AS, Zahra MH, Tlou GM, Monnye M. The current status of the alternative use to antibiotics in poultry production: an African perspective. *Antibiotics.* (2020) 9:1–18. doi: 10.3390/antibiotics9090594
- Paintsil EK, Ofori LA, Akenten CW, Fosu D, Ofori S, Lamshöft M, et al. Antimicrobial usage in commercial and domestic poultry farming in two communities in the Ashanti region of Ghana. *Antibiotics.* (2021) 10:1–9. doi: 10.3390/antibiotics10070800
- Rahman MA, Parvin MS, Sarker RR, Islam MT. Effects of growth promoter and multivitamin-mineral premix supplementation on body weight gain in broiler chickens. *J Bangladesh Agril Univ.* (2012) 10:245–8. doi: 10.3329/jbau.v10i2.14914
- Peric L, Zikic D, Lukic M. Application of alternative growth promoters in broiler production. *Biotech Anim Husb.* (2009) 25:387–97. doi: 10.2298/BAH0906387P
- Engberg RM, Hedenann MS, Steinfeldt S, Jensen BB. Influence of Whole Wheat and Xylanase on Broiler Performance and Microbial Composition and Activity in the Digestive Tract. *Poultry Science.* (2004) 82:925–38. doi: 10.1093/ps/83.6.925
- Mehdi Y, Létourneau-Montminy MP, Gaucher M, Chorfi Y, Suresh G, Rouissi T, et al. Use of antibiotics in broiler production: Global impacts and alternatives. *Anim Nutr.* (2018) 4:170–8. doi: 10.1016/j.aninu.2018.03.002
- CSCRA. Canadian Antimicrobial Resistance Surveillance System—2016 Report. Government of Canada, France. (2016).
- Nhung NT, Chansiripornchai N, Carrique-Mas JJ. Antimicrobial resistance in bacterial poultry pathogens: a review. *Front Vet Sci.* (2017) 4:1–17. doi: 10.3389/fvets.2017.00126
- Christy M-L, Sampson M, Edson M, Anthony O. Antibiotic use in agriculture and its consequential resistance in environmental sources: potential public health implications. *Molecules.* (2018) 23:1–48. doi: 10.3390/molecules23040795
- Oniciuc E, Likotrafiti E, Alvarez-Molina A, Prieto M, Santos J, Alvarez-Ordóñez A. The present and future of whole genome sequencing (WGS) and whole metagenome sequencing (WMS) for surveillance of antimicrobial resistant microorganisms and antimicrobial resistance genes across the food chain. *Genes.* (2018) 9:1–28. doi: 10.3390/genes9050268

25. Carvalho IT, Santos L. Antibiotics in the aquatic environments: a review of the European scenario. *Environ Int.* (2018) 94:736–57. doi: 10.1016/j.envint.2016.06.025
26. Gonzalez RM, Angeles Hernandez JC. Antibiotic and synthetic growth promoters in animal diets: a review of impact and analytical methods. *Food Contr.* (2017) 72:255–67. doi: 10.1016/j.foodcont.2016.03.001
27. Dibner JJ, Richards JD. Antibiotic growth promoters in agriculture: history and mode of action. *Poult Sci.* (2005) 84:634–43. doi: 10.1093/ps/84.4.634
28. Melaku M, Zhong R, Han H, Wan F, Yi B, Zhang H. Butyric and citric acids and their salts in poultry nutrition: effects on gut health and intestinal microbiota. *Int J Mol Sci.* (2021) 22:1–17. doi: 10.3390/ijms221910392
29. Diarra MS, Malouin F. Antibiotics in canadian poultry productions and anticipated alternatives. *Front Microbiol.* (2014) 5:282. doi: 10.3389/fmicb.2014.00282
30. Chand N, Ihsanuddin, Khan RU. Replacement of soybean meal with yeast single cell protein in broiler ration: the effect on performance traits. *Pakistan J. Zool.* (2014) 46:1753–8.
31. Khan RU, Naz S, Dhama K. Chromium: pharmacological applications in heat stressed poultry. *Int J Pharmacol.* (2014) 10:213–317. doi: 10.3923/IJP.2014.213.217
32. Archawakulathep A, Kim CT, Meunsene D, Handijatno H, Hassim A, Rovira HR, et al. Perspectives on antimicrobial resistance in livestock and livestock products in Asian countries. *Wetachasan Sattawaphaet.* (2014) 44:5–13.
33. Yanhong JH, Benjamin JC. Reducing antibiotic use in livestock, China. *Bull World Health Organ.* (2020) 98:360–1. doi: 10.2471/blt.19.243501
34. Ziping Wu. Antibiotic use and antibiotic resistance in food producing animals in china. 13-15 November 2018. *OECD Conference Centre, Paris.* (2019). p. 1–32.
35. Aarestrup F. Sustainable farming: get pigs off antibiotics. *Nature.* (2012) 486:465–6. doi: 10.1038/486465a
36. Koirala A, Bhandari P, Shewade HD, Tao W, Thapa B, Terry R, et al. Antibiotic use in broiler poultry farms in Kathmandu valley of nepal: which antibiotics and why? *Trop Med Infect Dis.* (2021) 6:47. doi: 10.3390/tropicalmed6020047
37. Al-Mustapha AI, Adetunji VO, Heikinheimo A. Risk perceptions of antibiotic usage and resistance: a cross-sectional survey of poultry farmers in Kwara State, Nigeria. *Antibiotics.* (2020) 9:378. doi: 10.3390/antibiotics9070378
38. Imam T, Gibson JS, Foysal M, Das SB, Gupta SD, Fournié G, et al. Cross-sectional study of antimicrobial usage on commercial broiler and layer chicken farms in Bangladesh. *Front Vet Sci.* (2020) 7:576113. doi: 10.3389/fvets.2020.576113
39. Moal Aurélie. *Consistency Is Vital for Antibiotic-Free Poultry Production.* Paris: Adisseo. (2021).
40. Mousavi S, Ibrahim S, Aroua MK. Sequential nitrification and denitrification in a novel palm shell granular activated carbon twin-chamber upflow bioelectrochemical reactor for treating ammonium-rich wastewater. *Bioresour Technol.* (2012) 125:256–66. doi: 10.1016/j.biortech.2012.08.075
41. Shamlo R, Nasr J, Kheiri F. *Effects of various levels of pennyroyal (Mentha pulegium L) on carcass characteristics and serum cholesterol in broiler Res Opin Anim Vet Sci.* (2014) 4:453–7.
42. Asad S, Tahseen U, Sarzamin K, Rifat UK. Effect of organic acid supplementation on the performance and ileal microflora of broiler during finishing period, Pakistan J. Zool. (2012) 47:635–9.
43. Hayden DH, Karla AV, Lixin Z, A. Review of Antimicrobial Resistance in Poultry Farming within Low-Resource Settings. *Animals.* (2020) 10:1264. doi: 10.3390/ani10081264
44. Van Boeckel TP, Glennon EE, Chen D, Gilbert M, Robinson TP, Grenfell BT, et al. Reducing antimicrobial use in food animals. *Science.* (2017) 357:1350–2. doi: 10.1126/science.aao1495
45. Boamah VE, Agyare C, Odoi H, Dalsgaard A. Practices and Factors Influencing the Use of Antibiotics in Selected Poultry Farms in Ghana. *J Antimicrob.* (2016) 2:1–8. doi: 10.4172/2472-1212.1000120
46. Smith PW, Agbaje M, LeRoux-Pullen L, Van Dyk D, Debusho LK, Shittu A, et al. Implication of the knowledge and perceptions of veterinary students of antimicrobial resistance for future prescription of antimicrobials in animal health, South Africa. *J S Afr Veter Assoc.* (2019) 90:1–8.
47. Patterson JA, Burkholder KM. Application of prebiotics and probiotics in poultry production. *Poult Sci.* (2003) 82:627–31. doi: 10.1093/ps/82.4.627
48. Amaechi N, Amaeze PN. Effect of dietary chloroacetic acid as antibiotic replacer on the gastrointestinal microflora and gut morphology of weanling pigs. *Res Opin Anim Vet Sci.* (2012) 2:494–8.
49. Joerger RD. Alternatives to antibiotics: bacteriocins, antimicrobial peptides, and bacteriophages. *Poult Sci.* (2002) 82:640–7. doi: 10.1093/ps/82.4.640
50. Waldroup PW, Oviedo-Rondon EO, Fritts CA. Comparison of bio-mos® and antibiotic feeding programs in broiler diets containing copper sulfate. *Int J Poult Sci.* (2003) 2:28–31. doi: 10.3923/ijps.2003.28.31
51. Suganya T, Senthilkumar S, Deepa K, Muralidharan J, Gomathi G, Gobiraju S. Herbal feed additives in poultry. *Int J Sci Environ Technol.* (2016) 5:1137–45.
52. Qidong Z, Peng S, Bingkun Z, Ling k, Chuanpi X, Zhigang S. Progress on gut health maintenance and antibiotic alternatives in broiler chicken production. *Front Nutr.* (2021) 8:692839. doi: 10.3389/fnut.2021.692839
53. Jet SM, Florencia NS. Phytogetic feed additives as an alternative to antibiotic growth promoters in poultry nutrition. Advanced studies in the 21st century. *Animal Nutr.* (2021) 8:1–18. doi: 10.5772/intechopen.99401
54. Mohamed E, Mohamed T, Heba M, Amira M, Mohamed M, Gehan BA, et al. Alternatives to antibiotics for organic poultry production: types, modes of action and impacts on bird's health and production. *Poultry Sci.* (2022) 101:101696. doi: 10.1016/j.psj.2022.101696
55. Sakine Y, Ebru E, Reisli Z, Suzan Y. Effect of garlic powder on the performance, egg traits, and blood parameters of laying hens. *J Food Sci.* (2006) 86:1336–9. doi: 10.1002/jfsa.2515
56. Sandra DS, Doris D, Debrabrata B, Irene H. Botanical alternatives to antibiotics for use in organic poultry production. *Poult Sci.* (2015) 94:1419–30. doi: 10.3382/ps/pev014
57. Reda FM, El-Saadony MT, El-Rayes TK, Attia AI, El-Sayed SA, Ahmed SY, et al. Use of biological nano zinc as a feed additive in quail nutrition: biosynthesis, antimicrobial activity and its effect on growth, feed utilization, blood metabolites and intestinal microbiota. *Ital J Anim Sci.* (2021) 20:324–35. doi: 10.1080/1828051X.2021.1886001
58. Gunal M, Yayli G, Kaya O, Karahan N, Sulak O. The effects of antibiotic growth promoter, probiotic or organic acid supplementation on performance, intestinal microflora and tissue of broilers. *Poult Sci.* (2006) 5:149–55.
59. Polycarpo GV, Andretta I, Kipper M, Cruz-Polycarpo VC, Dadalt JC, Rodrigues PHM, et al. Meta-analytic study of organic acids as an alternative performance-enhancing feed additive to antibiotics for broiler chickens. *Poult Sci.* (2017) 96:3645–53. doi: 10.3382/ps/pex178
60. Biggs P, Parsons CM. The effects of several oligosaccharides on true amino acid digestibility and true metabolizable energy in cecectomized and conventional roosters. *Poult Sci.* (2007) 86:1161–5. doi: 10.1093/ps/86.6.1161
61. Awad WA, Bohm J, Razzazi-Fazeli E, Ghareeb K, Zentek J. Effect of addition of a probiotic microorganism to broiler diets contaminated with deoxyvalenol on performance and histological alterations of intestinal villi of broiler chickens. *Poult Sci.* (2006) 85:974–9. doi: 10.1093/ps/85.6.974
62. Viveros AA, Brenes M, Pizarro G, Castanb M. Effect of enzyme supplementation of a diet based on barley, and actoclave apparent digestibility, growth performance and gut morphology of broilers. *Anim Feed Sci Technol.* (1994) 48:237–51.
63. Sheiha AM, Abdelnour SA, Abd-El-Hack ME, Khafaga AF, Metwally KA, El-Saadony MT. Effects of dietary biological or chemical-synthesized nanoselenium supplementation on growing rabbits exposed to thermal stress. *Animals.* (2020) 10:430. doi: 10.3390/ani10030430
64. Reda FM, El-Saadony MT, Elnesr SS, Alagawany M, Tufarelli V. Effect of dietary supplementation of biological curcumin nanoparticles on growth and carcass traits, antioxidant status, immunity, and caecal microbiota of Japanese quails. *Animals.* (2020) 10:754. doi: 10.3390/ani10050754
65. Puvaca NV, Glamocic D, Lević J, Peric L, Milic D. Beneficial effects of phytoadditives in broiler nutrition. *World's Poult Sci J.* (2013) 69:27–34. doi: 10.1017/S0043933913000032
66. Bravo D, Pirgozliev V, Rose SPA, A. mixture of carvacrol, cinnamaldehyde, and capsicum oleoresin improves the energy utilization and growth performance of broiler chickens fed a maize-based diet. *J Anim Sci.* (2014) 92:1531–6. doi: 10.2527/jas.2013-6244

67. Pirgozliev V, Beccaccia A, Rose SP. Partitioning of dietary energy of chickens fed maize- or wheat-based diets with and without a commercial blend of phyto-genic feed additives. *J Ani Sci.* (2015) 93:1695–702. doi: 10.2527/jas.2014-8175
68. Madhupriya V, Shamsudeen P, Raj Manohar G, Senthilkumar S, Soundarapandiyam V, Moorthy M. Phyto feed additives in poultry nutrition - A review. *Int J Sci Environ Technol.* (2018) 7:815–22.
69. Abdelnour SA, Swelum AA, Salama A, Al-Ghadi MQ, Qattan SYA, Abd ElHack ME, et al. The beneficial impacts of dietary phycocyanin supplementation on growing rabbits under high ambient temperature. *Ital J Anim Sci.* (2020) 19:1046–56. doi: 10.1080/1828051X.2020.1815598
70. Ogbuewu IP, Okoro VM, Mbajiorgu CA. Meta-analysis of the influence of phytobiotic (pepper) supplementation in broiler chicken performance. *Trop Anim Health Prod.* (2020) 52:17–30. doi: 10.1007/s11250-019-02118-3
71. Abd Elkader AM, Labib S, Taha TF, Althobaiti F, Aldhahrani A, Salem HM, Saad HM, Ibrahim AFM. Phyto-genic compounds from avocado (*Persea Americana L.*) extracts; antioxidant activity, amylase inhibitory activity, the therapeutic potential of type 2 diabetes. *Saudi J Biol Sci.* (2021) 29:1428–33. doi: 10.1016/j.sjbs.2021.11.031
72. Abdel-Moneim AME, El-Saadony MT, Shehata AM, Saad AM, Aldhumri SA, Ouda SM, et al. Antioxidant and antimicrobial activities of *Spirulina platensis* extracts and biogenic selenium nanoparticles against selected pathogenic bacteria and fungi. *Saudi J Biol Sci.* (2021) 29:1197–209. doi: 10.1016/j.sjbs.2021.09.046
73. Ashour EA, Farsi RM, Alaidarous BA, Abdel-Moneim AME, El-Saadony MT, Osman AO, et al. Impacts of dietary supplementation of phycocyanin powder on growth performance, carcass traits, blood chemistry, meat quality and gut microbial activity of broilers. *Ital J Anim Sci.* (2021) 20:1357–72. doi: 10.1080/1828051X.2021.1924087
74. Peng QY, Li JD, Li Z, Duan ZY, Wu YP. Effects of dietary supplementation with oregano essential oil on growth performance, carcass traits, and jejunal morphology in broiler chickens. *Anim Feed Sci Technol.* (2016) 214:148–53. doi: 10.1016/j.anifeedsci.2016.02.010
75. Toghyani M, Toghyani M, Gheisari A, Ghalamkari G, Eghbalsaid S. Evaluation of cinnamon and garlic as antibiotic growth promoter substitutions on performance, immune responses, serum biochemical and haematological parameters in broiler chicks. *Livest Sci.* (2011) 138:167–73. doi: 10.1016/j.livsci.2010.12.018
76. Mpofo DA, Marume U, Mlambo V, Hugo A. The effects of *Lippia javanica* dietary inclusion on growth performance, carcass characteristics, and fatty acid profiles of broiler chickens. *Anim Nutri.* (2016) 2:160–7. doi: 10.1016/j.aninu.2016.05.003
77. Kirubakaran A, Moorthy M, Chitra R, Prabakar G. Influence of combinations of fenugreek, garlic, and black pepper powder on production traits of the broilers. *Vet World.* (2016) 9:470–4. doi: 10.14202/vetworld.2016.470-474
78. Goodarzi M, Nanekarani S. Effects of feeding mentha *Pulegium L.* As an alternative to antibiotics on the performance of broilers. *APCBEE Procedia.* (2014) 8:53–8. doi: 10.1016/j.apcb.2014.01.079
79. Landy N, Ghalamkari G, Toghyani M. Performance, carcass characteristics, and immunity in broiler chickens fed dietary neem (*Azadirachta Indica*) as an alternative for an antibiotic growth promoter. *Livest Sci.* (2011) 142:305–9. doi: 10.1016/j.livsci.2011.08.017
80. Zhang ZF, Cho JH, Kim IH. Effects of *Bacillus subtilis* Ubt-Mo2 on growth performance, relative immune Organ weight, gas concentration in excreta, and intestinal microbial shedding in broiler chickens. *Livest Sci.* (2013) 155:343–7. doi: 10.1016/j.livsci.2013.05.021
81. Toghyani M, Mosavi Sk, Modaresi M, Landy N. Evaluation of kefir as a potential probiotic on growth performance, serum biochemistry and immune responses in broiler chicks. *Animal Nutri.* (2015) 1:305–9. doi: 10.1016/j.aninu.2015.11.010
82. Mountzouris KC. Phyto-genic and probiotic feed additives for broilers: Evidence for growth performance links with gut performance indices. Pages 107–116 in Proceedings of the 2016 World Nutrition Forum. Erber Ag, Austria (2016).
83. Platel K, Srinivasan K. Digestive stimulant action of spices: a myth or reality? *Indian J Med Res.* (2004) 119:167–79.
84. Wati T, Ghosh TK, Syed B, Haldar S. Comparative efficacy of a phyto-genic feed additive and an antibiotic growth promoter on production performance, caecal microbial population and humoral immune response of broiler chickens inoculated with enteric pathogens. *Anim Nutri.* (2015) 1:213–9. doi: 10.1016/j.aninu.2015.08.003
85. Tazi SM, Mukhtar MA, Mohamed KA, Tabidi MH. Effect of using black pepper as natural feed additive on performance and carcass quality of broiler chicks. *Global advanced research. J Agri Sci.* (2014) 4:108–13.
86. Frankic T, Volj CM, Salobir J, Rezar V. Use of herbs and spices and their extracts in animal nutrition. *Acta Agric Slov.* (2009) 94:95–102.
87. Cuppett SL, Hall CA. Antioxidant activity of Labiatae. *Adv Food Nutri Res.* (1998) 42:245–71. doi: 10.1016/S1043-4526%2808%2960097-2
88. Jariyawanachaiikul W, Chaveerach P, Chokesajjawatee N. Antimicrobial activity of Thai-Herbal plants against food-borne pathogens *E. Coli*, *S. Aureus*, and *C. Jejuni*. *Agri. Agri Sci Procedia.* (2016) 11:20–4. doi: 10.1016/J.AASPRO.2016.12.004
89. Huyghebaert G, Ducatelle R, Van Immerseel F. An update on alternatives to antimicrobial growth promoters for broilers. *Vet J.* (2011) 187:182–8. doi: 10.1016/j.tvjl.2010.03.003
90. Li HL, Zhao PY, Lei Y, Hossain MM, Kim IH. Phytoncide, phyto-genic feed additive as an alternative to conventional antibiotics, improved growth performance and decreased excreta gas emission without adverse effect on meat quality in broiler chickens. *Livest Sci.* (2015) 181:1–6. doi: 10.1016/j.livsci.2015.10.001
91. Alceick AH, Bozkurt M, Çabuk M. The effect of a mixture of herbal essential oils, an organic acid or a probiotic on broiler performance. *S Afr J Anim Sci.* (2004) 34:217–22.
92. Miguel MG. Antioxidant and anti-inflammatory activities of essential oils: a short review. *Molecules.* (2010) 15:9252–87. doi: 10.3390/molecules15129252
93. Hammer KA, Carson CF. *Antibacterial and Antifungal Activities of Essential Oils.* In *Lipids and Essential Oils as Antimicrobial Agents; First edition*, NJ, USA, Hoboken: John Wiley & Sons, Ltd. (2011). p. 255–306.
94. Joshua JF, Bhaskar G, Sami D. Phyto-genic feed additives improve broiler feed efficiency via modulation of intermediary lipid and protein metabolism-related signaling pathways. *Poult Sci.* (2021) 100:1–11. doi: 10.1016/j.psj.2020.12.060
95. Orłowski S, Flees J, Greene ES, Ashley D, Lee SO, Yang FL, et al. Effects of phyto-genic additives on meat quality traits in broiler chickens. *J Anim Sci.* (2018) 96:3757–67. doi: 10.1093/jas/sky238
96. Flees J, Greene E, Ganguly B, Dridi S. Phyto-genic feed and water-additives improve feed efficiency in broilers via modulation of (an)orexic hypothalamic neuropeptide expression. *Neuropeptides.* (2020) 81:102005. doi: 10.3390/ani11030750
97. Lambert RJW, Proteus NS, Proteus JC, Nychas G-JE. A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol, and carvacrol. *J Appl Microbiol.* (2001) 91:453–62.
98. Yang C, Chowdhury MAK, Hou Y, Gong J. Phyto-genic compounds as alternatives to in-feed antibiotics: potentials and challenges in application. *Pathogens.* (2015) 4:137–56. doi: 10.3390/pathogens4010137
99. Zeng Z, Zhang S, Wang H, Piao X. Essential oil and aromatic plants as feed additives in non-ruminant nutrition: a review. *J Anim Sci Biotechnol.* (2015) 6:1–10. doi: 10.1186/s40104-015-0004-5
100. Soliman KM, Badeaa RI. Effect of oil extracted from some medicinal plants on different mycotoxigenic fungi. *Food Chem Toxicol.* (2002) 40:1669–75. doi: 10.1016/S0278-6915%2802%2900120-5
101. Burt S. Essential oils: their antibacterial properties and potential applications in foods- a review. *Int J Food Microbiol.* (2004) 94:223–53. doi: 10.1016/j.ijfoodmicro.2004.03.022
102. Peng M, Salaheen S, Biswas D. Animal health: global antibiotic issues. In *Encyclopedia of Agriculture and Food Systems.* Van Alfen, NK, Ed Oxford, UK: Academic Press. (2014) 1:346–57. doi: 10.1016/B978-0-444-52512-3.00187-X
103. French D. Chapter five- advances in clinical mass spectrometry. In *Advances in Clinical Chemistry.* Makowski, GS, Ed. San Francisco, CA, USA; Elsevier. (2017) 79:153–98. doi: 10.1016/bs.acc.2016.09.003
104. Chahardoli A, Jalilian F, Memariani Z, Farzaei MH, Shokoohinia Y. Analysis of organic acids. Sanches Silva A, Nabavi SF, Saedi M, Nabavi SM, Eds. In

- Recent Advances in Natural Products Analysis*. Amsterdam, The Netherlands: Elsevier; (2020). p. 767–823. doi: 10.1016/b978-0-12-816455-6.0026-3
105. Yadav AS, Kolluri G, Gopi M, Karthik K, Malik YS, Dhama K. Exploring alternatives to antibiotics as health promoting agents in poultry- a review. *J Exp Biol Agric Sci*. (2016) 4:368–83. doi: 10.18006/2016.4%2835%29.368.383
 106. Hermans D, De Laet M. Reaching genetic potential with medium chain fatty acids (MCFAs). *Int Poult Prod*. (2014) 22:7–9.
 107. Hajati H. Application of organic acids in poultry nutrition. *Int J Avian & Wildlife Biol*. (2018) 3:324–329.
 108. Adil S, Banday MT, Bhat GA, Mir MS, Rehman M. Effect of dietary supplementation of organic acids on performance, intestinal histomorphology, and serum biochemistry of broiler chicken. *Veter Med Int*. (2010) 8:1–7. doi: 10.4061/2010/479485
 109. Hassan HMA, Mohamed MA, Youssef AW, Hassan ER. Effect of using organic acids to substitute antibiotic growth promoters on performance and intestinal microflora of broilers. *Asian-Aust. J Anim Sci*. (2010) 23:1348–53. doi: 10.5713/AJAS.2010.10085
 110. Qaisrani S, Van Krimpen M, Kwakkel R, Verstegen M, Hendriks W. Diet structure, butyric acid, and fermentable carbohydrates influence growth performance, gut morphology, and cecal fermentation characteristics in broilers. *Poult Sci*. (2015) 94:2152–64. doi: 10.3382/ps/pev003
 111. Diether NE, Willing BP. Microbial fermentation of dietary protein: an important factor in diet-microbe-host interaction. *Microorganisms*. (2019) 7:1–14. doi: 10.3390/microorganisms7010019
 112. Ikker P, Dirkwager A, Fledderus J, Trevisi P, Le Huërou-Luron I, Lallès J, et al. Dietary protein and fermentable carbohydrates contents influence growth performance and intestinal characteristics in newly weaned pigs. *Livest Sci*. (2007) 108:194–7. doi: 10.1016/j.livsci.2007.01.057
 113. Partanen KH, Morz Z. Organic acids for performance enhancement in pig diets. *Nutr Res Rev*. (1999) 12:117–45. doi: 10.1079/095442299108728884
 114. Suiryanrayna MV, Ramana JV, A. Review of the effects of dietary organic acids fed to swine. *J Anim Sci Biotechnol*. (2015) 6:45. doi: 10.1186/s40104-015-0042-z
 115. Christian L, Mellor S. The use of organic acids in animal nutrition, with special focus on dietary potassium deformity under European and Austral-Asian conditions. *Recent Adv Anim Nutr Aus*. (2011) 4:123–30.
 116. Lan Y, Verstegen MWA, Tamminga S, Williams BA. The role of the commensal gut microbial community in broiler chickens. *Worlds Poult Sci J*. (2005) 61:95–104. doi: 10.1079/WPS200445
 117. Araujo, Robert GAC, Polycarpo GV, Adriano B, Kelry MS, Gabriela V, et al. Performance and economic viability of broiler chickens fed with probiotic and organic acids in an attempt to replace growth-promoting antibiotics. *Braz J Poult Sci*. (2019) 21:1–7.
 118. Christian L. Effect of dietary sodium deformity on growth performance, nutrient digestibility, gut health, and profitability in broilers. 20th European Symposium on Poultry Nutrition Poster presentation. Available online at: http://en.engormix.com/MA-poultry_industry/events/20th-european-symposium-poultry-nutrition-2015-t2254.htm
 119. Naseri KG, Rahimi S, Khaki P. Comparison of the effects of probiotics, organic acid, and medicinal plant on *Campylobacter jejuni* challenged broiler chickens. *J Agric Sci Technol*. (2012) 14:1485–96.
 120. Tschirner K, Susenbeth A, Wolfram S. Influence of Sangrovit supplementation on nitrogen balance and feed intake in growing pigs. *Ninth Symposium Vitamins and Additives in the Nutrition of Man and Animal*. Jena: Friedrich Schiller University (2003). p. 45.
 121. Vukic Vramjes M, Wenk C. Influence of Dietary Enzyme Complex on the Performance of Broilers Fed on Bites with and without Antibiotic Supplementation. *Br Poult Sci*. (1995) 36:265–75. doi: 10.1080/00071669508417774
 122. Bafundo KW, Cox LA, Bywater R. Review Lends Perspective to Recent Scientific Findings on virginiamycin, antibiotic resistance debate. *Feed Stuffs*. (2002) 75:26–7.
 123. Lan RX Li SQ, Zhao Z, An L. Sodium butyrate as an effective feed additive to improve growth performance and gastrointestinal development in broilers. *Vet Med Sci*. (2020) 6:491–9. doi: 10.1002/vms3.250
 124. Mohammadagheri N, Najafi R, Najafi G. Effects of dietary supplementation of organic acids and phytase on performance and intestinal histomorphology of broilers. *Vet Res Forum*. (2016) 7:189–95.
 125. Chowdhury R, Islam K, Khan MJ, Karim MR, Haque MN, Khatun M, et al. Effect of citric acid, avilamycin, and their combination on the performance, tibia ash, and immune status of broilers. *Poult Sci*. (2009) 88:1616–22. doi: 10.3382/ps.2009-00119
 126. Leeson S, Namkung H, Antongiovanni M, Lee EH. Effect of butyric acid on the performance and carcass yield of broiler chickens. *Poult Sci*. (2005) 84:1418–22. doi: 10.1093/ps/84.9.1418
 127. Panda AK, Rao SVR, Raju MV, Sunder GS. Effect of butyric acid on performance, gastrointestinal tract health and carcass characteristics in broiler chickens. *Asian Australas J Anim Sci*. (2009) 22:1026–31. doi: 10.5713/ajas.2009.80298
 128. Namkung H, Yu H, Gong J, Leeson S. Antimicrobial activity of butyrate glycerides toward *salmonella typhimurium* and *clostridium perfringens*. *Poult Sci*. (2011) 90:2217–22. doi: 10.3382/ps.2011-01498
 129. Song B, Li H, Wu Y, Zhen W, Wang Z, Xia Z, et al. Effect of microencapsulated sodium butyrate dietary supplementation on growth performance and intestinal barrier function of broiler chickens infected with necrotic enteritis. *Anim Feed Sci Technol*. (2017) 232:6–15. doi: 10.1016/j.anifeedsci.2017.07.009
 130. Kaczmarek SA, Barri A, Hejdysz M, Rutkowski A. Effect of different doses of coated butyric acid on growth performance and energy utilization in broilers. *Poult Sci*. (2016) 95:851–9. doi: 10.3382/ps/pev382
 131. Naela M, Reda M. Korany. Studying the effect of formic acid and potassium deformity on performance, immunity and gut health of broiler chickens. *Animal Nutr*. (2016) 2:296–302. doi: 10.1016/j.aninu.2016.08.003
 132. Kim GB, SeoC MYH, KimI K. Paik Effect of dietary prebiotic supplementation on the performance, intestinal microflora, and immune response of broilers. *Poult Sci*. (2011) 90:75–82. doi: 10.3382/ps.2010-00732
 133. Rehman A, Arif M, Sajjad N, Al-Ghadi MQ, Alagawany M, Abd El-Hack ME, et al. Dietary effect of probiotics and prebiotics on broiler performance, carcass, and immunity. *Poult Sci*. (2020) 99:6946–53. doi: 10.1016/j.psj.2020.09.043
 134. Saminathan M, Chin CS, Kalavathy R, Norhani A, Yin WH. Effects of dietary prebiotics, probiotic and synbiotics on performance, caecal bacterial populations and caecal fermentation concentrations of broiler chickens. *J Sci Food Agric*. (2014) 94:341–8. doi: 10.1002/jsfa.6365
 135. Froebel LK, Jalukar S, Lavergne TA, Lee JT, Duong T. Administration of dietary prebiotics improves growth performance and reduces pathogen colonization in broiler chickens. *Poult Sci*. (2019) 98:6668–76. doi: 10.3382/ps/pez537
 136. Emmanuel U, Medani E, Edwin P, Apeh A, Mohammed AQ, Harriet G, et al. Influence of dietary supplementation of autolyzed whole yeast and yeast cell wall products on broiler chickens. *Asian-Australas J Anim Sci*. (2020) 33:579–87. doi: 10.5713/ajas.19.0220
 137. Midilli M, Kocabach M, Alp N, Mughal O, Turan N, Yilmaz H, et al. Effects of dietary probiotic and prebiotic supplementation on growth performance and serum IgG concentration of broilers A J. *An Sci*. (2008) 38:21–7.
 138. Kazi KI. Atiqul Md, Mahedul Md, Khaled MS, Kamrul Md, Mohammad AM. Effects of selected probiotics and synbiotics on growth performance and blood-biochemical changes in broiler chickens. *J Bangladesh Agril Univ*. (2021) 19:471–6. doi: 10.5455/JBAU.120923
 139. Airat K, Fail K, Vyacheslav K, Hamit T, Maksim R, Aleksandra A, et al. Effect of normosil probiotic supplementation on the growth performance and blood parameters of broiler chickens. *Indian J Pharmaceut Edu Res*. (2020) 54:1046–55. doi: 10.5530/ijper.54.4.199
 140. Deraz SF. Synergetic effects of multispecies probiotic supplementation on certain blood parameters and serum biochemical profile of broiler chickens. *J Anim Health Prod*. (2018) 6:27–34. doi: 10.17582/JOURNAL.JAHP/2018/6.1.27.34
 141. Kumprecht I, Zobac P. The effect of probiotic preparations containing *Saccharomyces cerevisiae* and *Enterococcus faecium* in diets with different levels of beta-vitamins on chicken broiler performance. *Czech J Anim Sci UZPI*. (1998) 43:63–70.
 142. Kulshreshtha G, Rathgeber B, Stratton G, Thomas N, Evans F, Critchley A, et al. Feed supplementation with red seaweeds, *Chondrus*

- crispus, and *Sarcoditheca gaudichaudii*, affects performance, egg quality, and gut microbiota of layer hens. *Poult Sci.* (2014) 93:2991–3001. doi: 10.3382/ps.2014-04200
143. Murate LS, Paião FG, de Almeida AM, Berchieri Jr A, Shimokomaki M. Efficacy of prebiotics, probiotics, and synbiotics on laying hens and broilers challenged with *Salmonella enteritidis*. *J Poult Sci.* (2015) 52:52–6. doi: 10.2141/jpsa.0130211
 144. Davani-Davari D, Negahdaripour M, Karimzadeh I, Seifan M, Mohkam M, Masoumi SJ, et al. Prebiotics: definition, Types, sources, mechanisms, and clinical applications. *Foods.* (2019) 8:92. doi: 10.3390/foods8030092
 145. Wassie T, Niu K, Xie C, Wang H, Xin W. Extraction techniques, biological activities and health benefits of marine algae *Enteromorpha Prolifera* polysaccharide. *Front Nutr.* (2021) 8:747928. doi: 10.3389/fnut.2021.747928
 146. Coppa GV, Zampini L, Galeazzi T, Gabrielli O. Prebiotics in human milk: a review. *Dig Liver Dis.* (2006) 38:291–4. doi: 10.1016/S1590-8658(07)60013-9
 147. Bird AR, Conlon MA, Christophersen CT, Topping DL. Resistant starch, large bowel fermentation and a broader perspective of prebiotics and probiotics. *Benef Microbes.* (2010) 1:423–31. doi: 10.3920/BM2010.0041
 148. Al-Sultan S, Abdel- Raheem SM, El- Ghareeb WR, Mohamed MHA. Comparative effects of using prebiotic, probiotic, symbiotic and acidifier on growth performance, intestinal microbiology and histomorphology of broiler chicks Japanese. *J Veterinary Res.* (2016) 64:S187–95.
 149. Amrit PK, Sonali B, Daljeet SD, Eugenie N, Natália C-M, Kamil K, et al. Plant prebiotics and their role in the amelioration of diseases. *Biomolecules.* (2021) 11:1–25. doi: 10.3390/biom11030440
 150. Dallies N, Francois J, Paquet V. A. new method for quantitative determination of polysaccharides in the yeast cell wall. Application to the cell wall defective mutants of *Saccharomyces cerevisiae*. *Yeast.* (1998) 14:1297–306. doi: 10.1002/(SICI)1097-0061(1998100)14:14%3C1297::AID-YEA310%3E3.0.CO;2-L
 151. Sun HY, Jiang ZN, Shen X, Xu SX. Report on the use of veterinary antibiotics in China in 2018. *China Anim Health.* (2019) 21:8–9.
 152. Gibson GR, Fuller R. Aspects of *in vitro* and *in vivo* research approaches directed toward identifying probiotics and prebiotics for human use. *J Nutr.* (2000) 130:391–5. doi: 10.1093/jn/130.2.391s
 153. Doyle ME. *Alternatives to Antibiotic Use for Growth Promotion in Animal Husbandry*; Food Research Institute. Madison, WI: University of Wisconsin-Madison (2001).
 154. Baurhoo B, Phillip L, Ruiz-Feria CA. Effects of purified lignin and mannan oligosaccharides on intestinal integrity and microbial populations in the ceca and litter of broiler chickens. *Poult Sci.* (2007) 86:1070–8. doi: 10.1093/ps/86.6.1070
 155. Petersen CB. Comparative effects of ZooLac, Bio-MOS, and Bio-Pro on the performance of broilers to 36 days. *Poster In Biotechnology in the Feed Industry Proc Alltechs 14th Annual Symposium; Lyons, TP, Ed.* Nicholasville, KY, USA: Archivos de Medicina Veterinaria. (1998).
 156. Shashidhara RG, Devegowda G. Effect of dietary mannan oligosaccharide on broiler breeder production traits and immunity. *Poult Sci.* (2003) 82:1319–25. doi: 10.1093/ps/82.8.1319
 157. Sinovec Z, Markovic R. Use of pre-biotics in poultry nutrition. *Biotechnol Stoc.* (2005) 21:235–9.
 158. Spring P. *Effects of Mannan oligosaccharide on Different Cecal Parameters and on Cecal Concentrations of enteric Pathogens in Poultry*. PhD Thesis, Zurich, Switzerland: Swiss Fed Inst Tech. (1996).
 159. Mazanko MS, Gorlov IF, Prazdnova EV, Makarenko MS, Usatov AV, Bren AB, et al. Bacillus probiotic supplementations improve laying performance, egg quality, hatching of laying hens, and sperm quality of roosters. *Probiotics Antimicrob Proteins.* (2018) 10:367–73. doi: 10.1007/s12602-017-9369-4
 160. Neupane D, Nepali DB, Devkota N, Sharma MP, Kadaria IP. Effect of Probiotics on production and egg quality of dual-purpose chicken at Kathmundo in Nepal. *Bangladesh J Anim Sci.* (2019) 48:29–35. doi: 10.3329/bjas.v48i1.44556
 161. Yaqoob M, Abd El-Hack ME, Hassan F, El-Saadony MT, Khafaga A, Batiha G, Yehia N, Elnesr S, Alagawany M, El-Tarabily KA, Wang M. The potential mechanistic insights and future implications for the effect of prebiotics on poultry performance, gut microbiome, and intestinal morphology. *Poult Sci.* (2021) 100 7:101143. doi: 10.1016/j.psj.2021.101143
 162. Po-Yun T, Woo KK. Review: roles of prebiotics in intestinal ecosystem of broilers. *Front Vet Sci.* (2018) 5:245. doi: 10.3389/fvets.2018.00245
 163. Yang Y, Iji PA, Kocher A, Mikkelsen LL, Choct M. Effects of mannanoligosaccharide and fructooligosaccharide on the response of broilers to pathogenic *Escherichia coli* challenge. *Br Poult Sci.* (2008) 49:550–9. doi: 10.1080/00071660802290408
 164. Chee SH, Iji PA, Choct M, Mikkelsen LL, Kocher A. Characterization and response of intestinal microflora and mucins to manno-oligosaccharide and antibiotic supplementation in broiler chickens. *Br Poult Sci.* (2010) 51:368–80. doi: 10.1080/00071668.2010.503477
 165. Walker AW, Ince J, Duncan SH, Webster LM, Holtrop G, Ze X, et al. Dominant and diet-responsive groups of bacteria within the human colonic microbiota. *ISME J.* (2011) 5:220–30. doi: 10.1038/ismej.2010.118
 166. BogusŁawska-Tryk M, Piotrowska A, Burlikowska K. Dietary fructans and their potential beneficial influence on health and performance parameters in broiler chickens. *J Cent Eur Agric.* (2012) 13:270–88. doi: 10.5513/JCEA01/13.2.1045
 167. Troy EB, Kasper DL. Beneficial effects of *Bacteroides fragilis* polysaccharides on the immune system. *Front Biosci-Landmark.* (2010) 15:25–34. doi: 10.2741/3603
 168. Bonos E, Christaki E, Abraham A, Soutlos N, Florou-Paneri P. The influence of mannan oligosaccharides, acidifiers and their combination on cecal microflora of Japanese quail (*Coturnix japonica*). *Anaerobe.* (2011) 17:436–9. doi: 10.1016/j.anaerobe.2011.05.006
 169. Tizard IR. *Veterinary Immunology*. 9th ed St Louis, MO: Elsevier. (2013).
 170. Kogut MH. The gut microbiota and host innate immunity: regulators of host metabolism and metabolic diseases in poultry? *J Appl Poult Res.* (2013) 22:637–46. doi: 10.3382/japr.2013-00741
 171. Gustavo PA-M, Sandy A, Laura MB, Larissa CZ, Ricardo W, Karina RB. Pattern recognition receptors and the host cell death molecular machinery. *Front Immunol.* (2018) 9:1–19. doi: 10.3389/fimmu.2018.02379
 172. Indikova I, Humphrey TJ, Hilbert F. Survival with a helping hand: campylobacter and microbiota. *Front Microbiol.* (2015) 6:1–6. doi: 10.3389/fmicb.2015.01266
 173. Shini S, Zhang D, Aland RC Li X, Dart PJ, Callaghan MJ, Speight RE, et al. Probiotic *Bacillus amyloliquefaciens* H57 ameliorates subclinical necrotic enteritis in broiler chicks by maintaining intestinal mucosal integrity and improving feed efficiency. *Poult Sci.* (2020) 99:4278–93. doi: 10.1016/j.psj.2020.05.034
 174. Plavnik I, Scott ML. Effects of additional vitamins, minerals, or brewer's yeast upon leg weaknesses in broiler chickens. *Poult Sci.* (1980) 59:459–64. doi: 10.3382/ps.0590459
 175. Anadón A, Martínez-Larrañaga MR, Martínez M. Probiotics for animal nutrition in the European Union. regulation and safety assessment. *Regul Toxicol Pharmacol.* (2006) 45:91–5. doi: 10.1016/j.yrtph.2006.02.004
 176. Al-Khalaifa H, Al-Nasser A, Al-Surayee T, Al-Kandari S, Al-Enzi N, Al-Sharrah T, et al. Effect of dietary probiotics and prebiotics on the performance of broiler chickens. *Poult Sci.* (2019) 98:4465–79. doi: 10.3382/ps/pez282
 177. Guyard-Nicodème M, Keita A, Quesne S, Amelot M, Požévara T, Le Berre B, et al. Efficacy of feed additives against *Campylobacter* in live broilers during the entire rearing period. *Poult Sci.* (2016) 95:98–305. doi: 10.3382/ps/pev303
 178. Peralta-Sánchez JM, Martín-Platero AM, Ariza-Romero JJ, Rabelo-Ruiz M, Zurita-González MJ, Baños A, et al. Egg production in poultry farming is improved by probiotic bacteria. *Front Microbiol.* (2019) 10:1042. doi: 10.3389/fmicb.2019.01042
 179. Qamar A, Waheed J, Hamza A, Mohyuddin SG, Lu Z, Namula Z, et al. Chen JJ. The role of intestinal microbiota in chicken health, intestinal physiology, and immunity. *J Anim Plant Sci.* (2020) 31:342–51. doi: 10.36899/JAPS.2021.2.0221
 180. Wu Y, Wang B, Zeng Z, Liu R, Tang L, Gong L, et al. Effects of probiotics *Lactobacillus plantarum* 16 and *Paenibacillus polymyxa* 10 on intestinal barrier function, antioxidative capacity, apoptosis, immune response, and biochemical parameters in broilers. *Poult Sci Poultry Science Association Inc.* (2019) 98:5028–39. doi: 10.3382/ps/pez226
 181. Singh AK, Tiwari UP, Berrocoso JD, Dersjant-Li Y, Awati A, Jha R. Effects of a combination of xylanase, amylase and protease, and probiotics on

- major nutrients including amino acids and non-starch polysaccharides utilization in broilers fed different level of fibers. *Poult Sci.* (2019) 98:5571–81. doi: 10.3382/ps/pez310
182. Wealleans AL, Walsh MC, Romero LF, Ravindran V. Comparative effects of two multi-enzyme combinations and a *Bacillus* probiotic on growth performance, digestibility of energy and nutrients, disappearance of non-starch polysaccharides, and gut microflora in broiler chickens. *Poult Sci.* (2017) 96:4287–97. doi: 10.3382/ps/pex226
 183. De Cesare A, Caselli E, Lucchi A, Sala C, Parisi A, Manfreda G, et al. Impact of a probiotic-based cleaning product on the microbiological profile of broiler litters and chicken caeca microbiota. *Poult Sci.* (2019) 98:3602–10. doi: 10.3382/ps/pez148
 184. Patel SG, Raval AP, Bhagwat SR, Sadrasaniya DA, Patel AP, Joshi SS. Effects of probiotics supplementation on growth performance, feed conversion ratio, and economics of broilers. *J Ani Res.* (2015) 5:155–60.
 185. Mizak L, Gryko R, Kwiatek M, Parasion S. Probiotics in animal nutrition. *Zycie Weter.* (2012) 87:736–42.
 186. Bajagai YS, Klieve AV, Peter JD, BW L. Probiotics in animal nutrition-production, impact, and regulation. *Makkar HPS, editor FAO Anim Prod Heal FAO Animal Production and Health.* (2016) 179:1–89.
 187. Markowiak P, Slizewska K. The role of probiotics, prebiotics, and synbiotics in animal nutrition. *Gut Pathog BioMed Central.* (2018) 10:1–20. doi: 10.1186/s13099-018-0250-0
 188. Zommiti M, Chikindas ML, Ferchichi M. Probiotics-live biotherapeutics: a story of success, limitations, and future prospects-not only for humans. Probiotics antimicrob proteins. *Probio Antimicro Prot.* (2020) 12:1266–89. doi: 10.1007/s12602-019-09570-5
 189. Khan RU, Naz S. The applications of probiotics in poultry production. *Worlds Poult Sci J.* (2013) 69:621–32. doi: 10.1017/S0043933913000627
 190. AlFatah MA. Probiotic modes of action and its effect on biochemical parameters and growth performance in poultry. *Iran J Appl Anim Sci.* (2020) 10:9–15.
 191. Sandvang D, Skjoet-Rasmussen L, Cantor MD, Mathis GF, Lumpkins BS, Blanch A. Effects of feed supplementation with 3 different probiotic *Bacillus* strains and their combination on the performance of broiler chickens challenged with *Clostridium perfringens*. *Poult Sci.* (2021) 100:1–10. doi: 10.1016/j.psj.2021.01.005
 192. Abd El-Hack ME, Alaidarous BA, Farsi RM, Abou-Kassem DE, El-Saadony MT, Saad AM, et al. Impacts of supplementing broiler diets with biological curcumin, zinc nanoparticles, and *Bacillus licheniformis* on growth, carcass traits, blood indices, meat quality, and cecal microbial load. *Animals.* (2021) 11:1–24. doi: 10.3390/ani11071878
 193. Celina EB, Tamiris NSS. Probiotics as a Promising Additive in Broiler Feed: Advances and Limitations, *Advances in Poultry Nutrition Research.* (2021).
 194. Wang C, Shi C, Su W, Jin M, Xu B, Hao L, et al. Dynamics of the physicochemical characteristics, microbiota, and metabolic functions of soybean meal and corn mixed substrates during two-stage solid-state. *App Environmen Sci.* (2020) 5:1–17. doi: 10.1128/mSystems.00501-19
 195. Alagawany, Elnesr, Farag. The role of exogenous enzymes in promoting growth and improving nutrient digestibility in poultry. *IJVR.* (2018) 19:157–64.
 196. Ojha BK, Singh PK, Shrivastava N. Enzymes in the Animal Feed Industry. In *Enzymes in Food Biotechnology.* Mohammed, K., Ed.; Cambridge, MA, USA: Academic Press (2019). p. 93–109. doi: 10.1016/B978-0-12-813280-7.00007-4
 197. Velázquez-De Lucio BS, Hernández-Domínguez EM, Villa-García M, Díaz-Godínez G, Mandujano-González V, Mendoza-Mendoza B, et al. Exogenous enzymes as zootechnical additives in animal feed: a review. *Catalysts.* (2021) 11:851. doi: 10.3390/catal11070851
 198. Ravindran V. Feed enzymes: the science, practice, and metabolic realities. *J Appl Poult Res.* (2013) 22:628–36. doi: 10.3382/japr.2013-00739
 199. Amerah A, Romero L, Awati A, Ravindran V. Effect of exogenous xylanase, amylase, and protease as single or combined activities on nutrient digestibility and growth performance of broilers fed corn/soy diets. *Poult Sci.* (2017) 96:807–16. doi: 10.3382/ps/pew297
 200. Shimaa A, Mohamed A, Ahmed A, Ahmed A. Saleh, Shafika A, Doaa M, et al. Effect of dietary supplementation of alpha-galactosidase on the growth performance, ileal digestibility, intestinal morphology, and biochemical parameters in broiler chickens. *BMC Vet Res.* (2020) 16:144. doi: 10.1186/s12917-020-02359-7
 201. Rao S, Raju M, Nagalakshmi D, Prakash B, Paul S. Effect of supplementation of graded concentrations of xylanase and α -amylase on performance, slaughter variables and energy digestibility in broiler chicken fed corn-soybean meal based diet. *J App Poult Res.* (2021) 30:100139. doi: 10.1016/j.japr.2021.100139
 202. Cowieson A, Bedford M, Ravindran V. Interactions between xylanase and glucanase in maize-soy-based diets for broilers. *Br Poult Sci.* (2010) 51:246–57. doi: 10.1080/00071661003789347
 203. Rambabu D, Reddy Ravinder V, Qudratullah S, Reddy M, Reddy K. Effect of exogenous enzyme supplementation on certain growth and carcass characteristics of broiler chicks. *Indian J Poult Sci.* (2011) 46:189–94.
 204. Saeed M, Ayaşan T, Alagawany M, MEA El-HackMA, Abdel-Latif, AK Patra. The role of β -mannanase (Hemicell) in improving poultry productivity, health and environment. *Braz J Poult Sci.* (2019) 21:1–8. doi: 10.1590/1806-9061-2019-1001
 205. Alhidary IA, Abdelrahman MA, Albadani H, Khan RU, Selvaggi M, Laudadio V, et al. Impact of microbial protease enzyme and dietary crude protein levels on growth and nutrients digestibility in broilers over 15–28 days. *Animals.* (2021) 11:2499. doi: 10.3390/ani11092499
 206. Alshamiri MMA, Ali SAM, Abdalla HO, Ahmed HB. The effect of supplementing different levels of phytase enzyme on performance, some carcass properties and economics of broiler chickens. *Agrobiological Records.* (2021) 4:14–22. doi: 10.47278/journal.abr/2020.025
 207. Adeola O, Cowieson AJ. Board-invited review: Opportunities and challenges in using exogenous enzymes to improve non-ruminant animal production. *J Anim Sci.* (2011) 89:3189–218. doi: 10.2527/jas.2010-3715
 208. Ugwuanyi JO. Enzymes for nutritional enrichment of agro-residues as livestock feed. In: Gurpreet SD, Surinder K, editors. *Agro-Industrial Wastes as Feedstock for Enzyme, Production.* Cambridge, MA: Academic Press. (2016). pp. 233–60. doi: 10.1016/B978-0-12-802392-1.00010-1
 209. Doskovic V, Bogosav, Jevic-Boskovic S, Pavlovski Z, Milosevic B, Skrbic Z, et al. Enzymes in broiler diets with special reference to protease. *World's Poult Sci J.* (2013) 69:343–60. doi: 10.1017/S0043933913000342
 210. Khattak FM, Pasha TN, Hayat Z, Mahmud A. Enzymes in poultry nutrition. *J Anim Plant Sci.* (2006) 16:1–2.
 211. Mabelebele M, Gous RM, Siwela M, O'Neil H, Iji P. Performance of broiler chickens fed South African sorghum-based diets with xylanase. *S Afr J Anim Sci.* (2017) 47:679–87.
 212. Babalola TO, Apata DF, Atteh JO. Effect of β -xylanase supplementation of boiled castor seed meal-based diets on the performance, nutrient absorbability and some blood constituents of pullet chicks. *Trop Sci.* (2006) 46:216–23. doi: 10.1002/ts.181
 213. Café MB, Borges CA, Fritts CA, Waldroup PW. Avizyme improves performance of broilers fed corn-soybean meal-based diets. *J Appl Poult Res.* (2002) 11:29–33. doi: 10.1093/japr/11.1.29
 214. Ohimain EI, Ofongo R. Enzyme supplemented poultry diets: benefits so far-A review. *Int J Adv Biotechnol Res.* (2014) 3:31–9.
 215. Tuoying AO. *Exogenous Enzymes and Organic Acids in the Nutrition of Broiler Chicks: Effects on Growth Performance and in vitro and in vivo Digestion.* Ph D Thesis, The Graduate School University of Kentucky (2005).
 216. Jia W, Slominski BA, Bruce HL, Blank G, Crow G, Jones O. Effects of diet type and enzyme addition on growth performance and gut health of broiler chickens during sub clinical clostridium perfringens challenge. *Poult Sci.* (2009) 88:132–40. doi: 10.3382/ps.2008-00204
 217. Knudsen KEB. Fibre and non-starch polysaccharide content and variation in common crops used in broiler diets. *Poult Sci.* (2014) 93:2380–93. doi: 10.3382/ps.2014-03902
 218. Courtin C, Broekaert W, Swenen K, Lescaort O, Onagbesan O, Buyse J, et al. Dietary inclusion of wheat bran arabinoxylooligosaccharides induces beneficial nutritional effects in chickens. *Cereal Chem.* (2008) 85:607–13. doi: 10.1094/CCHEM-85-5-0607
 219. Chen H, Liu S, Xu XR, Zhou GJ, Liu SS, Yue WZ. Antibiotics in the coastal environment of the hailing bay region, South China Sea: spatial distribution, source analysis and ecological risks. *Mar Pollut Bull.* (2015) 95:365–673. doi: 10.1016/j.marpollbul.2015.04.025

220. Zhang L, Xu J, Lei L, Jiang Y, Gao F, Zhou G. Effects of Xylanase supplementation on growth performance, nutrient digestibility and non-starch polysaccharide degradation in different sections of the gastrointestinal tract of broilers. *Asian Austr J Ani Sci.* (2014) 27:855–86. doi: 10.5713/ajas.2014.14006
221. Lee S, Apajalahti K, Vienola G, Gonzalez-Ortiz C, Bedford M. Age and dietary xylanase supplementation affects ileal sugar residues and short chain fatty acid concentration in the ileum and caecum of broiler chickens. *Ani Feed Sci Technol.* (2017) 234:29–42. doi: 10.1016/j.anifeedsci.2017.07.017
222. Ravangard A, Houshmand M, Khajvi K, Naghihi R. Performance and cecal bacteria counts of broilers fed low protein diets with and without a combination of probiotic and prebiotic. *Br J Poul Sci. Nutrition.* (2017) 3:75–82. doi: 10.1590/1806-9061-2016-0319
223. Shengru W, Li T, Niu H, Zhu Y, Liu Y, Duan Y, et al. Effects of glucose oxidase on growth performance, gut function, and cecal microbiota of broiler chickens. *Poult Sci.* (2019) 98:828–41. doi: 10.3382/ps/pey393
224. Simbaya J, Slominski B, Guenter W, Morgan A, Campbell L. The effects of protease and carbohydrase supplementation on the nutritive value of canola meal for poultry: *in vitro* and *in vivo* studies. *Anim Feed Sci Technol.* (1996) 61:219–34. doi: 10.1016/0377-8401(95)00939-6
225. Zanella I, Sakomura N, Silversides F, Figueirido A, Pack M. Effect of enzyme supplementation of broiler diets based on maize and soybeans. *Poult Sci.* (1999) 78:561–8. doi: 10.1093/ps/78.4.561
226. Cowieson J, Adeola O. Carbohydrases, protease, and phytase have an additive beneficial effect in nutritionally marginal diets for broiler chicks. *Poult Sci.* (2005) 84:1860–7. doi: 10.1093/ps/84.12.1860

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Ayalew, Zhang, Wang, Wu, Qiu, Qi, Tekeste, Wassie and Chanie. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.